



University of Groningen

Glucose and glycogen metabolism in cancer

de Heer, Ellen

DOI: 10.33612/diss.797819115

IMPORTANT NOTE: You are advised to consult the publisher's version (publisher's PDF) if you wish to cite from it. Please check the document version below.

Document Version Publisher's PDF, also known as Version of record

Publication date: 2023

Link to publication in University of Groningen/UMCG research database

Citation for published version (APA): de Heer, E. (2023). *Glucose and glycogen metabolism in cancer: potentials for intervention and patient selection.* [Thesis fully internal (DIV), University of Groningen]. University of Groningen. https://doi.org/10.33612/diss.797819115

Copyright

Other than for strictly personal use, it is not permitted to download or to forward/distribute the text or part of it without the consent of the author(s) and/or copyright holder(s), unless the work is under an open content license (like Creative Commons).

The publication may also be distributed here under the terms of Article 25fa of the Dutch Copyright Act, indicated by the "Taverne" license. More information can be found on the University of Groningen website: https://www.rug.nl/library/open-access/self-archiving-pure/taverneamendment.

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

Downloaded from the University of Groningen/UMCG research database (Pure): http://www.rug.nl/research/portal. For technical reasons the number of authors shown on this cover page is limited to 10 maximum.

Y. C.

Mapping heterogeneity in glucose uptake in metastatic melanoma using quantitative ¹⁸F-FDG PET/CT analysis

Ellen C. de Heer, Adrienne H. Brouwers, Ronald Boellaard, Wim J. Sluiter, Gilles F. H. Diercks, Geke A. P. Hospers, Elisabeth G. E. de Vries and Mathilde Jalving



EJNMMI Res. 2018;8:101

ABSTRACT

Background

Metastatic melanoma patients can have durable responses to systemic therapy and even long-term survival. However, a large subgroup of patients does not benefit. Tumour metabolic alterations may well be involved in the efficacy of both targeted and immunotherapy. Knowledge on in vivo tumour glucose uptake and its heterogeneity in metastatic melanoma may aid in upfront patient selection for novel (concomitant) metabolically targeted therapies. The aim of this retrospective study was to provide insight into quantitative ¹⁸F-fluorodeoxyglucose positron emission tomography/computed tomography (¹⁸F-FDG PET/CT) parameters and corresponding intra- and inter-patient heterogeneity in tumour ¹⁸F-FDG uptake among metastatic melanoma patients. Consecutive, newly diagnosed stage IV melanoma patients with a baseline ¹⁸F-FDG PET/CT scan performed between May 2014 and December 2015 and scheduled to start first-line systemic treatment were included. Volume of interests (VOIs) of all visible tumour lesions were delineated using a gradient-based contour method, and standardized uptake values (SUVs), metabolically active tumour volume (MATV) and total lesion glycolysis (TLG) were determined on a per-lesion and per-patient basis. Differences in quantitative PET parameters were explored between patient categories stratified by BRAFV600 and RAS mutational status, baseline serum lactate dehydrogenase (LDH) levels and tumour programmed death-ligand 1 (PD-L1) expression.

Results

In 64 patients, 1143 lesions \geq 1 ml were delineated. Median number of lesions \geq 1 ml was 6 (range 0–168), median maximum SUVpeak 9.5 (range 0–58), median total MATV 29 ml (range 0–2212) and median total TLG 209 (range 0–16,740). Per-patient analysis revealed considerable intra- and inter-patient heterogeneity. Maximum SUVs, MATV, number of lesions and TLG per patient did not differ when stratifying between *BRAFV600* or *RAS* mutational status or PD-L1 expression status, but were higher in the patient group with elevated LDH levels (> 250 U/I) compared to the group with normal LDH levels (P < 0.001). A subset of patients with normal LDH levels also showed above median tumour ¹⁸F-FDG uptake.

Conclusions

Baseline tumour ¹⁸F-FDG uptake in stage IV melanoma is heterogeneous, independent of mutational status and cannot be fully explained by LDH levels. Further investigation of the prognostic and predictive value of quantitative ¹⁸F-FDG PET parameters is of interest.

BACKGROUND

Novel therapies, especially immunotherapy, have revolutionized the treatment of stage IV metastatic melanoma over the past decade. One-year overall survival (OS) rates have improved to 50–75% and a subset of patients shows durable responses¹. Still, a considerable number of patients do not respond, especially those with elevated serum lactate dehydrogenase (LDH) levels¹.

The metabolic reprogramming that characterizes cancer cells may well be involved in the efficacy of antitumour immune responses^{2,3}. Cancer cells metabolize a substantial amount of the consumed glucose through glycolysis only—even under aerobic conditions—in order to generate sufficient biomass for rapid cellular proliferation^{4,5}. Novel therapeutic agents interfering with this altered glucose metabolism have shown hints of anticancer activity in (pre)clinical studies, for example in breast cancer, non-small cell lung cancer and glioblastoma^{6,7}. Additionally, preclinical data suggest metabolically targeted therapies can improve antitumour immune response and susceptibility to adjuvant chemo- and radiotherapy⁸⁻¹⁰. In patients, however, such treatments can result in toxicity in highly glucose-dependent healthy tissues, such as the kidney^{7,11}. Furthermore, recent in vitro studies demonstrate that not all melanomas rely on altered glucose metabolic pathways to the same extent^{12,13}. This underlines the need for upfront selection of patients with highly glucose-dependent tumours in order to maximize the benefit of (concomitant) metabolic therapies and ensure a sufficiently broad therapeutic window.

