

# Hyperin up-regulates miR-7031-5P to promote osteogenic differentiation of MC3T3-E1 cells

Dongchen Qian<sup>1,2\*</sup>, Yueyue Chen<sup>3\*</sup>, Xusheng Qiu<sup>4</sup>, Baohua Zhu<sup>2</sup>, Lin Zhang<sup>2</sup>, Yifeng Yan<sup>2</sup> and Yixin Chen<sup>4</sup>

<sup>1</sup>Department of Orthopedic, Nanjing Drum Tower Hospital Clinical College of Nanjing University of Chinese Medicine, <sup>2</sup>Department of Orthopedic, The Second Affiliated Hospital of Nanjing University of Chinese Medicine, <sup>3</sup>Department of Immunology and Rheumatology, Nanjing Hospital of Chinese Medicine Affiliated to Nanjing University of Chinese Medicine and <sup>4</sup>Department of Orthopedic, Nanjing Drum Tower Hospital, The Affiliated Hospital of Nanjing University Medical School, Nanjing, PR China

\*These authors contributed equally to this work

**Summary.** Objective. To investigate the effects of Hyperin (Hyp) on osteogenic differentiation of MC3T3-E1 cells.

Methods. Differentially expressed miRNA was screened by miRNA Microarray. miR-7031-5P overexpression and knockdown MC3T3-E1 cell models were constructed by transfecting miR-7031-5P mimics and inhibitor. Alizarin red staining (ARS) assay was used to observe the formation of mineralized nodules in MC3T3-E1 cells. ALP activity was detected by using ALP detection kit. Western blot assay was used to examine the changes in osteogenic differentiation-related proteins. The relationship between miR-7031-5P and Wnt7a was revealed by dual luciferase report experiments.

Results. We found that miR-7031-5P was up-regulated in MC3T3-E1 cells after Hyp treatment. The results indicated that compared with the untreated group, Hyp promoted the formation of mineralized nodules and the alkaline phosphatase (ALP) activity of MC3T3-E1 cells via overexpressing miR-7031-5P. Besides, elevated miR-7031-5P increased OPN, COL1A1, and Runx2 mRNA expression. More importantly, Wnt7a was identified as the downstream target gene of miR-7031-5P promoting osteogenic differentiation of MC3T3-E1 cells.

Conclusions. Hyp up-regulated miR-7031-5P to promote osteogenic differentiation of MC3T3-E1 cells by targeting Wnt7a.

**Key words:** Hyp, MC3T3-E1, Wnt7a, Osteogenic differentiation, miR-7031-5P

## Introduction

Primary osteoporosis (POP) is a natural physiological degenerative disease, which is characterized by the deterioration of bone microstructure and the decrease of bone density (Ensrud and Crandall, 2017). Studies have shown that once the role of bone resorption is greater than bone formation, the balance between bone formation and bone resorption is broken, resulting in bone loss and bone defect diseases. Bone diseases often cause short-term or long-term physical pain and even disability (Harvey et al., 2010). Besides, bone defects lead to various complications, which usually affect clinical efficacy and are a challenge for orthopedic surgeons (Pepe et al., 2019). On the other hand, promoting osteogenic differentiation has been a crucial strategy for bone loss and bone defect disease repair (Wang et al., 2016). Osteoporosis is the consequence of altered bone metabolism resulting in the systemic reduction of bone strength and increased risk of fragility fractures. MicroRNAs (miRNAs) regulate gene expression on a post-transcriptional level and are known to take part in the control of bone formation and bone resorption (Weilner et al., 2015; Pizzicannella et al., 2019). In addition, Wnt signaling disruption in osteoblastic-lineage cells leads to bone formation defects in osteoporosis (Jing et al., 2018). In recent years, herbal remedies have been found to play roles in regulating the proliferation and differentiation of human mesenchymal stromal cells leading to osteogenic differentiation (Udalamattha et al., 2016). The whole differentiation process of osteoblasts includes mesenchymal stem cells, pre-osteoblasts, mature osteoblasts, and bone cells (Rodan, 1998; Rachner et al., 2011). MC3T3 is a C57BL/6 osteoblast derived from suckling mice. MC3T3-E1, as a subgenus of MC3T3, belongs to the pre-osteoblastic cell line (Wang et al., 2010). In addition, some studies have found that MC3T3-E1 was related to the occurrence, development, and metastasis of certain

*Corresponding Author:* Yixin Chen, Department of Orthopedic, Nanjing Drum Tower Hospital, The Affiliated Hospital of Nanjing University Medical School, Nanjing, Jiangsu Province, 210000, China. e-mail: cheniyixin93@126.com

www.hh.um.es. DOI: 10.14670/HH-18-579



malignant bone tumors (Yang et al., 2016). Besides, an increasing number of documents have demonstrated that MC3T3-E1 cells have active biological properties, and their proliferation and differentiation are essential for the repair of bone loss and bone defect diseases (Choi et al., 1996).

Hyperin (Hyp) as a flavonol glycoside compound, also known as quercetin-3-O- $\beta$ -D galactopyranoside, is widely present in various plants such as Garciniaceae, Leguminosae, Rhododendronaceae, and Euonymae (Wang et al., 2000). Since Nair et al. isolated Hyp from red-osier dogwood in 1960, scholars at home and abroad began to study Hyp and found that Hyp has analgesia, anti-oxidation, and protective effects on myocardium and liver (Nair and Rudloff, 1960; Wang et al., 1996). However, the roles of Hyp in osteogenic differentiation have not been reported in the published documents.

As such, in this study, we found that Hyp treatment made miR-7031-5P expression in MC3T3-E1 cells up-regulated, and then explored the mechanism of Hyp's effects on the osteogenic differentiation of MC3T3-E1 cells. The results indicated that Hyp treatment promoted the formation of mineralized nodules and the alkaline phosphatase (ALP) activity of MC3T3-E1 cells via miR-7031-5P overexpression. Besides, Wnt7a was identified as the downstream target gene of miR-7031-5P promoting osteogenic differentiation of MC3T3-E1. Our study provides a pharmacological basis for Hyp's treatment of bone loss diseases, to better improve the quality of life of patients with bone tissue damage.

## Materials and methods

### Cell culture

Mouse embryonic osteoblast precursor cell line (MC3T3-E1 cells) was purchased from Sciencell Corporation (Carlsbad, California, USA). Hyp was provided by Shanghai Enzyme Link Biotechnology Co., Ltd (Shanghai, China). The cells were cultured in DMEM medium (Gibco, #1868985) with Fetal Bovine Serum (Gibco, #10091-148), placing in an incubator (SANYO, Cat. No. MCO-175) with 37°C containing 5% CO<sub>2</sub>. In terms of osteogenic differentiation, MC3T3-E1 cells were treated with osteogenic medium (containing 10% Fetal Bovine Serum, 5 mmol/L  $\alpha$ -sodium glycerophosphate, 50  $\mu$ g/ml ascorbic acid and 10<sup>-8</sup> mol/L dexamethasone) for 2 to 3 days to induce

osteogenic differentiation. MC3T3-E1 cells cultured in complete medium served as a negative control group.

