

Feasibility of using reflectance spectroscopy for the analysis of bio-element concentrations in Antarctic ornithogenic sediments

Liu Xiaodong(刘晓东)¹, Sun Jing(孙静)², Sun Liguang(孙立广)¹, Liu Wenqi(刘文齐)³ and Wang Yuhong(王玉红)⁴

1 Institute of Polar Environment, University of Science and Technology of China, Hefei 230026, China

2 Department of Earth Sciences, Dartmouth College, Hanover, NH 03755, USA

3 Instruments' Center for Physical Science, University of Science and Technology of China, Hefei 230026, China

4 National Institutes of Health, Bethesda, MD, 20892, USA

Received October 28, 2009

Abstract Compared with traditional chemical analysis methods, reflectance spectroscopy has the advantages of speed, minimal or no sample preparation, non-destruction, and low cost. The present study explored the application of the reflectance spectroscopy within near ultraviolet-visible-near infrared region to predict bio-element compositions in the ornithogenic sediments from the maritime Antarctic. A total of 106 samples were taken from four ornithogenic sediment cores on the Ardley Island of Antarctica, 68 samples were used for building calibration equation, and 38 for prediction of nine bio-elements including P, Ca, Cu, Zn, Se, Sr, Ba, F and S. Three multivariate statistical analysis techniques, including stepwise multiple linear regression (Stepwise-MLR), principal component regression (PCR) and partial least squares regression (PLS) were used to develop mathematical relationships between the spectral data and the chemical reference data. The results showed that the regression models constructed by PCR and PLS models have no significant differences, and obviously superior to Stepwise-MLR. The correlations between spectra-predicted and chemically analyzed concentrations of nine bio-elements are statistically significant, and the concentration-versus-depth profiles predicted from reflectance spectra using PLS calibration model are consistent with those from actual chemical analysis. These results demonstrated the feasibility of using reflectance spectroscopy to infer bio-element concentrations in the ornithogenic sediments, and thus it is suggested that the reflectance spectroscopy could provide a rapid and valuable technique to indirectly identify whether the sediments were influenced by penguin droppings in the Antarctic region.

Key words Reflectance spectroscopy, Ornithogenic sediments, Multivariate statistical analysis, Bio-element, Antarctic.

1 Introduction

Reflectance spectroscopy, a non-destructive technique, has been considered as an alternative or complementary method for compositional analysis of various chemical constituents in soil, sediment and biological samples, and for assessment of environmental quality^[1-12]. Recently, there is a growing interest in using reflectance spectroscopy for palaeolimnological and palaeoenvironmental studies, since this method offers several special advantages such as speed of determination, minimal or no preparation of samples, non-destructive analysis, no consumption of reagents, and low costs of analysis^[13-22].

Wavelength position and height of the spectral peaks from reflectance spectroscopy cannot directly be used for determinations of chemical compositions, because the spectra are too complex and the peaks in the near infrared region are broad and overlapping. Instead, the spectra have to be calibrated using samples in which the constituents of interest are known from reference methods. Useful information from the spectra can be extracted using chemometric techniques such as stepwise multiple linear regression (Stepwise-MLR) or other multivariate calibration methods such as principal component regression (PCR), and partial least squares (PLS) regression^[23].

In recent years, numerous studies showed that the remnants of ancient penguin droppings in the lake sediments near penguin colonies may be identified by their geochemical characteristics, which may provide continuous information about historical population changes of penguin^[24-29]. Nine elements including sulfur (S), phosphorus (as P_2O_5), calcium (as CaO), copper (Cu), zinc (Zn), selenium (Se), strontium (Sr), barium (Ba) and fluorine (F) have been found to be enriched and significantly correlated with each other in the sediments amended by penguin guanos, and their assemblage is identified to be an important geochemical signal for indication of impact from penguin droppings or guano soils in Antarctica^{[24][30]}. Besides these elemental proxies, isotopic and organic geochemical proxies have also been successfully used to reconstruct the long-term variations in historical penguin populations^[31-33]. Since these multi-proxies are usually not determined on the same aliquot and the samples are taken from remote Antarctic areas, there is a common trouble of insufficient amount of samples for chemical analyses in the laboratories. Therefore, there is a need for faster and more efficient and economical methods to facilitate reconstruction of palaeorecords in the Antarctic ornithogenic sediments and to improve the understanding of the effects of climate change on penguin ecology.

According to the issued reports, reflectance spectroscopy has rarely been used for scientific research in maritime Antarctic. In this paper, four sediment cores influenced by penguin droppings on the Ardley Island were selected and the corresponding reflectance spectroscopy signals were analyzed with the main objective to examine the feasibility of using reflectance spectroscopy within near ultraviolet-visible-near infrared region to determine nine bio-elements concentrations in the sediments. If this can be done successfully, reflectance spectroscopy will allow numerous chemical constituents in core sections to be determined rapidly and simultaneously, and thus greatly reduce the cost for certain chemical analyses. Thus, in the future this approach may provide a rapid technique to identify whether the sediments are impacted by penguin

droppings in the Antarctica, and further to reconstruct the palaeoecological evolution recorded in the Antarctic ornithogenic sediments.

2 Study Area

The Ardley Island (62°13'S, 58°56'W), a 2 km long and 1.5 km wide island, is about 500 m to the east of Fildes Peninsula, Maxwell Bay, King George Island and connected with the Fildes Peninsula by a sandy dam (Fig. 1). The Great Wall Station of China is located about 0.5 km to the west of the island. The study area has a cold oceanic climate, characteristic of maritime Antarctica. According to the meteorological records from the Great Wall Station, the mean annual precipitation is about 630 mm, the annual average relative humidity is about 90%, and the mean annual air temperature is around $-2.6\text{ }^{\circ}\text{C}$ with a winter low at $-26.6\text{ }^{\circ}\text{C}$, and summer high at $11.7\text{ }^{\circ}\text{C}$. It is free of snow and ice during the summer. Geologically, the island mainly consists of tertiary andesitic and basaltic lavas and tuffs together with raised beach terraces. The topography of the island is relatively flat with the highest elevation of 70 m. Seventy-eighty percent of the island is covered by vegetation, predominantly consisting of mosses and lichens.

