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Cervical dystonia: a disorder of the midbrain network for covert attentional orienting

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While the pathogenesis of cervical dystonia remains unknown, recent animal and clinical experimental studies have indicated its probable mechanisms. Abnormal temporal discrimination is a mediational endophenotype of cervical dystonia and informs new concepts of disease pathogenesis. Our hypothesis is that both abnormal temporal discrimination and cervical dystonia are due to a disorder of the midbrain network for covert attentional orienting caused by reduced gamma-aminobutyric acid (GABA) inhibition, resulting, in turn, from as yet undetermined, genetic mutations. Such disinhibition is (a) *subclinically* manifested by abnormal temporal discrimination due to prolonged duration firing of the visual sensory neurons in the superficial laminae of the superior colliculus and (b) *clinically* manifested by cervical dystonia due to disinhibited burst activity of the cephalomotor neurons of the intermediate and deep laminae of the superior colliculus. Abnormal temporal discrimination in unaffected first-degree relatives of patients with cervical dystonia represents a subclinical manifestation of defective GABA activity both within the superior colliculus and from the substantia nigra pars reticulata. A number of experiments are required to prove or disprove this hypothesis.

Keywords: cervical dystonia, temporal discrimination, covert attention, GABA, superior colliculus

INTRODUCTION

Cervical dystonia, characterized by sustained or intermittent neck muscle contractions causing abnormal head movements, is the most common form of adult-onset idiopathic isolated focal dystonia (AOIFD) and may be sporadic or familial (1). The pathogenesis of cervical dystonia is unknown (2). In patients with cervical dystonia, a number of neurophysiological and neuroimaging abnormalities have been described as endophenotypes implying a relationship to causal mechanisms [reviewed in Ref. (3)]. We have argued that many of these abnormalities are secondary endophenotypes resulting from disease manifestation; only mediational endophenotypes, including abnormal temporal discrimination, found in unaffected first-degree relatives as well as patients, are relevant to understanding disease pathogenesis (4, 5) (Table 1). The features of a mediational endophenotypes are (a) it is an expression

of a genetic mutation, necessarily present prior to disease onset, (b) it reflects disease susceptibility and is not altered by disease expression or severity, and (c) it is more penetrant than the phenotype (6). The attributes of abnormal temporal discrimination which indicate that it is a mediational endophenotype of cervical dystonia are listed in Table 1.

Mediational endophenotypes may elucidate mechanisms of disease pathogenesis not obvious from the phenotype. Shared pathological mechanisms for both disordered temporal discrimination and cervical dystonia have become evident over the past 10 years from highly sophisticated piecemeal investigations in diverse disciplines including primate and sub-primate neurophysiology, anatomy, chemistry, psychology, and clinical research. Using this evidence, we propose a unified theoretical model to explain the etiopathogenesis of cervical dystonia.

HYPOTHESIS

We postulate that cervical dystonia is a disorder of a midbrain network for covert attentional orienting involving both the sensory and motor laminae of the superior colliculus. The mediational

Abbreviations: AOIFD, adult-onset idiopathic isolated focal dystonia; DLSC, intermediate and deep laminae of the superior colliculus; SLSC, superficial laminae of the superior colliculus; SNpr, substantia nigra pars reticulata; TDT, temporal discrimination threshold; WFV, wide field vertical.

Table 1 | A summary of the evidence from experimental clinical and laboratory studies suggesting that an abnormal TDT is a mediational endophenotype of cervical dystonia.

Sensitivity: the TDT is abnormal in more than 80–90% of patients with various idiopathic AOIFD phenotypes, and sensitivity is 97% in cervical dystonia (7)

Specificity: the specificity of an abnormal TDT is 98–100% (5, 7)

Autosomal dominant transmission: abnormal TDTs show autosomal dominant transmission in multiplex AOIFD families (8) and in families of sporadic cervical dystonia (9). Unaffected obligate heterozygotes have abnormal TDTs (8)

Age-related and gender-related penetrance: in unaffected first-degree female relatives, an abnormal TDT is fully penetrant by 48 years of age. In male relatives, there is 40% penetrance after 25 years of age (5)

Putaminal enlargement: unaffected relatives (of patients with cervical dystonia) with abnormal TDTs have larger putaminal volumes by VBM than relatives with normal TDTs (8). Putaminal enlargement is found in patients with blepharospasm (10) and musicians with task-related dystonia (11)

Putaminal activation: unaffected first-degree relatives (of patients with cervical dystonia) with abnormal TDTs have reduced putaminal activation in a temporal discrimination test compared to relatives with normal TDTs (5). Perceptual certainty in a temporal discrimination task is associated with putaminal activation; the putamen is involved early in a temporal processing task (12)

AOIFD, adult-onset isolated focal dystonia; TDT, temporal discrimination threshold; VBM, voxel-based morphometry.

endophenotype, abnormal temporal discrimination and the phenotype, cervical dystonia, are caused by defective inhibition of sensory and motor neurons in the superior colliculus. This disinhibition is due to the effects of reduced gamma-aminobutyric acid (GABA) activity, both from the substantia nigra pars reticulata (SNpr) and GABAergic interneurons within the superior colliculus. Abnormal temporal discrimination, a subclinical marker of this disinhibition, results from prolonged duration firing of visual sensory neurons in the superficial layers of the superior colliculus (SLSC). The abnormal involuntary movement characteristic of cervical dystonia results from *subsequent secondary* disinhibition of cephalomotor neurons in the intermediate and deep layers of the superior colliculus (DLSC). A summary of the arguments deployed in support of this hypothesis is outlined in **Table 2**.