### Hyp treated MC3T3-E1 cells to screen differentially expressed miRNA

In the preliminary experiments, we treated MC3T3-E1 cells using Hyp with different concentration gradients (0, 20, 40, 80, 100, 120, 150, 200  $\mu$ mol/L). The optimal concentration of Hyp was defined as 80  $\mu$ mol/L by measuring the degree of cell differentiation, the ALP activity, and the formation of mineralized nodules in cells (Figs. 1-3). The experiment was divided into two groups: MC3T3-E1 cells with Hyp (80  $\mu$ mol/L) and the control group (MC3T3-E1 cells without Hyp). After Hyp treatment for 7 days, the differentially expressed miRNA between the Hyp treatment group and control group was screened by Agilent Mouse miRNA Microarray Kit (Release 21.0, 8x60K).

### RNA extraction and qRT-PCR

The total RNA was isolated according to the manufacturer's protocol of the Trizol reagent (Sigma, St. Louis, MO, USA), and the production was reversely transcribed to obtain cDNA using the Hiscript QRT supermix kit (Vazyme, Nanjing, Jiangsu, China). The system of qRT-PCR was 10  $\mu$ L according to SYBR Green Mastermix Kit (Vazyme, Nanjing, Jiangsu, China). U6 was used as an internal reference gene. The relative expression of mRNA was calculated by the 2<sup>- $\Delta\Delta$ Ct</sup> method. The primer sequences (5'-3') are listed in Table 1.

### The construction of miR-7031-5P overexpression and knockdown cell models

MC3T3-E1 cells in logarithmic growth period were seeded in a 24-well plate and cultured in serum-free  $\alpha$ -MEM medium without dual antibodies to make the cell confluence reach 50%. 5  $\mu$ L miR-7031-5P mimics and inhibitor were added into the 24-well plate at room temperature for 3-5 min. 1  $\mu$ L Lipofectamine 2000 was subsequently added and cultured for 5 min at room temperature. After 24h of transfection, qRT-PCR was used to determine the transfection efficiency. The sequences of miR-7031-5P overexpression and knockdown were as follows: miR-7031-5P over-

Table 1. Primers used for qRT-PCR

Gene	Forward primer sequence	Reverse primer sequence
U6	CTCGCTTCGGCAGCAC	AACGCTTCACGAATTTGCGT
MIR-7031-5P	GTGGGAGCAGCCTTGAAT	GTGCAGGGTCCGAGGT
OPN	CTATCCAGCCACCTTCACTACA	TCGCCAGACAGACTCATCCA
COL1A1	TGCTCGTGGAAATGATGGTG	GGAGCACCATTGGCACCTTT
RUNX	TGGCTTACGGACTGAGGT	TTGGAACCTGCCTGACTG
Wnt7a	CTGGCCTTCCACTCTCAGA	AGTCGCGGGCTGTAAGTAG

Hyperin promotes MC3T3-E1 cells' osteogenic differentiation

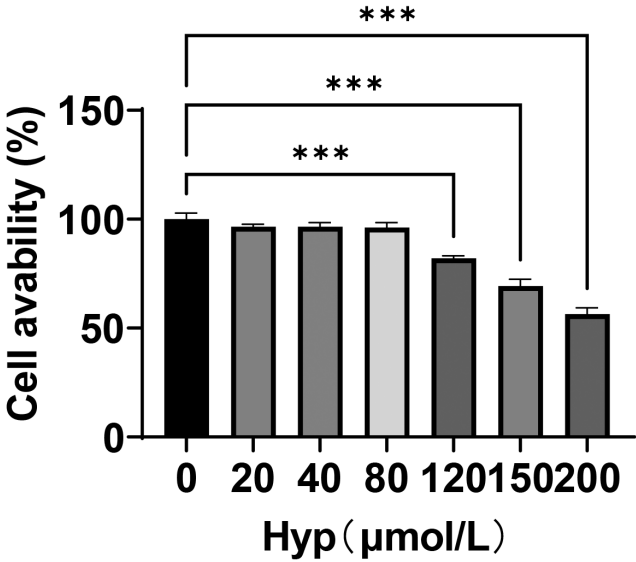


Fig. 1. The different concentrations of Hyp on MC3T3-E1 cell viability. \*\*\*P<0.001.

expression- ACATATAGGGTTCTTGCAGCTTGCC  
 GAGAACTGTTCCCTCAGGTGGGAAGTTGTT  
 GCAAGAGAAGGATGCTCTGTGCCTCCTTCTG  
 GAGAACTACAGACAACCTGAGAGGCCTGAAG  
 GGTGGGAGCAGCCTTGAATGACCAACCCTCTG  
 CCCTCTCCTAGAGCTCAAAGCCCAAGCTCCAAG  
 AGGCCTGTTCCAGCATCCGGGAGGTCCGACCAT  
 GTACACGCCTGGAGATGCCTGACAACCTCTACA  
 CCTTTGTGTTGATTTTTT; miR-7031-5P knockdown-  
 TCCCACCCTTCAGGCCTCTCAGGCTTCTCCCACC  
 CTTTCAGGCCTCTCAGGCTTCTCCCACCCTTCAGG  
 CCTCTCAGGCTTCTCCCACCCTTCAGGCCTCTCA  
 GGCTTCTCCCACCCTTCAGGCCTCTCAGGCTTCT  
 CCCACCCTTCAGGCCTCTCAGGCTTCTTTTTT.

Alizarin red staining (ARS)

After MC3T3-E1 cells transfected with miR-7031-5P mimics and inhibitor were treated with Hyp, the culture medium with or without osteogenic induction fluid was added to culture cells for 14 or 21 days. 2 mL of 4% paraformaldehyde was added, and then discarded