Ardley Island is one of the most important penguin colonies in the maritime Antarctic region. In 1991, this island has been declared as Antarctic Specially Protected Area by the Protocol on Environmental Protection to the Antarctic Treaty. During the breeding period every summer, the number of penguins on this island was about 10218; the major species were Gentoo (*Pygoscelis papua*, 74%), Adelie (*Pygoscelis adeliae*, 21%) and Chinstrap (*Pygoscelis antarctica*, 5%)^[31,34]. It is estimated that penguins on the Ardley Island discharge about 139 t droppings based on the hypothesis that each penguin excretes 84.5 g droppings (dry weight) a day during the breeding period^[27,30]. Droppings are transferred and deposited in the lakes or depressions by ice or snowmelt water, and those ancient penguin waste products preserved in the sediments record the historical information of penguin population change^[24].

3 Materials and Methods

3.1 Sample collection and chemical data

A total of 106 samples were collected from four sediment cores (Y2, Y2-4, Y4 and AD3) on the Ardley Island for spectral and chemical analyses. For comparison, 5 soil samples (N1) and 3 pure guano samples (AP) were analyzed for spectral characteristics. The sampling sites of four sediment cores are shown in Figure 1. The sediment cores Y2 and Y2-4 are collected from the same lake Y2. The lithological descriptions of these sediment cores are given in the references^[24,28,30], in which studies have showed that all the sediment cores are influenced by penguin droppings. In the laboratory, the sediment cores were sectioned at the intervals of 1-2 cm, and the sub-samples were air-dried, sieved and then ground to powder for chemical analysis. The chemical methods, as well as the concentrations of nine bio-elements including S, P,

Ca, Cu, Zn, Se, Sr, Ba and F in the sediment subsamples, are reported in detail in the references mentioned above. The soil samples(N1) were affected neither by penguin droppings nor by biological action, and they are natural products of local bed-rock weathering^[35].

3.2 *Spectral measurement and transformation*

Prior to spectral analysis, the collected sediment subsamples were ground in a mortar

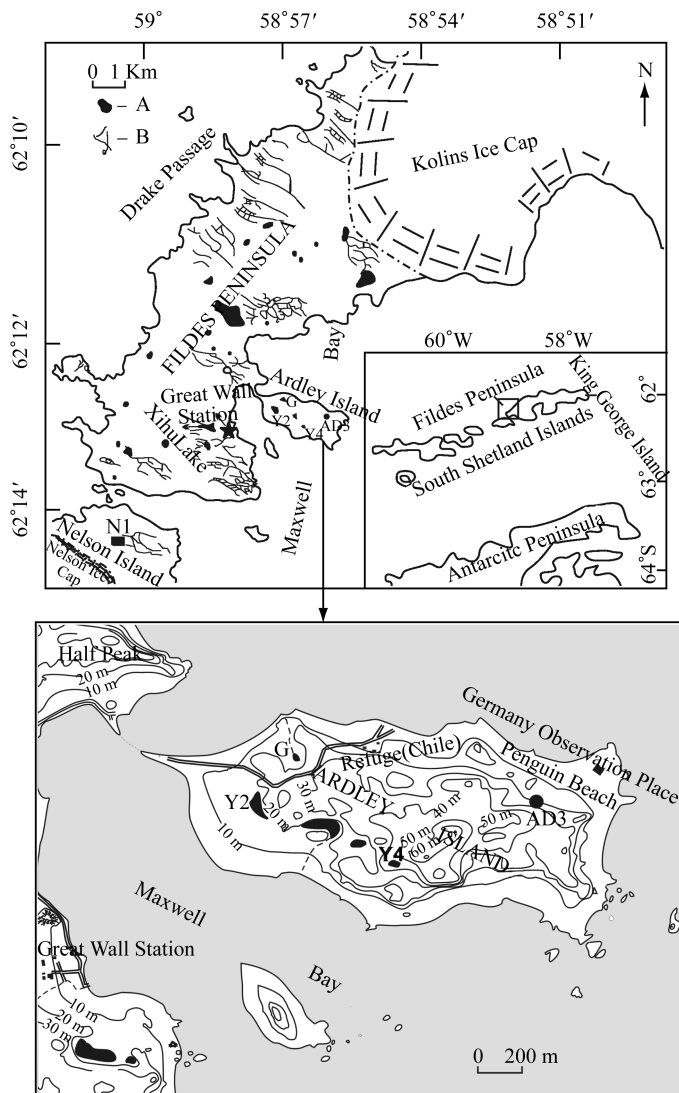


Fig. 1 Studied area and sampling sites (Y2, Y4 and AD3). In the top panel, marker A shows lakes, marker B shows network of meltwater channels. In the bottom panel, the contour interval is 10 meter (From Liu *et al.*, 2005).

to pass a 0.074 mm sieve, and then dried at 105 °C for 2 hours. For each sample, approximately 1 g powder was packed into a measuring cell, and subsequently the spectral reflectance was recorded in Shimadzu SolidSpec-3700 UV-VIS-NIR Recording Spectrophotometer, configured with UV-Probe software. All the samples were scanned over a wavelength range of 200-2600 nm covering the near ultraviolet-visible-near infrared region at 1 nm interval. A total of 2401 data points were collected for each spectrum and it took less than 10 minutes to complete a scan. Samples were read against an external polyethylene (zero absorbance) reference, which was read alternately with the samples. The reference spectrum was automatically subtracted from each sample spectrum and the resulting spectrum automatically recorded. To

establish a robust prediction model, several spectral pretreatments were applied to the reflectance spectra to optimize the accuracy. Using UV-Probe software, the original reflectance (r) data could be transformed to first (r') or second (r'') derivative, reciprocal ($1/r$) and absorbance ($\log 1/r$) data.

3.3 Model construction and validation

To establish and test mathematical relationships between spectral data and the corresponding chemical reference data, two sets of samples were created firstly: calibration set and prediction set. For each chemical constituent, stepwise multiple linear regression, principal component regression and partial least squares regression methods were used to find and test the mathematical relationship, then their calibration results were compared.

The first method is stepwise multiple linear regression (Stepwise-MLR). Considering that the reflect peaks (or absorb valleys) are the most significant (see below), the reflect peaks of each spectra curve and removed the overlapping ones were picked out. A total of 121 peaks were selected to perform the regression analyses and establish the mathematical relationship between these independent variables with reference chemical data, the dependent variables. Five mathematical regression equations using the independent variable of the raw optical data (r) and four derived data (r' , r'' , $1/r$, $\log(1/r)$) and the calibration data set were computed by the Stepwise-MLR option of MATLAB 7.1; r' and r'' represent the first and second order derivatives of r . These computations generated a number of statistical parameters such as R^2 (square of the regression coefficient) and RMSE (root mean squared error of calibration) ^[18]. The regression equations from the calibration data set were used to predict the relevant chemical constituents of the prediction set. The predicted values were compared with the measured reference values using several statistics, such as R , the coefficient of linear correlation, and SEP, the standard error of prediction (the standard deviation of the points about the 1 : 1 line). For each constituent, the regression equation yielding the highest R and the lowest SEP was accepted as the best ones. The regression equation was further evaluated by two additional statistical quality parameters: the residual prediction deviation (RPD) and ratio error range (RER). The RPD is the ratio of the standard deviation (SD) of the reference chemistry values for the prediction set over the SEP. The RER is the ratio of the range of the reference chemistry values in the prediction set over the SEP. In agricultural applications, RPD values >5 are desirable, >3 are acceptable, and between 2.5–3 are suitable for screening; RER should be greater than 10^[16,17].