MIDBRAIN NETWORK FOR COVERT ATTENTIONAL ORIENTING

Attentional networks in the brain can be divided into: (i) an overt (top-down) attentional network utilizing volitional eye movements to orient the visual system and (ii) a covert (bottom-up) network, which shifts attention to locations by head movements with or without saccade initiation (13). The midbrain covert attentional network captures change in the environment and alerts the individual to a salient stimulus, which requires inspection and action that may be important for survival. A significant node in this network is the superior colliculus, a complex sensorimotor

Table 2 | An outline summary of some of the more important experimental evidence from clinical and animal studies supporting the hypothesis that both abnormal temporal discrimination and adult-onset isolated focal dystonia are due to a disorder of reflex covert orienting caused by defective GABAergic inhibition in the superior colliculus.

Hypothesis: pathological disinhibition in the midbrain network for covert attentional orienting due to deficient GABAergic activity causes

(a) *Subclinically*, abnormal temporal discrimination due to disordered visual processing in the SLSC

(b) *Clinically*, cervical dystonia due to disinhibited prolonged burst activity of cephalomotor neurons in the DLSC

Key observations in support of the hypothesis

Sensory aspects

(1) The midbrain network for covert attentional orienting is a primitive, highly conserved system for detecting salient environmental change, inspecting the change, and responding rapidly and appropriately

(2) Wide field visual sensory neurons in the SLSC, via the retinotectal pathway, detect environmental visual change and respond by transient discharges to the pre-motor neurons for saccade generation and head turning

(3) Focal inactivation of the SLSC causes loss of covert visual attention in the visual field represented by the inactivated part of the SLSC

(4) Inhibitory GABAergic activity (within the superior colliculus and from the SNpr) limits the duration of the transient response in visual sensory neurons in the SLSC

(5) A moving or sudden luminant visual stimulus elicits a time-locked electromyographic response in cervical muscles involved in ipsilateral head turning

Motor aspects

(1) Movement is initiated by the striatum through release from the tonic inhibition exerted by the SNpr. A core neurophysiological feature of dystonia is reduced inhibition at all levels of the CNS; the most probable cause is defective GABAergic inhibition

(2) The oculomotor and cephalomotor pre-motor neurons of the DLSC for saccade generation and head turning are tonically inhibited by the SNpr. Release from that inhibition allows prolonged burst discharges of premotor neurons

(3) Oculomotor premotor neurons are gated by omnipause neurons; the cephalomotor premotor neurons are not gated

(4) Stimulation of cephalomotor neurons in the DLSC causes ipsilateral head turning in monkeys via the tectospinal and tecto-reticulospinal fiber tracts terminating in the cervical cord

(5) A unilateral lesion of the SNpr in macaques causes a movement disorder resembling cervical dystonia. A further lesion in the superior colliculus abolishes/attenuates the movement disorder

This argument is expanded and fully referenced in the text. AOIFD, adult-onset isolated focal dystonia; DLSC, intermediate and deep laminae of the superior colliculus; SLSC, superficial laminae of the superior colliculus; SNpr, substantia nigra pars reticulata; TDT, temporal discrimination threshold.

brainstem structure, which acts as a sentinel for detecting sudden environmental change and responding to that change. The superior colliculus integrates multimodal sensory information from visual, auditory, and tactile sources; generates outputs for gaze, head, and arm movement; and sends priority signals to the substantia nigra pars compacta and the intralaminar nucleus of the thalamus (14).

SUPERIOR COLLICULUS – ANATOMY

The superior colliculus is a laminated structure with seven alternating gray and white layers (15). The SLSC, laminae I–III, receive direct visual input from the retina and indirect visual input from many cortical structures. The DLSC, laminae IV–VII, contain sensory cells responsive to multiple sources and premotor neurons with outputs to a number of structures described below. A significant intrinsic population of GABAergic inter-neurons (16) modulates the activity of both the visual sensory receptive cells in the SLSC and the premotor neurons in the DLSC (17, 18).

SUPERIOR COLLICULUS – INPUTS AND OUTPUTS

SENSORY INPUTS

The SLSC contain sensory cells, which receive visual inputs from the visual cortex and the retinotectal pathway. Magnocellular retinal ganglion cells convey responses to luminance change and movement in the visual field via the retinotectal pathway; about 10% of retinal ganglion cells project to the SLSC (19). It has been shown that the visual sensory cells of the SLSC also respond to chromatic S-cone stimuli, but at a longer latency than the faster retinotectal input; this longer latency reflects the retinogeniculo-cortical-collicular pathways (20–22). Among a variety of sensory neurons in the SLSC, wide field sensory neurons respond to salient environmental luminance change and movement; most neurons in the SLSC show an early 40–70 ms transient response to visual stimuli (23). Wide field sensory neurons in the SLSC have direct interlaminar connections with the premotor neurons in the DLSC considered to be the direct visuomotor pathway responsible for orienting responses (saccade generation and head turning) at extremely short latency (discussed below) (24, 25).

The SLSC is retinotopically organized; focal inactivation of a visual sensory area in the SLSC prevents macaques from detecting an odd-ball salient stimulus in the inactivated visual field and biases saccade selection away from that area (26, 27). Conversely, electrical stimulation of a focal area of the SLSC improves performance in a visual attention task in monkeys and favors saccades directed to the visual field corresponding to the stimulation site, indicating evidence for a role of the superior colliculus in the control of attention (28–30).

