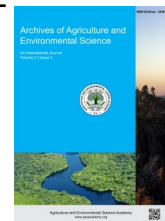




e-ISSN: 2456-6632

This content is available online at AESA


Archives of Agriculture and Environmental Science  
Journal homepage: journals.aesacademy.org/index.php/aaes



ORIGINAL RESEARCH ARTICLE



## Genotypic variability and genetic parameters for root yield, dry matter and related traits of cassava in the Guinea Savannah ecological zone of Ghana

Joseph Adjebeng-Danquah<sup>1\*</sup> , Kwabena Acheremu<sup>1</sup>, Emmanuel Boache Chamba<sup>1</sup>, Issah Alidu Abukari, Ayishetu Sumaila<sup>1</sup>, Richard Yaw Agyare<sup>1</sup>, Isaac Kodzo Amegbor<sup>1</sup>, Freda Ansaah Agyapong<sup>1</sup>, Ophelia Asirifi Amoako<sup>1</sup>, Flora Christine Amagloh<sup>1</sup>, Yussif Baba Kassim<sup>1</sup> and Elizabeth Parkes<sup>2</sup>

<sup>1</sup>Council for Scientific and Industrial Research - Savanna Agricultural Research Institute, P. O. Box TL 52, Tamale, GHANA

<sup>2</sup>International Institute of Tropical Agriculture, PMB 5320, Oyo Road, Ibadan, NIGERIA

\*Corresponding author's E-mail: barchus2003@yahoo.com

### ARTICLE HISTORY

Received: 07 July 2023

Revised received: 25 August 2023

Accepted: 14 September 2023

### Keywords

Adaptability

Agronomic performance

Dry matter content

Genotype stability index

Starch yield

### ABSTRACT

The aim of this study was to assess the agronomic performance and genetic parameters governing storage root yield and related traits in cassava genotypes in order to identify superior genotypes. The study involved 18 elite cassava genotypes which were arranged in a randomized complete block design with three replications and assessed for storage root yield and yield components (12 months after planting) in the Guinea savannah ecology of Ghana for three seasons. Analysis of variance indicated significant ( $p < 0.05$ ) genotype and year main effects for fresh and dry root yields, dry matter content, starch yield and harvest index. Genotype  $\times$  year effect was significant ( $p < 0.05$ ) for fresh root yield, dry root yield and starch yield. Estimates of the variance components revealed greater genotypic influence for starch yield, fresh and dry root yields implying the potential for genetic gain with selection for these traits among the genotypes. Relatively high (69 %) broad sense heritability estimate was observed for dry storage root yield indicating the depth of genetic influence. Path coefficient analysis revealed a direct positive effect of dry matter content on dry storage root yield whilst dry storage root yield had direct positive effect on starch yield suggesting the possibility of indirect selection for starch yield through dry storage root yield. The study revealed ample genotypic variability among the cassava genotypes to warrant selection. Four genotypes, IBA 070134, IBA 419, IBA 950289 and IBA 980581 were identified for high and stable fresh and dry root yields for multilocational testing.

©2023 Agriculture and Environmental Science Academy

**Citation of this article:** Adjebeng-Danquah, J., Acheremu, K., Chamba, E. B., Abukari, I. A., Sumaila, A., Agyare, R. Y., Amegbor, I. K., Agyapong, F. A., Amoako, O. A., Amagloh, F. C., Kassim, Y. B., & Parkes, E. (2023). Genotypic variability and genetic parameters for root yield, dry matter and related traits of cassava in the Guinea Savannah ecological zone of Ghana. *Archives of Agriculture and Environmental Science*, 8(3), 310-318, <https://dx.doi.org/10.26832/24566632.2023.080306>

### INTRODUCTION

Cassava (*Manihot esculenta* Crantz) is a perennial shrub belonging to the *Euphorbiaceae* family and widely cultivated in the tropics particularly South America, Africa and the Pacific for its starchy roots (Lopez-Diago *et al.*, 2018). Due to its high productivity under adverse conditions, all year-round availability and compatibility with different farming systems, it is consid-

ered an ideal food security crop (Okeleye *et al.*, 2001). The crop possesses different adaptive morphological and physiological attributes that make it resilient in dry ecologies where most crops grown for food and feed would fail (Adjebeng-Danquah *et al.*, 2020; Okogbenin *et al.*, 2013; El-Sharkawy and Cadavid, 2002; El-Sharkawy, 2007; Lenis *et al.*, 2006) It is therefore an ideal feed and food security crop in such ecologies (El-Sharkawy, 2004; Hillocks, 2002). Despite the high potential

of cassava as an industrial raw material particularly starch and animal feed industry (Ceballos *et al.*, 2004; Ceballos *et al.*, 2012), past breeding efforts in the Guinea savannah ecology have focused on developing early bulking varieties and varieties with good food quality with adaptation to the short growing season of the Guinea Savannah agroecological zone of Ghana (Adjebeng-Danquah *et al.*, 2012; Adu-Gyamfi *et al.*, 2016; Osei *et al.*, 2002). Genotypes that fail to meet these objectives are discarded at an earlier stage of the breeding programme. This has thus limited the broad scale utilization of the crop to only food uses and minor processing. However, certain cassava varieties have high dry matter content which makes them ideal for industrial processing. Dry matter content is highly polygenic trait in cassava and is very useful for the acceptance or rejection of cassava varieties by most end-users including processors (Prasannakumari *et al.*, 2021). High dry matter cassava varieties have been useful in starch production since high dry matter content has been found to be positively associated with high starch content (Maraphum *et al.*, 2021; Teye *et al.*, 2011). Starch from cassava is one of the most preferred and cheapest sources of starch and has relatively better pasting properties, good texture, good stability and swelling power (Amelework and Bairu, 2022). This makes it ideal as raw material for a number of industries. The starch also has numerous applications in the paper, textile, pharmaceutical (as excipient), adhesives, food (as thickener), water treatment (as coagulant), and polymer industries (Hernández-Carmona *et al.*, 2017; Koopmans, 2005) as well as in the production of biopolymeric materials and food packaging film (Tumwesigye *et al.*, 2017).

The full economic potential of cassava for industrial use can be exploited if high yielding productive and industry-preferred cassava varieties could be identified for cultivation. The productivity of cassava and suitability for different products depends on several factors including variety and environmental factors at the time of production (Amelework *et al.*, 2023; Amelework and Bairu, 2022). Several studies have reported significant genetic variation among cassava genotypes in terms of dry matter content and starch properties which can be influenced by the environment and genotype  $\times$  environment interaction (Agunbiade and Ighodaro, 2010; Ayetigbo *et al.*, 2018; Eke *et al.*, 2007; Nuwamanya *et al.*, 2009; Omodamiro *et al.*, 2007). Dry matter content as well as the quality and quantity of starch obtained depend on the genotype, environmental conditions and the interaction between a number of factors (Adjebeng-Danquah *et al.*, 2016; Eke *et al.*, 2007). It has thus become important to evaluate and select cassava varieties not only for their food quality and uses, but also as an industrial commodity. Since most traits of economic importance in cassava are subject to influence by genotype, environment and genotype  $\times$  environment interaction (Amelework *et al.*, 2023; Aghogho *et al.*, 2022; Bakare *et al.*, 2022; Uchendu *et al.*, 2022), it is difficult to predict genotypic performance in different years or environments. As a result, genotypes have to be evaluated for several years or across several environments to assess their stability for these traits (Haldavankar *et al.*, 2009). Genotypes that exhibit less

variation in performance relative to a check variety across years and/or environments, are considered to be stable (Olivoto, *et al.*, 2019). After these multi-year or multi-environment evaluations, the successful genotypes that show superior performance are considered as suitable candidates that have the potential to thrive under the different environmental conditions on farmers' fields and across years. The objective of this study was to assess the genotypic variability for agronomic performance, genetic parameters and stability for storage root yield, dry matter content, root starch content and related traits among 18 elite cassava genotypes in the Guinea Savannah ecology of Ghana.

