



## The role of surfactants and biosurfactants in the wound healing process

Ohadi, M., Forootanfar, H., Dehghannoudeh, N., Banat, I. M., & Dehghannoudeh, G. (2023). The role of surfactants and biosurfactants in the wound healing process: a review. *Journal of Wound Care*, 32(Sup4a), xxxix-xlvi. Advance online publication. <https://doi.org/10.12968/jowc.2023.32.Sup4a.xxxix>

[Link to publication record in Ulster University Research Portal](#)

**Published in:**  
Journal of Wound Care

**Publication Status:**  
Published (in print/issue): 08/04/2023

**DOI:**  
[10.12968/jowc.2023.32.Sup4a.xxxix](https://doi.org/10.12968/jowc.2023.32.Sup4a.xxxix)

**Document Version**  
Author Accepted version

**General rights**  
Copyright for the publications made accessible via Ulster University's Research Portal is retained by the author(s) and / or other copyright owners and it is a condition of accessing these publications that users recognise and abide by the legal requirements associated with these rights.

**Take down policy**  
The Research Portal is Ulster University's institutional repository that provides access to Ulster's research outputs. Every effort has been made to ensure that content in the Research Portal does not infringe any person's rights, or applicable UK laws. If you discover content in the Research Portal that you believe breaches copyright or violates any law, please contact [pure-support@ulster.ac.uk](mailto:pure-support@ulster.ac.uk).

1 **The role of surfactants and biosurfactants in wound healing process: A Review**

2

3 Mandana Ohadi<sup>a</sup>, Hamid Forootanfar<sup>b</sup>, Negar Dehghannoudeh<sup>c</sup>, Ibrahim M Banat<sup>d</sup>, Gholamreza  
4 Dehghannoudeh<sup>a\*</sup>

5

6

7 *<sup>a</sup>Pharmaceutics Research Center, Institute of Neuropharmacology, Kerman University of Medical*

8 *Sciences, Kerman, Iran*

9 *<sup>b</sup>Pharmaceutical Sciences and Cosmetic Products Research Center, Kerman University of*

10 *Medical Sciences, Kerman, Iran*

11 *<sup>c</sup> Faculty of Arts and Science, University of Toronto, Toronto, Ontario, Canada*

12 *<sup>d</sup>School of Biomedical Sciences, Faculty of Life & Health Sciences, University of Ulster, Coleraine*

13 *BT52 ISA, Northern Ireland, UK*

14

---

15 *\*Corresponding author: Tel.: +98-34-31325015; fax: +98-34-31325003 email: addresses:*

16 *ghr\_dehghan@kmu.ac.ir (G. Dehghan-Noudeh), gholamreza.dehghannoudeh@utoronto.ca.*

17

18

19

20

21

22

23

24

25

26

27

28

29

30

31 **Graphical Abstract**

32

33

34

35

36

37

38

39

40

41

42

43

44

45

46

47

48

49

50

51

52

53

54

55

56

57

58

59

60

61

62

63

64

65

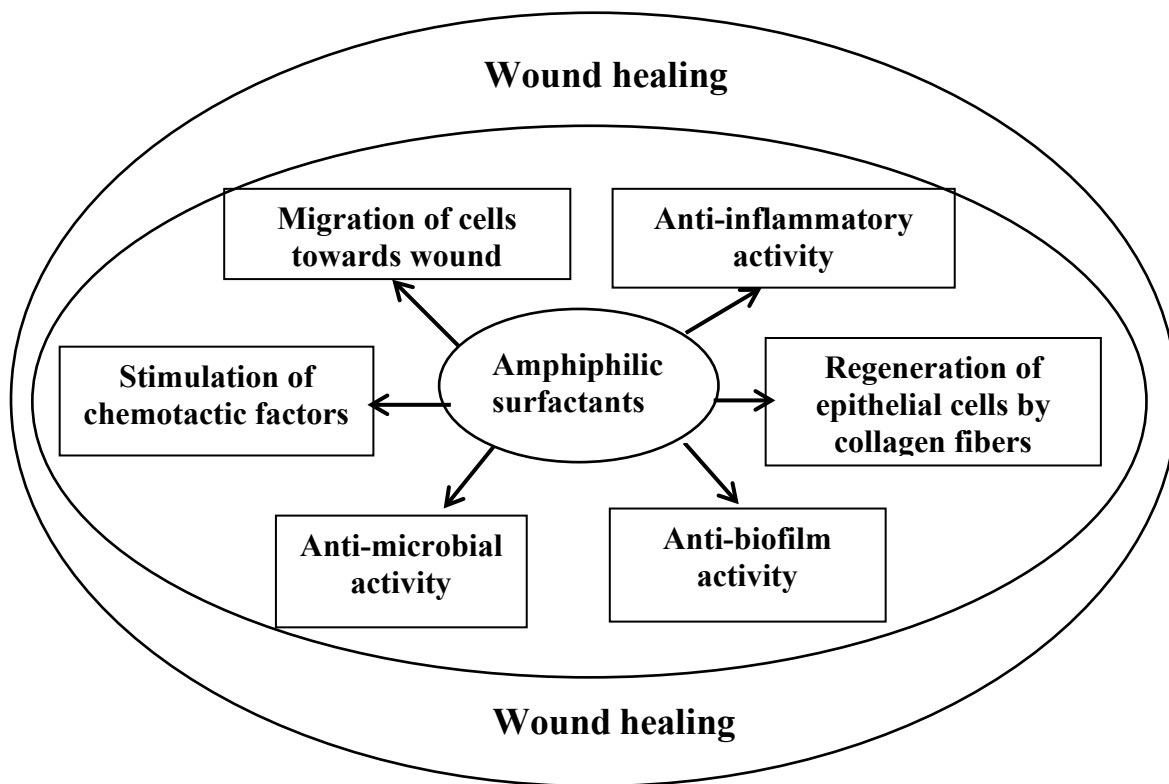
66

67

68

69

70



71 **Abstract**

72  
73 Wound healing refers to the complex process of restoring the forms and functions of damaged  
74 tissues. Multiple growth factors and released cytokines tightly regulate the site of the wound. The  
75 healing processes can be disrupted by any alteration that would aggravate the damage and lengthen  
76 the repair process. Some of the conditions that may impair wound healing include infections and  
77 inflammation. Surfactants are amphiphilic compounds widely used in various formulations  
78 including detergents, food, pharmaceutical, and cosmetic industries. Biosurfactants therefore are  
79 surface-active compounds produced by biological agents particularly yeast or bacteria and  
80 represent a safer and environmentally preferred alternative to chemical surfactants. Numerous  
81 studies have targeted surface-active molecules as wound healing agents for their anti-  
82 inflammatory, antioxidant, and antibacterial potential. This review focuses on surface-active  
83 molecules used in wound healing activities and analyzes their effectiveness and mechanisms of  
84 action.