Although most experimental work relates to the visual system, deeper laminae of the superior colliculus receive other salient tactile and auditory inputs (15). Thus, both the SLSC and the DLSC receive multisensory inputs relating to environmental change, detect that change, and integrate the sensory information into a required appropriate motor output for immediate inspection or avoidance through motor outputs to the eye, neck, and arm muscles.

CHOLINERGIC INPUTS TO THE SUPERIOR COLLICULUS

Cholinergic inputs from the pedunculopontine tegmental nucleus to the intermediate layers of the superior colliculus primarily induce excitation of the motor output (31). When nicotine was applied *in vitro* to a rodent superior colliculus, with stimulation from the optic tract, sub-threshold excitatory post-synaptic potentials, recorded in the DLSC neurons, were depolarized and exhibited bursting responses (25). Nicotine injected into the superior colliculus of macaques had been shown to increase the frequency of express saccades with a shortened saccadic reaction time of approximately 100 ms (32). Recently, it has been demonstrated that the major function of the cholinergic input to the GABAergic neurons in the DLSC is excitatory and this may regulate the spike timing of the oculomotor and cephalomotor premotor neurons (33).

SUBSTANTIA NIGRA PARS RETICULARIS INPUT TO THE SUPERIOR COLLICULUS

Since mid-1980s, it was indicated that the striato-nigro-collicular pathway is one of the neural circuits through which the basal ganglia can influence both ocular and cephalic motricity. The SNpr provides a tonically active GABAergic inhibitory outflow from the striatum to the DLSC (34, 35). In rats, stimulation of the SNpr causes both short latency and short duration inhibition in the response of the tectospinal neurons to spontaneous and peripherally evoked discharges (36). Tectospinal neurons discharge vigorously in response to SNpr silencing by intranigral GABA. Intra-striatal injection of glutamate, which silences SNpr, causes the tectospinal cells to discharge. This striatally induced firing of the tectospinal tract is sensitive to a GABA antagonist, intranigral bicuculline (37). In monkeys, a substantial number of neurons in the medial SNpr are concerned with orienting behavior including postural and motor mechanisms during visually triggered arm movements, (38) whereas the lateral part is involved with oculomotor control (34).

MOTOR OUTPUT OF SUPERIOR COLLICULUS

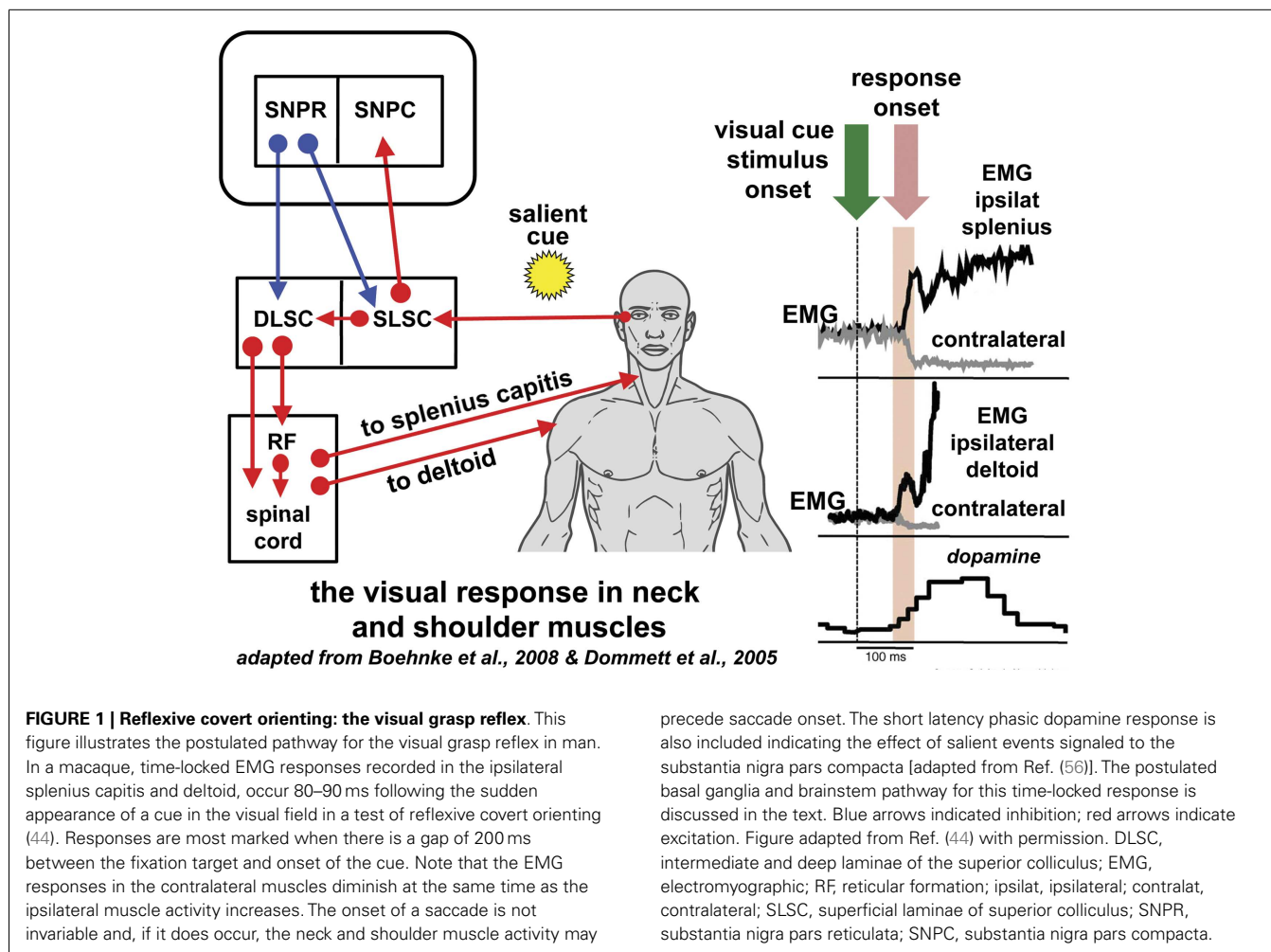
While the classical concept of the motor function of the superior colliculus is that of a saccade generator, recent studies in macaques have highlighted other motor functions.

