

See discussions, stats, and author profiles for this publication at: <https://www.researchgate.net/publication/367091540>

Adsorption challenge in the PDMS-based microfluidic systems for drug screening application

Conference Paper · November 2022

CITATIONS

0

READS

113

12 authors, including:



Reza Mohammadigharehbagh
Eskisehir Osmangazi University

57 PUBLICATIONS 446 CITATIONS

[SEE PROFILE](#)



Ozlem Tomsuk

7 PUBLICATIONS 53 CITATIONS

[SEE PROFILE](#)



Hamed Ghorbanpoor
Eskisehir Osmangazi University

44 PUBLICATIONS 185 CITATIONS

[SEE PROFILE](#)



Aliakbar Ebrahimi
Eskisehir Osmangazi University

27 PUBLICATIONS 45 CITATIONS

[SEE PROFILE](#)

Some of the authors of this publication are also working on these related projects:



Development of Electrochemical Biosensors [View project](#)



Design of Advanced Three-Dimensional (3D) Eco-Friendly Fiber Reinforced Bio-Composite Structures [View project](#)



FIBER &
POLYMER
RESEARCH INSTITUTE

GEBZE
TEKNİK ÜNİVERSİTESİ



İTÜ



Türkiye Bilimler Akademisi
TURKISH ACADEMY OF SCIENCES

11

ULUSLARARASI LİF VE POLİMER ARAŞTIRMALARI SEMPOZYUMU

11TH INTERNATIONAL FIBER AND POLYMER RESEARCH SYMPOSIUM

4 - 5 Kasım
November 2022

Gebze Teknik Üniversitesi
Gebze Technical University

GTÜ Kongre ve Kültür Merkezi
GTU Congress and Cultural Center

www.ulpas.org

Biy- ve Fonksiyonel Lifler | Bio- & Functional Fibers

BİLDİRİ KİTAPÇIĞI BOOK OF PROCEEDINGS

ISBN: 978-605-9332-12-5

Adsorption challenge in the PDMS-based microfluidic systems for drug screening application

MOHAMMADIGHAREHBAGH, Reza^{1,2}, TOMSUK, Özlem^{2,3}, GHORBANPOOR, Hamed^{2,4}, EBRAHIMI, Aliakbar², ABDULLAYEVA, Nuran^{2,5}, GASIMZADE, Nigar^{2,5}, KOÇ, Yücel^{2,6}, ÖZEL, Ceren^{2,5}, GHORBANI, Aynaz^{2,5}, DEMIR, Bahar², UYSAL, Onur^{2,5}, DOĞAN GUZEL, Fatma⁷, EKER SARIBOYACI, Ayla^{2,5}, AVCI, Huseyin^{2,8,9*}

¹Eskişehir Osmangazi University, Faculty of Science and Letter, Department of Physics, Eskişehir, Turkey

²Cellular Therapy and Stem Cell Production Application and Research Center (ESTEM), Eskişehir Osmangazi University, Eskişehir, Turkey

³Department of Graduate School of Natural and Applied Sciences, Biotechnology and Biosafety, Eskişehir Osmangazi University, Eskişehir, Turkey

⁴Department of Biomedical Engineering, Eskişehir Osmangazi University, Eskişehir, Turkey

⁵Department of Stem Cell, Eskişehir Osmangazi University, Eskişehir, Turkey

⁶Department of Chemical Engineering, Eskişehir Osmangazi University, Eskişehir, Turkey

⁷Department of Biomedical Engineering, Ankara Yildirim Beyazıt University, Ankara, Turkey

⁸Department of Metallurgical and Materials Engineering, Eskişehir Osmangazi University Eskişehir, Turkey

⁹Translational Medicine Application and Research Center, Eskişehir Osmangazi University, Eskişehir, Turkey

*Corresponding author: havci@ogu.edu.tr

SUMMARY

Drug screening is one of the demand areas due to close and direct dependency on human health. On the other hand, recently microfluidic systems have been increasingly used for drug development and screening purposes. However, this system has some challenges such as adsorption issue which can affect pharmacokinetic–pharmacodynamic (PK-PD) of the drugs. Thus, in this research, the issue was characterized and evaluated by UV-Vis spectrophotometry and FTIR spectroscopy devices as a model drug of cisplatin. Despite of strong relationship between logP and adsorption, and the very low value of logP in the drug candidate, the results for both apical and basal planes of the microfluidic chip confirmed the adsorption. In the UV-Vis spectrophotometry, the basal plane show 5%, and 10% higher adsorption compared to apical and control polydimethylsiloxane (PDMS)-based microfluidic. Additionally, the FTIR patterns were a good coincide with UV-Vis results.