The second method, principal component regression (PCR) is a two-stage process. First, it minimizes the number of independent components required to describe the variations over the entire spectrum and between spectra. This technique enables a total of 2401 spectral points to be reduced to a few principal components (PCs), where the PCs describe the spectral variance over all the samples. Second, these PCs are regressed against known property data (measured concentration), and then regression models are constructed. In this study, the method of singular value

decomposition (SVD) in MATLAB 7.1 were used and the method of “validation set prediction” was chosen to evaluate the precision of regression^[36]. Regression models were further evaluated by the statistic parameters mentioned above.

The third method, partial least squares regression (PLS), is widely used for multivariate calibration and has been applied in various applications for chemical analysis. PLS decomposes spectral data (X) and sediment property data (Y) into a new smaller set of latent variables and their scores that best describe all the variance in the data^[3]. PLS was performed using ParLeS software version 3.1, which is specially designed for chemometric analysis of spectroscopic data^[37]. In the present study the same optimal spectral indices was used as PCR. Following the procedure of Malley *et al.* (1999)^[16], cross validation, also known as ‘jackknifing’ or ‘leave-one-out’, was used to pick out the best number of factors for regression. The PLS can be performed to maximize the size of the calibration data set and to predict all of the samples. It used the mathematical treatment and number of PLS factors from the previous steps to develop calibrations. The procedure was repeated 68 times until each sample is predicted. Regression models were further evaluated by the statistics mentioned above (R, SEP, RPD and RER) to ensure they are robust enough for predicting concentration data of nine bio-elements from the reflectance spectral.

4 Results and discussion

4.1 Distributions of elements and spectral characteristics

Reflectance spectra of representative samples from the four ornithogenic sediment cores are given in Figure 2. It can be seen from this figure that the shape of the spectra curves is almost identical for all four samples, but the reflectance values are notably different. All the ornithogenic sediments have high reflectance intensity in the near-infrared light region (800–2500 nm). The ornithogenic sediments are predominately composed of the weathering products from bedrocks and the penguin guano, and the guano contains much more organic matter than the weathering products, thus the input of penguin guano could significantly influence the content of organic matter in the sediments^[26,32]. Numerous studies have shown that the attributes of organic materials are important for NIR spectral feature^[18,20]. It is assumed that the penguin guano enriched with organic matter is the dominant influencing factor on the reflectance spectra of these ornithogenic sediments.

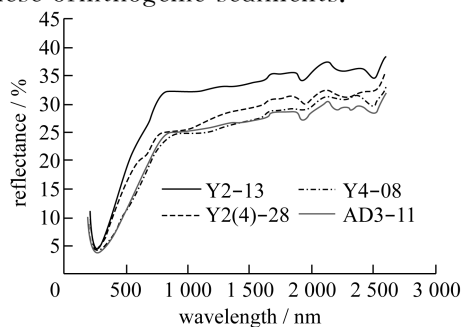


Fig. 2 Reflectance spectra of representative samples from the four sediment cores.

Some trace elements in sediments and soils generally do not respond to the incident light directly, but they may be detectable in the spectrum signatures because they can form complex with organic matter, and be associated with moieties such as hydroxides, sulfides, carbonates or oxides that are detectable, and be absorbed to clays that absorb light in some wavelength range^[2,8,20]. Sun *et al.* (2000, 2001, 2004)^[24,25,30] reported that nine inorganic elements including S, P, Ca, Cu, Zn, Se, Sr, Ba and F were enriched and significantly correlated with each other in the soils or sediments amended by penguin guanos. The assemblage of P, Sr, Cu, Zn, Se, Ca, F, Ba and S is suggested to be an important geochemical marker for penguin droppings or the soils and sediments impacted by them in the maritime Antarctic. Thus, it can be reasonably inferred that the reflecting intensity of the ornithogenic sediments is related to the concentrations of the nine bio-elements.

In this study, the spectra data that represent the same data sets for all the nine bio-elements were chosen. Using the same procedure for chemical data processing, spectra are sorted from lowest to highest reflectance values and divided into two groups. Two-thirds of the spectra were used as a calibration set, and the remaining as a predication set. Each set represented approximately the full range of concentrations. Two-thirds of the spectra were used as calibration set, the rest as predication set, and they consist of 68 (except for element Ba, n=67) and 38 samples, respectively. Chemically determined concentrations of nine bio-elements (P_2O_5 , CaO, Cu, F, Ba, S, Zn, Sr, and Se) in both sets are given in Table 1, and they are of large variations, indicating variable influence of penguin droppings on the sediments^[24,28]. The values of standard deviation (SD) and range are used to compute the statistic of RPD and RER in the model calibration, respectively.

Table 1. Chemically determined concentrations of nine bio-elements in the calibration and prediction sets

| Element | Units(g/g) | set | min | max | mean | range | SD | n |
|----------|------------|------|------|--------|--------|--------|--------|----|
| P_2O_5 | 10^{-2} | Cal | 0.74 | 15.52 | 6.372 | 14.78 | 3.688 | 68 |
| | | Pred | 0.64 | 13.45 | 6.472 | 12.81 | 4.339 | 38 |
| CaO | 10^{-4} | Cal | 5.23 | 19.80 | 9.931 | 14.57 | 3.453 | 68 |
| | | Pred | 5.18 | 18.35 | 10.304 | 13.17 | 4.455 | 38 |
| Cu | 10^{-4} | Cal | 0.62 | 9.95 | 3.600 | 9.33 | 2.105 | 68 |
| | | Pred | 0.65 | 7.65 | 3.619 | 7.00 | 2.335 | 38 |
| F | 10^{-4} | Cal | 3.92 | 123.90 | 40.200 | 119.58 | 27.690 | 68 |
| | | Pred | 3.21 | 83.13 | 38.037 | 79.92 | 28.501 | 38 |
| Ba | 10^{-4} | Cal | 0.93 | 2.54 | 1.635 | 1.61 | 0.474 | 67 |
| | | Pred | 0.95 | 2.42 | 1.509 | 1.48 | 0.404 | 38 |
| S | 10^{-4} | Cal | 1.40 | 178.00 | 38.990 | 176.60 | 42.520 | 68 |
| | | Pred | 1.30 | 18.40 | 52.550 | 182.70 | 57.360 | 38 |
| Zn | 10^{-4} | Cal | 0.52 | 8.35 | 3.424 | 7.83 | 2.010 | 68 |
| | | Pred | 0.35 | 7.52 | 3.447 | 7.17 | 2.367 | 38 |
| Sr | 10^{-4} | Cal | 3.69 | 19.30 | 9.191 | 15.61 | 4.779 | 68 |

