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*CORRESPONDENCE Marcos Adriano Garcia Campos gyleanes@alumni.usp.br

[†]These authors have contributed equally to this work and share first authorship

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Malignant lung PEComa (clear cell tumor): rare case report and literature review

Marcos Adriano Garcia Campos^{1*†}, Lucas Fernandes Vasques^{2†}, Rafael Goulart de Medeiros^{2†}, Érico Murilo Monteiro Cutrim¹, Ana Júlia Favarin¹, Sarah Rebecca Machado Silva¹, Gyl Eanes Barros Silva³, Marcelo Padovani de Toledo Moraes¹, Mariana Lopes Zanatta¹ and Diego Aparecido Rios Queiróz¹

¹Department of Internal Medicine, Hospital of Medical School of São Paulo State University, Botucatu, Brazil, ²Department of Medicine, University of Taubate Faculty of Medicine, Taubaté, Brazil, ³Laboratory of Immunofluorescence and Electron Microscopy, University Hospital of Federal University of Maranhão, São Luís, Brazil

Clear cell tumors of the lung (CCTL), or "sugar tumors" of lung, are very uncommon lesions and are mostly benign perivascular epithelioid cell (PEC) tumors with no specific morphologic features. Fewer than 100 cases have been reported; the aggressive nature demonstrated in sporadic reports has rarely been described in the literature. Although the course is generally described as benign, eight reported cases showed malignant behavior. We report a case of a PEC with a malignant presentation in a young man, correlating the main characteristics of the tumor with other cases reported in the literature to better elucidate this rare presentation. We also performed a literature review of reports on benign and malignant CCTL cases, with a focus on clinical, imaging, and immunohistochemical differentiation. CCTLs are rare tumors that require histopathological and immunohistochemical confirmation; to date, criteria that can predict malignant evolution are lacking.

KEYWORDS

PEComa, sugar tumor, pulmonary cancer, cathepsin K, HMB-45

Introduction

Clear cell tumors, or "sugar tumors", are very uncommon lesions (1) and are mostly benign perivascular epithelioid cell (PEC) tumors (2, 3). PEC tumors are a rare group of mesenchymal neoplasms histologically and immunohistochemically distinct from other neoplasms, with an abundance of periodic acid-Schiff (PAS)-positive glycogen (3). They are expressed in the bladder (4), uterus (5), breast (6), ovary (7), vagina (8), kidney (9), prostate (10), pancreas (11), jejunum (12), soft tissues (13) and lung (14). Due to this wide diversity, these tumors lack specific morphologic features (2).

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The lungs are an uncommon site for PEC tumors, known as clear cell tumors of lung (CCTL) (15), and such PEC tumors in the lung are usually benign, with malignancy exceedingly rare (9). Although fewer than 100 cases have been reported, sporadic reports have described CCTLs with very rare aggressive nature (3). A silent, indolent, and asymptomatic presentation may pose a diagnostic challenge for pathologists and oncologists (16).

CCTLs exhibit nested histological features similar to other PEC tumors. They consist of uniform round to polygonal epithelioid cells with clear or eosinophilic cytoplasm and well-defined borders. CCTLs are surrounded by prominent thin-walled vascular channels. Although the course of CCTL is typically described as benign, eight cases have shown malignant behavior (17). We present a case of CCTL with a malignant presentation, correlating the main characteristics of the tumor with other cases reported in the literature to better elucidate this rare presentation.

Case description

A 49-year-old man, hypertensive and a smoker, presented to the primary healthcare department with complaints of dyspnea on exertion and daily chest pain for the past year, with gradual worsening in the last three months, in addition to a weight loss of 10 kg. The patient denied coughing, hemoptysis, fever, or a family history of cancer.

Computed tomography (CT) of the chest showed a solid, calcified, upper left paratracheal lesion (191 cm³) with central necrosis in close contact with the ipsilateral subclavian artery without a cleavage plane, in addition to multiple randomly distributed nodules in the lung parenchyma and enlarged lymph nodes in the para-aortic chain and thin left pleural effusion



FIGURE 1

Initial coronal (A) and axial (B) computed tomography pulmonary angiogram reconstruction of the chest performed showing a mass in upper left paratracheal side (white arrow in A, B) associated with multiple randomly distributed nodules in the lung parenchyma (black arrow in C, D), the largest of them in the middle third of the lung right, enlarged lymph nodes in the para-aortic chain and thin left pleural effusion (black asterisk in C, D). (Figure 1). A lithic image of the T1 vertebral body indicated possible secondary neoplastic involvement; however, bone scintigraphy and positron emission tomography-CT (PET-CT) revealed a low probability of bone metastasis. No secondary lesions were observed in any of the abdominal organs.

The anatomopathological examination of the transbronchial biopsy showed the presence of neoplasia caused by epithelioid cells with abundant clear and granular cytoplasm, well-defined cell borders, and round and hyperchromatic nuclei, without the presence of prominent nucleoli (Figure 2). There were mild cellular atypia and foci of hemorrhage, necrosis, and absence of significant mitotic activity. Immunohistochemical (IHC) analysis revealed negativity for pan-cytokeratin (AE1, AE3, TTF1, RCC, MiTF, S-100, CK7, CK8, CK20, and desmin) and positivity for muscle markers and melanocyte antibodies (HMB-45, smooth actin muscle, CD10, and Melan-A) and Cathepsin K. These findings were indicative of a CCTL.

The challenges of verifying diagnosis and scheduling medical appointments, combined with suboptimal patient adherence, led to delayed treatment initiation. Only, five months after the biopsy, the patient underwent 2000 cGy doses of radiotherapy targeting the left lung in five fractions over one week and three cycles of doxorubicin 75 mg/m² within three months. However, owing to his compromised performance status and the presence of multiple nodules, surgical resection could not be performed. He presented with deterioration of the clinical picture and progression of pulmonary nodules in number and size two months after starting treatment, in addition to an increase in mediastinal lymph nodes and bilateral pleural effusion.