PURPOSE

The main purpose of this research is to prepare a PDMS-based microfluidic platform for the investigation of surface adsorption in the target drug analysis experiments. Today, this system has frequently been implemented as an alternative to animal-based trials for the drug development and study of disease.

Failure in the similarities between animals and human resulted to the attraction and motivation of researchers to rapidly move toward microfluidics-based organ-on-a-chip system [1-7]. On the other hand, using new and state-of-art technologies lead to new hurdles such as surface adsorption. Therefore, in this work, we focused on the adsorption issue. Finally, the prepared system is characterized by various devices for evaluation adsorption effect on the obtained results.

INTRODUCTION

Drug development and screening in recent years get more attention due to their significance and effect on human beings. Till today, conventional animal-based methods are performed for introducing a new drug in the market, but these routes faced big challenges i.e., 10-15 years to develop as well as spending approximately 2.6 billion dollars per each drug [8]. To overcome the challenges, crucial needs for emerging new platforms have not been ignorable. Therefore, the microfluidic organ-on-a-chip (OOC) system was discovered [9] and has been under improvement by diverse research groups [10-12]. The minimum cost of developing and using drugs from the initial stage to the market, portability, ease of access, best similarities with human structure, and reliability of the obtained results compared to animal trials are

classified as some advantages of this system [11, 13]. By benefiting from the advantages, different drug investigations have been carried out for OOCs applications [14]. In this research, the above-mentioned platform was performed for simulation liver-on-a-chip (LOC) and cisplatin was used as a drug candidate for the investigation of adsorption.

Regarding to the literature, diverse polymer types of materials have been implemented in microfluidic platforms [15, 16]. Currently, PDMS was the most attractive one concerning to the other owing to transparency, almost non-toxicity, best compatibility in biological applications in addition to flexibility, permeability and inexpensively [17-19].

Despite using the PDMS-based LOC facilitating and paving the way for a drug study, on the other side, other drawbacks, for instance, adsorption and absorption of small molecules due to the hydrophobic property of PDMS surface, are one of the reason to be investigated. To hinder these important barriers, therefore, some research groups provide coating, doping, grafting, and extraction effective routes [20, 21]. More detail about these methods was reviewed and exists in the literature [22, 23]. In addition to the above-mentioned pathways, in some cases, the researchers completely replaced PDMS polymer with alternatives such as polylactic [24], poly(methyl methacrylate) (PMMA), poly(carbonate) (PC) [25], high-density polyethylene (HDPE), and low-density polyethylene (LDPE) [26], and so on.

EXPERIMENTAL

Material: In this work, Cisplatin (Cas No: 15663-27-1, Product No: 479306) was used as a model drug. PDMS and PMMA were utilized for the fabrication of apical and basal channels of the chip, and to fix as well as prevent any misorientation in the channels and membrane. Polyethylene terephthalate (PET) membrane without a cell line was selected for this research as a preliminary investigation.

Analysis: Fourier transform infrared spectroscopy (FTIR), and ultraviolet-visible (UV-Vis) spectrophotometry was employed for in detail characterization of the molecular structure and existence of chemical groups on the chip's channels and the samples. The FTIR and UV-Vis spectrophotometry was performed in the 200-4000 cm^{-1} , and 190-1100 nm ranges, respectively.

Method: The apical and basal planes of the chip were initially poured into the home-made prepared mold by mixing the prepolymer and cross-linker with 10:1 w/w. Then the well-known soft lithography followed in the clean room; vacuumed the molded PDMS, and finally heat for overnight for the curing process. After curation,

the prepared polymer was unmolded and kept for the chip formation procedure.

The prepared polymers were carefully cleaned with acetone a few times, then a high-resolution camera was employed for precise in-situ mounting of the basal and apical channels, and basement membrane. The drug candidate in our case passed through the basal channel. The schematic of this process was visualized in Fig. 1.