85 **Keywords:** Surfactants; Wound healing; Adjuvant; Biosurfactants

86  
87  
88  
89  
90  
91  
92  
93  
94  
95  
96  
97  
98  
99  
100  
101  
102  
103

104 **Introduction:**

105

106 A wound may be considered as damage to the continuity of the skin epithelium or mucosa caused

107 by physical or thermal injury or a disease. Tissue regeneration and development which results in

108 wound healing takes place in four overlapping stages, including haemostasis and inflammation,

109 migration, proliferation, and maturation [1]. The stages and their biochemical and physiological

110 effects occurs in a specific sequence, at a distinct time, continuing for a specific duration at an

111 optimum intensity [2]. Wound healing is affected by several factors and interfering with one or

112 more stages in the process, leads to inappropriate or impaired tissue repair. Considering the

113 duration and nature of the repair process, wounds can be identified as acute or chronic wounds [3].

114 The former is an injury to the skin occurring suddenly due to accidents, burns and chemical or

115 mechanical injuries. They usually heal in a predictable timeframe commonly through 8 - 12 weeks

116 according to the severity or depth of the damage. Chronic wounds are those that usually cannot be

117 healed within the expected time frame of 12 weeks and often reoccur due to disruptions in the

118 orderly sequence of wound healing stages. These wounds cannot be healed due to repeated tissue

119 insults or physiological reasons, such as diabetes, malignancies, persistent infections, improper

120 primary therapy and other parameters linked to the patient. Chronic wounds include diabetic foot,

121 decubitus and leg ulcers [3]. It is expected to use 1–3% of the drugs indexed in western

122 pharmacopoeias for topical use on wounds [4]. There are several medications and ointments for

123 wound healing worldwide, of which surfactants are widely applied for removing debris, indicating

124 their function as cleaners. Surfactants for wound healing have been long used and can be found in

125 several wound cleansers used for wound cleaning and irrigation/hydration [5]. They act due to

126 their micelles' formation abilities, in which polymer chains including the hydrophilic head and

127 hydrophobic tail generate a hydrophilic outer shell and hydrophobic center. Such chemistry is

128 helpful for several cleaning processes [6]. Nonetheless, reports on using surfactants against other  
129 delayed factors affecting wound healing is somewhat limited. Therefore, this review will explore  
130 the literature surfactants' uses and their mods of action in managing the wound healing process.

131  
132 **Factors affecting wound healing**

133  
134 Bacterial biofilms seem to have a major role, their effects on chronic wounds have been widely  
135 considered in recent literature. They can be seen in 60 to 80% of wounds, and based on a previous  
136 meta-analysis; their presence is confirmed in 78.2% of chronic wounds [6]. They can persist in  
137 chronic wounds causing prolonged inflammation and consequently, delayed healing and enhance  
138 the risk of infection. Thus, they are crucial in the majority of chronic non-healing skin wounds [5,  
139 6]. Pathogenic bacteria like *Pseudomonas aeruginosa*, *Staphylococcus aureus*, *Streptococcus*  
140 *pyogenes* and others such as *Proteus*, *Clostridium* and *Coliform* species are detrimental to wound  
141 healing. Improper measures for the management of infected wounds result in cellulitis (cell  
142 inflammation), bacteraemia and septicaemia which can be fatal [7]. The formation of hypertrophic  
143 scars due to increased collagen synthesis if the cells persist at the site in the last phase of the wound  
144 healing process is also challenging [8]. On the other hand, wound healing may be delayed due to  
145 the generation of reactive oxygen species (ROS) and reactive nitrogen species (RNS) at the wound  
146 area because of oxidative stresses, lipid oxidation, DNA damage, and inactivation of enzymes  
147 involved in free radical scavenging. Therefore, wound healing drugs with antioxidant activity are  
148 often used [9, 10].

149 **Synthetic versus natural surfactants**

150 Chemically synthesized surfactants are surface-active amphiphilic compounds capable of reducing  
151 the surface tension between liquids and the different state of matter; gas, liquid and solid. They do  
152 this by forming micellar structures encompassing the hydrophilic and hydrophobic parts of these

153 molecules [11]. This results in increases of the surface humidity and materials solubility or  
154 miscibility that would otherwise not be possible without. Surfactants can be categorized according  
155 to their behavior in aqueous solution, in which each category is defined according to the charge on  
156 the hydrophilic head of the surfactant molecule. Cationic surfactants have a positive charge (e.g.  
157 quaternary ammonium compounds), anionic surfactants have a negative charge (e.g. soap,  
158 detergents, sodium dodecyl sulfate), non-ionic are uncharged (e.g. poloxamer, Tween 80, Triton-  
159 X) and amphoteric surfactants (e.g. Betaine) simultaneously carrying an anionic and a cationic  
160 hydrophilic group which are able to form cation or anion depending on pH changes and ambient  
161 conditions [12].

162 Natural surfactants also known as ‘biosurfactants’ have several amphipathic molecules  
163 characterized by special chemical structures naturally synthesized by different microorganisms  
164 [13]. However, as opposed to chemically produced surfactants, they are categorized according  
165 their head group which is often a sugar or protein molecule, chemical composition or overall  
166 molecular weight. Low molecular weights include glycolipids and lipopeptides while high  
167 molecular weight include polysaccharides, proteins, and lipoproteins surfactants [14]. Generally,  
168 the amphiphilic and polyphilic high molecular weight polymers have been shown to be more useful  
169 in stabilizing emulsions acting as emulsifiers, whereas the low molecular weight ones with simpler  
170 structures have better abilities to reduce surface activity. Hydrophilic moiety of natural surfactants  
171 commonly includes an acid, peptide cations or anions, mono-, di- or polysaccharides, whereas  
172 unsaturated or saturated hydrocarbons or fatty acids typically represent their hydrophobic moiety.  
173 Glycolipids and lipopeptides are the main microbial surfactants that have been investigated and  
174 explored [13]. The former includes rhamnolipids produced by *Pseudomonas aeruginosa*,  
175 sophorolipids produced by *Candida/Starmerella bombicola* and mannosylerythritol lipids

176 synthesized by *Ustilago* sp. or *Pseudozyma* yeasts with mono- or disaccharides associated with  
177 long-chain aliphatic acids or hydroxyaliphatic acids. The latter also includes surfactin, iturin, and  
178 fengicyn cyclic lipopeptides generated by *Bacillus* species as antibiotic or antimicrobial molecules  
179 [12, 14].

## 180 **Use of synthetic surfactants in wound healing**

### 181 *Surfactants as wound cleaning*

182 Surfactants are added to wounds as a hydration/irrigation agents and cleansers incorporated into  
183 surgical scrub solutions. The former includes surfactants that can properly clean and remove debris  
184 from the wound. An investigation involving 289 patients to compare the use of betaine surfactant-  
185 based saline solution impacts on bed preparation and inflammation in chronic wounds showed  
186 decreased inflammation and increased granulation tissue production and wound closure [15]. A  
187 retrospective assessment comparing the same hydration solution achieved by Ringer's solution or  
188 saline with and without surfactant on venous leg ulcers healing level, reported a 97% faster healing  
189 level in the surfactant (Betaine) containing solutions as oppose to controls without surfactants [16].  
190 Studies have also indicated the effectiveness of surfactant-containing irrigation solutions in wound  
191 cleansing, demonstrating their importance in standard care protocols whether used with or without  
192 antimicrobial agents [6, 16]. In another investigation carried out by Burnett. et al [17] surfactants  
193 containing betaine were shown to facilitate wound cleansing and autolytic debridement, as well as  
194 supporting wound healing at the cellular level.