One month later, the patient was admitted to the emergency room with dyspnea, desaturation, and severe chest pain. Additionally, he had mild respiratory alkalosis associated with microcytic and hypochromic anemia without leukocytosis or normal platelet levels. He had normal electrolyte and renal function, increase in transaminase levels by about three times the upper normal value, and slight hypoalbuminemia, without changes in bilirubin or coagulogram. C-reactive protein was 8.9 mg/dl (reference value, < 1 mg/dl), with no reports of infectious symptoms. Serological tests for hepatitis, HIV and syphilis were negative. Chest radiography and CT revealed a significant progression of the lung masses (Figure 3). There was no evidence of other complications such as massive pleural effusion, pulmonary embolism, or pneumothorax, which would justify respiratory worsening in addition to the progression of tumor masses. Owing to the rapid progression of his condition, the patient underwent palliative sedation and died approximately 48 h after admission. A timeline specifying the entire clinical evolution of the patient is shown in Figure 4.

Discussion

Herein, we present a rare case of malignant CCTL. It is an uncommon and usually benign neoplasm composed of uniform round to polygonal epithelioid cells with clear or eosinophilic cytoplasm, well-defined borders, and prominent thin-walled



FIGURE 2

Histological sections of lung biopsy showing in hematoxylin-eosin staining the presence of clear cell tumors represented by epithelioid cells with abundant clear and granular cytoplasm, with well-delimited cell borders and round and hyperchromatic nuclei, discreet cell atypia and absence of significant mitotic activity (A). The immunohistochemical study revealed co-expression of markers: HMB45 (B), smooth muscle actin (C), Melan-A (D) and Cathepsin K (E), and negativity for cytokeratins CK7, CK8 and CK20 (F, G, H, respectively).

vascular channels (1), as observed in our case. Throughout the clinical and imaging evaluation, the malignant presentation of the patient was clear.

CCTL has no established epidemiology, clinical characteristics, diagnoses, or treatments. It is commonly confused with lung carcinoma or metastases from clear-cell carcinoma of the kidney (18). Patients are usually > 50 years old, but cases from 9 to 75 years of age have been reported (15, 19), with a similar prevalence between the sexes (20).

Among the benign manifestations of CCTL, an asymptomatic condition with an accidental diagnosis is common. However, as the size of this neoplasm can vary from several millimeters to approximately 16 cm (15), symptoms such as chest pain, cough,



FIGURE 3

Radiographic images showing extensive pulmonary masses on chest X-ray (A) and computed tomography of the chest in coronal (B) and axial planes (C, D), suggesting disease progression. A solid tumor, calcified upper left parasternal lesion with central necrosis, close contact with ipsilateral subclavian artery, without a cleavage plane and a tumor with similar characteristics on the lower right.

shortness of breath, hemoptysis, or fever may be observed in larger masses, especially if the size exceeds 2.2 cm (21–24).

We performed a literature review of the cases of benign and malignant CCTL (Table 1). Cases of CCTL with malignant behavior during the course of the disease and its outcomes have been reported sporadically. Certain characteristics, such as diameter greater than 2.5-3.0 cm, presence of symptoms, metastases, extensive necrosis, and abundant mitoses, are associated with a malignant presentation (24). Of the 76 benign occurrences found, necrosis was present in only 6.5% of the cases and mitotic activity was present in 4.0%. The largest mean mass diameter was 2.8 cm. In malignant cases, necrosis and mitotic activity were significantly more frequent, occurring in 50% and 12.5% respectively, and the mean diameter of the mass was 6.7 cm, which is 2.3 times greater than that of benign masses.

The site that was most affected by the primary nodule was the lower lobe of the lung. In this case, an extensive lesion was observed in the left upper lobe, characterized by necrosis and abundant vascularization, in addition to other scattered nodules in the parenchyma.

CCTL has already been associated with other diseases, such as tuberous sclerosis complex (79), Birt-Hogg-Dube Syndrome, and essential thrombocythemia (26, 27). However, in the malignant cases, it was not possible to determine the presence of risk factors strongly related to CCTL (Table 2).

PEC tumors can manifest in the bladder, uterus, breast, ovary, vulva, vagina, kidney, prostate, pancreas, jejunum and soft tissues, often with benign behavior (2, 14). PEC tumors originating in soft tissues and gynecological have some findings suggestive of malignancy (8), being indicative in the presence of 2 or more criteria: > 5 cm, infiltration, high-grade atypia, necrosis, vascular invasion, mitoses > 1/50 high power fields. Some authors (80) suggest greater specificity for PEC tumors of gynecological origin with at least 4 or more of these characteristics. But WHO followed the guidance of at least 3 criteria for malignancy (81).

