

1 **GUT METABOLOMIC PROFILES IN PEDIATRIC**
2 **ULCERATIVE COLITIS PATIENTS PRIOR TO AND**
3 **AFTER RECEIVING FECAL MICROBIOTA**
4 **TRANSPLANTS**

5
6 Parastou S. Khalessi Hosseini¹, Beibei Wang², Yihui Luan², Fengzhu Sun^{3,*}, Sonia
7 Michail^{3,4}

8
9 ¹Los Angeles County – University of Southern California

10 ²Shandong University

11 ³University of Southern California

12 ⁴Children’s Hospital of Los Angeles

13
14 *Corresponding author Sonia Michail

15 4650 Sunset Blvd., MS #78

16 Los Angeles, CA 90027 Phone: 323-361-2181

17 Email:

18 Sonia.michail@hotmail.c

19 om Fax: 323-361-3718

20
21 Author contributions:

22 Conceptualization, P.S.K.H. and S.M.; Methodology, P.S.K.H. and S.M.; Formal
23 Analysis, B.W. and F.S.; Data Curation, P.S.K.H. and B.W.; Writing-Original Draft,

24 P.S.K.H. and S.M.;



CAMBRIDGE
UNIVERSITY PRESS

This peer-reviewed article has been accepted for publication in Gut Microbiome but has not yet been copy-edited or typeset so may be subject to change during the production process. The article is considered published and may be cited using its DOI: 10.1017/gmb.2023.15

Gut Microbiome is co-published by Cambridge University Press and The Nutrition Society. This is an Open Access article, distributed under the terms of the Creative Commons Attribution-NonCommercial-NoDerivatives licence (<http://creativecommons.org/licenses/by-nc-nd/4.0/>), which permits non-commercial re-use, distribution, and reproduction in any medium, provided the original work is unaltered and is properly cited. The written permission of Cambridge University Press must be obtained for commercial re-use or in order to create a derivative work.

25 Writing-Reviewing and Editing, P.S.K.H., S.M., B.W., Y.L., and F.S.,
26 Supervision, S.M. and F.S., Funding Acquisition, S.M.

27 **Background:** Ulcerative colitis (UC) is immune-mediated inflammation of the colonic
28 mucosa. Gut microbiota dysbiosis may play a significant role in disease pathogenesis
29 by causing shifts in metabolomic profiles within the gut.

30 **Aims:** To identify differences and trends in the metabolomic profile of pediatric UC
31 patients pre- and post-FMT.

32 **Methods:** Forty-six pediatric patients with mild to moderate UC and 30 healthy
33 pediatric patients were enrolled in this study. Baseline stool samples were collected
34 prior to FMT initiation and at months 1,3, 6, and 12 post-FMT. Pediatric Ulcerative
35 Colitis Activity Index (PUCAI) scores were calculated at baseline and months 1, 3, 6,
36 and 12 after FMT.

37 **Results:** The average Bray Curtis dissimilarities to healthy subjects decreased after
38 FMT. In PCoA plots, UC patients' centroids drew nearer to healthy individuals. The
39 variance explained by phenotype (Healthy versus UC) reduced and remained
40 significant. From 1-3 months after FMT, PUCAI trends were statistically significant
41 and decreasing. PUCAI scores remain flat starting 6 months after FMT.

42 **Conclusion:** This study concludes that pediatric UC patients have a significantly
43 different baseline metabolite profile than healthy controls. Although time-limited,
44 FMT significantly altered these metabolite profiles and shifted them towards that of
45 healthy controls.

46 Introduction:

47 Ulcerative Colitis (UC) is a type of inflammatory bowel disease (IBD)
48 characterized by chronic immune-mediated intestinal inflammation of the colonic
49 mucosal layer. Approximately 20-30% of patients become symptomatic and are
50 diagnosed with UC before 18 years of age (Abraham, 2012). This early presentation can
51 increase the risk of long-term physical and psychological sequelae in affected pediatric
52 patients.

53 There is significant interest in the role of the colonic microbiota in both UC
54 pathogenesis and management. It is hypothesized that dysbiosis within the gut can shift
55 metabolite profiles and cause an imbalance between anti- and pro-inflammatory
56 mediators. An interplay between these metabolites, genetics, and the environment could
57 be an inciting factor and impact the disease course. Research continues to be limited on
58 this topic and is primarily centered around adult UC patients. Given their unique and
59 more severe disease course, pediatric-specific research is needed, as this could reflect
60 differences in metabolite profiles (Tamboli et al., 2004; Lepage et al., 2008; Gever et al.,
61 2014; Scoville et al., 2018).

62 Metabolite profiles can reflect the gut microbiome, as many metabolites are
63 byproducts of microbiome metabolism. For instance, UC patients with reduced
64 *Ruminococcaceae* within the gut commonly have reduced levels of lithocholic acid and
65 deoxycholic acid (Sinha et al., 2020). A metabolite's protective versus detrimental role is
66 determined by complex host-microbe interaction and different regulatory pathways (Diab
67 et al., 2019; Shores et al., 2011; Staley et al., 2017).

68 Fecal microbiota transplants (FMT) could be a potential therapeutic option in
69 pediatric UC patients by reducing dysbiosis and shifting the colonic ecosystem towards
70 healthy donor levels. However, currently, there is a gap in knowledge of the baseline
71 metabolite profiles of pediatric UC patients, as well as how these profiles change
72 following FMT. This study aims to help define the pediatric UC patient's metabolomic
73 profile and describe individual metabolite trends post-FMT.

74

75 METHOD

76 **Materials:**

77 **Donors**

78 The universal donor subject is an identified healthy volunteer ≥ 16 to ≤ 21 years old, has
79 a BMI > 18.5 and < 25 , has not been diagnosed with any chronic illness, is on a regular
80 diet, and has not been taking any prescription, over-the-counter therapies, or probiotics
81 for at least 3 months.

82 The universal donor subject has a negative serum HIV, Hepatitis A, B (Davidovics et
83 al.,2019; Relman et al., 2013) and C, syphilis, and negative stool studies for culture,
84 multi-drug resistant organisms, Ova and parasites, *C. difficile*, Giardia, and
85 *Cryptosporidium* in accordance with recent guidelines endorsed by the American
86 Gastroenterological Association (Bakken et al.,2011; Owens et al., 2013).

87 **Sample population**

88 *Inclusion criteria*

89 Pediatric patients who have been diagnosed with mild to moderate ulcerative colitis. Mild
90 to moderate disease was based on a Pediatric Ulcerative Colitis Activity Index (PUCAI)
91 score of 10- 64.

92 *Exclusion criteria*

93 Children with known resistance to steroid therapy, immunomodulators, and biologics or
94 on a steroid dose greater than 0.5 mg/kg/day. Additionally, any child with recent dose
95 change in medications, allergy or intolerance to mesalamine or 5-ASA products,
96 evidence of infectious colitis, a concurrent infection that required anti-microbial therapy,
97 recently received probiotic preparations, recent or current pregnancy, currently
98 breastfeeding, renal or liver dysfunction, congenital or acquired immunodeficiency due
99 to conditions other than ulcerative colitis, recently received chemotherapy, recent
100 diagnosis with HIV, or inability to give informed consent/assent.

101

102 **FMT preparation and administration**

103 Heterologous (feces from healthy donor transplanted into person with ulcerative colitis)
104 and autologous (collection of feces during patient's healthy state for later use) were
105 collected. All heterologous and autologous stool samples were collected on-site,
106 transported on ice, and processed within two hours. The filtered healthy human donor
107 subject stool solution was homogenized in sterile normal saline. Fifty grams of processed
108 fecal material was infused into the terminal ileum through the working channel of the
109 instrument during colonoscopy to allow delivery of the transplanted microbiome to the
110 entire colon.

111

112 Sample Collection

113 Baseline stool samples from 44 different patients were collected prior to administration
114 of FMT. Follow-up stool samples were collected at months 1, 3, 6, and 12 after receiving
115 one infusion of FMT. Stools were immediately collected, transported on ice, and placed
116 into a -80C freezer until profiling.

