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Exploration of mycorrhizal fungi as potential biofertilizer in the management of plant biotic and abiotic stresses

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Article History	Abstract					
Received: 27Aug 2023	A deve ender and the life of (AMT) and formal formation the solution					
Revised: 28Sept 2023	Arbuscular mycorrnizal fungi (AMF) are fungi found in the soil					
Accepted: 06Oct 2023	and it can significantly enhance plant nutrient uptake and					
	increase resistance to various environmental stresses.					
	Arbuscular mycorrhizal symbiosis is the most common non- pathogenic symbiosis in the soil and is found in 80% of vascular plant roots. Most of AM fungi species belong to the sub-phylum Glomeromycotina within the phylum					
	Mucoromycota. Arbuscular mycorrhizal (AM) fungi not only					
	enhance the phosphorus supply to plants but also boost the					
	absorption of zinc, copper, nitrogen and iron. AM fungi limit					
	the uptake of Na and Cl. AM fungal hyphae make significant					
	contributions in enhancing soil structure and its ability to retain					
	water. Additionally, these fungi demonstrate resilience against					
	certain root diseases and display a tolerance to drought					
	conditions. Arbuscular mycorrhizal (AM) fungi serve as crucial					
	endosymbionts, playing a significant role in enhancing plant productivity and contributing to the overall functioning of					
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	ecosystems. Then importance is paramount in the context of					
CC License	sustainable crop enhancement.					
CC-BY-NC-SA 4.0	Keywords- Mycorrhiza, Photobionts, Disease, Biotic, Abiotic					

Introduction

The word "mycorrhiza" comes from the Greek roots "myco" for "fungus" and "rhiza" for "root" (Latef *et al.*, 2016). Most terrestrial plant have symbiotic association with fungi in their roots. This

common symbiosis known as mycorrhiza, assist as networks for the movement of nutrients and energy between plants and soils (Mohammadi et al., 2011). Mycorrhizal associations, which aid in the absorption of nutrients from soil, are formed between hyphal fungus and the underground organs of plants in this plant part termed as photobiont and fungal hyphae as mycobiont (Brundertt, 2002; Huey et al., 2020). Because mycorrhizae have an impact on plant diversity and productivity, mycorrhizal symbiosis is crucial to ecosystem health. The symbiosis besides being present in almost all plant in healthy vegetative condition is a form of "Biological fertilization" (Gianinazzi et al., 2010) which has very good result in case of biotic or abiotic stresses condition (Ganugi et al., 2019). Mycorrhizal relationships generally increase plant productivity; however, this is not always the case. Depending on the environment, symbiosis can involve a variety of species interactions, from mutualism to parasitism. Mycorrhizae are classified into two main groups i.e., ectomycorrhizae and endomycorrhizae (based on the structure of hyphae). Those mycorrhizal association in which hyphae of the fungi do not penetrates the individual cells within roots of plant is termed as ectomycorrhizae while endomycorrhizal fungi are those which invades the cell membrane by piercing the cell wall (Szabo et al., 2014). Vesicular-arbuscular mycorrhizal fungi (VAM) and soil fungi are synonymous terms used to refer to arbuscular mycorrhizal fungi (Vogelsang et al., 2004). VAM fungi found in the soil and it can significantly enhance plant nutrient uptake and increase resistance to various environmental stresses (Sun et al., 2018). Most of AM fungi species belong to the sub-phylum Glomeromycotina within the phylum Mucoromycota (Spatafora et al., 2016). Within this sub-phylum there are four distinct orders of AMF: Glomerales, Archaeosporales, Paraglomerales and Diversisporales, comprising a total of 25 genera (Redecker et al., 2013). These fungi are considered obligate biotrophs, relying on the intake of plant photosynthetic products (Bago et al., 2000) and lipids to complete their life cycle (Jiang et al., in 2017). Close to 90% of plant varieties, which includes flowering plants, bryophytes, and ferns, have the capability to establish mutually beneficial associations with AMF (Zhu et al., 2010 and Ahanger et al., 2014). VAM fungi not only increase the uptake of phosphorus supply to crop plant but also helps in the absorption of minor nutrients (Hart and Forsythe, 2012). AM fungi limit the uptake of Na and Cl. AM fungal hyphae make significant contributions in enhancing soil structure and its ability to retain water (Candido et al., 2015). Additionally, these fungi demonstrate resilience against certain root diseases and display a tolerance to drought conditions. Arbuscular mycorrhizal (AM) fungi serve as crucial endosymbionts, playing a significant role in enhancing plant productivity and contributing to the overall functioning of ecosystems. Their importance is paramount in the context of sustainable crop enhancement (Begum et al., 2019). Arbuscular mycorrhizal fungi, also referred to as "biofertilizers," are soil-borne fungi that may substantially boost plant nutrient intake and resistance to a number of abiotic stress conditions (Begum et al., 2019). At least 90% of plants are connected to fungi, and mycorrhizalfungi are significantly the most prevalent of them. A fungus and a green plant have a symbiotic association known as mycorrhiza. The plant creates organic compounds like sugars through photosynthesis and gives them to the fungus, which then gives the plant water and mineral nutrients like phosphorus that it absorbs from the soil (Milton et al., 2021).

Mycorrhiza's significance in nutrient uptake and mobilization

Plants can absorb nutrients from the soil in two different ways (Smith et al., 2011). One is a plantpathway, where no fungus is present and nutrients are directly absorbed by plant roots, and the other is a mycorrhizal channel, where nutrients are taken up by the extraradical mycelium of the associated fungus (Harrison et al., 2002). The mycorrhizal route is employed when the mycorrhizal fungus colonizes the roots of the plant. About 15 essential macro- and micronutrients necessary for the growth of the plant can be absorbed and transported by mycorrhizal fungus (Fig. 1). AMFs thrive in temperate and arid settings where P is frequently a limiting factor because they are exceptionally good at mobilizing inorganic phosphorus (P) (Teotia et al., 2017). By transporting nutrients (especially P) from the soil to the crop through their external mycelium, AMFs can increase nutrient uptake (Sun et al., 2022). The plant and fungus species have an impact on how much P is ultimately absorbed through a common pathway. Numerous mycorrhizal fungi perform a crucial function in mobilizing mineral nutrients from unavailable organic substrates, mineral particles, and rock surfaces, in addition to improving the absorption of mineral nutrients by plants in the soil. Along with mobilizing nutrients, mycorrhizal fungi also serve as an essential C sink in the soil, contributing significantly to the cycling of such mineral elements (Teotia et al., 2017). In high latitude and longitude or in other rocky environment mycorrhizal fungi helps to obtain the nutrients from primary rock surfaces (Milton et al., 2021). A study done by (Sun et al., 2022) in maize reflects that the application of AMFs helps to increase the shoot P, K, Ca, Mg, Mn and Zn contents.