SACCADE GENERATION (OCULOMOTOR PREMOTOR NEURONS)

Saccades are driven by brief, high-frequency, bursts from the DLSC to the brain stem burst generator (39). This oculomotor drive is potently inhibited by the pontine omnipause neurons. These tonically prevent the premature execution of eye movements until the DLSC output reaches a threshold, which is usually associated with the high-frequency burst. Low-frequency DLSC activity does not influence eye movement because of this gating by the omnipause neurons (40). Visually guided saccades have latencies of 150–350 ms; both humans and monkeys can generate express saccades with latencies of 70–100 ms in monkeys and 80–120 ms in humans (41).

HEAD-TURN GENERATION (CEPHALOMOTOR PREMOTOR NEURONS)

Initial studies in cats demonstrated that stimulation of the DLSC elicited early head movements without saccades (42) (Figure 1).



In macaques, a number of experiments in the last 15 years have revealed that the DLSC contains premotor neurons solely concerned with head-turn generation; this motor pathway has not been examined in human. In macaques, the sudden presentation of a bright target in the temporal visual field stimulates a discharge of DLSC premotor neurons and a time-locked burst of activity in ipsilateral neck muscles with an extremely short latency (55–95 ms after the visual stimulus), regardless of the ensuing saccadic reaction time (43, 44) (**Figure 1**). This has been termed the “*visual grasp reflex*” (44). When a gaze shift (head plus eye movement) does follow, the neck muscle electromyographic response increases significantly prior to the higher velocity head movement associated with the gaze shift reflecting a correlation with movement preparation (45, 46). Low-frequency stimulation of DLSC neurons generates head turns without saccades in macaques and, in one study, 26% of DLSC neurons sampled were head-only neurons (42, 45, 47–50). In contradistinction to the oculomotor pathway, an absent or significantly weaker gating mechanism on the head pathway permits the generation of head-only movements that can precede gaze shifts (48, 51–53). Monkeys trained to make head-only movements to reward (without gaze shifts) showed DLSC neuronal bursts 20 ms before neck EMG activity (46); the classical

DLSC neurons involved in gaze shift (head and eye movement) are not active in such head-only movements (54). In the sub-human primate, the representation of the position of head on body in the superior colliculus is used to compute oculomotor movement in saccade execution (55).

UPPER LIMB MOTOR CONTROL FROM THE SUPERIOR COLLICULUS

The primate superior colliculus also contains neurons which, when stimulated, produce arm movements independent of gaze shift (57–59); these neurons affect the shoulder and proximal arm muscles (57) and even hand muscles (60). In humans performing a reaching task with one arm following the sudden appearance of a visual target stimulus, there was a time-locked response in shoulder muscles at a latency of <100 ms indicating a rapid neural pathway linking visual input to arm motor out; this seemed most effective during reflexive movement tasks (61). The DLSC is the primary source of this short latency response.

OTHER SUPERIOR COLLICULUS OUTPUT TARGETS

These targets include substantia nigra pars compacta, SNpr, and the intralaminar nucleus of the thalamus (14, 62).

ANATOMY OF THE PATHWAYS FOR THE HEAD-TURN RESPONSE TO SUPERIOR COLLICULUS STIMULATION

In macaques, anatomical studies have shown that projections to downstream oculomotor and neck muscle structures originate in separate laminae of the stratum griseum intermediale in the DLSC (63, 64) (**Figure 2**). The *tectospinal tract* is a very conserved structure present in all mammals, although it is better developed in predatory than in less predatory species (65, 66). It originates in the DLSC and terminates almost exclusively on inter-neurons in the upper cervical cord whose motor neurons innervate neck muscles, and in the lower cervical enlargement innervating the forearm and hand muscles (65, 67, 68). Another important tectospinal target is the propriospinal neurons in the C3–C4 segments of the cervical cord, which govern target reaching of the forelimb (69). The *tecto-reticulospinal tract* also originates in the DLSC of the SC, involves the tegmental and pontine reticular formation and also ends in the upper cervical spinal cord (67, 68, 70–73) contacting, at least in part, the motor neurons of head extensor muscles. The cells of origin of the tectospinal and the tecto-reticulospinal tracts overlap to some degree in the DLSC (73). Electrical stimulation of the DLSC and the dorsal tegmentum in the cat activates reticulospinal neurons (74) that in turn activate motor neurons of the flexor and extensor muscles of the hind and forelimb (75). In the macaque, tecto-reticulospinal pathways to the neck and hand muscles have been demonstrated (76); neck muscle EMG activity following a visual stimulus is conveyed by the tecto-reticulospinal pathways (77). In cats, this pathway displays bursts of activity time-locked to visual target presentation (78). Similar burst discharges in response to visual stimuli have been found in tecto-reticular neurons in rats (79).