JUNE 8, 2022 Bronchoscopy and lung mass biopsy: insufficient sample, possibility of granulomatous lymphadenitis JUNE 15, 2022 New bronchoscopy and lung mass biopsy: clear cell tumors of the lung. Histornatholowy (below (HE staining)	First evaluation with oncologist. First CT below
0	End of staging: no secondary lesions. PET-CT below
OCTOBER 31, 2022	
Start of radiotherapy (400 cGy/fraction)	
NOVEMBER 7. 2022	24 22
End of radiotherapy (5 fractions)	
	 NOVEMBER 22, 2022 First cycle of doxorubicin (139 mg) DECEMBER 14, 2022 Second cycle of doxorubicin (139 mg) JANUARY 5, 2023 Third cycle of doxorubicin (120 mg), dose reduced due to mucositis
JANUARY 28, 2023	Hemoglobin 8.2 g/dl Aspartate transaminase
Admission to emergency room due to respiratory failure, with evidence of progression of lung masses. Differential diagnoses such as infection and pulmonary embolism were excluded. Chest RX above. Laboratory tests on the side	Image: Property of the system Property of the system Hematocrit 26.2% Alanine aminotransferase 15 U/l Leukocytes 6.2 / mm3 Urea 32 mg/dl Platelets 558.000 / mm3 Creatinine 0.20 mg/dl Total Proteins 5.3 g/dl Total Bilirubin 1.00 mg/dl Albumin 2.5 g/dl C-reactive protein 8.9 mg/dl
	JANUARY 30, 2023 Definition of palliative care. Patient

radiotherapy and hospitalization until the outcome of the case are also specified.

There are currently no well-defined diagnostic methods for malignant CCTL. Zhao et al. proposed major and minor criteria for differentiating between benign and malignant manifestations (17). Imaging characteristics are nonspecific, and the tumor can appear in any lobe (although more commonly in the bilateral lower lobes) with a well-demarcated nodule without cavitation or calcification (24). The presence of a single and intraparenchymal pulmonary nodule is common to several diseases in addition to CCTL, such as

bronchogenic carcinoma and oat cell carcinoma, others tumors such as angiomyolipoma, and with metastases from clear cell carcinoma of the kidney, breast, liver, ovary, uterus and cervix, adrenal and melanoma (28, 82). Due to the nonspecificity of the image, histopathological and immunohistochemical is highly recommended for the CCTL characterization (19)

At optical microscopy the CCTL cells are round or oval, have clear cytoplasm strongly stained by PAS due to the presence of TABLE 1 Comparison of features between benign and malignant cases of perivascular epithelioid cells (PEC) tumors.

Characteristic	Benign (81) ^a n (%)	Malignant (8) ^b n (%)	
Sex			
Male	45 (55.5)	3 (37.5)	
Female	36 (44.4)	5 (62.2)	
Range age	49 (10 – 75) years	60 (49 – 75) years	
Symptoms			
Not reported	31 (38.2)	2 (25.0)	
Cough	7 (8.6)	3 (37.5)	
Chest pain	13 (16.0)	2 (25.0)	
Dyspnoea	8 (9.8)	0 (0.0)	
Absente	26 (32.0)	0 (0.0)	
Pulmonary tumor site			
Lower right	16 (19.7)	2 (25.0)	
Lower left	22 (27.1)	4 (50.0)	
Upper left	14 (17.2)	0 (0.0)	
Superior right	13 (16.0)	1 (12.5)	
Middle right	6 (7.4)	1 (12.5)	
Middle left	4 (4.9)	0 (0.0)	
Range of the largest diameter of the lesion	2.78 (0.4 - 12) cm	6.7 (3 - 16.2) cm	
Concurrent nodules			
Single node	75 (92.5)	5 (62.2)	
Two nodules	2 (2.4)	1 (12.5)	
More than two nodules	4 (4.9)	2 (25.0)	
Borders			
Regular	65 (80.2)	5 (62.2)	
Irregular	4 (4.9)	3 (37.5)	
Comorbidities			
Not reported	9 (11.1)	3 (37.5)	
COPD	4 (4.9)	0 (0.0)	
SAH	8 (9.8)	0 (0.0)	
Dyslipidemia	3 (3.7)	0 (0.0)	
Essential Thrombocythemia	2 (2.4)	0 (0.0)	
Absente	46 (56.7)	5 (62.2)	
Smoking			
Not reported	33 (40.7)	2 (25.0)	
Not reported Present	33 (40.7) 19 (23.4)	2 (25.0) 0 (0.0)	

(Continued)

TABLE 1 Continued

Characteristic	Benign (81) ^a n (%)	Malignant (8) ^b n (%)				
Biomarkers						
HMB45	48 (59.2)	5 (62.2)				
Vimentin	21 (25.9)	4 (50.0)				
S-100	22 (27.1)	3 (37.5)				
Melan-A	15 (18.5)	3 (37.5)				
CD34	11 (13.5)	1 (12.5)				
NSE	11 (13.5)	0 (0.0)				
SMA	4 (4.9)	1 (12.5)				
CD56	2 (2.4)	1 (12.5)				
Necrosis						
Not reported	25 (30.8)	1 (12.5)				
Present	5 (6.1)	4 (50.0)				
Absent	51 (62.9)	3 (37.5)				
Mitotic activity						
Not reported	16 (19.7)	1 (12.5)				
Present	3 (3.7)	1 (12.5)				
Absent	62 (76.5)	6 (75.0)				
Vascularization						
Not reported	18 (22.3)	-				
Present	55 (67.9)	6 (75.0)				
Absent	8 (9.8)	2 (25.0)				

COPD, chronic obstructive pulmonary disease; SAH, systemic arterial hypertension. ^aReferences (1, 3, 18, 19, 21, 24–78). ^bReferences (14, 17, 23, 36, 74–77).

References (14, 1/, 23, 36, /4-//).

glycogen, mild nuclear pleomorphism and a characteristic plexus of prominent sinusoidal vessels, but mitotic activity and necrosis are absent (19, 24). This characteristic pattern helps to differentiate CCTL from other conditions, such as angiomyolipoma and adrenal carcinoma (28, 82).