## MATERIALS AND METHODS

### Description of the study site

The study was carried out at the CSIR-Savanna Agricultural Research Institute research fields, Nyankpala from 2016 to 2018. The study area is located in the Guinea savannah agroecological zone which is characterized by a single rainy season which normally begins in April and ends in October or November. Intermittent dry spells often lasting two to three weeks occur within the growing season (Alua *et al.*, 2018). Average annual rainfall is estimated to be about 1033 mm. Average annual temperature is 28.1 °C and relative humidity of 61 % also occurs in the area. Average potential evapotranspiration (1720 mm) normally exceeds annual rainfall (Agyemang and Abdul-korah, 2014; EPA, 2003). The soil of the study areas was a well-drained ferric luvisol, locally known as the Tingoli series (FAO-UNESCO, 1977).

### Germplasm and planting materials used for the study

Eighteen cassava genotypes were used for the study. These genotypes comprised 16 elite genotypes obtained from the breeding programme of the International Institute of Tropical Agriculture (IITA). They were evaluated alongside two farmer preferred varieties, "Biabasse" and "Eskamaye". Biabasse is a landrace widely cultivated in cassava growing communities in northern Ghana whilst Eskamaye was formally released as a variety in 2002 by the Savannah Agricultural Research Institute of the Council for Scientific and Industrial Research, Nyankpala, Ghana and has a yield potential of 16-23 t ha<sup>-1</sup> (Osei *et al.*, 2002).

### Land preparation, experimental design and planting

The land was ploughed and harrowed after which ridges were manually raised with hoes using inter-row spacing of one meter. The cassava genotypes representing the treatments were arranged in a randomized complete block design (RCBD) with three replications. Each plot consisted of four rows with five stands within a row giving a total of 20 stands in a plot resulting in an estimated population of 10,000 plants per hectare. Cassava stakes measuring approximately 25 – 30 cm were planted on top of the ridges using intra row of one meter. Manual weed control was carried out as and when necessary. Reshaping of ridges was carried out after each weeding to avoid exposing the roots.

### Data collection

Harvesting was done at 12 months after planting on the two middle rows. Data were recorded on fresh root weight (kg) and fresh shoot weight (kg). Fresh root yield ( $t\ ha^{-1}$ ), harvest index and dry matter content (%) were computed according to the procedure described in Adjebeng-Danquah *et al.* (2016). Dry root yield ( $t\ ha^{-1}$ ) was obtained by multiplying the fresh root yield by the dry matter content (%). Starch content was estimated using the specific gravity method as described by Fukuda *et al.* (2010). Starch yield ( $t\ ha^{-1}$ ) was computed as the product of starch content and fresh storage root yield ( $t\ ha^{-1}$ ).

### Data analysis

The data was subjected to analysis of variance using Genstat Statistical package version 12.1 (Payne *et al.*, 2009). Treatment means were separated using the standard error of the difference ( $p < 0.05$ ). Genotypes were considered as fixed factors whilst years were considered as random factors. Estimates of variance components were obtained from the observed mean squares by equating with their respective expected mean squares (Ewa *et al.*, 2017; Ntawuruhunga and Dixon, 2010). Genotypic coefficient of variation ( $CV_g$ ), environmental coefficient of variation ( $CV_e$ ), and relative coefficient of variation ( $CV_r$ ) and broad sense heritability were then estimated (Adjebeng-Danquah *et al.*, 2017; Oliveira *et al.*, 2021).

### Variance components:

$$CV_g (\%) = \left( \frac{\sqrt{\sigma_g^2}}{X} \right) * 100$$

$$CV_p (\%) = \left( \frac{\sqrt{\sigma_p^2}}{X} \right) * 100$$

$$CV_r = \frac{CV_g}{CV_e}$$

Where:  $CV_g$  = Genotypic coefficient of variation;  $CV_e$  = environmental coefficient of variation;  $CV_r$  = relative coefficient of variation;  $X$  = grand mean

$$\sigma_g^2 = \text{genotypic variance: } \sigma_g^2 = MSg - \frac{MSe}{r}$$

$$\sigma_p^2 = \text{phenotypic variance: } \sigma_p^2 = \sigma_g^2 + \frac{\sigma_{g \times y}^2}{Y} + \frac{\sigma_e^2}{Y \times R}$$

Where:  $MSg$  = Genotypic mean square;  $MSe$  = Error mean square;  $\sigma_{g \times y}^2$  = Genotype  $\times$  year variance;  $\sigma_y^2$  = Year variance;  $Y$  = number of years;  $R$  = number of replications.

$$\text{Broad sense heritability: } H^2 = \frac{\sigma_g^2}{\sigma_p^2} \times 100$$

### Estimation of the AMMI stability value

To assess the different genotypes for stable performances over the years of evaluation, the additive main effect multiplicative

interaction (AMMI) analysis was performed using Genstat 12.1 (Payne *et al.*, 2009). Subsequently, AMMI stability value and genotype stability index were estimated (Purchase *et al.*, 2000). The AMMI stability value (ASV) was calculated as follows:

$$ASV = \sqrt{\left[ \frac{IPCA1_{\text{Sum of squares}}}{IPCA2_{\text{Sum of squares}}} (IPCA1_{\text{Score}}) \right]^2 + (IPCA2_{\text{Score}})^2}$$

Where:  $\frac{IPCA1_{\text{Sum of squares}}}{IPCA2_{\text{Sum of squares}}}$  = the weight given to the IPCA1-value by dividing the IPCA1 sum of squares (from the AMMI analysis of variance table) by the IPCA2 sum of squares. The larger the IPCA score, either negative or positive, the more adapted a genotype is to a certain environment. Smaller ASV scores indicate a more stable genotype across environments (Farshadfar *et al.*, 2011).

Genotype stability index (GSI) was also calculated using the sum of the ranking based on yield and ranking based on the AMMI stability value.

$$GSI = RASV + RG$$

Where:  $RASV$  = the rank of the genotypes based on the AMMI stability value and  $RG$  = the rank of the genotypes based on performance across environments.

GSI incorporates both mean yield and stability in a single criterion. Low values of both parameters show desirable genotypes with high mean yield and stability (Adjebeng-Danquah *et al.*, 2017; Bose *et al.*, 2014; Tumuhimbise *et al.*, 2014).

### Correlation and path analysis

The information on the interrelationships between traits is important in the selection of key secondary traits for crop improvement. Both phenotypic and genotypic correlation analyses were carried out to determine the relationship between the different traits based on sequential stepwise multiple regressions (Adu *et al.*, 2016). For the path analysis, storage root yield, starch content and the other agronomic traits measured were examined based on stepwise multiple analysis and illustrated using path diagram.