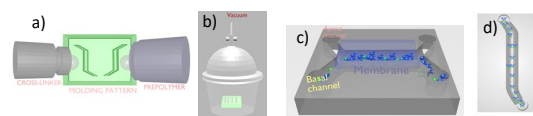


Figure 1. The schematic representative of preparation of the chip by using soft lithography

EXPERIMENTAL RESULTS AND DISCUSSIONS

The FTIR spectrum of the chip is displayed in Fig. 2. Also, the magnified section in the measured range is illustrated. It is obviously seen the intensity of the transmittance in the apical and basal plane is lower than control PDMS. Significantly, the transmittance value in the apical plane is higher than in the basal plane which demonstrated adsorption of the employed cisplatin drug when moving through the channel in the PDMS surface. A similar discussion was also reported by other works [22, 27]. In addition, it was noticed that there is no change in the location of the peaks that emerged in the control, basal and apical planes of the PDMS chip.

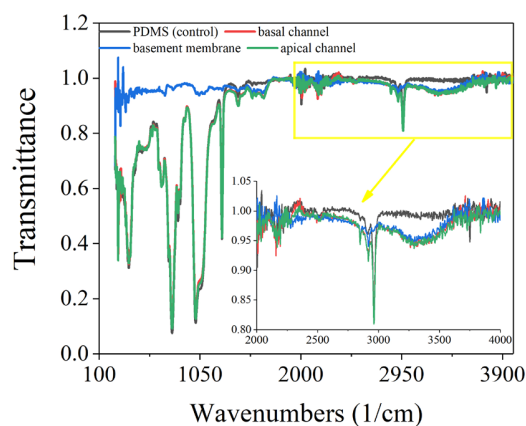


Figure 2. The FTIR spectrum of the basal, apical, and basement membrane of the fabricated chip using cisplatin drug after 7 days

The UV-Vis spectrophotometry of the control, basal and apical planes of the PDMS were depicted in Fig.

3. The change in the transmittance values of both channels is clearly clarified. The mean values of the transmittance were calculated 130%, 125%, 120% for control, apical, and basal PDMS planes in the 300-1100 nm wavelength range. It is seen that the minimum transparency was obtained in the basal plane which supported the FTIR results. To as best of our knowledge, there is a reverse relationship between transmittance and absorbance. So, this reduction exhibited adsorption of the drug onto the PDMS surface causing a reduction in the transmittance.

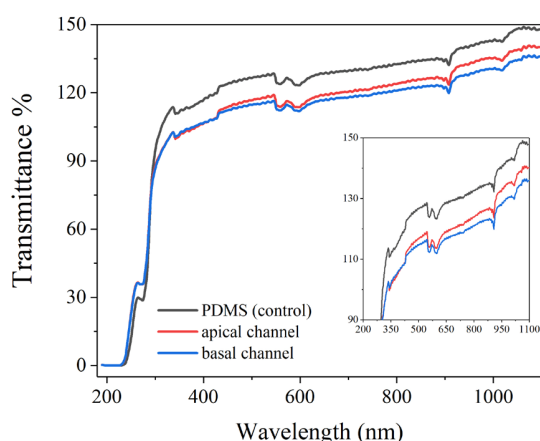


Figure 3. The UV-Vis spectrum of the fabricated chip using cisplatin as a model drug after 7 days

The adsorption of drugs in these types of experiments is highly related to the octanol/water concentration or partition coefficient or logP. The exact value of this parameter is known for most drugs but, should depend on experiment conditions^[28]. The parameter can alter from -4 to 5 value, that the higher value declared lipophilic properties of a drug. This increase in the logP value inferred higher adsorption in PDMS while regarding the works published by V. Novohradsky et al. the cisplatin has logP = -2.25^[29] demonstrates lower adsorption by the PDMS surfaces among other drugs. The adsorption in this system also probably due to the low amount of sample.