VISUAL RESPONSE IN THE SUPERIOR COLLICULUS

Among others, the Okazaki laboratory of Tadashi Isa has been extremely active in examining the microcircuitry of the superior colliculus (**Figure 3**). The predominant cell type in the superficial layer of the superior colliculus is the wide field vertical (WFV) cell, mainly in the stratum opticum of the SLSC; these cells have characteristic dendritic trees and are luminance and motion sensitive (**Figure 3**). In response to a visual stimulus, most of the neurons in the SLSC exhibit transient “ON” responses within 50 ms of the stimulus onset (80). With a persistent visual stimulus, most of these cells enter a pause phase and then only discharge again when the visual stimulus is switched off. This sequence “ON–PAUSE–OFF” (**Figure 3**) in the WFV neurons is characteristic of the response in the superior colliculus to the switching on and off of the visual stimulus or the response to movement. This is the mechanism by which the superior colliculus acts as a detector of salient stimuli (**Figure 3**).

GABAergic INHIBITION IN THE SUPERIOR COLLICULUS AND NORMAL TEMPORAL DISCRIMINATION

Application of a GABA_B antagonist to an electrode recording SLSC neuronal responses to optic fiber inputs in mice prolongs the “ON” burst duration. With blocking of both GABA_A and GABA_B receptors, there is excessive and prolonged burst activity in both the “ON” and “OFF” phases with loss or attenuation of the normal “PAUSE” phase (80) (**Figure 3**).

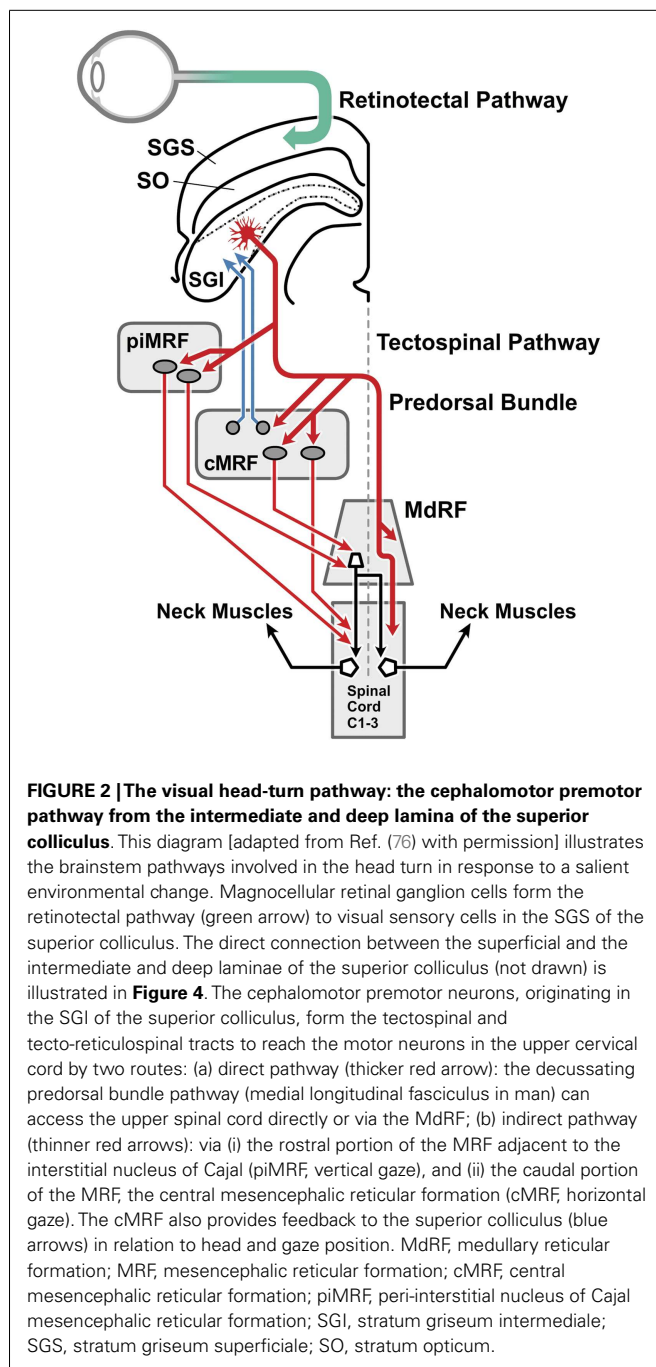


FIGURE 2 | The visual head-turn pathway: the cephalomotor premotor pathway from the intermediate and deep laminae of the superior colliculus.