Tumor cells show reactivity for HMB-45 in about 90% of cases, similar to most mesenchymal tumors, and do not express cytokeratins (AE1/AE3, CK7, CK8 and CK20) and epithelial membrane antigen (EMA) in 90% of cases, a fact that facilitates differentiation from other types of tumors (18, 19, 28), as observed in our case. The immunoreactivity of renal cell carcinoma to EMA, cytokeratins, CD10 and RCC are useful for the differentiation of CCTL, despite having vimentin as a positive marker in common. However, in the case of atypical melanomas, positivity to HMB-45 and S-100 present in both, does not allow elucidation of the diagnosis (28). Cathepsin K, the main marker for differentiating PEC tumors from other neoplasms, was diffusely positive in 5 cases of CCTL described in the literature (82), demonstrating the superiority of this marker over HMB-45 in differentiating CCTL

Reference	Sex/Age	Diagnostic exam	Treatment	Survival
Gaffey M.J. 1990 (36)	Male/66	unreported imaging exam Excisional biopsy	Surgical resection	Metastatic lesions remaining after 13 years. Death after 4 years of recurrence
Jeremy R.P. 2006 (77)	Female/53	CT Excisional biopsy Immunohistochemistry	Surgical resection Resection of brain metastases	Not reported
Ting Ye 2010 (76)	Female/50	CT Excisional biopsy Immunohistochemistry	Surgical resection	Not reported
Benedict Yan 2011 (23)	Female/75	CT Excisional biopsy Immunohistochemistry	Surgical resection	No signs of recurrence after 10 years. Death without clear cause
Hyun-ju Lim 2013 (14)	Male/60	PET/CT Excisional biopsy Immunohistochemistry	Surgical resection	Metastatic lesions remaining after 1 year
Yinghua Song 2017 (75)	Female/49	CT Excisional biopsy Immunohistochemistry	Surgical resection	Clinically stable during follow-up
Jiyong Wu 2019 (74)	Male/67	CT Excisional biopsy Immunohistochemistry	Not reported	Not reported
Jikai Zhao 2019 (17)	Male/54	CT Excisional biopsy Immunohistochemistry	Surgical resection Chemotherapy (paclitaxel and carboplatin)	Clinically stable during follow-up

TABLE 2 Malignant cases of perivascular epithelioid cells (PEC) tumors reported in literature.

CT, computed tomography; PET-CT, positron emission tomography-CT.

from other lung tumors. Thus, the co-expression of HMB-45 and cathepsin K markers ensures a more accurate diagnosis.

Furthermore, other markers such as S-100, vimentin, CD34, Melan-A, microphthalmia factor, neuron-specific enolase, cathepsin B, and CD1a can be expressed (18, 20, 28). CD10 may be expressed in other types of non-pulmonary PEComas (83, 84). Our patient tested positive for HMB-45, Melan-A, smooth muscle actin, and CD10, and tested negative for cytokeratins.

Complete surgical resection is probably the most effective therapeutic method at present (85). Radiotherapy may be considered for patients with surgical contraindications (20). Chemotherapy should be administered for malignant manifestations, as in the reported case. CCTLs upregulate the mammalian target of rapamycin (mTOR) signaling pathway; therefore, mTOR inhibitors can be used in such cases (86, 87).

Despite being described in the literature, mitotic activity and some biomarkers may not differ between malignant and benign CCTLs. Except for lesion size, clinical characteristics and complementary examinations cannot predict a worse development. Close follow-up can identify when CCTL cases may exhibit unexpected and malignant behaviors.

Conclusion

Herein, we report the clinical evolution of a patient with malignant CCTL. Although this form of presentation is extremely

rare in the literature, it is not yet possible to establish clinical, immunohistochemical, or histopathological criteria to predict the prognosis and possible malignant evolution. Although described in some studies, mitotic activity does not differentiate between benign and malignant tumors. The size of the initial lesion, presence of multiple lesions, and patient's clinical evolution seem to be the only comparative parameters.

Data availability statement

The original contributions presented in the study are included in the article/supplementary material. Further inquiries can be directed to the corresponding author.

Ethics statement

Written informed consent was obtained from the proxy for the patient described in the manuscript.

Author contributions

MC: Conceptualization, Methodology, Writing – review & editing. LV: Data curation, Formal Analysis, Investigation, Writing – original draft. RM: Data curation, Formal Analysis, Investigation, Writing – original draft. ÉM: Conceptualization, Methodology, Writing – original draft. AF: Conceptualization, Methodology, Writing – original draft. SS: Conceptualization, Methodology, Writing – original draft. GS: Supervision, Visualization, Writing – review & editing. MM: Data curation, Visualization, Writing – review & editing. MZ: Data curation, Investigation, Visualization, Writing – review & editing. DQ: Conceptualization, Investigation, Supervision, Writing – review & editing.

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Medical School of São Paulo State University (Unesp), Botucatu Campus

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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References

1. Liebow AA, Castleman B. Benign clear cell ("sugar") tumors of the lung. Yale J Biol Med (1971) 43:213-22.

2. Travis WD, Brambilla E, Nicholson AG, Yatabe Y, Austin JHM, Beasley MB, et al. The 2015 world health organization classification of lung tumors: impact of genetic, clinical and radiologic advances since the 2004 classification. J Thorac Oncol Off Publ Int Assoc Study Lung Cancer (2015) 10:1243–60. doi: 10.1097/JTO.00000000000000630

3. Shen L, Lin J, Ren Z, Wang B, Liu Y, Yuan J, et al. Clear cell tumor of the lung could be aggressive: a case report and review of the literature. *J Cardiothorac Surg* (2020) 15:177. doi: 10.1186/s13019-020-01224-w

4. Pann CC, Yu IT, Yang AH, Chiang H. Clear cell myomelanocytic tumor of the urinary bladder. *Am J Surg Pathol* (2003) 27(5):689-92. doi: 10.1097/00000478-200305000-00013