## RESULTS AND DISCUSSION

### Combined analysis of variance for storage root yield and yield components

Recent demand for cassava as an industrial crop has moved the crop from a staple food security crop to industrial raw material (Ceballos *et al.*, 2020). This has necessitated the development of new varieties not only with good cooking quality but with industry-preferred attributes such as high dry matter and high starch content which are preferred by most cassava industries (Dankwa and Peprah, 2019). This study sought to evaluate and identify cassava genotypes with high storage root yield and high dry matter content for industrial processing.

**Table 1.** Mean squares for six traits measured on 18 cassava genotypes evaluated at Nyankpala from 2016 – 2018.

Source of variation	d.f.	FSRY (t ha <sup>-1</sup> )	DM (%)	DSRY (t ha <sup>-1</sup> )	Stch (%)	Stchylid (t ha <sup>-1</sup> )	HI
Replication	2	135.06	16.10	13.96	12.39	5.67	375.42
Genotype	17	130.75***	66.58***	22.77***	32.55***	7.57***	189.02*
Year	2	955.08***	278.93***	51.64***	15.05ns	48.95***	857.57***
Genotype × Year	34	42.82*	15.16ns	5.11*	10.11ns	2.35*	130.84ns
Error	106	29.47	13.40	3.03	7.82	1.27	89.28
Total	161						

\*, \*\*, \*\*\* = significant at  $P < 0.05$ ,  $P < 0.01$  and  $P < 0.001$  respectively, ns = not significant ( $P > 0.05$ ). FSRY = fresh storage root yield (t ha<sup>-1</sup>), DM% = dry matter content, DSRY = dry storage root yield, Stch% = starch content of storage roots, Stchylid (t ha<sup>-1</sup>) = starch yield (t ha<sup>-1</sup>), HI = Harvest index.

**Table 2.** Mean ( $\pm$ ), estimates of variance components, heritability, phenotypic and genotypic coefficient of variation for five traits from 18 cassava genotypes evaluated from 2016-2018.

Parameters	Fresh root yield (t ha <sup>-1</sup> )	Dry matter content (%)	Dry root yield (t ha <sup>-1</sup> )	Starch content (%)	Starch yield (t ha <sup>-1</sup> )	Harvest index (%)
Mean ( $\pm$ STDEV)	19.90 $\pm$ 7.47	32.91 $\pm$ 4.81	6.50 $\pm$ 1.70	19.30 $\pm$ 3.33	3.86 $\pm$ 1.68	59.91 $\pm$ 11.03
$\sigma_g^2$	16.01	7.59	2.76	3.87	0.96	25.62
$\sigma_{g \times y}^2$	4.45	0.20	0.23	0.25	0.12	4.62
$\sigma_y^2$	9.82	4.47	1.01	2.61	0.42	29.76
$\sigma_p^2$	30.28	12.26	4.00	6.73	1.50	60.00
$H_b^2$	52.87	61.96	69.02	57.50	63.86	42.70
CV <sub>p</sub> (%)	28.05	10.64	30.82	13.44	31.76	12.93
CV <sub>g</sub> (%)	20.39	8.37	25.61	10.19	25.38	8.45
CV <sub>e</sub> (%)	15.97	6.42	15.47	8.37	16.87	9.11
CV <sub>r</sub>	1.28	1.30	1.65	1.22	1.50	0.93

Key:  $\sigma_g^2$  = genotypic variance,  $\sigma_{g \times y}^2$  = genotype × environment variance,  $\sigma_y^2$  = Year (environmental) variance,  $\sigma_p^2$  = phenotypic variance,  $H_b^2$  = broad sense heritability, CV<sub>p</sub> (%) = Phenotypic coefficient of variation, CV<sub>g</sub> (%) = Genotypic coefficient of variation, CV<sub>e</sub> (%) = environmental coefficient of variation, CV<sub>r</sub> = relative coefficient of variation.

Analysis of the results indicated highly significant ( $p < 0.001$ ) variability among the cassava genotypes for storage root yield, dry matter content and harvest index over the three years of study. Genotype × year interaction effect was also significant ( $p < 0.05$ ) for fresh root yield, dry root yield and starch yield (Table 1). The effect of year was also highly significant ( $p < 0.001$ ) for all the traits except for starch content which was not significant ( $p > 0.05$ ). Several studies have suggested that the success of any plant breeding programme depends on the extent of genotypic variability present in the germplasm assembled (Salgotra and Chauhan, 2023; Swarup et al., 2021; Varshney et al., 2021; Cobb et al., 2019). The observed significant genotype effect for the different traits suggests the presence of ample genotypic variability for these traits indicating the chance of success in terms selection for best genotypes for these traits (Oliveira et al., 2021).

#### Estimates of genetic parameters and broad sense heritability

Estimates of variance components indicated that genotypic variances ( $\sigma_g^2$ ) for all the traits were greater than their corresponding variances due to year ( $\sigma_y^2$ ) and genotype × year interaction ( $\sigma_{g \times y}^2$ ) except for harvest index (Table 2). Harvest index had a greater year variance than the genotype and genotype × year variances. These results though similar to the observations of Oliveira et al. (2021), are contrary to the findings of Silva et al. (2019) who recorded higher error or environmental variance than variance due to genotype in 49 cassava accessions

evaluated under drought conditions. This implies that these parameters vary with the specific population used. Harvest index had a greater year variance than the genotype and genotype × year variances suggesting that, the conditions in a particular year had a greater influence on the amount of dry matter partitioned into the storage roots relative to the above ground biomass. It also suggests that the performance of genotype(s) could vary with different years. Therefore, multilocation trials need to be considered within the target region to understand, exploit and/or minimize the adverse effect of years and genotype × environment interaction when selecting for genotypes based on harvest index. Earlier studies have reported strong influence of years or environments on harvest index and such influence tends to increase under unfavorable environmental conditions (Adjebeng-Danquah et al., 2020; Baafi and Safo-Kantanka, 2008). For a clonally propagated crop like cassava, desirable traits to be considered for selection must have higher genotypic influence which ensures more reliability than traits with higher environmental influences for simple direct selection to be possible (Tuberosa, 2012). Broad sense heritability estimates ranged from low (43 %) to medium (69 %) for harvest index and dry root yield respectively (Table 2). In the case of traits with low broad sense heritability estimates, direct selection within this population may not be effective and will result in slow progress (Li et al., 2020; Diniz and Oliveira, 2019). It will thus require the use of secondary traits that are strongly linked to these traits of interest.

**Table 3.** Phenotypic correlation (below diagonal) and genetic correlation (above diagonal) among traits for 18 cassava genotypes evaluated over three years at Nyankpala.