Additionally, by increasing the availability of phosphorus to the host plant, AMFs have an impact on soil phosphatase enzymatic activity and soil physicochemical parameters. The versatile function of mycorrhiza depicted in Figure 1. Mycorrhizal maize roots have considerably greater alkaline phosphatase activity than non-mycorrhizal maize roots (Sun *et al.*, 2022). Not only AMFs help in the uptake of macronutrients, but they also help in the absorption of micronutrients like Fe, Cu, Zn, etc (Teotia *et al.*, 2017). AMF expands its absorption network past the rhizosphere's nutritional depletion zones, enabling access to a broader area of soil (Smith *et al.*2011).



Fig. 1: Versatile functions of mycorrhiza in crop plants

Mycorrhiza's significance to biotic stress resistance

AMF contributes to disease resistance by competing for colonization sites and enhancing the plant's defensive mechanism. AMF colonization has a protective effect known as mycorrhizainduced resistance (MIR), which offers systemic defense against a variety of attackers and shares traits with induced systemic resistance (ISR) after root colonization by non-pathogenic rhizobacteria and systemic acquired resistance (SAR) after infection by pathogens. AMF causes plants to produce more antioxidant enzymes, which can help them fight off infections and other stresses (Diagne et al., 2020). Among the diseases that can be resisted by using AMFs are those that affect plants, such as the migratory nematode Pratylenchus penetrans and the sedentary nematode Meloidogyne incognita (Diagne et al., 2020). The Significance of mycorrhizal fungi of different species in Agriculture tabulated in Table 1. Another example is the charcoal root-rot disease in soybeans, which can be resisted by using AMF inoculation (Spagnoletti et al., 2020). Mycorrhizae can aid in the rebuilding of tissues following attacks, therefore the improvement in plant development can be beneficial. Compared to the application of water or non-mycorrhizal root exudates, the use of mycorrhizal root exudates further decreased nematode invasion in mycorrhizal plants and temporarily paralyzed nematodes. There aren't many studies on how AMF affects herbivorous insects. Some AMFs inhibit an insect's ability to chew. For instance, mycorrhizal infection improved the resistance of leaves to the chewing insect Arctia caja in Plantago lanceolate L (Diagne et al., 2020).

Mycorrhiza's significance in drought tolerance

One of the main factors that can significantly lower plant productivity is drought (Posta *et al.*, 2020). AMF has been shown to enhance plant performance under drought stress (Balestrini *et al.*, 2018). By reducing drought and tolerating it, mycorrhizal plants cope with water shortages (Bernardo *et al.*, 2019). Drought moderation is refereed by unintended AMF advantages and increased liquid intake, whereas drought tolerance is mediated by direct AMF advantages that improve the plant's natural ability to cope with stress (Posta *et al.*, 2020). Underacute conditions of drought, the symbiotic association of diverse plants with mycorrhizal fungi can increase the root size and efficiency, leaf area index and biomass (Begum *et al.*, 2019). It's possible that better access to tiny soil pores, increased surface area for water absorption given by AMF hyphae, or improved apoplastic water flow are the causes of the improvement in plant fitness brought on by AMF (Diagne *et al.*, 2020). Abscisic acid is regulated by the administration of AMF, which affects how stomata function. Stomatal conductance and other associated physiological processes are controlled by ABA reactions (Diagne *et al.*, 2020). Stomatal closure is induced by ABA, which also lowers cell water loss (Ouledali *et al.*, 2019).

Mycorrhiza's significance in heavy metal bioremediation

Multiple studies revealed that AMF had spread to more than 80% of the plants growing onmining sites (Wang *et al.*, 2017). They have good capacity to improve the defense system in crop plants and encourage plant growth and development, AMFs are widely thought to sustenance plant formation in soil with heavy metal contamination. AMF can remove heavy metals by "metal-binding" hyphae, which lowers the bioavailability of elements including Cu, Pb, Co, Cd and Zn (Diagne *et al.*, 2020).

Glomalin is a glycoprotein that AMF hyphae can secrete (Herath *et al.*, 2021). It improves growth, yield, and nutrient status by binding heavy metals in the mantle hyphae and corticalcells' cell walls, preventing their uptake (Begum *et al.*, 2019).

Mycorrhiza's significance in salinity tolerance

Soil salinization is a well-known environmental issue that poses a serious danger to the world'sfood security (Begum *et al.*, 2019). Saline areas have been known to naturally contain AMF (Beltrano *et al.*, 2013). In plants exposed to salinity, AMF colonization enhances stomatal conductance and lessens oxidative damage (Pedranzani *et al.*, 2015). In plants grown under saline stress, inoculation of AMF was also seen to promote the accumulation of different organic acids, leading to an up-regulation of the osmoregulation process (Begum *et al.*, 2019). For instance, *F. mosseae* inoculation of tomato plants with salty water irrigation enhanced plant biomass, fruit fresh output, and the amount of P, K Cu, Fe and Zn in the shoots. In a different study, the same AMF colonized plant roots, lowering Na levels while increasing the activity of numerous enzymes related to reducing salt stress (Diagne *et al.*, 2020)

Mycorrhiza's significance in coping with extreme temperatures

One of the most significant environmental factors that might impair plant development and productivity is temperature (Zhu *et al.*, 2011). Plants with AMF inoculation typically grow better under heat stress than those without it (Begum *et al.*, 2019). It is widely accepted that AMF enhances plant performance to withstand temperature (heat or cold) stress by improving food and water intake, photosynthetic capacity and efficiency, shielding plants from oxidative damage, and increasing osmolyte accumulation (Diagne *et al.*, 2020). AMF can also help the host plant retain moisture, produce more secondary metabolites, which strengthens the plant's immune system, and produce more protein, which benefits the plant resist to low temperature stress (Begum *et al.*, 2019). Overall mycorrhizal fungi help crop plants in root elongation, water absorption at higher temperature (Mathur *et al.*, 2020).

