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Prediction of the Free Energy of Binding for Cyclodextrin-Steroid Complexes: Phase Solubility and Molecular Dynamics Studies

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Abstract

Steroid hormones play a crucial role in the body by acting as chemical messengers. They are, however, poorly soluble in water, and cyclodextrins can increase their solubility thus leading to increased bioavailability when used in drug formulations. Accuracy in the prediction of the free energy of binding of cyclodextrin/steroid inclusion complexes with simulation is important because of the potential value it brings by providing low-cost predictions of the real-life behavior of the cyclodextrin/steroid inclusion complex and the potential for high-through-put screening. Many computational methods exist, and it is therefore important to understand the ability of current theoretical models to accurately predict the free energy of binding for these inclusion complexes.

We focused specifically on the estimation of the free energy of binding of inclusion complexes of four steroids: Hydrocortisone, dexamethasone, prednisolone, and 6 α -methylprednisolone with native α -CD, β -CD, γ -CD, hydroxypropyl- β -CD, and sulfobutylether- β -CD by phase solubility as well as with α , β , and γ -CD by simulations. The simulations were assessed with both docking and the molecular mechanics combined with the generalized Born and surface area (MM/GBSA) continuum solvation approach. Considering the phase solubility diagram, hydroxypropyl- β -CD and sulfobutylether- β -CD dissolved more steroids in the higher concentration range as expected. The assessment of the free energy of binding obtained from the phase solubility and theory showed that the MM/GBSA method has shown promise in reliably generating accurate predictions in the field of calculating the free energy of binding of steroids/cyclodextrins with a correlation coefficient (R^2) = 0.9.

Keywords: Cyclodextrin, Phase solubility, Steroid, The free energy of binding, Molecular Docking Simulation, Inclusion complex

Declarations

Conflict of interest The authors declare that they have no conflict of interest.

1. Introduction

Computational modeling has emerged as a powerful and cost-effective strategy for predicting properties of guest-host inclusion complexes [1] such as structures and the free energy of binding. Although computational techniques have the potential to reduce laboratory costs by predicting experimental outcomes, careful and critical evaluation of such models must be carried out. Accordingly, finding better and more reliable experimental sources of data as well as setting more realistic expectations for computational modeling should increase confidence in such predictions and ultimately accelerate research.

One important guest-host system is the steroid hormone-cyclodextrin system, which have found important applications due to the ability of cyclodextrin (CD) to change the apparent behavior of the included guest molecule. Steroid hormones are a group of organic compounds that act as chemical messengers and play an essential role in the body [2], [3]. Corticosteroids are a group of steroid hormones containing four cycloalkane rings that are linked together [4]. Hydrocortisone, dexamethasone, prednisolone, and 6 α -methylprednisolone belongs to the glucocorticoid group of

corticosteroids (Fig. 1). They are very important for protecting against stress, shock, and inflammation and these recent years they have also been used for treatment of COVID-19 patients [5], [6].

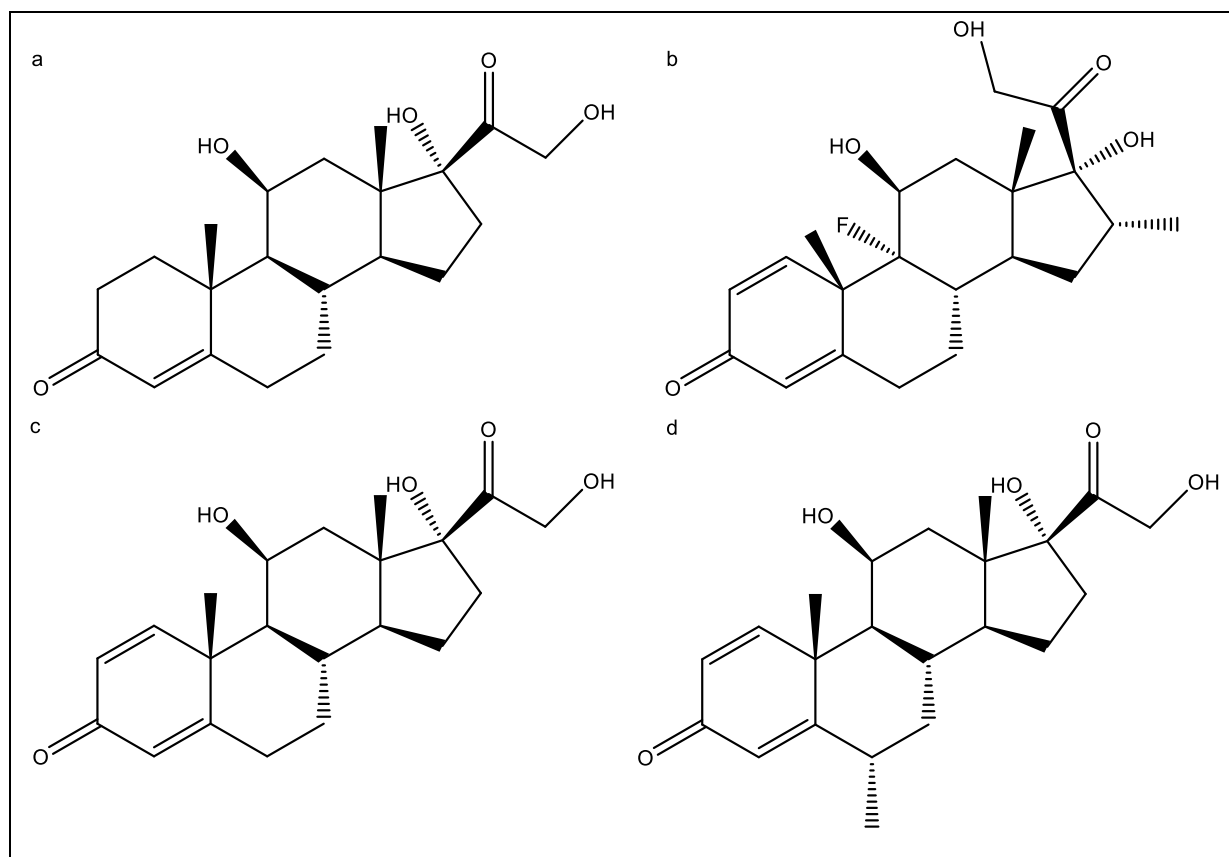


Fig. 1 The chemical structures of a) Hydrocortisone b) Dexamethasone c) Prednisolone d) 6α-methylprednisolone.