Traits	FSRY (t ha <sup>-1</sup> )	DM (%)	DSRY (t ha <sup>-1</sup> )	Stch (%)	Stchylid (t ha <sup>-1</sup> )	HI
<b>Genotypic correlation</b>						
FSRY (t ha <sup>-1</sup> )	-	-0.15ns	0.93***	0.09ns	0.91***	0.28***
DM (%)	0.58**	-	0.21**	0.81***	0.17*	0.13ns
DSRY (t ha <sup>-1</sup> )	0.98***	0.74***	-	0.37***	0.97***	0.33***
Stch (%)	0.35ns	0.99***	0.56*	-	0.47***	0.04ns
Stchylid (t ha <sup>-1</sup> )	0.94***	0.84***	0.98***	0.76***	-	0.26***
HI	0.93***	0.61**	0.86***	0.49*	0.89***	-
<b>Phenotypic correlation</b>						

Key: \*, \*\*, \*\*\* = significant at  $P < 0.05$ ,  $P < 0.01$  and  $P < 0.001$  respectively, ns = not significant ( $P > 0.05$ ), FSRY = Fresh storage root yield (t ha<sup>-1</sup>), DM = Dry matter content (%), DSRY = dry storage root yield (t ha<sup>-1</sup>), Stch = starch content of storage roots (%), Stchylid = starch yield (t ha<sup>-1</sup>), HI = Harvest index.

**Table 4.** Performance of genotypes for six traits of 18 cassava genotypes evaluated in Nyankpala.

Genotype	FSRY (t ha <sup>-1</sup> )	DM (%)	DSRY (t ha <sup>-1</sup> )	Stch (%)	Stchylid (t ha <sup>-1</sup> )	HI (%)
Biabasse	15.89	36.66	5.73	21.76	3.45	60.27
Eskamaye	18.03	31.62	5.70	18.11	3.28	61.90
IBA 010034	17.15	33.72	5.77	19.68	3.39	57.99
IBA 010040	19.11	32.00	6.05	19.35	3.69	56.55
IBA 011368	17.87	30.67	5.36	17.89	3.19	57.98
IBA 011371	13.94	28.09	3.88	16.62	2.31	52.27
IBA 020431	19.89	35.72	7.01	21.75	4.33	60.99
IBA 020452	18.76	35.94	6.61	21.42	4.04	55.19
IBA 061635	17.64	29.64	5.23	16.43	2.95	53.82
IBA 070134	32.28	36.03	11.60	20.01	6.47	66.91
IBA 30572	19.35	32.74	6.13	18.88	3.61	57.90
IBA 419	23.20	35.26	8.01	21.66	5.04	65.76
IBA 9102324	21.07	32.40	7.00	19.44	4.33	65.47
IBA 950289	21.13	28.04	5.82	15.78	3.32	62.20
IBA 961089A	21.09	32.03	6.67	18.44	3.88	65.83
IBA 980505	19.20	31.57	5.87	18.42	3.47	57.82
IBA 980581	21.58	35.80	7.62	21.53	4.64	54.64
IBA 993073	20.95	34.46	6.85	20.21	4.13	64.88
Mean	19.90	32.91	6.50	19.30	3.86	59.91
SED	2.45	1.73	0.82	1.32	0.53	4.45

Key: FSRY = Fresh storage root yield (t ha<sup>-1</sup>), DM = Dry matter content (%), DSRY = dry storage root yield (t ha<sup>-1</sup>), Stchylid = starch yield (t ha<sup>-1</sup>), HI = Harvest index.

Phenotypic coefficient of variation ( $CV_p$  %) ranged from 10.64 to 31.76 % for dry matter content and starch yield respectively. Genotypic coefficient of variation ( $CV_g$  %) varied between 8.37 and 25.61 % for dry matter content and dry root yield, respectively. Though phenotypic coefficients of variation were generally higher than the corresponding genotypic coefficients of variation ( $CV_g$  %), the magnitude of the differences varied with the different traits. This demonstrated the extent of the effect of the environment (different years) on the different traits. The environmental coefficient of variation ( $CV_e$  %) values of all the traits apart from harvest index, were lower than their corresponding genotypic coefficient of variation ( $CV_g$  %). The relative coefficient of variation ( $CV_r$ ) also known as relative variation index was greater than one for all traits except harvest index. This suggests that direct selection could be based on these traits (apart from harvest index) to select elite cassava genotypes for further improvement provided they are positively correlated with traits of economic importance (Diniz and Oliveira, 2019).

For traits with higher environmental influence, indirect selection based on secondary traits (with high heritability) could be more effective (Oliveira *et al.*, 2021).

#### Phenotypic and genotypic correlation analysis

Storage root yield in cassava is a complex quantitatively inherited trait which is associated with several physiological and morphological traits that are in turn highly influenced by the conditions prevailing in the growing environment (Adjebeng-Danquah *et al.*, 2020). It is therefore necessary to identify secondary traits that are positively related to the traits of interest to use them as indirect selection criteria (Rao *et al.*, 2017). The phenotypic correlation analysis showed highly significant associations (below the diagonal) between the different traits (Table 3). Fresh root yield was positively and highly correlated with dry root yield ( $r_p = 0.93$ ), starch yield ( $r_p = 0.91$ ) and harvest index ( $r_p = 0.28$ ) with which it had a weak association. A strong positive and highly significant correlation was found between dry root yield and starch yield ( $r_p = 0.97$ ).

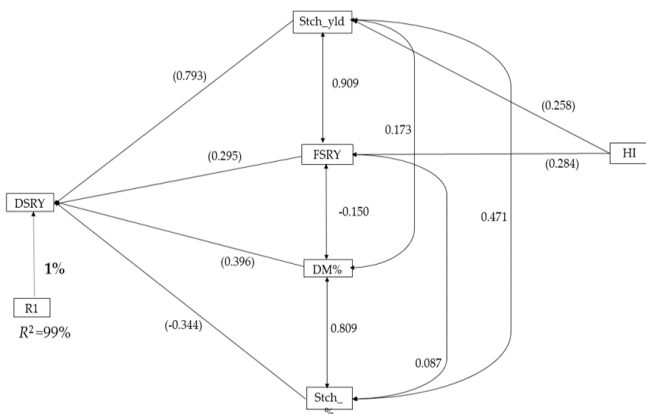
Dry matter content was also significantly and positively correlated with starch content ( $r_p = 0.81$ ) and starch yield agreeing with the findings of previous studies which also reported a significant direct positive correlation between dry matter content and starch content in cassava (Maraphum et al., 2021; Diniz and Oliveira, 2019; Oliveira et al., 2015). However, the correlations between harvest index and dry matter content as well as harvest index with starch content were not significant. The genotypic correlation analysis (above the diagonal) also indicated highly significant associations between the different traits. Fresh storage root yield was positively and highly correlated with harvest index ( $r_g = 0.93$ ), starch yield ( $r_g = 0.94$ ) and dry storage root yield ( $r_g = 0.98$ ). A strong positive and highly significant correlation was found between dry matter content and starch content ( $r_g = 0.99$ ), starch yield ( $r_g = 0.84$ ) and dry storage root yield ( $r_g = 0.74$ ). This observation agrees with the findings of Amelework and Bairu (2021) who reported strong association between dry matter content and starch content in cassava. However, there was a weak positive correlation between fresh storage root yield and starch content ( $r_g = 0.35$ ), and an almost perfect correlation between starch yield and dry storage root yield ( $r_g = 0.98$ ). The significant and positive associations among the different traits indicate that direct selection for any of the traits

would indirectly lead to the improvement of the associated traits simultaneously.