AMF Species	Host plant	Importance	Response of plant after AMFs	References	
			inoculation		
			High Alkaline phosphatase, enlargement		
Diversispora	Zea mays L.	Nutrient uptake	of root	Sun et al. (2022)	
eburnea			surface, increase in photosynthetic		
			pigment, increase in intake of Phosphorus		
			and magnesium		
Gigaspora margarita	ita Lotus japonicus L. Nutrient uptake Phosphorus concentrations in plant shoots		Zhang et al. (2015)		
			and roots were significantly		
			increased		
Glomus versiforme,			Decreased Cd concentrations in shoots		
Rhizophagus	Lonicera japonica	Heavy Metals tolerance	and roots, reduced Cd concentrations in	Jiang et al. (2016)	
intraradices	Thunb.		shoots but increased Cd concentrations in		
roots		roots			
Glomus	Trigonella		Increased antioxidant enzymes		
monosporum, G.	foenum-	Heavy Metals tolerance	activities and malondialdehyde	Abdelhameed and	
clarum, Gigaspora	graecum L.	content.		Rabab (2019)	
nigra, and					
Acaulospora laevis					
Rhizophagus			Increased leaf length, plant height, leaf		
intraradices,	Zea mays	Extreme temperature	e temperature number, chlorophyll a, photosynthetic		
Funneliformis		tolerance	rate, stomatal		
mosseae, F.			conductance, and transpiration rate		
geosporum					
G. versiforme, R.	Hordeum vulgare L	Extreme temperature	Increasing the survival rate,	Hajiboland <i>et al</i> .	
<i>irregularis</i> tolerance		tolerance	alleviation of low-	(2019)	
			temperature stress		
Rhizophagus	S. lycopersicum		Improved shoot dry weight, stomatal		
irregularis	Lactucasativa	Drought stress tolerance	conductance, photosystem II efficiency,	Ruiz-Lozano et al.	
	Linn		ABA and strigolactone contents	(2015)	
			Increased osmotic potential, chlorophyll		
Glomus mosseae	Triticum aestivum	Drought stress tolerance	content and fluorescence, activities of	Rani (2016)	
			antioxidant enzymes,		
			ascorbic acid, enzymes of N and P		

Table 1. Significance of mycorrhizal fungi of different species in Agriculture

			(1.1' 1	
			metabolism, and	
			contents of N, P, and K	
			Production of antimicrobial compounds	
			from the mycorrhizal root that arrested	
Glomus sp.	L. esculentum	Biotic stress tolerance	the mycelial growth of the fungal	Kumari et al. (2019)
			pathogen (Fusarium oxysporum	
			f. sp. lycopersici), reduced the disease	
			incidence, increased the plant growth,	
			dry weight, N, P, K	
			content, chlorophyll content and yield of	
			the plant	
			Higher resistance against	
F. mosseae	Solanum	Biotic stress tolerance	<i>Cladosporium fulvum</i> infection,	Wang <i>et al.</i> (2017)
	lycopersicum L.		higher fresh and dry weight,	
			increases in total chlorophyll	
			contents and netphotosynthesis rate	
			Increased stomatal conductance,	
R. irregularis	Digitaria eriantha	Salinity stress tolerance	antioxidant enzymesactivities (CAT et	Pedranzani et al.
	Steud.		APX), jasmonate content, and reduced	(2015)
			root and shoot hydrogen peroxide	
			accumulation	
Glomus etunicatum.				
Glomus	Cucumis sativus L	Salinity stress tolerance	Increased biomass, photosynthetic	Hashem <i>et al.</i> (2018)
intraradices Glomus	Chelinits Suirrus Er	Summy substituted	nigment synthesis and enhanced	2010)
mossoga			antiovidant anzumas	
mosseae			antioxidant enzymes	

Impact of arbuscular mycorrhizas in plant nutrient and growth

In over 90% of plants, according to van der Heijden et al. (2015), Mycorrhizal fungi frequently escape unnoticed in biodiversity surveys, despite their significance for plant life histories (Soudzilovskaia *et al.*, 2020). This is due to the fact that most mycorrhizal fungi frequently fail to form fruiting bodies, making them difficult to spot in the wild (Egli 2011; Büntgen *et al.*, 2013). AMF are fungi that live in the soil and have been shown to dramatically increase plant nutrient uptake and resilience to a variety of abiotic stressors (Sun *et al.*2018). The sub-phylum Glomeromycotina of the phylum Mucoromycota is where the bulk of AMF species are found (Spatafora *et al.*, 2016). This sub-phylum contains 25 species and four orders of AMF, namely Glomerales, Archaeosporales, Paraglomerales and Diversisporales (Redecker *et al.*, 2013). These fungi can develop and aid plants in absorbing nutrients and water. Plants feed fungi by sending sugars from their leaves. Mycorrhizae can also enhance root surface area, which enables plants to absorb water and nutrients more effectively from a large soil volume (Nadeem *et al.*, 2014). Numerous studies have demonstrated that, in contrast to their non-mycorrhizal counterparts, mycorrhizal plants typically modified their drought physiology later during soil drying or in somewhat drier soils (Augé 2001; Augé *et al.*, 2015). The comparison between colonization of AM fungi and without AM fungi illustrated in Figure 2.



Fig. 2: Comparison between colonization of AM fungi and without AM fungi

In an effort to identify the underlying mechanisms, studies have revealed a complex reorganization of the mycorrhizal plant's response to water stress, including sustained stomatal opening (Augé et al., 2015), higher plant water potentials (Abdalla and Ahmed 2021; Porcel and Ruiz-Lozano 2004), differential expression and activation of root aquaporins (Sharma et al., 2021), altered osmolytes. Leaching, mineralization, and nitrification are only a few of the chemical reactions in soils that release H+ ions and regulate soil pH (Neina, 2019). It is challenging to identify the precise mechanism by which the biological community and biological activities of the soil are controlled by pH (Neina, 2019). Changes in the physical-chemical characteristics of the soil have an impact on these fungi's abundance, richness and community composition.