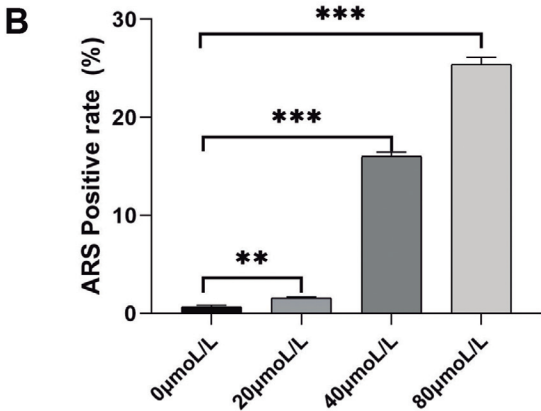
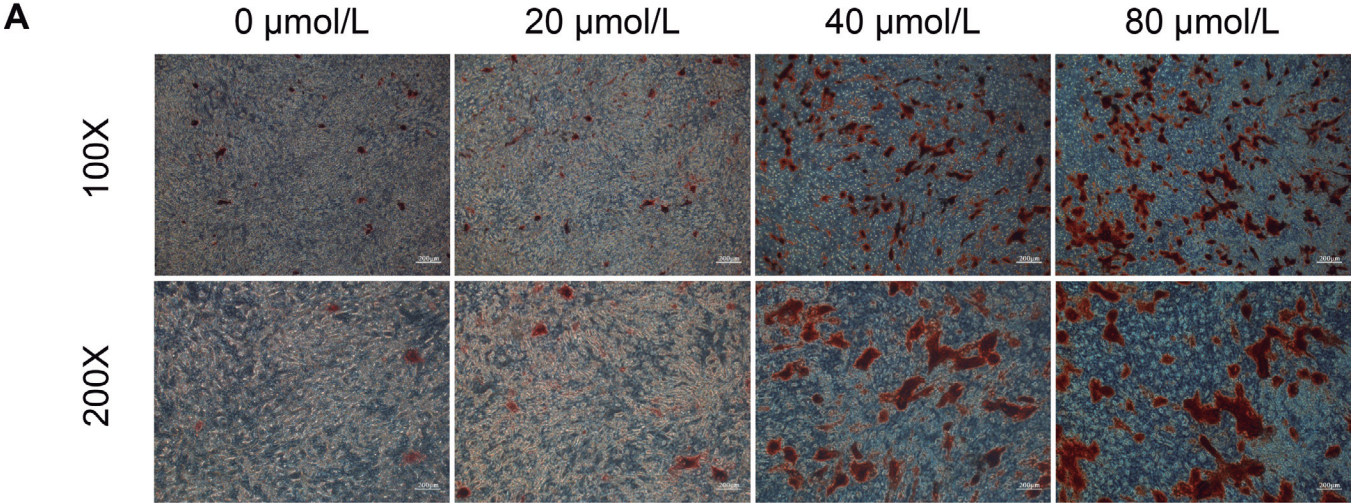


Fig. 2. Hyp promoted the formation of mineralized nodules of MC3T3-E1 cells in a dose-dependent manner. A. The formation of mineralized nodules of MC3T3-E1 cells with a dose gradient of Hyp was detected by ARS assay. B. The quantification of relative ARS positive rate under a dose gradient Hyp treatment in MC3T3-E1 cells. n=6 independent capture area, \*\*P<0.05, \*\*\*P<0.01.

### Hyperin promotes MC3T3-E1 cells' osteogenic differentiation

after 15 min. After that, 2 mL ARS staining solution was used to detect the formation of mineralized nodules of MC3T3-E1 cells in different groups. Finally, the cells were observed under an inverted microscope and pictures were taken.

#### ALP activity detection

After MC3T3-E1 cells transfected with miR-7031-5P mimics and inhibitor were treated with Hyp, the culture medium with or without osteogenic induction fluid was added to culture cells for 7 days. The ALP activity of MC3T3-E1 cells was detected using ALP activity detection kit (Jiancheng, Nanjing). After adding the chromogenic reagent to the cells, they were placed in an enzyme-linked immunoassay at 520 nm to measure the absorbance value. Finally, the ALP activity in MC3T3-E1 cells was calculated.

#### Dual-Luciferase report experiment

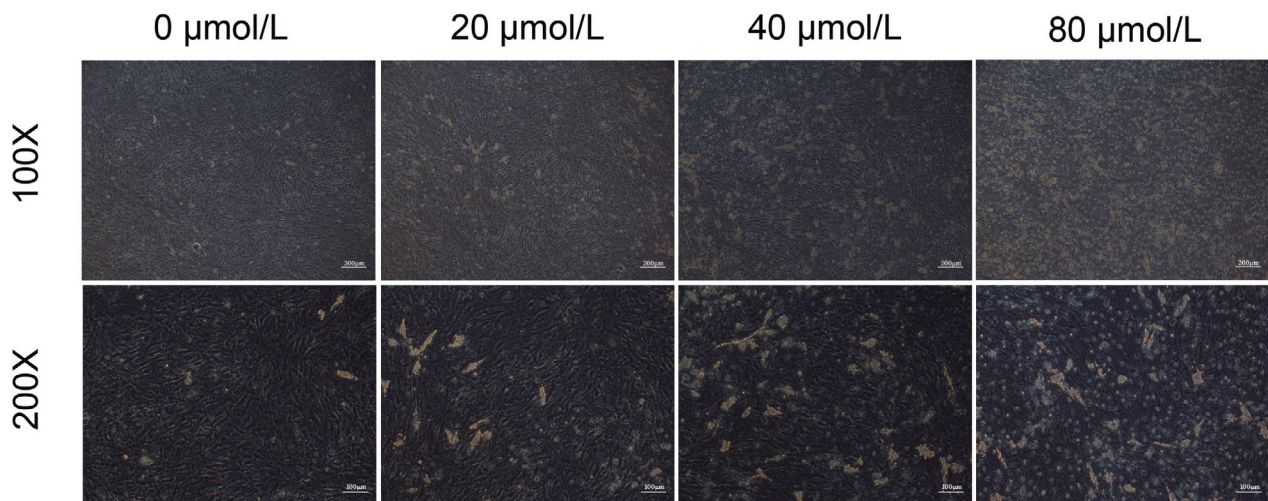
The Dual-luciferase report gene plasmid was

transfected into MC3T3-E1 cells by Lipofectamine 2000. According to the manufacturer's protocol of Promega Dual-Luciferase system, a 96-well plate was prepared by adding diluted Passive Lysis Buffer, and MC3T3-E1 cells after Hyp treatment were harvested and centrifuged at 12000 rpm for 10 min to collect cell supernatant. After that, 100  $\mu$ L Luciferase Assay Reagent II (LAR II) (Luciferase Assay Reagent, Progenia) was added into the above 96-well plate. Then, 20  $\mu$ L cell lysate was added, and the value of Firefly luciferase was measured and recorded, which was used as the internal reference value. Finally, 100  $\mu$ L Stop and Glo<sup>®</sup> Reagent (Luciferase Assay Reagent, Progenia) was added and the Renilla luciferase was recorded as the luminescence value of the reporter gene.

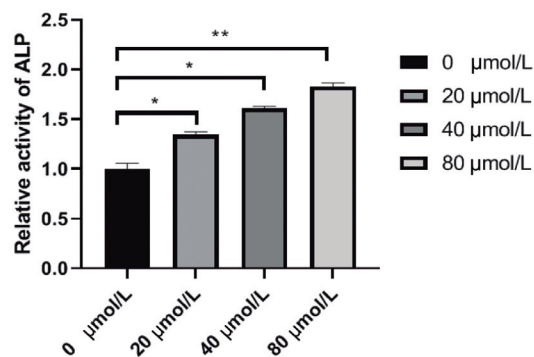
#### Western blot assay (WB)

After Hyp treatment, MC3T3-E1 cells were harvested to collect the total proteins. At the same time, 10% SDS-PAGE was used to segregate the total proteins and transferred into PVDF membranes followed by blocking

**A**



**B**



**Fig. 3.** Hyp enhanced the alkaline phosphatase (ALP) activity of MC3T3-E1 cells in a dose-dependent manner. **A.** The ALP activity of MC3T3-E1 cells with a dose gradient of Hyp was detected by ALP activity kit. **B.** The quantification of relative activity of ALP under a dose gradient Hyp treatment in MC3T3-E1 cells. n=6 independent capture areas, \* $P$ <0.05, \*\* $P$ <0.01.