5. Vang R, Kempson RL. Perivascular epithelioid cell tumor ('PEComa') of the uterus: a subset of HMB-45-positive epithelioid mesenchymal neoplasms with an uncertain relationship to pure smooth muscle tumors. *Am J Surg Pathol* (2002) 26:1–13. doi: 10.1097/00000478-200201000-00001

6. Kim H, Kim J, Lee SK, Cho EY, Cho SY. TFE3-expressing perivascular epithelioid cell tumor of the breast. J Pathol Transl Med (2019) 53:62-5. doi: 10.4132/jptm.2018.08.30

7. Anderson AE, Yang X, Young RH. Epithelioid angiomyolipoma of the ovary: a case report and literature review. *Int J Gynecol Pathol Off J Int Soc Gynecol Pathol* (2002) 21:69–73. doi: 10.1097/00004347-200201000-00013

8. Folpe AL, Mentzel T, Lehr H-A, Fisher C, Balzer BL, Weiss SW. Perivascular epithelioid cell neoplasms of soft tissue and gynecologic origin: a clinicopathologic study of 26 cases and review of the literature. *Am J Surg Pathol* (2005) 29:1558–75. doi: 10.1097/01.pas.0000173232.22117.37

9. Martignoni G, Pea M, Reghellin D, Zamboni G, Bonetti F. PEComas: the past, the present and the future. *Virchows Arch Int J Pathol* (2008) 452:119–32. doi: 10.1007/s00428-007-0509-1

10. Pan C-C, Yang A-H, Chiang H. Malignant perivascular epithelioid cell tumor involving the prostate. *Arch Pathol Lab Med* (2003) 127:e96–8. doi: 10.5858/2003-127-e96-MPECTI

11. Zamboni G, Pea M, Martignoni G, Zancanaro C, Faccioli G, Gilioli E, et al. Clear cell "Sugar" Tumor of the pancreas: A novel member of the family of lesions characterized by the presence of perivascular epithelioid cells. *Am J Surg Pathol* (1996) 20:722. doi: 10.1097/00000478-199606000-00010

12. Yanai H, Matsuura H, Sonobe H, Shiozaki S, Kawabata K. Perivascular epithelioid cell tumor of the jejunum. *Pathol - Res Pract* (2003) 199:47-50. doi: 10.1078/0344-0338-00353

13. Mai KT, Belanger EC. Perivascular epithelioid cell tumour (PEComa) of the soft tissue. *Pathol (Phila)* (2006) 38:415–20. doi: 10.1080/00313020600922504

14. Lim H, Lee HY, Han J, Choi YS, Lee KS. Uncommon of the uncommon: Malignant perivascular epithelioid cell tumor of the lung. *Korean J Radiol* (2013) 14:692–6. doi: 10.3348/kjr.2013.14.4.692

15. Tazelaar HD, Batts KP, Srigley JR. Primary extrapulmonary sugar tumor (PEST): a report of four cases. *Mod Pathol Off J U S Can Acad Pathol Inc* (2001) 14:615–22. doi: 10.1038/modpathol.3880360

16. Chakrabarti A, Bandyopadhyay M, Purkayastha B. Malignant perivascular epitheloid cell tumour (PEComa) of the lung – a rare entity. *Innov Surg Sci* (2017) 2:39–42. doi: 10.1515/iss-2016-0032

17. Zhao J, Teng H, Zhao R, Ding W, Yu K, Zhu L, et al. Malignant perivascular epithelioid cell tumor of the lung synchronous with a primary adenocarcinoma: one

case report and review of the literature. BMC Cancer (2019) 19:235. doi: 10.1186/s12885-019-5383-0

 Olivencia-Yurvati AH, Rodriguez AE. Clear cell "Sugar" Tumor of the lung: benign or Malignant? Int Surg (2015) 100:924–6. doi: 10.9738/INTSURG-D-14-00021.1

19. Tsilimigras DI, Bakopoulos A, Ntanasis-Stathopoulos I, Gavriatopoulou M, Moris D, Karaolanis G, et al. Clear cell "sugar tumor" of the lung: Diagnostic features of a rare pulmonary tumor. *Respir Med Case Rep* (2018) 23:52–4. doi: 10.1016/j.rmcr.2017.12.001

20. Ntanasis-Stathopoulos I, Kyriazoglou A, Tsilimigras DI, Tzanninis IG, Liontos M, Dimopoulos MA, et al. Clear cell sugar tumor of the lung; a systematic review for a rare entity. *J BUON Off J Balk Union Oncol* (2021) 26:17–32.

21. Gora-Gebka M, Liberek A, Bako W, Szumera M, Korzon M, Jaskiewicz K. The "sugar" clear cell tumor of the lung—clinical presentation and diagnostic difficulties of an unusual lung tumor in youth. *J Pediatr Surg* (2006) 41:e27–9. doi: 10.1016/j.jpedsurg.2006.02.035

22. Zarbis N, Barth TFE, Blumstein NM, Schelzig H. Pecoma of the lung: a benign tumor with extensive 18F-2-deoxy-d-glucose uptake. *Interact Cardiovasc Thorac Surg* (2007) 6:676–8. doi: 10.1510/icvts.2007.154039

23. Yan B, Yau EX, Petersson F. Clear cell 'sugar' tumour of the lung with Malignant histological features and melanin pigmentation – the first reported case. *Histopathology* (2011) 58:498–500. doi: 10.1111/j.1365-2559.2011.03773.x

24. Wang G, Zhang D, Diao X, Wen L. Clear cell tumor of the lung: a case report and literature review. *World J Surg Oncol* (2013) 11:247. doi: 10.1186/1477-7819-11-247