According to Bowles *et al.* (2016), the formation of a hyphal network by plant roots and the AMF considerably improves roots' access to a vast soil surface area and improves plant growth. By enhancing the availability and transport of different nutrients, AMF enhance plant nutrition (Rouphael et al., 2015). AMF enhance soil quality by affecting the texture and structure of the soil, which benefits plant health (Zou et al., 2016; Thirkell et al., 2017). According to Paterson et al. (2016), fungal hyphae can hasten the breakdown of soil organic materials. The present review focuses on the role of AMF as bio-fertilizers in the regulation of plant growth and development with improved nutrient uptake under stressful environments, as well as the extent to which AMF can enhance plant growth under stressful environments. This is due to the significance of AMF and the research advancements related to their applications in agriculture.

According to Brundrett's definition from 2002, mycorrhizae's modified absorptive organs, which mostly consist of plant roots (photobiont) and fungus hyphae (mycobiont), develop mutualistic interactions with one another. Nutrient transfer between the organisms is the primary goal of this connection (Brundrett, 2002). AMF and plants were known to coexist 400 million years ago (Selosse et

al., 2015). Such connections are made through a series of biological processes that have a range of beneficial consequences on both agricultural and natural biotas (Van der Heijden *et al.*, 2015).

The role of mycorrhizal fungi in plant disease management

In present days plant pathogens are a major cause for increasing losses in agriculture. There are four main methods to control these pathogens, those are traditional methods, physical methods, chemical methods, biological methods. Many synthetic chemicals are used in chemical method as an instant way to control these pathogens, whereas these chemical compounds are a threat to sustainability of natural resources. Biological method is a method in which pathogens can be controlled with least damage to the environment. In past few years mycorrhizae has attained interest of many researchers due to its positive advances in terms of protection against plant diseases. Mycorrhizae is a symbiotic association between plant roots and fungi. It absorbs nutrients and water for plants and in turn plants provide shelter and photosynthetic supplements to it. Mycorrhiza belongs to phylum: Glomeromycota and genus: Glomus. Mycorrhizal fungi are of three types, they are endomycorrhiza, ectomycorrhiza and endo-ectomycorrhiza. Arbuscular mycorrhizae are endomycorrhizal fungi which are most commonly found in the environment.

Controlling plant diseases is very important in present day agriculture and food storage but currently used methods are eradicating both disease-causing pathogens along with non-target organisms Brimner *et al.* (2003). The efficiency of mycorrhizal fungi in plant disease management tabulated in Table 2.

Pathogen	Impact	Species	Example	Reference
1. Fungi	Positive	Glomus	Soyabean	Zamboline <i>et</i>
		mossae	infected by	al. (1983).
			Rhizoctonia	
			solani and M.	
			phaseolina	
2. Bacteria	Positive	G. mosseae or	Mulberry	Sharma <i>et al</i> .
		G.	infected by P.	(1995).
		fasciculatum	<i>syringae</i> pv.	
			Mori	
			Causing	
			bacterial blight	
3. Nematodes	Positive	G. mosy	Nematode	Liu et al.
			infection in	(2012).
			tobacco by	
			Meloidogyne	
			incognita and	
			T. basicola	

Table 2. Mycorrhizal fungi in disease management

4. Viruses	Negative	G.	Yellow mosaic	Jayaram et al.
		fasciculatum	Bigeminy virus	(1995).
	Cause: increase		in mung bean	
	in virus			
	multiplications			
	due to enhanced			
	phosphorous			
	levels			

Signalling pathways between fungi and plant resulted in identification of nutrient transporters that revealed cellular processes portraying symbiosis Bonfante *et al.* (2010).

Increase in absorption of phosphorous by plant is one of the earliest proposed mechanisms of AMF-mediated pathogen or disease tolerance that is still applicable Xavier *et al.* (2004). As a result of mycorrhizal association there is a change in root exudate composition followed by change in permeability of root membrane of the plant Graham *et al.* (1981) resulting in alternation of rhizosphere microbial equilibrium Brejda *et al.* (1998). Mycorrhizal plants can tolerate plant pathogens and reduce root damage pathogens as mycorrhizae can increase plant nutrition and health Declerck *et al.* (2002). Arbuscular mycorrhizal fungi can reduce the damage caused by bacteria, fungi, nematodes and other pathogens of cucumber, tomato, olive, strawberry, mandarin orange, melon, soybean, maize, potato, banana, and other plants Weng *et al.* (2022). For example, in fungal infestation soybean plants without mycorrhizal association infected with *Rhizoctonia solani* and *M. phaseolina* has shown lower shoot and root weigh along with less plant height whereas plants with mycorrhizal association were able to tolerate infestation of pathogens Zamboline *et al.* (1983). The mechanism of action of mycorrhizal fungi depicted in Figure 3.

The mechanisms exhibited by AMF to protect host from pathogens are competition for colonization sites, enhanced nutrient absorption, damage compensation of plant, morphological and anatomical changes in roots and change in microbial composition in rhizosphere, yet in viral infections mycorrhizae associated plants are more susceptible to viruses than non-mycorrhizal plants as enhanced phosphorous levels.



Fig.3: Various mode of action of mycorrhizal fungi

In mulberry, inoculation of mycorrhizae along with 60 -90 kg of phosphorous per hectare reduced the infestation of bacterial blight Sharma *et al.* (1995). In tobacco, Pre-inoculation of plants with *G. mosy* can reduce the number of *Meloidogyne incognita* and *T. basicola* propagules and resulting in increase of plant resistance towards pathogenic nematodes Liu *et al.* (2012). The usage of AMF along with other beneficial microorganisms at once, has reduced the occurrence of plant diseases and damage. Namely, *Trichoderma* used along with AMF in different combinations has given different results in plant disease control Martinez *et al.* (2011).

Conclusion

With the introduction of mycorrhizal fungi, they showed as a valuable substitute towards the indiscriminate use of chemicals either it is fertilizer or fungicides. Mycorrhizal fungi also help in enhancing photosynthetic rate and other gas exchange-related traits, as well as increased water uptake Mycorrhiza play a versatile role in up taking the essential nutrients (macro & micro) from the soil *viz.*, nitrogen, phosphorus and many more. They also provide protection from biotic and abiotic factors. AMF sustainably improves plant growth, productivity and also helps to reduce the diseases causes by plant pathogenic agents. So use of mycorrhizal fungi in crop plants could be more beneficial in comparison to chemicals because pesticides harming the colonization of microbiome, environment and human health's.

