

The Influence of Climate and Landscape on *Allocapnia mohri*  
Distribution in Arkansas

A thesis submitted in partial fulfillment  
of the requirements for the degree of  
Master of Science in Biology

by

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Bachelor of Science in Biology, 2016

August 2023  
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## Abstract

Stoneflies are a globally threatened aquatic insect order. In Arkansas, a diverse group of winter stonefly (Capniidae: *Allocapnia*) have not been surveyed since the 1980s, likely because species-level identification requires the rarely-collected adult form. *Allocapnia mohri*, a regional endemic, was previously commonly found in mountainous, intermittent streams from the Ouachita Mountains ecoregion north to the Ozark Highlands, but no species distributional models including land use or climate variables exist to our knowledge. We collected adults from 70 stream reaches from the historic Arkansas range from November to April 2020 and 2021. We modeled distributions using random forest (RF) models populated with landscape, climate, and both data to determine which were most predictive of species presence. Correlations between landscape or climate variables and presence were examined using multiple logistic regression. The landscape RF models performed better than the climate or landscape + climate RF models. *A. mohri* presence sites tended to have a greater elevation, a lower mean July temperature, and a greater percentage of very slow infiltration soils in the watershed, compared to absence sites. *A. mohri* was absent at the Ouachita Mountains sites and may be experiencing a range contraction or migration northward.

## **Acknowledgements**

I would like to thank my lab members, Camryn Larson, Sam Dias, and Zachary Tipton for their help with revisions and stonefly collection. I would like to thank my husband, Adam Annaratone, for his companionship and assistance on all my field sampling trips. I would like to thank Dr. John Tyler Fox for his help with coding and statistics.

This project was funded in part by the State Wildlife Grants Program (Grant#AR-T-F19AF01234) of the U.S. Fish and Wildlife Service through an agreement with Arkansas Game and Fish Commission.

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## Published Papers

Chapter 1- The Influence of Climate and Landscape on *Allocapnia mohri* Distribution in Arkansas

1. Annaratone B, Larson C, Prater C, Dowling A, Magoulick DD, Evans-White MA. Predicting Habitat and Distribution of an Interior Highlands Regional Endemic Winter Stonefly (*Allocapnia mohri*) in Arkansas Using Random Forest Models. *Hydrobiology*. 2023; 2(1):196-211. <https://doi.org/10.3390/hydrobiology2010013>

## I. Introduction

Stoneflies (Plecoptera) are a highly diverse macroinvertebrate group encompassing more than 1000 species and representing 99 genera and ten families in North America [1]. Though the plecopteran group is expansive, little species-specific data on habitat, feeding habits, and distributions exist, and several existing records need updating to more accurately estimate their population status and to manage their conservation. This information is essential as Plecoptera are estimated to be one of the most endangered groups of insects globally [2,3]. Possible reasons for this include the alteration of plecopteran habitat by anthropogenic influences, such as climate and land use changes [3]. Here we model distributions of an endemic stonefly species to close the gap in species-specific stonefly knowledge, and to investigate whether climate, landscape, or a combination of factors could be affecting spatial distributions of stoneflies.

Several insect species, including the nymphs of some species of stonefly, are reliant on allochthonous resources [4], and the energy transported by these terrestrial materials, along with the energy acquired through autochthonous resources, is the primary source of matter and energy for many streams, particularly low order forested streams [5–7]. Shredding stoneflies will consume autumn-shed leaves, then these nymphs are subject to predation, thereby using the energy acquired through allochthonous sources. [8]. This cycling of allochthonous material into the stream makes stoneflies a vital component of the stream food web [9]. The reliance on fallen leaves for food adds a habitat requirement of course particulate organic matter (CPOM) for shredder stoneflies in this macroinvertebrate group, and several stonefly species have strict water oxygenation and temperature requirements [10,11], increasing the habitat needs for these sensitive plecopteran nymphs. One such group of threatened plecopterans is the winter stonefly.

Stoneflies require specific environmental conditions, such as stream water temperatures and hydroperiod [12], and these factors will be affected by global climate change [13].

Ephemeroptera, Plecoptera, and Trichoptera (EPT) richness has displayed a decline of 81.6% worldwide, coming to an average loss of 1.9% loss of EPT richness each year. This 1.9% loss in richness mirrors the 1.88 °C increase in water temperature over the 40 years [14]. Several stonefly species have a thermal cue for egg hatching, and if stream temperatures do not reach a thermal low-end cue, nymphs will not hatch, causing extinction of species in certain thermally inadequate streams [12]. The effect of changing hydroperiod can greatly affect stoneflies, as they spend most of their lives as aquatic nymphs. Plecopterans are sensitive to benthic habitat loss, due in part to their need to lay eggs inside refugia within the stream (i.e., aquatic plants and woody debris), and their need to emerge using this refugia [12]. In some streams, stonefly nymphs can retreat to the hyporheic zone in dry periods, and it was found that several stonefly species even have diapausing eggs to outlast the dry stream conditions [15], though for many species and for many systems, the hyporheic zone is not as important as refugia. Small stoneflies have been found to aggregate on stones, and larger stoneflies aggregate in leaf patches during high flow, as well as a general movement toward areas with flow in drying summer months [16].

Land-use change is another key driver of declines in stonefly populations [3]. Loss of riparian forest will likely amplify increases in stream water temperatures [17], reducing forested area in catchments, which may reduce stonefly habitat availability [18], and this, along with climate change, may cause reductions in stonefly populations due to their strict thermal requirements [10]. Further, standing crops of coarse benthic organic detritus can be directly related to the secondary production of shredding macroinvertebrates, such as stoneflies [19,20]. So, the loss of

forested land in catchments can be strongly linked to a lower standing crop of litter detritus and likely reduced food availability for shredding stoneflies.

Forested land is often converted to agricultural or urban land uses that may also contribute to stonefly population declines and local species extinctions. Anthropogenic activities and land usage around stream systems directly influence the physical and chemical characteristics of the streams [21], including the amounts of sewage inputs, salts from road de-icing, and agricultural fertilizer and pesticides that seep into streams [22–24]. Anthropogenic activities can also result in impervious soil surrounding streams, leading to slow infiltration of water [25], causing an increase in speed and volume of storm runoff [26], and an increase in pollutants, ultimately causing a degradation of the aquatic habitat [27] and a loss of benthic macroinvertebrate diversity [28].

Anthropogenic nutrient inputs often lead to an increase in primary production [29], and this eutrophication can lead to low levels of dissolved oxygen (DO) during certain times of the day [29], thereby killing sensitive species in the stream, such as stoneflies [30]. This increase in nutrients can also stimulate microbial decomposition of leaf litter, the food source of shredders such as stoneflies [31]. While microbial colonization is thought to improve the nutritional quality of leaf litter for shredders, this is not always the case [32–35]. Further, elevated microbial decomposition may reduce the quantity of coarse detrital carbon available for food in stream ecosystems [36]. These effects can compound over time and be detrimental to stonefly species in the long-term.

Plecopterans are sensitive to these stream impurities and to microhabitat disturbances, which makes them an important biological model to indicate water quality [37]. Most of the studies on benthic macroinvertebrates in relation to water quality evaluations have dealt with the richness of



the EPT community because these groups are intolerant to pollutants in water bodies, but in the EPT community, plecopterans are often the most sensitive to changes in water quality [38].

While the common causes of global-scale stonefly declines are known, a finer-scale, regional perspective is needed for conservation of sensitive groups, such as winter stoneflies. Winter stoneflies (Order: Plecoptera) emerge from the nymph stage in the winter months, and they are known as “clean-water insects” due to their need for highly oxygenated water [39]. A genus of winter stoneflies (Family: Capniidae, Genus: *Allocapnia*) emerges from the nymph stage to a small-winged adult across a wide variety of intermittent to perennial streams on vertical surfaces in and around the water body and lacks stream-to-stream vagility [12]. The Arkansas Level III Ecoregions known as the Ozark Highlands, Boston Mountains, Arkansas Valley, and Ouachita Mountains have a high diversity of *Allocapnia*. During the last regional census in the late 1980’s, seven endemic species of *Allocapnia* were found in Arkansas [12], several of which have recently been listed as species of greatest conservation need (SGCN) according to the Arkansas Game and Fish Commission’s 2019 Arkansas Wildlife Action Plan (AWAP).

*Allocapnia mohri* is a regional endemic that was commonly collected in the last 1980’s census [12], making it a good possible candidate for assessing important habitat factors and for distribution modeling. *A. mohri* populations during the previous census occurred broadly across Strahler stream orders one through five and were also found broadly across sites with differing flow permanence, from intermittent sites with a completely dry stream bed or only pools with no flowing water for part of the year to perennial sites [12]. However, they were not detected at perennial sites with a significant groundwater flow [12]. This may be due to groundwater-based streams lacking the necessary temperature cue for egg hatching, whereas runoff dominated streams get cold enough with ice and snow melt entering the stream to cue hatching.

During the last widespread survey of *Allocapnia* in Arkansas, geospatial climate and land use data for stream catchments were not readily available [12] making it difficult to determine what factors are associated with *A. mori* distributions in the region. Species distribution models (SDMs) are empirical models relating field presence or abundance data to environmental predictor variables [40]. The use of SDMs has gained popularity in the past 20 years in ecological fields as geospatial mapping data with high resolution has become more readily available to conservation biology, making it possible to gauge the effect of climate, land use, and other environmental changes on the distribution of rare and understudied species [41].