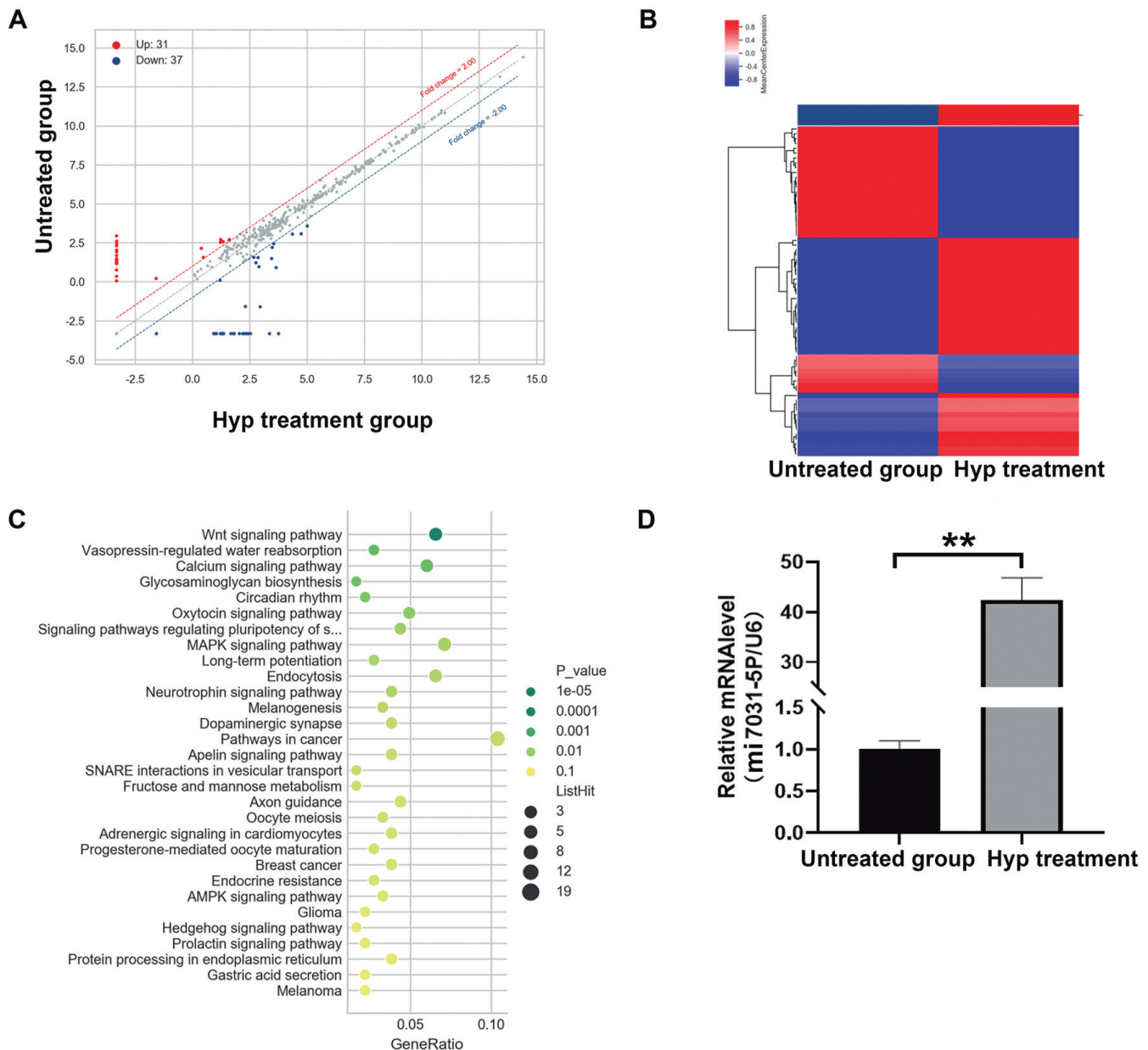
## Hyperin promotes MC3T3-E1 cells' osteogenic differentiation

with TBST solution containing 5% skim milk at room temperature for 1h. Next, the membranes were incubated with primary and secondary antibodies and washed with TBST solution three times (10 min/time). Finally, the ECL+plus™ Western blotting system kit was used for color rendering and X-ray imaging was captured. The primary antibodies used in WB were as follows: OPN (1:1000, abcam, #ab11503), COL1A1 (1:1000, Beyotime, #AF1840), RUNX2 (1:1000, CST, #8486s) and GAPDH

(1:3000, Bioworld, #AP0063). The secondary antibody was Goat Anti-Rabbit (1:3000, Beyotime, #A0208), Goat Anti-Mouse (1:3000, Beyotime, #A0216) and Donkey Anti-Goat (1:3000, Beyotime, #A0181).

### Statistical analysis

All data were analyzed by using GraphPad Prism 8 (San Diego, CA, USA) and SPSS 19.0 (IBM, SPSS,



**Fig. 4.** Screening of differentially expressed miRNAs. **A.** The differentially expressed miRNAs were evaluated between Hyp treatment group and untreated group by scatter plots. **B.** The differentially expressed miRNAs were evaluated between Hyp treatment group and untreated group by cluster analysis. **C.** The pathways related to differentially expressed genes were evaluated by Kyoto Encyclopedia of Genes and Genomes (KEGG) analysis. **D.** miR-7031-5P expression level in MC3T3-E1 cells after Hyp treatment was revealed by qRT-PCR. Results were presented as mean  $\pm$  SD. \*\* $P < 0.01$ .

