

Spatial summation in the glaucomatous macula: a link with retinal ganglion cell damage

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Abstract

Purpose: to test whether functional loss in the glaucomatous macula is characterised by an enlargement of Ricco's area (RA) through the application of a computational model linking retinal ganglion cell (RGC) damage to perimetric sensitivity

Methods: one eye from each of 29 visually-healthy subjects <40 years old, 30 glaucoma patients and 20 age-similar controls was tested with a 10-2 grid with stimuli of five different area sizes. Structural estimates of point-wise RGC density were obtained from Optical Coherence Tomography scans. Structural and functional data from the young healthy cohort were used to estimate the parameters of a computational spatial summation model to generate a template. The template was fitted with a Bayesian hierarchical model to estimate the latent RGC density in glaucoma patients and age-matched controls.

We tested two alternative hypotheses: fitting the data by translating the template horizontally (H_1 : change in RA) or vertically (H_2 : loss of sensitivity without change in RA). Root Mean Squared Error (RMSE) of the model fits to perimetric sensitivity were compared. 95%-Confidence Intervals were bootstrapped. The dynamic range of the functional and structural RGC density estimates was denoted by their 1st and the 99th percentile.

Results: the RMSE was 2.09 [1.92-2.26] under H_1 and 2.49 [2.24-2.72] under H_2 ($p < 0.001$). The average dynamic range for the structural RGC density estimates was only 11% that of the functional estimates.

Conclusions: macular sensitivity loss in glaucoma is better described by a model in which RA changes with RGC loss. Structural measurements have limited dynamic range.

1 Introduction

2 Glaucoma is characterized by progressive loss of the visual field (VF) as a consequence of damage to,
3 and death of, Retinal Ganglion Cells (RGCs).^{1,2} VF damage is usually detected and monitored with
4 Standard Automated Perimetry (SAP), in which circular stimuli of constant area and duration are
5 modulated in luminance on a uniform background at different VF locations. The test aims to
6 estimate, for each location, the stimulus luminance that represents the just noticeable difference
7 from the background luminance. This is expressed as VF sensitivity, where decibel units measure the
8 attenuation of the brightest stimulus (higher dB indicating dimmer stimuli). Despite a long-
9 established understanding that perimetric sensitivity is associated with RGC density,³⁻⁶ in that they
10 co-vary in disease such as glaucoma, their exact relationship has proven difficult to elucidate.

11 Useful insights into the pathophysiology of visual loss in glaucoma can be gathered by studying how
12 perimetric sensitivity changes with stimulus area. For a given duration and background luminance,
13 sensitivity is known to increase with the area of the stimulus (*spatial summation*)⁷. The change in
14 sensitivity is steeper and directly proportional to the area of the stimulus (*complete spatial*
15 *summation*) up to a certain critical area (Ricco's area, or the area of complete spatial summation).
16 After this point, sensitivity still increases with stimulus area but by a smaller amount (*partial*
17 *summation*). Ricco's area is known to enlarge with eccentricity and different stimulating conditions
18 and it has been hypothesized that a critical number of RGCs underlies Ricco's area across different
19 eccentricities⁸⁻¹⁴, this varying with adaptation level¹⁵. Similar scaling of Ricco's area with RGC density
20 has been hypothesized to hold true with RGC loss in glaucoma¹⁶. Redmond et al. demonstrated that
21 Ricco's area is enlarged in glaucoma, which can account for the difference in sensitivity between
22 patients and healthy controls for conventional Goldmann III stimuli¹⁶. Antwi-Boasiako et al showed
23 similar results in non-human primates¹⁷.

24 The use of computational models has been pivotal to the understanding of these phenomena.
25 Swanson et al.¹⁸ showed that spatial summation phenomena can be reproduced by a two-stage
26 hierarchical process involving RGC density as well as the spatial tuning of cortical filters, which can
27 be independent of the underlying density of RGCs. Further research by Pan & Swanson suggested
28 that probability summation across RGCs cannot explain spatial summation of perimetric stimuli,
29 whereas it may be explained instead by cortical pooling by multiple spatial mechanisms¹⁹. We have
30 recently proposed a computational model able to reproduce the interaction between stimulus area
31 and duration in the response of a synthetic RGC mosaic in healthy observers²⁰. In that work, we also
32 hypothesised, in partial agreement with Swanson et al.²¹, that the retinal input would determine the
33 selection of different cortical filters, altering spatial summation. We hypothesised that this retinal
34 input could also be altered by a change in the density of RGCs. Under this assumption, we showed
35 that our model would be able to reproduce the results presented by Redmond et al.¹⁶ in glaucoma.

36 Glaucoma damage in the macula has been documented extensively in the literature^{22,23}, but has
37 gained increasing attention in recent years after reports that it can be affected in early disease,²⁴⁻²⁶
38 albeit often going undetected clinically until later in the condition,^{27,28} and that it affects quality of
39 life of patients at all stages of disease²⁹. In the healthy eye, sensitivity measures with the Goldmann
40 III stimulus adopted in SAP (0.43 deg in diameter) in photopic conditions are determined by
41 complete spatial summation only outside the central 15 degrees^{8-10,21}. This means that early macular
42 damage from glaucoma would produce only small changes in SAP sensitivity until a very large
43 proportion of RGCs is lost^{16,18,30,31}. Despite its relevance, only two studies investigated spatial
44 summation in the glaucomatous macula, one in non-human primates¹⁷ and one in glaucoma
45 patients^{17,32}. However, they limited their analysis to early damage. Moreover, the investigation in
46 glaucoma patients³² only correlated sensitivity with coarse RGC count estimates from Optical

47 Coherence Tomography (OCT) imaging, rather than attempting to model the underlying latent
48 process of damage.

49 In the current study, we wished to test the hypothesis that changes in sensitivity in the macula of
50 patients with glaucoma could be explained by a change in the spatial scale used by the visual system
51 that relates to RGC loss or damage. Here, we perform five separate SAP examinations, each with a
52 different fixed-area luminance-modulated stimulus on a 10-2 grid, in eyes with glaucoma with
53 different levels of damage and age-similar healthy control eyes, as well as in young healthy eyes. We
54 then compare our functional RGC density estimates derived from the spatial summation model with
55 structural estimates from high-density OCT scans, to determine the extent to which VF damage can
56 be predicted from clinical measures of tissue loss in the macula.

57 Methods

58 Study population

59 Data were collected in the eye clinic at Santi Paolo e Carlo Hospital – University of Milan, Milan, Italy
60 and in the glaucoma clinic at IRCCS Fondazione G.B. Bietti, Rome, Italy.

61 Thirty young healthy participants were recruited among staff and students on a voluntary basis.
62 Inclusion criteria for this cohort were: 1) age between 18 and 40 years; 2) best corrected visual
63 acuity (BCVA) of 0 logMAR or better; 3) Intraocular pressure (IOP) < 21 mmHg; 4) no evidence of
64 ocular disease on preliminary ophthalmoscopic examination; 5) no history or evidence of systemic
65 disease that might affect the VF or compromise the execution of the test. Individuals were excluded
66 if the macular or optic nerve head (ONH) OCT scans collected for the study showed any signs of
67 ocular disease (details of the imaging and macular testing protocols are reported later). A 24-2
68 Swedish Interactive Thresholding Algorithm (SITA) VF test was performed for descriptive purposes
69 for the study but was not used to assess inclusion.

70 Glaucoma patients and the age-similar healthy participants were recruited on a voluntary basis.
71 Glaucoma patients' charts were screened by clinicians in order to identify potentially eligible
72 candidates. To meet eligibility criteria, patients were required to have a confirmed clinical diagnosis
73 of open angle glaucoma (which could include pseudoexfoliative and pigment dispersion glaucoma),
74 regardless of the integrity of their VF. Glaucoma patients were stratified by level of damage
75 according to the Mean Deviation (MD) value from their most recent reliable (FP < 15%) 24-2 SITA
76 test and classified as early (MD better than -6 dB), moderate (MD between -6 dB and -12 dB) or
77 advanced (MD worse than -12 dB), with the aim of recruiting 10 participants for each class. Other
78 inclusion criteria were: 1) age greater than 18 years; 2) BCVA of 0.2 logMAR or better; 3) no history
79 or evidence of other ocular or systemic diseases, other than glaucoma, that might affect the VF or
80 compromise the execution of the test. Age-matched controls were recruited among members of
81 staff and patients' spouses, partners and relatives. Inclusion criteria were the same as for the
82 healthy young cohort, but with no upward age limit and the requirement for VA to be better than or
83 equal to 0.2 logMAR.