117

118 PUCAI scoring

119 PUCAI scores were calculated at baseline and at months 1, 3, 6, and 12 post-FMT.

120

121 COVID-19 Pandemic

122 All samples were collected prior to the COVID-19 pandemic.

123

124 Metabolomic profiling

125 Fecal samples were analyzed at the UC Davis West Coast Metabolomics Center using
126 untargeted metabolomics by gas chromatography-time of flight-mass spectrometry (GC-
127 TOF-MS). Metabolites were identified by comparison to the BinBase database
128 (Nusbaum et al., 2018). Signal intensities were obtained for 230 metabolites which were
129 included in subsequent analyses.

130

131 Statistical analysis

132 Data analysis was performed in R version 4.1.2 (Team, 2021). Bray-Curtis
133 dissimilarities (Bray and Curtis, 1957) for the metabolomics data were calculated using
134 ‘vegdist’ function in the ‘vegan’ package (version 2.5.7) (Oksanen et al., 2020) after
135 normalizing the metabolomics profiles to relative abundances. A principal coordinates
136 analysis (PCoA) (Dray et al., 2006) based on Bray-Curtis dissimilarities was
137 implemented using the ‘pcoa’ function in the ‘ape’ package (version 5.6.2) (Paradis and
138 Schliep, 2019) to project samples into two-dimensional Euclidean space.
139 Quantifications of variance explained by different variables were calculated using
140 permutational multivariate analysis of variance (PERMANOVA) (Anderson, 2001)

141 with the 'adonis' function in the 'vegan' package (version 2.5.7) (Oksanen et al., 2020)
142 based on Bray-Curtis dissimilarities. To avoid problems related to variable ordering, the
143 total variance explained by each variable was evaluated independently of other variables
144 and thus should be regarded as the total variance explainable by that variable (Lloyd-
145 Price et al., 2019). The corresponding significances were assessed using permutational
146 tests with 1,000 permutations.

147 Differential abundance analysis of metabolites was conducted by MaAsLin 2
148 (Microbiome Multivariable Associations with Linear Models) (Mallick et al., 2021). It
149 provided a coherent paradigm through a multi-model framework with arbitrary
150 coefficients and contrasts of interest and had been shown to produce more consistent
151 results across different datasets (Nearing et al., 2022). Metabolites with very low
152 variance across all samples (below half the median of all feature-wise variances) were
153 removed from the following analysis. We entered log-transformed metabolomics
154 profiles into the primary ‘Maaslin2’ function within the ‘MaAsLin2’ R package (version
155 1.8.0) (Mallick et al., 2021). And then fit a linear model for each feature. BH FDR
156 corrected p values by Wald test were produced.

- 157 1. For identifying the differential expressed metabolites between Healthy controls and
158 UC cases at different time points after FMT, log-transformed abundances were fit
159 with the following per-feature linear fixed-effects model: $\text{Feature} \sim (\text{intercept}) +$
160 $\text{phenotype} + \text{gender} + \text{age} + \text{ethnicity}$, (1) where phenotype (Healthy / UC with
161 Healthy as the reference group), gender (female / male), and ethnicity (Hispanic /
162 non-Hispanic) were category variables, and age was a continuous variable.
163 Significant associations were defined as those with BH-FDR q value of the
164 corresponding coefficient below the threshold of 0.05.
- 165 2. For recognizing the differential expressed metabolites between any two of the time
166 points after FMT of UC cases, log-transformed abundances were fit with the
167 following per-feature linear mixed-effects model: $\text{Feature} \sim (\text{intercept}) + \text{time point} +$
168 $\text{age} + \text{gender} + \text{ethnicity} + \text{pancolitis}$
169 $+ \text{CDI history} + \text{FMT type} + \text{medication} + (1|\text{subject})$, (2) where the subject was
170 included as a random effect to account for the correlations in the repeated measures.
171 Timepoint (any pairs of the combinations of baseline, 1 month after FMT, 3 months
172 after FMT, 6 months after FMT, 12 months after FMT, with the previous time point
173 as the reference group), gender (female / male), ethnicity (Hispanic / non-Hispanic),
174 pancolitis (no / yes), CDI history (no / yes), FMT type (autologous / heterologous),
175 and medication (no / yes) were category variables. And age was a continuous
176 variable. Significant associations were defined as those with BH-FDR q value of the

177 corresponding coefficient below the threshold of 0.05.
178 3. In Figure 3, time points, CDI history, medication, ethnicity, and FMT type explained
179 a significant variance of metabolites in UC cases. To figure out the influence of these
180 confounding factors in the FMT process for UC cases, we fit the following per-
181 feature linear fixed-effects model within each time point. Medications were not
182 involved in the analysis due to their complicated prescriptions, however patient
183 receiving concurrent antimicrobial

184 medications, probiotics, received or are receiving chemotherapy were excluded from
185 the study.. Feature ~ (intercept) +ethnicity + CDI history + FMT type, (3) where
186 ethnicity (Hispanic / non-Hispanic), CDI history (no / yes), and FMT type
187 (autologous / heterologous) were category variables. Significant associations were
188 defined as those with BH-FDR q value of the corresponding coefficient below the
189 threshold of 0.25.

190

191 **RESULTS:**

192 **Demographics**

193 Key patient information and demographics can be found in Table 1. In total, we included
194 30 healthy children and 46 pediatric UC patients. UC patients received FMTs. In this
195 paper, we focused on metabolome profiles for UC patients at 5 different time points,
196 including baseline (before FMT), 1 month after FMT, 3 months after FMT, 6 months
197 after FMT, and 12 months after FMT. The average age of 30 healthy children was
198 approximately 14 years, while pediatric UC patients were approximately 16 years of
199 age. There were differences between the ethnicities for healthy and UC subjects. As
200 shown in Table 1, all the healthy individuals were non- Hispanics, while 52% of UC
201 patients were non-Hispanics.

202 Of the children with UC, 30 out of 46 patients had pancolitis, and half had CDI
203 (*Clostridioides difficile* infection) histories. 12 pediatric UC patients received autologous
204 FMT, while the rest received heterologous FMT. In addition, 42 out of 46 patients received
205 one or several therapies before FMT, such as proton pump inhibitors, biologic therapy,
206 immunomodulators, 5- aminosalicylates, and antibiotics. For simplicity, we considered
207 whether they were taking medications or not.

208

209

210

211

212

213

214

215

216

217
218
219
220
221
222
223
224
225

	Healthy	Ulcerative Colitis
Number of patients	30	46
Age (average)	14.43	16.41
Gender (males)	14	27
Ethnicity (Hispanic)	0	22
Pancolitis (yes)	NA	30
Number of patients having CDI history	NA	23
Number of patients having autologous FMT	NA	12
Endoscopic Mayo score (average)	NA	2.26
Number of patients taking medications	NA	42
Average BMI	NA	22

Table 1: Demographics of pediatric healthy controls and UC cases

226

227

Donor Microbiota Profile

228

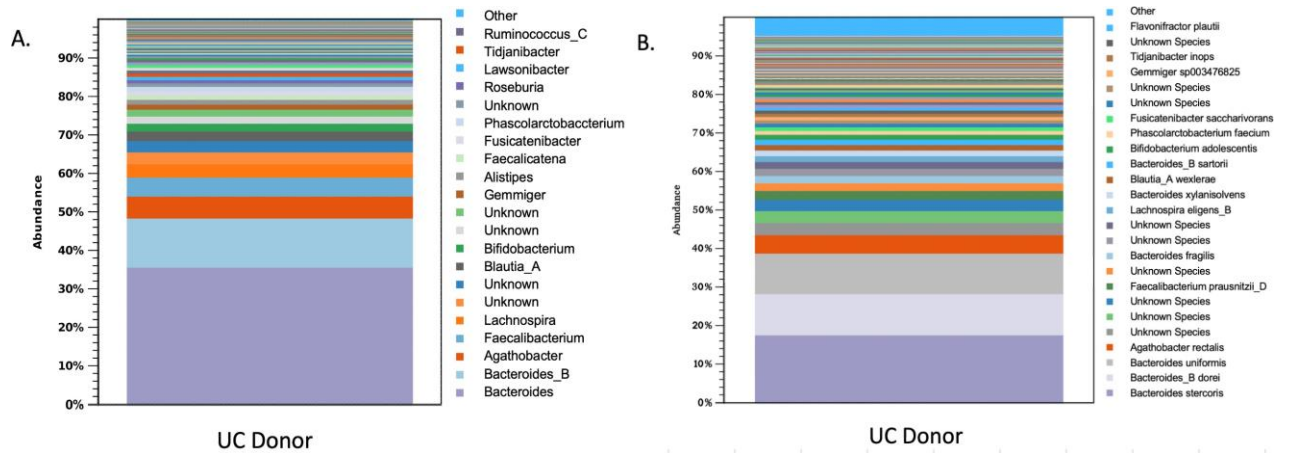
229

The microbiota had a Shannon diversity index of around 6.5, with the dominant microbiota consisting of *Firmicutes* and *Bacteroidetes*. There was also an abundance of *Faecalibacterium prausnitzii*, lactobacilli, *Bacteroides*, and *bifidobacterial* (Figure 1).