Metastatic melanoma is clinically renowned for its high uptake of the glucose analogue ¹⁸F-fluorodeoxyglucose (¹⁸F-FDG) on positron emission tomography/computed tomography (PET/CT) scans. Whole-body ¹⁸F-FDG PET/CT is therefore part of standard care staging procedures at baseline in stage IV disease, where it is used in a qualitative fashion to provide information on the presence and location of metastases. However, quantitative ¹⁸F-FDG PET/CT scan analysis has been completely unvisited in stage IV melanoma so far and could provide a wealth of knowledge on quantitative tumour glucose uptake in vivo, potentially useful for upfront patient selection for metabolically targeted therapies. The aim of this retrospective study was to provide an overview of tumour ¹⁸F-FDG uptake and corresponding intra- and inter-patient heterogeneity in metastatic melanoma patients using quantitative ¹⁸F-FDG PET/CT scan analysis.

PATIENTS AND METHODS

Patients

Patients for this retrospective study were selected from a prospectively maintained database containing all melanoma patients registered at the Department of Medical Oncology of the University Medical Center Groningen (UMCG), the Netherlands, from 2012 onwards. All patients \geq 18 years of age with histologically proven cutaneous or mucosal metastatic melanoma (American Joint Committee on Cancer [AJCC] 7th edition stage IV melanoma¹⁴) without prior systemic treatment and with a baseline ¹⁸F-FDG PET/CT scan performed between May 2014 and December 2015 were eligible for inclusion (n = 108). Exclusion criteria were unknown or inadequate adherence to European Association of Nuclear Medicine (EANM) PET/CT scan acquisition guidelines¹⁵ (e.g. PET/CT scan not performed at our hospital) (n = 26), no indication for start of first-line systemic treatment within 2 months of baseline PET/CT scan (n = 10), concurrent malignancy or other malignancy within the previous 10 years (n = 5) and/or no PET-positive lesions (n = 3). Ultimately, 64 patients were included (SUPPL. FIGURE 1). The Medical Ethics Committee approved the study. Consultation of the local objection registry verified that none of the selected patients had objected to use of their personal data for research purposes. Patients were pseudonymized, and data were stored on a secured server following local data management regulations.

¹⁸F-FDG PET/CT imaging

¹⁸F-FDG PET/CT scans were acquired using a Siemens Biograph mCT PET/CT system (Siemens/CTI, Knoxville, TN) accredited by the European Association of Nuclear Medicine (EANM) Research Limited (EARL). Scan acquisition and reconstructions were performed following the recommendations of the EANM guideline for oncology ¹⁸F-FDG imaging¹⁵. Patients were instructed to fast and avoid exercise at least 4–6h prior to intravenous ¹⁸F-FDG injection at an activity of 3 MBq/kg. Serum glucose levels before tracer injection were < 8.3 mmol/l. Whole-body PET/CT scanning (from the top of the skull to the bottom of the feet) was performed 60 min after ¹⁸F-FDG injection with 1–3 min per bed position. Prior to the PET acquisition, patients underwent a low-dose CT (LD-CT) scan during tidal breathing for attenuation correction (80–140 kVp, quel. ref. 30 mAs and pitch of 1).

¹⁸F-FDG PET/CT scan analysis and volume of interest delineation

All PET/CT scans were initially reported by a nuclear medicine physician as part of routine patient care. Quantitative scan analysis and identification and delineation of all tumour lesions for this study were performed by one investigator (EH) and verified by a board-certified nuclear medicine physician with expertise in melanoma (AB).

PET(/CT) and gradient PET images were displayed side-to-side, and volume of interests (VOIs) were delineated on the gradient PET images using a gradient-based manual contouring method (in-house developed software program). Gradient PET images are derived directly from reconstructed PET images and depict the relative change in counts between neighbouring voxels (Δ standardized uptake value [SUV]), which is typically the highest around tumour borders. Gradient PET images consequently provide an image where the borders of the lesion are most intense. Use of gradient PET data enables a (manual) VOI delineation method where lesion border location is independent of colour scale, in contrast to manual contouring on regular PET images. Additional motives for choosing gradient-based delineation were a lack of systematic delineation studies in metastatic melanoma and inaccuracy of EARL-recommended semi-automatic delineation methods for delineation of large heterogeneous tumour lesions or small yet highly ¹⁸F-FDG-avid lesions¹⁵.