### 195 *Surfactants as antibiofilm agent*

196 The presence of biofilms in wounds can reduce healing rates and increase the chances of  
197 reinfection which often results in the development of chronic wounds [6]. Biofilms are able to  
198 form rapidly in wounds, as demonstrated by Kennedy et al. [18] who visualized biofilm formation



199 in burn wounds during 7–31 days post injury. Nakagami et al. [19] also demonstrated early signs  
200 of biofilm formation in infected wounds 3–7 days post injury. Biofilms have also been shown to  
201 cause chronic inflammation in wounds, as the elevated levels of cytokines produced by  
202 macrophages in response to the biofilm typically leads to increased participation/accumulation of  
203 immune cells at the site [20]. This causes the over-production of proteases and ROS, which break  
204 down the proteins involved in the wound-healing process [21].

205 There are several examples of synthetic surfactants used in wound care with the most well-  
206 researched being poloxamers and betaines [5, 6]. Poloxamers are non-ionic, synthetic surfactants  
207 with a central hydrophobic chain of polyoxypropylene as well as two hydrophilic polyoxyethylene  
208 chains. The chains length can be adjusted to produce different types of poloxamers [5, 6].  
209 Poloxamer 188 has been shown to inhibit biofilm generation and development by *S. aureus* or  
210 *Acinetobacter baumannii* persisting in the wound following treatment in an *ex vivo* porcine skin  
211 model [22]. Additionally, wound dressing solution of poloxamer containing 1% silver sulfadiazine  
212 (SSD) has shown effectiveness in wounds that are more likely to be infected or those with clinical  
213 infection symptoms [23, 24]. Another case series reported such uses to be associated with  
214 favorable healing rates and reduction in pain when compared to standard care procedures [24].  
215 Romic et al. [25] also showed that poloxamer 407 could reduce biofilm production through  
216 disrupting the attachment of *Staphylococcus epidermidis* to the wound surface. The effectiveness  
217 of surfactants on biofilm control has also been demonstrated in *in vitro* investigations by Yang et  
218 al. [21] who evaluated the effectiveness of a wound dressing containing surfactant in porcine skin  
219 explants. They reported that biofilm reduction to undetectable levels occurred one day following  
220 therapy and after cleaning the skin model using poloxamer 188-moistened gauze.

221 In another study, Percival et al. [26] reported the advantages of using concentrated poloxamer-  
222 containing surfactant in breaking down, dispersing and inhibition of *P. aeruginosa*, *Enterococcus*  
223 spp., *S. epidermidis*, and methicillin resistant *S. aureus* biofilms. *In vivo* investigations by Howell  
224 et al. [27] demonstrated that pretreatment with a poloxamer-based surfactant prior to using iodine-  
225 containing surgical scrub improved povidone iodine effectiveness. However, limited information  
226 is available about poloxamer- based surfactants in clinical settings. Topical use of these surfactants  
227 is greatly tolerable and has been well accepted by patients and do not negatively affect the general  
228 wound healing process [6]. Interestingly, they also improved healing in full- thickness rat excisions  
229 wound model [6]. Plurogel<sup>®</sup> (Medline Industries Inc) is an example of a wound gel containing the  
230 surfactant Poloxamer 188, which has shown its capability in reducing the inflammatory effects  
231 caused by biofilms through modulating the secretion of pro-inflammatory cytokines. It provides a  
232 moisture barrier and cleansing impact assisted by forming micelle gel matrixes [20]. The  
233 micellular component within the Poloxamer gel solutions exists in an extensively disordered state  
234 at room temperature, which forms a thin flowing gel. At increased temperatures, the micelle core  
235 is dehydrated, forming a more ordered crystalline gel state. As a result, when applying liquid  
236 poloxamer to the human body, subjecting it to slightly higher body temperatures quickly produces  
237 a more solid gel structure while at lower temperatures, disorganization occurs in the micelles and  
238 the gel flows like a liquid again [6].

239

240

241

242 ***Combining surfactants and antimicrobials***

243

244 To further enhance surfactants impacts on biofilm removal, they are often combined with  
245 antimicrobials. For example, the surfactant Poloxamer 188 has been combined with SSD  
246 antimicrobial and was shown to eliminate all viable bacteria from the skin within 3 days of  
247 application [21]. Additionally, the use of the synthetic surfactant undecylenamidopropyl betaine  
248 and the antimicrobial polyhexanide, showed to result in biofilm elimination and dramatic  
249 improvements in wound healing in 7 out of 10 patients within 3 weeks [28]. The combination of  
250 0.1% polyhexanide and 0.1% betaine was also used to investigate their ability to manage infected  
251 wounds. The results showed a 5.3-5.8 log reduction in the numbers and prevalence of *S.*  
252 *epidermidis*, *P. aeruginosa*, *Candida albicans*, *S. aureus*, *Enterococcus faecalis*, and *Escherichia*  
253 *coli* in addition to several other strains commonly found in wound biofilms [28].

#### 254 **Surfactants' mechanism of action**

255 Surfactants are able to act as wound healing agent by various mechanisms including as facilitators  
256 at the liquid/liquid interface (water and oil) or as solid/liquid interface due to their hydrophilic and  
257 hydrophobic mobile structures. For instance, they are able to breakdown the water/oil interface  
258 promoting emulsification and holding oil in suspension. Water-insoluble molecules gather near  
259 the hydrophobic groups and the spherical micelle components are formed in a concentration and  
260 temperature-dependent manner [12]. Accordingly, debris from the wound are continuously  
261 trapped, resulting in a rinsing action. Surfactants also reduce the interfacial tension between the  
262 wound bed and cleansing liquid; therefore, a close connection is formed between the cleansing  
263 liquid and the wound bed, which can facilitate separating loose and nonviable tissues and microbial  
264 pathogens from the viable wound bed preventing biofilm production and inhibiting the existing  
265 more persistent biofilms. [26].

266

## 267 **Wound treatment using different biosurfactant formulations**

268 Cosmetic, wound care and pharmaceutical industries always endeavor to enhance the preservation  
269 and solubilization of the active components in their formulations; therefore, they choose  
270 surfactants and preservatives, which mostly include chemical compounds that may occasionally  
271 increase irritant or allergic reactions. Consequently, alternative natural-based products have been  
272 recently sought after and considered as replacements in cosmetic and pharmaceutical industries  
273 [29, 30]. Accordingly, some of the biocompatible preservatives with reduced side effects compared  
274 to their chemical counter parts are often considered as alternatives [31]. Thus, some biosurfactants  
275 with preservative and emulsifying properties were more commonly used. Some of these  
276 biosurfactants used as solubilizers have an additional inherent antimicrobial and anti-adhesive  
277 properties. In addition, biosurfactants are readily biodegradable due to their lipids, proteins,  
278 peptides or carbohydrates structures which make them more appealing for many applications.  
279 Several biosurfactants have potential to be used for dermatological purposes, including wound  
280 healing. They also occasionally have some synergetic activity with some antibiotics, enhancing  
281 their solubilization and antimicrobial activities in addition to have some antimicrobial and  
282 antibiofilm activities themselves [32-34]. Although, the number of studies on antimicrobial,  
283 antiadhesion, and anti-inflammatory properties of different biosurfactants have been increasing,  
284 their uses in wound healing remain quite limited.