25. Gupta I, Bhalla AS, Mallick S, Mohan A, Jain D. An enigma of a rare pulmonary neoplasm in bronchial brushings. *Cytopathol Off J Br Soc Clin Cytol* (2021) 32:534–7. doi: 10.1111/cyt.12971

26. Yazak V, Sargin G, Yavasoglu I, Kadikoylu G, Tataroglu C, Bozkurt G, et al. Lung clear "Sugar" Cell tumor and JAK V617F positive essential thrombocythemia: a simple coincidence? *Mediterr J Hematol Infect Dis* (2013) 5:e2013021. doi: 10.4084/MJHID.2013.021

27. Sen S, Senturk E, Kuman NK, Pabuscu E, Kacar F. PEComa (Clear cell "Sugar" Tumor) of the lung: A benign tumor that presented with trombocytosis. *Ann Thorac Surg* (2009) 88:2013–5. doi: 10.1016/j.athoracsur.2009.05.041

28. Policarpio-Nicolas ML, Covell J, Bregman S, Atkins K. Fine needle aspiration cytology of clear cell "sugar" tumor (PEComa) of the lung: Report of a case. *Diagn Cytopathol* (2008) 36:89–93. doi: 10.1002/dc.20647

29. Becker NH, Soifer I. Benign clear cell tumor ("sugar tumor") of the lung. *Cancer* (1971) 27:712–9. doi: 10.1002/1097-0142(197103)27:3<712::aid-cncr2820270330>3.0.co;2-b

30. Hoch WS, Patchefsky AS, Takeda M, Gordon G. Benign clear cell tumor of the lung. *ultrastructural study. Cancer* (1974) 33:1328–36. doi: 10.1002/1097-0142(197405) 33:5<1328::aid-cncr2820330518>3.0.co;2-j

31. Harbin WP, Mark GJ, Greene RE. Benign clear-cell tumor ("Sugar" Tumor) of the lung: a case report and review of the literature. *Radiology* (1978) 129:595–6. doi: 10.1148/129.3.595

32. Makek M, Rüttner JR. Benigner hellzelliger lungentumor (»Sugar tumor«). Langenbecks Arch Für Chir (1979) 348:177–81. doi: 10.1007/BF01239445

33. Andrion A, Mazzucco G, Gugliotta P, Monga G. Benign clear cell ('Sugar') tumor of the lung A light microscopic, histochemical, and ultrastructural study with a review of the literature. *Cancer* (1985) 56:2657–63. doi: 10.1002/1097-0142(19851201) 56:11<2657::AID-CNCR2820561121>3.0.CO;2-U

34. Nakanishi K, Kawai T, Suzuki M. Benign clear cell tumor of the lung a histopathologic study. *Pathol Int* (1988) 38:515–22. doi: 10.1111/j.1440-1827.1988.tb02324.x

35. Termeer A, Arkenbout PM, Lacquet LK, Cox AL. Benign clear cell tumour of the lung–intermediate filament typing as a tool in differential diagnosis. *Eur Respir J* (1988) 1:288–90. doi: 10.1183/09031936.93.01030288

36. Gaffey MJ, Mills SE, Askin FB, Ross GW, Sale GE, Kulander BG, et al. Clear cell tumor of the lung. A clinicopathologic, immunohistochemical, and ultrastructural study of eight cases. *Am J Surg Pathol* (1990) 14:248–59. doi: 10.1097/00000478-199003000-00006

37. Mishina T, Suzuki I, Fujino M, Watanabe N, Akita H, Narita Y, et al. A benign clear cell tumor of the lung that grew gradually over five years. *Nihon Kyobu Shikkan Gakkai Zasshi* (1995) 33:765–70.

38. Takanami I, Kodaira S, Imamura T. The use of transbronchial lung biopsy to establish a diagnosis of benign clear cell tumor of the lung: Report of a case. *Surg Today* (1998) 28:985–7. doi: 10.1007/s005950050268

39. Hashimoto T, Oka K, Hakozaki H, Kuraoka S, Otani S, Kawano H, et al. Benign clear cell tumor of the lung. *Ultrastruct Pathol* (2001) 25:479-83. doi: 10.1080/019131201753343511

40. Kula* Ö, Bekçi* TT, Şenol* T, Pazarli* P. Akciğerin iyi huylu clear cell tümörü. Turk Klin Arch Lung (2002) 3:67–9.

41. Masur Y, Becker HP, Fackeldey V. [Sugar tumor. Rare differential diagnosis of pulmonary clear cell tumor after renal carcinoma]. *Urol Ausg A* (2003) 42:1250–4. doi: 10.1007/s00120-003-0376-6

42. Santana ANC, Nunes FS, Ho N, Takagaki TY. A rare cause of hemoptysis: benign sugar (clear) cell tumor of the lung. *Eur J Cardiothorac Surg* (2004) 25:652–4. doi: 10.1016/j.ejcts.2004.01.022

43. Drebber U, Ortmann M, Kasper HU, Kern MA, Fischer UM, Nemat AT, et al. Klarzelltumor der lunge. *Pathol* (2005) 26:378–82. doi: 10.1007/s00292-005-0755-4

44. Jordá Aragón C, Froufe Sánchez A, Padilla Alarcón J. Benign clear cell tumor of the lung. Arch Bronconeumol (2005) 41:59. doi: 10.1016/s1579-2129(06)60396-0

45. Adachi Y, Kitamura Y, Nakamura H, Taniguchi Y, Miwa K, Horie Y, et al. Benign clear (sugar) cell tumor of the lung with CD1a expression. *Pathol Int* (2006) 56:453–6. doi: 10.1111/j.1440-1827.2006.01988.x

46. Kavunkal AM, Pandiyan MS, Philip MA, Parimelazhagan KN, Manipadam MT, Cherian VK. Large clear cell tumor of the lung mimicking Malignant behavior. *Ann Thorac Surg* (2007) 83:310–2. doi: 10.1016/j.athoracsur.2006.04.038

47. Ayadi-Kaddour A, Ben Slama S, Braham E, Abid L, Djilani H, Ismail O, et al. Clear cell tumor of the lung. A case report with review of the literature. *Rev Pneumol Clin* (2006) 62:395–8. doi: 10.1016/s0761-8417(06)75474-8

48. Shiran M, Tan G, Arunachalam N, Sabariah A, Pathmanathan R. Clear cell "sugar" tumour of the lung: a case report. *Malays J Pathol* (2006) 28:113–6.