A region of interest (ROI) was manually drawn around each tumour lesion on consecutive transaxial slices. Subsequently, the observer adjusted a %-threshold based on maximum SUV (SUVmax) until the VOI borders optimally corresponded with the location of the steepest gradient on the gradient PET images as judged visually. SUVmax, mean SUV (SUVmean), peak SUV (SUVpeak, i.e. a 1.2-cm³ spheric region positioned to yield the highest average value), metabolically active tumour volume (MATV) and total lesion glycolysis (TLG, the product of SUVmean and MATV) were determined for each VOI. SUVs were corrected for serum glucose level and lean body mass according to the Janmahasatian

TABLE 1 | Patient characteristics.

Characteristic	All patients (n = 64)
Gender	
Male	40 (62.5%)
Female	24 (37.5%)
Age (years) at baseline PET/CT	59 (45-69) (range 25-80)
o o	AE (70 3%)
1	7 (10 9%)
>2	7 (11.0%)
Missina	5 (7.8%)
Histology primary melanoma	
Cutaneous	47 (73.4%)
Mucosal	4 (6.3%)
Primary melanoma unknown/missing	13 (20.3%)
M-stage at baseline PET/CT	
M1a	1 (1.6%)
M1b	2 (3.1%)
MIC	61 (95.3%)
1	3 (4 7%)
2	6 (9 4%)
>2	55 (85.9%)
Organ involvement	
(Sub)cutaneous	39 (60.9%)
Lymph nodes	54 (84.4%)
Lungs	40 (62.5%)
Muscular	25 (39.1%)
Skeletal	39 (60.9%)
	24 (37.5%) 30 (46.9%)
Other ^c	14 (21 9%)
Brain metastases ^d	14 (21.370)
Yes	22 (34.4%)
¹⁸ F-FDG-avid ^e	11 (17.2%)
Not ¹⁸ F-FDG-avid	11 (17.2%)
No	35 (54.7%)
Missing	7 (10.9%)
BRAF mutation status	74 640 400
BRAFV600 mutation	31 (48.4%)
No BRAFV600 mutation	33 (51.6%)
RAS mutation ^f	15 (23.4%)
No R4S mutation	49 (76 6%)
Baseline serum LDH (U/I)	246 (192–327) (range 92–11.371)
Normal	35 (54.7%)
Elevated ⁹	28 (43.7%)
>1-2× ULN	23 (35.9%)
>2×ULN	5 (7.8%)
Missing	1 (1.6%)
Interval between baseline PET/CT and LDH measurement (days)	0 (–7 to +3) (range – 39 to +11)

Data are displayed as n (%) or median (interquartile range). Including brain metastases Number of patients with lesions in the abdominal cavity/peritoneum (n = 27; 42.2% of all patients), adrenal gland (n = 12; 18.8%), bowel (n = 6; 9.4%), spleen (n = 3; 4.7%), kidney (n = 2; 3.1%), gallbladder (n = 1; 1.6%), stomach (n = 1; 1.6%), rectum (n = 1; 1.6%) and/or pancreas (n = 1; 1.6%) Number of patients with lesions in the vaginal or nasal mucosa (n = 4; 6.3%), myelum (n = 1; 1.6%), shoulder joint (n = 2; 3.1%), breast (n = 2; 3.1%), pericardium (n = 3; 4.7%), heart (n = 2; 3.1%) and/or abdominal or thoracic wall of undetermined tissue of origin (n = 2; 3.1%) dBased on MRI brain (n = 53) or, when missing, contrast enhanced CT (n = 4) el.e. distinguishable from normal brain tissue NRAS (n = 14) and KRAS (n = 1) el.e. > 250 U/I. LDH = lactate dehydrogenase; ULN = upper limit of normal

formula¹⁵.

Lesions with an MATV < 1 ml were excluded from the final quantitative analysis to prevent partial volume effects. PET parameters were analysed on a per-patient, per-location and per-lesion basis. Patient's maximum SUV and median SUV reflect respectively the highest and median value derived from all lesions \geq 1 ml within that patient. Interquartile range (IQR) SUVpeak was derived from the SUVpeaks of all individual lesions delineated in one patient as a measure for intra-patient ¹⁸F-FDG uptake heterogeneity. Total MATV or total TLG equals the sum of respectively MATV or TLG of all lesions \geq 1 ml within that patient.

CT and brain MRI scan analysis

Previously, PET-negative (i.e. with SUVmax < 1.5) melanoma metastases have been described, and we excluded three eligible patients upfront due to the presence of only PET-negative lesions¹⁶. Therefore, we aimed to evaluate the first 20 included patients for the presence of PET-negative lesions with a diameter ≥ 1 cm on baseline contrast-enhanced CT (ce-CT) scan performed within 1 month of the baseline PET/CT. ce-CT scan was available in 12 of the 20 patients and revealed only 2 additional ¹⁸F-FDG PET-negative lesions ≥ 1 cm on top of the total of 491 PET-positive lesions > 1 ml in these patients (0.4%). Due to this limited additional value, ce-CT analysis was omitted for the remaining patients.