84 Written informed consent was obtained from all participants. The study adhered to the tenets of the
85 Declaration of Helsinki and was approved by local ethics committees (Comitato Etico Milano Area 1 -
86 code OCU_SSSF; Comitato Etico Centrale IRCCS Lazio N. 90/19/FB).

87 Study protocol

88 All healthy participants underwent an ophthalmoscopic examination and measurement of their
89 BCVA and IOP (Goldmann Applanation Tonometry) in order to confirm eligibility. Their BCVA was not

90 tested beyond 0 logMAR. BCVA and IOP were not recorded for the study and only used to assess the
91 exclusion criteria. Axial length and corneal curvature were measured with an IOLMaster (Carl Zeiss
92 Meditec, Dublin, USA) and recorded for the study.

93 Only one eye per participant was included in the study. Where both eyes of healthy controls were
94 eligible, one was chosen arbitrarily by the researcher for testing. In the glaucoma cohort, if the two
95 eyes were classified as having a different stage of glaucoma, one was chosen to populate the
96 severity group, as needed. Otherwise, one was chosen arbitrarily by the researcher.

97 Standard Automated Perimetry

98 All VF tests were performed with a Humphrey Field Analyzer (HFA, Carl Zeiss Meditec, Dublin, USA).
99 Participants' near correction was used where required. For young healthy participants, near
100 correction was used according to their preference. All healthy participants underwent a 24-2 SITA
101 Standard test to obtain MD and Pattern Standard Deviation (PSD) values for descriptive purposes
102 and for the purposes of disease severity classification.

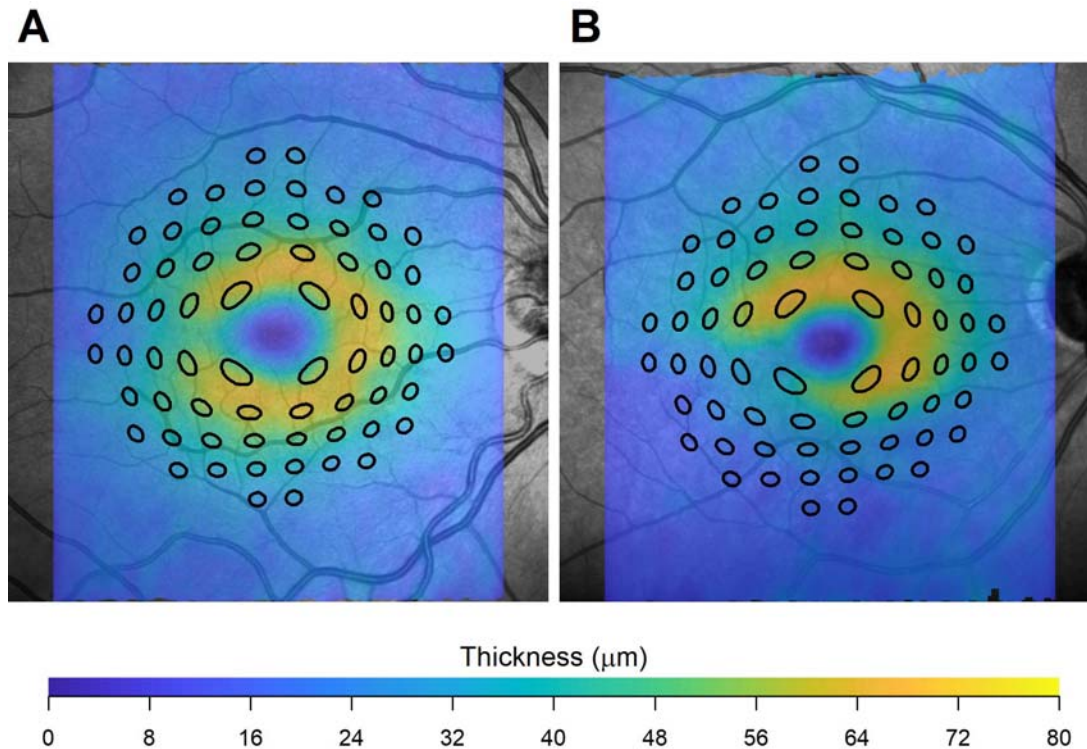
103 Separate macular perimetric tests were performed with a 10-2 grid, Full-threshold strategy, each
104 with a different Goldmann stimulus diameter (in degrees): G-I (0.10); G-II (0.21); G-III (0.43); G-IV
105 (0.86); G-V (1.72). The order of these tests was randomized following a computer generated
106 sequence of tests, one for each subject. For the young healthy cohort, the G-I test was repeated
107 twice, because results with this stimulus were expected to be more variable²⁰. For glaucoma patients
108 and age matched controls, the G-III test was performed twice instead, to produce a more reliable
109 estimate of the age-corrected sensitivity loss, because normative databases in the HFA are only
110 available for the G-III stimulus. All participants performed a total of six 10-2 SAP tests. Based on
111 previous literature for full-threshold tests,³³ reliability of the tests was only assessed with the
112 percentage of FP errors (< 33%). For the healthy participants, a limit of 33% on false negative errors
113 was also set. The operator was instructed to carefully monitor the participants and ensure good
114 fixation throughout the test. If unreliable, the test, but not the participant, was excluded from
115 analysis. Fixation losses were not used to determine good fixation because of their poor reliability as
116 a fixation metric³³.

117 OCT imaging

118 Spectral Domain OCT (SD-OCT) imaging was performed with a Spectralis SD-OCT (Heidelberg
119 Engineering, Heidelberg, Germany). A circumpapillary Retinal Nerve Fibre Layer (cp-RNFL) scan and a
120 high-density macular cube (121 vertical B-scans, 30 x 25 degrees) were acquired. These scans were
121 inspected by an ophthalmologist (the author, GM) to confirm the absence of any abnormality in the
122 healthy cohorts and of any ocular disease other than glaucoma in the glaucoma cohort. Scans were
123 judged of sufficient quality if all the layers could be clearly identified in the central 15 degrees
124 around the fovea. No scans were removed because of poor quality.

125 Macular volumes were then exported in RAW binary format (.vol) using the Heidelberg Eye Explorer
126 platform and read into R (R Foundation for Statistical Computing, Vienna, Austria). This file
127 contained raw image files and segmentations of retinal layers, including the Inner limiting
128 membrane (ILM), Bruch's membrane (BM), the RNFL, and Ganglion Cell Layer (GCL). These
129 segmentations were checked for errors by an ophthalmologist (the author, GM) and corrected were
130 needed. Retinal thickness and GCL thickness maps were generated and processed as previously
131 described to obtain localised estimates of the number of RGCs underlying each stimulus area at all
132 locations in the 10-2 grid^{20,30,34}. Briefly, the fovea was automatically located via template matching
133 on the retinal thickness map. The GCL thickness map was transformed into a RGC density map with
134 histology data from Curcio and Allen³⁵ using a method proposed by Raza and Hood.³⁶ This method

135 accounts for eccentricity because the histology-derived volumetric density varies at different
136 positions on the retina. The area covered by the stimuli was displaced and distorted to account for
137 RGC displacement according to a revised version of the model proposed by Drasdo et al.^{30,34,37}
138 (**Figure 1**). Note that our method for displacement is different from the one used by a similar
139 previous study in the field,³² and produces different RGC counts especially in the parafoveal region.
140 However, our method was confirmed to be accurate.^{34,38} All calculations were performed in visual
141 degrees because we have previously shown that, under a spherical expansion model of the eye,
142 calculations of RGC density in visual degrees are unaffected by axial length³⁴. There is anatomical³⁴
143 and psychophysical³⁹ evidence to support a spherical expansion model, at least for moderate
144 refractive errors.