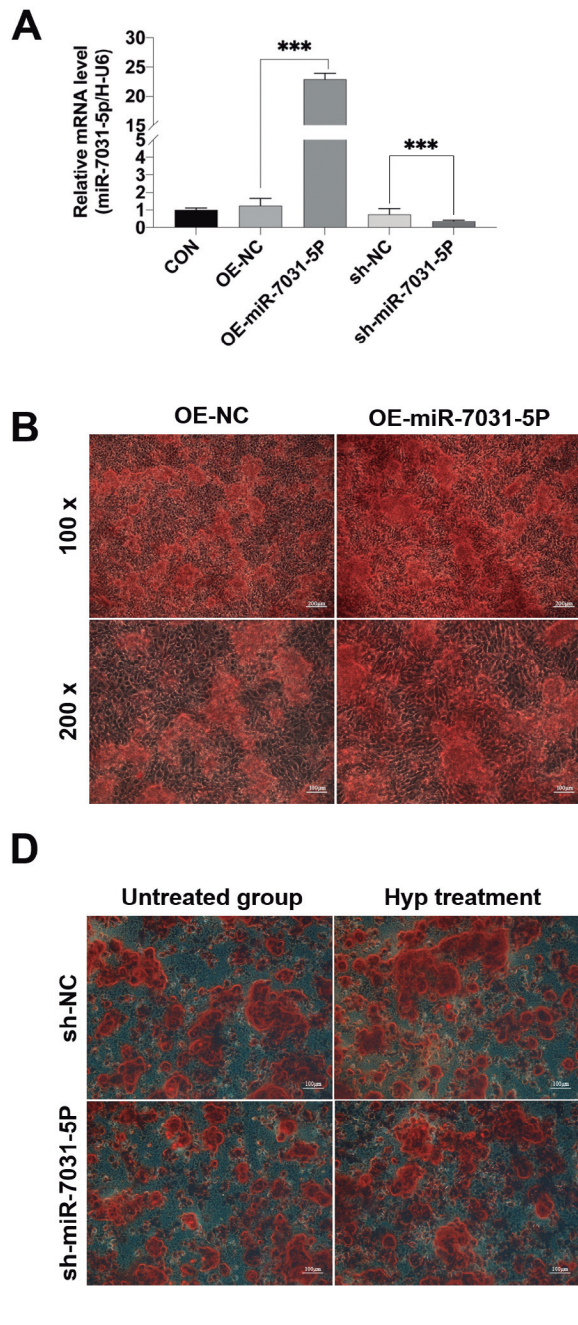


*Hyperin promotes MC3T3-E1 cells' osteogenic differentiation*

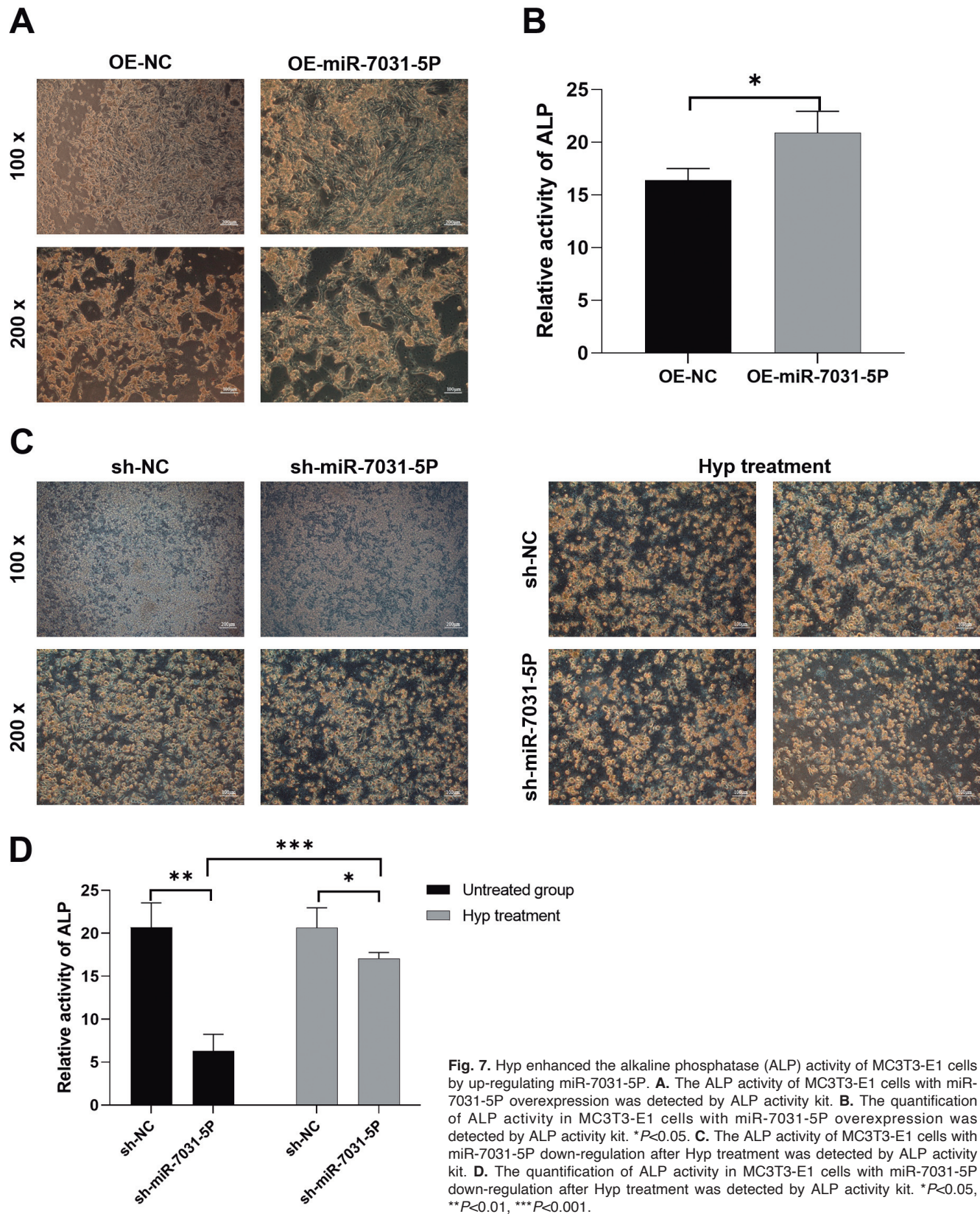
above studies, indicating that Hyp promoted ALP activity in MC3T3-E1 cells by up-regulating miR-7031-5P.

*Hyp up-regulated OPN, COL1A1 and Runx2 expression in MC3T3-E1 cells via miR-7031-5P overexpression*

In this section, we wanted to evaluate osteogenic differentiation-related protein levels in lentivirus infected MC3T3-E1 cells. We noticed that, from qPCR assay, OPN, COL1A1 and Runx2 mRNA expression was evidently increased in the OE-miR-7031-5P group compared with the OE-NC group, while decreased OPN, COL1A1 and Runx2 were noted in the sh-miR-7031-5P group (Fig. 8A). Finally, combining the published literature with bioinformatics online analysis (TargetsCan), we again identified Wnt7a as the downstream target gene of miR-7031-5P promoting osteogenic differentiation of MC3T3-E1. Additional dual luciferase report experiments demonstrated that there was an interaction between miR-7031-5P and Wnt7a (Fig. 8B). Furthermore, we detected Wnt7a



**Fig. 6.** Hyp promoted the formation of mineralized nodules of MC3T3-E1 cells via miR-7031-5P overexpression. **A.** The transfection efficiencies of miR-7031-5P mimics and inhibitor in MC3T3-E1 cells were evaluated through qRT-PCR. **B.** The formation of mineralized nodules of MC3T3-E1 cells with miR-7031-5P overexpression was detected by ARS assay. **C.** The quantification of relative ARS positive rate of MC3T3-E1 cells with miR-7031-5P overexpression was detected by ARS assay. **\*\*** $P < 0.01$ . **D.** The formation of mineralized nodules of MC3T3-E1 cells with miR-7031-5P down-regulation after Hyp treatment was detected by ARS assay. **E.** The quantification of relative ARS positive rate of MC3T3-E1 cells with miR-7031-5P down-regulation after Hyp treatment by ARS assay. **\*** $P < 0.05$ .