This diagram [adapted from Ref. (76) with permission] illustrates the brainstem pathways involved in the head turn in response to a salient environmental change. Magnocellular retinal ganglion cells form the retinotectal pathway (green arrow) to visual sensory cells in the SGS of the superior colliculus. The direct connection between the superficial and the intermediate and deep laminae of the superior colliculus (not drawn) is illustrated in **Figure 4**. The cephalomotor premotor neurons, originating in the SGI of the superior colliculus, form the tectospinal and tecto-reticulospinal tracts to reach the motor neurons in the upper cervical cord by two routes: (a) direct pathway (thicker red arrow): the decussating predorsal bundle pathway (medial longitudinal fasciculus in man) can access the upper spinal cord directly or via the MdRF; (b) indirect pathway (thinner red arrows): via (i) the rostral portion of the MRF adjacent to the interstitial nucleus of Cajal (piMRF; vertical gaze), and (ii) the caudal portion of the MRF, the central mesencephalic reticular formation (cMRF; horizontal gaze). The cMRF also provides feedback to the superior colliculus (blue arrows) in relation to head and gaze position. MdRF, medullary reticular formation; MRF, mesencephalic reticular formation; cMRF, central mesencephalic reticular formation; piMRF, peri-interstitial nucleus of Cajal mesencephalic reticular formation; SGI, stratum griseum intermediale; SGS, stratum griseum superficiale; SO, stratum opticum.

GABAergic SLSC inter-neurons, following a visual stimulus, activate GABA_B receptors and limit the duration of both “ON” and “OFF” responses (81, 82). GABAergic inhibition of activity in excitatory neurons prevents unnecessarily prolonged burst activities in the SLSC local circuit, which affects the burst firing in the DLSC through direct signal transmission from the superficial to the deeper layer of the SC (24, 81, 83, 84). This mechanism may endow the SLSC with the ability to detect the appearance, but not the persistent presentation, of an object in the visual field, allowing the SLSC to function as a saliency detector, which might be modulated by cholinergic inputs (85).

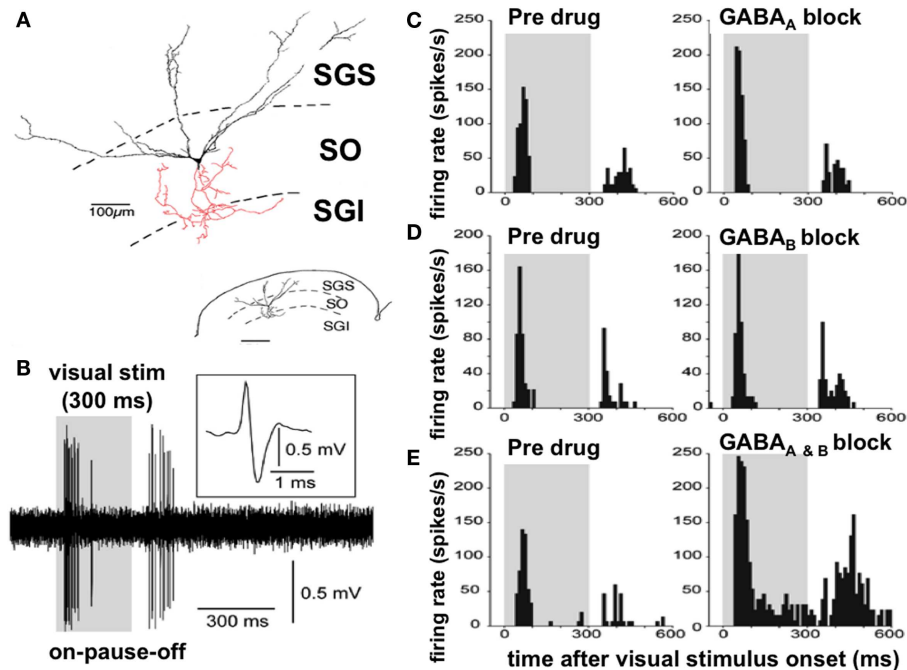


FIGURE 3 | Discharge properties of the wide field vertical cells in the superficial lamina of the superior colliculus: GABA_A and GABA_B receptors cooperatively shape transient “ON” responses. (A) A typical WFV cell is illustrated in the SGS and SO; both form part of the superficial laminae of the superior colliculus, contacting a premotor neuron (in red) in the SGI (part of the DLSC). Inset: small-scale drawing of same section. (B) Typical ON–PAUSE–OFF pattern response in a WFV cell to a static visual stimulus lasting 300 ms. Five traces are superimposed. The inset shows the expanded trace of a single spike with a positive–negative sequence due to close apposition of the recording electrode to the cell. Gray-shaded areas in the panels indicate duration of visual stimulus presentation. (C) Local

application of the GABA_A receptor antagonist significantly increased the peak-firing rate but did not affect “ON” response duration. (D) Local application of the GABA_B receptor antagonist significantly prolonged the “ON” response duration, but did not affect the peak-firing rate. (E) Local application of both GABA_A and GABA_B receptor antagonists significantly increased the peak-firing rate and prolonged the “ON” response duration. Figure adapted from an original study kindly provided by Professor Tadashi Isa, from Ref. (80) with permission. GABA, gamma-aminobutyric acid; WFV, wide field vertical; SGS, stratum griseum superficiale; SGI, stratum griseum intermediale; SO, stratum opticum; DLSC, intermediate and deep laminae of the superior colliculus.