High physiological background ¹⁸F-FDG uptake prevents accurate detection and quantification of brain metastases. Therefore, the presence of brain lesions was additionally evaluated on baseline cerebral MRI scans or cerebral ce-CT. Quantitative data from brain lesions were not incorporated in per-patient PET parameters. When brain lesions were measurable (longest axis on MRI > 1 cm according to Response Assessment in Neuro-Oncology Brain Metastases [RANO-BM] criteria¹⁷) and ¹⁸F-FDG-avid, SUVpeak and SUVmax were measured.

Data acquisition

Patient and tumour characteristics, baseline serum LDH levels and respectively tumour *BRAF* and *RAS* mutation status were retrospectively determined from the electronic patient file. Pre-treatment serum LDH levels were derived from the date closest to the baseline PET/CT scan. When pre-treatment archival tumour biopsies for a distant metastasis were available, PD-L1 immunohistochemistry (IHC) was performed as previously described elsewhere using the 22C3 anti-PD-L1 antibody (DAKO, Merck) on Ventana BenchMark ULTRA platform¹⁸. Tissue derived from primary melanomas, local recurrences, in-transit cutaneous metastases or lymph node metastases was excluded. Scoring was performed by two board-certified pathologists (GFHD, NAH) and performed according to the manufacturer's instructions.

Statistical analysis

Variables were assessed for normal distribution by Q-Q plots. Independent Mann-Whitney U tests were used to assess differences in PET parameters between LDH, *BRAF* and *RAS* groups, respectively, and Kruskal-Wallis tests for differences between metastatic locations and PD-L1 expression groups. Spearman's rank correlation was used for the correlation between lesion MATV and SUVpeak. A P-value < 0.05 (two-sided) was considered statistically significant. Statistical analysis was performed using SPSS Statistics, version 23.0 (IBM

Corp., Armonk, NY).

RESULTS

PET parameters on a per-patient basis

Patient characteristics are presented in TABLE 1. *BRAFV600* mutational status did not differ between patients with normal or elevated serum LDH (42.9% vs. 57.1%; P = 0.260). Patient's maximum SUVpeak showed a broad range (0–58; median 9.5) between patients (FIGURE 1 and TABLE 2). Furthermore, intra-patient ¹⁸F-FDG uptake heterogeneity was observed, with SUVpeak IQR ranging from 0 to 42.4 (median 2.1). The number of lesions, SUVs, total MATV and total TLG (per-patient basis) did not differ between *BRAFV600* mutant vs. wild-type patients (TABLE 3), *RAS* mutated vs. wild-type patients and *BRAFV600/RAS* mutant vs. *BRAFV600+RAS* wild-type patients (data not shown). Patients with an elevated LDH level (> 250 U/I) had more lesions ≥ 1 ml (median 17 vs. 4, P < 0.001), a higher total MATV (127 vs. 14 ml, P < 0.001), higher maximum SUVpeak (13.3 vs. 8.7, P = 0.011), SUVmax (15.8 vs. 11.3, P = 0.026) and SUVmean (9.0 vs. 6.0, P = 0.009) and higher total TLG (1180 vs. 67, P < 0.001) (TABLE 3). Of the 13 tumour specimens that were available for PD-L1 IHC, 4 showed < 1% PD-L1 expression, 3 1–49% and 6 ≥ 50%. PD-L1 expression status did not correlate with any of the PET parameters (data not shown).

	All patients (n = 64)	Range	
No. of lesions			
All	18 (11–51)	1–417	
≥1 ml ^{a,b}	6 (2-16)	0-168	
SUVpeak			
Maximum	9.5 (5.5–15.5)	0-58.3	
Median	4.3 (3.2-8.6)	0-25.2	
SUVpeak interquartile range ^c	2.1 (0-5.1)	0-42.4	
SUVmax			
Maximum	11.8 (7.3-18.0)	0-67.2	
Median	6.1 (4.1-11.5)	0-37.6	
SUVmean			
Maximum	7.2 (4.7-10.7)	0-30.2	
Median	4.3 (3.0-7.3)	0-18.5	
Total MATV (ml)	29.2 (12.2-234)	0-2212	
Total TLG	209 (46.2-1510)	0-16,740	

 TABLE 2 | 18F-FDG PET tumour lesion parameters on a per-patient basis.

Data are displayed as median (interquartile range). ^aThree patients had only lesions < 1 ml. ^bl.e. all lesions included in quantitative analyses. ^cl.e. interquartile range of the different SUVpeaks measured within one patient, measure of intra-patient heterogeneity.

PET parameters on a per-lesion and per-location basis

In total, 3408 tumour lesions were delineated, of which 1143 had an MATV \geq 1 ml. Median lesion SUVpeak was 5.0 (range 0–58), median MATV was 2.4 ml (range 1.0–1921) and median TLG 11 (range 1.1–11,206) (SUPPL. TABLE 1). Lesion SUVpeak and MATV were mode-rately correlated (correlation coefficient 0.521, P < 0.001). The highest numbers of separate lesions were observed in bone (n = 504, 44% of all lesions \geq 1 ml), liver (n = 241, 21%) and lymph nodes (n = 125, 11%) (SUPPL. FIGURE 2A), and total measured MATV was highest in the abdomen (5683 ml, 39%) followed by bone (3864 ml, 27%) and lymph nodes (2321 ml, 16%) (SUPPL. FIGURE 2B). No major differences between metastatic locations concerning individual lesion's MATV and SUVpeak were observed (SUPPL. FIGURE 3).