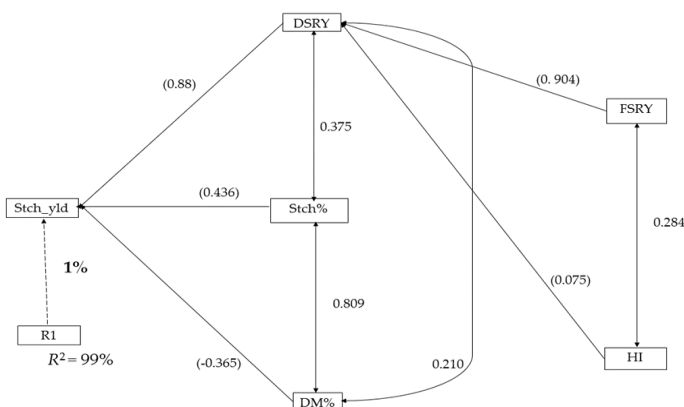
### Path analysis

The use of secondary traits in indirect selection is one of the most effective ways of selecting traits with low heritability or traits that are difficult to measure (Li et al., 2020; Diniz and Oliveira, 2019). Path analysis was used to determine the direct and indirect relationships between dry storage root yield, starch yield and the other related traits in a systematic manner. First, dry storage root yield was considered as the primary trait of interest or resultant (dependent) variable (Figure 1) whilst all the other traits including fresh storage root yield, starch content, starch yield, dry matter content and harvest index were considered as the independent or causal variables. Starch yield (0.793), fresh storage root yield (0.295) and dry matter content (0.396) all had direct positive effect on dry storage root yield whilst starch content had a direct negative effect (-0.344) on dry storage root yield. Harvest index had a positive indirect effect on dry storage root yield through fresh storage root yield (0.284) and starch yield (0.258). Strong positive correlation was found between starch yield and fresh storage root yield (0.909), dry matter content and starch content (0.809) but moderate correlation between starch yield and starch content (0.471). Though strong direct effect of fresh root yield on starch yield had been reported in an earlier study (Diniz and Oliveira, 2019), the findings from the current study rather revealed an indirect relationship between fresh storage root yield and starch yield through dry storage root yield. They further suggested that high root yield could be used as an efficient trait for targeting high starch yield. From our study, it would be important to consider dry matter content alongside storage root yield when targeting genotypes with high starch yield potential.

Secondly, starch yield was considered as the resultant (dependent) variable (Figure 2) whilst all the other traits including fresh root yield, starch content, dry matter content, dry storage root yield and harvest index were considered as the independent or causal variables. Dry storage root yield, starch content and dry matter content were classified as first order traits which had direct relationship with starch yield. Fresh root yield and harvest index had indirect effect on starch yield. Dry storage root yield had the strongest direct positive effect (0.88) on starch yield which was followed by starch content (0.436). Dry matter content (%) had a negative direct effect (-0.365) on starch yield. However, dry matter content had an indirect positive effect on starch yield through starch content (0.809). Fresh storage root yield had a strong positive indirect effect on starch yield through dry storage root yield (0.904) whilst a weak indirect relationship was observed between harvest index and starch yield through dry storage root yield.



**Figure 1.** Path diagram showing direct and indirect relationship between dry storage root yield and other traits. FSRY = Fresh storage root yield ( $t\ ha^{-1}$ ), DM = Dry matter content (%), DSRY = dry storage root yield ( $t\ ha^{-1}$ ), Stchylid = starch yield ( $t\ ha^{-1}$ ), HI = Harvest index, R1 = Residual;  $R^2$  = Coefficient of determination.



**Figure 2.** Path diagram showing direct and indirect relationship between starch yield and other traits. FSRY = Fresh storage root yield ( $t\ ha^{-1}$ ), DM = Dry matter content (%), DSRY = dry storage root yield ( $t\ ha^{-1}$ ), Stchylid = starch yield ( $t\ ha^{-1}$ ), HI = Harvest index, R1 = Residual;  $R^2$  = Coefficient of determination.

### Average performance of the cassava genotypes for six traits

Average fresh root yield across the three years ranged between  $13.94\ t\ ha^{-1}$  (IBA 011371) and  $32.28\ t\ ha^{-1}$  (IBA 070134) with a mean yield of  $19.90\ t\ ha^{-1}$  (Table 4).

**Table 5.** Ranking of 18 cassava genotypes for fresh root yield (FSRY), dry root yield (DSRY) and starch yield over three years based on AMMI stability value (ASV) and genotype stability index (GSI) at 12 months after planting.

Genotype	FSRY (t ha <sup>-1</sup> )	ASV	RASV	Rank FRY	Rank sum	GSI Rank	DSRY (t ha <sup>-1</sup> )	ASV	RASV	Rank DRY	Rank sum	GSI Rank	StchYld (t ha <sup>-1</sup> )	ASV	RASV	Rank StchYld	Rank sum	GSI Rank
Biabasse	15.89	0.44	4	17	21	10	5.73	0.06	1	14	15	6	3.45	0.09	4	12	16	7
Eskamaye	18.03	1.05	8	13	21	9	5.69	0.95	14	15	29	18	3.28	0.74	11	15	26	17
IBA010034	17.15	1.39	11	16	27	17	5.77	0.91	13	13	26	16	3.39	0.78	13	13	26	15
IBA010040	19.11	18.42	17	11	28	18	6.05	2.53	18	10	28	17	3.69	3.70	18	9	27	18
IBA011368	17.87	1.65	12	14	26	16	5.36	0.19	5	16	21	10	3.19	0.31	7	16	23	12
IBA011371	13.94	0.81	7	18	25	15	3.88	0.21	6	18	24	14	2.31	0.24	6	18	24	13
IBA020431	19.89	2.92	15	8	23	11	7.01	0.40	8	4	12	3	4.33	0.68	10	5	15	5
IBA020452	18.76	0.36	3	12	15	6	6.61	0.85	12	8	20	9	4.04	0.87	14	7	21	10
IBA061635	17.64	1.16	9	15	24	12	5.23	0.35	7	17	24	15	2.95	0.31	8	17	25	14
IBA070134	32.28	0.69	6	1	7	3	11.60	0.40	9	1	10	2	6.47	0.44	9	1	10	3
IBA30572	19.35	3.72	16	9	25	14	6.13	0.17	4	9	13	4	3.61	0.18	5	10	15	6
IBA419	23.20	0.18	2	2	4	1	8.01	1.01	15	2	17	8	5.04	1.07	15	2	17	8
IBA9102324	21.07	1.95	13	6	19	8	6.99	1.73	17	5	22	12	4.33	1.95	16	4	20	9
IBA950289	21.13	0.06	1	4	5	2	5.82	0.58	11	12	23	13	3.32	0.75	12	14	26	16
IBA961089A	21.09	2.72	14	5	19	7	6.67	0.58	10	7	17	7	3.88	0.01	1	8	9	2
IBA980505	19.20	0.68	5	10	15	5	5.87	0.16	3	11	14	5	3.47	0.04	2	11	13	4
IBA980581	21.58	1.26	10	3	13	4	7.62	0.09	2	3	5	1	4.64	0.05	3	3	6	1
IBA993073	20.95	20.38	18	7	25	13	6.85	1.28	16	6	22	11	4.13	2.00	17	6	23	11

Key: FSRY = Fresh storage root yield (t ha<sup>-1</sup>), DSRY = Dry storage root yield (t ha<sup>-1</sup>), StchYld = starch yield (t ha<sup>-1</sup>), ASV = AMMI stability value, GSI = genotype stability index.