145
146 **Figure 1.** Test locations of the 10-2 grid distorted and displaced to cover the corresponding area on the
147 ganglion cell layer thickness map in a healthy eye (A) and an eye with glaucoma (B). This example is for a G-V
148 stimulus, for ease of visualization.

149 [Spatial summation model](#)

150 A previously described summation model²⁰ was used to generate a template to fit the sensitivity vs
151 stimulus area data. The summation model is described in more detail in the **Appendix**. In brief, the
152 model integrates the total retina input, which is the product of stimulus area, stimulus duration, RGC
153 density and Cone-to-RGC convergence ratio at a specific location. For this application, the stimulus
154 duration was fixed at 200ms. The model predicts a biphasic relationship between retinal input and
155 sensitivity, with a gradual transition from total to partial summation (**Figure 2**). The model accounts
156 for the Cone-to-RGC convergence ratio because we found, in previous experiments and
157 calculations,⁴⁰ that the spatial summation response profile (and Ricco's area) did not scale perfectly
158 with the number of RGCs at different eccentricities, but that the number of RGCs needed to be
159 weighted by the number of cones converging onto each RGC. Because different classes of RGCs tile
160 the retina with independent and partially overlapping mosaics, we only consider Parasol (or

161 magnocellular) OFF RGCs (P-OFF-RGCs) for our calculations^{41,42} because P-RGCs have been shown to
162 be preferentially stimulated by briefly flashed stimuli.^{43,44} However, for a given location, the effect of
163 stimulus area can be explained by a change in the number of RGCs being stimulated. This indicates a
164 scaling of recruited cortical filters with the amount of total retinal input, at least in healthy
165 observers. Note that we do not attribute any specific role to OFF-RGCs, although a preferential
166 involvement of this sub-class of RGCs has been suggested in glaucoma⁴⁵. This sub-class was simply
167 chosen to model a hexagonal mosaic of non-overlapping RGCs^{37,42} and because OFF-RGCs are the
168 most abundant in the human retina⁴⁶. Modelling ON-RGCs would have no material effect on our
169 results other than proportionally scaling the underlying RGC density in the model. Structural density
170 of P-OFF-RGCs were obtained as a proportion of the total structural RGC density estimates using the
171 equations provided by Drasdo et al.^{34,37}.

172 In the current study, we wanted to test the hypothesis that such a cortical filter scaling would also
173 occur with RGC damage in glaucoma. This can be done by testing whether the change in sensitivity
174 from RGC damage in glaucoma could be explained by a simple horizontal shift of a summation
175 template predicted by the model, similarly to what was reported by Redmond et al.¹⁶ This
176 corresponds to a change in Ricco's area (**Figure 2**). To test this hypothesis, we made two
177 assumptions:

- 178 1) RGC death and dysfunction would be indistinguishable, meaning that the model would not
179 be able to distinguish whether the reduced input is provided by a smaller number of fully
180 functional cells or a larger amount of dysfunctional cells⁴⁶.
- 181 2) The change in sensitivity would be predominantly a consequence of RGC loss and not of
182 photoreceptor damage, media opacity or other conditions.

183 An alternative hypothesis was to assume *no change in spatial scaling*. This corresponds to modelling
184 the change in sensitivity in glaucoma as a vertical shift in the summation template, i.e. change in
185 sensitivity without any change in Ricco's area. Note that the actual value of Ricco's area is not
186 reported as part of the results because it is not relevant for testing our hypothesis and because it is
187 not univocally defined for a summation curve with a smooth transition from total to partial
188 summation.

189 The model template was calibrated with data from the young healthy cohort and tested on
190 glaucoma patients and age matched controls.

191 Model calibration

192 The model has three parameters (see Formula in the **Appendix**): α determines the vertical offset of
193 the template (in \log_{10} scale); τ determines the transition from total to partial summation; κ
194 determines the slope of the partial summation portion of the curve (slope = $1/\kappa$). The model was
195 calibrated with RGC count estimates and perimetric sensitivity values from the healthy young
196 cohort. The RGC count estimates are more likely to be accurate in this group because of the low
197 likelihood of retinal damage and the close similarity in age with the retinae in the original histology
198 dataset by Curcio and Allen³⁵.

199 The parameters were estimated via numerical optimization (*fminsearch* function in Matlab R2018b,
200 The Mathworks, Natick, USA) and 95%-Confidence Intervals (CIs) for the parameters were computed
201 via bootstrap, resampling individual eyes rather than observations to preserve the correlation
202 structure of the data. The calibrated model was used to generate a template to fit the rest of the
203 data and test our hypothesis, as explained in the next section.

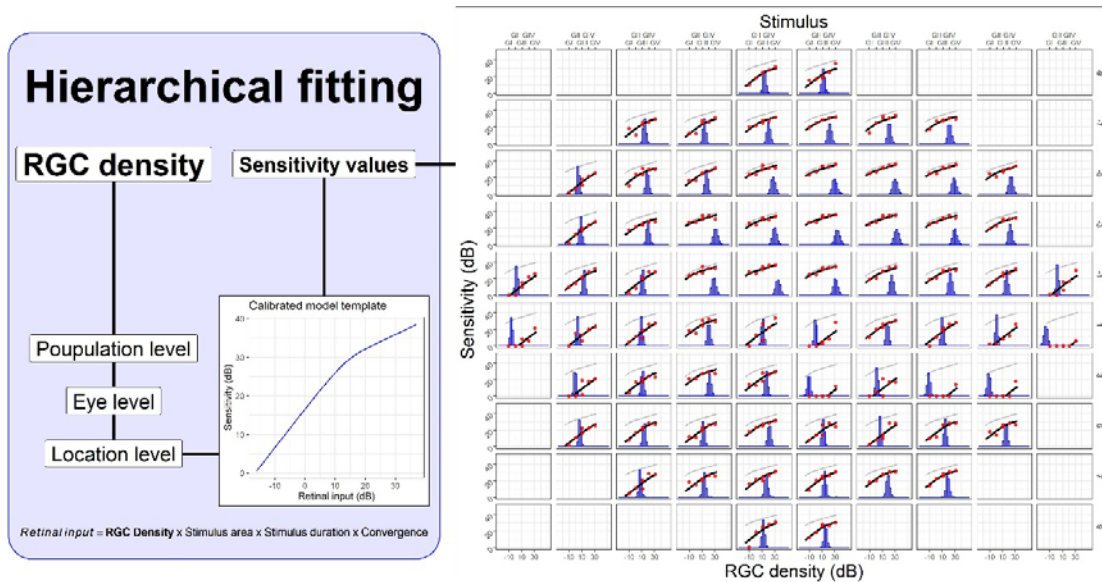
204 Template fitting to glaucoma patients and controls

205 Both the main and alternative hypothesis (*spatial scaling vs no spatial scaling* in glaucoma) can be
206 tested by fitting the summation template to the perimetric data with different assumptions. Fitting
207 the template presents significant challenges, especially because of the involvement of eyes with
208 advanced damage. The main technical issues are the presence of censored data, because the HFA is
209 not capable of presenting stimuli with luminance greater than $3,185 \text{ cd/m}^2$ (0 dB), and a consequent
210 lack of sensitivity values for more damaged locations. This can, on the one hand, bias the estimates.
211 On the other hand, it makes it difficult to obtain stable estimates for these locations when only few
212 sensitivity values are available at this level of damage. Bayesian computation and hierarchical
213 models can offer a solution because data censoring can be easily incorporated in complex models,
214 avoiding the bias from censored data (i.e. sensitivities $< 0 \text{ dB}$), and estimates at individual locations
215 can be made more robust by efficiently distributing information across different levels of the
216 hierarchy.