Brain metastases were present in 22 patients, and 16 had measurable disease according to RANO-BM criteria¹⁷. In 11 of these patients, brain metastases were visible as hypermetabolic lesions on ¹⁸F-FDG PET/CT. Median SUVpeak and SUVmax of these lesions were 7.2 (range 4.8–36.8) and 9.0 (6.5–45.4), respectively.

	LDH groups ^a		P-value	BRAFV600 groups		P-value
	NORMAL [⊾] (N = 35)	ELEVATED (N = 28)		WILD-TYPE (N = 33)	MUTANT (N = 31)	
No. of lesions						
All	13 (7-17)	46 (20-140)	< 0.001	17 (10-34)	18 (11-64)	0.510
≥1 ml°	4 (2-6)	17 (7-48)	< 0.001	6 (3-14)	6 (2-26)	0.984
SUVpeak						
Maximum	8.7 (4.4-13.1)	13.3 (7.1-23.5)	0.011	10.1 (5.6-18.9)	8.8 (5.2-13.9)	0.310
Median	3.9 (2.7-7.4)	5.5 (3.5-8.9)	0.203	5.3 (3.3-9.0)	4.0 (3.2-8.3)	0.317
SUVpeak interquartile range ^d	1.2 (0-3.3)	3.3 (1.5-6.7)	0.002	2.7 (0-5.1)	2.0 (0-4.7)	0.380
SUVmax						
Maximum	11.3 (6.1-16.6)	15.8 (9.0-27.7)	0.026	13.0 (7.3-22.2)	11.6 (7.2-17.4)	0.344
Median	5.3 (3.7-9.4)	8.0 (4.9-12.0)	0.171	7.4 (4.5-11.7)	5.3 (4.0-11.7)	0.394
SUVmean						
Maximum	6.0 (4.1-8.7)	9.0 (6.1-15.4)	0.009	8.4 (4.7-12.3)	6.9 (4.7-8.8)	0.274
Median	3.9 (2.8-6.2)	5.3 (3.4-7.6)	0.128	4.8 (3.2-7.9)	3.9 (2.9-7.1)	0.256
Total MATV (ml)	14 (6-65)	127 (29-512)	< 0.001	44 (10-185)	29 (13-238)	0.861
Total TLG	67 (18-448)	1180 (200-2998)	< 0.001	281 (43-1541)	199 (64-1568)	0.984

TABLE 3	¹⁸ F-FDG PE	T lesion	parameters	on a	a per-patient	basis,	stratified	by L	DH or	BRAFV600) mutatio	n
status.												

Data are displayed as median (interquartile range). *One patient had a missing LDH value. b Three patients with normal LDH had only lesions <1 ml. *All lesions included in subsequent quantitative analyses ⁴Interquartile range of the different SUVpeaks measured within one patient, measure of intra-patient heterogeneity. LDH = lactate dehydrogenase

Overall survival

Following the baseline PET/CT scan, patients commenced standard systemic treatment consisting of either immune checkpoint inhibition, BRAF(/MEK) inhibition and/or dacarbazine chemotherapy. Given the various systemic treatments used, overall survival analysis was performed for exploratory purposes only (Kaplan-Meier overall survival curves in SUPPL. FIGURE 4).

DISCUSSION

We show major intra- and inter-patient heterogeneity in tumour lesion ¹⁸F-FDG uptake among metastatic melanoma patients. Presence of tumours with above median ¹⁸F-FDG uptake was independent of tumour mutational status and did not fully coincide with high serum LDH level. This suggests that tumour ¹⁸F-FDG uptake is an independent feature and that ¹⁸F-FDG PET parameters might be suitable as a selection tool for novel metabolic therapies.

This is the first large study providing an overview of intra- and inter-patient differences in tumour glucose consumption in metastatic melanoma patients using quantitative who-



Mapping heterogeneity in ¹⁸F-FDG uptake in melanoma



le-body imaging of ¹⁸F-FDG uptake. Previous melanoma studies on ¹⁸F-FDG PET/CT imaging focused on its diagnostic accuracy for qualitative lesion detection and/or used quantitative parameters derived from the primary melanoma or only a limited number of (the most intense) lesions for response evaluation or prognostic models. By performing quantitative evaluation of all tumour lesions, we highlight the utility of ¹⁸F-FDG PET/CT in demonstrating heterogeneity of glucose uptake among metastatic melanoma patients. Preliminary estimates of the influence of tumour ¹⁸F-FDG uptake on survival support further prospective investigation as a prognostic biomarker.