| | | | | | | | | |
|----|-----------|------|------|-------|--------|-------|--------|----|
| | | Pred | 3.84 | 18.46 | 9.163 | 14.62 | 5.422 | 38 |
| Se | 10^{-6} | Cal | 1.44 | 54.90 | 17.823 | 53.46 | 14.905 | 68 |
| | | Pred | 1.69 | 49.70 | 18.716 | 48.01 | 17.772 | 38 |

4.2 Construction of calibration models

Table 2 summarizes the results of optimal Stepwise-MLR model and statistical parameters. As shown in this table, the prediction quality strongly depends on data preprocessing. For example, the best results were achieved using first order derivative (r') for Sr, Cu and Se, reciprocal ($1/r$) for S and Zn, and original spectra (r) for P_2O_5 and Ba. The optimal models achieved high precisions of regression and validation for each bio-element, and all the coefficients of regression calibrations, R^2 , are close to or exceed 0.95. The correlation coefficients, R , between the reflectance spectrum predicated and chemically determined concentrations is higher than 0.95, except for Ba ($R=0.894$). RPD are >2.5 for most elements, and RER are >10 for P, Ca, Cu, F, S and Zn. The wavelengths upon which the Stepwise-MLR calibrations are based are listed in Table 2, they indicate the regions of the spectrum used in the prediction, and are different for each element. This result is consistent with the observations by Malley *et al.* (1999)^[16] and Malley and Williams (1997)^[2]. Generally, the results of model reconstruction by the Stepwise-MLR calibration are satisfactory for most bio-elements, indicating that the spectral method has potential to predict rapidly the concentrations of nine bio-elements in the ornithogenic sediments in the maritime Antarctic.

Table 2. Prediction accuracy, mathematical treatments model, and used wavelengths for the determination of nine bio-element concentrations by stepwise-MLR

| Element | P_2O_5 | CaO | Cu | F | Ba | S | Zn | Sr | Se |
|---------------|----------|---------|--------|---------|-------|--------|--------|-------|-------|
| R^2 | 0.983 | 0.963 | 0.973 | 0.981 | 0.947 | 0.955 | 0.961 | 0.933 | 0.948 |
| RMSE | 0.523 | 0.713 | 0.375 | 4.060 | 0.121 | 9.424 | 0.415 | 1.333 | 3.583 |
| R | 0.962 | 0.972 | 0.971 | 0.963 | 0.894 | 0.976 | 0.969 | 0.951 | 0.959 |
| SEP | 1.179 | 1.059 | 0.578 | 7.686 | 0.192 | 11.902 | 0.588 | 1.630 | 5.611 |
| RPD | 3.681 | 4.209 | 4.043 | 3.708 | 2.107 | 4.819 | 4.026 | 3.327 | 3.167 |
| RER | 10.868 | 12.442 | 12.120 | 10.398 | 7.692 | 15.351 | 12.194 | 8.970 | 8.556 |
| Model | r | log 1/r | r' | log 1/r | r | 1/r | 1/r | r' | r' |
| Wavelength/nm | 844 | 669 | 669 | 654 | 260 | 654 | 654 | 844 | 844 |
| | 876 | 1720 | 844 | 844 | 669 | 669 | 669 | 1407 | 866 |
| | 1730 | 1916 | 929 | 1021 | 844 | 844 | 1720 | 1715 | 1400 |
| | 1741 | 1934 | 1400 | 1916 | 1021 | 859 | 1838 | 1724 | 2250 |
| | 1855 | 2108 | 1709 | 1939 | 1916 | 1846 | 2409 | 2213 | 2346 |
| | 1922 | 2132 | 1720 | 2209 | 2102 | 2144 | 2491 | 2230 | 2351 |
| | 1939 | 2213 | 2102 | 2400 | 2235 | | | 2346 | 2504 |
| | 2209 | 2480 | 2292 | 2495 | 2240 | | | 2367 | |
| | 2400 | 2487 | 2351 | 2510 | 2245 | | | 2427 | |
| | | | 2510 | | 2250 | | | | |

2363

2395

The statistical parameters of the final models using the PCR method are summarized in Table 3. Calibrations developed by PCR for all bio-elements produced good relationships with $R_s > 0.92$ and $RPDs > 2.5$. Most RER values are beyond 10 except for Ba. Compared with the results of Stepwise-MLR calibrations (Table 2), PCR calibrations have the higher correlation coefficients R_s , lower SEPs, larger RPDs and RERs for the same inorganic element. Thus, PCR calibrations utilizing the whole spectrum data (2401 spectral points) were clearly superior to Stepwise-MLR characteristic of segmental spectra analyses.

Table 3. Regression statistics and parameter estimates for the determination of nine bio-element concentrations by PCR method

| element | P ₂ O ₅ | CaO | Cu | F | Ba | S | Zn | Sr | Se |
|---------|-------------------------------|------------------|------------------|------------------|------------------|------------------|------------------|------------------|------------------|
| units | 10 ⁻² | 10 ⁻² | 10 ⁻⁴ | 10 ⁻⁴ | 10 ⁻⁴ | 10 ⁻⁴ | 10 ⁻⁴ | 10 ⁻⁴ | 10 ⁻⁶ |
| Model | r | log 1/r | r' | log 1/r | r | 1/r | 1/r | r' | r' |
| R | 0.978 | 0.983 | 0.977 | 0.975 | 0.924 | 0.984 | 0.987 | 0.979 | 0.963 |
| SEP | 0.890 | 0.862 | 0.496 | 6.105 | 0.161 | 9.875 | 0.374 | 1.094 | 4.668 |
| RPD | 4.876 | 5.171 | 4.710 | 4.668 | 2.513 | 5.809 | 6.322 | 4.955 | 3.807 |
| RER | 14.395 | 15.287 | 14.119 | 13.090 | 9.176 | 18.501 | 19.150 | 13.360 | 10.284 |