### 285 ***Lipopeptide biosurfactants as wound healing agent***

286 Zouari et al. [29] studied *in vitro* antioxidant properties and the wound healing effect of *Bacillus*  
287 *subtilis* SPB1 lipopeptide biosurfactant on excision wound areas in experimental rats and reported  
288 a remarkable enhancement in wound closure rate compared to control and CICAFLORA™-  
289 administrated animals. Biopsies treatment using lipopeptide biosurfactant showed completely re-

290 epithelized wounds associated with excellent epidermal reproduction. The free-radical scavenging  
291 effect of the lipopeptide biosurfactant was capable of preventing inflammation and improving  
292 tissue generation, re-epithelization and differentiation of the epidermis [35]. Also, lipopeptide  
293 biosurfactant can inhibit multidrug-resistance bacteria [36] and act against phytopathogenic fungi  
294 [37]. Wound healing effects of investigated lipopeptide biosurfactants might be ascribed their  
295 ability for reducing oxidative stress through preventing ROS generation. Ohadi et al. [9] indicated  
296 that the lipopeptide biosurfactants formed by *Acinetobacter junii* B6 improved scavenging free  
297 radicals properties and enhanced histopathological remission in rats.

#### 298 ***Glycolipids biosurfactants as wound healing agent***

299 Gupta et al. [1] evaluated improved wound healing in rat tissue *in vivo* using glycolipids generated  
300 by *Bacillus licheniformis* SV1 containing ointment and found re- epithelization and fibroblast cell  
301 proliferation in the primary phase of wound healing leading to higher rate of deposition of collagen  
302 in the next phases. Rhamnolipids were also used for the re-epithelizing of mucous membrane  
303 tissues, especially to treat and prevent gum disorder and improve periodontal regeneration [38].  
304 Lower levels of rhamnolipids have also been able to inhibit the phagocytic actions of macrophages  
305 allowing improved control the inflammatory phase. Stipceviv et al. [38] investigated the wound  
306 healing properties of di-rhamnolipid BAC-3 formulated as an ointment and used topically on full-  
307 thickness burn wounds in healthy rats covering 5% of the body surface. They also noted that the  
308 BAC-3 di- rhamnolipid was well-tolerated as daily subcutaneous injection, through 7 days in  
309 female mice at the maximum of 120 mg/ (kg day) [38]. In another study, Sana et al [30] assessed  
310 the wound healing activity of rhamnolipid generated by *P. aeruginosa* C2. They found that  
311 rhamnolipid could accelerate wound healing through their antimicrobial activities. This study also  
312 suggested healing support *via* increasing protein, DNA, hexosamine content, and reduced tumor

313 necrosis factor-alpha (TNF- $\alpha$ ) amount. Decreasing level of TNF- $\alpha$  in rhamnolipid group might  
314 lead to reduction in inflammation and collagen formation. Lydon et al. [33] investigated very pure  
315 micelle-producing nonacetylated acidic sophorolipids containing 90% C18 congener and reported  
316 that acidic sophorolipids was appropriate for use in antimicrobial creams for reducing wound  
317 infection risk through healing. In another study, Sophorolipids were also reported to act as  
318 stimulators of skin fibroblast metabolism contributing to skin restructuring, repair and protection  
319 [39]. Sophorolipids act as desquamating and depigmenting agents by eliminating the affected  
320 surface of the epidermis protective layer as a part of the wound healing process. A summary of  
321 wound healing activities for different biosurfactants are listed in Table 1.

322

323

324

325

326

327

328

329

330

331

332 Table1. Summary of wound healing activity of biosurfactants

Type of biosurfactant	Strain	Biological activity	Ref
Lipopeptide	<i>B. subtilis</i> SPB1 LP	Anti-inflammatory Antimicrobial	[29]
Lipopeptide	<i>A. junii</i> B6	Anti-inflammatory Antioxidant Antimicrobial	[9]
Lipopeptide	<i>Bacillus mojavensis</i> A21	Anti-inflammatory Antioxidant	[10]
Glycolipid	<i>B. licheniformis</i> SV1	Anti-inflammatory Antioxidant Antimicrobial	[1]
Glycolipid	<i>Starmerella bombicola</i>	Antimicrobial	[33]
Glycolipid	<i>Candida bombicola</i>	Depigmenting	[39]
Glycolipid	<i>P. aeruginosa</i> C2	Antimicrobial	[30]

333

334 **Biosurfactants adjuvant/synergetic activities facilitating wound healing**

335 Using biosurfactants for medical purposes has been increasingly considered in the past few years  
336 due to some of their unique properties [34]. Such properties are related to their amphiphilic nature  
337 increasing the surface humidity and the solubility of materials and an ability to decrease the  
338 liquids/surface tension, leading to a higher penetration of fluids, such as solvents and  
339 antimicrobials [33]. Reduction of the interfacial tension between immiscible liquids molecules  
340 makes them more ‘slippery’, resulting in a lower risk of adhesion to surfaces. Accordingly,

341 biosurfactants, allows some target molecules to be carried away upon the application of irrigating  
342 compounds such as water or saline through creating micellular structures facilitating the cleaning  
343 processes [5]. In addition to these properties, biosurfactants can act as an adjuvant or synergetic  
344 agent for enabling faster wound healing [33] in addition to their advantageous antimicrobial and  
345 antibiofilm activities [32, 34]. Investigating the effect of acidic sophorolipid as an adjuvant  
346 antibiotic effect to facilitate wound healing, Lydon et al. [32] concluded that acidic sophorolipids  
347 antibacterial, antibiofilm, and antibiotic adjuvant effects supports further consideration for  
348 applications to reduce chronic wounds infections. In another study, Tabbene et al [40] evaluated  
349 the lipopeptide bacillomycin D and amphotericin B effects separately and in combination to  
350 inhibit *C. albicans* biofilm production and accelerating keratinocyte cell migration. They showed  
351 the effectiveness of the lipopeptide bacillomycin D in combination with the common amphotericin  
352 B in inhibiting biofilm production and reducing cell viability as well as improving the closure of  
353 ‘pseudo-wounds’.