230

231

232



233

234

Figure 1: Shows the microbiota profile of donors by (A.) genus and (B.) species.

235

236

237

Metabolomes of pediatric UC patients shifted toward healthy profiles after FMT

238

239

240

241

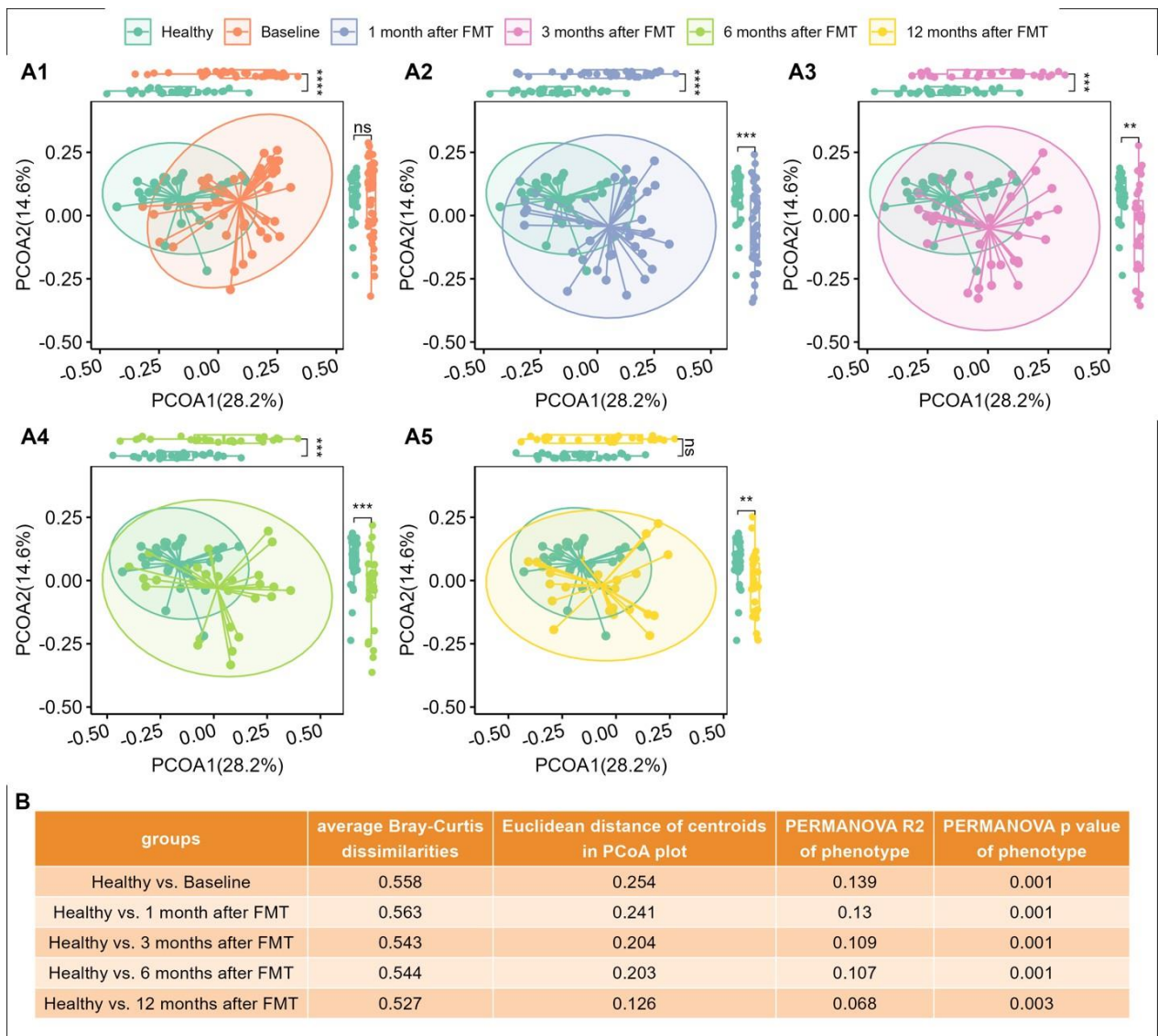
242

We computed the Bray Curtis distance between any pair of samples and used two-dimensional PCoA plots to visualize the samples based on metabolomes profiles. **Figure 2 A1** shows that Healthy and UC baseline samples clustered into two groups according to their phenotypes.

Wilcoxon tests revealed that the PCoA1 of UC patients significantly differed from that of healthy individuals (**Figure 2 A1**), although the PCoA2 did not vary statistically because of overdispersion (**Figure 2 AI**).

245

246 Figure 1B shows that the average Bray-Curtis dissimilarities, the Euclidean distances
 247 between the centroids, and the variance explained by the different groups between the
 248 healthy controls and the UC patients decrease with time after FMT. The last column
 249 of Figure 1C shows that although still significant, the variance explained by
 250 phenotype (Healthy versus UC) was reduced. Figure 2 clearly shows that
 251 metabolomes of pediatric UC patients shifted toward healthy profiles after FMT.



252 Figure 2: The metabolomic profiles of UC patients progressed to healthy levels after
253 FMT. **A1- A5**, PCoA plots based on metabolomics (Bray Curtis dissimilarities on relative
254 abundance) for Healthy controls versus UC cases at Baseline (**A1**), Healthy controls
255 versus UC cases at 1 month after FMT, (**A2**), Healthy controls versus UC cases at 3
256 months after FMT (**A3**), Healthy controls versus UC cases after 6 months after FMT
257 (**A4**), and Healthy controls versus UC cases after 12 months after FMT (**A5**), separately.
258 Boxplots of PCoA and PCoA2 were shown in the margins of PCoA plots. Wilcoxon rank
259 sum tests were used to compare the differences between Healthy and UC subjects at
260 different time points after FMT, with ns (not significant) for $p > 0.05$, * for $p \leq 0.05$, **
261 for $p \leq 0.01$, *** for $p \leq 0.001$, and **** for $p \leq 0.0001$. **B**, Quantitative differences
262 between Healthy and UC subjects at different time points, including average Bray- Curtis
263 dissimilarities between two groups, Euclidean distance between centroids of two groups
264 in PCoA plots (**A1-A5**), variance explained (R²) by phenotype (Healthy versus UC) and
265 respective p-value determined by PERMANOVA on metabolomics (Bray Curtis distance
266 on relative abundance).