Compared to previous studies, we found a higher proportion of bone metastases and a lower incidence of lung and soft tissue metastases. Two previous studies in metastatic melanoma using ¹⁸F-FDG PET/CT \pm other imaging methods qualitatively report metastases predominantly to the lung, liver, lymph nodes and skin/soft tissue^{19,20}. This difference might be explained by differing patient populations, especially since our study also included patients with an unknown primary melanoma and subsequent widespread (skeletal) metastases (n = 8), as opposed to the study performed by Schoenewolf et al.¹⁹. Furthermore, we excluded lesions with an MATV < 1 ml to minimize partial volume effects. Three patients had only lesions with an MATV < 1 ml, which all concerned metastases in the lymph nodes, lung, subcutis and/or muscles. The small MATV at these locations, resulting in the exclusion of these lesions for the analysis, further explains the smaller fraction of soft tissue, lymph node and subcutaneous lesions in our PET-based study.

BRAFV600 mutant melanoma cells rely heavily on glycolysis with high glycolytic rates induced by activation of the mitogen-activated protein kinase (MAPK) pathway^{21,22}. *BRAFV600* wild-type melanomas (approximately 50% of melanomas) often have alternative mutations in the MAPK-pathway including *RAS* or *MEK1/2* that are also associated with glycolytic dependency and increased glucose uptake²³⁻²⁵. In thyroid carcinoma, *BRAFV600E* tumours show increased expression of glucose transporter (GLUT) and higher SUVs compared to *BRAFV600* wild-type tumours^{26,27}. We found no difference in tumour glucose uptake and MATV between patients with and without a *BRAFV600* or *RAS* mutation. Overexpression of other proteins stimulating glucose consumption in the *BRAF/RAS* wild-type population may explain this observation. Potentially relevant proteins include MEK1/2 (8% of melanomas), involved in the MAPK-pathway²⁵, and mTOR (10.4% of primary melanomas) or PDK1, involved in the PI3K-Akt-mTOR pathway^{28,29}. Furthermore, patient's *BRAF* status is determined based on one tumour tissue sample, not uncommonly the (excised) primary melanoma, and consequently does not necessarily represent the mutational status of all metastases within a patient³⁰.

Patients with an elevated serum LDH level — a well-established prognostic biomarker for both worse survival and poor treatment response — had higher tumour ¹⁸F-FDG uptake as well as higher metabolic tumour volume compared to those with normal LDH levels. However, we also observed tumours with high ¹⁸F-FDG in patients with (still) relatively low MATV and normal LDH levels. Moreover, several patients with an elevated LDH level had only tumour lesions with relatively low ¹⁸F-FDG uptake. LDH is a cytoplasmic enzyme that catalyses the interconversion of pyruvate and lactate downstream of glycolysis. A high LDH serum level is generally regarded as a marker of cell damage or necrosis, but the exact source of serum LDH levels is unknown. The biological role of LDH in glucose metabolism

has also been suggested as an underlying mechanism and in vitro data suggest differential reliance on aerobic glycolysis and oxidative phosphorylation between patients with normal and elevated serum LDH^{3,13}. Unfortunately, meaningful multivariate approaches to unravel the interrelations between tumour volume, tumour glucose consumption and a proposed metabolic factor underlying serum LDH levels were prohibited by collinearity in our data.

Metabolic targeting may constitute a promising novel approach for patients with tumours with high glucose uptake identified by ¹⁸F-FDG PET. Furthermore, glycolysis results in extracellular accumulation of lactate and low pH, which are known to impair immune cell function and contribute to an immunosuppressive tumour microenvironment^{3, 5}. Metabolic interference combined with immunotherapy might thus be attractive for improving immunotherapy response, for instance in the poorly responding group of metastatic melanoma patients with elevated LDH levels. Metabolic cancer therapies have numerous specific metabolic targets and so far, studies into the correlation between melanoma expression of specific glycolytic transporters and enzymes, such as GLUT1 and hexokinase (HK), and ¹⁸F-FDG uptake are limited and contradictive^{31,32}. New studies are needed to integrate tumour ¹⁸F-FDG uptake and other clinical biomarkers with tumour dependence upon specific metabolic pathways and targetable metabolic transporters and enzymes.

Limitations of our study include its retrospective nature and patient heterogeneity in treatment, which allowed only preliminary estimates of the influence of tumour PET parameters on survival. The lack of ce-CT in several patients and its more detailed anatomical lesion information could have resulted in erroneous inclusion of physiological PET-positive lesions or exclusion of malignant PET-positive lesions, respectively. Since tumour measurements were performed on PET images only, necrotic areas (observed in three patients) and brain metastases (n = 22) are not incorporated in the MATV.

CONCLUSIONS

Tumour ¹⁸F-FDG uptake is heterogeneous within and among metastatic melanoma patients. High ¹⁸F-FDG uptake is independent of BRAF/RAS mutation status and does not fully correlate with serum LDH levels. This suggests ¹⁸F-FDG PET metabolic parameters could serve as an (additional) selection tool for melanoma patients potentially benefiting from metabolic therapies. Further investigation of the prognostic and predictive value of quantitative ¹⁸F-FDG PET parameters is warranted.

ACKNOWLEDGMENTS

The authors acknowledge JH van Snick and NA 't Hart for their technical assistance.