#### 354 **Biosurfactants skin penetration enhancement techniques**

355 Several studies indicated the effect of amino acids and peptides on reversing the cutaneous  
356 symptoms of aging and having a secondary advantage of improving the wound healing process  
357 [41]. Such peptides consist of a small sequence amino acid chains possibly stimulating  
358 angiogenesis, granulation tissue synthesis, and new collagen production. Nonetheless, proteins and  
359 peptides are hydrophilic molecules that are frequently charged at physiological pH conditions.  
360 Their molecular weights are varied from small peptides (300Da) to proteins (>1000kDa). They  
361 cause weak skin permeation and although they have high potency, are mostly therapeutically  
362 ineffective when administered transdermally due to poor permeation. To overcome the skin barrier  
363 and facilitate easier permeability of bio-drugs several skin penetration enhancement methods have



364 been evaluated. Formulation and chemical enhancement methods can be used for the delivery of  
365 small peptides used in dermatological and cosmetic applications [42]. The positive effects of  
366 coupling tetrapeptide attached to a short hydrocarbon chain can improve transepidermal delivery  
367 of protein inhibitors. Attached oligopeptides linked to a fatty acid for example can improve oil  
368 solubility of peptides resulting in a favorable skin penetration [43]. Cutaneous absorption of  
369 interferon alpha ( $\text{INF}\alpha$ ) attached to different palmitoyl molecules has been reported to occur at  
370 nearly five to six times that of the parent peptides [44]. Lipopeptides biosurfactants containing 8-  
371 24 carbon atoms made by a hydrophilic or hydrophilic peptide chain have been reported to  
372 stimulate the production of crucial constituents of the skin matrix (collagen and elastin) when  
373 added to the skin fibroblast culture [45].

#### 374 **Mechanism of wound healing by microbial surfactant**

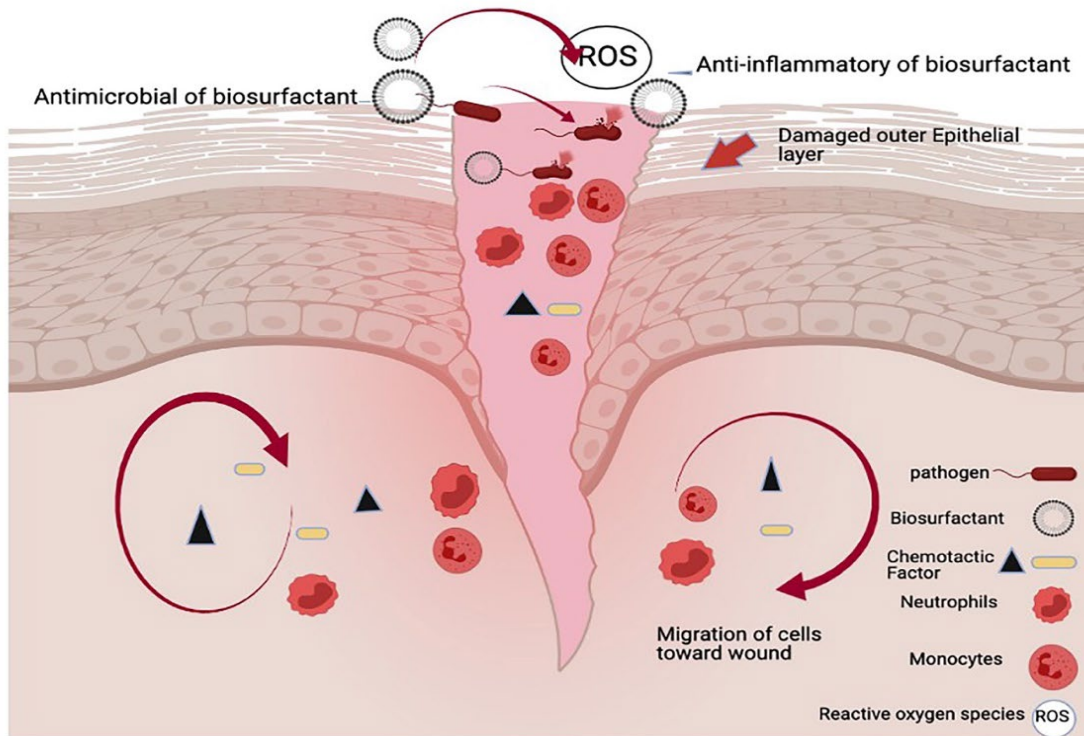
375 Wound repair is a dynamic series of events, in which adhered pathogens are killed, chemotactic  
376 factors stimulated, inflammatory cells are migrated and injured dermal layer is remodeled.  
377 Cellular cross-talking reactions are also initiated inside and outside the wound region that begins  
378 the repair of injured epithelial layers. The repair speed however, is affected by pathogenic  
379 infections and poor immune reactions. Therefore, the “elimination of pathogens” is of great  
380 importance for effective wound treatment [29]. Biosurfactants can enhance bacterial cell  
381 membrane permeability as well as cellular metabolites leakage preventing microbial infections.  
382 This antimicrobial effect enhances early wound healing [1, 46]. The mechanism of wound  
383 healing by biosurfactants is schematically illustrated in Fig 1. Using biosurfactants as constituent  
384 can disinfect the wound areas from potential pathogens and stimulate numerous chemotactic  
385 factors which generates chemotactic signals using different inflammatory cells, such as  
386 monocytes, endothelial cells, neutrophils, fibroblast cells at the wound area [38, 47]. These cells

387 migrate to the wound area via circulating blood capillaries, collecting pathogens and dead  
388 epithelial cells through phagocytosis and acting as protective materials of cellular repair systems  
389 enhancing wound healing activities at the injured area. Fibroblast cells can be activated to begin  
390 synthesizing collagen types I and II proteins that cross-link forming a thick layer of connective  
391 tissue simultaneously (Fig. 1) [48]. The connective tissue rich in collagen is crucial for re-  
392 epithelization and neovascularization process at the wound site, which is helpful in the early  
393 wound contraction. In the Underlying connective tissues in the wound site, the epidermal part  
394 gets smaller leading to bringing the wound edges nearer to each other and assisting remodeling  
395 process of the tissue [49, 50]. Hence, biosurfactants can be regarded as a transdermal substitute  
396 causing intensive connective tissue remodeling and a greater frequency of re-epithelization with  
397 improved wound healing effects. In a recent study, Mehrabani et al. [51] investigated the wound  
398 healing mechanism of lipopeptide biosurfactants. They showed that lipopeptide biosurfactants  
399 increase the rate of wound site angiogenesis through making improvements in the levels of HIF-  
400  $1\alpha$  and VEGF protein expression. Lack of HIF- $1\alpha$  and VEGF protein expression is partly  
401 responsible for poor wound healing. However, the functional role of biosurfactants in increasing  
402 the expression of HIF- $1\alpha$  and VEGF protein is not well known. In normoxia, prolyl hydroxylase  
403 domain proteins (PHDs) trigger the hydroxylation of two proline residues in the oxygen  
404 degradation domain of HIF- $\alpha$  trigger leading to its degradation. In addition to  $O_2$ , PHDs also  
405 require other cofactors to be activated including  $Fe^{+2}$ . Therefore, through the inhibition of  
406 PHDs, iron chelating agents could be used to stabilize HIF- $\alpha$  and protect it from further  
407 degradation. The elevating effect on HIF- $1\alpha$  and its downstream genes may well be associated  
408 with the iron chelating capacity of biosurfactants. Hemlata et al. [52] demonstrated the iron  
409 chelating ability of a glycolipid biosurfactant produced by *Stenotrophomonas maltophilia* NBS-