Table 4 summarizes the statistic parameters for PLS calibration. The predicted concentrations by PLS for P₂O₅, CaO, Cu, F, S, Zn and Sr have $R_s > 0.97$, $RPDs > 4.0$ and $RERs > 12$, indicative of good calibrations. The calibration for Se gives R of 0.956, RPD of 3.472 and RER of 9.381. The poorest calibration is for Ba, it gives a high correlation coefficient R of 0.914, but both the RPD at 2.421 and RER at 8.842 are lower than desirable. Compared with the results of Stepwise-MLR calibrations (Table 2), PLS calibrations have higher correlation coefficients R_s , lower SEPs, larger RPDs and RERs. Thus, PLS calibrations utilizing whole spectrum data (2401 spectral points) are also clearly superior to Stepwise-MLR calibration method. Compared with the result of PCR calibrations, it is found that the differences among the statistics R, SEP, RPD and RER are very small. Due to the overlapping of absorption peaks and the collinearity of the absorbance at different wavelengths, methods using single wavelengths such as Stepwise-MLR are usually less successful than principal component regression (PCR) or partial least squares regression (PLS), which utilize the whole spectrum to extract the spectral fingerprint of a chemical compound. Therefore, it is suggested that the reflectance spectral method utilizing mathematical treatment models PCR and PLS has the potential to rapidly predict the concentrations of nine bio-elements in the ornithogenic sediments of the maritime Antarctic.

Table 4. Mathematical treatment model, optimal number of factors, and prediction statistics for the determination of nine bio-element concentrations by PLS

| Element | P ₂ O ₅ | CaO | Cu | F | Ba | S | Zn | Sr | Se |
|---------|-------------------------------|-----|----|---|----|---|----|----|----|
|---------|-------------------------------|-----|----|---|----|---|----|----|----|

| Model | r | log 1/r | r' | log 1/r | r | 1/r | 1/r | r' | r' |
|---------------|--------|---------|--------|---------|-------|--------|-------|--------|-------|
| No of factors | 17 | 16 | 7 | 14 | 9 | 10 | 10 | 8 | 8 |
| R | 0.976 | 0.973 | 0.972 | 0.975 | 0.914 | 0.982 | 0.987 | 0.977 | 0.956 |
| SEP | 0.953 | 1.035 | 0.542 | 6.196 | 0.167 | 10.363 | 0.386 | 1.132 | 5.118 |
| RPD | 4.555 | 4.302 | 4.305 | 4.600 | 2.421 | 5.535 | 6.127 | 4.790 | 3.472 |
| RER | 13.447 | 12.719 | 12.905 | 12.899 | 8.842 | 17.631 | 18.56 | 12.916 | 9.381 |

Figure 3 gives loadings of the first factors of reflectance spectral data from PLS vs. wavelength for nine bio-elements. The loadings indicate wavelength regions where variance was used in computing the PLS calibration equations. The results showed that the first PLS factor for all nine bio-elements account for the vast majority of the variance in their concentrations (93%, 87%, 88%, 96%, 87%, 82%, 59%, 90%, 73% for Ba, Zn, P₂O₅, CaO, Cu, F, S, Sr and Se, respectively) and the loadings are virtually identical (Fig. 3). This indicates that the organic matter from the penguin droppings responsible for the variability in the bio-element levels is of the same quality for all bio-elements^[2]. Furthermore, for all nine bio-elements, the loading versus wavelength profiles of the first factor are very consistent with those curves of reflectance spectrum of the sediment samples (Fig. 2), and this confirms that the reflectance spectrum intensity of the ornithogenic sediments is predominantly controlled by the concentration changes of nine bio-elements, which depend on the amount of penguin dropping input.

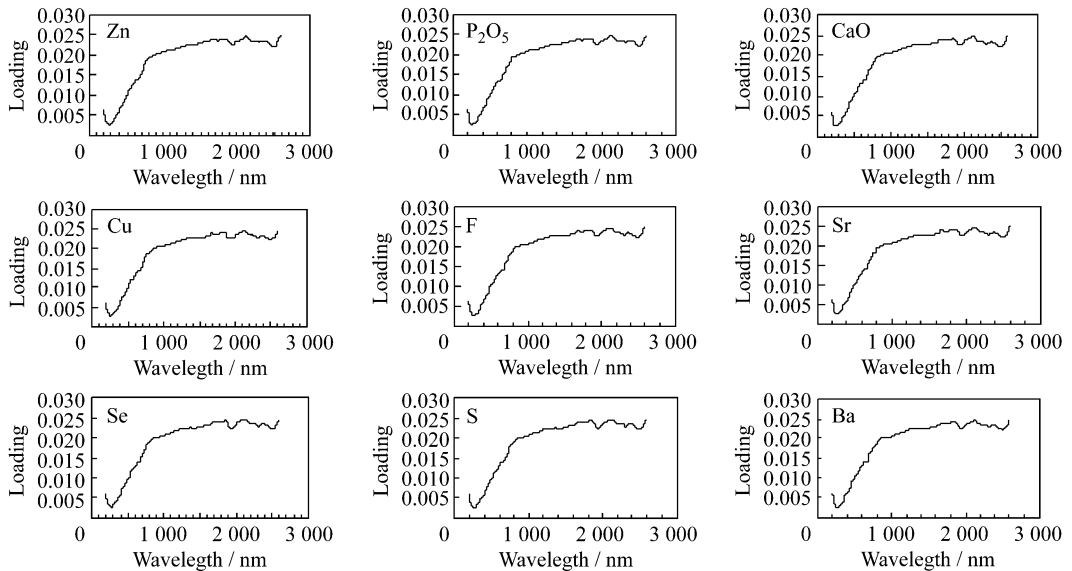


Fig. 3 Loadings of the first factor of reflectance spectral data from PLS vs. wavelength for nine bio-elements.

4.3 Comparison of NIR-predicated and reference bio-element concentrations

As suggested previously, PLS has advantages in reducing noise, detecting the compositions of unknown samples that are not represented by the calibration model,

and obviating the need for wavelength selection; and it tends to improve model stability and increase prediction accuracy^[3]. Therefore, up to now PLS is widely used for multivariate calibration and chemical analysis. In present, the calibration equations constructed by PLS approach were used to predict chemical constituents of each sample for the cores Y2 and Y4, and then compare NIR-predicated and reference core profiles.