One particular way of creating distribution maps that is gaining popularity is Random Forest (RF) modeling. Random Forest modeling is a group of tree predictors based on classification and regression tree (CART). The trees depend on the value of a random, independently sampled vector, so the method reduces bias [42]. Random Forest makes predictions by machine learning and then utilizes out-of-bag samples for model validation [43]. This method is excellent at minimizing overfitting due to its random nature and is effective at modeling spatial data even though it is not a spatial method. Random Forest modeling is superior to other popular SDM procedures, such as Mahalanobis Typicalities, a method adopted from remote sensing analyses. It is comparable to Maxent, a statistical mechanics approach, when one has a small-to-moderate number of the collected species presence/absence records ( $n = 38-94$ ), and in species with low dispersal abilities [44]. The average number of records of each species of *Allocapnia* sampled in the previous census was 39 [12], and Plecoptera have reduced or absent flight capabilities, so Random Forest will likely work well for modeling winter stonefly species.

No SDMs exist for *A. mohri* to our knowledge. However, it was found to be the most widely distributed species of *Allocapnia* in Arkansas in the 1980's (Figure 1A). Though this species used

to be the most common through Arkansas, the species range has not been evaluated since the 1990s census. Also, due to the lack of geospatial data, no distribution models including land use and landcover or climate variables have been developed for *A. mohri*. Thus, our study objective was to recensus the Arkansas population and use the data to spatially model *A. mohri* distributions in Arkansas using RF models. Landscape, climate, and a combined landscape and climate data set were used to populate our RF model to determine which factors were most predictive of species presence. We then used multiple logistic regression to examine four hypotheses:

Hypothesis H1-*A. mohri* will be more likely to occur in mountainous regions and higher elevations [45]. Hypothesis H2- *A. mohri* will differ across landuse types. Specifically, we expect that the species should be more common in areas with high percentages of forested land [46] because leaf litter inputs are an important food source for detritivorous stoneflies [47] and will decline with shifts to other landuse types. Hypothesis H3-*A. mohri* will be more common in streams with high rainfall and low water temperatures [46]. Hypothesis H4-*A. mohri* will be less common in streams with a slow soil infiltration rate, due to their sensitivity to stream impurities [12].

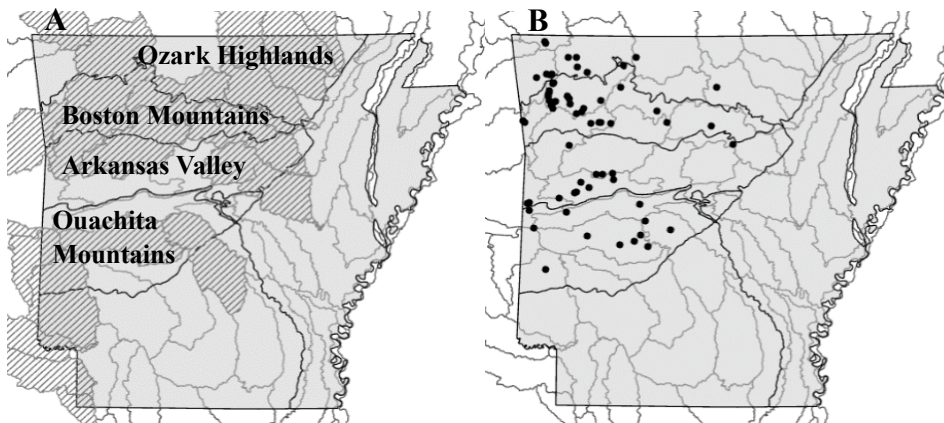


Figure 1. (A) Historic distribution of *A. mohri* across Omernik Level III ecoregions. The HUC8 units that overlapped with *A. mohri* presence watersheds from Poulton and Stewart, 1991 are hatched. The HUC8 units in Arkansas that did not have *A. mohri* presence are outlined. (B) Locations of the 70 reaches sampled in the present study. The northernmost site coordinates are (36.44117572, -94.22998045), and the southernmost site coordinates are (34.68526175, -94.32049797). Sample reaches fell within the following HUC8 units: Ouachita Headwaters, Upper Saline, Fourche La Fave, Poteau, Petit Jean, Frog-Mulberry, Dardanelle Reservoir, Robert S. Kerr Reservoir, Beaver Reservoir, Bull Shoals Lake, Elk, Little Red, Middle White, Buffalo, Upper Ouachita, Little Missouri, Lower Little, Illinois.

## II. Materials and Methods

### 2.1. Study Design

Our resulting RF SDMs are some of the first published for a North American plecopteran species (but see [48]), and the landscape RF model performed better than the climate or combined RF models. The MLR modeling indicated both landscape and climate variables were important for predicting presence. *Allocapnia mohri* was absent at the Ouachita Mountains sites in 2020 and 2021, possibly indicating range changes [12].

Our study area included 70 sample reaches stratified across the Level II Ecoregion Ozark/Ouachita–Appalachian Forests, which lies within the Level I Ecoregion, the Eastern Temperate Forests. The sample reaches were further spread into the following Level III Arkansas Ecoregions: Ozark Highlands (n = 15), Boston Mountains (n = 29), Arkansas Valley (n = 13), and Ouachita Mountains (n = 13) (Figure 1). These Level III ecoregions historically had high stonefly diversity compared to other ecoregions in Arkansas [12]. Sample locations were based on historical studies reporting presence of *Allocapnia* genera [12]. We also stratified sample sites by Strahler stream order with several representing 1st order (n = 28), 2nd order (n = 15), 3rd order (n = 10), 4th order streams (n = 10), 5th order (n = 6), and 6th order (n = 1) (Table 1). Finally, the hydrologic flow regime was evaluated using the GeoCrawler Google Earth

application (application located in Appendix B <https://doi.org/10.1002/rra.2838>). Due to the high amount of headwater streams sampled, and the stream locale belonging mainly to the Eastern Temperate Forests, most study sites had a high probability of belonging to the intermittent flashy flow regime (n = 46) [49].

Table 1. The range, median, and mean of each variable across our study sites.

<b>Variables</b>	<b>Range</b>	<b>Median</b>	<b>Mean</b>
Stream Order	1st - 6th	2	2.37
Mean Width (m)	0.66 - 6.52	2.6	2.99
Mean Depth (cm)	5.56 - 41.40	14.72	15.63
Elevation (m)	155-598	402	376.11
%Forested	8.61 - 98.6	86.27	75.92
%Agriculture	0.00 - 51.57	7.67	12.78
%Urban	0.23 - 57.73	5.04	10.99
%High Infiltration Soils	0.00 - 66.76	2.09	4.30
%Moderate Infiltration Soils	0.00 - 84.21	19.25	25.13
%Low Infiltration Soils	0.00 - 100.00	16.73	26.49
%Very Slow Infiltration Soils	0.00 - 96.10	39.02	39.57
Total Annual Precipitation (cm)	110.0 - 155.3	126.0	125.33
Total February Precipitation (cm)	5.9 - 10.8	7.9	7.86
Total May Precipitation (cm)	12.7 - 17.5	14.7	14.8
Annual Mean Water Temperature (°C)	14.2 - 16.9	14.6	15.1
January Mean Water Temperature (°C)	2.1 - 5.6	2.8	3.3
July Mean Water Temperature (°C)	25.1 - 27.6	25.9	26.1

## 2.2. Local Site Characterization

We measured the length and five wetted widths along evenly spaced transects for each habitat unit (e.g., riffles, pools, runs) at wadable sites in August and September 2020–2022. Stream depth was measured at each width transect by taking five measurements across the cross-section of the stream. Each sample stream reach was 200 m in length, allowing all reaches to measure at least six times the wetted width of the stream, and each reach included at least one pool and riffle

sequence [50]. Discharge was measured at the base of the reach using the mid-section method [51] at wadable sites with no U.S. Geological Survey (USGS) gaging station. Otherwise, gaging station discharge was used.

Stream permanence was also estimated according to two separate metrics—one method described by Poulton and Stewart (1991) [12] and the other by Sheldon and Warren (2009) [52] in August and September of 2020–2022, which is generally the driest time of the year. The first method involved giving streams a ranking of A through D depending on the dryness over time, with an A ranking given to streams with a dry bed for part of the year and a D ranking given to streams with a permanent flow through the year along with a significant underground source. The second metric was a stream drying metric wherein we measured the linear extent of visible surface water in the stream reach in marked 50 m sections on each visit, then converted the wetted extent to a proportion, and calculated a time-weighted annual average for each site. This stream drying metric is primarily an index of time without surface flow, whereas the former gives a large-scale idea of the stream permanence. Using both metrics gives a clearer idea of the flow characteristics of each stream.

### *2.3. Landscape-Level Site Characterization*

We collected two main types of landscape data: table and raster. Tabular climate and watershed data was collected virtually for all sites via Model My Watershed [53], and it was used in Multiple Logistic Regression (MLR) models (see Statistical analyses and modeling section). Several high-resolution landscape-level rasters were collected for the state of Arkansas to create distribution models of the species (see Statistical analyses and modeling section), including a land use raster (NLCD 2016) [54], a soil variable raster (gNATSGO) [55], and a large-scale environmental data raster (WorldClim) [56].