Average dry matter content on the other hand varied from 28.04 to 36.66 % for IBA 950289 and Biabasse, respectively, with a mean of 32.91 %. Genotype IBA 070134 had the highest dry root yield (11.60 t ha<sup>-1</sup>) which was superior to the local variety, Biabasse (5.73 t ha<sup>-1</sup>) and the improved check, Eskamaye (5.70 t ha<sup>-1</sup>). Two other genotypes, IBA 419 (8.01 t ha<sup>-1</sup>) and IBA 980581 (7.62 t ha<sup>-1</sup>) had significantly higher dry root yield than the checks. Average starch content ranged between 15.78 % for IBA 950289 and 21.76 % for Biabasse with a mean of 19.30 %. Average starch yield ranged from 2.31 t ha<sup>-1</sup> to 6.47 t ha<sup>-1</sup> for IBA 0110371 and IBA 070134 respectively with a mean of 3.86 t ha<sup>-1</sup>. The average harvest index across years was 59.91 % and ranged between 52 % and 66.91 % for IBA 011371 and IBA 070134 respectively. Besides genotypic performance, year effect was also significant for all the traits except starch content which was not influenced by the effect of year. Additionally, genotype × year interaction effect was significant for fresh root yield, dry root yield and starch yield. This suggests the tendency of the different genotypes to produce variable performances in different years depending on the environmental conditions. Several authors have suggested that significant genotype × environment interaction arises due to differential ranking of genotypic performances in different environments or years and can confound selection (Amelework et al., 2023; Aghogho et al., 2022; Bakare et al., 2022; Ebem et al., 2021; Osei et al., 2018). It could also arise from unequal variances (heterogenous variances) among genotypic means when evaluated in different environments (Amelework et al., 2023; Bakare et al., 2022). Therefore, multilocal testing of such genotypes is necessary to identify genotypes that would exhibit consistent performance for such traits.

### Ranking of genotypes based on storage root yield and starch yield

From the study, fresh root yield, dry root yield and starch yield were highly influenced by genotype × year interaction effect suggesting high variability of these traits in the different years. This necessitated the stability analysis to identify genotypes that would remain consistent in different years in terms of performance for these traits. The AMMI stability value and genotype stability index were used to rank the performance of genotypes based on these traits. The Additive Main effect and Multiplicative Interaction (AMMI) stability value (ASV) ranked genotypes based on the least score. Low scores represent the most stable genotypes. Genotypes IBA 419 (23.2 t ha<sup>-1</sup>), IBA 950289 (21.13 t ha<sup>-1</sup>), IBA 070134 (32.28 t ha<sup>-1</sup>) and IBA 980581 (21.58 t ha<sup>-1</sup>) combined high fresh root yield with high stability and were ranked 1-4 (Table 5). Genotypes IBA 950289, IBA 419 and IBA 070134 were also found to combine good yield with stability. Other introduced genotypes (IBA 980581, IBA 9102324, IBA 993073 and IBA 961089A) combined high fresh and dry root yields with good stability. Genotypes IBA 993073 (20.95 t ha<sup>-1</sup>) and IBA 020431 (19.89 t ha<sup>-1</sup>) though relatively high yielding, were less stable compared to the above genotypes.

Since stability per se does not connote desirable performance unless the genotype combines it with high yield (Farshadfar et al., 2011), the genotype stability index (GSI) is used to select genotypes that combine high yield with stability. Genotypes with the lowest GSI score are ranked highest and are most desired since they combine high yield with stability (Amelework et al., 2023). In terms of dry root yield, genotypes IBA 980581 (7.62 t ha<sup>-1</sup>), IBA 070134 (11.60 t ha<sup>-1</sup>) and IBA 020431 (7.01 t ha<sup>-1</sup>) also combined high dry root yield with high stability and were therefore ranked 1-3. These genotypes exhibited consistent performances over the three years of study. For starch yield, two genotypes IBA 980581 (4.64 t ha<sup>-1</sup>) and IBA 070134 (6.47 t ha<sup>-1</sup>) had superior starch yield compared to the farmer preferred variety, Biabasse (3.45 t ha<sup>-1</sup>) and the improved variety Eskamaye (3.28 t ha<sup>-1</sup>) and were also stable over the three years.

## Conclusion

The study revealed significant genotypic variability for fresh root yield, dry root yield and starch yield to warrant selection among the cassava genotypes for these traits. Estimates of genetic parameters indicated strong genotypic influence for fresh root yield, dry root yield and starch yield. Genotype × year interaction effect was significant for fresh root yield, dry root yield and starch yield which necessitates multilocal testing to identify genotypes that are stable for these traits. Four genotypes, IBA 070134, IBA 419, IBA 950289 and IBA 980581 showed high and stable performance for fresh and dry root yields over the three seasons' on-station testing making them suitable candidates for multilocal testing to assess their overall adaptability across the Guinea savannah ecology of Ghana.

## Conflicts of interest

The authors declare no conflict of interest.

## ACKNOWLEDGMENTS

The authors duly acknowledge the support of the Management and staff of the CSIR-Savanna Agricultural Research Institute, Nyankpala for the support in carrying out this study. The International Institute of Tropical Agriculture (IITA), Ibadan, Nigeria is also duly acknowledged and appreciated for providing the germplasm used for this work.

**Open Access:** This is an open access article distributed under the terms of the Creative Commons Attribution NonCommercial 4.0 International License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author(s) or sources are credited.

## REFERENCES

Adjebeng-Danquah, J., Asante, I. K., Manu-Aduening, J., Agyare, R. Y., Gracen, V. E.,