MECHANISM OF ABNORMAL TEMPORAL DISCRIMINATION IN DYSTONIA

Excessive firing in both the “ON” and “OFF” phases of the visual response in the SLSC and the consequent reduction of the “PAUSE” phase, caused by loss of GABA inhibition, results in increased discharge duration and loss of sharpness of the offset of the neuronal response (Figure 3). The effect of the increased duration of both the “ON” and “OFF” phases of the discharge in the WFV cells would increase the duration of the inter-stimulus interval before two asynchronous stimuli would seem, to the observer, as being separate in time. Thus, we postulate that impaired GABAergic activity in the SLSC would result in an inability to discriminate two visual stimuli in short succession and result in abnormal (prolonged) temporal discrimination.

CONTROL OF CEPHALOMOTOR OUTPUT FROM THE SUPERIOR COLLICULUS

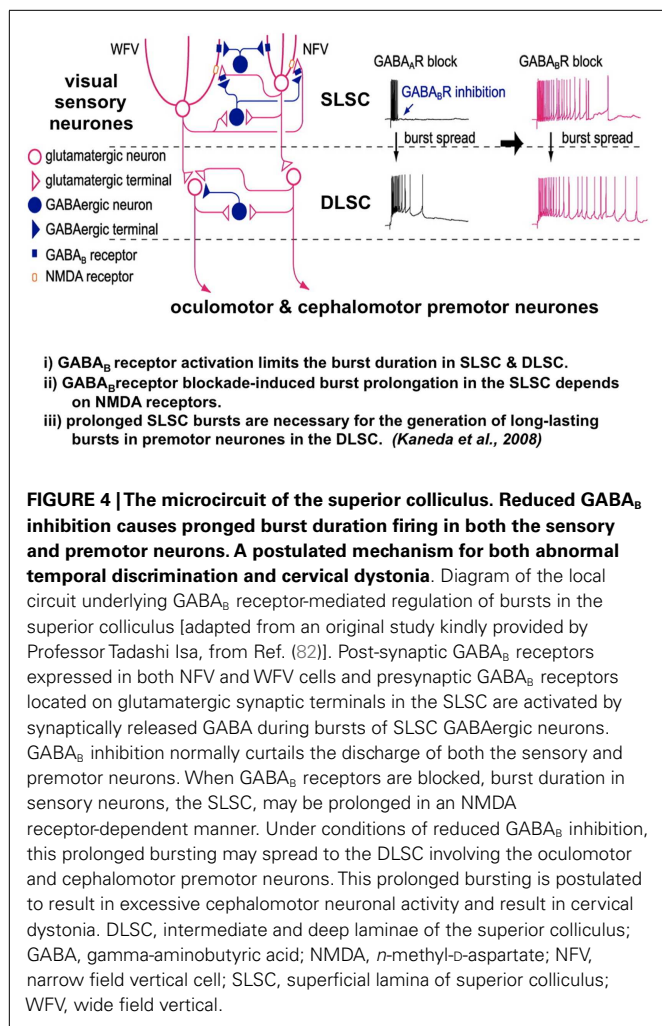
DISINHIBITION OF THE SUPERIOR COLLICULUS AND CEPHALOMOTOR NEURONS

The vertical interlaminar connection from the SLSC to the DLSC requires disinhibition from the GABAergic system for the signal to transmit in the pathway (24, 86) (Figure 4). The

relationship between GABAergic inhibition from the SNpr and the intrinsic GABAergic inter-neurons of the superior colliculus has been examined by Isa and his colleagues. Neuroanatomical studies have shown that SNpr input to the superior colliculus contacts both glutaminergic motor cells in the DLSC and GABAergic inter-neurons. GABA_B receptor activation limits both the burst duration of the visual cells in the SLSC and the premotor neurons in the DLSC. Importantly, prolonged bursts in the visual sensory cells in the SLSC, caused by GABA_B receptor blockade, are necessary for the generation of long-lasting bursts in the premotor neurons in the DLSC (Figure 4) (81, 82).

DISINHIBITION OF THE SUPERIOR COLLICULUS IN THE GENERATION OF EXPERIMENTAL CERVICAL DYSTONIA IN PRIMATES

In monkeys, experimental evidence has shown over the last 25 years that (a) unilateral SNpr lesions induced head turning/tilting to the opposite side (38), (b) muscimol (a GABA agonist) injection into the left SNpr induced a severe contralateral torticollis in 10/12 injections (87), and (c) intranigral muscimol induced a contralaterally directed torticollis from central and posterior sites of the SNpr in 3/3 monkeys (88).



Further to their initial experiments, Karen Gale's group examined the effect of blocking the DLSC (89). In four macaques pretreating the DLSC with muscimol, before lesioning the SNpr in the same animal, prevented or attenuated the development of the experimentally induced cervical dystonia (laterocollis in three animals and torticollis in one). Thus, reduction of the inhibitory GABAergic input from the SNpr to the superior colliculus in macaques causes a movement disorder resembling cervical dystonia. Blocking the motor output from DLSC by pretreating the superior colliculus prevents the movement disorder (89).

DISINHIBITION OF THE SUPERIOR COLLICULUS CAUSES BOTH DYSTONIA AND ABNORMAL TEMPORAL DISCRIMINATION

A core neurophysiological feature of dystonia is reduced inhibition at all levels of the CNS; the most probable cause of this is defective GABAergic inhibition (90). It thus might be reasonably postulated that the link between abnormal temporal discrimination and cervical dystonia is reduced GABAergic inhibition affecting the superior colliculus. Such impaired inhibition results in increased excitability and abnormal burst firing of visual sensory neurons

in the SLSC; the subsequent increased excitability of the cephalomotor cells of the DLSC causes experimental cervical dystonia. Thus, it would be reasonable to postulate that abnormal temporal discrimination in both patients with cervical dystonia and their unaffected first-degree relatives is a subclinical marker of defective GABA inhibition in the superior colliculus.