#### 2.4. *A. mohri* Presence

Adult stonefly sampling started in early December and continued through mid-April during the 2020–2021 and 2021–2022 field seasons to match the emergence periods of *A. mohri*. Several approaches have been used to determine presence of *Allocapnia* populations in the past [12,52,57,58]. None of these methods have been evaluated to determine detection probability, which requires multiple samples over the emergence period. We chose to focus on adult collections, and all sample reaches were visited at least nine times across the emergence period to allow for evaluation of detection probability for future research. We searched for adults in all areas of the 200 m reaches but particularly focused searches on compacted leaves and rocks in riffles, tree trunks, woody debris, and leaf litter debris piles on the side of the stream for 20 minutes per reach. All adults were collected by hand with forceps and immediately preserved in 95% ethanol, transported back to the laboratory, sexed, and identified to species using a regional taxonomic key [12]. The species were then placed on a distributional map.

#### 2.5. Statistical Analyses and Modeling

We chose Random Forest (RF) to determine the possible distribution of a species of *Allocapnia mohri* relative to using MaxEnt and Mahalanobis because of its reduced bias with moderate to large sample size and transparent readout [43]. The RF species distribution models in this study were created in ArcGIS [59], which also produced out-of-bag score, mean squared error, Matthews Correlation Coefficient, and F1 statistics to evaluate model accuracy. The out-of-bag (OOB) error is the average error for each tree, calculated using predictions from the trees that do not contain it in their respective bootstrap sample. This score provides information on the expected performance on new, unseen data, and generally, smaller OOB scores correspond with better models. The mean squared error (MSE) is a measure of the prediction accuracy of the

model. There are no set acceptable limits for MSE, but in general the lower the MSE, the more accurate the model. F1 scores are a weighted average of the precision and recall of a model, where an F1 score reaches its best value at 1 and worst score at 0. The Matthews Correlation Coefficient scores are based on results for all four confusion matrix categories (true positives, false negatives, true negatives, and false positives), and a higher score corresponds to a better result. It scores proportionally both to the size of positive and negative elements in the dataset, making it a great metric for unbalanced data [60,61].

Three different RF models were created to parse out effects of land cover and climatic data. Each model was trained with 80% of the collected presence and absence data, and 20% of the data was used to validate and test the models. Model 1 was the “Landscape Model”, which included all land use and soil variable rasters along with an elevation raster. Model 2 was the “Climate Model”, which included only climatic raster data. Model 3 was the “Combined Model”, which included soil, land cover, and climatic variable rasters.

Random forest models are limited in that they do not depict the way that variables relate to one another, and they do not determine if the variables are significantly affecting *A. mohri* presence. Therefore, they are not appropriate for testing hypotheses about important habitat factors describing the landscape or climate niche of *A. mohri* in headwater streams. To resolve this issue, the data were evaluated using a multiple logistic regression (MLR). The MLR models were created in R (glm{stats})[62]. A total of four separate models were created based on the four established hypotheses for this study (Table 2). The Hypothesis 1 (H1) model evaluated the ecoregion and elevation variables in the study to see if *A. mohri* was more common in mountainous regions and higher elevations. The Hypothesis 2 (H2) model included land use data to determine if *A. mohri* will differ across land use types. The Hypothesis 3 (H3) model included



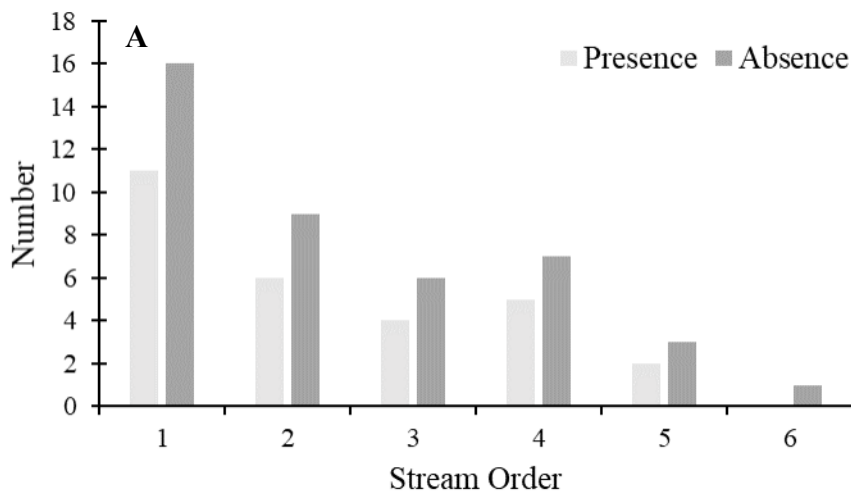
annual precipitation total and annual mean temperature variables to determine if *A. mohri* is more common in streams with high rainfall and low water temperatures. The Hypothesis 4 (H4) model included water infiltration rate variables to determine if *A. mohri* would be less common in streams with impacted soils. All landscape and climate predictor variables were tested for model assumptions including binomial distribution, independence of error terms, linear relationship between  $\text{logit}(Y)$  and  $X$ , and lack of collinearity among predictors. After checking for assumptions, some variables were excluded from analysis due to collinearity issues (correlation among predictor variables), and the variables that lacked logistic restraints and were determined to be of highest biological importance in the model were retained. All models were then compared and ranked using Akaike Information Criterion (AIC) to determine which model was best at predicting *A. mohri* presence, and Hosmer–Lemeshow goodness-of-fit tests and likelihood-ratio tests were run to compare all models to the global model.

**Table 2.** Variables used in Multiple Logistic Regression analyses grouped by hypothesis. N/A indicates that no variable was removed from the final model due to correlation and/or collinearity.

Model	Variables included in MLR models	Removed Due to Correlation and/or Collinearity
H1 Model	Ecoregion, Max Elevation	Average Elevation, Minimum Elevation, Elevation
H2 Model	%Urban	%Forest, %Agriculture
H3 Model	Annual Precipitation Total (cm), July Mean Water Temperature, January Mean Water Temperature, May Total Precipitation	Annual Mean Temperature, May Total Temperature, February Total Precipitation
H4 Model	High Infiltration, Moderate Infiltration, Very Slow Infiltration	Slow Infiltration

### III. Results

We collected and identified 3463 *A. mohri* individuals, with 2519 males and 944 females. This species began emerging around the end of November with the first *A. mohri* collection being made on November 23, 2020, nearly coinciding with the previously stated early December start of the emergence period for *A. mohri* [12]. We found *A. mohri* adults at 28 of our 70 sample reaches. Adults were found across stream orders one through five with no bias towards a particular stream order (Pearson's chi-square = 3.98,  $p = 0.55$ , Figure 2A). *Allocaenia mohri* was not found at our 6th order stream site. *Allocaenia mohri* presence differed across sites with a bias towards the Boston Mountain ecoregion (Pearson's chi square = 12.045,  $p = 0.07$ , Figure 2B) and was not found in the Ouachita Mountain sites. Adults were also found across all hydrologic permanence categories with no bias towards a particular category (Pearson's chi-square = 2.01,  $p = 0.57$ , Figure 2C).



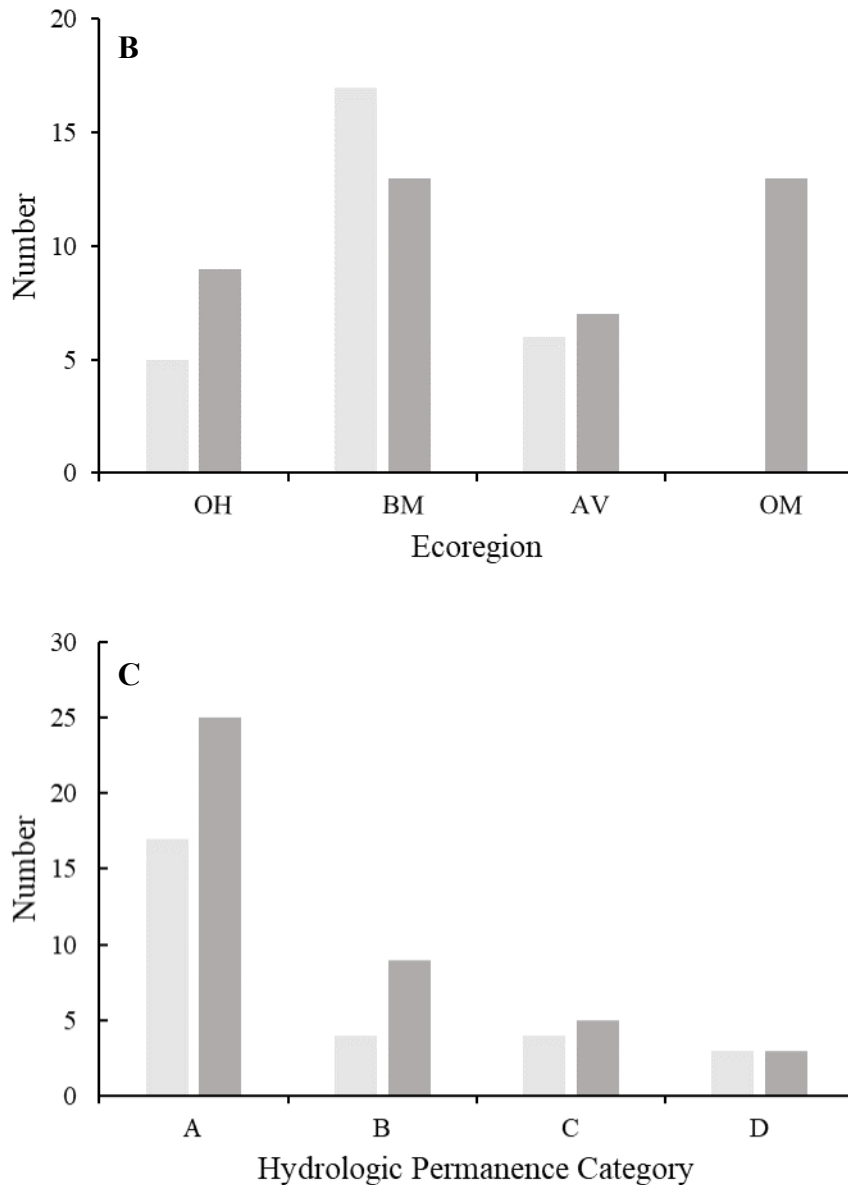


Figure 2. The distribution of *A. mohri* present and absent sites across stream orders (A), Omernik level III Ecoregions (B) of Ozark Highlands (OH) Boston Mountains (BM), Arkansas Valley (AV), and Ouachita Mountains (OM), and Hydrologic Permanence Categories (C); Poulton and Stewart, 1991).