217 Details about the implementation of the Bayesian hierarchical model for this study are reported in
218 the **Appendix**. In brief, for the main hypothesis (*spatial scaling*), the model estimated the density of
219 RGCs at each location, in \log_{10} -scale, by optimising the horizontal shift of the template to fit the
220 observed sensitivity values for each stimulus area (**Figure 2**). The first level of the hierarchy was the
221 population level, modelling the average RGC count. This was then propagated at the eye level and
222 then at each location. The eye and location levels can be considered nested Gaussian random
223 effects. Because of the hierarchical structure, all the data were fitted concomitantly and the
224 estimate at each location was also informed by the data at other locations within the same eye and
225 by the general behaviour of the population. The template was used as a link function to model the
226 expected sensitivity at each stimulus area given the modelled RGC density estimate. The response
227 variable was the sensitivity, which was censored at 0 dB. Note that using a link function for the
228 expected sensitivity is different from modelling an inverse transformation of the data. The fitting
229 process also modelled a vertical shift of the template at the population level, to optimise the
230 average centration of the template. The alternative hypothesis (*no change in spatial scaling*) was
231 implemented with a similar model. In this case, the hierarchical parameter was the vertical shift of
232 the template and the horizontal shift (Ricco's area) was only modelled at the population level. This
233 fitting process assumes no change in spatial scaling across subjects, while the change in sensitivity is
234 only modelled through the vertical shift of the template.

235 Note that it is not possible to model a vertical and a horizontal shift of the template simultaneously,
236 because the solution would be undefined in locations for which the tested stimulus area sizes do not
237 encompass Ricco's area. For example, a location for which all tested stimulus sizes are smaller than
238 Ricco's area can be fitted by arbitrary combinations of vertical and horizontal shifts of the template.
239 Therefore, we used the alternative hypothesis of no spatial scaling as a comparator to assess the
240 significance of our results under the main hypothesis (see next section). Normally, statistical
241 significance can be assessed by quantifying the uncertainty around parameters' estimates. However,
242 because each version of the model is forced to fit the data with either a horizontal or a vertical shift
243 of the template, the parameter estimate associated with the modelled shift is likely to be
244 significantly different from zero (no shift) in both cases and cannot be used to accept or reject the
245 tested hypothesis.

246



247

248 **Figure 2.** Schematic illustrating the hierarchical fitting process for the template. The template shown on the
 249 left is shifted horizontally to match the data. The example on the right shows the result of the fit. The top
 250 horizontal axis reports the stimulus size. The bottom horizontal axis refers to the histograms, which represent
 251 the estimated Retinal Ganglion Cell (RGC) density (in dB) for each location. The histograms show all the
 252 iterations of the Bayesian fitting procedure. The red dots are the measured sensitivity, the black lines are the
 253 shifted templates (the original “healthy” template is reported in light gray).

254 Data analysis

255 All data, including those from the young healthy cohort, were used in the fitting, but only data from
 256 the glaucoma patients and age-similar healthy controls were used to calculate goodness of fit
 257 statistics. The R^2 was calculated for the sensitivity predicted with the template fitted at each location
 258 and expressed as the percentage of variance explained. Confidence intervals for the R^2 were
 259 calculated via bootstrap (1000 samples) using the subject as the resampling unit. The Root Mean
 260 Squared Error (RMSE) was also calculated, for comparison with the structural predictions (see
 261 below).

262 The structure-function analysis was performed in a similar fashion, using the point-wise structural
 263 RGC density, calculated as described above, with estimates of GCL thickness from the SD-OCT scans
 264 (calculated as the average density from the five different stimulus sizes). However, because there
 265 was no fitting involved in the structure-function predictions, only the RMSE was calculated. Both
 266 RGC density estimates were expressed in dB ($10 \cdot \log_{10}(\text{Density})$). We also calculated the dynamic
 267 range for the structural and functional density estimates as the width of the 2.5% - 97.5% interval, to
 268 report the structural floor effect. All the analyses were performed in R.

269 When referring to estimates of the total retinal input, we will use the term *functional retinal input* to
 270 refer to the total retinal input calculated with local RGC density values estimated by fitting the
 271 functional data. The *structural retinal input* was instead calculated using structurally derived local
 272 RGC density values.

273 Results

274 Study population

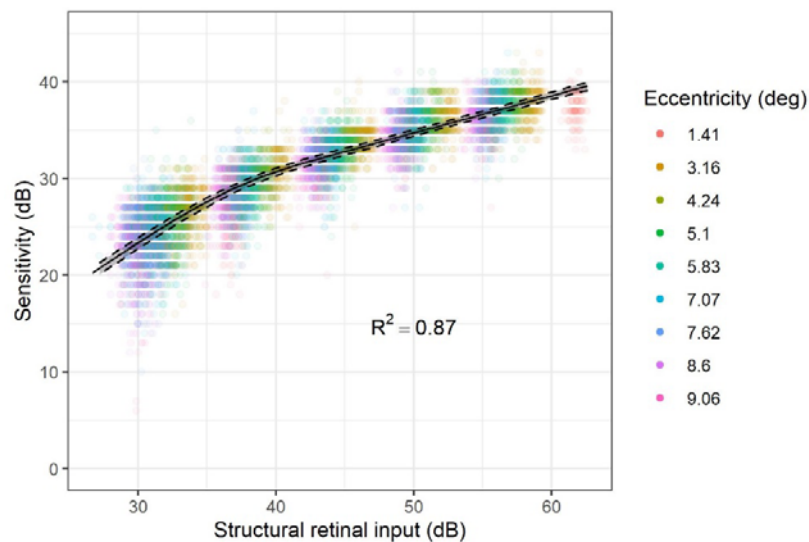
275 Descriptive statistics for the sample are reported in **Table 1**. One individual in the healthy cohort was
 276 excluded because they completed only two of the six tests. None of the tests was unreliable.

	Healthy < 40 years old (N = 29)	Age matched controls (N = 20)	Glaucoma		
			Early (N = 10)	Moderate (N = 10)	Advanced (N = 10)
Age (age)	28 (3)	62 (11)	66 (9)	59 (10)	62 (11)
AL (mm)	24.40 (1.05)	24.00 (0.94)	23.56 (0.65)	24.75 (1.35)	23.71 (1.18)
24-2 MD (dB)	-0.67 (0.91)	0.16 (1.36)	-2.26 (1.56)	-8.21 (2.13)	-18.51 (5.78)
24-2 PSD (dB)	1.45 (0.37)	1.91 (0.58)	3.24 (1.60)	11.10 (2.35)	11.61 (1.99)
cpRNFL (µm)	96.8 (9.2)	93.8 (9.5)	72.0 (10.4)	61.3 (15.4)	47.1 (6.9)
WRT (µm)	311.1 (13.8)	303.1 (13.9)	290.5 (17.7)	280.8 (16.2)	275.5 (8.7)
GCL (µm)	39.6 (3.10)	37.1 (3.2)	31.8 (4.9)	26.8 (5.4)	23.2 (3.8)
RGCs (dB)	5.58 (0.03)	5.54 (0.04)	5.47 (0.08)	5.39 (0.10)	5.32 (0.08)

277 **Table 1.** Descriptive statistics of the sample reported as Mean (Standard deviation). AL = Axial Length; MD =
 278 Mean Deviation; PSD = Pattern Standard Deviation; cpRNFL = circumpapillary Retinal Nerve Fibre Layer; WRT =
 279 Whole Retinal Thickness; GCL = Ganglion Cell Layer; RGCs = Retinal Ganglion Cell count (in 10^*log_{10} scale). The
 280 structural metrics are total or average values calculated within the central 10 degrees from the fovea.

281 Model calibration

282 The parameter estimates for the model fitted in the young healthy cohort were (Mean [95% - CIs]): α
 283 = 1.42 [1.29, 1.57]; $log_{10}(\tau) = 3.58$ [3.44, 3.70]; $\kappa = 2.59$ [2.45, 2.78] (corresponding to a partial
 284 summation slope of 0.39 [0.36, 0.40]). The slope was notably different from the commonly chosen
 285 0.25 ($p < 0.001$)^{19,21} but not dissimilar to the 0.369 reported by Antwi-Boasiako et al. ($p = 0.146$)¹⁷.
 286 The result of the fitting is shown in **Figure 3**.