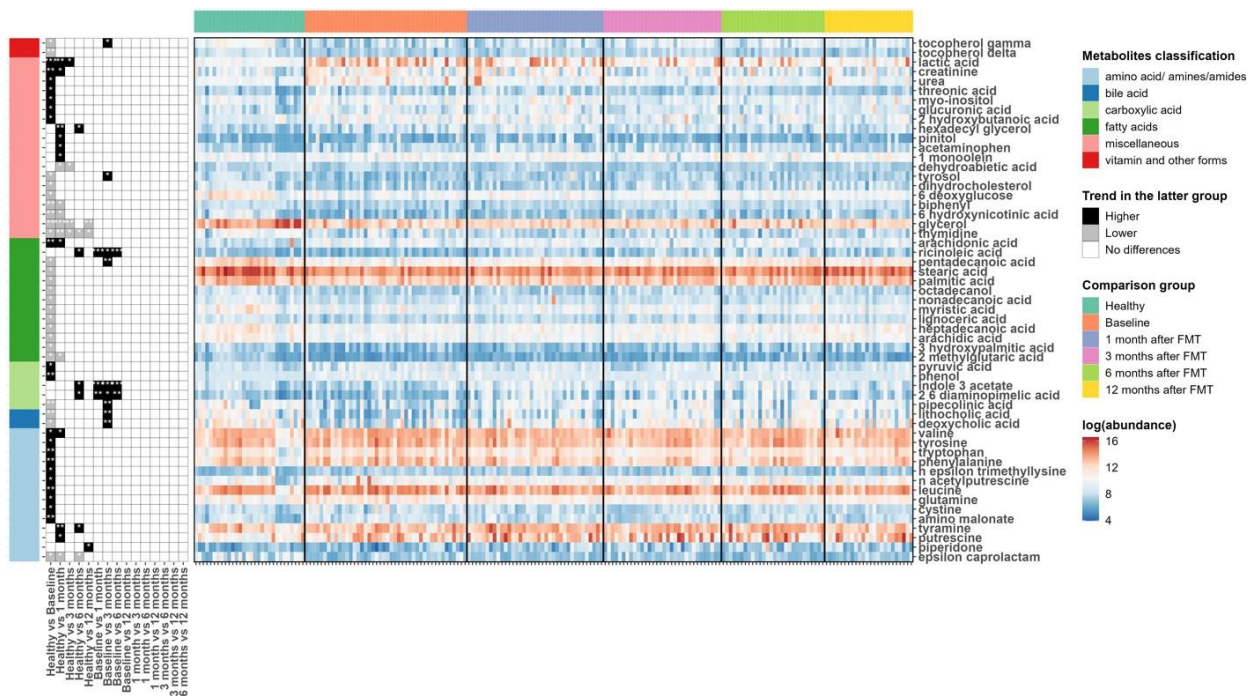
267 Metabolites changes in pediatric UC patients

268

269 We next used a linear fixed-effects model (equation (1)) by MaAsLin 2 to identify
270 metabolites associated with pediatric UC. A total of 230 metabolites were tested and
271 metabolites with Benjamini-Hochberg FDR less than 0.05 are shown in **Figure 2**.
272 Among the differentially abundant metabolites, 10 amino acids (amino malonate,
273 cysteine, glutamine, leucine, n- acetylputrescine, n-epsilon-trimethyllysine,
274 phenylalanine, tryptophan, tyrosine, and valine), 2- carboxylic acid (phenol and pyruvic
275 acid), 1 fatty acid (arachidonic acid), and 7 miscellaneous (2-hydroxybutanoic acid,
276 creatinine, glucuronic acid, lactic acid, myo-inositol, threonic acid, and urea) were
277 significantly higher in UC patients compared to healthy individuals. While 1 amino
278 acid (epsilon-caprolactam), 2 bile acids (deoxycholic acid and lithocholic acid), 1
279 carboxylic acid (pipercolinic acid), 11 fatty acids (2-methylglutaric acid, 3-
280 hydroxypalmitic acid, arachidic acid, heptadecanoic acid, lignoceric acid, myristic acid,
281 nonadecanoic acid, octadecanol, palmitic acid, pentadecanoic acid, and stearic acid), 7
282 miscellaneous (6-deoxyglucose, 6-hydroxynicotinic acid, biphenyl, dehydrocholesterol,
283 glycerol, thymidine, and tyrosol), and 2 vitamin and other forms (delta-tocopherol and
284 gamma-tocopherol) tended to be reduced in UC patients.

285 Linear mixed-effects models (equation (2)) were fitted to recognize the metabolites with
286 significant changes post-FMT in UC patients. Many metabolites were noted to increase
287 in UC patients after FMT when compared to their baseline metabolomic profiles. Indole-
288 3-acetate, 2,6- diaminopimelic acid, and ricinoleic acid quickly responded to FMT and
289 kept sustained growth for 6 months after one FMT infusion . Deoxycholic acid,
290 lithocholic acid, 3 hydroxyphenyl acetic acid, phenylacetic acid, pipercolinic acid,
291 pentadecanoic acid, 3,4-hydroxyphenyl propionic

292 acid, 3-hydroxybenzoic acid, dihydro-3-coumaric acid, glutaric acid, indole-3-propionic
 293 acid, ribose, tyrosol, nicotinic acid, and gamma-tocopherol increased for the first 3 months
 294 post one FMT infusion. Phenylacetic acid continued to increase for a total of 6 months
 295 after FMT. At 12 months after one FMT infusion, most metabolites in UC patients
 296 increased, with the exception of glycerol and thymidine which decreased.



297
 298 Figure 3: Heatmaps showing the log-transformed abundance of differentially abundant
 299 metabolites identified by MaAsLin2. Metabolites were grouped according to their
 300 classifications (left bar), and the samples were grouped by their phenotypes and time
 301 points after FMT for UC patients (top bar). The panel in the left column indicated the
 302 coefficients and BH-FDR corrected q values for the coefficients from the linear fixed
 303 effects model (Healthy versus UC, with Healthy as the reference group) or linear mixed
 304 effects model (UC versus UC at different time points, with UC patients at the previous
 305 time point as reference group), with black for higher abundances in the latter group, grey
 306 for lower in the latter group, and white for no significant differences between two groups,
 307 and * for FDR corrected q value < 0.05, ** for FDR corrected q value < 0.01, and ***
 308 for FDR corrected q value < 0.001. The panel in the right column showed the log-
 309 transformed abundance of differentially abundant metabolites.

310 **The effects of FMT were time-limited**

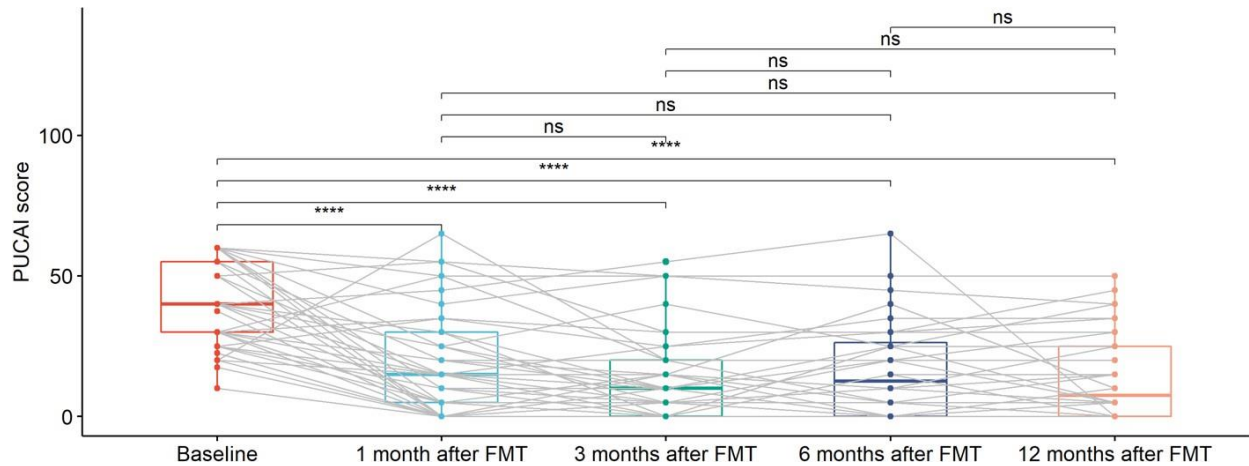
311 PUCAI scores were calculated at baseline and at months 1, 3, 6, and 12 after FMT.