Random Forest models estimated the potential distribution of *A. mohri* across Arkansas. The Landscape Model (Figure 3A) had an out-of-bag score of 0.409, an F1 score of 0.80, and MSE of 4.382, and a MCC of 0.74. Further evaluation of the visual present/absent prediction output of the RF model shows that 19 known present sites fall within the produced predicted bounds and

31 known absent sites correctly fall within the produced predicted absent bounds, meaning the visual prediction output for the Landscape model correctly sorted 50 present and absent sites out of 70 (71.4%).

The Climate Model (Figure 3B) had an out-of-bag score of 0.864, an F1 score of 0.62, a MSE of 1.173, and a MCC of 0.33 (Table 3). Further evaluation of the visual present/absent prediction output of this RF model shows that 23 known present sites fall within the produced predicted bounds and 31 known absent sites fall inside the produced predicted absent bounds, meaning the visual prediction output for the Climate model correctly sorted 54 present and absent sites out of 70 (77.1%).

The Combined Model (Figure 3C) had an out-of-bag score of 0.114, an F1 score of 0.60, a MSE of 1.373, and a MCC of 0.43 (Table 3). Further evaluation of the visual present/absent prediction output of this RF model shows that 24 known present sites fall within the produced predicted bounds and 30 known absent sites fall inside the produced predicted absent bounds, meaning the visual prediction output for the Landscape model correctly sorted 54 present and absent sites (77.1%).

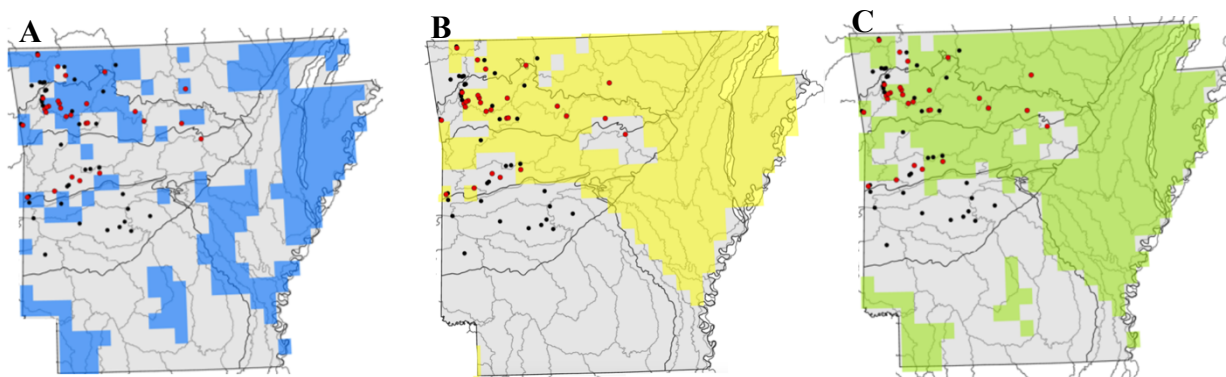


Figure 3. The Random Forest distribution models for *Allocapnia mohri* plotted with the known species distribution. *A. mohri* presence (red circle) and absence (+) sites are marked. The areas that the Landscape distribution model predicted would have *A. mohri* presence are marked in blue (A).

The areas that the Climate distribution model predicted would have *A. mohri* presence are marked in yellow (B). The combined Landscape and Climate model prediction is in green (C).

Table 3. Random Forest model scores for Out-of-Bag (OOB), F1, Mean Squared Error (MSE), Matthews Correlation Coefficient (MCC), Accuracy, and Validation Accuracy variables grouped by hypothesis.

Model	Out of Bag Score	F1 Score	MSE	MCC	Accuracy	Validation Accuracy
Model 1: Landscape	0.409	0.80	4.382	0.74	0.98	0.87
Model 2: Climate	0.864	0.62	1.173	0.33	0.91	0.67
Model 3: Climate + Landscape	0.114	0.60	1.373	0.43	0.93	0.73

Next, MLR models were used to test our study hypotheses. The H1 model showed no elevation or ecoregion variable was significant in predicting *A. mohri* presence ( $p > 0.05$ ) (Table 4). The H2 model was the best model according to AIC ranking. This H2 model only consisted of a single variable, urban land percentage. The H2 model showed that percent urban land use variable was significant in predicting *A. mohri* presence ( $p < 0.05$ , chi-squared = 0.07, Table 4), and correlation between urban land and species presence was negative (Figure 4). The H3 model showed that three temperature variables were significant in predicting *A. mohri* presence (Table 4): January water temperature ( $p < 0.05$ , chi-square = 0.05) (Figure 4B), July water temperature ( $p < 0.05$ , chi-square = 0.03) (Figure 4C), and annual temperature ( $p < 0.05$ , chi-square = 0.05) (Figure 4D); all temperature variables were negatively correlated with species presence (Fig. 4). The results from the H4 model revealed that very slow soil infiltration was significant in predicting *A. mohri* presence ( $p < 0.05$ , chi-square = 0.03; Table 4), and the correlation between *A. mohri* presence and very slow soil infiltration was positive (Figure 4E).

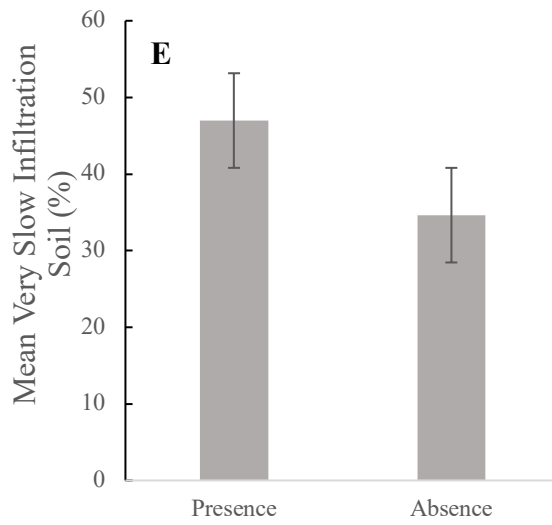
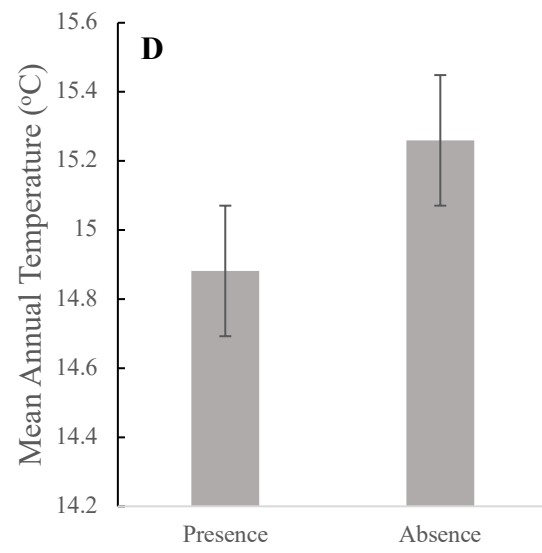
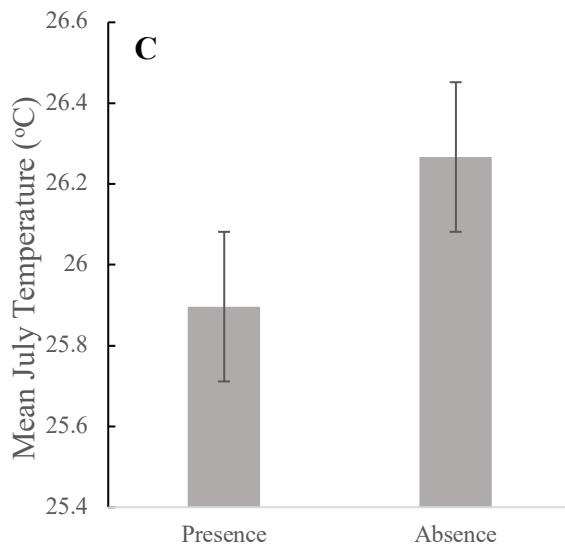
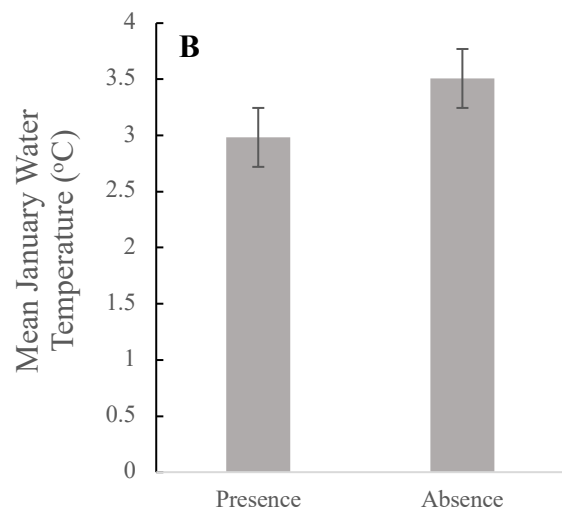
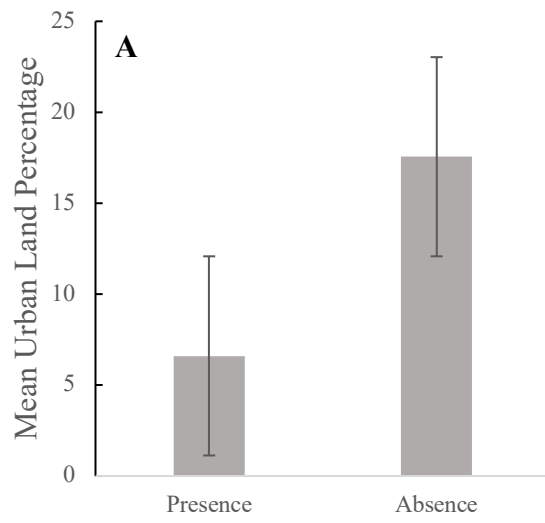