287

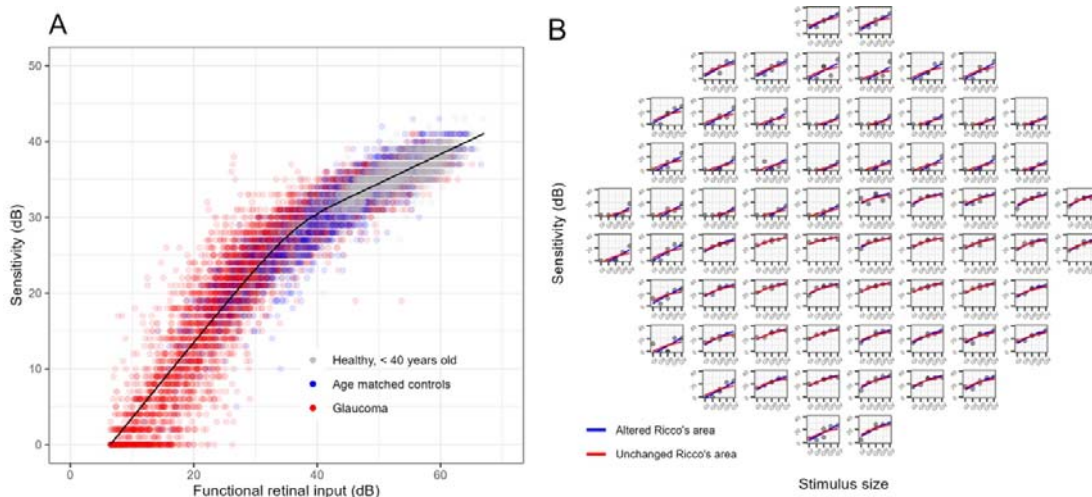
288 **Figure 3.** Results of the calibration procedure of the template on the data from the young healthy cohort. The
 289 Dashed lines represent the 2.5%-97.5% confidence bands for the template estimated via bootstrap. The data
 290 are clustered due the different stimulus diameters used.

291 **Template fitting**

292 The horizontal shift of the template (which assumes a change in Ricco’s area from RGC damage)
 293 explained 95.2% [95%-CIs: 94%, 96.2%] of the overall variance in the data, a significant improvement
 294 over assuming no change in Ricco’s area ($p < 0.001$). **Table 2** reports the R^2 and RMSE values for the
 295 healthy subjects and the glaucoma patients at different stages of damage. **Figure 4** shows the fitting
 296 results. **Supplementary Figure 1** shows the same results for each location (horizontal shift). The
 297 average error per subject for the horizontal shift of the template was not significantly affected by
 298 age (linear regression, $p = 0.819$), indicating that modelling a change in Ricco’s area was able to
 299 account for most of the effect of ageing. The differences in accuracy between the two alternative
 300 models were more evident in the glaucoma cohort with intermediate damage, where a transition
 301 from partial to complete summation would be more evident if RGC damage was indeed causing a
 302 change in Ricco’s area. **Supplementary Figure 2** shows the fitting error, stratified by sensitivity, of
 303 the two alternative models compared to the test-retest noise. Fitting the template with a horizontal
 304 shift produced the closest error to the test-retest noise, consistently below that obtained with a
 305 vertical shift.

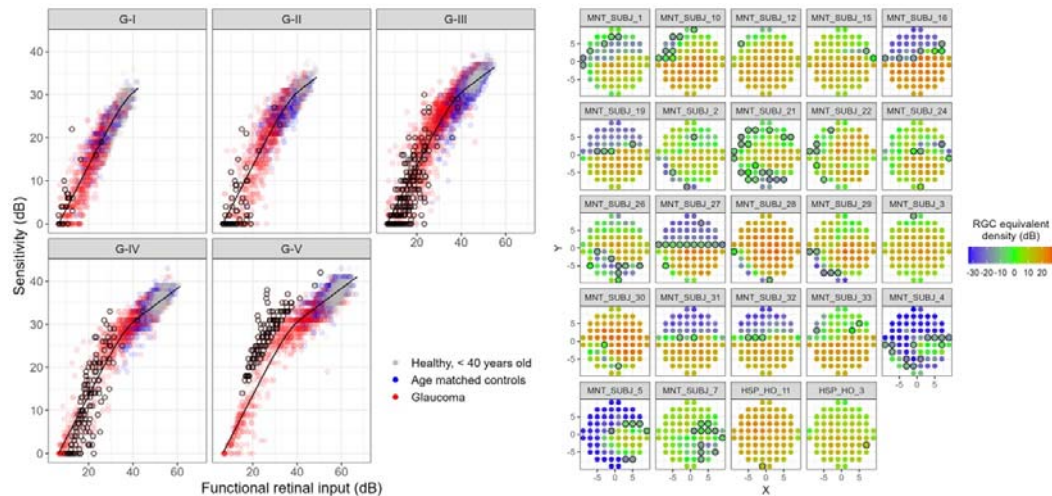
Group	Estimate [95%-CIs]					
	Altered Ricco’s area		Unchanged Ricco’s area		Improvement (%)	
	R^2 (%)	RMSE (dB)	R^2 (%)	RMSE (dB)	R^2	RMSE
All	95.2 [93.9-96.1]	2.09 [1.92-2.26]	93.2 [91.5-94.5]	2.49 [2.24-2.72]	2.1 [1.6-2.7]	15.9 [12.6-18.3]
Healthy	91.3 [90.4-92.1]	1.56 [1.44-1.71]	89.8 [88.8-90.8]	1.69 [1.58-1.83]	1.7 [0.8-2.5]	7.7 [4.0-11.5]
Glaucoma						
Early	91.6 [89.5-93.1]	2.21 [1.74-2.64]	88.4 [86.6-90.0]	2.59 [1.99-3.10]	3.4 [2.4-4.2]	14.5 [9.70-18.7]
Moderate	93.2 [90.9-95.3]	2.96 [2.50-3.39]	89.6 [85.4-93.1]	3.66 [2.98-4.29]	3.9 [2.1-6.2]	19.2 [14.3-22.5]
Advanced	95.3 [93.7-96.3]	2.99 [2.70-3.29]	92.3 [89.1-94.3]	3.83 [3.33-4.32]	3.1 [1.9-5.0]	21.8 [17.1-25.2]

306 **Table 2.** R^2 and Root Mean Squared Error (RMSE) statistics for the hierarchical fitting of the template. The 95%
 307 Confidence Intervals were estimated via bootstrap. These statistics exclude the data from the young healthy
 308 cohort used for calibration. Improvement was calculated as percent increase in R^2 and percent reduction in
 309 RMSE fitting a horizontal shift of the template over fitting a vertical shift. All improvements were significant (p
 310 < 0.001).



311
 312 **Figure 4. A)** Results of the template fitting via horizontal shift on the overall sample. For this graph, the
 313 observations from each location were shifted horizontally according to their estimated parasol OFF Retinal
 314 Ganglion Cell (RGC) density. **B)** Example (one eye with glaucoma) comparing the fit obtained via horizontal
 315 (altered Ricco’s area) and vertical (unchanged Ricco’s area) shift of the template.

316 When broken down into different stimulus sizes, some locations appeared to have their sensitivity
 317 underestimated by the model for the largest stimuli. We identified these locations as those that
 318 were greater than 97.5% of the prediction error (4.9 dB) above the prediction with the G-V stimulus
 319 (**Figure 5**). The sensitivity for these locations also appeared to increase more steeply than predicted
 320 by complete summation for smaller stimulus sizes^{47,48}. We hypothesized that this could be a
 321 consequence of testing at the edge of scotomas. When plotted in the 10-2 grid, these locations were
 322 in fact mostly located in regions of sharp change in the modelled RGC density estimates (**Figure 5**).
 323 We further tested this hypothesis by simulating the response from an RGC mosaic with a sharp
 324 change in cell density and we were able to reproduce the same behaviour (**Supplementary Figure 3**).



325
 326 **Figure 5.** Fitting results split by stimulus size (left panels). The observations circled in black are those that
 327 exceeded the 97.5% limit of the prediction error for the G-V stimulus. The same locations are reported on the
 328 map on the right, representing the modelled RGC density.

329 Structure-function relationship

330 The structural and functional estimates of RGC density are plotted in **Figure 6**. The overall
 331 agreement was poor (**Table 3**), mostly due to the limited dynamic range of the structural estimates,
 332 which was, on average, only 11% ($\pm 2\%$) of the functional estimates.