Steroids are poorly soluble in water which limit their use as efficient drugs. CDs are relatively small cyclic molecules, which can participate in the host (H)-guest (G) interactions to form 1:1 complex (HG) in water (Eq. 1).



CDs are typically used to bring more drug, including steroid hormones, into solutions or increase the dissolution rate of solid formulations leading to increased bioavailability [7]. They are obtained from starch and are considered for a large number of administration routes [8]. CDs have been shown to have good chemical stability and are in general non-toxic to humans in food and medicines [9], [10]. α -, β - and γ -CD consist of 6, 7, or 8 glucopyranose units, respectively, connected through α -1,4-D glycosidic linkages [11] as shown in Fig. 2. The truncated cone-like structure of CDs presents

a hydrophilic exterior formed by hydroxyl groups while the interior is relatively hydrophobic due to the structure of the pyranose rings. This allows the CDs to act as host molecules for smaller guests or parts of larger molecules.

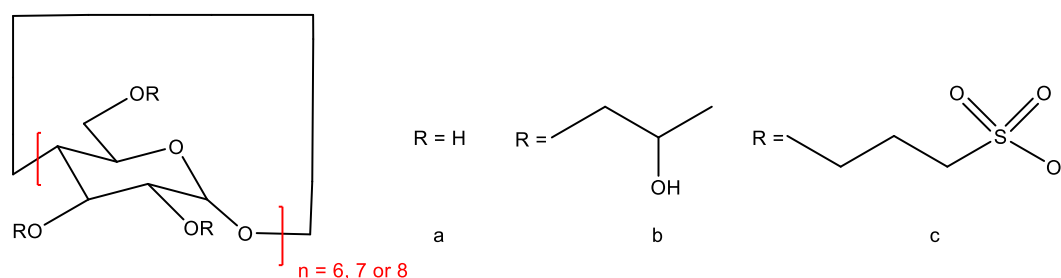


Fig. 2 Structure of α -, β -, and γ -CD consisting of $n=6, 7$ and 8 α -1,4-D-glucopyranose units in the macrocycle, respectively. R refers to the structures of native α -CD, β -CD, and γ -CD (a) or HP-CD (a and b) or SBE-CD (a and c), respectively

CDs can be chemically modified by substitution of some (or all) of the hydroxyl groups with various side chains as shown in Fig. 2. Known derivatives that are already used for drug delivery in approved and marketed products are hydroxypropyl CDs (HP-CD) and sulfobutylether CDs (SBE-CD) where the β -CD versions of the two derivatives (HP- β -CD and SBE- β -CD) are leading in the market [7].

Studies of the equilibrium (Eq. 1) are important for understanding the effects CDs exert on different drugs. Various experimental methods can be used for this purpose where the most common include phase solubility [12]–[15] or titration based methods relying on UV-Vis absorption, fluorescence [16], or NMR [14], [16]. In general, a straightforward model assuming a simple stoichiometry (e.g. 1:1) is fitted to the obtained data to predict key parameters of the equilibrium, including the stability constant (K) and Gibb's free energy (ΔG). In some studies, theoretical calculations from molecular dynamic (MD) or molecular mechanics (MM) are included to back up the experimental data, i.e., investigate structure and dynamics of the inclusion complex and validate the free energies of binding [17]–[19].

The explosive increase of data-based techniques raises an important question: To what extent are current theoretical models able to reproduce experimental the free energies of binding? We will in this study address this question by

computing the free energy of binding by a range of computational modeling methods and comparing them to values determined experimentally (by phase solubility method) using steroid-CD equilibria as models.

2. Methods

2.1. Materials

α -, β - and γ -CD (pharmaceutical grade) were a gift from Wacker Chemie AG (Burghausen, Germany). Hydrocortisone 98.0%, dexamethasone 98.0%, prednisolone 99% and 6α -methylprednisolone 98.0% were purchased from TCI, Combi-Blocks, Sigma-Aldrich and TCI, respectively. Hydroxypropyl β -CD (KLEPTOSE[®] HPB Oral grade) DS 4.38 (average MW 1389.37 Da) was a gift from Roquette (Lestrem, France). SBE- β -CD (Captisol) DS 6.3 (average MW 2131.46 Da) was kindly donated by Ligand Pharmaceutical (San Diego, United States), Milli-Q water (Millipore, Billerica, MA, USA) was used to prepare CD solutions and high-performance liquid chromatography (HPLC) mobile phases. Acetonitrile used to prepare the HPLC mobile phases was purchased from Sigma-Aldrich.

2.2. Phase-Solubility Studies

Phase-solubility studies were performed in accordance with the Higuchi and Connors method [20]. All CDs were dried under vacuum at 110°C prior to use. An excessive amount of hydrocortisone, dexamethasone, prednisolone, and 6α -methylprednisolone (approximately 5-10 times the amount of CDs (in units of mol)) were added to aqueous solutions containing various concentrations of CDs (0-0.009 M for β -CD, 0-0.1 M for α -, γ -, HP- β -CD and SBE- β -CD). The suspensions were stirred for 24 and 48 hours to reach equilibrium at 23°C. The mixture was centrifuged (Sigma model (1-15PK) with 12124 rotor, Taufkirchen, Germany) at 14.000 rpm for 20 minutes at room temperature to separate the soluble and insoluble fractions. The resulting supernatant was collected with syringe and needle and was filtered with a 0.45 μ m Phenex[™]-RC membrane 15mm Syringe Filters (Phenomenex, Aschaffenburg, Germany) before HPLC analysis. 1000 μ L of the filtered solution was diluted with 250 μ L of the same starting solution without steroid to prevent precipitation.

The amount of steroids in solution was quantified using HPLC. A Dionex (Germering, Germany) setup equipped with P 680A LPG-4 HPLC Pump and ASI-100 Automated Sample Injector with Dionex UVD 170U UV/VIS Detector and Kinetex C18 column (150 \times 4.6 mm, 5 μ m; Alltech, Værløse, Denmark). The isocratic mobile phase was acetonitrile/water (10/90, 20/80, 10/90 and 10/90 for hydrocortisone, dexamethasone, prednisolone, 6α -methylprednisolone, respectively)

with a flow rate of 1.0 ml/min. The eluent was monitored at 210 nm for hydrocortisone and 245 nm for the other steroids. The data were recorded and treated using Chromeleon™ 6.8 SR15 program from Thermo Scientific (Göteborg, Sweden). As you can see in Fig. 3, Linear external calibration curves in the concentration range from 0.01 to 0.20 mg/mL for hydrocortisone, 0.01 to 0.10 mg/mL for dexamethasone and 0.01 to 0.07 mg/mL for both prednisolone and 6 α -methylprednisolone were used.