410 11. They suggested that cells may be protected against oxidative stress through making use of the  
411 iron chelating capacity of the produced biosurfactant. Also, previous studies have demonstrated  
412 that iron chelators may positively affect the angiogenesis process through increasing the  
413 expression of VEGF and HIF 1- $\alpha$  proteins [53]. ROS and RNS generation on the wounded area  
414 can delay the healing process by initiating oxidative stresses, LPO, DNA damage, and  
415 inactivating free radical scavenger enzymes. The anti-inflammatory activity of a range of  
416 biosurfactants has been investigated in various studies [9, 10, 29]. Biosurfactants' scavenging  
417 activity might be attributed to their hydrophilic and lipophilic effects that may improve the  
418 radical scavenging abilities [54]. Biosurfactants have reported as antibiofilm agents although the  
419 anti-biofilm effect mechanism is still unclear [32]. However, most biosurfactants are able to  
420 enhance bacterial surface hydrophobicity as well as de- stabilize lipid packing because of their  
421 amphipathic nature. Finally, such alterations improve the membrane cells permeability and  
422 consequently, reduce microbial adhesion toward solid surfaces [46]. In addition, the mechanism  
423 of anti-biofilm effect via microbial surfactant collected from lactic acid bacteria to *S. aureus*  
424 CMCC 26003 has been reported by Yan et al. [55]. Biofilm-associated genes expressions, such  
425 as *cidA*, *icaA*, *dltB*, *agrA*, *sortaseA* and *sarA* are affected by biosurfactants and interfere with  
426 signaling molecules released by the quorum sensing systems which may prevent biofilm  
427 development.

428 Diabetic wound healing is usually delayed by many factor, including high TNF- $\alpha$  expression and  
429 low TGF- $\beta$  expression which can affect the formation of new epithelial and collagen as the main  
430 goal of wound healing process. The increasing TNF- $\alpha$  expression is associated with inhabitation  
431 of cell migration, failure fibroblast proliferation, and inhabitation of angiogenesis resulting in the  
432 failure of diabetic wound healing. Raihanah et al. [56] reported that biosurfactants containing

433 dialkyl alginate cream decreased TNF- $\alpha$  expression and increase TGF- $\beta$  expression, and  
434 reepithelization.



435  
436 **Fig1.** Biosurfactants mediating *in vivo* wound repair. Figure created using BioRender  
437 (<https://biorender.com/>).

### 438 Challenges and recommendations for future research

439  
440 A comprehensive literature search was carried out using the ISI Web of Science and PubMed  
441 search engines. Both databases were selected in order to cover all of the published peer-reviewed  
442 literature. Several studies have demonstrated the efficacy of synthetic and natural surfactants in  
443 the wound healing. Nonetheless, considering the benefits of biodegradability and the generation  
444 of renewable-resource substrates, biosurfactants may possibly replace their chemically produced  
445 compounds. Biosurfactants have not been widely used due to their high production and purification  
446 costs [57]. Biosurfactants-producing strain' physiology, genetics, downstream processing and

447 purification should be investigated to reduce their production costs [58]. It is also difficult to  
448 compare between studies because of variations in wound types and size, animal models,  
449 biosurfactants types, and biosurfactants mixtures (congeners variations). To increase our  
450 knowledge about the possible advantages or disadvantages of these compounds in wound healing,  
451 data regarding the constitution of biosurfactants mixtures and standardizing considered  
452 experimental wound healing model are vital [34, 59]. Additionally, the toxicity of a new  
453 biosurfactant needs to be addressed for certifying their immunity to using in cosmetics and wound  
454 care industry [9, 60]. Limited of clinical data on the use of biosurfactants in human volunteers is  
455 major challenge in preparing safe formulations in wound healing. To construct a sustainable  
456 society, several programs are now underway worldwide, of which the introduction of green  
457 technology is challenging. Regarding the today's social and technological circumstances, using  
458 biosurfactants, as environmentally friendly and highly functional components, remains an  
459 attractive proposition.

460

461

462

463

464

465

466

467

468

469

470

471

472

473

474

475 **References:**

476

477 [1] S. Gupta, N. Raghuwanshi, R. Varshney, I.M. Banat, A.K. Srivastava, P.A. Pruthi, V.J.B.

478 Pruthi, *Pharmacotherapy, Accelerated in vivo wound healing evaluation of microbial glycolipid*

479 *containing ointment as a transdermal substitute*, 94 (2017) 1186-1196.

480 [2] A. Bianchera, O. Catanzano, J. Boateng, L.J.T.D. Elviri, W.H. Applications, *The Place of*

481 *Biomaterials in Wound Healing*, (2020) 337-366.

482 [3] S. Dhivya, V.V. Padma, E.J.B. Santhini, *Wound dressings—a review*, 5(4) (2015).

483 [4] E. Yadav, D. Singh, P. Yadav, A.J.B. Verma, *Pharmacotherapy, Antioxidant and anti-*

484 *inflammatory properties of Prosopis cineraria based phenolic rich ointment in wound healing*,

485 108 (2018) 1572-1583.

486 [5] H. Tyldesley, A. Salisbury, R. Chen, M. Mullin, S.J.W.I. Percival, *Surfactants and their role*

487 *in biofilm management in chronic wounds*, 10(1) (2019).

488 [6] S.L. Percival, D. Mayer, M. Malone, T. Swanson, D. Gibson, G.J.J.o.w.c. Schultz,

489 *Surfactants and their role in wound cleansing and biofilm management*, 26(11) (2017) 680-690.

490 [7] G. Sganga, T. Spanu, G. Bianco, B. Fiori, E. Nure, G. Pepe, T. D'inzeo, M. Lirosi, F.

491 Frongillo, S. Agnes, *Bacterial bloodstream infections in liver transplantation: etiologic agents*

492 *and antimicrobial susceptibility profiles*, *Transplantation proceedings*, Elsevier, 2012, pp. 1973-

493 1976.

494 [8] C. Shen, L. Jiang, H. Shao, C. You, G. Zhang, S. Ding, T. Bian, C. Han, Q.J.S.r. Meng,  
495 Targeted killing of myofibroblasts by biosurfactant di-rhamnolipid suggests a therapy against  
496 scar formation, 6(1) (2016) 1-10.

497 [9] M. Ohadi, H. Forootanfar, H.R. Rahimi, E. Jafari, M. Shakibaie, T. Eslaminejad, G.  
498 Dehghannoudeh, Antioxidant Potential and Wound Healing Activity of Biosurfactant Produced  
499 by *Acinetobacter junii* B6, *Curr. Pharm. Biotechnol* (2017).

500 [10] H.B. Ayed, S. Bardaa, D. Moalla, M. Jridi, H. Maalej, Z. Sahnoun, T. Rebai, P. Jacques, M.  
501 Nasri, N.J.P.B. Hmidet, Wound healing and in vitro antioxidant activities of lipopeptides mixture  
502 produced by *Bacillus mojavensis* A21, 50(6) (2015) 1023-1030.