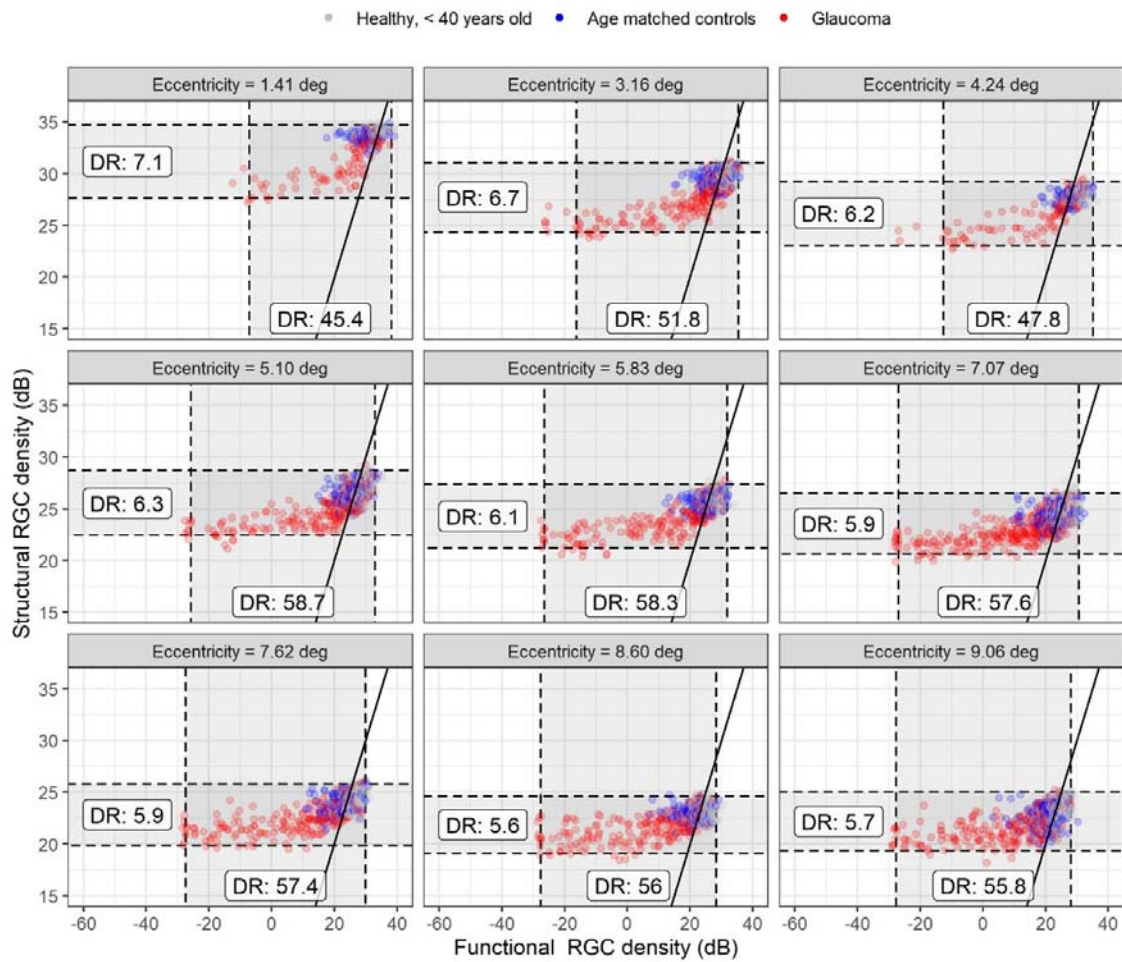
333 Using the template to predict the sensitivity from the structural RGC estimates generally provided
 334 poor prediction accuracy (**Table 3**). These predictions are reported in **Figure 7**. These predictions
 335 were improved, as expected, by only analysing locations where sensitivity with a G-I stimulus was
 336 greater than 10 dB. This latter sub-analysis was performed for comparison with the work of Antwi-
 337 Boasiako et al.¹⁷

Structural RMSE (dB) [95%-CIs]			
Group	Sensitivity, all locations	Sensitivity, locations ≥ 10 dB	Functional RGC Density
All	10.6 [8.4-12.5]	3.5 [2.9-3.7]	14.3 [11-17.6]
Healthy	3.0 [2.1-3.9]	3.0 [2.2-3.7]	4.0 [2.5-5.3]
Early	5.9 [3.7-7.5]	3.1 [2.4-3.8]	7.2 [4.5-9.3]
Glaucoma	11.8 [9.2-14.1]	4.2 [3.3-4.6]	15.2 [11.6-18.5]
Advanced	18.8 [15.7-21.8]	4.8 [3.3-4.9]	26.5 [20.7-31.8]

338 **Table 3.** Root Mean Squared Error (RMSE) for structure-function predictions. For sensitivity, structural
 339 predictions were generated using the spatial summation template with structural estimates of the parasol OFF
 340 Retinal Ganglion Cell (RGC) number as an input. For the RGC density estimates, we report the RMSE of

341 structural estimates of local parasol OFF RGC density predicting the corresponding functional estimates from
342 the fitting of the template.

343

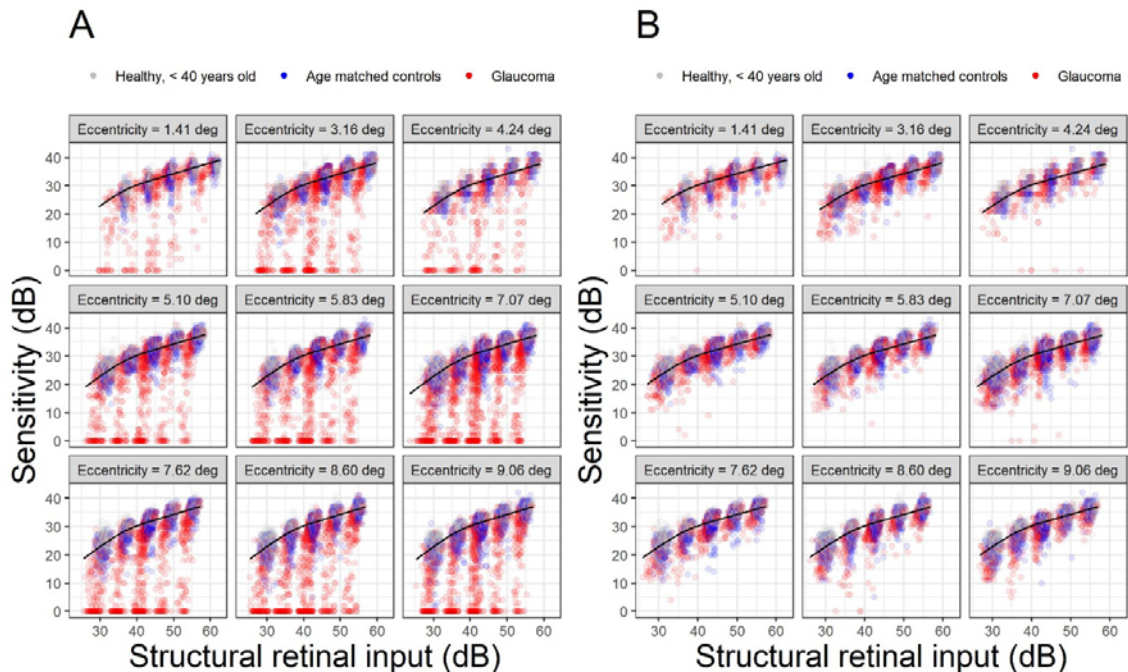


344

345 **Figure 6.** Structural and functional estimates of the Parasol OFF Retinal Ganglion Cell (RGC) density at each
346 location. The solid line indicates the identity. The dashed line represents the dynamic range (DR) of the
347 structural and functional estimates.

348

349



350

351 **Figure 7.** Structure-function predictions based on the template for the whole sample (A) and for locations
 352 where sensitivity was > 10 dB with a G-I (B). The structural retinal input was calculated identically to the
 353 functional retinal input, but using structural estimates of local parasol OFF retinal ganglion cell density instead
 354 of the functional ones, derived from fitting the template (as in Figure 4 and 5). This is identical to the retinal
 355 input calculated for the young healthy cohort for calibration (Figure 3), which was also derived from structure.