The apparent stability constant ($K_{1:1}$) was determined from the phase-solubility diagrams using the linear model described in Eq. 2: [20], [21]

$$K_{1:1} = \frac{\text{slope}}{S_0(1 - \text{slope})} \quad (2)$$

Here, the slope is obtained from the linear relationship between the concentration of steroids and the concentration of CDs, and the intrinsic solubility S_0 of the steroid was assessed in pure water in the absence of CD [22]. All phase-solubility results are reported as means with a standard error based on four experiments ($n = 4$).

The change in Gibbs free energy ($\Delta G_{bind,exp}$) of the steroid/CD equilibria obtained from the phase solubility experiments was derived from Eq. 3:

$$\Delta G_{bind,exp} = -RT \ln K_{1:1} \quad (3)$$

Here, R is the ideal gas constant (8.314 J K⁻¹ mol⁻¹) and T is the temperature (296 K).

2.3. Molecular Simulations

2.3.1. Preparation of Molecular Structures

The starting structures of native α -, β -, and γ -CD were generated with the tleap program in the AMBER 16 software [23]. Structures of hydrocortisone, dexamethasone, prednisolone and 6 α -methylprednisolone (Fig. 1) were prepared with the 2D sketcher in Maestro [24] in Schrödinger 2021-3 software [24] and minimized without solvent using the OPLS4 force field [25].

2.3.2. Molecular Docking

Molecular docking simulations were performed with Glide [26]–[28] to obtain ΔG_{bind} for the inclusion complexes of hydrocortisone, dexamethasone, prednisolone, and 6 α -methylprednisolone with α -, β - and γ -CD, respectively. Stable conformations of each steroid were found using a conformational search with enhanced torsional sampling enabled [29]. The docking simulations were performed by choosing the centroid of the CDs as the center for the docking grid. Both

standard precision (SP) and extra precision (XP) [28] docking simulations were carried out. Mean and standard error of the simulated Gibbs free energies were obtained from 5 independent docking simulations assuming 1:1 interaction to assess the free energy of binding for each steroid/CDs.

2.3.3. Molecular Mechanics with Generalized Born and Surface Area Method

We also employed the molecular mechanics energies in combination with the Generalized Born (GB) and Surface Area (SA) method (MM/GBSA) [30], [31] on inclusion complexes to obtain the free energies of binding of the four ligands and the three native CDs. This was done with the AMBER 16 software using the one-trajectory approach where a single molecular dynamics (MD) simulation is carried out on the inclusion complex (HG , eq 1) and used to compute ΔG : [30]

$$\Delta G_{bind} = \langle G_{HG} - G_H - G_G \rangle_{HG}. \quad (4)$$

Here, the free energies (G_{HG} , G_H and G_G) are all computed according to

$$G_X = E_{bnd} + E_{el} + E_{vdw} + G_{pol} + G_{np} - TS. \quad (5)$$

The free energy of X in Eq. 5 is calculated from bonded (E_{bnd}), electrostatic (E_{el}) and van der Waals (E_{vdw}) terms of the respective part of the system. The polar solvation energy, G_{pol} , is obtained solving the GB model (igb=5) [32] and non-polar contribution to the free energy of binding, G_{np} , was estimated from the solvent accessible surface area [33]. The entropy, S , was computed using a normal mode analysis [34].

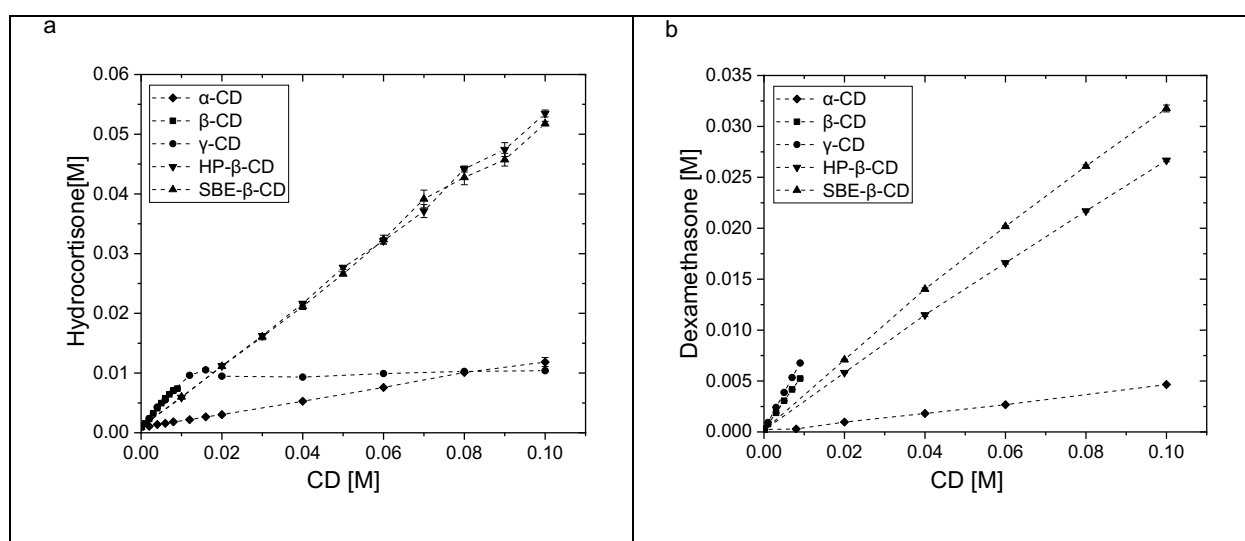
For each steroid, we performed two simulations with the polar groups of each steroid pointing either down through the primary side (orientation-b) or up through the secondary side (orientation-a). The ligands were parameterized with the GAFF forcefield using the tleap program in AMBER. All CDs were parameterized with the GLYCAM06 carbohydrate force field [35]. The final inclusion complex was solvated in a cubic box of TIP3P [36] water with at least 14 Å of boundary between the CD and the periodic walls. To equilibrate the systems in AMBER, we initially minimized each structure with steepest descent for a maximum of 1000 steps or until the root mean square deviation on the gradient was below 10^{-3} kcal mol⁻¹ Å⁻¹. Due to observed instabilities in preliminary tests of the molecular dynamic simulations, we chose a conservative equilibration protocol. Unless otherwise noted, we harmonically restrained the atoms of both steroid and CD with a force constant of 100 kcal mol⁻¹ Å⁻² to their initial positions. We subsequently performed 100 ps simulation in the NVT at 10 K ensemble with a step size of 1 fs followed by 2 ns of NPT simulation where the temperature was increased linearly to 300 K while keeping the pressure constant at 1 bar and a step size of 2 fs. In the final part of the equilibration, we performed 10 ns of equilibration in the NPT ensemble where the force constant for the restraints were