The reference concentration data of nine bio-elements in the ornithogenic sediments and those predicated by PLS on UV-VIS-NIR are compared in Figure 4. As shown in this figure, the slopes of the measured vs. predicated regression lines for all the nine bio-elements are not significantly different from 1, indicating that this technique does not over or underestimate the chemical properties of ornithogenic sediments.

To visually compare the concentration-versus-depth profiles determined by reference chemistry and reflectance spectral methods, the concentrations of nine bio-elements in all the subsamples from the sediment cores Y2 and Y4 were computed using the PLS calibration

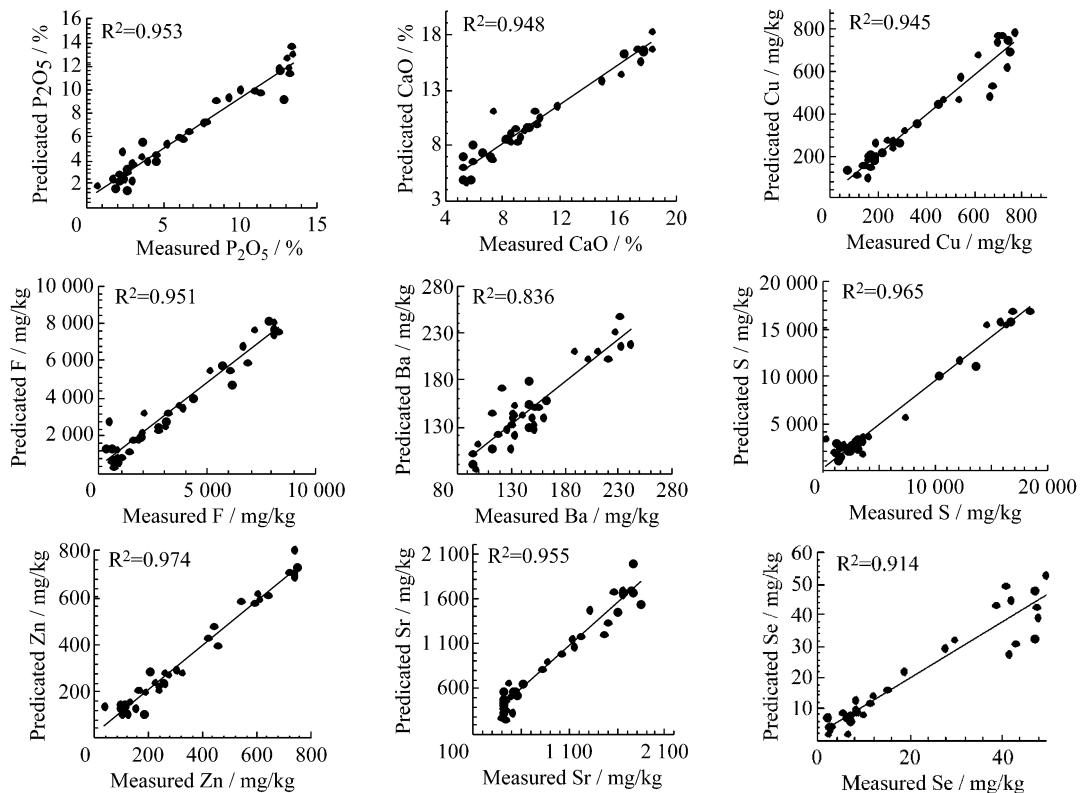


Fig. 4 Correlations between the concentrations determined by PLS and reference chemistry of nine bio-elements.

model. The measured and inferred values for Y2 and Y4 sediment cores are plotted versus depth in Figure 5 and Figure 6, respectively. As shown in these two figures,

with the exception of Ba in Y4 (Fig. 6), which has a relatively low correlation coefficient of 0.35 ($p > 0.01$), all the reflectance spectrum-predicated profiles of nine bio-elements are significantly correlated with those by chemistry reference ($p < 0.01$). This implies that the overall trends in bio-element concentrations in the ornithogenic sediment cores on the Ardley Island can be reconstructed from reflectance spectrum without chemical measurements. For Ba in the core Y4, the marked difference is likely due to the fact that Ba in the Y4 lacustrine sediments is associated with lithogenic elements, not bio-source ones^[28].

5 Conclusions

In this paper, four Antarctic ornithogenic sediment cores were taken as study samples, and measured by both chemical and spectral methods. Based on the Stepwise-MLR, PCR and PLS approaches, mathematical relationships between the spectral and the chemical reference data were developed. The correlation structure reflects clearly the prediction results:

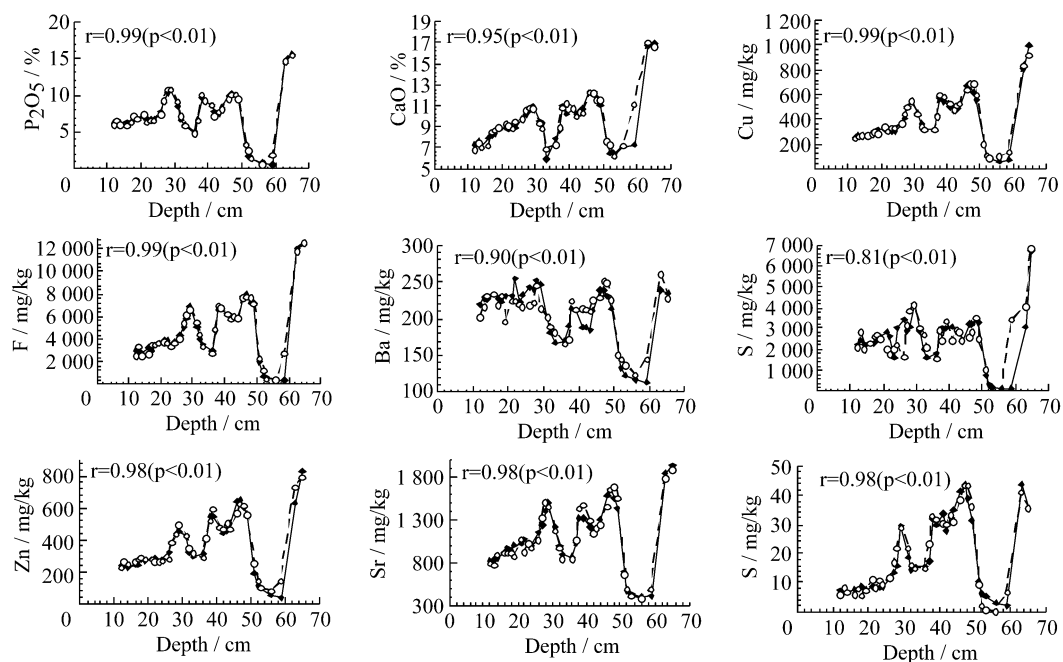


Fig. 5 Comparison of concentration versus depth profiles determined by reference chemistry (solid dot line) and PLS-predicted by reflectance spectroscopy (hollow dot line) for nine bio-elements in the Y2 core.