Conflict of Interest

The authors declare they have no conflict of interest.

References

- 1. Abdelhameed, R. E., and Rabab, A. M. (2019). Alleviation of cadmium stress by arbuscular mycorrhizal symbiosis. Int. J. Phytoremed. doi: 10.1080/15226514.2018.1556584
- 2. Ahanger, M.A., Tyagi, S.R., Wani, M.R., Ahmad, P. (2014). "Drought tolerance: role of organic osmolytes, growth regulators, and mineral nutrients," in Physiological mechanisms and

adaptation strategies in plants under changing environment, vol. 1. Eds. Ahmad, P., Wani, MR (New York, NY: Springer), 25–55.

- 3. Augé, R.M. (2001). Water relations, drought and vesicular-arbuscular mycorrhizal symbiosis. Mycorrhiza, 11(1):3–42
- 4. Augé, R.M., Toler, H.D., Saxton, A.M. (2015). Arbuscular mycorrhizal symbiosis alters stomatal conductance of host plants more under drought than under amply watered conditions: a meta-analysis. Mycorrhiza 25(1):13–24
- 5. Bago, B., Pfeffer, P.E., and Shachar-Hill, Y. (2000). Carbon metabolism and transport in arbuscular mycorrhizas. Plant Physiol. 124, 949–958.
- Balestrini, R., Lumini, E. (2018). Focus on mycorrhizal symbioses. Appl. Soil Ecol., 123, 299– 304.
- 7. Battini, F., Grønlund, M., Agnolucci, M., Giovannetti, M., Jakobsen, I. (2017). Facilitation of phosphorus uptake in maize plants by mycorrhizosphere bacteria. Sci. Rep., 7, 4686.
- Begum, N., Qin, C., Ahanger, M.A., Raza, S., Khan, M.I., Ashraf, M., Ahmed, N. and Zhang, L. (2019). Role of arbuscular mycorrhizal fungi in plant growth regulation: implications in abiotic stress tolerance. Frontiers in plant science, 10, 1068.
- 9. Beltrano, J., Ruscitti, M., Arango, M., Ronco, M. (2013). Effects of arbuscular mycorrhiza inoculation on plant growth, biological and physiological parameters and mineral nutrition in pepper grown under different salinity and p levels. J. Soil Sci. Plant. Nutr., 13, 123–141.
- Bernardo, L., Carletti, P., Badeck, F., Rizza, F., Morcia, C., Ghizzoni, R., Rouphael, Y., Colla, G., Terzi, V., Lucini, L. (2019). Metabolomic responses triggered by arbuscular mycorrhiza enhance tolerance to water stress in wheat cultivars. Plant. Physiol. Biochem., 137, 203–212.
- 11. Birhane, E., Sterck, F., Fetene, M., Bongers, F., Kuyper, T. (2012). Arbuscular mycorrhizal fungi enhance photosynthesis, water use efficiency, and growth of frankincense seedlings under pulsed water availability conditions. Oecologia, 169, 895–904.
- 12. Bonfante, P., Genre, A. (2010). Mechanisms underlying beneficial plant-fungus interactions in mycorrhizal symbiosis. Nature Communications, 1:48
- 13. Bowles, T.M., Barrios-Masias, F.H., Carlisle, E.A., Cavagnaro, T.R., and Jackson, L.E. (2016). Effects of arbuscular mycorrhizae on tomato yield, nutrient uptake, water relations, and soil carbon dynamics under deficit irrigation in field conditions. Sci.Total Environ, 566, 1223–1234.
- 14. Bowles, T.M., Barrios-Masias, F.H., Carlisle, E.A., Cavagnaro, T.R., Jackson, L.E. (2016). Effects of arbuscular mycorrhizae on tomato yield, nutrient uptake, water relations, and soil carbon dynamics under deficit irrigation in field conditions. Sci. Total Environ. 566, 1223–1234.
- 15. Brejda, J.J., Moser, L.E., Vogel, K.P. (1998) Evaluation of switchgrass rhizosphere microflora for enhancing seedling yield and nutrient uptake. Agron Journal, 90:753-758
- 16. Brimner, T.A., & Boland, G.J. (2003). A review of the non-target effects of fungi used to biologically control plant diseases. Agriculture, ecosystems & environment, 100(1), 3-16.
- 17. Brundrett, M.C. (2002) Coevolution of roots and mycorrhizas of land plants. New Phytol, 154:275–304.

- 18. Büntgen, U., Peter, M., Kauserud, H., Egli, S. (2013) Unraveling environmental drivers of a recent increase in Swiss fungi fruiting. Global Change Biol, 19(9):2785–2794.
- 19. Calevo, J., & Duffy, K.J. (2023). Interactions among mycorrhizal fungi enhance the early development of a Mediterranean orchid. Mycorrhiza, 33(4), 229-240.
- Candido, V., Candido, Campanelli, G., D'Addabbo, T., Castronuovo. D., Perniola, M., Camele, I. (2015) Growth and yield promoting effect of artificial mycorrhization on field tomato at different irrigation regimes Sci Hortic, 187:35-43
- 21. Cao, J., Wang, C., Huang, Y. (2015). Interactive impacts of earthworms (Eisenia fetida) and arbuscular mycorrhizal fungi (Funneliformis mosseae) on the bioavailability of calcium phosphates. Plant Soil, 396, 45–57
- Caradonia, F., Francia, E., Morcia, C., Ghizzoni, R., Moulin, L., Terzi, V., Ronga, D. (2019). Arbuscular Mycorrhizal Fungi and Plant Growth Promoting Rhizobacteria Avoid Processing Tomato Leaf Damage during Chilling Stress. Agronomy, 9, 299.
- 23. Deckmyn, G., Meyer, A., Smits, M. M., Ekblad, A., Grebenc, T., Komarov, A., & Kraigher, H. (2014). Simulating ectomycorrhizal fungi and their role in carbon and nitrogen cycling in forest ecosystems. Canadian Journal of Forest Research, 44(6), 535–553.
- 24. Declerck, S., Risede, JM., Ruflikiri, G., and Delvaux, B. (2002). Effects of arbuscular mycorrhizal fungi on severity of root rot of banana caused by Cylindrocladium spathiphylli. Plant Pathol 51,109–115.
- 25. Diagne, N., Ngom, M., Djighaly, P. I., Fall, D., Hocher, V., & Svistoonoff, S. (2020). Roles of arbuscular mycorrhizal fungi on plant growth and performance: Importance inbiotic and abiotic stressed regulation. *Diversity*, *12*(10), 370.
- 26. Egli, S. (2011). Mycorrhizal mushroom diversity and productivity—an indicator of forest health. Ann for Sci 68(1),81–88.
- 27. Frew, A., Powell, J.R., Glauser, G., Bennett, A.E., Johnson, S.N (2018). Mycorrhizal fungi enhance nutrient uptake but disarm defences in plant roots, promoting plant-parasitic nematode populations. Soil Biol. Biochem. 2018, 126, 123–132.
- 28. Ganugi, P., Masoni, A., Pietramellara, G., Benedettelli, S. (2019). A Review of Studies from the Last Twenty Years on Plant–Arbuscular Mycorrhizal Fungi Associations and Their Uses for Wheat Crops. Agronomy, 9(12), 840.
- 29. George, E., Marschner, H., & Jakobsen, I. (1995). Role of arbuscular mycorrhizal fungiin uptake of phosphorus and nitrogen from soil. Critical Reviews in Biotechnology, 15(3-4), 257-270.
- Gianinazzi, S., Gollotte, A., Binet, M.N., van Tuinen, D., Redecker, D., Wipf, D. (2010). Agroecology: the key role of arbuscular mycorrhizas in ecosystem services. Mycorrhiza, 20(8), 519-30.
- 31. Graham, J.H., Leonard, R.T., Menge, J.A. (1981). Membrane mediated decrease in root exudation responsible for inhibition of vesicular-arbuscular mycorrhiza formation. Plant Physiol, 68, 548-552