lowered to $10 \text{ kcal mol}^{-1} \text{ \AA}^{-2}$. We finally performed the production with no restraints for a total of 5 ns simulation in the NPT ensemble. A total of 100 frames were saved for postprocessing with MM/GBSA.

3. Results and Discussion

3.1. Phase-Solubility Studies

Both A_L and B_S -type phase-solubility diagrams display a linear initial slope (Figs. 3 and 4) and we use this to estimate $K_{1:1}$ by fitting the data to Eq. 2. Comparison of samples harvested at 24 h and 48 h showed no significant change in amount of drug in solution, signifying that the equilibrium was established already after 24 h of incubation. In Fig. 3, we present our phase-solubility results of the four steroids in complex with various hosts in concentrations up to 0.10 M. We initially used hydrocortisone to explore the experimental behavior. For example, a B_S -type plot is observed for γ -CD/hydrocortisone, as shown in Fig. 3a. γ -CD concentration up towards 0.012 M shown an initial increasing linear trend whereafter we observe a plateau in the concentration of hydrocortisone brought into solution. Therefore, $K_{1:1}$ for γ -CD/hydrocortisone was calculated from the data in the concentration interval from 0 to 0.012 M. Since the initial linear slope on the B_S diagrams and A_L -type diagrams all had slopes less than unity, formation of 1:1 complex within the range of concentrations studied is likely [37], [38] and consistent with previous results [14], [39]. Based on these preliminary results for hydrocortisone the concentration ranges for all subsequent experiments were changed to 0–0.009 M for β - and γ -CD and 0–0.100 M for α -, HP- β -CD, and SBE- β -CD. The number of data points used in the experiments were decreased from 11 to 6 for the remaining phase solubility studies.



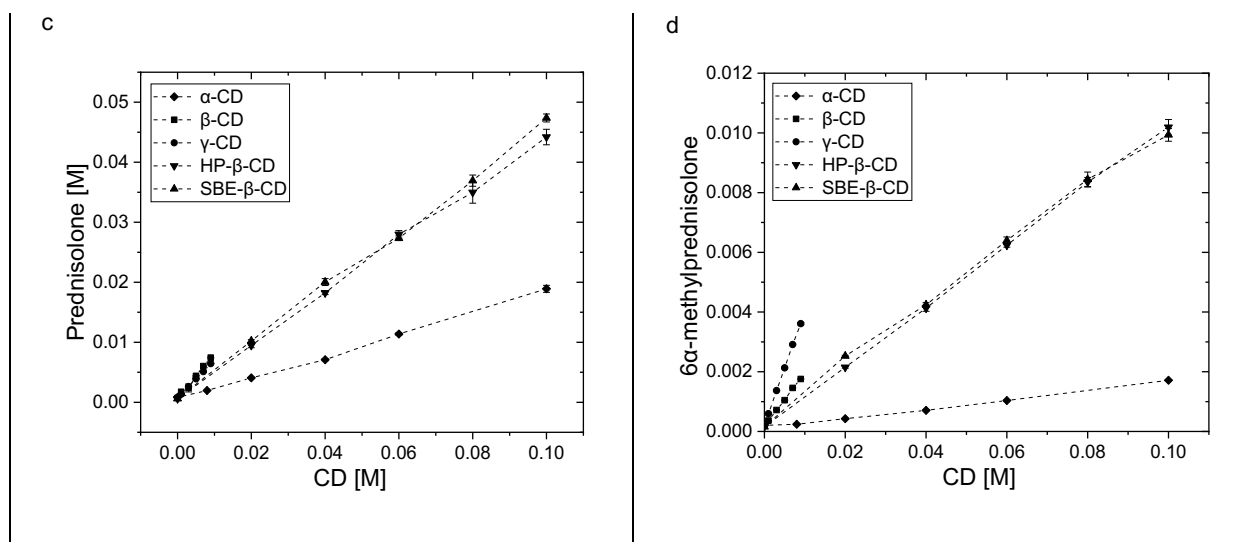


Fig. 3 Phase-solubility diagrams of a) hydrocortisone, b) dexamethasone, c) prednisolone, and d) 6 α -methylprednisolone in solutions of α -, β -, γ -, HP- β -CD or SBE- β -CD at 23 $^{\circ}$ C. Data are expressed as the mean \pm S.D (n=4)

Assuming a 1:1 complex stoichiometry, the stability constants for the inclusion complex of hydrocortisone, dexamethasone, prednisolone, and 6 α -methylprednisolone/CDs were calculated (Table 1). The stability constants of hydrocortisone with all five CDs varied from $125 \pm 6 \text{ M}^{-1}$ to $3509 \pm 206 \text{ M}^{-1}$. The complex between hydrocortisone and β -CD exhibited the strongest stability constants compared to the other CDs (Table 1). The solubility of β - and γ -CD are relatively limited and typically makes poorly soluble complexes resulting in B_s type diagrams [40]. The HP- β -CD and SBE- β -CD phase solubility diagrams with the various steroids were linear in a larger concentration range as expected for heterogeneous CD derivatives (Fig. 3). The amorphous nature of HP- β -CD and SBE- β -CD leads to higher solubility of both the CD derivative itself and its inclusion complexes [41]. As shown in Table 1, the stability constants found between the steroids and β -CD is relatively higher than those found for HP- β -CD and SBE- β -CD. This is probably due to the presence of side chains in HP- β -CD and SBE- β -CD causing a narrowing of the β -CD cavity opening and steric hindrance. SBE- β -CD have a slightly higher stability constant compared to HP- β -CD, possibly due to the longer side chain or the polar head group or both [39], [42], [43].