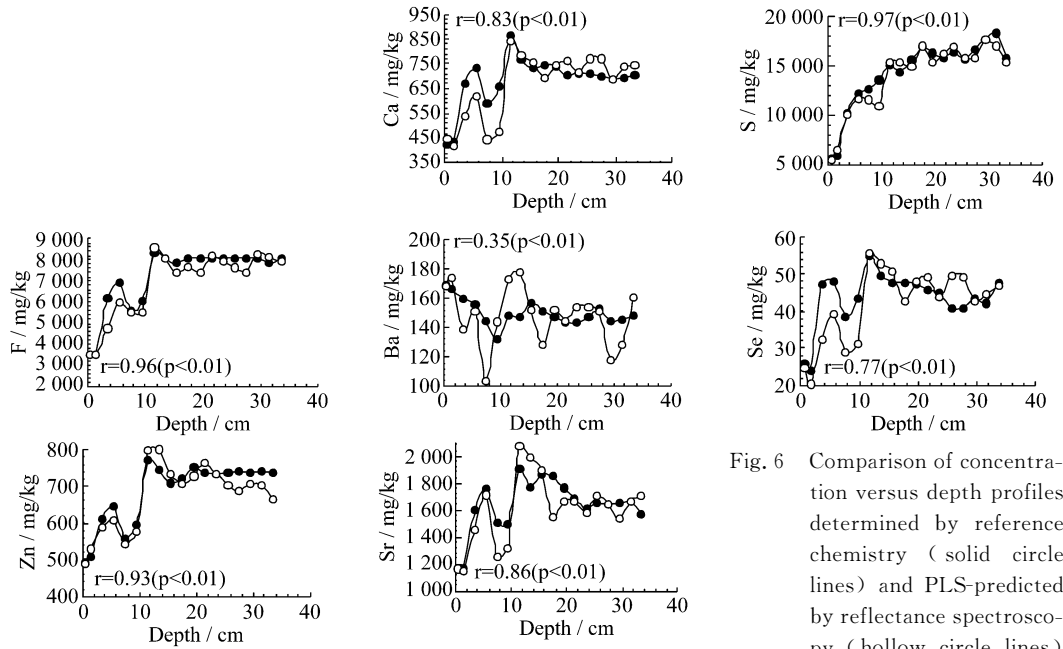


Fig. 6 Comparison of concentration versus depth profiles determined by reference chemistry (solid circle lines) and PLS-predicted by reflectance spectroscopy (hollow circle lines) for nine bio-elements in the Y4 lacustrine sediments.

the reflectance spectra of the sediments are significantly correlated with the nine bio-element concentrations (S, P, Ca, Cu, Zn, Se, Sr, Ba) and these elements are predicted more precisely by PCR and PLS. The correlation coefficients between PLS-predicted and chemically analyzed concentrations of these nine bio-elements are very high, and the predicted concentration-versus-depth profiles were very consistent with those from actual chemical analysis. These results demonstrated that it was feasible to predict bio-element compositions in the maritime Antarctic using the rapid, cost-effective, non-destructive and simultaneous technique of reflectance spectroscopy. Thereby this technique has the potential of being a valuable tool for the palaeo-ecological process study of penguins, other seabirds and mammals in Antarctic region through augmenting sampling density.

Acknowledgments We would like to thank Chinese Antarctic and Arctic Administration of National Oceanic Bureau for the logistic support for field sampling. This study was supported by the National Natural Science Foundation (Grant Nos. 40876096, 40606003 and 40730107), the young fund for strategic research of Chinese polar sciences from CAAA (No. 20070202), open research fund from SOA Key Laboratory for Polar Science (KP2007002) and special fund for excellent PhD thesis of CAS.

References

- [1] Nilsson MB, DÅbakk E, Korsman T, Renberg I (1996): Quantifying relationships between near-infrared reflectance spectra of lake sediments and water chemistry, *Environ. Sci. Technol.*, 30: 2586 – 2590.
- [2] Malley DF, Williams PC (1997): Use of near-infrared reflectance spectroscopy in prediction of heavy metals in freshwater sediment by their association with organic matter, *Environ. Sci. Technol.*, 31: 3461 – 3467.
- [3] Kooistra L, Wehrens R, Leuven RSEW., Buydens LMC (2001): Possibilities of visible-near-infrared spectroscopy for the assessment of soil contamination in river floodplains. *Analytica Chimica Acta*, 446: 97 – 105.
- [4] Pasquini C (2003): Near Infrared Spectroscopy: Fundamentals, Practical Aspects and Analytical Applications. *J. Braz. Chem. Soc.*, 14(2): 198 – 219.
- [5] Font R, Del Río M, Vélez D, Montoro R, De Haroa A (2004): Use of near-infrared spectroscopy for determining the total arsenic content in prostrate amaranth. *Science of the Total Environment*, 327: 93 – 104.
- [6] Sørensen LK, Dalsgaard S (2005): Determination of clay and other soil properties by near infrared spectroscopy. *Soil Sci. Soc. Am. J.*, 69: 159 – 167.
- [7] Wu YZ, Chen J, Ji JF, Tian QJ, Wu XM (2005): Feasibility of Reflectance Spectroscopy for the Assessment of Soil Mercury Contamination. *Environ. Sci. Technol.*, 39: 873 – 878.
- [8] Richardson AD, Reeves III JB (2005): Quantitative reflectance spectroscopy as an alternative to traditional wet lab analysis of foliar chemistry: near-infrared and mid-infrared calibrations compared. *Can. J. For. Res.*, 35: 1122 – 1130.
- [9] Wolfe AP, Vinebrooke RD, Michelutti N, Rivard B, Das B (2006): Experimental calibration of lake-sediment spectral reflectance to chlorophyll α concentrations: methodology and paleolimnological validation. *J Paleolimnol*, 36: 91 – 100.
- [10] Cohen MJ, Prenger JP, DeBusk WF (2005): Visible-near infrared reflectance spectroscopy for rapid, nondestructive assessment of wetland soil quality. *J. Environ. Qual.*, 34: 1422 – 1434.