356 Discussion

357 We evaluated the hypothesis that changes in perimetric sensitivity from modelled RGC damage or
 358 loss in the glaucomatous macula could be explained by a change in the spatial scaling of the
 359 response of visual system. We tested this by fitting experimental perimetric data in human observers
 360 (patients with glaucoma and healthy controls) with a template that models a change in Ricco's area,
 361 and showed that a horizontal shift of the template, modelling an enlargement of Ricco's area, could
 362 explain 95% of the overall variance in the data. This explained the data significantly better than a
 363 vertical shift of the template, which would model a change in sensitivity without a change in Ricco's
 364 area. We then showed that the local functional loss was not entirely captured by structural
 365 measurements from SD-OCT.

366 Our findings support the hypothesis that RGC damage from glaucoma produces a perimetric
 367 functional loss that can be explained by an enlargement of Ricco's area¹⁶. This was speculated to be
 368 a consequence of the loss of RGCs, leading to the hypothesis that Ricco's area would scale with RGC
 369 density, to include a constant number of RGCs. In general, this hypothesis has been shown to hold
 370 true in healthy eyes when tested at different eccentricities⁸⁻¹⁴ and in glaucoma patients when tested
 371 with computational models similar to the one used in this work²¹. However, Swanson et al.²¹ showed
 372 that the extent of Ricco's area depends on the spatial scale of the cortical filters, regardless of the
 373 underlying density of RGCs. In fact, previous work has shown that the extent of Ricco's area (and
 374 thus the number of RGCs underlying a Ricco's area scaled stimulus¹⁵) at any given location can be
 375 altered, in healthy observers, by stimulation conditions, such as background luminance⁴⁹⁻⁵¹, duration
 376 of the stimulus^{20,50,52,53} or by high frequency background noise¹⁹. This makes it clear that VF

377 sensitivity cannot be explained solely by RGC density and likely also involves further processing at a
378 cortical level.

379 Redmond et al.¹⁶ provided experimental evidence of such a change in the spatial scaling occurring in
380 patients with glaucoma. However, the same phenomenon has not been extensively investigated in
381 advanced glaucomatous damage and in the macular region. While there is no specific reason to
382 expect spatial summation to behave differently in the macula, its impact would be the greatest in
383 this region for standard perimetry with a G-III stimulus^{8-10,21,30}. This is because the high initial RGC
384 density in the healthy macula would determine a transition between partial and total summation, as
385 RGCs are lost in glaucoma. Moreover, the macula allows direct individualised point-wise structural
386 OCT measurements, which are not usually available for the more peripheral retina. One study by
387 Yoshioka et al.³² investigated the effect of spatial summation on the association between perimetric
388 sensitivity and retinal structure in the macula of eyes with early glaucomatous damage and showed
389 that it is improved with smaller stimulus-sizes. This is compatible with our findings, since smaller
390 stimulus sizes would operate under complete spatial summation in both healthy and glaucomatous
391 eyes, making the slope of the relationship between the number of RGCs and sensitivity steeper. One
392 important difference was the method used to displace the stimuli to account for RGC displacement,
393 which, in the case of Yoshioka et al.³², was later shown to yield less accurate results, especially in the
394 parafoveal region³⁴. This was then also confirmed by the same group in later work⁵⁴. More recently,
395 a detailed analysis has been presented by Antwi-Boasiako et al.¹⁷, who studied the relationship
396 between macular RGC counts and perimetric sensitivity in non-human primates with experimental
397 glaucoma. Antwi-Boasiako et al.¹⁷ also analysed their data within the framework of spatial
398 summation. Some of their results were confirmed in our study. Importantly, the partial summation
399 slope estimated by our data (0.39, corresponding to an exponent κ of 2.59) was very close to their
400 estimate (0.369). This is noteworthy, because there is still uncertainty about the most accurate
401 choice of slope to describe partial summation for perimetric stimuli in studies of this kind. In
402 computational models of sensitivity, this mainly depends on the choice of the spatial filter and of the
403 Minkowski summation exponent κ ¹⁹. Common choices for the exponent are between 2 and 4. For
404 most symmetric filter choices (except some Gaussian derivatives used to model cortical responses),
405 these values correspond to a partial summation slope of 0.5 (Piper's law) and 0.25. An exponent of 4
406 was used in a previous implementation of our model²⁰ and by others^{19,21}. However, an intermediate
407 value for the exponent seems more reasonable given the experimental results from this work and
408 Antwi-Boasiako et al.¹⁷.

409 Differently from Antwi-Boasiako et al.¹⁷, we found that structural measurements were not able to
410 fully characterize functional damage, owing to their reduced dynamic range (**Figure 6**). One factor
411 that could explain this discrepancy is that Antwi-Boasiako et al.¹⁷ had access to histology-derived
412 RGC counts in both healthy and glaucomatous eyes to calibrate their structural models, which would
413 naturally improve accuracy. In contrast, we only relied on a limited histology data in healthy human
414 subjects provided by Curcio and Allen³⁵. Additionally, it is unclear from their paper whether Antwi-
415 Boasiako et al.¹⁷ accounted for RGC displacement by simply moving the center of the 10-2 stimuli, as
416 in Yoshioka et al.³², or whether they applied the displacement to the edge of the stimulus (**Figure 1**).
417 This is relevant because, despite yielding correct RGC counts in healthy eyes and in early damage,
418 our method of displacement, by its nature, amplifies the floor-effect, since non-functional residual
419 tissue is summed over a larger area, especially in the parafovea. Finally, the level of damage in
420 Antwi-Boasiako et al. was in general less advanced than in our dataset, with the lowest sensitivity
421 values being approximately 10 dB. Indeed, restricting our analysis to locations with a sensitivity > 10
422 dB with a G-I stimulus resulted in a great improvement in the RMSE for structure-function estimates
423 (**Table 3** and **Figure 7**). Nevertheless, our results find ample confirmation in previous literature^{36,55,56}

424 documenting a structural floor-effect at around 10 dB of sensitivity loss in the macula and
425 confirming that structurally derived estimates offer only a partial description of RGC loss and
426 damage occurring in glaucoma. All these aspects, including the increased level of perimetric noise at
427 more advanced damage, contributed to the poor RMSE in the structure-function predictions
428 reported in **Table 3**.

429 Our findings have important implications for the interpretation of macular perimetric damage in
430 glaucoma. The first important aspect is that it confirms a change in the spatial scale of the response
431 following RGC loss or damage, which corresponds to an enlargement of Ricco's area. As previously
432 stated, the exact value of Ricco's area is irrelevant for testing our hypothesis and is not univocally
433 defined for curves with a smooth transition from total to partial summation. However, Ricco's area is
434 a useful concept to describe changes in spatial scaling, and here it is used as synonymous of *spatial*
435 *scale*. One thing that should be noted is that previous work mostly focussed on the relationship
436 between the number of RGC receptive fields covered by the stimulus and perimetric response.
437 According to this view, the response of the visual system would scale to include a constant number
438 of RGCs at Ricco's area^{16,17}. Our interpretation differs slightly, because the total retinal input in our
439 summation model would not differentiate between reduced input from RGC loss or dysfunction.
440 Differentiating between these two contributions would require additional investigations. Adaptive
441 optics OCT imaging has shown promising results allowing direct visualization of RGCs in healthy
442 subjects⁵⁷ and glaucoma patients⁵⁸ and could be used to more precisely quantify the density of RGCs.
443 Functional tests, such as high contrast grating stimuli, could be used for the same scope^{46,59-62}.