Table 1. Apparent stability constants ($K_{1:1}$), ΔG and characteristics of the phase solubility diagrams obtained for hydrocortisone, dexamethasone, prednisolone, and 6 α -methylprednisolone in inclusion complexes with α -, β -, γ -, HP- β -CD or SBE- β -CD, respectively, at 23 $^{\circ}$ C.

Cyclodextrin		Hydrocortisone				
				Phase		
	Slope	R ²	K _{1:1} (M ⁻¹)	solubility ΔG [kJ mol ⁻¹]	Literature K _{1:1} (M ⁻¹) ^a	Literature ΔG [kJ mol ⁻¹] ^a
α-CD	0.1	0.999	125 ± 6	-11.88 ± 0.10	72 [39]	-10.52
					57 [44]	-9.95
β-CD	0.7	0.999	3509 ± 206	-20.09 ± 0.15	1360 [39]	-17.76
					3803 ^g [37]	-20.29
					2683 [45]	-19.43
γ-CD	0.7	0.996	2389 ± 79 ^b	-19.14 ± 0.09	1720 [44]	-18.34
					1970 [39]	-18.68
					2240 [44]	-18.99
HP-β-CD	0.5	0.999	1111 ± 30	-17.26 ± 0.067	3290 [46]	-19.94
					1450 [39]	-17.92
					1000 [40]	-17.01
					1010 [47]	-17.03
SBE-β-CD	0.5	0.997	1155 ± 34	-17.35 ± 0.07	1700 [22]	-18.31
					2056 ^h [48]	-18.78
					1580 [39]	-18.13
					2516 ^{h, c} [48]	-19.28
Dexamethasone						
α-CD	0.04	0.995	234 ± 2	-13.42 ± 0.37	240 [12]	-13.49
β-CD	0.5	0.999	5570 ± 152	-21.23 ± 0.18	5340 [12]	-21.13
					730 ± 150 ^d [49]	-16.23
γ-CD	0.7	1	13618 ± 287	-23.43 ± 0.05	12887 ^f [50]	-23.30
					1210 [12]	-17.48
					980 [51]	-16.96
					920 ^e [51]	-16.80

					2311 [38]	-19.07
					2070 [12]	-18.80
HP- β -CD	0.2	0.999	2182 \pm 46	-18.92 \pm 0.05	2193 ^j [52]	-18.94
					800 [22]	-16.46
					2221 ^{j, h} [52]	-18.97
SBE- β -CD	0.3	0.998	2879 \pm 40	-19.60 \pm 0.034	2880 [12]	-19.61
Prednisolone						
α -CD	0.2	0.997	278 \pm 9	-13.85 \pm 0.47	288 \pm 7 [14]	-13.94
					298 [44]	-14.02
β -CD	0.7	0.997	3242 \pm 32	-19.89 \pm 0.02	3479 \pm 139 [14]	-20.08
					3600 [44]	-20.16
γ -CD	0.6	0.999	2092 \pm 36	-18.82 \pm 0.04	1783 \pm 48 [14]	-18.43
					3240 [44]	-19.90
HP- β -CD	0.4	0.999	1286 \pm 5	-17.62 \pm 0.01	2428 \pm 56 [14]	-19.19
					960 [53]	-16.91
					1319 ^h [48]	-17.69
SBE- β -CD	0.5	0.998	1382 \pm 64	-17.80 \pm 0.10	2680 \pm 62 ^c [14]	-19.43
					1513 ^c [53]	-18.02
					1821 ^{h, c} [48]	-18.48
6α-methylprednisolone						
α -CD	0.02	0.995	77 \pm 1	-10.69 \pm 0.30	98 \pm 3 [14]	-11.29
β -CD	0.17	0.999	1049 \pm 14	-17.12 \pm 0.03	1022 \pm 30 [14]	-17.06
γ -CD	0.4	0.999	3026 \pm 65	-19.72 \pm 0.05	2539 \pm 96 [14]	-19.30
					3038 \pm 45 [54]	-19.74
HP- β -CD	0.1	0.999	725 \pm 30	-16.21 \pm 0.20	580 \pm 16 [14]	-15.67
					563 ^h [48]	-15.59
					630 \pm 18 [54]	-15.87
SBE- β -CD	0.1	0.997	700 \pm 15	-16.12 \pm 0.10	706 \pm 19 ^c [14]	-16.15

	726 ^{h, c} [48]	-16.22
	720 ± 84 ⁱ [54]	-16.20

- a. All literature $K_{1:1}$ values were estimated from phase-solubility diagrams, except for the few that are noted.
- b. The stability constant calculated from the linear range from 0 to 0.012 M.
- c. (SBE)_{7M}- β -CD
- d. Obtained by isothermal titration calorimetry analysis
- e. With an excipient concentration of 0.1% (w/v) hydroxypropyl methylcellulose (HPMC)
- f. Dexamethasone/ γ CD complex in 20 mM HEPES buffer containing 140 mM NaCl and 2 mM CaCl₂ (pH 7.4 at 34 ± 1 °C).
- g. In 0.1 M phosphate buffer at pH = 7 at 0.1 M NaCl and 25 °C.
- h. Determined by UV-vis spectroscopy
- i. SBE_{4,6}- β -CD
- j. Obtained at 25 °C