Figure 4. The mean and standard error of urban land percentage (A), January water temperatures (B), July water temperatures (C), annual temperatures (D), and the percentage of very fine soils in the watershed (E) for sites where *Allocapnia mohri* was present and absent.

Table 4. Results from the Multiple Logistic Regression (MLR) showing values for p and chi-square, then Akaike Information Criterion (AIC) ranking for four hypotheses related to *Allocapnia mohri* presence/absence.

Model	df	Chi-Squared	p-Value	AIC Score
H1 Model	3	0.03	P > 0.05	96.57
H2 Model	2	0.07	P < 0.05	90.01
H3 Model	2	0.05	P < 0.05	92.37
H4 Model	2	0.03	P < 0.05	96.52

#### IV. Discussion

Stoneflies are a major component to aquatic diversity [47]. Many species are globally imperiled [3,63], and yet little is known about species-specific habitat requirements and tolerances to changing land use and climate variables. Our study provides one of the first RF species distributional models for a North American plecopteran species. It also provides more detailed habitat and climate sensitivity information and updates distribution data that can aid conservation of a regional endemic winter stonefly species.

A 1980's census provided some information about *A. mohri*'s habitat [12] that is generally consistent with the present study's findings, but our study adds to their results. We also found *A. mohri* broadly inhabited stream orders and hydrologic permanence categories. Specifically, *A. mohri* has been historically found at intermittent lower-order sites that have a completely dry stream bed or only have pools during parts of the year; therefore, they may have desiccation-resistant, diapausing eggs [64]. They also occurred at higher-order perennial sites; we found them at perennial sites that also have a significant underground spring source. *Allocapnia mohri*

was historically found in first through fifth-order streams but not in sixth-order streams. We found this same pattern. However, we only censused one sixth-order stream, which was located in the Ouachita Mountains. So, our study has limited power evaluating species presence in larger rivers.

Curiously, we did not find *A. mohri* in any of our 16 Ouachita Mountains stream reaches.

*Allocaonia mohri* was found in Ouachita Mountains streams in the 1980's [12]. Further, it was found in the Little Glazypeau Creek and Alum Fork of the Saline River drainages in 2000–2001 [52], and in Lick Creek of the Petit Jean River drainages in 2022 (DeWalt et. al., Unpublished Data). We have study reaches in these watersheds and did not find them despite visiting the reaches nine times during the winter emergence period. Their absence in the Ouachita Mountains ecoregion may be a sampling artefact, but it is possible that their southern range is shifting northward.

The Random Forest model using only landscape predictor variables was the best model of *A. mohri* distribution according to the MCC and F1 scores (Table 3). The Landscape model scored the highest in MCC out of all models and this metric in particular is an appropriate metric for this dataset, being that it works proportionally to the size of both positive and negative elements in the data, and the dataset that was used was uneven, with much more absence values than presence values. The Landscape model had the best accuracy in correctly predicting *A. mohri* absence when evaluating the incorrectly sorted present and absent sites in the RF prediction output. However, the Landscape model incorrectly sorted 20 present and absent sites out of the total 70 sample sites and incorrectly predicted that *A. mohri* should be present in the Ouachita Mountains ecoregion.



The Climate model was determined to be less accurate than the Landscape model according to the MC and, F1 scores, though it was the most accurate at sorting the present sites in the RF prediction output and had only 16 incorrectly sorted present and absent sites out of 70 sample sites. Though the Climatic model had the lowest MSE out of all three models, those scores test how accurately the model predicts the data it was given, meaning our Climate model did a great job in predicting the sites that we already knew about. The purpose of our distributional models was to predict the distribution of *A. mohri* outward into the rest of the state, which the Climate RF model does not do as well as the Landscape RF model, according to the other metrics used. The Climate model did correctly predict that *A. mohri* would not be present in the Ouachita Mountains ecoregion.

Elevation is known to be an important component for plecopterans, as stonefly presence increases as elevation increases [12,45,65,66], and this is even true for rare stonefly species, like *Beloneuria jamesae* [67]. Elevation has also been found to be an important factor in stonefly abundance [45], showing that mountain streams are beneficial for plecopterans as a whole. This link between elevation and species presence could be due to high elevation streams being a lower stream size order (1–3) [68] with riparian vegetation contributing allochthonous inputs to the stream, which shredding macroinvertebrates utilize. The forested canopy also shades the stream and keeps water temperatures cooler compared to areas further downstream [69]. Though we found that no elevation or ecoregion was significant in predicting *A. mohri* species presence, this may be due to the small range of elevation in our study (Table 1). In the future, studies should incorporate a wider range of elevation to determine if higher elevation streams are more likely to have *A. mohri*.

Catchment urban land cover percentages were found in the highest ranked model according to AIC (Table 4), and the Arkansas Valley ecoregion had the lowest percentage of urban land of all ecoregions (Figure 4A). As impervious cover, such as that found in urban land development of roads, houses, etc. increases, EPT richness decreases [70]. This is due in part to the nutrient additions that urban areas add to streams. Dissolved N and P additions to streams proliferate growth of decomposers, which will break down allochthonous materials, leaving less detrital material for the shredder species [34, 35]. This loss in EPT richness is also due to the lack of overhead tree canopy [60]. Forested habitat loss is one major driver of decline in stonefly populations [3], and if land in a watershed is transformed to majority urban, this could have devastating effects on the EPT community, including *A. mohri*.

Environmental temperature variables were found to be significant in predicting *A. mohri* presence (Table 4). Stoneflies have been shown to be sensitive to stream temperature, and as water temperatures increase, streams become less hospitable to stoneflies [71]. Water temperature can be important to winter stonefly life cycles. For example, diapausing eggs may require a thermal cue to hatch [12]. Streams with average July temperatures reaching 26.4°C did not tend to have *A. mohri* present (Figure 4C). It is possible that these temperatures did not get low enough to cue eggs or nymphs [12]. Mean January and July temperatures increased as one moves from the Ozark Highlands south to the Ouachita Mountains ecoregion (Figures 5B and C, respectively). The Ouachita Mountains streams had a higher minimum and maximum mean temperature across the year than other ecoregions, as well as a higher mean annual temperature (Figure 4D), possibly explaining the absence of *A. mohri*. Higher elevation streams in the region may be a refuge from warmer stream temperatures [72]. Future studies should endeavor to sample some higher elevation streams in the Ouachita ecoregions to see if they might be *A.*

*mohri* refuges. If these higher elevation streams act as shelter for temperature sensitive species, such as *Allocapnia mohri*, they might not be a haven for much longer. As climate change increases the temperatures of the vulnerable cool mountain streams, cold water adapted species like *Allocapnia mohri* will lose their thermal refuges [72] and eventually be replaced by more thermal generalist species, affecting the diversity of freshwater ecosystems [73–76].