ID	PUCA	EM	P1	P3	P6	P12	ID	PUC	EM	P1	P3	P6	P12
	I	S						AI	S				
1	60	3	50	50	50	50	26	55	2	0	0	0	0
2	60	3	55	50	45	40	27	60	3	25	40	25	25
3	20	1	15	0	0	10	28	55	3	0	0	15	15
4	20	1	15	0	0	10	29	60	3	55	–	–	–
5	30	2	35	30	30	35	30	25	2	10	5	0	5
6	30	2	35	30	30	35	31	40	3	5	5	10	5
7	60	2	0	0	15	0	32	60	3	–	–	–	–
8	50	2	15	10	25	40	33	30	2	50	20	5	5
9	25	1	15	15	0	0	34	50	1	15	5	20	5
10	40	2	30	0	25	5	35	25	2	10	0	15	0
11	60	2	10	10	0	0	36	50	2	0	5	0	5
12	60	2	5	0	0	5	37	60	3	25	5	5	0
13	10	1	0	0	0	10	38	10	1	0	0	0	0
14	35	1	0	0	0	10	39	40	3	15	15	10	25
15	25	3	35	25	35	35	40	30	3	20	20	20	20
16	60	3	40	50	45	40	41	30	3	15	25	30	45
17	30	2	0	10	0	5	42	30	3	20	15	40	10
18	50	3	55	30	30	30	43	40	3	15	25	30	45
19	55	2	20	20	5	0	44	35	3	0	0	0	0
20	20	2	0	15	10	15	45	40	3	0	0	0	0
21	25	3	5	15	5	15	46	20	1	0	0	0	0
22	40	3	45	55	65	0	47	15	1	0	0	0	0
23	40	1	25	10	20	30	48	60	3	0	0	0	0
24	20	2	65	20	20	20	49	30	1	20	10	25	5
25	30	3	5	5	5	5	50	30	1	20	15	40	10

312 **Table 2:** Baseline PUCAI and Endoscopic Mayo scores for 50 pediatric UC patients. PUCAI
 313 scores at months 1, 3, 6, and 12 after FMT initiation. EMS=Endoscopic Mayo Score, P1=PUCAI
 314 score month 1, P3=PUCAI score month 3, P6=PUCAI score month 6, P12=PUCAI score month
 315 12.

316

317 As shown in **Figure 4**, there was a statistically significant decreasing trend of PUCAI scores at
 318 1 month after FMT, which continued until 3 months after FMT. PUCAI scores remain flat
 319 starting 6 months after FMT. These may imply that the effects of FMT were time limited.

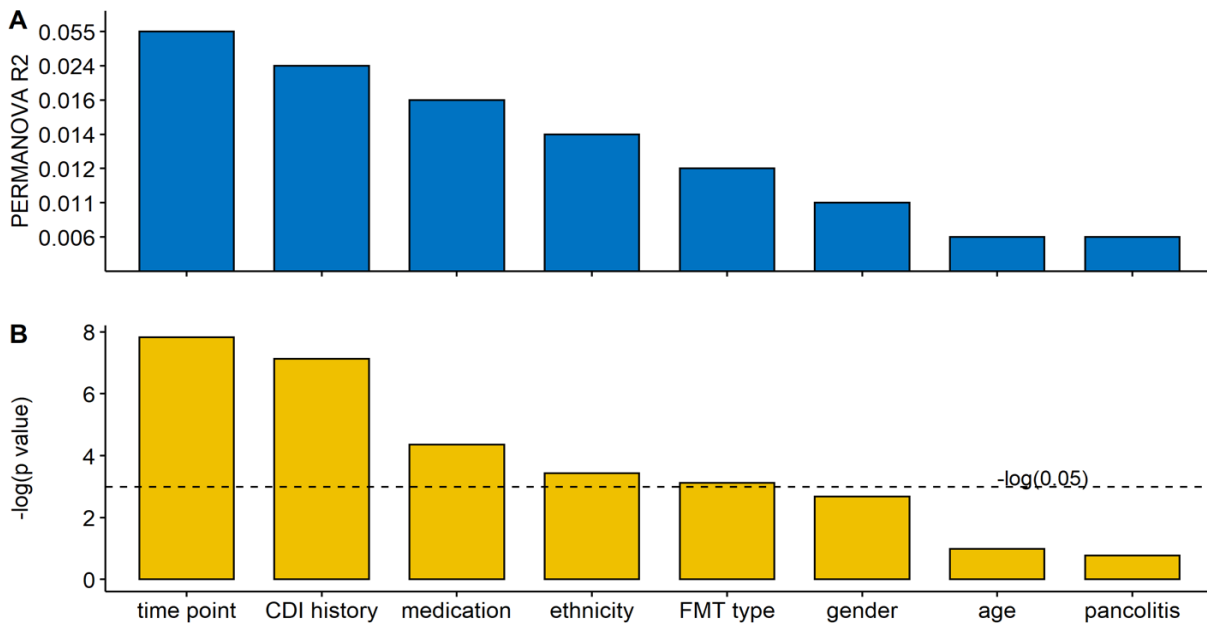


320

321 Figure 4: Boxplots of PUCAI scores for UC patients at different time points, including baseline,
 322 1 month after FMT, 3 months after FMT, 6 months after FMT, and 12 months after FMT. The
 323 gray lines in the figure mark the trajectories of each patient over time. Pairwise comparisons
 324 were performed using paired Wilcoxon rank sum tests, with ns (not significant) for $p > 0.05$ and
 325 **** for $p \leq 0.0001$. Two patients with missing data (UC012 with 3, 6, 12 months after FMT
 326 missing and UC015 with 1, 3, 6, 12 months after FMT missing) were not included in the analysis.

327 **CDI history, ethnicity, and FMT type influenced the FMT response of**
 328 **pediatric UC patients**

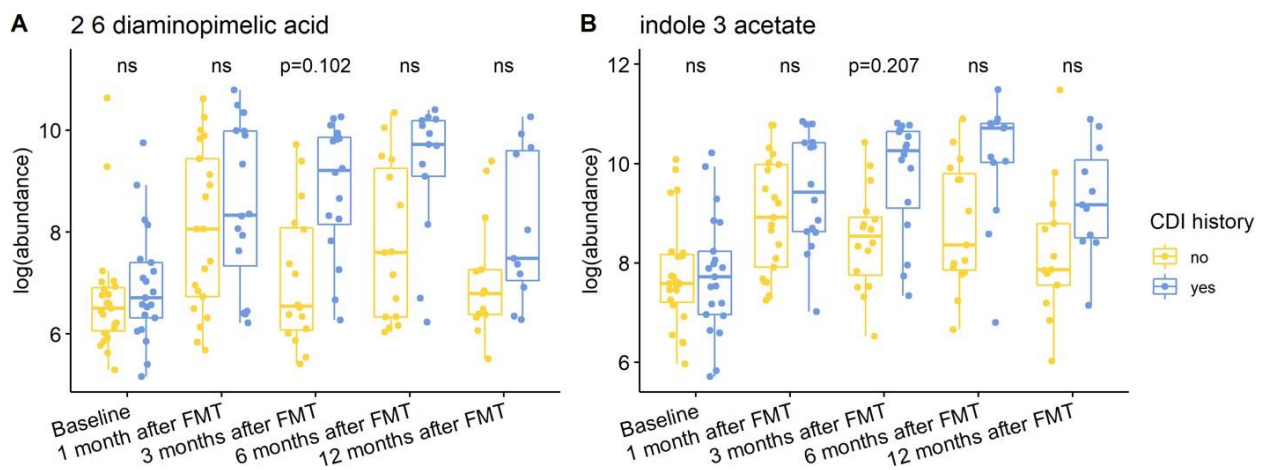
329 Focusing on UC patients, we analyzed the impact of confounding factors such as age and
 330 gender. A total of 8 different metadata were collected, including time point, CDI history,
 331 medication, ethnicity, FMT type, gender, age, and pancolitis. R2 values and the p-values
 332 from PERMANOVA analysis for each variable are shown in **Figure 5**. Timepoint had
 333 the largest interaction with gut metabolite composition. The other significant confounding
 334 factors were CDI history, medication, ethnicity, and FMT type. Medications were not
 335 involved in the analysis due to their complicated prescriptions. We fit a linear model
 336 (equation (3)) within each time point to determine their influence on the FMT process.
 337



338
 339 **Figure 5: Multivariate analysis showing the amount of inferred variance explained (R2)**
 340 **(A) by each covariate and respective p-value (B) determined by PERMANOVA on**
 341 **metabolomics (Bray Curtis distance on relative abundance). The variance explained by**
 342 **each variable was calculated independently of other variables (the sole variable in the**
 343 **model) to avoid issues related to variable ordering. Time points, CDI history, medication,**
 344 **ethnicity, and FMT type explained a significant but limited fraction of UC patients' total**
 345 **variation in metabolomics.**

346 Individuals with a history of CDI maintained relatively higher levels of 2,6-
 347 diaminopimelic acid and indole-3-acetate post-FMT (**Figure 6**). The abundance of these
 348 two metabolites was also higher in Hispanic patients 3 months post-FMT (**Figure 7**).
 349 Threonic acid was significantly lower in Hispanic patients at 3 months post-FMT. In
 350 contrast, UDP-glucuronic acid was significantly higher in Hispanics at 3 months after
 351 FMT. Lactose decreased in autologous FMT while 2,6-diaminopimelic and fructose
 352 increased in autologous FMT 3 months after FMT (**Figure 8**).