- MOHAMMADIGHAREHBAGH, Reza (0000-0002-0333-487X)
- TOMSUK, Özlem (0000-0001-7201-686X)
- GHORBANPOOR, Hamed (0000-0002-2665-8172)
- EBRAHIMI, Aliakbar (0000-0001-6437-7796)
- ABDULLAYEVA, Nuran (0000-0002-9945-6686)
- GASIMZADE, Nigar (0000-0001-6291-2337)
- KOÇ, Yücel (0000-0002-8301-5595)
- ÖZEL, Ceren (0000-0002-5648-3174)
- GHORBANI, Aynaz (0000-0001-5516-027X)

- DEMIR, Bahar (0000-0002-2444-8833)
- UYSAL, Onur (0000-0001-6800-5607)
- DOĞAN GUZEL, Fatma (0000-0001-7200-4615)
- EKER SARIBOYACI, Ayla (0000-0003-4536-9859)
- AVCI, Huseyin (0000-0002-2475-1963)

CONCLUSIONS

The cisplatin was successfully used as a model drug for understanding adsorption in the PDMS-based microfluidic system. The chip was fabricated using PMDS and PET-based membrane without a cell line for at least 7 days. Regarding the suitable analyses which were performed in this work, despite lower logP in the selected drug the adsorption was observed. In both UV-Vis and FTIR measurements, the obtained results were confirmed to reduce the intensity of the transmittance values of the basal, apical channels of the prepared chip. This research investigation highly recommended the usage of diverse routes introduced in this work for improvement in this challenge of obtaining reliable and repeatable results.

SUGGESTIONS FOR FURTHER WORK

The adsorption by the PDMS-based microfluidics leads to a serious change in the results obtained after the tests, then according to the works in the literature and our best knowledge, it is highly recommended to use a layer between the selected drug and PDMS surface or altered the surface properties of the used polymer material. This process has effectively and significantly behaved as a barrier for reduction or in some cases minimizing the adsorption to overcome the challenge and access close and reliable results for widespread of the platform in more applications.

ACKNOWLEDGEMENTS / SOURCES OF FUNDING

This study was supported by Turkish Scientific and Technological Council (TÜBİTAK 1004-Regenerative and Restorative Medicine Research and Applications) under the grant numbers of 20AG003 and 20AG031. The authors also gratefully acknowledge Eskişehir Osmangazi University for financial support (Scientific Research Foundation, grant number TOA-2022-2307).

The authors were declared thanks to Associate Prof. Dr. Murat Ardic for using UV-Vis spectrophotometry.