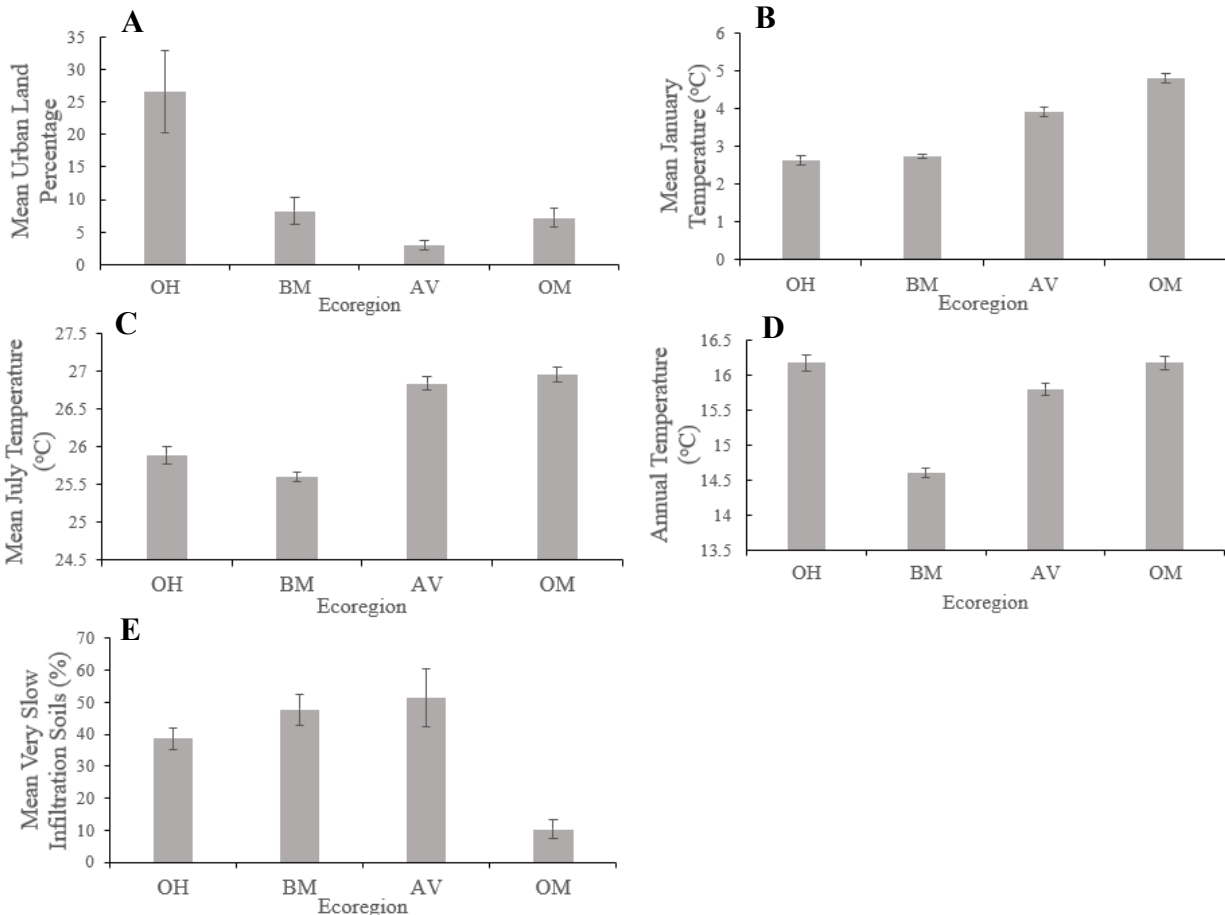


Figure 5. The mean + 1SE of Urban land use percentage (A), mean January (B), July (C) and annual (D) stream temperatures during our study period and percentage of very slow infiltration soils (E) in study reach catchments grouped by ecoregions. Ecoregions move from north (left) to south (right). OH = Ozark Highlands, BM = Boston Mountains, AV= Arkansas Valley, OM = Ouachita Mountains.

Catchment soil infiltration category percentages were found to be significant in predicting *A.*

*mohri* presence (Table 4). This result may be due to the inherent soil texture found in each

ecoregion. Soil texture refers to the percentage of sand, silt, and clay, which are the major factors

affecting water infiltration. Species presence dropped to zero in the Ouachita Mountains ecoregion, which had the highest infiltration values of all visited ecoregions (Figure 5D). Soil in the Arkansas Valley, Boston Mountains, and Ozark High-lands ecoregions had a higher proportion of clay (a very small-pored soil) compared to the Ouachita Mountains, which has more sandy soil. Therefore, this correlation between very slow infiltration and species presence could just be due to the correlation between soil type and ecoregion.

## V. Conclusions

Our resulting RF SDMs are some of the first published for a North American plecopteran species. Though the landscape RF model performed better than the climate or combined RF models, our MLR modeling indicates that both climate and landscape factors are important habitat variables controlling *A. mori* distributions. This discrepancy could be due to the differing types of data used by the RF and MLR models, and the general approach that these models used. The RF models used rasters that had gridded satellite data, whereas the MLR data was watershed-specific, higher resolution data, and this could have made the RF models less reliable overall. Also, since the RF models used rasters consisting of large sets of data, the amount of predictor variables in that set of data were not reduced before creating a distributional map. When developing the MLR models, however, it was easier to determine which precise variables were significant to species presence, and which variables were diminishing the accuracy of the model. As high-resolution satellite imaging becomes more readily available, more accurate rasters including landscape and climate variables can be used to improve upon these species distribution models.

Even though the landscape-level soil infiltration categories led to a strong MLR model, these variables have likely not changed much over the last thirty years as *A. mohri* shifted from present

to absent in censuses occurring in the Ouachita Mountains. The Landscape RF model also clearly predicts *A. mohri* should be present there, but the Climate RF model does not. Additional surveys for adults in the Ouachita Mountains stream reaches within predicted presence watersheds based on the RF model, particularly those with greater maximum elevation, could help determine if these reaches may be a refuge for *A. mohri* populations.

Climate change and associated warming stream waters can be a significant threat to macroinvertebrate taxa and can be responsible for species losses [3,65,71,77]. Warming stream temperatures may be causing losses of *A. mohri* in streams at the southern edge of its range (e.g., the Ouachita Mountains), shifting its distributions northwards. Evidence of range shifts due to effects of climate change has been observed in other macroinvertebrate species [76,78], and could be expected to occur for sensitive species in a continually warming climate.

## VI. References

1. DeWalt, R.E.; Ower, G.D. Ecosystem Services, Global Diversity, and Rate of Stonefly Species Descriptions (Insecta: Plecoptera). *Insects* **2019**, *10*, 99.  
<https://doi.org/10.3390/insects10040099>.
2. Fochetti, R.; De Figueroa, J.M.T. Notes on diversity and conservation of the European fauna of Plecoptera (Insecta). *J. Nat. Hist.* **2006**, *40*, 2361–2369.  
<https://doi.org/10.1080/00222930601051386>.
3. Sánchez-Bayo, F.; Wyckhuys, K. Worldwide decline of the entomofauna: A review of its drivers. *Biol. Conserv.* **2019**, *232*, 8–27. <https://doi.org/10.1016/j.biocon.2019.01.020>.

4. Allan, J.D.; Castillo, M.M.; Capps, K.A. *Stream Ecology: Structure and Function of Running Waters*, 3th ed.: Springer: Cham, Switzerland, 2021.
5. Fisher, S.G.; Likens, G.E. Energy Flow in Bear Brook, New Hampshire: An Integrative Approach to Stream Ecosystem Metabolism. *Ecol. Monogr.* **1973**, *43*, 421–439.  
<https://doi.org/10.2307/1942301>.
6. Hornick, L.E.; Webster, J.R.; Benfield, E.F. Periphyton Production in an Appalachian Mountain Trout Stream. *Am. Midl. Nat.* **1981**, *106*, 22. <https://doi.org/10.2307/2425132>.
7. Martínez, A.; Larrañaga, A.; Pérez, J.; Basaguren, A.; Pozo, J. Leaf-litter quality effects on stream ecosystem functioning: A comparison among five species. *Fundam. Appl. Limnol.* **2013**, *183*, 239–248. <https://doi.org/10.1127/1863-9135/2013/0514>.
8. Perry, W.B.; Benfield, E.F.; Perry, S.A.; Webster, J.R.; Energetics, Growth, and Production of a Leaf-Shredding Stonefly in an Appalachian Mountain Stream. *J. North Am. Benthol. Soc.* **1987**, *6*, 12–25. <https://doi.org/10.2307/1467520>.
9. Thorp, J.H.; Rogers, D.C. Chapter 22—Stoneflies: Insect Order Plecoptera. In *Field Guide to Freshwater Invertebrates of North America*; Thorp, J.H., Rogers, D.C., Eds.; Academic Press: Boston, MA, USA, 2011; pp. 199–204.

10. Zwick, P. Phylogenetic System and Zoogeography of the Plecoptera. *Annu. Rev. Entomol.* **2000**, *45*, 709–746. <https://doi.org/10.1146/annurev.ento.45.1.709>.
11. Maguire, T.J.; Mundle, S.O.C. Citizen Science Data Show Temperature-Driven Declines in Riverine Sentinel Invertebrates. *Environ. Sci. Technol. Lett.* **2020**, *7*, 303–307. <https://doi.org/10.1021/acs.estlett.0c00206>.
12. Poulton, B.C.; Stewart, K.W. The stoneflies of the Ozark and Ouachita Mountains (Plecoptera). *Mem. Am. Entomol. Soc.* **1991**, *38*, 1–116.
13. IPCC, 2013: *Climate Change 2013: The Physical Science Basis. Contribution of Working Group I to the Fifth Assessment Report of the Intergovernmental Panel on Climate Change*; Cambridge University Press: Cambridge, UK; New York, NY, USA, 2013.
14. Sánchez-Bayo, F.; Wyckhuys, K.A.G. Further evidence for a global decline of the entomofauna. *Austral Entomol.* **2020**, *60*, 9–26. <https://doi.org/10.1111/aen.12509>.
15. Stanford, J.A.; Gaufin, A.R. Hyporheic Communities of Two Montana Rivers. *Science* **1974**, *185*, 700–702. <https://doi.org/10.1126/science.185.4152.700>.
16. Lancaster, J.; Hildrew, A.G. Flow Refugia and the Microdistribution of Lotic Macroinvertebrates. *J. North Am. Benthol. Soc.* **1993**, *12*, 385–393.