353



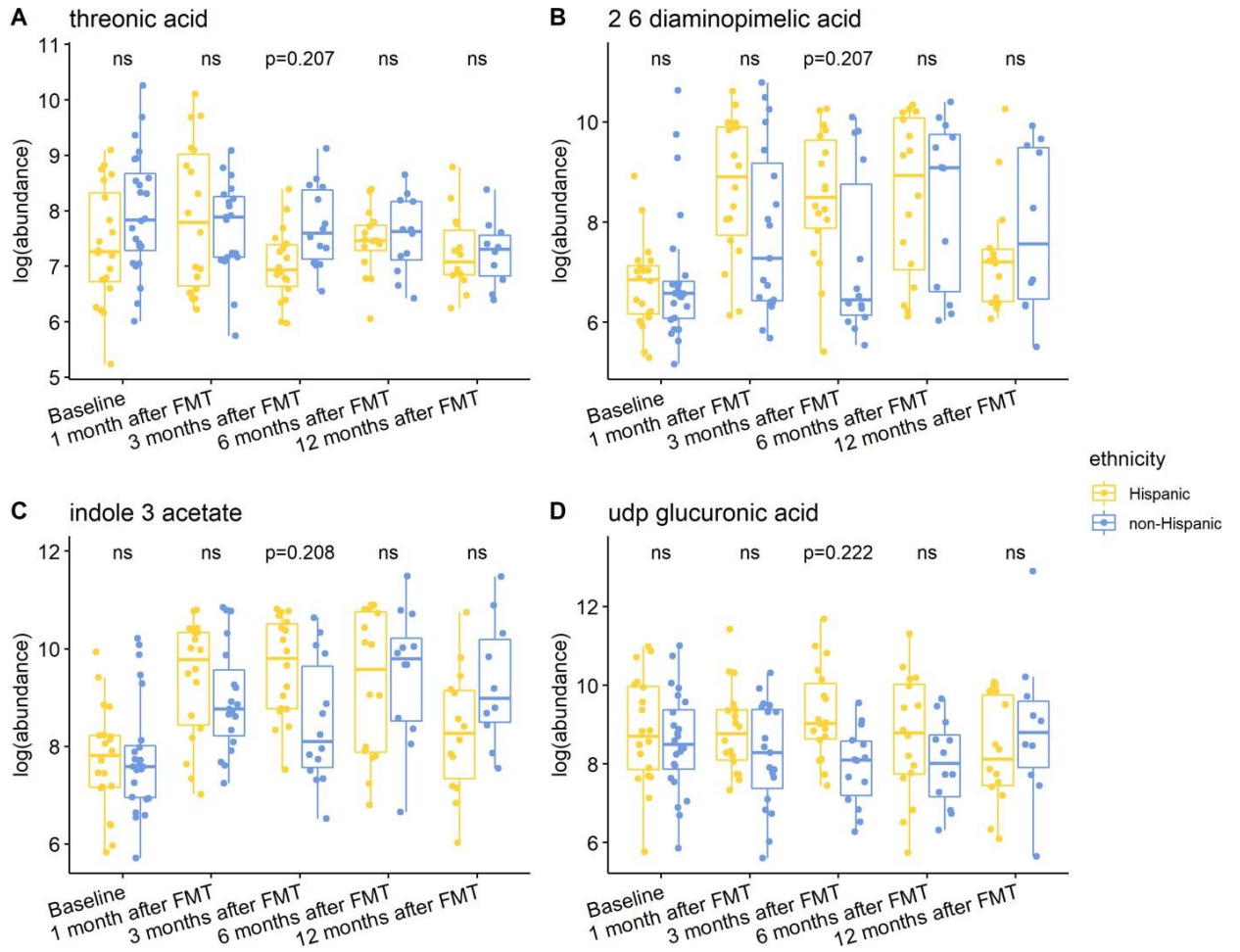
354

355 Figure 6: Boxplots of the log-transformed abundance of CDI history related metabolites
 356 for UC patients at different time points, including baseline, 1 month after FMT, 3 months
 357 after FMT, 6 months after FMT, and 12 months after FMT. The differences between
 358 patients with and without CDI history were tested using a linear model (equation (3))
 359 within each time point, with ns for not significant, and BH-FDR corrected q value
 360 annotated.

361

362

363



364

365

366

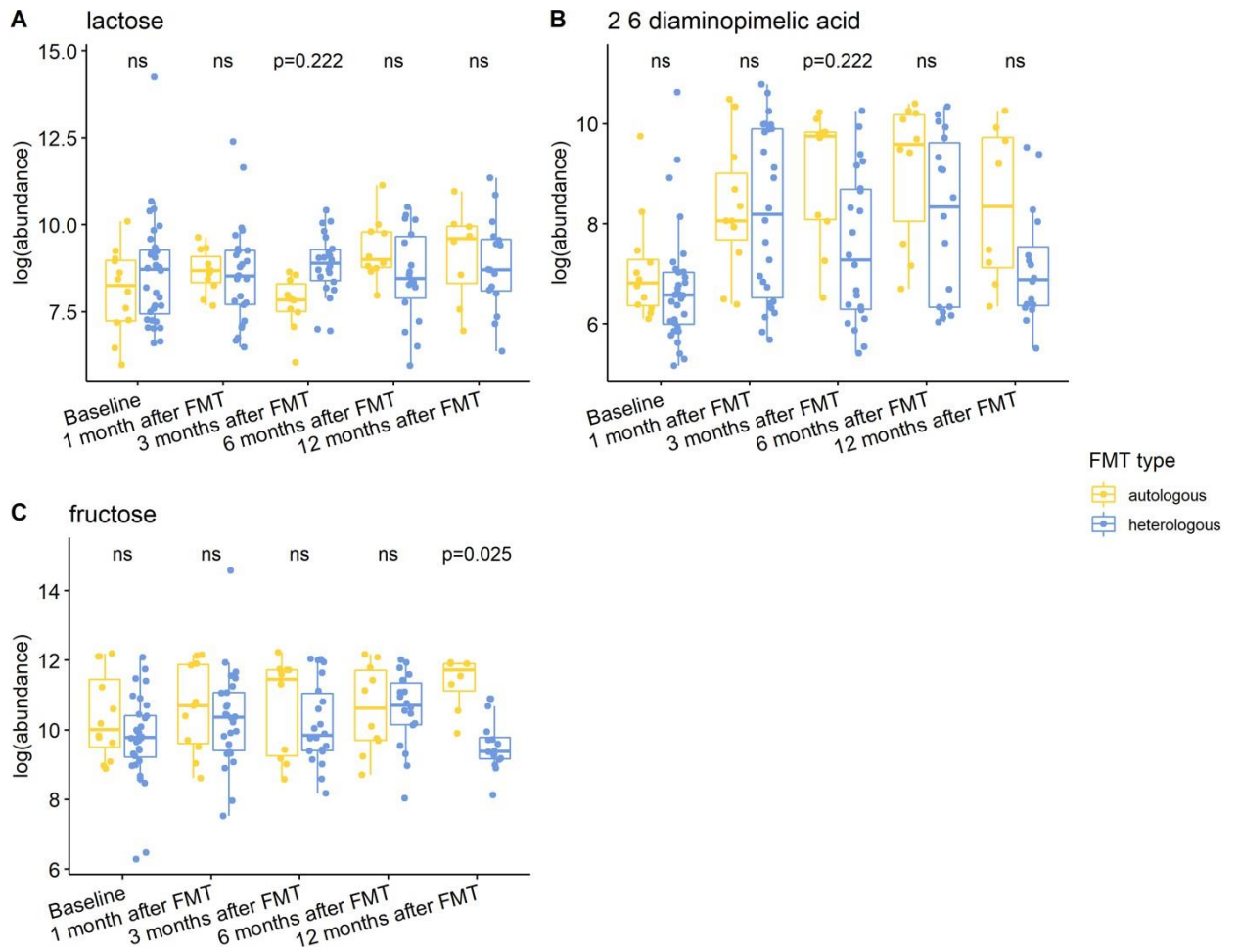
367

368

369

Figure 7: Boxplots of the log-transformed abundance of ethnicity related metabolites for UC patients at different time points, including baseline, 1 month after FMT, 3 months after FMT, 6 months after FMT, and 12 months after FMT. The differences between Hispanic and non-Hispanic were tested using a linear model (equation (3)) within each time point, with ns for not significant, and BH-FDR corrected q value annotated.