49. Hirata T, Otani T, Minamiguchi S. Clear cell tumor of the lung. Int J Clin Oncol (2006) 11:475–7. doi: 10.1007/s10147-006-0604-3

50. Edelweiss M, Gupta N, Resetkova E. Preoperative diagnosis of clear cell "sugar" tumor of the lung by computed tomography-guided fine-needle biopsy and core-needle biopsy. *Ann Diagn Pathol* (2007) 11:421-6. doi: 10.1016/j.anndiagpath.2006.12.015

51. Kim WJ, Kim SR, Choe YH, Lee KY, Park SJ, Lee HB, et al. Clear cell "Sugar" Tumor of the lung: A well-enhanced mass with an early washout pattern on dynamic contrast-enhanced computed tomography. *J Korean Med Sci* (2008) 23:1121–4. doi: 10.3346/jkms.2008.23.6.1121

52. Nehra D, Vagefi PA, Chang H, Louissaint A, Mino-Kenudson M, Ryan DP. Clear cell sugar tumor of the lung masquerading as tuberculosis in a pediatric patient. *J Thorac Cardiovasc Surg* (2010) 139:e108–9. doi: 10.1016/j.jtcvs.2009.03.037

53. Mizobuchi T, Masahiro N, Iwai N, Kohno H, Okada N, Nakada S. Clear cell tumor of the lung: surgical and immunohistochemical findings. *Gen Thorac Cardiovasc Surg* (2010) 58:243–7. doi: 10.1007/s11748-009-0513-2

54. Vijayabhaskar R, Mehta SS, Deodhar KK, Pramesh CS, Mistry RC. PEComa of the lung. J Cancer Res Ther (2010) 6:109–11. doi: 10.4103/0973-1482.63548

55. Han B, Jiang G, Wang H, He W, Liu M, Song N. Benign clear cell tumor of the lung. Ann Thorac Surg (2010) 89:2012–4. doi: 10.1016/j.athoracsur.2009.10.017

 Kalkanis A, Trianti M, Psathakis K, Mermigkis C, Kalkanis D, Karagkiouzis G, et al. A clear cell tumor of the lung presenting as a rapidly growing coin lesion: is it really a benign tumor? *Ann Thorac Surg* (2011) 91:588–91. doi: 10.1016/j.athoracsur.2010.07.043

57. Neri S, Ishii G, Aokage K, Hishida T, Yoshida J, Nishimura M, et al. Multiple perivascular epithelioid cell tumors: clear cell tumor of the lung accompanied by angiomyolipoma of the liver. *Ann Thorac Cardiovasc Surg Off J Assoc Thorac Cardiovasc Surg Asia* (2014) 20 Suppl:453–6. doi: 10.5761/atcs.cr.12.02179

58. Sanai Raggad S, Zouaoui A, Mehiri N, Cherif J, Zakhama H, Kchir MN, et al. [Sugar tumor]. *Tunis Med* (2011) 89:202–5.

59. Das S, Cherian SV, Das N, Hamarneh WA, Singh Garcha A, Preet Singh P, et al. A 52-year-old smoker with an incidental pulmonary nodule. *Chest* (2012) 141:1346–50. doi: 10.1378/chest.11-2242

60. Xu Q, Lu C, Li L, Xu K. Clear cell tumor of the lung. *Med (Baltimore)* (2016) 95: e4492. doi: 10.1097/MD.00000000004492

61. Gunji-Niitsu Y, Kumasaka T, Kitamura S, Hoshika Y, Hayashi T, Tokuda H, et al. Benign clear cell "sugar" tumor of the lung in a patient with Birt-Hogg-Dubé syndrome: a case report. *BMC Med Genet* (2016) 17:85. doi: 10.1186/s12881-016-0350-y 62. Chang M, Lim D, Genovesi M. Clear cell "sugar" tumor of the lung: a case report and review of the literature. AME Case Rep (2018) 2:40. doi: 10.21037/acr.2018.08.03

63. Yeon EK, Kim JI, Won KY, Lee HN. Growth pattern change of a benign clear cell "sugar" tumor of the lung: Serial imaging surveillance over seven years. *Oncol Lett* (2018) 15:8652–4. doi: 10.3892/ol.2018.8391

64. Sjniari M, Miele E, Stati V, Di Cristofano C, Diso D, Pecoraro Y, et al. Early clear cell "sugar" lung cancer management: A case report and a brief literature review. *Thorac Cancer* (2019) 10:1289–94. doi: 10.1111/1759-7714.13077

65. Page S, Yong MS, Sinha A, Saxena P. Clear cell sugar tumour: a rare tumour of the lung. ANZ J Surg (2020) 90:E54–6. doi: 10.1111/ans.15218

66. Onorati I, Kambouchner M, Martinod E. A pigmented lung perivascular epithelioid cell tumor. *Am J Respir Crit Care Med* (2021) 203:1023-4. doi: 10.1164/rccm.202004-1358IM

67. Yu S, Zhai S, Gong Q, Hu X, Yang W, Liu L, et al. Perivascular epithelioid cell tumor of the lung: A case report and literature review. *Thorac Cancer* (2022) 13:2542–53. doi: 10.1111/1759-7714.14583

68. Donato UM, Ferguson K. Perivascular epithelioid cell tumor (PEComa) of the lung in a 56-year-old female patient: a case report. *Cureus* (2022) 14:e29246. doi: 10.7759/cureus.29246

69. Buero A, Tchercansky A, Olivera Lopez S, Pankl LG, Chimondeguy DJ, Lyons GA, et al. PEComa pulmonar. *Med BAires* (2022) 82(6):963–6.