<https://doi.org/10.2307/1467619>.

17. Pusey, B.J.; Arthington, A. Importance of the riparian zone to the conservation and management of freshwater fish: A review. *Mar. Freshw. Res.* **2003**, *54*, 1–16.  
<https://doi.org/10.1071/mf02041>.
18. Burdon, F.J.; McIntosh, A.; Harding, J.S. Habitat loss drives threshold response of benthic invertebrate communities to deposited sediment in agricultural streams. *Ecol. Appl.* **2013**, *23*, 1036–1047. <https://doi.org/10.1890/12-1190.1>.
19. Wallace, J.B.; Eggert, S.L.; Meyer, J.L.; Webster, J.R. Effects of Resource Limitation on a Detrital-Based Ecosystem. *Ecol. Monogr.* **1999**, *69*, 409–442. [https://doi.org/10.1890/0012-9615\(1999\)069\[0409:EORLOA\]2.0.CO;2](https://doi.org/10.1890/0012-9615(1999)069[0409:EORLOA]2.0.CO;2).
20. Cross, W.F.; Wallace, J.B.; Rosemond, A.D.; Eggert, S.L. Whole-System Nutrient Enrichment Increases Secondary Production in a Detritus-Based Ecosystem. *Ecology* **2006**, *87*, 1556–1565. [https://doi.org/10.1890/0012-9658\(2006\)87\[1556:WNEISP\]2.0.CO;2](https://doi.org/10.1890/0012-9658(2006)87[1556:WNEISP]2.0.CO;2).
21. Likens, G.E.; Bormann, F.H.; Johnson, N.M.; Fisher, D.W.; Pierce, R.S. Effects of Forest Cutting and Herbicide Treatment on Nutrient Budgets in the Hubbard Brook Watershed-Ecosystem. *Ecol. Monogr.* **1970**, *40*, 23–47. <https://doi.org/10.2307/1942440>.



22. Lee, J.-W.; Lee, S.-W.; An, K.-J.; Hwang, S.-J.; Kim, N.-Y. An Estimated Structural Equation Model to Assess the Effects of Land Use on Water Quality and Benthic Macroinvertebrates in Streams of the Nam-Han River System, South Korea. *Int. J. Environ. Res. Public Health* **2020**, *17*, 2116. <https://doi.org/10.3390/ijerph17062116>.
23. Waschbusch, R.J.; Selbig, W.; Bannerman, R.T. Sources of phosphorus in stormwater and street dirt from two urban residential basins in Madison. *Wisconsin* **1994**, *95*, 1999. <https://doi.org/10.3133/wri994021>.
24. Omernik, J.M. *The Influence of Land Use on Stream Nutrient Levels*; US Environmental Protection Agency, Office of Research and Development, Corvallis Environmental Research Laboratory, Eutrophication Survey Branch; Corvallis, Oregon USA; 1976.
25. Paul, M.J.; Meyer, J.L. Streams in the Urban Landscape. In *Urban Ecology: An International Perspective on the Interaction Between Humans and Nature*; Marzluff, J.M.; Shulenberger, E.; Endlicher, W.; Alberti, M.; Bradley, G.; Ryan, C.; ZumBrunnen, C.; Simon, U., Eds.; Springer US: Boston, MA, USA, 2008; pp. 207–231.
26. Booth, D.B.; Jackson, C.R. URBANIZATION OF AQUATIC SYSTEMS: DEGRADATION THRESHOLDS, STORMWATER DETECTION, AND THE LIMITS OF MITIGATION. *JAWRA J. Am. Water Resour. Assoc.* **1997**, *33*, 1077–1090. <https://doi.org/10.1111/j.1752-1688.1997.tb04126.x>.

27. Freeborn, J.R.; Sample, D.J.; Fox, L.J. RESIDENTIAL STORMWATER: METHODS FOR DECREASING RUNOFF AND INCREASING STORMWATER INFILTRATION. *J. Green Build.* **2012**, *7*, 15–30. <https://doi.org/10.3992/jgb.7.2.15>.
28. King, R.S.; Scoggins, M.; Porras, A. Stream biodiversity is disproportionately lost to urbanization when flow permanence declines: Evidence from southwestern North America. *Freshw. Sci.* **2016**, *35*, 340–352. <https://doi.org/10.1086/684943>.
29. Dodds, W.K. Trophic state, eutrophication and nutrient criteria in streams. *Trends Ecol. Evol.* **2007**, *22*, 669–676. <https://doi.org/10.1016/j.tree.2007.07.010>.
30. Stringfellow, W.; Herr, J.; Litton, G.; Brunell, M.; Borglin, S.; Hanlon, J.; Chen, C.; Graham, J.; Burks, R.; Dahlgren, R.; et al. Investigation of river eutrophication as part of a low dissolved oxygen total maximum daily load implementation. *Water Sci. Technol.* **2009**, *59*, 9–14. <https://doi.org/10.2166/wst.2009.739>.
31. Suberkropp, K. Annual production of leaf-decaying fungi in a woodland stream. *Freshw. Biol.* **1997**, *38*, 169–178. <https://doi.org/10.1046/j.1365-2427.1997.00203.x>.
32. Evans-White, M.A.; Haggard, B.E.; Scott, J.T. A Review of Stream Nutrient Criteria Development in the United States. *J. Environ. Qual.* **2013**, *42*, 1002–1014. <https://doi.org/10.2134/jeq2012.0491>.

33. Halvorson, H.M.; Scott, J.T.; Sanders, A.J.; Evans-White, M.A. A stream insect detritivore violates common assumptions of threshold elemental ratio bioenergetics models. *Freshw. Sci.* **2015**, *34*, 508–518. <https://doi.org/10.1086/680724>.
34. Howard-Parker, B.; White, B.; Halvorson, H.M.; Evans-White, M.A. Light and dissolved nutrients mediate recalcitrant organic matter decomposition via microbial priming in experimental streams. *Freshw. Biol.* **2020**, *65*, 1189–1199. <https://doi.org/10.1111/fwb.13503>.
35. Eckert, R.A.; Halvorson, H.M.; Kuehn, K.A.; Lamp, W.O. Macroinvertebrate community patterns in relation to leaf-associated periphyton under contrasting light and nutrient conditions in headwater streams. *Freshw. Biol.* **2020**, *65*, 1270–1287. <https://doi.org/10.1111/fwb.13473>.
36. Rosemond, A.D.; Benstead, J.P.; Bumpers, P.M.; Gulis, V.; Kominoski, J.S.; Manning, D.W.P.; Suberkropp, K.; Wallace, J.B. Experimental nutrient additions accelerate terrestrial carbon loss from stream ecosystems. *Science* **2015**, *347*, 1142–1145. <https://doi.org/10.1126/science.aaa1958>.
37. Stewart, K.W.; Stark, B.P. Plecoptera (Chapter 14). Pp 311-384. In *An Introduction to the Aquatic Insects of North America*, 4th ed.; Merritt, R.W., Cummins, K.W., Berg, M.B., Ed.; Kendall/Hunt Publishing Company, Dubuque, Iowa, USA, 2007; 1158p.

38. Ab Hamid, S.; Rawi, C.S. Application of Aquatic Insects (Ephemeroptera, Plecoptera and Trichoptera) in Water Quality Assessment of Malaysian Headwater. *Trop. Life Sci. Res.* **2017**, *28*, 143–162. <https://doi.org/10.21315/tlsr2017.28.2.11>.
39. McCafferty, W.P.; Provonsha, A.V. *Aquatic Entomology: The Fishermen's and Ecologists' Illustrated Guide to Insects and Their Relatives*; Jones and Barlett Publishers. Inc.: Boston, MA, USA, 1983.
40. Guisan, A.; Zimmermann, N.E. Predictive habitat distribution models in ecology. *Ecol. Model.* **2000**, *135*, 147–186. [https://doi.org/10.1016/s0304-3800\(00\)00354-9](https://doi.org/10.1016/s0304-3800(00)00354-9).
41. Guisan, A.; Thuiller, W. Predicting species distribution: Offering more than simple habitat models. *Ecol. Lett.* **2005**, *8*, 993–1009. <https://doi.org/10.1111/j.1461-0248.2005.00792.x>.
42. Breiman, L. Random forests. *Mach. Learn* **2001**, *45*, 5–32. <https://doi.org/10.1023/A:1010933404324>.
43. Breiman, L. (1984). *Classification and Regression Trees*. United Kingdom: CRC Press.
44. Hernandez, P.A.; Franke, I.; Herzog, S.K.; Pacheco, V.; Paniagua, L.; Quintana, H.L.; Soto, A.; Swenson, J.J.; Tovar, C.; Valqui, T.; et al. Predicting species distributions in poorly-studied landscapes. *Biodivers. Conserv.* **2008**, *17*, 1353–1366.

<https://doi.org/10.1007/s10531-007-9314-z>.