REFERENCES

1. Ugurel S, Röhmel J, Ascierto PA, Flaherty KT, Grob JJ, Hauschild A, et al. Survival of patients with advanced metastatic melanoma: the impact of novel therapies. Eur J Cancer. 2016;53:125-134.

2. Brand A, Singer K, Koehl GE, Kolitzus M, Schoenhammer G, Thiel A, et al. LDHA-associated lactic acid production blunts tumor immunosurveillance by T and NK cells. Cell Metab. 2016;24:657-671.

3. Blank CU, Haanen JB, Ribas A, Schumacher TN. The "cancer immunogram" Science. 2016;352:658–660.

 Vander Heiden MG, Cantley LC, Thompson CB. Understanding the Warburg effect: the metabolic requirements of cell proliferation. Science. 2009;324:1029–1033.

5. Pavlova NN, Thompson CB. The emerging hallmarks of cancer metabolism. Cell Metab. 2016;23:27-47.

 Michelakis ED, Sutendra G, Dromparis P, Webster L, Haromy A, Niven E, et al. Metabolic modulation of glioblastoma with dichloroacetate. Sci Transl Med. 2010;2:31ra34.

7. Martinez-Outschoorn UE, Peiris-Pagés M, Pestell RG, Sotgia F, Lisanti MP. Cancer metabolism: a therapeutic perspective. Nat Rev Clin Oncol. 2017;14:11–31.

8. Vartanian A, Agnihotri S, Wilson MR, Burrell KE, Tonge PD, Alamsahebpour A, et al. Targeting hexokinase 2 enhances response to radio-chemotherapy in glioblastoma. Oncotarget. 2016;7:69518-69535.

9. Bénéteau M, Zunino B, Jacquin MA, Meynet O, Chiche J, Pradelli LA, et al. Combination of glycolysis inhibition with chemotherapy results in an antitumor immune response. Proc Natl Acad Sci U S A. 2012;109:20071–20076.

10. Ganapathy-Kanniappan S, Geschwind JF. Tumor glycolysis as a target for cancer therapy: progress and prospects. Mol Cancer. 2013;12:152.

11. Garon EB, Christofk HR, Hosmer W, Britten CD, Bahng A, Crabtree MJ, et al. Dichloroacetate should be considered with platinum-based chemotherapy in hypoxic tumors rather than as a single agent in advanced non-small cell lung cancer. J Cancer Res Clin Oncol. 2014;140:443–452.

12. Shestov AA, Mancuso A, Lee SC, Guo L, Nelson DS, Roman JC, et al. Bonded cumomer analysis of human melanoma metabolism monitored by 13C NMR spectroscopy of perfused tumor cells. J Biol Chem. 2016;291:5157-5171.

13. Ho J, de Moura MB, Lin Y, Vincent G, Thorne S, Duncan LM, et al. Importance of glycolysis and oxidative phosphorylation in advanced melanoma. Mol Cancer. 2012;11:76.

14. Balch CM, Gershenwald JE, Soong SJ, Thompson JF, Atkins MB, Byrd DR, et al. Final version of 2009 AJCC melanoma staging and classification. J Clin Oncol. 2009;27:6199–6206.

15. Boellaard R, Delgado-Bolton R, Oyen WJ, Giammarile F, Tatsch K, Eschner W, et al. FDG PET/CT: EANM procedure guidelines for tumour imaging: version 2.0. Eur J Nucl Med Mol Imaging. 2015;42:328–354.

16. Strobel K, Dummer R, Husarik DB, Pérez Lago M, Hany TF, Steinert HC. High-risk melanoma: accuracy of FDG PET/CT with added CT morphologic information for detection of metastases. Radiology. 2007;244:566–574.

17. Lin NU, Lee EQ, Aoyama H, Barani IJ, Barboriak DP, Baumert BG, et al. Response assessment criteria for brain metastases: proposal from the RANO group. Lancet Oncol. 2015;16:270-278.

18. Ilie M, Khambata-Ford S, Copie-Bergman C, Huang L, Juco J,

Hofman V, et al. Use of the 22C3 anti-PD-L1 antibody to determine PD-L1 expression in multiple automated immunohistochemistry platforms. PLoS One. 2017;12:e0183023.

19. Schoenewolf NL, Belloni B, Simcock M, Tonolla S, Vogt P, Scherrer E, et al. Clinical implications of distinct metastasizing preferences of different melanoma subtypes. Eur J Dermatology. 2014;24:236-241.

20. Frauchiger AL, Mangana J, Rechsteiner M, Moch H, Seifert B, Braun RP, et al. Prognostic relevance of lactate dehydrogenase and serum S100 levels in stage IV melanoma with known BRAF mutation status. Br | Dermatol. 2016;174:823-830.

21. Hall A, Meyle KD, Lange MK, Klima M, Sanderhoff M, Dahl C, et al. Dysfunctional oxidative phosphorylation makes malignant melanoma cells addicted to glycolysis driven by the (V600E)BRAF oncogene. Oncotarget. 2013;4:584-599.