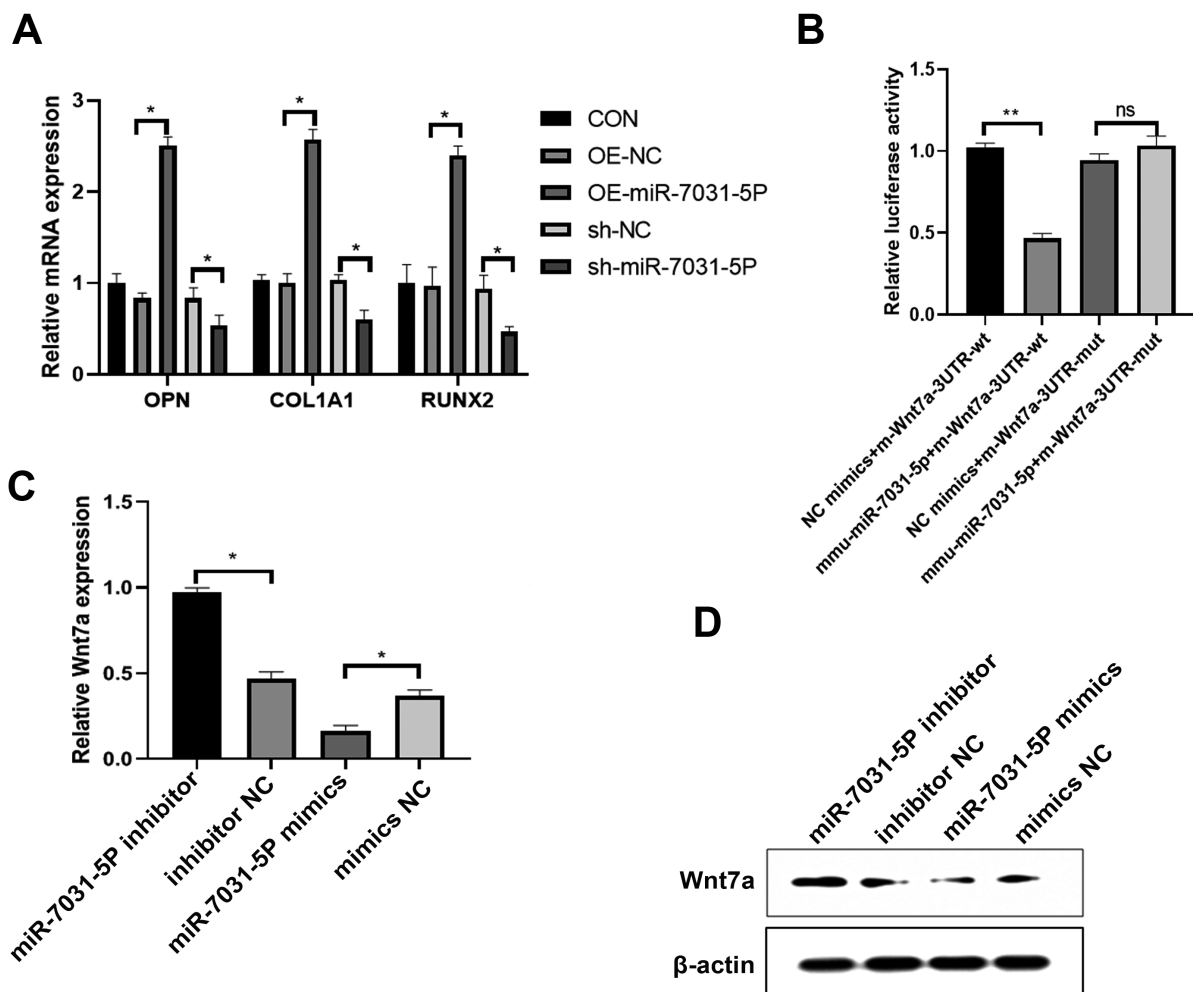
*Hyperin promotes MC3T3-E1 cells' osteogenic differentiation*

mRNA and protein levels in miR-7031-5P overexpression and knockdown MC3T3-E1 cells. As expected, elevated miR-7031-5P down-regulated Wnt7a mRNA and protein levels, while silencing miR-7031-5P up-regulated Wnt7a mRNA and protein levels (Fig. 8C,D). As per the results presented so far, we concluded that Hyp up-regulated miR-7031-5P to promote osteogenic differentiation of MC3T3-E1 cells by targeting Wnt7a.

### Discussion

miRNAs are closely related to the osteogenic differentiation of bone marrow mesenchymal stem cells (BMSCs), and play an important regulatory role in the process of bone tissue metabolism (Long, 2011). A large number of studies have shown that a variety of intracellular and extracellular signal transmission is

involved in the process of osteogenic differentiation, such as miRNA and signal pathways (Hankenson et al., 2015). For example, the level of miR-29b increases in the process of osteogenic differentiation, which can down-regulate the expression of COL4A2, COL5A3 and COL1A1, eventually promoting osteogenic differentiation (Li et al., 2009). An important cause of bone loss and bone defect diseases is that osteogenic differentiation is inhibited or weakened, which is in turn associated with the abnormal expression of miRNAs (Arfat et al., 2015; Hata and Kang, 2015). In this current study, 80  $\mu\text{mol}\cdot\text{L}^{-1}$  Hyp was used to treat MC3T3-E1 cells. The results indicated that miR-7031-5P expression level was markedly up-regulated in MC3T3-E1 cells after Hyp treatment. Besides, the ability of mineralized nodule formation and the ALP activity in MC3T3-E1 cells after Hyp treatment was significantly increased, suggesting that Hyp promoted the osteogenic



**Fig. 8.** Exploration of downstream mechanisms of miR-7031-5P regulating the osteogenic differentiation of MC3T3-E1 cells. **A**, Hyp up-regulated OPN, COL1A1 and Runx2Runx2 expression in MC3T3-E1 cells via miR-7031-5P overexpression **B**. The relationship between miR-7031-5P and Wnt7a was revealed by the dual-luciferase report experiments. **C**, **D**. Wnt7a mRNA and protein levels were determined in MC3T3-E1 cells with miR-7031-5P overexpression and down-regulation by qRT-PCR (**C**) and WB (**D**) assays. \* $P < 0.05$ , \*\* $P < 0.01$ .

differentiation of MC3T3-E1 cells via up-regulating miR-7031-5P. More importantly, we found that Wnt7a was the potential downstream target gene of miR-7031-5P inhibiting osteogenic differentiation of MC3T3-E1.