References

1. AVCI, H., et al., Recent advances in organ-on-a-chip technologies and future challenges: a review. *Turkish Journal of Chemistry*, 2018. 42(3): p. 587-610.
2. Zhang, Y.S., et al., Multisensor-integrated organs-on-chips platform for automated and continual in situ monitoring of organoid behaviors. *Proceedings of the National Academy of Sciences*, 2017. 114(12): p. E2293-E2302.
3. Shin, S.R., et al., Aptamer-based microfluidic electrochemical biosensor for monitoring cell-secreted trace cardiac biomarkers. *Analytical chemistry*, 2016. 88(20): p. 10019-10027.
4. Shin, S.R., et al., Label-Free and Regenerative Electrochemical Microfluidic Biosensors for Continual Monitoring of Cell Secretomes. *Advanced Science*, 2017. 4(5): p. 1600522.
5. Shaegh, S.A.M., et al., Rapid prototyping of whole-thermoplastic microfluidics with built-in microvalves using laser ablation and thermal fusion bonding. *Sensors and Actuators B: Chemical*, 2018. 255: p. 100-109.
6. Ceren, Ö., et al., Investigation of 3D Culture of Human Adipose Tissue-Derived Mesenchymal Stem Cells in a Microfluidic Platform. *Eskişehir Technical University Journal of Science and Technology A-Applied Sciences and Engineering*, 2021. 22(8th ULPAS-Special Issue 2021): p. 85-97.
7. Bayir Garbioglu, D., et al., Determination of therapeutic agents efficiencies of microsatellite instability high colon cancer cells in post-metastatic liver biochip modeling. *The FASEB Journal*, 2021. 35(9): p. e21834.
8. Serras, A.S., et al., A critical perspective on 3D liver models for drug metabolism and toxicology studies. *Frontiers in cell and developmental biology*, 2021. 9: p. 626805.
9. Dittrich, P.S. and A. Manz, Lab-on-a-chip: microfluidics in drug discovery. *Nature reviews Drug discovery*, 2006. 5(3): p. 210-218.
10. Korner, J.L., E.B. Stephenson, and K.S. Elvira, A bespoke microfluidic pharmacokinetic compartment model for drug absorption using artificial cell membranes. *Lab on a Chip*, 2020. 20(11): p. 1898-1906.
11. Forigua, A., et al., Recent advances in the design of microfluidic technologies for the manufacture of drug releasing particles. *Journal of Controlled Release*, 2021. 333: p. 258-268.
12. Ghorbanpoor, H., et al., A fully integrated rapid on-chip antibiotic susceptibility test—A case study for *Mycobacterium smegmatis*. *Sensors and Actuators A: Physical*, 2022. 339: p. 113515.
13. ÖZEL, C., et al., Investigation of Mesenchymal cells in the Microfluidic Cell Culture Device.
14. Didarian, R., et al. Investigation of Polar and Nonpolar Cyclotides Separation from Violet Extract Through Microfluidic Chip. 8. in *International Fiber and Polymer Research Symposium*. 2021.
15. Halldorsson, S., E. lucumi, r. Gomez-Sjoberg and r. MT Fleming. *Biosens. Bioelectron*, 2015. 63: p. 218-231.
16. Song, S. and K.Y. Lee, Polymers for microfluidic chips. *Macromolecular Research*, 2006. 14(2): p. 121-128.
17. Carter, S.-S.D., et al., PDMS leaching and its implications for on-chip studies focusing on bone regeneration applications. *Organs-on-a-Chip*, 2020. 2: p. 100004.
18. Sano, E., et al., Generation of Tetrafluoroethylene–Propylene Elastomer-Based Microfluidic Devices for Drug Toxicity and Metabolism Studies. *ACS omega*, 2021. 6(38): p. 24859-24865.
19. Özel, C., et al., Design Alginate Based Blends for Living Composite Fibers to Promote Wound Healing. *Eskişehir Technical University Journal of Science and Technology A-Applied Sciences and Engineering*, 2021. 22(SPECIAL ISSUE 2021).
20. Vickers, J.A., M.M. Caulum, and C.S. Henry, Generation of hydrophilic poly (dimethylsiloxane) for high-performance microchip electrophoresis. *Analytical chemistry*, 2006. 78(21): p. 7446-7452.
21. Schneider, M.H., Y. Tran, and P. Tabeling, Benzophenone absorption and diffusion in poly (dimethylsiloxane) and its role in graft photo-polymerization for surface modification. *Langmuir*, 2011. 27(3): p. 1232-1240.
22. Almutairi, Z., C.L. Ren, and L. Simon, Evaluation of polydimethylsiloxane (PDMS) surface modification approaches for microfluidic applications. *Colloids and Surfaces A: Physicochemical and Engineering Aspects*, 2012. 415: p. 406-412.
23. Rodrigues, P., et al., Critical Analysis of the Absorption/Adsorption of Small Molecules by Polydimethylsiloxane in Microfluidic Analytical Devices.
24. Ongaro, A.E., et al., Polylactic is a sustainable, low absorption, low

- autofluorescence alternative to other plastics for microfluidic and organ-on-chip applications. *Analytical chemistry*, 2020. 92(9): p. 6693-6701.
25. Soper, S.A., et al., Surface modification of polymer-based microfluidic devices. *Analytica Chimica Acta*, 2002. 470(1): p. 87-99.
 26. Han, X., et al., Polymer-based microfluidic devices: A comprehensive review on preparation and applications. *Polymer Engineering & Science*, 2022. 62(1): p. 3-24.
 27. Reese, W.M., et al., Facile macrocyclic polyphenol barrier coatings for PDMS microfluidic devices. *Advanced Functional Materials*, 2020. 30(48): p. 2001274.
 28. Raveendran, R., et al., Pt (IV) derivatives of cisplatin and oxaliplatin with phenylbutyrate axial ligands are potent cytotoxic agents that act by several mechanisms of action. *Chemical Science*, 2016. 7(3): p. 2381-2391.
 29. Novohradsky, V., et al., New insights into the molecular and epigenetic effects of antitumor Pt (IV)-valproic acid conjugates in human ovarian cancer cells. *Biochemical pharmacology*, 2015. 95(3): p. 133-144.