- Hajiboland, R., Joudmand, A., Aliasgharzad, N., Tolrá, R., Poschenrieder, C. (2019). Arbuscular mycorrhizal fungi alleviate low-temperature stress and increase freezing resistance as a substitute for acclimation treatment in barley. Crop. Pasture Sci., 70, 218–233.
- Harrison, M.J., Dewbre, G.R., Liu, J. (2002). A phosphate transporter from Medicago truncatula involved in the acquisition of phosphate released by arbuscularmycorrhizal fungi. Plant Cell 14, 2413–2429
- 34. Hart, M.M., Forsythe, J.A. (2012). Scientia Horticulture Using arbuscular mycorrhizal fungi to improve the nutrient quality of crops; nutritional benefits in addition to phosphorus. *Sci Hortic* (*Amsterdam*) 148, 206–214.
- 35. Hashem, A., Alqarawi, A. A., Radhakrishnan, R., Al-Arjani, A. F., Aldehaish, H. A., Egamberdieva, D., et al. (2018). Arbuscular mycorrhizal fungi regulate the oxidative system, hormones and ionic equilibrium to trigger salt stress tolerance in Cucumis sativus L. Saudi J. Biol. Sci. 25 (6), 1102–1114.
- 36. Herath, B. M. M. D., Madushan, K. W. A., Lakmali, J. P. D., & Yapa, P. N. (2021).
- Jiang, Q.-Y., Zhuo, F., Long, S.-H., Zhao, H.-D., Yang, D.-J., Ye, Z.-H., Li, S.-S., Jing, Y.-X. (2-16). Can arbuscular mycorrhizal fungi reduce Cd uptake and alleviate Cd toxicity of Lonicera japonica grown in Cd-added soils? Sci. Rep., 6, 21805.
- Jiang, Y. N., Wang, W. X., Xie, Q. J., Liu, N., Liu, L. X., Wang, D. P. (2017). Plants transfer lipids to sustain colonization by mutualistic mycorrhizal and parasitic fungi. Science 356, 1172– 1175.
- 39. Khan, Y., Yang, X., Zhang, X., Yaseen, T., Shi, L., Zhang, T (2021). Arbuscular mycorrhizal fungi promote plant growth of Leymus chinensis (Trin.) Tzvelev by increasing the metabolomics activity under nitrogen addition. Grassl Sci, 67, 128–138.
- 40. Kumari, S.M.P & Prabina, B.J. Protection of Tomato, Lycopersicon esculentum from Wilt Pathogen, Fusarium oxysporum f.sp. lycopersici by Arbuscular Mycorrhizal Fungi, Glomus sp. Int. J. Curr. Microbiol. Appl. Sci. 2019, 8, 1368–1378.
- 41. Latef, A.A.H.A., Hashem, A., Rasool, S. et al. Arbuscular mycorrhizal symbiosis and abiotic stress in plants. A review. J. Plant Biol, 59, 407–426 (2016).
- 42. Liu, R.J., Dai, M., Wu, X., Li, M., Liu, X.Z.(2012). Suppression of the root-knot nematode Meloidogyne incognita (Kofoid & White) Chitwood] on tomato by dual inoculation with arbuscular mycorrhizal fungi and plant growth-promoting rhizobacteria. Mycorrhiza, 22, 289–296.
- 43. Mansfield, T. M., Albornoz, F. E., Ryan, M. H., Bending, G. D., & Standish, R. J. (2023). Niche differentiation of Mucoromycotinian and Glomeromycotinian arbuscular mycorrhizal fungi along a 2-million-year soil chronosequence. Mycorrhiza, 1-14.
- 44. Martínez-Medina, A., Roldán, A., Pascual, J.A. (2011). Interaction between arbuscular mycorrhizal fungi and Trichoderma harzianum under conventional and low input fertilization field condition in melon crops: Growth response and Fusarium wilt biocontrol. Appl. Soil Ecol, 47, 98–105
- 45. Mathur, S., Jajoo, A. (2020). Arbuscular mycorrhizal fungi protects maize plants from

high temperature stress by regulating photosystem II heterogeneity. Ind Crop Prod, 143, 111934.