Growing evidence has confirmed that Wnt plays a key role in the process of osteogenic differentiation of human mesenchymal stem cells (de Boer et al., 2004). The canonical Wnt- $\beta$ -catenin signaling pathway seems to explain most of the effects of Wnt signaling in the skeleton (Canalis, 2013). The canonical Wnt signaling pathway is mediated by  $\beta$ -catenin (Maeda et al., 2019). The non-canonical signaling pathway is a generic term used for pathways not mediated by  $\beta$ -catenin. Ligands such as Wnt5a and Wnt11 activate the Wnt/ $\text{Ca}^{2+}$  and Wnt/PCP pathways without the induction of intracellular  $\beta$ -catenin accumulation. In the Wnt/ $\text{Ca}^{2+}$  pathway, the increased intracellular concentration of  $\text{Ca}^{2+}$  activates calmodulin-dependent protein kinase II (CaMK II) and protein kinase C (PKC). In the Wnt/planar cell polarity (PCP) pathway, small G proteins such as Rac and Rho are activated to enhance cell motility as well as determining the direction and localization of cilia (Baron and Kneissel, 2013; Karner and Long, 2017). Noncanonical Wnt signaling also regulates chondrocyte differentiation (Bradley and Drissi, 2010). Crosstalk of prototypical canonical (Wnt3a), and non-canonical (Wnt5a) Wnts leads to functional antagonism during osteogenic differentiation (Baksh et al., 2007). Furthermore,  $\beta$ -catenin plays a central role in regulating osteogenic differentiation of MSCs in inflammatory microenvironments through both the canonical Wnt/ $\beta$ -catenin pathway and the noncanonical Wnt/ $\text{Ca}^{2+}$  pathway (Liu et al., 2011). WNT7A forms a receptor complex with LRP6 and FZD5 to activate the canonical WNT signaling pathway (Lan et al., 2019). In stem cells, Wnt7a was revealed to promote differentiation of mesenchymal stem cells into bone (Yang et al., 2021) and bone mesenchymal stem cells osteogenic differentiation (Li et al., 2015). Msx2 exerts bone anabolism in part by reducing Dkk1 expression and enhancing Wnt7a and Wnt7b in primary mouse bone cells (Cheng et al., 2008). However, on the other hand, the pro-inflammatory cytokine IL-1 $\beta$  has been reported to induce Wnt5a and Wnt7a expression in primary culture articular chondrocytes. It was also revealed that Wnt7a induces dedifferentiation and inhibits NO-induced apoptosis of primary culture articular chondrocytes (Li et al., 2015). So the function of Wnt7a would be different in different cell types. In the present study, we identified Wnt7a as a potential target gene of miR-7031-5P. Hyp upregulated miR-7031-5P expression in MC3T3-E1 cells. We speculated that Hyp promoted osteogenic differentiation through miR-7031-5p/Wnt7a program. However, miR-7031-5p has so many potential target genes and Wnt7a was only a middle regulator that could regulate downstream genes of osteogenesis. So we still didn't know the exact intricate regulatory network involved in Hyp mediated osteogenesis. That should be researched more deeply in the future.

Many studies have shown that miRNAs are important epigenetic regulators of Wnt signaling genes during bone differentiation (Amjadi-Moheb and Akhavan-Niaki, 2019). In detail, in the process of osteogenic differentiation mediated by the Wnt signaling pathway, miRNAs bind to target genes to regulate Runx2 expression and ALP activity, thereby regulating the Wnt signaling pathway and osteogenic differentiation. For example, Zhang et al. found that miR-355-5p down-regulated DKK1, enhanced the Wnt signaling pathway, increased the expression levels of osteogenic differentiation-related factors such as Runx2, and promoted the osteogenic differentiation of BMSCs (Zhang et al., 2011). Su et al. demonstrated that miR-26a promoted osteogenic differentiation of BMSCs through the Wnt/ $\beta$ -catenin signaling pathway (Su et al., 2015). MiR-29a-3p had been found to inhibit proliferation and osteogenic differentiation of human BMSCs via targeting FOXO3 and repressing Wnt/ $\beta$ -catenin signaling pathway signaling in steroid-associated osteonecrosis (Wang et al., 2022).

In conclusion, the results of this study showed that Hyp played an important role in osteogenic differentiation of MC3T3-E1, mainly through miR-7031-5P overexpression to promote osteogenic differentiation, which may provide a promising therapy option for bone-related diseases.

---

*Acknowledgements.* This work was financially supported by the Nanjing Medical Science and Technique Development Foundation (grant numbers JQX16021 and QRX17050) and the Inner-hospital Project of the Second Affiliated Hospital of Nanjing University of Chinese Medicine (SEZ202025).

*Author contributions.* Dongchen Qian, Yueyue Chen, and Yixin Chen designed this research. Xusheng Qiu and Yueyue Chen performed the experiments. Baohua Zhu, Quanhong Yang, and Yifeng Yan conducted the data processing and analysis. Dongchen Qian completed the manuscript which was reviewed by Yixin Chen. All the authors have confirmed the submission of this manuscript.

*Conflict of interests.* The authors declare that they have no conflict of interest.

---

## References

- Amjadi-Moheb F. and Akhavan-Niaki H. (2019). Wnt signaling pathway in osteoporosis: Epigenetic regulation, interaction with other signaling pathways, and therapeutic promises. *J. Cell. Physiol.* 234, 14641-14650.
- Arfat Y., Xiao W.Z., Ahmad M., Zhao F., Li D.J., Sun Y.L., Hu L., Zhihao C., Zhang G., Iftikhar S., Shang P., Yang T.M. and Qian A.R. (2015). Role of microRNAs in osteoblasts differentiation and bone disorders. *Curr. Med. Chem.* 22, 748-758.
- Baksh D., Boland G.M. and Tuan R.S. (2007). Cross-talk between Wnt signaling pathways in human mesenchymal stem cells leads to functional antagonism during osteogenic differentiation. *J. Cell. Biochem.* 101, 1109-1124.
- Baron R. and Kneissel M. (2013). Wnt signaling in bone homeostasis and disease: From human mutations to treatments. *Nat. Med.* 19,