- [11] Xia XQ, Mao YQ, Ji JF, Ma HR, Chen J, Liao QL (2007): Reflectance spectroscopy study of Cd contamination in the sediments of the Changjiang River, China. *Environ. Sci. Technol.*, 41: 3449 – 3454.
- [12] Wang L, Okin GS, Wang J, Epstein H (2007): Predicting leaf and canopy ¹⁵N compositions from reflectance spectra. *Geophysical Research Letters*, 34: L02401, doi: 10.1029/2006GL028506.
- [13] Korsman T, Nilsson M, Öhman J, Renberg I (1992): Near-infrared reflectance spectroscopy of sediments: a potential method to infer the past Ph of lakes. *Environ. Sci. Technol.*, 26: 2122 – 2126.
- [14] Korsman T, Nilsson MB, Landgren K, Renberg I (1999): Spatial variability in surface sediment composition characterized by near-infrared (NIR) reflectance spectroscopy. *J Paleolimnol.*, 21: 61 – 71.
- [15] Foley WJ, McIlwee A, Lawler I, Aragonés L, Woolnough AP, Berding N (1998): Ecological applications of near infrared reflectance spectroscopy—a tool for rapid, cost-effective prediction of the composition of plant and animal tissues and aspects of animal performance. *Oecologia*, 116: 293 – 305.
- [16] Malley DF, Röncke H, Findlay D, Zippert B (1999): Feasibility of using near-infrared reflectance spectroscopy for the analysis of C, N, P, and diatoms in lake sediments. *J Paleolimnol.*, 21: 295 – 306.
- [17] Malley DF, Lockhart L, Wilkinson P, Hauser B (2000): Determination of carbon, carbonate, nitrogen, and phosphorus in freshwater sediments by near-infrared reflectance spectroscopy: rapid analysis and a check on conventional analytical methods. *J Paleolimnol.*, 24: 415 – 425.
- [18] Rosén P, Dábakk E, Renberg I, Nilsson M, Hall R (2000): Near-infrared spectrometry (NIRS): a new tool for inferring past climatic changes from lake sediments. *The Holocene*, 10: 161 – 166.
- [19] Das B, Vinebrooke RD, Sanchez-Azofeifa A, Rivard B, Wolfe AP (2004): Inferring sedimentary chlorophyll concentrations with reflectance spectroscopy: a novel approach to reconstructing historical changes in the trophic status of mountain lakes. *Can. J. Aquat. Sci.*, 62: 1067 – 1078.
- [20] Chang CW, You CF, Huang CY, Lee TQ (2005): Rapid determination of chemical and physical properties in marine sediments using a near-infrared reflectance spectroscopic technique. *Applied Geochemistry*, 20: 1637 – 1647.
- [21] Rosén P (2005): Total organic carbon (TOC) of lake water during the Holocene inferred from lake sediments and near-infrared spectroscopy (NIRS) in eight lakes from northern Sweden. *Biogeochemistry*, 76: 503 – 516.
- [22] Michelutti N, Wolfe AP, Vinebrooke RD, Rivard B (2005): Recent primary production increases in arctic lakes. *Geophysical Research Letters* 32, L19715, doi:10.1029/2005GL023693.
- [23] Chang CW, Laird DA, Mausbach MJ, Hurburgh CR (2001): Near-infrared reflectance spectroscopy – principal components regression analyses of soil properties. *Soil Sci. Soc. Am. J.*, 65: 480 – 490.
- [24] Sun LG, Xie ZQ, Zhao JL (2000): A 3,000-year record of penguin populations. *Nature*, 407 – 858.
- [25] Sun LG, Zhu RB, Yin XB, Liu XD, Xie ZQ, Wang YH (2004): A geochemical method for reconstruction of the occupation history of penguin colony in the maritime Antarctic. *Polar Biology*, 27: 670 – 678.
- [26] Sun LG, Xie ZQ, Liu XD, Yin XB, Zhu RB (2006): Eco-geology in Antarctic ice-free areas. Science Press, Beijing: 1 – 306 (in Chinese).
- [27] Sun LG, Xie ZQ (2001): Relics: penguin population programs. *Science progress.*, 84(1): 31 – 44.
- [28] Liu XD, Sun LG, Xie ZQ, Yin XB, Wang YH (2005): A 1300-year record of penguin populations at Ardley Island in the Antarctica, as deduced from the geochemical data in the ornithogenic lake sediments. *Arctic Antarctic and Alpine Research*, 37(4): 490 – 498.
- [29] Liu XD, Sun LG, Xie ZQ, Yin XB, Zhu RB, Wang YH (2007): Preliminary Geochemical Record for the Pre-Historical Seabird Population in “Mochou” Lake Sediments on the Larsemann Hills,

- East Antarctica. *Boreas*, 36: 182 – 197.
- [30] Sun LG, Xie ZQ, Zhao JL(2001): The sediments of lake on the Ardley Island, Antarctic; Identification of penguin-dropping soil. *Chinese Journal of Polar Science*, 12(1): 1 – 8.
- [31] Sun LG, Zhu RB, Liu XD, Xie ZQ, Yin XB, Zhao SP, Wang YH (2005): HCl-soluble $^{87}\text{Sr}/^{86}\text{Sr}$ ratio in the sediments impacted by penguin or seal excreta as a proxy for the size of historical population in the maritime Antarctic. *Marine Ecology-Progress Series*, 303: 43 – 50.
- [32] Liu XD, Li HC, Sun LG, Yin XB, Zhao SP, Wang YH (2006): $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ in the ornithogenic sediments from the Antarctic maritime as paleoecological proxies during the past 2000 Years. *Earth and Planetary Science Letters*, 243: 424 – 438.
- [33] Wang JJ, Wang YH, Wang XM, Sun LG(2007): Penguins and vegetations on Ardley Island, Antarctica: evolution in the past 2,400 years. *Polar Biol.*, 30:1475 – 1481.
- [34] Trivelpiece WZ, Trivelpiece SG, Volkman NJ(1987): Ecological segregation of Adelie, Gentoo, and Chinstrap penguins at King George Island, Antarctica. *Ecology*, 68: 351 – 361.
- [35] Liu XD, Sun LG, Yin XB (2004): Textural and geochemical Characteristics of proglacial sediments: a case study in the foreland of Nelson Ice Cap, Antarctica. *Acta Geological Sinica*, 78: 970 – 981.
- [36] Lu WZ, Yuan HF, Xu GT, Qiang DM(2000): Analytical Technology of Modern Near-Infrared Spectroscopy. Beijing: China Petrochemical Press, 1 – 260(in Chinese).
- [37] Rossel RAV (2008): ParLeS: Software for chemometric analysis of spectroscopic data. *Chemometrics and Intelligent Laboratory Systems*, 90: 72 – 83.