- 46. Mathur, S., Sharma, M. P., and Jajoo, A. (2016). Improved photosynthetic efficacy of maize Zea mays plants with arbuscular mycorrhizal fungi (AMF) under high temperature stress. J. Photochem. Photobiol. B 180, 149–154.
- 47. Mei, L., Yang, X., Cao, H., Zhang, T., Guo, J. (2019). Arbuscular mycorrhizal fungi alter plant and soil C: N: P stoichiometries under warming and nitrogen input in a semiarid meadow of China. Int. J. Environ. Res. Public Health, 16, 397
- 48. Merckx, V.S.F.T. (2012). Mycoheterotrophy: the biology of plants living on fungi. *Mycoheterotrophy Biol Plants Living Fungi*.
- 49. Milton, Mikhil & Kumar, Sarvesh & Singh, A & Kumar, Vinai & Bisarya, Dipti. (2021). Mycorrhizae and their importance in agriculture. JETIR 8, 201-206.
- 50. Miransari, M. (2013). Soil microbes and the availability of soil nutrients. Acta Physiol. Plant, 35, 3075–3084.
- Mohammadi, Khosro & Khalesro, Shiva & Sohrabi, Yousef & Heidari, Gholamreza. (2011). A Review: Beneficial Effects of the Mycorrhizal Fungi for Plant Growth. J. Appl. Environ. Biol. Sci. 1, 310-319.
- 52. Mustafa, G., Randoux, B., Tisserant, B., Fontaine, J., Magnin-Robert, M., Sahraoui, A.L.H., Reignault, P. (2016). Phosphorus supply, arbuscular mycorrhizal fungal species, and plant genotype impact on the protective efficacy of mycorrhizal inoculation against wheat powdery mildew. Mycorrhiza, 26, 685–697.
- 53. Nadeem, S.M., Ahmad, M., Zahir, Z.A., Javaid, A., Ashraf, M. (2014). The role of mycorrhizae and plant growth promoting rhizobacteria (PGPR) in improving crop productivity under stressful environments. Biotechnol Adv, 32,:429–448.
- 54. Neina, D. (2019). The role of soil pH in plant nutrition and soil remediation. Appl Environ Soil Sci, 5794869,:1–9.
- 55. Nuria, F. (2019). Arbuscular mycorrhizas as key players in sustainable plant phosphorus acquisition: An overview on the mechanisms involved Plant science, 280, Pages 441-447.
- 56. Ouledali, S.; Ennajeh, M.; Ferrandino, A.; Khemira, H.; Schubert, A.; Secchi, F. (2019). Influence of arbuscular mycorrhizal fungi inoculation on the control of stomata functioning by abscisic acid (ABA) in drought-stresse do live plants, 121, 152–158.
- 57. Paterson, E., Sim, A., Davidson, J., and Daniell, T. J. (2016). Arbuscular mycorrhizal hyphae promote priming of native soil organic matter mineralization. Plant Soil, 408, 243–C254.
- 58. Pauwels, R., Graefe, J., Bitterlich, M. (2023). An arbuscular mycorrhizal fungus alters soil water retention and hydraulic conductivity in a soil texture specific way. Mycorrhiza, 33(3), 165-179.
- Pedranzani, H., Rodríguez-Rivera, M., Gutierrez, M., Porcel, R., Hause, B., Ruiz-Lozano, J.M. (2015). Arbuscular mycorrhizal symbiosis regulates physiology and performance of Digitariaeriantha plants subjected to abiotic stresses by modulatingantioxidant and jasmonate levels. Mycorrhiza, 26, 141–152.

- 60. Pedranzani, H., Rodríguez-Rivera, M., Gutierrez, M., Porcel, R., Hause, B. & Ruiz-Lozano, J.M. (2015). Arbuscular mycorrhizal symbiosis regulates physiology and performance of Digitaria eriantha plants subjected to abiotic stresses by modulating antioxidant and jasmonate levels. Mycorrhiza, 26, 141–152.
- 61. Posta, K. Duc, N. (2019). Benefits of Arbuscular Mycorrhizal Fungi Application to Crop
Production under Water Scarcity.Fungi Application to Crop
10.5772/intechopen.86595.

- 62. Rani, B. (2016). Effect of arbuscular mycorrhiza fungi on biochemical parameters in wheat Triticum aestivum L. under drought conditions. Doctoral dissertation, CCSHAU, Hisar.
- Redecker, D., Schüssler, A., Stockinger, H., Stürmer, S.L., Morton, J. B., Walker, C. (2013). An evidence-based consensus for the classification of arbuscular mycorrhizal fungi (Glomeromycota). Mycorrhiza 23 (7), 515–531.
- Redecker, D., Schüssler, A., Stockinger, H., Stürmer, S. L., Morton, J. B., and Walker, C. (2013). An evidence-based consensus for the classification of arbuscular mycorrhizal fungi (Glomeromycota). Mycorrhiza 23 (7), 515–531.
- 65. Rodriguez, R.J., Henson, J., Van Volkenburgh E., Hoy, M., Wright, L., Beckwith, F., Kim, Y.O., Redman, R.S., (2008). Stress tolerance in plants via habitat-adapted symbiosis. ISME J., 2(4):404-16.
- Rouphael, Y., Franken, P., Schneider, C., Schwarz, D., Giovannetti, M., and Agnolucci, M. (2015). Arbuscular mycorrhizal fungi act as bio-stimulants in horticultural crops. Sci. Hort, 196, 91–108.
- 67. Ruiz-Lozano, J.M., Aroca, R., Zamarreño, Á.M., Molina, S., Andreo-Jimenez, B., Porcel, R., García-Mina, J.M., Ruyter-Spira, C., López-Ráez, J.A. (2015). Arbuscular mycorrhizal symbiosis induces strigolactone biosynthesis under drought and improves drought tolerance in lettuce and tomato. PlantCell Environ. 39, 441–452.
- Salam, E. A., Alatar, A., El-Sheikh, M. A. (2017). Inoculation with arbuscular mycorrhizal fungi alleviates harmful effects of drought stress on damask rose. Saudi J. Biol. Sci, 25 (8), 1772–1780.
- Santander, C., Sanhueza, M., Olave, J., Borie, F., Valentine, C., Cornejo, P. (2019). Arbuscular mycorrhizal colonization promotes the tolerance to salt stress in lettuce plantsthrough an efficient modification of ionic balance. J. Soil Sci. Plant Nutr, 19 (2), 321–331.
- Satar, D., Hemmatinezhad, B. (2015). Review of Application and Importance of Ectomycorrhiza Fungi and their Role in the Stability of Ecosystems. Biosciences Biotechnology Research Asia. 12. 153-158.
- 71. Selosse, M. A., Strullu-Derrien, C., Martin, F. M., Kamoun, S., Kenrick, P. (2015). Plants, fungi and oomycetes: a 400-million years affair that shapes the biosphere. New Phytol. 206, 501–506.
- 72. Sharma, S., Dohroo , N.P., Sharma, S. (1997). Management of ginger yellows through organic amendment, fungicide seed treatment and biological methods. Ind Cocoa Arecanut Spice J, 21,:29-30
- 73. Smith, S.E., Read, D.J. (2008). Mineral nutrition, toxic element accumulation and water relations of arbuscular mycorrhizal plants. In Mycorrhizal Symbiosis; Academic Press: Cambridge, MA, USA, pp, 145–148
- 74. Smith, S.E., Smith, F.A. (2011). Roles of arbuscular mycorrhizas in plant nutrition and growth: New paradigms from cellular to ecosystem scales. Biol. Annu. Rev. Plant, 62,:

227-250

- 75. Soudzilovskaia, N.A., Vaessen, S., Barcelo, M., He, J., Rahimlou, S., Abarenkov, K., Brundrett, M.C., Gomes, S.I.F., Merckx, V. and Tedersoo, L. (2020). FungalRoot: global online database of plant mycorrhizal associations. New Phytol, 227,: 955-966
- Spagnoletti, F.N., Cornero, M., Chiocchio, V., Lavado, R.S., Roberts, I.N. (2020). Arbuscular mycorrhiza protects soybean plants against Macrophomina phaseolina even under nitrogen fertilization. Eur. J.Plant. Pathol, 156, 839–849.
- Spatafora , J.W., Chang, Y., Benny, G.L., Lazarus, K., Smith, M.E., Berbee, M.L, Bonito, G., Corradi , N., Grigoriev , I., Gryganskyi, A., James, T.Y., O'Donnell, K., Roberson, R.W., Taylor, T.N., Uehling, J., Vilgalys, R., White, M.M., Stajich, J.E. (2016). A phylum-level phylogenetic classification of zygomycete fungi based on genome-scale data. Mycologia, 108(5), 1028-1046.
- 78. Sun, J., Jia, Q., Li, Y., Zhang, T., Chen, J., Ren, Y., Dong, K., Xu, S., Shi, N.-N & Fu, S. (2022). Effects of Arbuscular Mycorrhizal Fungi and Biochar on Growth, Nutrient Absorption, and Physiological Properties of Maize (Zea mays L.). J. Fungi, 8, 1275.
- 79. Sun, J., Jia, Q., Li, Y., Zhang, T., Chen, J., Ren, Y., Dong, K., Xu, S., Shi, N.-N., Fu, S. (2022). Effects of Arbuscular Mycorrhizal Fungi and Biochar on Growth, Nutrient Absorption, and Physiological Properties of Maize (Zea mays L.). J. Fungi, 8, 1275.
- Sun, Z., Song, J., Xin, X., Xie, X., & Zhao, B. (2018). Arbuscular mycorrhizal fungal proteins 14-3-3- are involved in arbuscule formation and responses to abiotic stresses during AM symbiosis. Front. Microbiol. 5, 9–19.
- 81. Szabo, K., Böll, S & Erős-Honti, ZS. (2014). Applying artificial mycorrhizae in planting urban trees. Appl Ecol.12:835–853.
- Teotia, P., Kumar, M., Prasad, R., Kumar, V., Tuteja, N., & Varma, A. (2017). Mobilization of micronutrients by mycorrhizal fungi. Mycorrhiza-function, diversity, state of the art, 9-26.
- 83. Thirkell, T. J., Charters, M. D., Elliott, A. J., Sait, S. M & Field, K. J. (2017). Are mycorrhizal fungi our sustainable saviours considerations for achieving food security. J. Ecol. 105, 921–929.
- 84. Tyub, Sumira ., Kamili , Azra ., Reshi , Zafar ., Mearaj , Syed., Mokhdomi, Taseem ., Bukhari , Shoiab ., Wafai , Asrar ., Amin, Asif & Qadri, Raies. (2016). Ectomycorrhizae: Activity and Growth. European Academic Research. IV. 4481-4505.
- 85. Vogelsang, KM ., Bever, JD ., Griswold , M & Schultz PA (2004) The use of mycorrhizal fungi in erosion control applications. Contract 1–150
- Wang, F. Occurrence of arbuscular mycorrhizal fungi in mining-impacted sites and their contribution to ecological restoration: Mechanisms and applications. Crit. Rev. Environ. Sci. Technol. 2017, 47, 1–57.
- 87. Wang, Y.-Y., Yin, Q.-S., Qu, Y., Li, G.-Z & Hao, L. (2017). Arbuscular mycorrhizamediated resistance in tomato against Cladosporium fulvum -induced mould disease. J. Phytopathol, 166, 67–74.

- 88. Weng, W., Yan, J., Zhou, M., Yao, X., Gao, A., Ma, C., Cheng, J. and Ruan, J. (2022). Roles of arbuscular mycorrhizal fungi as a biocontrol agent in the control of plant diseases. Microorganisms, 10(7), p.1266.
- 89. Xavier, L. J., & Boyetchko, S. M. (2004). Arbuscular mycorrhizal fungi in plant disease control. Mycology series, 21, 183-194.
- 90. Xin, Zhang., Baodong, Chen., & Ryo Ohtomo (2015). Mycorrhizal effects on growth, P uptake and Cd tolerance of the host plant vary among different AM fungal species, Soil Science and Plant Nutrition, 61:2, 359-368.
- Zambolin L, Schenck NC. Reduction of the effects of pathogenic root-infecting fungi on soybean by the mycorrhizal fungus, Glomus mosseae. Phytopathology. 1983; 73, 1402-1405.
- 92. Zhu, X. C., Song, F. B., and Xu, H. W. (2010a). Arbuscular mycorrhizae improve low temperature stress in maize via alterations in host water status and photosynthesis. Plant Soil. 331, 129–137. doi: 10.1007/s11104-009-0239-z
- 93. Zhu, X.-C., Song, F.-B., Liu, S.-Q & Liu, T.-D. (2011). Effects of arbuscular mycorrhizal fungus on photosynthesis and water status of maize under high temperature stress. Plant. Soil, 346, 189–199.
- 94. Zou, Y. N., Srivastava, A. K., & Wu, Q. S. (2016). Glomalin: a potential soil conditioner for perennial fruits. Int. J. Agric. Biol. 18, 293–297.