503 [11] C.E. Drakontis, S.J.C.O.i.C. Amin, I. Science, *Biosurfactants: Formulations, Properties, and*  
504 *Applications*, (2020).

505 [12] L.R.J.J.o.c. Rodrigues, i. science, *Microbial surfactants: fundamentals and applicability in*  
506 *the formulation of nano-sized drug delivery vectors*, 449 (2015) 304-316.

507 [13] M. Ohadi, A. Shahravan, N. Dehghannoudeh, T. Eslaminejad, I.M. Banat, G.J.D.D.  
508 Dehghannoudeh, *Development, Therapy, Potential Use of Microbial Surfactant in*  
509 *Microemulsion Drug Delivery System: A Systematic Review*, 14 (2020) 541.

510 [14] I.M. Banat, A. Franzetti, I. Gandolfi, G. Bestetti, M.G. Martinotti, L. Fracchia, T.J. Smyth,  
511 R.J.A.m. Marchant, *biotechnology, Microbial biosurfactants production, applications and future*  
512 *potential*, 87(2) (2010) 427-444.

513 [15] A. Bellingeri, F. Falciani, P. Trapedini, A. Moscatelli, A. Russo, G. Tino, P. Chiari,  
514 A.J.J.o.W.C. Peghetti, *Effect of a wound cleansing solution on wound bed preparation and*  
515 *inflammation in chronic wounds: a single-blind RCT*, 25(3) (2016) 160-168.

516 [16] A.E. Andriessen, T.J.W.a.c.o.c.r. Eberlein, practice, Assessment of a wound cleansing  
517 solution in the treatment of problem wounds, 20(6) (2008) 171-175.

518 [17] C.L. Burnett, W.F. Bergfeld, D.V. Belsito, R.A. Hill, C.D. Klaassen, D. Liebler, J.G. Marks  
519 Jr, R.C. Shank, T.J. Slaga, P.W.J.I.j.o.t. Snyder, Final report of the Cosmetic Ingredient Review  
520 Expert Panel on the safety assessment of cocamidopropyl betaine (CAPB), 31(4\_suppl) (2012)  
521 77S-111S.

522 [18] P. Kennedy, S. Brammah, E.J.B. Wills, Burns, biofilm and a new appraisal of burn wound  
523 sepsis, 36(1) (2010) 49-56.

524 [19] G. Nakagami, H. Sanada, J. Sugama, T. Morohoshi, T. Ikeda, Y.J.W.r. Ohta, regeneration,  
525 Detection of *Pseudomonas aeruginosa* quorum sensing signals in an infected ischemic wound: an  
526 experimental study in rats, 16(1) (2008) 30-36.

527 [20] A.-M. Salisbury, D. Mayer, R. Chen, S.L.J.A.i.w.c. Percival, Efficacy of Concentrated  
528 Surfactant-Based Wound Dressings in Wound Repair and Biofilm Reduction, 7(9) (2018) 315-  
529 322.

530 [21] Q. Yang, C. Larose, A.C. Della Porta, G.S. Schultz, D.J.J.I.w.j. Gibson, A surfactant-based  
531 wound dressing can reduce bacterial biofilms in a porcine skin explant model, 14(2) (2017) 408-  
532 413.

533 [22] Q. Yang, G.S. Schultz, D.J.J.J.o.B.C. Gibson, Research, A surfactant-based dressing to treat  
534 and prevent *Acinetobacter baumannii* biofilms, 39(5) (2018) 766-770.

535 [23] J.S. Black, D.B.J.P.S.N. Drake, A prospective randomized trial comparing silver  
536 sulfadiazine cream with a water-soluble polyantimicrobial gel in partial-thickness burn wounds,  
537 35(1) (2015) 46-49.



538 [24] C. Zölß, J.D.J.I.w.j. Cech, Efficacy of a new multifunctional surfactant-based biomaterial  
539 dressing with 1% silver sulphadiazine in chronic wounds, 13(5) (2016) 738-743.

540 [25] M.D. Romić, M.Š. Klarić, J. Lovrić, I. Pepić, B. Cetina-Čižmek, J. Filipović-Grčić,  
541 A.J.E.J.o.P. Hafner, Biopharmaceutics, Melatonin-loaded chitosan/Pluronic® F127 microspheres  
542 as in situ forming hydrogel: An innovative antimicrobial wound dressing, 107 (2016) 67-79.

543 [26] S.L. Percival, D. Mayer, A.M.J.W.R. Salisbury, Regeneration, Efficacy of a surfactant-  
544 based wound dressing on biofilm control, 25(5) (2017) 767-773.

545 [27] J.M. Howell, T.O. Stair, A.W. Howell, D.J. Mundt, A. Falcone, S.R.J.T.A.j.o.e.m. Peters,  
546 The effect of scrubbing and irrigation with normal saline, povidone iodine, and cefazolin on  
547 wound bacterial counts in a guinea pig model, 11(2) (1993) 134-138.

548 [28] A.J.B.J.o.n. Horrocks, Prontosan wound irrigation and gel: management of chronic wounds,  
549 15(22) (2006) 1222-1228.

550 [29] R. Zouari, D. Moalla-Rekik, Z. Sahnoun, T. Rebai, S. Ellouze-Chaabouni, D.J.B. Ghribi-  
551 Aydi, Pharmacotherapy, Evaluation of dermal wound healing and in vitro antioxidant efficiency  
552 of Bacillus subtilis SPB1 biosurfactant, 84 (2016) 878-891.

553 [30] S. Sana, S. Datta, D. Biswas, B. Auddy, M. Gupta, H.J.W.m. Chattopadhyay, Excision  
554 wound healing activity of a common biosurfactant produced by Pseudomonas sp, 23 (2018) 47-  
555 52.

556 [31] K. Liu, Y. Sun, M. Cao, J. Wang, J.R. Lu, H.J.C.O.i.C. Xu, I. Science, Rational design,  
557 properties and applications of biosurfactants: a short review of recent advances, (2019).

558 [32] I.M. Banat, M.A.D. De Rienzo, G.A.J.A.m. Quinn, biotechnology, Microbial biofilms:  
559 biosurfactants as antibiofilm agents, 98(24) (2014) 9915-9929.

560 [33] H.L. Lydon, N. Baccile, B. Callaghan, R. Marchant, C.A. Mitchell, I.M.J.A.a. Banat,  
561 chemotherapy, Adjuvant antibiotic activity of acidic sophorolipids with potential for facilitating  
562 wound healing, 61(5) (2017) e02547-16.

563 [34] L. Fracchia, C. Ceresa, I.M. Banat, Biosurfactants in cosmetic, biomedical and  
564 pharmaceutical industry, Microbial Biosurfactants and their Environmental and Industrial  
565 Applications, CRC Press2019, pp. 258-287.

566 [35] N. Jemil, H.B. Ayed, A. Manresa, M. Nasri, N.J.B.m. Hmidet, Antioxidant properties,  
567 antimicrobial and anti-adhesive activities of DCS1 lipopeptides from *Bacillus methylotrophicus*  
568 DCS1, 17(1) (2017) 144.