45. Grubbs, S.; Sheldon, A.L. The stoneflies (Insecta, Plecoptera) of the Talladega Mountain region, Alabama, USA: Distribution, elevation, endemism, and rarity patterns. *Biodivers. Data J.* **2018**, *6*, e22839. <https://doi.org/10.3897/bdj.6.e22839>.
46. Ross, H.H.; Ricker, W.E. *The Classification, Evolution, and Dispersal of the Winter Stonefly Genus Allocapnia*; University of Illinois Press: Urbana, IL, USA, 1971; Volume 45.
47. Merritt, R.W.; Cummins, K.W.; Berg, M.B. *An Introduction to the Aquatic Insects of North America*, 5th Ed.; Kendall/Hunt Publishing Company: USA, 2019.
48. Cao, Y.; DeWalt, R.E.; Robinson, J.L.; Tweddale, T.; Hinz, L.; Pessino, M. Using Maxent to model the historic distributions of stonefly species in Illinois streams: The effects of regularization and threshold selections. *Ecol. Model.* **2013**, *259*, 30–39. <https://doi.org/10.1016/j.ecolmodel.2013.03.012>.
49. Leasure, D.R.; Magoulick, D.D.; Longing, S.D. Natural Flow Regimes of the Ozark-Ouachita Interior Highlands Region. *River Res. Appl.* **2014**, *32*, 18–35. <https://doi.org/10.1002/rra.2838>.

50. Newbury, R.; Bates, D.; Alex, K.L. Restoring Habitat Hydraulics with Constructed Riffles. *Earth Space Sci* **2013**, *194*, 353–366. <https://doi.org/10.1029/2010gm000978>.
51. Gore, J.A. *Chapter 3—Discharge Measurements and Streamflow Analysis*; Academic Press: Cambridge, MA, USA, 2006.
52. Sheldon, A.L.; Jr, M.L. Warren. Filters and templates: Stonefly (Plecoptera) richness in Ouachita Mountains streams, U.S.A. *Freshw. Biol.* **2009**, *54*, 943–956. <https://doi.org/10.1111/j.1365-2427.2008.02144.x>.
53. Stroud Water Research Center. (2020). Model My Watershed [Software]. Available online: <https://wikiwatershed.org/> (accessed on 9 December 2022).
54. Dewitz, J. *National Land Cover Dataset (NLCD) 2016 Products*; U.S. Geological Survey Data Release: 2019. Reston, Virginia. Available online: <https://doi.org/10.5066/p96hhbie> (accessed on 15 December 2022).
55. Soil Survey Staff, 2020. The Gridded National Soil Survey Geographic (gNATSGO) Database for Arkansas. United States Department of Agriculture, Natural Resources Conservation Service. Available online: <https://nrcs.app.box.com/v/soils> (accessed on 10 December 2022).

56. Fick, S.E.; Hijmans, R.J. WorldClim 2: New 1-km spatial resolution climate surfaces for global land areas. *Int. J. Climatol.* **2017**, *37*, 4302–4315. <https://doi.org/10.1002/joc.5086>.
57. Tarter, D.C.; Chaffee, D.L.; Grubbs, S.A. Revised Checklist of The Stoneflies (Plecoptera) Of Kentucky, U.S.A. *Entomol. News* **2006**, *117*, 1–10. [https://doi.org/10.3157/0013-872X\(2006\)117\[1:RCOTSP\]2.0.CO;2](https://doi.org/10.3157/0013-872X(2006)117[1:RCOTSP]2.0.CO;2).
58. McRoberts, T.; Grubbs, S. Effects of stream permanence on stonefly (Insecta, Plecoptera) community structure at Mammoth Cave National Park, Kentucky, USA. *Biodivers. Data J.* **2021**, *9*, e62242. <https://doi.org/10.3897/bdj.9.e62242>.
59. *ESRI, 2020*; ArcGIS Pro Version 2.7.2: Redlands, CA USA, 2020.
60. Chicco, D.; Jurman, G. The advantages of the Matthews correlation coefficient (MCC) over F1 score and accuracy in binary classification evaluation. *BMC Genom.* **2020**, *21*, 6. <https://doi.org/10.1186/s12864-019-6413-7>.
61. Boughorbel, S.; Jarray, F.; El Anbari, M. Optimal classifier for imbalanced data using Matthews Correlation Coefficient metric. *PLoS ONE* **2017**, *12*, e0177678. <https://doi.org/10.1371/journal.pone.0177678>.
62. R Core Team. *R: A Language and Environment for Statistical Computing*; R Foundation for Statistical Computing: Vienna, Austria, 2019. Available online: <https://www.R-project.org/>

(accessed on 4 December 2022).

63. Reid, A.J.; Carlson, A.K.; Creed, I.F.; Eliason, E.J.; Gell, P.A.; Johnson, P.T.J.; Kidd, K.A.; MacCormack, T.J.; Olden, J.D.; Ormerod, S.J.; et al. Emerging threats and persistent conservation challenges for freshwater biodiversity. *Biol. Rev.* **2019**, *94*, 849–873.  
<https://doi.org/10.1111/brv.12480>.
64. Snellen, R.K.; Stewart, K.W. The Life Cycle of *Perlesta placida* (Plecoptera: Perlidae) in an Intermittent Stream in Northern Texas<sup>1</sup>. *Ann. Entomol. Soc. Am.* **1979**, *72*, 659–666.  
<https://doi.org/10.1093/aesa/72.5.659>.
65. Sheldon, A.L. Possible climate-induced shift of stoneflies in a southern Appalachian catchment. *Freshw. Sci.* **2012**, *31*, 765–774. <https://doi.org/10.1899/11-135.1>.
66. Garcia-Raventós, A.; Viza, A.; de Figueroa, J.M.T.; Riera, J.L.; Múrria, C. Seasonality, species richness and poor dispersion mediate intraspecific trait variability in stonefly community responses along an elevational gradient. *Freshw. Biol.* **2017**, *62*, 916–928.  
<https://doi.org/10.1111/fwb.12912>.
67. Sheldon, A.L.; Grubbs, S.A. Distributional ecology of a rare, endemic stonefly. *Freshw. Sci.* **2014**, *33*, 1119–1126. <https://doi.org/10.1086/678049>.



68. Strahler, A.N. Quantitative analysis of watershed geomorphology. *Eos Trans. Am. Geophys. Union* **1957**, *38*, 913–920. <https://doi.org/10.1029/tr038i006p00913>.
69. Vannote, R.L.; Minshall, G.W.; Cummins, K.W.; Sedell, J.R.; Cushing, C.E. The River Continuum Concept. *Can. J. Fish. Aquat. Sci.* **1980**, *37*, 130–137. <https://doi.org/10.1139/f80-017>.
70. King, R.S.; Scoggins, M.; Porras, A. (2016). Stream biodiversity is disproportionately lost to urbanization when flow permanence declines: evidence from southwestern North America. *Freshwater Science*, *35*(1), 340–352. doi:10.1086/684943
71. Hotaling, S.; Shah, A.A.; Dillon, M.E.; Giersch, J.J.; Tronstad, L.M.; Finn, D.S.; Woods, H.A.; Kelley, J.L. Cold Tolerance of Mountain Stoneflies (Plecoptera: Nemouridae) from the High Rocky Mountains. *West. North Am. Nat.* **2021**, *81*, 54–62. <https://doi.org/10.3398/064.081.0105>.
72. Mulholland, P.J.; Marzolf, E.R.; Webster, J.R.; Hart, D.; Hendricks, S.P. Evidence that hyporheic zones increase heterotrophic metabolism and phosphorus uptake in forest streams. *Limnol. Oceanogr.* **1997**, *42*, 443–451. <https://doi.org/10.4319/lo.1997.42.3.0443>.
73. Burgmer, T.; Hillebrand, H.; Pfenninger, M. Effects of climate-driven temperature changes on the diversity of freshwater macroinvertebrates. *Oecologia* **2006**, *151*, 93–103.

<https://doi.org/10.1007/s00442-006-0542-9>.

74. Durance, I.; Ormerod, S.J. Climate change effects on upland stream macroinvertebrates over a 25-year period. *Glob. Chang. Biol.* **2007**, *13*, 942–957. <https://doi.org/10.1111/j.1365-2486.2007.01340.x>.
75. Hering, D.; Schmidt-Kloiber, A.; Murphy, J.; Lücke, S.; Zamora-Muñoz, C.; López-Rodríguez, M.J.; Huber, T.; Graf, W. Potential impact of climate change on aquatic insects: A sensitivity analysis for European caddisflies (Trichoptera) based on distribution patterns and ecological preferences. *Aquat. Sci.* **2009**, *71*, 3–14. <https://doi.org/10.1007/s00027-009-9159-5>.
76. Domisch, S.; Jähnig, S.C.; Haase, P. Climate-change winners and losers: Stream macroinvertebrates of a submontane region in Central Europe. *Freshw. Biol.* **2011**, *56*, 2009–2020. <https://doi.org/10.1111/j.1365-2427.2011.02631.x>.
77. Shokri, M.; Cozzoli, F.; Vignes, F.; Bertoli, M.; Pizzul, E.; Basset, A. Metabolic rate and climate change across latitudes: Evidence of mass-dependent responses in aquatic amphipods. *J. Exp. Biol.* **2022**, *225*, jeb244842. <https://doi.org/10.1242/jeb.244842>.
78. Péliissié, M.; Johansson, F.; Hyseni, C. Pushed Northward by Climate Change: Range Shifts With a Chance of Co-occurrence Reshuffling in the Forecast for Northern European Odonates. *Environ. Èntomol.* **2022**, *51*, 910–921. <https://doi.org/10.1093/ee/nvac056>.