370
371
372



373
374
375
376
377
378
379

Figure 8: Boxplots of the log transformed abundance of FMT type related metabolites for UC patients at different time points, including baseline, 1 month after FMT, 3 months after FMT, 6 months after FMT, and 12 months after FMT. The differences between patients taking autologous FMT and patients taking heterologous FMT were tested using a linear model (equation (3)) within each time point, with ns for not significant, and BH-FDR corrected q value annotated.

380

DISCUSSION

381

382

383

384

385

386

387

388

389

390

391

392

393

394

395

396

397

398

399

400

401

402

403

404

Fatty Acids

405

406

407

408

409

Previous studies have investigated the role of dietary FAs in the pathophysiology of IBD (Ananthakrishnan et al., 2014). Our study found eleven FAs that declined in UC patients compared to healthy controls. These fatty acids included one methyl branched (2-methylglutaric acid), ten saturated long chain fatty acids (3-hydroxypalmitic acid, arachidic acid, heptadecanoic acid, lignoceric acid, myristic acid, nonadecanoic acid,

410 octadecanoyl, palmitic acid, pentadecanoic acid, and stearic acid). One polyunsaturated
411 fatty acid (arachidonic acid) was higher in UC patients than healthy controls.

412

413 *Long chain fatty acids*

414 It has been theorized that the length of FAs plays a crucial role in its effect on gut health,
415 but few have highlighted the role of LCFAs (Chunxiang and Zhang, 2019), defined as
416 FAs with a carbon chain length of 13-21 (Galli, 2009). Our study identified eleven
417 LCFAs of interest in UC patients, including ten saturated and one unsaturated
418 LCFAs. Most statistically significant LCFAs began trend towards healthy control levels
419 within 1-6 months post-FMT, suggesting that FMT can alter the gut's LCFA profile.

420

421 There has also been debate regarding the impact of saturated versus unsaturated LCFAs
422 in strengthening the intestinal barrier (Berengere Benoit et al., 2015). A study done by
423 Benoit et al., showed that saturated LCFAs, such as palmitic acid and palm oil, enhanced
424 MUC2 synthesis and promoted differentiation of goblet cells which could be beneficial
425 to intestinal health in IBD (Berengere Benoit et al., 2015). When evaluating the trends of
426 our saturated LCFAs, we found that palmitic acid and its derivative 3-hydroxypalmitic
427 acid were declined in UC patients compared to healthy controls.

428

429 Arachidonic acid, a polyunsaturated LCFAs, was found to be higher in UC patients
430 compared to healthy controls. There have been mixed theories on the role of
431 polyunsaturated FAs in the mucosal health of ulcerative colitis patients. Arachidonic acid
432 has been documented in previous literature to be elevated in ulcerative colitis patients
433 due to impact of inflammation on the composition of phospholipids in colonic mucosa
434 (Nishida et al., 1987). Previous literature has also found that monounsaturated fatty acids
435 (MUFA) may play a role in the regulation of gut microbiota and inflammation (Flavia
436 Galvao Candido et al., 2017). Additional studies have found cis-palmitoleic acid
437 decreases inflammation in the gut of UC patients by increasing the expression of HNF4 α
438 and HNF4 γ (Hernandez et al., 2017).

439

440

Methyl-branched Fatty Acid

441

442

443

444

445

446

447

448

449

450

451

452

453

Amino acids

454

455

456

457

458

459

460

461

462

463

464

465

466

Vitamins

467

468

Vitamins have been theorized to play a role in intestinal barrier function and modulation of gut microbiota. Tocopherol/Vitamin E has previously been linked to protecting

469 intestinal barrier function and modulating the gut microbiota (Liua et al., 2021). Our study
470 found a baseline deficiency in Tocopherol with a gradual rise in levels after FMT. These
471 deficiencies may indicate a lack of protective metabolites in the gut of UC patients.

472

473

474 **Secondary Bile Acids**

475 Secondary bile acids have been theorized to have anti-inflammatory and cytoprotective
476 actions and have been a metabolite of interest as a potential therapeutic option. Our study
477 also found a deficiency in deoxycholic and lithocholic acid at baseline in UC patients,
478 which may suggest a lack of protective effects. Ward et al., investigated the impact of
479 secondary bile acids on cytokine release from colonic epithelial cells. They found that
480 lithocholic acid potently inhibited epithelial cytokine release and protected against
481 mucosal inflammation (Ward et al.,2017).

482

483 **Nucleosides and Nucleobases**

484 DNA nucleosides, specifically thymidine, and their impaired incorporation into DNA
485 have previously been linked to colitis regardless of disease severity (Alpers et al.,1980).
486 However, our study found that at baseline UC patients were deficient in thymidine.

487

488 **Conclusion**

489 This study showed that baseline metabolite profiles in UC pediatric patients are different
490 from healthy subjects. Additionally, we demonstrated that FMT alters metabolite
491 profiles. Our data suggested a time-dependent trend from UC-type to healthy profiles
492 after FMT. FMT may be effective in altering the inflammatory state of the gut by
493 increasing the abundance of anti-inflammatory metabolites and decreasing pro-
494 inflammatory metabolites. Future studies should investigate the timing and need for
495 repeat FMT and follow their trends. Overall, our study was notable for a significant
496 baseline difference in the metabolomic profiles and a significant difference at follow-up
497 months 1, 3, 6, and 12 when comparing UC to healthy controls. These differences were

498 primarily seen with FAs, AAs, nucleosides/nucleobases, vitamins, and bile acids. Further
499 studies with larger sample populations are needed to identify significant differences in
500 specific metabolites and their trends post-FMT.

501

502 **Limitations**

503

504 Limitations of the study include inability to control for patient's environment and diet at
505 home, loss of data points due to loss of follow-up, and insufficient data on very early onset
506 ulcerative coli **Acknowledgement**

507 We thank UC Davis West Coast Metabolomics Center for their expertise and support in
508 metabolite. Author Contributions

509 All authors made significant contributions to the paper, including the conception and
510 design of the study, acquisition of data, analysis, and interpretation of data, drafting the
511 article or revising it critically for important intellectual content, and final approval of
512 the version to be submitted.

513

514 **Financial Support:**

515

516 This work was supported by the National Institutes of Health [grant number
517 R01HD081197 to S.M.]; the National Center for Advancing Translational Science [grant
518 numbers UL1TR001855, UL1TR000130]; and the Biostatistics Core of the Saban
519 Research Institute at Children's Hospital Los Angeles.

520 **Conflict of interest:**

521 Authors Parastou Khalessi Hosseini, Beibei Wang, Fengzhu Sun, Yihui Luan, and Sonia
522 Michail certify that they have NO affiliations with or involvement in any organization or
523 entity with any financial interest (such as honoraria; educational grants; participation in
524 speakers' bureaus; membership, employment, consultancies, stock ownership, or other
525 equity interest; and expert testimony or patent-licensing arrangements), or non-financial
526 interest (such as personal or professional relationships, affiliations, knowledge or beliefs)
527 in the subject matter or materials discussed in this manuscript

528 **Data Sharing**

529

530 Details regarding access to data, materials, protocols, and software will be made
531 available upon request.

532
533
534
535
536
537
538
539
540
541
542
543
544
545
546
547
548
549
550
551
552
553
554
555
556
557
558
559
560
561
562