- & Offei, S. K. (2020). Genotypic variability in some morpho-physiological traits in different environments and their relationship with cassava (*Manihot esculenta* Crantz) root yield. *International Journal of Agronomy*, 1–19.
- Adjebeng-Danquah, J., Acheremu, K., Asante, S. K., Kusi, F., & Osei, C. (2012). Evaluation of early bulking cassava accessions for high yield potential for the Guinea savanna Zone of Ghana. *Ghana Journal of Agricultural Science*, 45(2), 61–70.
- Adjebeng-Danquah, J., Gracen, V. E., Offei, S. K., Asante, I. K., & Manu-Aduening, J. (2016). Genetic variability in storage root bulking of cassava genotypes under irrigation and no irrigation. *Agriculture and Food Security*, 5, 1–12.
- Adjebeng-Danquah, J., Manu-Aduening, J., Gracen, V. E., Asante, I. K., & Offei, S. K. (2017). AMMI stability analysis and estimation of genetic parameters for growth and yield components in cassava in the forest and Guinea savannah ecologies of Ghana. *International Journal of Agronomy*, 1–10.
- Adu-Gyamfi, R., Osei, C., & Anadumba, E. (2016). Yield and earliness in bulking of some introduced cassava genotypes Under moist savanna. *UDS International Journal of Development*, 3(1), 20–28.
- Adu, G. B., Akromah, R., Abdulai, M. S., Obeng-Antwi, K., Alidu, H., & Tengan, K. M. L. (2016). Trait association for improved grain yield of extra-early maturing maize hybrids evaluated in the forest and transitional zones of Ghana. *Australian Journal of Crop Science*, 10(8), 1127–1135.
- Aghogho, C. I., Eleblu, S. J. Y., Bakare, M. A., Kayondo, I. S., Asante, I., Parkes, E. Y., Kulakow, P., Offei, S. K., & Rabbi, I. (2022). Genetic variability and genotype by environment interaction of two major cassava processed products in multi-environments. *Frontiers in Plant Science*, 17, 1–17.
- Agunbiade, S. O., & Ighodaro, O. M. (2010). Variation in the physical, chemical and physico-functional properties of starches from selected cassava cultivars. *New York Science Journal*, 3(4), 48–53.
- Agyemang, I., & Abdul-korah, R. (2014). Strategies to combat desertification in Northern Region of Ghana: The role of Environmental Protection Agency. *Physical Sciences Research International*, 2(2), 35–43.
- Alua, M. A., Pehrah, K., Thomas, G., & Achana, W. (2018). Climate change implications for crop farming in Ghana's semi-arid Guinea savannah. *International Journal of Development and Sustainability*, 7(9), 2334–2349.
- Amelework, A. B., Bairu, M. W., Marx, R., Laing, M., & Venter, S. L. (2023). Genotype × environment interaction and stability analysis of selected cassava cultivars in South Africa. *Plants*, 12(13), 1–13.
- Amelework, A. B., & Bairu, M. W. (2022). Advances in genetic analysis and breeding of cassava (*Manihot esculenta* Crantz): A Review. *Plants*, 11(1617), 1–19.
- Ayetigbo, O., Latif, S., Abass, A. & Müller, J. (2018). Comparing characteristics of root, flour and starch of biofortified yellow-flesh and white-flesh cassava variants, and sustainability considerations: A review. *Sustainability*, 10(9), 1–32.
- Baafi, E. & Safo-Kantanka, O. (2008). Agronomic evaluation of some local elite and released cassava varieties in the forest and transitional ecozones of Ghana. *Asian Journal of Agricultural Research*, 2(1), 32–36.
- Bakare, M. A., Kayondo, S. I., Aghogho, C. I., Wolfe, M. D., Parkes, E. Y., Kulakow, P., Egesi, C., Rabbi, I. Y., & Jannink, J. L. (2022). Exploring genotype by environment interaction on cassava yield and yield related traits using classical statistical methods. *PLoS One*, 17(7), 1–24.
- Bose, L. K., Jambhulkar, N. N., Pande, K., & Singh, O. N. (2014). Use of AMMI and other stability statistics in the simultaneous selection of rice genotypes for yield and stability under direct-seeded conditions. *Chilean Journal of Agricultural Research*, 74(1), 3–9.
- Ceballos, H., Rojanaridpiched, C., Phumichai, C., Becerra, L. A., Kittipadukul, P., Iglesias, C. & Gracen, V. E. (2020). Excellence in cassava breeding: Perspectives for the future. *Crop Breeding, Genetics and Genomics*, 2(2), 1–31.
- Ceballos, H., Iglesias, C. A., Pérez, J. C., & Dixon, A. G. O. (2004). Cassava breeding: Opportunities and challenges. *Plant Molecular Biology*, 56, 503–516.
- Ceballos, H., Kulakow, P., & Hershey, C. (2012). Cassava Breeding: Current Status, Bottlenecks and the Potential of Biotechnology Tools. *Tropical Plant Biology*, 5, 73–87.
- Cobb, J. N., Juma, R. U., Biswas, P. S., Arbelaez, J. D., Rutkoski, J., Atlin, G., Hagen, T., Quinn, M. & Ng, E. H. (2019). Enhancing the rate of genetic gain in public-sector plant breeding programs: lessons from the breeder's equation. *Theoretical and Applied Genetics*, 132, 627–645.
- Dankwa, K. O., & Pehrah, B. B. (2019). Industrialization of cassava sector in Ghana: Progress and the role of developing of high starch cassava varieties. *Ghana Journal of Agricultural Science*, 54(2), 79–85.
- Diniz, R. P., & Oliveira, E. J. (2019). Genetic parameters, path analysis and indirect selection of agronomic traits of cassava germplasm. *Anais Da Academia Brasileira de Ciências*, 91(3), 1–11.