569 [36] D. Ghribi, L. Abdelkefi-Mesrati, I. Mnif, R. Kammoun, I. Ayadi, I. Saadaoui, S. Maktouf,  
570 S.J.B.R.I. Chaabouni-Ellouze, Investigation of antimicrobial activity and statistical optimization  
571 of *Bacillus subtilis* SPB1 biosurfactant production in solid-state fermentation, 2012 (2012).

572 [37] I. Mnif, A. Grau-Campistany, J. Coronel-León, I. Hammami, M.A. Triki, A. Manresa,  
573 D.J.E.S. Ghribi, P. Research, Purification and identification of *Bacillus subtilis* SPB1 lipopeptide  
574 biosurfactant exhibiting antifungal activity against *Rhizoctonia bataticola* and *Rhizoctonia*  
575 *solani*, 23(7) (2016) 6690-6699.

576 [38] T. Stipcevic, A. Piljac, G.J.B. Piljac, Enhanced healing of full-thickness burn wounds using  
577 di-rhamnolipid, 32(1) (2006) 24-34.

578 [39] F.B. Concaix, Use of sophorolipids comprising diacetyl lactones as agent for stimulating  
579 skin fibroblast metabolism, Google Patents, 2003.

580 [40] O. Tabbene, S. Azaiez, A. Di Grazia, I. Karkouch, I. Ben Slimene, S. Elkahoui, M. Alfeddy,  
581 B. Casciaro, V. Luca, F.J.J.o.a.m. Limam, Bacillomycin D and its combination with

582 amphotericin B: promising antifungal compounds with powerful antibiofilm activity and wound-  
583 healing potency, 120(2) (2016) 289-300.

584 [41] M.P. Lupo, A.L.J.D.t. Cole, *Cosmeceutical peptides*, 20(5) (2007) 343-349.

585 [42] H.A. Benson, Elastic liposomes for topical and transdermal drug delivery, *Current drug*  
586 *delivery* 6(3) (2009) 217-226.

587 [43] M. Foldvari, M.E. Baca-Estrada, Z. He, J. Hu, S. Attah-Poku, M.J.B. King, a. biochemistry,  
588 *Dermal and transdermal delivery of protein pharmaceuticals: lipid-based delivery systems for*  
589 *interferon  $\alpha$* , 30(2) (1999) 129-137.

590 [44] M. Foldvari, M.E. Baca-Estrada, Z. He, J. Hu, S. Attah-Poku, M. King, *Dermal and*  
591 *transdermal delivery of protein pharmaceuticals: lipid-based delivery systems for interferon  $\alpha$* ,  
592 *Biotechnology and applied biochemistry* 30(2) (1999) 129-137.

593 [45] M. Kanlayavattanakul, N. Lourith, Lipopeptides in cosmetics, *International journal of*  
594 *cosmetic science* 32(1) (2010) 1-8.

595 [46] M. Ohadi, H. Forootanfar, G. Dehghannoudeh, T. Eslaminejad, A. Ameri, M. Shakibaie,  
596 M.J.M.p. Adeli-Sardou, Antimicrobial, anti-biofilm, and anti-proliferative activities of  
597 lipopeptide biosurfactant produced by *Acinetobacter junii* B6, 138 (2020) 103806.

598 [47] L. Yan, G. Liu, B. Zhao, B. Pang, W. Wu, C. Ai, X. Zhao, X. Wang, C. Jiang, D. Shao,  
599 *Novel Biomedical Functions of Surfactin A from Bacillus subtilis in Wound Healing Promotion*  
600 *and Scar Inhibition, Journal of Agricultural and Food Chemistry* (2020).

601 [48] S.A. Eming, P. Martin, M.J.S.t.m. Tomic-Canic, Wound repair and regeneration:  
602 mechanisms, signaling, and translation, 6(265) (2014) 265sr6-265sr6.

603 [49] J. Kelsoe, T. Greenwood, H. Akiskal, K.J.I.C.P. Akiskal, The genetic basis of affective  
604 temperament and the bipolar spectrum, 28 (2012) e5-e6.

605 [50] G.J.B.j.o.c.n. Gethin, Understanding the inflammatory process in wound healing, 17(Sup3)  
606 (2012) S17-S22.

607 [51] M. Mehrabani, M. Esmaceli-Tarzi, H. Forootanfar, M.H. Nematollahi, I.M. Banat, M.  
608 Ohadi, G. Dehghannoudeh, Lipopeptide Biosurfactant from *Acinetobacter junii* B6: A Promising  
609 Natural Surfactant for Promoting Angiogenesis, International Journal of Peptide Research and  
610 Therapeutics (2021) 1-7.

611 [52] B. Hemlata, J. Selvin, K. Tukaram, Optimization of iron chelating biosurfactant production  
612 by *Stenotrophomonas maltophilia* NBS-11, Biocatalysis and agricultural biotechnology 4(2)  
613 (2015) 135-143.

614 [53] J.A. Wright, T. Richards, S.K. Srari, The role of iron in the skin and cutaneous wound  
615 healing, Frontiers in Pharmacology 5 (2014) 156.

616 [54] C. Chen, T. Lin, Y.J.J.o.b. Shieh, bioengineering, Emulsification and antioxidation of  
617 biosurfactant extracts from Chinese medicinal herbs fermentation in vitro, 120(4) (2015) 387-  
618 395.

619 [55] X. Yan, S. Gu, X. Cui, Y. Shi, S. Wen, H. Chen, J.J.M.p. Ge, Antimicrobial, anti-adhesive  
620 and anti-biofilm potential of biosurfactants isolated from *Pediococcus acidilactici* and  
621 *Lactobacillus plantarum* against *Staphylococcus aureus* CMCC26003, 127 (2019) 12-20.

622 [56] C. Raihanah, N. Mahyani, K.J.J.I.K.I. Kintoko, Diabetic Wound Healing Biosurfactants  
623 Dialkyl Alginate Cream on TNF- $\alpha$  TGF- $\beta$  Expression, Reepithelization, and Collagenization,  
624 17(1) (2019) 72-80.

625 [57] M. Ohadi, G. Dehghannoudeh, M. Shakibaie, I.M. Banat, M. Pournamdari, H. Forootanfar,  
626 Isolation, characterization, and optimization of biosurfactant production by an oil-degrading

627 *Acinetobacter junii* B6 isolated from an Iranian oil excavation site, *Biocatalysis and Agricultural*  
628 *Biotechnology* 12 (2017) 1-9.

629 [58] E.O. Fenibo, S.I. Douglas, H.O. Stanley, A review on microbial surfactants: production,  
630 classifications, properties and characterization, *Journal of Advances in Microbiology* (2019) 1-  
631 22.

632 [59] R. Jahan, A.M. Bodratti, M. Tsianou, P. Alexandridis, Biosurfactants, natural alternatives to  
633 synthetic surfactants: Physicochemical properties and applications, *Advances in colloid and*  
634 *interface science* (2019) 102061.

635 [60] E.J. Gudiña, V. Rangarajan, R. Sen, L.R. Rodrigues, Potential therapeutic applications of  
636 biosurfactants, *Trends in pharmacological sciences* 34(12) (2013) 667-675.

637