The change in Gibbs free energy of the steroids/CD inclusion complexes (phase solubility ΔG in kJ mol⁻¹) at 23°C was calculated and reported in Table 1. The free energy of binding for the hydrocortisone/ β -CD inclusion complex was estimated to -20.09 ± 0.15 kJ mol⁻¹. This was -8.21, -0.95, -2.83, and -2.74 kJ mol⁻¹ more favorable than for hydrocortisone in complex with α -CD, γ -CD, HP- β -CD, and SBE- β -CD, respectively. Similarly, the free energy of binding of prednisolone/ β -CD was -6.00, -1.07, -2.27, and -2.09 kJ mol⁻¹ more favorable than for α -CD, γ -CD, HP- β -CD, and SBE- β -CD, respectively. We found that dexamethasone/ γ -CD displayed the most stable inclusion complex with the free energy of binding of -23.43 ± 0.05 kJ mol⁻¹. 6 α -methylprednisolone exhibited a similar pattern for the free energy of binding as for dexamethasone with strongest binding to γ -CD. The increased free energy of binding of both 6 α -methylprednisolone and dexamethasone towards γ -CD is rationalized below. For 6 α -methylprednisolone the added methyl group (Fig. 1) increases steric hindrance and is most likely the main reason for the less favorable free energy of binding when compared to prednisolone in complex with α -CD, β -CD, HP- β -CD, and SBE- β -CD. On the other hand, as γ -CD has a wider cavity and steric hindrance from the additional the methyl group may not play such a large role in inclusion complex formation. Therefore, we conclude that the presence of a methyl group in 6 α -methylprednisolone significantly affects the interaction of the steroid/CD as observed both in Fig. 3 and as presented in Table 1. Similar arguments can be made for dexamethasone which has a fluorine atom that also increases steric hindrance. The large difference in the free energy of

binding between some steroids/ β -CD and other CDs is most likely related to the fact that the size of steroids and the cavity size of β -CD is particularly favorable. For all steroid/CD inclusion complexes we found that α -CD had the least favorable free energy of binding. This is not unexpected due to the size of the steroids and since α -CD in general favor smaller ligands (e.g. linear alkanes) due to the small size of the cavity.

The stability constants found in the literature were also converted to the free energies of binding for comparison (Table 1). Most of the free energy of binding obtained from literature were estimated from phase-solubility diagrams, except a few that was obtained using ITC [49] and UV/Vis-spectroscopy [48], [52]. We found only few studies on the free energy of binding of α -CD with hydrocortisone, dexamethasone, prednisolone, and 6α -methylprednisolone inclusion complexes. We found four different the free energies of binding for hydrocortisone/ β -CD inclusion complex in the literature obtained using similar phase solubility protocols with the free energy of binding values ranging from -17.76 to -20.29 kJ mol⁻¹ [37], [39], [44], [45]. For Dexamethasone in complex with HP- β -CD and SBE- β -CD our obtained the free energy of binding of -18.92 and -19.60 kJ mol⁻¹ are in good agreement with the free energy of binding reported by Usayapant et. al. (-18.93 to -18.96 kJ mol⁻¹) [52] and Jansook et. al. (-19.60 kJ mol⁻¹) [12], respectively. Previous studies have examined the dexamethasone/ γ -CD inclusion complex with a broad distribution of obtained the free energies of binding. Compared to our results, previous results were -23.29 [50], -17.47 [12], -16.95 [51], and -16.79 [51] kJ mol⁻¹, deviate by 0.14, 5.59, 6.48 and 6.64 kJ mol⁻¹ to our result. The free energy of binding of dexamethasone/ β -CD in the literature was found to vary between -21.12 [12] and -16.23 [49] kJ mol⁻¹ with the first value being in very good agreement with our -21.23 \pm 0.18 kJ mol⁻¹ and the latter is underestimated by 5.00 kJ mol⁻¹. We do note that this free energy of binding was obtained with ITC (48).

Taken together, some of the obtained free energies of binding were comparable to previously reported results (Table 1), but different methods may have led to differences between the free energy of binding obtained in this study and the literature. These differences can be due to deviations in the temperature, the specific CD derivative used with various substitution degrees (SBE_{7M}- β -CD [14], [48], [53] and SBE_{4.6}- β -CD [54]), the use of buffer and other additives or the use of different protocols and experimental methods such as UV/Vis-spectroscopy and ITC [48], [49], [52]. We see that in some cases it can be difficult to reproduce the experimental work due to these listed differences and in order to have a consistent dataset for comparison with simulations we performed all experiments with a single consistent protocol.

3.2. Simulation methods

We have assessed to what extent the free energy of binding of the complexes could be reproduced with two end-point methods: docking [27] and MM/GBSA [30], [31]. We have compared the results with experimental data obtained using classical phase solubility studies as presented above. Docking is easily accessible and has seen a resurgence in use in high-through put virtual screening of large libraries for drug discovery [55] although the accuracy can vary across investigated systems [56] as it omits the hosts ability to adjust itself to each ligand. The MM/GBSA approach is based on molecular dynamics simulations and thus both host and ligand are able to adjust themselves during complexation to optimize binding. Due to the use of molecular dynamics, the computational cost is however vastly increased over the simpler docking approach.

3.2.1. Docking with Standard Precision

Docking poses obtained with Glide SP are illustrated in Fig. 4 for the four steroids when docked in α -, β -, and γ -CD. Here we observe that the ligands fit deep into the cavities of β - and γ -CD whereas only a small portion of each ligand is inside the cavity of α -CD. In Table 2, we present the free energies of binding obtained with Glide SP (ΔG_{bind}^{SP}). For hydrocortisone, we obtain ΔG_{bind}^{SP} values of -15.21 ± 0.80 kJ mol⁻¹, -16.01 ± 1.04 kJ mol⁻¹ and -16.76 ± 0.11 kJ mol⁻¹ for α -, β -, and γ -CD, respectively. Compared with the phase solubility experiments presented above these results overestimate the free energy of binding of hydrocortisone and α -CD by 3.3 kJ mol⁻¹. The results are underestimated by roughly 4 kJ mol⁻¹ for β -CD and by 2.4 kJ mol⁻¹ for γ -CD. Due to the structural similarity of all the tested ligands, it is not unexpected that we obtain similar results for dexamethasone, prednisolone, and 6 α -methylprednisolone where results for α -CD are typically overestimated by 2 to 4 kJ mol⁻¹ and both β - and γ -CD are underestimated by 1 to 5 kJ mol⁻¹.