444 The varying relationship between RGC damage and functional loss is especially important in the
445 macular region, because sensitivity to the widely used G-III, 200 ms stimulus would initially be
446 determined by partial summation, making the relationship with retinal structure shallow. As RGCs
447 are lost or damaged, the response would gradually transition into complete summation, where the
448 relationship between sensitivity and retinal structure becomes steeper. This implies that, for the
449 same percentage of RGC loss, changes in sensitivity would be much smaller early in the disease
450 compared to more advanced damage. This might make the detection of early damage, and similarly
451 early progression, more challenging^{6,63}. Other strategies employing smaller targets or shorter
452 durations for macular stimuli might make perimetric tests more efficient by testing always under
453 complete summation conditions, although this might limit the dynamic range of the test. Some of
454 these strategies have already been adopted in some home monitoring devices⁶⁴. Another approach
455 would be to modulate the area or duration of the target instead of the luminance. This approach
456 would take full advantage of the horizontal translation of the response profile observed in our data
457 and in previous publications^{16,63}, effectively testing the response at a fixed point of the summation
458 curve. Such an approach has been shown to maximise signal-to-noise ratio in glaucoma and to
459 reduce response variability compared to luminance modulation⁶³.

460 It should be noted that, while fitting a template and testing the spatial-scaling hypothesis did not
461 require a link to RGC density, modelling the retinal input and the effect of RGC loss provides a
462 linkage to an underlying biological substrate, offering a generalisable framework for interpreting the
463 results. For example, using a computational model of an RGC mosaic allowed us to provide a
464 possible explanation for the edge effect for larger perimetric stimuli observed in the data (see
465 **supplementary material**). Moreover, modelling changes in retinal input rather than simple
466 translations of 'healthy' summation functions for each tested location highlighted how changes in
467 spatial summation both across the healthy VF and as a consequence of damage can arise in the
468 context of different modifications to the same underlying biological substrate. It should finally be
469 highlighted that, because of how the spatial summation template was calculated (i.e. using

470 sensitivity values and estimated RGC counts in healthy subjects), the intrinsic linkage to the
471 underlying retinal input is present in our calculations, regardless of whether it is made explicit or not
472 in our interpretation of the results.

473 A better characterization of the relationship between RGC damage and perimetric sensitivity is also
474 useful to improve the correspondence between perimetric changes and structural damage observed
475 with imaging. As shown in this and previous work^{32,36}, both measurements can be reported in a log-
476 scale of RGC number. This could facilitate structure-function analyses for progression or enable
477 seamless integration of structurally derived metrics into perimetric strategies⁶⁵. One limitation,
478 however, is that structural metrics do not seem to have enough dynamic range, at least locally, to
479 capture the full extent of functional damage measured by perimetry. Although such a discrepancy
480 has been reduced by nonlinear estimates, such as with help of artificial intelligence⁶⁶⁻⁶⁸, structural
481 tests are unlikely to replace perimetry. An efficient integration of the two sources of information
482 seems, therefore, the most effective way of diagnosing and monitoring glaucoma.

483 A limitation of this work is that it was not possible to derive sensitivity estimates for all stimulus
484 areas at all tested locations, especially among patients with intermediate or advanced glaucoma.
485 This was expected given the technical limitations of the device (limited stimulus areas and fixed
486 duration), and addressed with the use of a hierarchical model, which allowed for more robust
487 estimates of RGC damage for locations where only limited data could be collected, and by
488 accounting for censoring at 0 dB. However, the estimates for these locations are necessarily less
489 precise and mostly reliant on the behaviour of the other locations within the same eye and on the
490 general trend of the overall population. For the same reason, it was not possible for us to model the
491 horizontal and vertical shift at the same time, because the fitting results would only be fully
492 constrained for locations that span both partial and complete summation with the available stimulus
493 diameters. For example, for locations exhibiting complete summation exclusively, the same fitting
494 result can be achieved by either a vertical or a horizontal translation of the template. However, this
495 would not affect the ability to compare our two alternative hypotheses. It is also important to note
496 that previous work, especially by Gardiner et al.^{69,70}, has shown poor correlation between accurate
497 sensitivity estimates derived from frequency of seen curves and clinical perimetry, especially for
498 values < 20 dB. In our analysis, however, we assumed that low sensitivities would still provide useful
499 information to test population-level hypotheses, especially in eyes with advanced glaucoma. We
500 provide, as **supplementary**, additional analyses supporting this assumption. Importantly, we show
501 that including sensitivity values ≤ 15 dB reduced the prediction error for the fitted model for
502 sensitivity values > 15 dB. This indicates that, in our data, locations with advanced damage improved
503 the precision of the model.

504 In our study, we could not control for the effect of optics on macular sensitivity. This could have
505 been influenced by age-related changes to refractive media. We controlled for this limitation by
506 comparing glaucoma with age similar controls. The effect of optics^{71,72} and ageing⁷³ on spatial
507 summation is still unclear. Redmond et al.⁷³ did not find any change in the critical area with age.
508 However, from our data, there does not seem to be any significant residual effect of ageing on
509 explaining the change in sensitivity once the change in spatial summation is accounted for. However,
510 our data does not allow us to test this hypothesis specifically and further, more targeted
511 investigations, are needed.

512 Appendix

513 Computational model

514 The model, as previously explained²⁰, predicts sensitivity as a function of the total retinal input,
515 which is the product of the number of RGC receptive fields that underlie the stimulus, the duration
516 of the stimulus presentation, and the cone-to-RGC convergence ratio at different eccentricities. This
517 was derived by combining Curcio and Allen's data^{35,74} and the RGC receptive field (RGC-RF) density
518 obtained from the equations provided by Drasdo et al.^{34,37}. In our previous analysis of spatial
519 summation data in healthy subjects²⁰, we showed that this weighted RGC-RF number, rather than
520 the raw count of RGC-RFs covered by the stimulus, were able to equate the spatial summation
521 curves at different eccentricities. The model uses a capacitor equation and continuous integration
522 over the input. A Minkowski exponent is used in the integration, similar to the vector summation
523 equation used by Pan and Swanson¹⁹. The model has three parameters that can be fitted: α
524 determines the vertical offset of the template (in \log_{10} scale); τ determines the transition from total
525 to partial summation; κ determines the slope of the partial summation portion of the curve (slope =
526 $1/\kappa$). The formula from Montesano et al.²⁰, with small modifications, is reported below:

$$R = 10^\alpha \left(\int_0^T M^\kappa * d(st) \right)^{1/\kappa}$$

527 Where R is the sensitivity in linear units ($10^{\text{dB}/10}$), M is the total retinal input filtered (convolved) with
528 a capacitor equation in the form

$$M = \exp(-st/\tau) \times S$$

529 Where S is a step function of the retinal input and is equal to 1 over a segment of st (an arbitrary
530 unit of spatio-temporal input) that indicates the extent of the total retinal input of the stimulus, i.e.
531 it becomes longer when more RGCs are stimulated or the same RGCs are stimulated for a longer
532 period of time. The symbol \times indicates the convolution operation.

533 Bayesian fitting

534 The fitting sought to find the optimal value of RGC density for each location that would give the best
535 fit for the template. Changing RGC density corresponds to a horizontal shift of template. RGC density
536 at each location was modelled as a hierarchical random effect, nested within another random effect
537 grouping locations from the same eye. A single global parameter also allowed a vertical offset of the
538 template to achieve the optimal fit in the overall sample. This offset was however very small (-0.23
539 dB). The same procedure was adopted to fit vertical shifts of the template at each location (i.e. no
540 change in Ricco's area), while a global parameter optimized the location of Ricco's area in the whole
541 sample (this offset was also small, -0.05 \log_{10} -units). Note that the template was not allowed to
542 move both horizontally and vertically at each eye/location because this would make the fitting
543 undetermined for all locations where sensitivity values showed no change in slope in the data,
544 because the same fit could be obtained by infinite combinations of vertical and horizontal shifts.

545 VF sensitivity was assumed to have a Normal distribution of the residuals, censored at 0 dB. Fitting
546 of the Bayesian model was achieved using JAGS (Just Another Gibbs Sampler⁷⁵) to run Markov Chain
547 Monte Carlo (MCMC) simulations, within the R environment (R Foundation for Statistical
548 Computing). Two parallel MCMCs were run for at least 5,000 iterations after 1,000 adaptation steps
549 and 5,000 burn-in iterations. The MCMCs were stopped if the Gelman-Rubin diagnostic was < 1.2 for
550 all the monitored parameters, indicating convergence⁷⁶. Prior distributions on the fixed effects were
551 non-informative Normal distributions with a precision of 0.01 (Variance = 100).

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