- Ebem, E. C., Afuape, S. O., Chukwu, S. C., & Ubi, B. E. (2021). Genotype  $\times$  environment interaction and stability analysis for root yield in sweet potato [*Ipomoea batatas* (L.) Lam]. *Frontiers in Agronomy*, 3, 1–14.
- Eke, J., Achinewhu, S. C., Sanni, L., Barimalaa, I. S., Maziya-Dixon, B. & Dixon, A. (2007). Seasonal variations in the chemical and functional properties of starches from local and improved cassava varieties in high rainfall region of Nigeria. *Journal of Food, Agriculture and Environment*, 5(3–4), 36–42.
- El-Sharkawy, M. A., & Cadavid, L. F. (2002). Response of cassava to prolonged water stress imposed at different stages of growth. *Experimental Agriculture*, 38(3), 333–350.
- El-Sharkawy, M. A. (2004). Cassava biology and physiology. *Plant Molecular Biology*, 56(4), 481–501.
- El-Sharkawy, M. A. (2007). Physiological characteristics of cassava tolerance to prolonged drought in the tropics: Implications for breeding cultivars adapted to seasonally dry and semiarid environments. *Brazilian Journal of Plant Physiology*, 19(4), 257–286.
- EPA. (2003). National action programme to combat drought and desertification. Environmental Protection Agency. Accra-Ghana. Final Report. 160 pp.
- Ewa, F., Nwofia, E., Egesi, C., Olanmi, B., & Okogbenin, E. (2017). Genetic variability, heritability and variance components of some yield and yield related traits in second backcross population (BC<sub>2</sub>) of cassava. *African Journal of Plant Science*, 11(6), 185–189.
- FAO-UNESCO. (1977). FAO-UNESCO soil map of the world, 1:5000000. Africa. *FAO Soil Bulletin*, VI(1), 299 pp.
- Farshadfar, E., Mahmodi, N., & Yaghotipoor, A. (2011). AMMI stability value and simultaneous estimation of yield and yield stability in bread wheat (*Triticum aestivum* L.). *Australian Journal of Crop Science*, 5(13), 1837–1844.
- Fukuda, W. M. G., Guevara, C. L., Kawuki, R., & Ferguson, M. E. (2010). *Selected morphological and agronomic descriptors for the characterization of cassava*. International Institute of Tropical Agriculture (IITA), Ibadan, Nigeria. 19 pp.
- Haldavankar, P. C., Joshi, G. D., Bhave, S. G., Klandekar, R. G., & Sawant, S. S. (2009). Stability of yield and yield attributing phenotypic characters in sweet potato. *Journal of Root Crops*, 35(1), 28–35.
- Hernández-Carmona, F., Morales-Matos, Y., Lambis-Miranda, H. & Pasqualino, J. (2017). Starch extraction potential from plantain peel wastes. *Journal of Environmental Chemical Engineering*, 5(5), 4980–4985.
- Hillocks, R. J. (2002). Cassava in Africa. in: Hillocks, R. J., Thresh, J. M. & Bellotti, A. (Eds.), *Cassava: utilisation* CABI Publishing: Oxon UK. pp. 41–54.
- Koopmans, A. (2005). Biomass energy demand and supply for South and South-East Asia - Assessing the resource base. *Biomass and Bioenergy*, 28(2), 133–150.
- Lenis, J. I., Calle, F., Jaramillo, G., Perez, J. C., Ceballos, H. & Cock, J. H. (2006). Leaf retention and cassava productivity. *Field Crops Research*, 95(2–3), 126–134.
- Li, M., Liu, Y., Wang, C., Yang, X., Li, D., Zhang, X., Xu, C., Zhang, Y., Li, W., & Zhao, L. (2020). Identification of traits contributing to high and stable yields in different soybean varieties across three Chinese latitudes. *Frontiers in Plant Science*, 10(1642), 1–14.
- Lopez-Diogo, L. F., Castillo, K., Vidal, M. V., Pasqualino, J., Meza-Castellar, P., & Lambis-Miranda, H. A. (2018). Evaluation of the production of starch from bitter cassava (*Manihot utilissima*) using different methodologies. *Chemical Engineering Transactions*, 65, 613–618.
- Maraphum, K., Saengprachatanarug, K., Wongpichet, S., Phuphuphud, A., Sirisomboon, P., & Posom, J. (2021). Modified specific gravity method for estimation of starch content and dry matter in cassava. *Heliyon*, 7(7), 1–10.
- Ntawuruhunga, P., & Dixon, A. (2010). Quantitative variation and interrelationship between factors influencing cassava yield. *Journal of Applied Biosciences*, 26, 1594–1602.
- Nuwamanya, E., Baguma, Y., Kawuki, R. S., & Rubaihayo, P. R. (2009). Quantification of starch physicochemical characteristics in a cassava segregating population. *African Crop Science Journal*, 16(3), 191–202.
- Okeleye, K. A., Ikeorgu, J. E. G., Melifonwu, A., Aihou, K., Maroya, N. G., Dosoo, E., Awah, E. T., Tumenteh, A., & Salau, R. A. (2001). Cassava-based cropping systems and use of inputs in different ecological zones of West and Central Africa. *African Journal of Root Crops*, 4(2), 13–17.
- Okogbenin, E., Setter, T. L., Ferguson, M., Mutegi, R., Ceballos, H., Olanmi, B., & Fregene, M. (2013). Phenotypic approaches to drought in cassava: Review. *Frontiers in Physiology*, 4(93), 1–15.
- Oliveira, C. R. S., Borel, J. C., Pereira, D. A., Carvalho, B. P., Medrado, E. S., Ishikawa, F. H., & Oliveira, E. J. (2021). Genetic parameters and path analysis for root yield of cassava under drought and early harvest. *Crop Breeding and Applied Biotechnology*, 21(3), 1–8.
- Oliveira, E. J., Aidar, S. T., Morgante, C. V., Chaves, A. R. M., Cruz, J. L., & Coelho Filho, M. A. (2015). Genetic parameters for drought-tolerance in cassava. *Pesquisa Agropecuária Brasileira*, 50(3), 233–241.
- Olivoto, T., Lúcio, A. D., Silva, J. A., Marchioro, V. S., Souza, V. Q., & Jost, E. (2019). Mean performance and stability in multi-environment trials I: Combining features of AMMI and BLUP techniques. *Agronomy Journal*, 111, 2949–2960.
- Omodamiro, R. M., Iwe, M. O., & Ukpabi, U. J. (2007). Pasting and functional properties of lafun and starch processed from some improved cassava genotypes in Nigeria. *Nigerian Food Journal*, 25(2), 122–129.
- Osei, C., Bagamsah, S. K., Asante, S. K., Alhassan, A. Y., Acheremu, K., Dei, D., Sallah, Y., Wilson, K., Eledi, E., Wawula, M., Abukari, S., Kakraba, M., Dixon, A. G. O., Ilona, P., Asiedu, R., & Ofori, F. (2002). Evaluation and selection of IITA-Improved cassava genotypes for cultivation in semi-arid Guinea savannah zone of Ghana. A report presented to the National Variety Release and Registration Committee of Ghana. 33 pp.
- Osei, M. K., Annor, B., Adjebeng-Danquah, J., Danquah, A., Danquah, E., Blay, E., & Adu-Dapaah, H. (2018). Genotype  $\times$  environment interaction: A prerequisite for tomato variety development. in: Nyaku S.T. and Danquah A. (Eds.), *Recent advances in tomato breeding and production*. Intechopen Limited, London, UK. pp. 93–113.
- Payne, R. W., Murray, D. A., Harding, S. A., Baird, D. B., & Soutar, D. M. (2009). *Genstat for Windows (12<sup>th</sup> Edition) Introduction*. VSN International, Hemel Hempstead. VSN International, Hemel Hempstead.
- Prasannakumari, V., Nair, A. G. H. & Mohan, C. (2021). Identification of quantitative trait loci (QTLs) conferring dry matter content and starch content in cassava (*Manihot esculenta* Crantz). *American Journal of BioScience*, 9, 1–9.
- Purchase, J. L., Hatting, H., & van Deventer, C. S. (2000). Genotype  $\times$  environment interaction of winter wheat (*Triticum aestivum* L.) in South Africa: II. Stability analysis of yield performance. *South African Journal of Plant and Soil*, 17(3), 101–107.
- Rao, B. B., Swami, D. V., Ashok, P., Kalyana Babu, B., Ramajayam, D., & Sasikala, K. (2017). Correlation and path coefficient analysis of cassava (*Manihot esculenta* Crantz) genotypes. *International Journal of Current Microbiology and Applied Sciences*, 6(9), 549–557.
- Salgotra, R. K., & Chauhan, B. S. (2023). Genetic diversity, conservation, and utilization of plant genetic resources. *Genes*, 14, 1–20.
- Silva, P. P., Sousa, M. B., & Oliveira, E. J. (2019). Prediction models and selection of agronomic and physiological traits for tolerance to water deficit in cassava. *Euphytica*, 215(4), 1–18.
- Swarup, S., Cargill, E. J., Crosby, K., Flagel, L., Kniskern, J., & Glenn, K. C. (2021). Genetic diversity is indispensable for plant breeding to improve crops. *Crop Science*, 61(2), 839–852.
- Teye, E., Asare, P. A., Amoah, R. S., & Tetteh, J. P. (2011). Determination of the dry matter content of cassava (*Manihot esculenta* Crantz) tubers using specific gravity method. *APRN Journal of Agricultural and Biological Science*, 6(11), 23–28.
- Tuberosa, R. (2012). Phenotyping for drought tolerance of crops in the genomics era. *Frontiers in Physiology*, 3(347), 1–26.
- Tumuhimbise, R., Melis, R., Shanahan, P., & Kawuki, R. (2014). Genotype  $\times$  environment interaction effects on early fresh storage root yield and related traits in cassava. *Crop Journal*, 2(5), 329–337.
- Tumwesigye, K. S., Sousa, A. R., Oliveira, J. C., & Sousa-Gallagher, M. J. (2017). Evaluation of novel bitter cassava film for equilibrium modified atmosphere packaging of cherry tomatoes. *Food Packaging and Shelf Life*, 13, 1–14.
- Uchendu, K., Njoku, D.N., Ikeogu, U.N., Dziridziyonyo, D., Tongoona, P., Offei, S., & Egesi, C. (2022). Genotype-by-environment interaction and stability of root mealiness and other organoleptic properties of boiled cassava roots. *Scientific Reports*, 12, 1–11.
- Varshney, R. K., Bohra, A., Yu, J., Graner, A., Zhang, Q., & Sorrells, M. E. (2021). Designing future crops: genomics-assisted breeding comes of age. *Trends in Plant Science*, 26(6), 631–649.