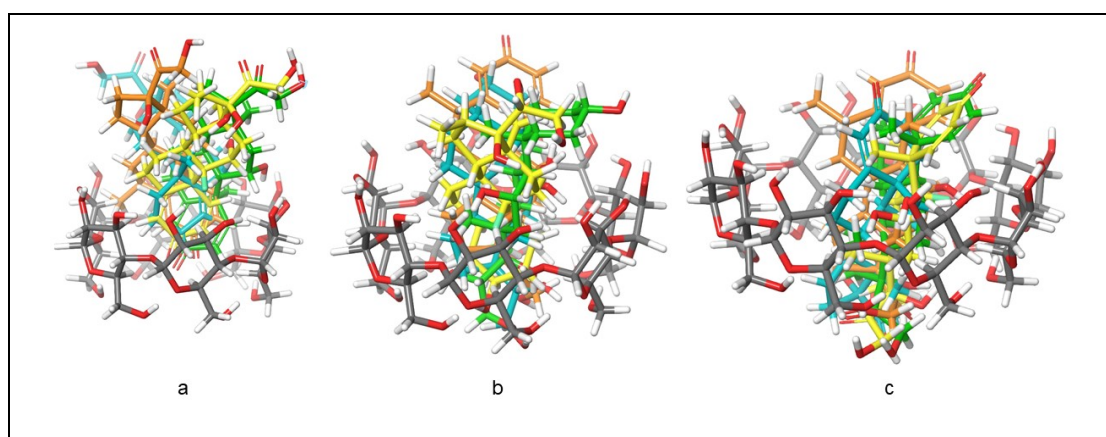


Fig. 4 The typical structures of 1:1 stoichiometry inclusion complex for hydrocortisone (blue), dexamethasone (yellow), prednisolone (green) and 6 α -methylprednisolone (orange) with a) α -CD, b) β -CD and c) γ -CD shown in gray

3.2.2. Docking with Extra Precision

The results of the Glide XP score (ΔG_{bind}^{XP}) are listed in Table 2 and shows that ΔG_{bind}^{XP} for hydrocortisone are 11.41 ± 0.21 kJ mol⁻¹, -15.49 ± 0.82 kJ mol⁻¹ and -14.93 ± 0.80 kJ mol⁻¹ α -, β - and γ -CD, respectively. The free energies of binding obtained with Glide XP are slightly less overestimated for α -CD with for example hydrocortisone/ α -CD is deviating only -0.47 kJ mol⁻¹ compared with $\Delta G_{bind,exp}$. For β - and γ -CD in complex with hydrocortisone the computed free energies of binding (ΔG_{bind}^{XP}) are underestimated by 4.60 and 4.21 kJ mol⁻¹, respectively. With Glide XP, β -CD was predicted to form the strongest inclusion complexes with the free energies of binding of -15.49 , -14.24 , -15.87 , and -16.77 kJ mol⁻¹ for hydrocortisone, dexamethasone, prednisolone, and 6 α -methylprednisolone, respectively.

Table 2. The results of the mean and standard error of $\Delta G_{bind,exp}$, ΔG_{bind}^{XP} , ΔG_{bind}^{SP} and $\Delta G_{bind}^{MM/GBSA}$ for the inclusion complexes between hydrocortisone, dexamethasone, prednisolone, and 6 α -methylprednisolone and native α -, β -, and γ -CD, as well as HP- β -CD and SBE- β -CD, respectively.

	$\Delta G_{bind,exp}$ (kJ mol ⁻¹)	ΔG_{bind}^{XP} (kJ mol ⁻¹)	ΔG_{bind}^{SP} (kJ mol ⁻¹)	$\Delta G_{bind}^{MM/GBSA}$ (kJ mol ⁻¹)
Hydrocortisone				
α -CD	-11.88 ± 0.10	-11.41 ± 0.21	-15.21 ± 0.82	-9.04 ± 0.88
β -CD	-20.09 ± 0.15	-15.49 ± 0.82	-16.01 ± 1.04	-21.42 ± 0.98
γ -CD	-19.14 ± 0.09	-14.93 ± 0.82	-16.76 ± 0.12	-19.39 ± 1.14
Dexamethasone				
α -CD	-13.04 ± 0.37	-10.33 ± 0.41	-15.18 ± 0.14	-5.86 ± 0.88
β -CD	-21.23 ± 0.18	-14.24 ± 0.09	-17.07 ± 0.92	-20.27 ± 1.00
γ -CD	-23.43 ± 0.05	-11.19 ± 0.32	-18.25 ± 0.18	-23.21 ± 1.107
Prednisolone				
α -CD	-13.85 ± 0.47	-11.07 ± 0.19	-15.96 ± 0.15	-11.10 ± 1.08
β -CD	-19.89 ± 0.02	-15.87 ± 0.72	-16.62 ± 1.52	-19.27 ± 0.95
γ -CD	-18.82 ± 0.04	-9.94 ± 0.93	-17.23 ± 0.37	-20.50 ± 1.09

6α- methylprednisolone				
α -CD	-10.69 \pm 0.30	-7.11 \pm 0.79	-15.78 \pm 0.36	-7.92 \pm 0.97
β -CD	-17.12 \pm 0.03	-16.77 \pm 0.31	-16.81 \pm 1.02	-16.18 \pm 1.04
γ -CD	-19.72 \pm 0.05	-16.36 \pm 0.59	-17.57 \pm 0.18	-21.20 \pm 1.13

3.2.3. MM/GBSA

Illustrations of orientation-a and orientation-b between β - and γ -CD/steroids obtained using MM/GBSA are presented in the Supporting Information. As you can see in the Supporting Information, we could not perform MM/GBSA for the orientation-a for α -CD with steroids and in the orientation-b the steroids are located on the wide edge of α -CD, meaning that the benzene ring did not penetrate deep into the cavity of α -CD. The obtained $\Delta G_{\text{bind}}^{\text{MM/GBSA}}$ for α -, β -, and γ -CD/steroids are shown in Table 2. $\Delta G_{\text{bind}}^{\text{MM/GBSA}}$ of hydrocortisone/ α -, β -, and γ -CD are -9.04 \pm 0.88, -21.42 \pm 0.98, and -19.39 \pm 1.14 kJ mol⁻¹, respectively. $\Delta G_{\text{bind}}^{\text{MM/GBSA}}$ is almost the same for α -CD/hydrocortisone compared to the $\Delta G_{\text{bind,exp}}$ obtained from solubility phase experiments but differs by about 2.85 kJ mol⁻¹ and 1.34 kJ mol⁻¹ for the β - and γ -CD/hydrocortisone inclusion complex, respectively.