## Hyperin promotes MC3T3-E1 cells' osteogenic differentiation

- 179-192.
- Bradley E.W. and Drissi M.H. (2010). WNT5A regulates chondrocyte differentiation through differential use of the CaN/NFAT and IKK/NF-KAPPAB pathways. *Mol. Endocrinol.* 24, 1581-1593.
- Canalis E. (2013). Wnt signalling in osteoporosis: Mechanisms and novel therapeutic approaches. *Nat. Rev. Endocrinol.* 9, 575-583.
- Cheng S.L., Shao J.S., Cai J., Sierra O.L. and Towler D.A. (2008). Msx2 exerts bone anabolism via canonical Wnt signaling. *J. Biol. Chem.* 283, 20505-20522.
- Choi J.Y., Lee B.H., Song K.B., Park R.W., Kim I.S., Sohn K.Y., Jo J.S. and Ryoo H.M. (1996). Expression patterns of bone-related proteins during osteoblastic differentiation in MC3T3-E1 cells. *J. Cell. Biochem.* 61, 609-618.
- de Boer J., Siddappa R., Gaspar C., van Apeldoorn A., Fodde R. and van Blitterswijk C. (2004). Wnt signaling inhibits osteogenic differentiation of human mesenchymal stem cells. *Bone* 34, 818-826.
- Ensrud K.E. and Crandall C.J. (2017). Osteoporosis. *Ann. Intern. Med.* 167, ITC17-ITC32.
- Hankenson K.D., Gagne K. and Shaughnessy M. (2015). Extracellular signaling molecules to promote fracture healing and bone regeneration. *Adv. Drug Deliv. Rev.* 94, 3-12.
- Harvey N., Dennison E. and Cooper C. (2010). Osteoporosis: Impact on health and economics. *Nat. Rev. Rheumatol.* 6, 99-105.
- Hata A. and Kang H. (2015). Functions of the bone morphogenetic protein signaling pathway through microRNAs (review). *Int. J. Mol. Med.* 35, 563-568.
- Jing H., Su X., Gao B., Shuai Y., Chen J., Deng Z., Liao L. and Jin Y. (2018). Epigenetic inhibition of Wnt pathway suppresses osteogenic differentiation of BMSCs during osteoporosis. *Cell Death Dis.* 9, 176.
- Karner C.M. and Long F. (2017). Wnt signaling and cellular metabolism in osteoblasts. *Cell. Mol. Life. Sci.* 74, 1649-1657.
- Lan L., Wang W., Huang Y., Bu X. and Zhao C. (2019). Roles of Wnt7a in embryo development, tissue homeostasis, and human diseases. *J. Cell. Biochem.* 120, 18588-18598.
- Li Q., Dou X. and Kan S. (2015). Function and mechanisms of Wnt7a in bone mesenchymal stem cells osteogenic differentiation. *Int. J. Biomed. Eng.* 38, 32-35.
- Li Z., Hassan M.Q., Jafferji M., Aqeilan R.I., Garzon R., Croce C.M., van Wijnen A.J., Stein J.L., Stein G.S. and Lian J.B. (2009). Biological functions of miR-29b contribute to positive regulation of osteoblast differentiation. *J. Biol. Chem.* 284, 15676-15684.
- Liu N., Shi S., Deng M., Tang L., Zhang G., Liu N., Ding B., Liu W., Liu Y., Shi H., Liu L. and Jin Y. (2011). High levels of  $\beta$ -catenin signaling reduce osteogenic differentiation of stem cells in inflammatory microenvironments through inhibition of the noncanonical Wnt pathway. *J. Bone. Miner. Res.* 26, 2082-2095.
- Long F. (2011). Building strong bones: Molecular regulation of the osteoblast lineage. *Nat. Rev. Mol. Cell Biol.* 13, 27-38.
- Maeda K., Kobayashi Y., Koide M., Uehara S., Okamoto M., Ishihara A., Kayama T., Saito M. and Marumo K. (2019). The regulation of bone metabolism and disorders by Wnt signaling. *Int. J. Mol. Sci.* 20, 5525.
- Nair G.V. and Rudloff E.V. (1960). Isolation of hyperin from red-osier dogwood (*Cornus stolonifera* michx.). *Can. J. Chem.* 38, 2531-2533.
- Pepe J., Cipriani C., Cecchetti V., Ferrara C., Della Grotta G., Danese V., Colangelo L. and Minisola S. (2019). Patients' reasons for adhering to long-term alendronate therapy. *Osteoporos. Int.* 30, 1627-1634.
- Pizzicannella J., Diomedede F., Gugliandolo A., Chiricosta L., Bramanti P., Merciaro I., Orsini T., Mazzon E. and Trubiani O. (2019). 3D printing PLA/Gingival stem cells/Evs upregulate miR-2861 and -210 during osteoangiogenesis commitment. *Int. J. Mol. Sci.* 20, 3256.
- Rachner T.D., Khosla S. and Hofbauer L.C. (2011). Osteoporosis: Now and the future. *Lancet* 377, 1276-1287.
- Rodan G.A. (1998). Control of bone formation and resorption: Biological and clinical perspective. *J. Cell. Biochem. (Suppl. 30-31)*, 55-61.
- Su X., Liao L., Shuai Y., Jing H., Liu S., Zhou H., Liu Y. and Jin Y. (2015). Mir-26a functions oppositely in osteogenic differentiation of BMSCs and ADSCs depending on distinct activation and roles of Wnt and BMP signaling pathway. *Cell Death Dis.* 6, e1851.
- Udalarnaththa V.L., Jayasinghe C.D. and Udagama P.V. (2016). Potential role of herbal remedies in stem cell therapy: Proliferation and differentiation of human mesenchymal stromal cells. *Stem Cell Res. Ther.* 7, 110.
- Wang W.Q., Ma C.G. and Xu S.Y. (1996). Protective effect of hyperin against myocardial ischemia and reperfusion injury. *Zhongguo. Yao. Li. Xue. Bao.* 17, 341-344.
- Wang, X.-W., Ma, C.-G., Xu and S.-Y. (2000). Hyperin. *Analgesic. Drugs Future* 25, 347-350.
- Wang A., Ding X., Sheng S. and Yao Z. (2010). Bone morphogenetic protein receptor in the osteogenic differentiation of rat bone marrow stromal cells. *Yonsei Med. J.* 51, 740-745.
- Wang C., Wang J., Li J., Hu G., Shan S., Li Q. and Zhang X. (2016). KDM5A controls bone morphogenetic protein 2-induced osteogenic differentiation of bone mesenchymal stem cells during osteoporosis. *Cell Death Dis.* 7, e2335.
- Wang C., Zhu M., Yang D., Hu X., Wen X. and Liu A. (2022). MiR-29a-3p inhibits proliferation and osteogenic differentiation of human bone marrow mesenchymal stem cells via targeting FOXO3 and repressing Wnt/ $\beta$ -catenin signaling in steroid-associated osteonecrosis. *Int. J. Stem. Cells.* 15, 324-333.
- Weilner S., Skalicky S., Salzer B., Keider V., Wagner M., Hildner F., Gabriel C., Dovjak P., Pietschmann P., Grillari-Voglauer R., Grillari J. and Hackl M. (2015). Differentially circulating miRNAs after recent osteoporotic fractures can influence osteogenic differentiation. *Bone* 79, 43-51.
- Yang D., Okamura H., Morimoto H., Teramachi J. and Haneji T. (2016). Protein phosphatase 2A Ca regulates proliferation, migration, and metastasis of osteosarcoma cells. *Lab. Invest.* 96, 1050-1062.
- Yang L., Li Q., Zhang J., Li P., An P., Wang C., Hu P., Zou X., Dou X. and Zhu L. (2021). Wnt7a promotes the osteogenic differentiation of human mesenchymal stem cells. *Int. J. Mol. Med.* 47, 94.
- Zhang J., Tu Q., Bonewald L.F., He X., Stein G., Lian J. and Chen J. (2011). Effects of miR-335-5p in modulating osteogenic differentiation by specifically downregulating Wnt antagonist DKK1. *J. Bone. Miner. Res.* 26, 1953-1963.