References

1. Abraham, B. P., Mehta, S., & El-Serag, H. B. (2012). Natural history of pediatric-onset inflammatory bowel disease: A systematic review. *Journal of Gastroenterology*, 46(7), 581–589.
2. Desreumaux, P., & Colombel, J. F. (2004). Dysbiosis in inflammatory bowel disease. *Gut*, 53(1), 1-4.
3. Lepage, P., Colombet, J., Marteau, P., Sime-Ngando, T., Doré, J., & Leclerc, M. (2008). Dysbiosis in inflammatory bowel disease: A role for bacteriophages? *Gut*, 57(3), 424-425.
4. Gevers, D., Kugathasan, S., Denson, L. A., Vázquez-Baeza, Y., Van Treuren, W., Ren, B., ... & Morgan, X. C. (2014). The treatment-naive microbiome in new-onset Crohn's disease. *Cell Host & Microbe*, 15(3), 382-392.
5. Scoville, E. A., Allaman, M. M., Brown, C. T., Motley, A. K., Horst, S. N., Williams, C. S., ... & Schwartz, D. A. (2018). Alterations in lipid, amino acid, and energy metabolism distinguish Crohn's disease from ulcerative colitis and control subjects by serum metabolomic profiling. *Metabolomics*, 14(1), 17.
6. Sinha, S. R., Haileselassie, Y., Nguyen, L. P., Tropini, C., Wang, M., Becker, L. S., ... & Namkoong, H. (2020). Dysbiosis-induced secondary bile acid deficiency promotes intestinal inflammation. *Cell Host & Microbe*.
7. Diab, J., Hansen, T., Goll, R., Stenlund, H., Jensen, E., Moritz, T., ... & Forsdahl, G. (2019). Mucosal Metabolomic Profiling and Pathway Analysis Reveal the Metabolic Signature of Ulcerative Colitis. *Metabolites*, 9(12), 291.
8. Shores, D. R., Binion, D. G., Freeman, B. A., & Baker, P. R. (2011). New insights into the role of fatty acids in the pathogenesis and resolution of inflammatory bowel disease. *Inflammatory Bowel Diseases*, 17(10), 2192-2204.
9. Staley, C., Weingarden, A. R., Khoruts, A., & Sadowsky, M. J. (2017). Interaction of gut microbiota with bile acid metabolism and its influence on disease states. *Applied Microbiology and Biotechnology*, 101, 47-64.
10. Davidovics, Z. H., et al. (2019). Fecal Microbiota Transplantation for Recurrent *Clostridium difficile* Infection and Other Conditions in Children: A Joint

- 563 Position Paper From the North American Society for Pediatric
564 Gastroenterology, Hepatology, and Nutrition and the European Society for
565 Pediatric Gastroenterology, Hepatology, and Nutrition. *Journal Pediatric*
566 *Gastroenterology and Nutrition*, 68(1), 130-143.
- 567 11. Relman, D. A., et al. (2013). Current Consensus guidance on donor screening
568 and stool testing for FMT. *Infectious Disease Society of America*, Arlington,
569 VA.
- 570 12. Bakken, J. S., et al. (2011). Treating *Clostridium difficile* infection with fecal
571 microbiota transplantation. *Clinical Gastroenterology and Hepatology*, 9(12),
572 1044-1049.
- 573 13. Owens, C., Broussard, E., & Surawicz, C. (2013). Fecal microbiota
574 transplantation and donor standardization. *Trends in Microbiology*, 21(9), 443-
575 445.
- 576 14. Nusbaum, D. J., et al. (2018). Gut microbial and metabolomic profiles after
577 fecal microbiota transplantation in pediatric ulcerative colitis patients. *FEMS*
578 *Microbiology Ecology*, 94(9).
- 579 15. Team, R. C. (2021). R: A Language and Environment for Statistical Computing.
- 580 16. Bray, J. R., & Curtis, J. T. (1957). An ordination of the upland forest
581 communities of southern Wisconsin. *Ecological Monographs*, 27(4), 326-349.
- 582 17. Oksanen, J., et al. (2020). *vegan: Community Ecology Package*.
- 583 18. Dray, S., Legendre, P., & Peres-Neto, P. R. (2006). Spatial modelling: A
584 comprehensive framework for principal coordinate analysis of neighbour
585 matrices (PCNM). *Ecological Modelling*, 196(3-4), 483-493.
- 586 19. Paradis, E., & Schliep, K. (2019). *ape 5.0: An environment for modern*
587 *phylogenetics and evolutionary analyses in R*.
- 588 20. Anderson, M. J. (2001). A new method for non-parametric multivariate analysis
589 of variance. *Austral Ecology*, 26(1), 32-46.
- 590 21. Lloyd-Price, J., et al. (2019). Multi-omics of the gut microbial ecosystem in
591 inflammatory bowel diseases. *Nature*, 569(7758), 655-662.
- 592 22. Mallick, H., et al. (2021). Multivariable association discovery in population-
593 scale meta-omics studies. *PLoS Computational Biology*, 17(11), e1009442.

- 594 23. Nearing, J. T., et al. (2022). Microbiome differential abundance methods
595 produce different results across 38 datasets. *Nature Communications*, 13(1), 1-
596 16.
- 597 24. Sokol, H., & Secher, T. (2020). Gut microbiota-derived metabolites as key
598 actors in inflammatory bowel disease. *Nature Reviews Gastroenterology &*
599 *Hepatology*, 17, 223–237.
- 600 25. Alam, M. T., et al. (2020). Microbial imbalance in inflammatory bowel disease
601 patients at different taxonomic levels. *Gut Pathogens*, 12.
- 602 26. Moayyedi, P., et al. (2015). Fecal Microbiota Transplantation Induces
603 Remission in Patients With Active Ulcerative Colitis in a Randomized
604 Controlled Trial. *Gastroenterology*, 149(1), 102-109.
- 605 27. Ananthakrishnan, A. N., et al. (2014). Long-term Intake of Dietary Fat and Risk
606 of Ulcerative Colitis and Crohn’s Disease. *Gut*, 63(5), 776-784.
- 607 28. Ma, C., & Zhang, H. (2019). The Role of Long-Chain Fatty Acids in
608 Inflammatory Bowel Disease. *Mediators of Inflammation*, 2019.
- 609 29. Galli, W. M., & Hornstra, G. (2009). Fat and fatty acid terminology, methods of
610 analysis and fat digestion and metabolism: A background review paper. *Annals*
611 *of Nutrition & Metabolism*, 55(1-3), 8-43.
- 612 30. Benoit, B., et al. (2015). Saturated and unsaturated fatty acids differently
613 modulate colonic goblet cells in vitro and in rat pups. *The Journal of Nutrition*,
614 145(8), 1754-1762.
- 615 31. Nishida, T., et al. (1987). Increased arachidonic acid composition of
616 phospholipids in colonic mucosa from patients with active ulcerative colitis.
617 *Gut*, 28(8), 1002-1007.
- 618 32. Cândido, F. G., et al. (2017). Impact of dietary fat on gut microbiota and low-
619 grade systemic inflammation: Mechanisms and clinical implications on obesity.
620 *International Journal of Food Sciences and Nutrition*, 68(2), 149-158.
- 621 33. Bueno-Hernández, N., et al. (2017). Effect of cis-palmitoleic acid
622 supplementation on inflammation and expression of HNF4 γ , HNF4 α and IL6 in
623 patients with ulcerative colitis. *Minerva Gastroenterol Dietol*, 63(3), 257-263.
- 624 34. Connors, J., & Van Limbergen, J. (2019). The Role of Succinate in the

- 625 Regulation of Intestinal Inflammation. *Nutrients*, 11(1), 25.
- 626 35. Ooi, M., et al. (2011). GC/MS-based profiling of amino acids and TCA cycle-
627 related molecules in ulcerative colitis. *Inflammation Research*, 60, 831-840.
- 628 36. Nie, C., et al. (2018). Branched chain amino acids: Beyond nutrition
629 metabolism. *International Journal of Molecular Sciences*, 19(4), 954.
- 630 37. Papada, E., et al. (2020). The Association of Plasma-Free Branched-Chain
631 Amino Acids with Disease Related Parameters in Ulcerative Colitis.
632 *Diagnostics*, 10(10), 798.
- 633 38. Liu, C. X., et al. (2021). Vitamin E alpha- and gamma-tocopherol mitigate
634 colitis, protect intestinal barrier function and modulate the gut microbiota in
635 mice. *Free Radical Biology and Medicine*, 163, 180-189.
- 636 39. Ward, J. B. J., et al. (2017). Ursodeoxycholic acid and lithocholic acid exert
637 anti-inflammatory actions in the colon. *Gastrointestinal and Liver Physiology*,
638 312(6), G550-G558.
- 639 40. Alpers, D. H., et al. (1980). Control of thymidine incorporation in mucosal
640 explants from patients with chronic ulcerative colitis. *Gastroenterology*, 78(3),
641 470-478.

642