The $\Delta G_{\text{bind}}^{\text{MM/GBSA}}$ difference for β -CD/6 α -methylprednisolone was 5.02 kJ mol⁻¹ compared to γ -CD/6 α -methylprednisolone, and the $\Delta G_{\text{bind}}^{\text{MM/GBSA}}$ difference for β -CD/prednisolone also was 1.23 kJ mol⁻¹ compared to γ -CD/prednisolone. The obtained $\Delta G_{\text{bind}}^{\text{MM/GBSA}}$ for the complex between the steroids and α -CD were -9.04, -5.86, -11.10 and -7.92 kJ mol⁻¹ for hydrocortisone, dexamethasone, prednisolone, and 6 α -methylprednisolone, respectively. In all MD

simulations, the steroids penetrated the CDs cavity completely except α -CD which is also corroborated by the low the free energies of binding obtained from both simulations and phase-solubility experiments.

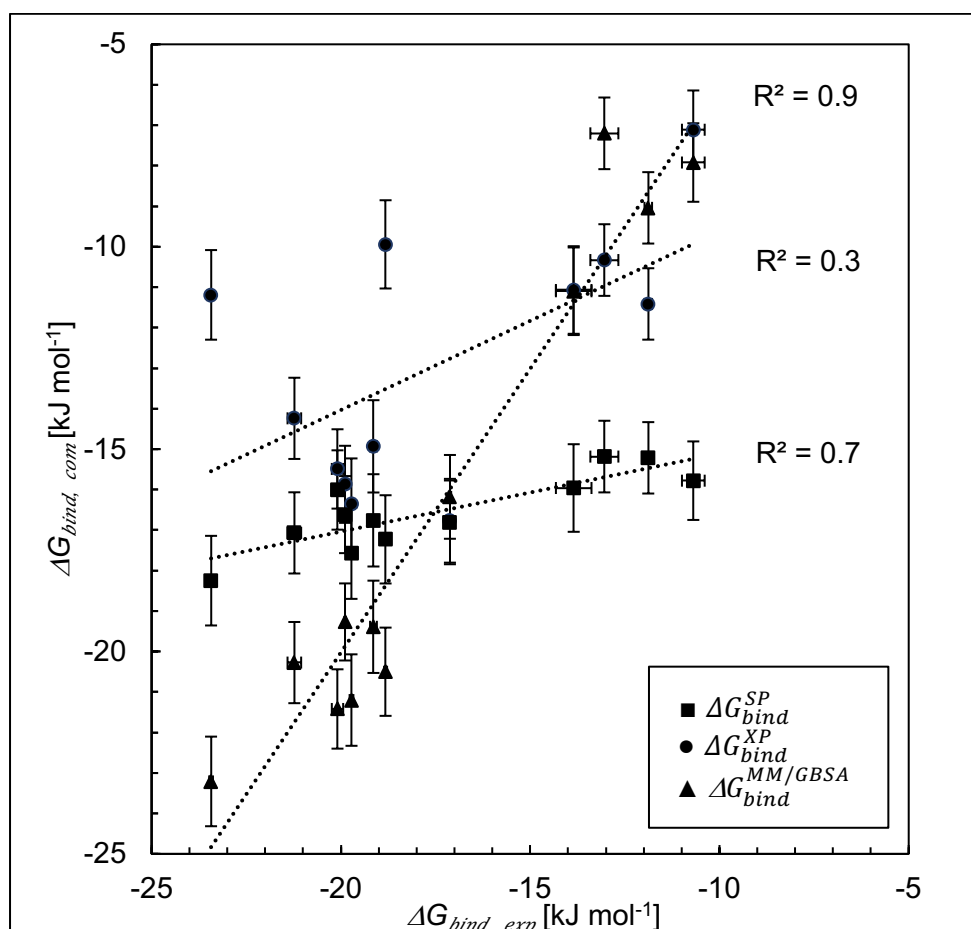


Fig. 5 Scatter plot of experimental $\Delta G_{bind,exp}$ (kJ mol⁻¹) versus simulated $\Delta G_{bind,com}$ (kJ mol⁻¹) [ΔG_{bind}^{SP} (SP): ■, ΔG_{bind}^{XP} (XP): ●, and $\Delta G_{bind}^{MM/GBSA}$: ▲] for α -CD, β -CD and γ -CD with hydrocortisone, dexamethasone, prednisolone, and 6 α -methylprednisolone with standard errors

3.3. Comparison of the free energies of binding obtained using Phase solubility Experiments and Simulations

In Fig. 5, a scatter plot of experimental the free energies of binding ($\Delta G_{bind,exp}$) versus the free energies of binding obtained from simulations ($\Delta G_{bind,com}$) for native CDs in complex with steroids is presented. For standard precision docking (ΔG_{bind}^{SP}), the curve is observed to show some correlation to experimental the free energies of binding with an R^2 value of 0.7. However, as discussed above in detail, the overestimation of binding with α -CD and underestimation for β -CD and γ -CD yields little predictive power of the model as shown in Fig. 5. Also, the free energy of binding for a set of

complexes between β -CD and 57 small organic molecules have been explored in previous studies with MM/GBSA, which showed a very weak correlation ($R^2 = 0.24$) [57]. For extra precision docking (ΔG_{bind}^{XP}) we observe no clear trend (R^2 value of 0.3) and although some the free energies of binding agree more with experiments, others agree less. However, For the MM/GBSA simulation results ($\Delta G_{bind}^{MM/GBSA}$), we observe a good correlation with experimental values across the wide range of native CDs, concluding with an R^2 value of 0.9.

Conclusion

The stability constants from the phase solubility studies obtained for steroids in complex with various CDs showed that complex stability was highly dependent on CD cavity size and type of substitution. HP- β -CD and SBE- β -CD formed weaker inclusion complexes compared to native β -CD, but they were capable of bringing more steroid in solution due to their higher intrinsic solubility and amorphous nature. All the data obtained from the simulations support the formation of steroid/CDs inclusion complexes in solution. The range of obtained free energies of binding varies depending on the simulation method used. We found that docking in general is not reliable when reproducing experimental the free energies of binding to CDs because it either overestimated the free energies of binding greatly for small cavities (α -CD), underestimated binding for larger cavities (β -, and γ -CD) or showed no systematic behaviour at all. We did, however, find that the MM/GBSA method based on molecular dynamics simulations was able to reproduce experimental the free energies of binding with good accuracy. The observed correlation between the computed free energies of binding and those obtained from phase solubility experiments leads us to conclude that simulations based on the MM/GBSA method may be a useful tool to study CD/steroid inclusion complexes before attempting to produce them in the laboratory.

Acknowledgments

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