70. Sale GF, Kulander BG. Benign clear cell tumor of lung with necrosis. *Cancer* (1976) 37:2355–8. doi: 10.1002/1097-0142(197605)37:5<2355::aid-cncr2820370527>3.0.co;2-4

71. Arafah MA, Raddaoui E, Alsheikh A, Hajjar WM, Alyousef F. Mimicry of sugar tumor and minute pulmonary meningothelial-like nodule to metastatic lung deposits in a patient with rectal adenocarcinoma. *Ann Saudi Med* (2013) 33:400–3. doi: 10.5144/ 0256-4947.2013.400

72. Fernandez A, Jatene FB, Zamboni M. Diagnóstico e estadiamento do câncer de pulmão. J Pneumol (2002) 28:219–28. doi: 10.1590/S0102-35862002000400006

73. Wang B, Xu X, Zhao Z. Clear cell "sugar" tumor of the lung: Diagnostic characteristics of a rare pulmonary tumor: A case report and review of literature. *Med (Baltimore)* (2023) 102:e33035. doi: 10.1097/MD.00000000033035

74. Wu J, Jiang L, Zhang F, Huang Y, Wang H. Malignant perivascular epithelioid cell tumor of lung on FDG PET/CT. *Clin Nucl Med* (2019) 44:469–71. doi: 10.1097/RLU.00000000002505

75. Song Y, Chen F, Zhang C, Lin X. Spindle cell subtype of pulmonary clear cell tumor with prominent calcification and Malignant potential. *Thorac Cancer* (2017) 8:530–4. doi: 10.1111/1759-7714.12457

76. Ye T, Chen H, Hu H, Wang J, Shen L. Malignant clear cell sugar tumor of the lung: patient case report. J Clin Oncol Off J Am Soc Clin Oncol (2010) 28:e626–628. doi: 10.1200/JCO.2010.29.6939

77. Parfitt JR, Keith JL, Megyesi JF, Ang LC. Metastatic PEComa to the brain. Acta Neuropathol (Berl) (2006) 112:349–51. doi: 10.1007/s00401-006-0105-5

78. Papla B, Demczuk S, Malinowski E. Benign clear-cell "sugar" tumor of the lunga case report. *Pol J Pathol Off J Pol Soc Pathol* (2003) 54:183–5.

79. Flieder DB, Travis WD. Clear cell "sugar" tumor of the lung: association with lymphangioleiomyomatosis and multifocal micronodular pneumocyte hyperplasia in a patient with tuberous sclerosis. *Am J Surg Pathol* (1997) 21:1242–7. doi: 10.1097/00000478-199710000-00017

80. Schoolmeester JK, Howitt BE, Hirsch MS, Dal Cin P, Quade BJ, Nucci MR. Perivascular epithelioid cell neoplasm (PEComa) of the gynecologic tract: clinicopathologic and immunohistochemical characterization of 16 cases. *Am J Surg Pathol* (2014) 38:176–88. doi: 10.1097/PAS.000000000000133

81. Bennett JA, Braga AC, Pinto A, Van de Vijver K, Cornejo K, Pesci A, et al. Uterine PEComas: A morphologic, immunohistochemical, and molecular analysis of 32 tumors. *Am J Surg Pathol* (2018) 42:1370–83. doi: 10.1097/PAS.00000000001119

82. Caliò A, Mengoli MC, Cavazza A, Rossi G, Ghimenton C, Brunelli M, et al. Cathepsin K expression in clear cell "sugar" tumor (PEComa) of the lung. *Virchows Arch* (2018) 473:55–9. doi: 10.1007/s00428-018-2325-1

83. Ueberschaar S, Goebeler M, Kneitz H. CD10-positive cutaneous PEComa: an extremely rare skin tumour. *Case Rep Dermatol* (2020) 12:192–8. doi: 10.1159/000510718

84. Armah HB, Parwani AV. Malignant perivascular epithelioid cell tumor (PEComa) of the uterus with late renal and pulmonary metastases: a case report with review of the literature. *Diagn Pathol* (2007) 2:45. doi: 10.1186/1746-1596-2-45

85. Eldessouki I, Gaber O, Riaz MK, Wang J, Abdel Karim N. Clinical presentation and treatment options for clear cell lung cancer: university of cincinnati A case series and literature review of clear cell lung cancer. *Asian Pac J Cancer Prev APJCP* (2018) 19:2373–6. doi: 10.22034/APJCP.2018.19.9.2373

86. Sanfilippo R, Jones RL, Blay J-Y, Le Cesne A, Provenzano S, Antoniou G, et al. Role of chemotherapy, VEGFR inhibitors, and mTOR inhibitors in advanced perivascular epithelioid cell tumors (PEComas). *Clin Cancer Res Off J Am Assoc Cancer Res* (2019) 25:5295–300. doi: 10.1158/1078-0432.CCR-19-0288

87. Subbiah V, Trent JC, Kurzrock R. Resistance to mammalian target of rapamycin inhibitor therapy in perivascular epithelioid cell tumors. *J Clin Oncol Off J Am Soc Clin Oncol* (2010) 28:e415. doi: 10.1200/JCO.2010.29.4678