22. Hardeman KN, Peng C, Paudel BB, Meyer CT, Luong T, Tyson DR, et al. Dependence on glycolysis sensitizes BRAFmutated melanomas for increased response to targeted BRAF inhibition. Sci Rep. 2017;7:42604.

23. Nazarian R, Shi H, Wang Q, Kong X, Koya RC, Lee H, et al. Melanomas acquire resistance to B-RAF(V600E) inhibition by RTK or N-RAS upregulation. Nature. 2010;468:973-977.

24. Kerr EM, Gaude E, Turrell FK, Frezza C, Martins CP. Mutant Kras copy number defines metabolic reprogramming and therapeutic susceptibilities. Nature. 2016;531:110-113.

25. Richtig G, Hoeller C, Kashofer K, Aigelsreiter A, Heinemann A, Kwong LN, et al. Beyond the BRAFV600E hotspot: biology and clinical implications of rare BRAF gene mutations in melanoma patients. Br | Dermatol. 2017;177:936-944.

26. Choi EK, Chong A, Ha JM, Jung CK, O JH, Kim SH. Clinicopathological characteristics including BRAF V600E mutation status and PET/CT findings in papillary thyroid carcinoma. Clin Endocrinol. 2017;87:73–79.

27. Yoon M, Jung SJ, Kim TH, Ha TK, Urm SH, Park JS, et al. Relationships between transporter expression and the status of BRAF V600E mutation and F-18 FDG uptake in papillary thyroid carcinomas. Endocr Res. 2016;41:64–69.

28. Yan K, Si L, Li Y, Wu X, Xu X, Dai J, et al. Analysis of mTOR gene aberrations in melanoma patients and evaluation of their sensitivity to PI3K-AKT-mTOR pathway inhibitors. Clin Cancer Res. 2016;22:1018–1027.

29. Pópulo H, Caldas R, Lopes JM, Pardal J, Máximo V, Soares P. Overexpression of pyruvate dehydrogenase kinase supports dichloroacetate as a candidate for cutaneous melanoma therapy. Expert Opin Ther Targets. 2015;19:733-745.

 Riveiro-Falkenbach E, Santos-Briz A, Ríos-Martín JJ, Rodríguez-Peralto JL. Controversies in intrapatient melanoma BRAFV600E mutation status. Am J Dermatopathol. 2017;39:291-295.

31. Park SG, Lee JH, Lee WA, Han KM. Biologic correlation between glucose transporters, hexokinase-II, Ki-67 and FDG uptake in malignant melanoma. Nucl Med Biol. 2012;39:1167-1172.

32. Yamada K, Brink I, Bissé E, Epting T, Engelhardt R. Factors influencing [F-18] 2-fluoro-2-deoxy-D-glucose (F-18 FDG) uptake in melanoma cells: the role of proliferation rate, viability, glucose transporter expression and hexokinase activity. J Dermatol. 2005;32:316-334.

SUPPLEMENTARY MATERIAL

	All lesions (n = 1143)	Range			
SUVpeak	5.0 (3.4-7.8)	0.7-58.3			
SUVmax	6.8 (4.6-10.7)	1.1-67.2			
SUVmean	4.6 (3.4-6.8)	0.7-30.2			
MATV (ml)	2.4 (1.4-6.1)	1.0-1921			
TLG	11.4 (5.8-34.0)	1.1-11206			
Data are displayed as median (interguartile range)					

SUPPL. TABLE 1 | ¹⁸F-FDG PET tumour lesion parameters on a per-lesion basis.





SUPPL. FIGURE 1 | Patient selection. Flow diagram showing the selection of eligible patients.

*One patient with active pulmonal lymphangioleiomyomatosis was excluded based on contradictive literature on differentiating lymphangioleiomyomatosis from tumor lesions on ¹⁸F-FDG PET/CT. † One patient with PET-negative diffuse bone marrow metastases (biopsy-confirmed) and two patients with only brain metastases. AJCC = American Joint Committee on Cancer



SUPPL. FIGURE 2 | Number of tumour lesions per metastatic location (A) and total MATV (B) per location. In total, 1143 tumour lesions \geq 1 ml were identified in 64 patients. The outer ring in (A) displays the distribution when lesions <1 ml are incorporated as well (total lesion n = 3408), showing only minor differences. Total MATV of all 1143 lesions was 14,560 ml (B). LN = lymph node.







SUPPL. FIGURE 4 | Kaplan-Meier overall survival estimates stratified by LDH levels and PET parameters.

Following baseline ¹⁸F-FDG PET/CT scan, 30 patients (46.9%) started with immunotherapy, 20 patients (31.3%) started with BRAF(/ MEK) inhibition, and 5 patients (7.8%) commenced dacarbazine chemotherapy. Nine patients (14.1%) did not receive any systemic treatment. Twenty-one of the included 64 patients (32.8%) were still alive at the time of analysis (17.9 months after the last included baseline PET/CT scan). Curves display overall survival of all patients (n = 64) stratified by normal vs. elevated (i.e. > 250 U/I) LDH levels and patient population median of respectively maximum SUVpeak (**A**), total MATV (**B**) and total TLG (**C**). LDH = lactate dehydrogenase.