CEREBELLAR LESIONS, DYSTONIA, AND TEMPORAL DISCRIMINATION

The role of cerebellar lesions in the genesis of secondary dystonia including cervical dystonia and blepharospasm has been recently highlighted (91, 92). Temporal discrimination is abnormal in cerebellar degeneration (93). It is likely that inputs from the cerebellum modulate the intrinsic microcircuit of the superior colliculus but the mechanisms have not been examined in animal models.

CLINICAL EVIDENCE OF ABNORMAL HEAD-TURN CONTROL IN CERVICAL DYSTONIA

In the last few years, a number of investigators have demonstrated abnormalities in head control and head turning in cervical dystonia. Given the difficulties with rapid alternating movements seen in dystonic muscles, this might not be unexpected. However, foveation delays, head on trunk bradykinesia, and truncal bradykinesia, exceeding that found in Parkinson's disease, are seen in cervical dystonia (94). Oculomotor function is normal in cervical dystonia but gaze control (combined head and eye movement) is defective because abnormal head turning interferes with saccadic function. A number of investigators have implicated the midbrain interstitial nucleus of Cajal in this defective mechanism (95, 96) but the preponderance of the experimental evidence described above indicates that defective inhibition within the superior colliculus and from the SNpr, disrupting the midbrain attentional orienting network, is the more probable mechanism of disease pathogenesis in cervical dystonia.

OTHER LATE-ONSET FOCAL DYSTONIA PHENOTYPES

Most work quoted in this study has addressed cervical dystonia and the cephalomotor system. However, it is likely that all the adult-onset focal dystonia phenotypes share similar etiological factors and pathogenic mechanisms (97). Anatomical studies indicate that premotor neurons originating in the DLSC form part of the tecto-reticulospinal and tectospinal tracts, and may affect facial, bulbar, forearm, and hand muscles, and thus be implicated in the genesis of cranial and focal hand dystonias (69, 76, 98–101) (Figure 2). Certainly abnormal temporal discrimination is found in all these focal dystonia phenotypes suggesting that similar pathogenic mechanisms may be involved (7, 102, 103).

CONCLUSION

We postulate that cervical dystonia and abnormal temporal discrimination are both due to a disorder of the midbrain network for attentional orienting, caused by impaired GABAergic mechanisms of inhibition of sensorimotor processing within the superior colliculus. This deficiency in GABAergic activity results in abnormal burst firing in the visual sensory cells in the superficial laminae of the superior colliculus, and thus the abnormal temporal discrimination found in patients with cervical dystonia and their

unaffected first-degree relatives. Prolonged duration firing of the visual sensory neurons is a necessary prior abnormality for the development of the hyperexcitability of the premotor neurons in the DLSC. These hyperexcitable DLSC premotor neurons, via the tecto-reticulospinal and tectospinal pathways, stimulate motor neurons in the upper cervical spinal cord resulting in the abnormal, jerky head spasms characteristic of cervical dystonia.

PROBLEMS AND FUTURE SOLUTIONS

Clearly, this hypothesis does not explain the cause of the defective GABAergic inhibition in the superior colliculus. It is likely that processes upstream in the basal ganglia in relation to dopamine processing and disordered D1/D2 receptor availability are responsible [reviewed in Ref. (104)].

In order to advance this hypothesis, a number of experiments are required. These experiments include: (a) examination of the visual response to salient stimuli in the neck muscles in patients with cervical dystonia and in their unaffected first-degree relatives with abnormal and normal temporal discrimination. (b) Examination of the fMRI response to looming stimuli in the superior colliculus in both patients with cervical dystonia and their unaffected relatives with and without abnormal temporal stimulation. (c) Magnetic resonance spectroscopic examination studies are needed for levels of GABA in the superior colliculus in patients with cervical dystonia and their relatives with and without abnormal temporal discrimination.

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REFERENCES

- Albanese A, Bhatia K, Bressman SB, DeLong MR, Fahn S, Fung VS, et al. Phenomenology and classification of dystonia: a consensus update. *Mov Disord* (2013) **28**:863–73. doi:10.1002/mds.25475
- Jinnah HA, Berardelli A, Comella C, Defazio G, DeLong MR, Factor S, et al. The focal dystonias: current views and challenges for future research. *Mov Disord* (2013) **28**:926–43. doi:10.1002/mds.25567
- Stamelou M, Edwards MJ, Hallett M, Bhatia KP. The non-motor syndrome of primary dystonia: clinical and pathophysiological implications. *Brain* (2012) **135**:1668–81. doi:10.1093/brain/awr224
- Hutchinson M, Kimmich O, Molloy A, Whelan R, Molloy F, Lynch T, et al. The endophenotype and the phenotype: temporal discrimination and adult-onset dystonia. *Mov Disord* (2013) **28**:1766–74. doi:10.1002/mds.25676
- Kimmich O, Molloy A, Whelan R, Williams L, Bradley D, Balsters J, et al. Temporal discrimination, a cervical dystonia endophenotype: penetrance and functional correlates. *Mov Disord* (2014). doi:10.1002/mds.25822
- Kendler KS, Neale MC. Endophenotype: a conceptual analysis. *Mol Psychiatry* (2010) **15**:789–97. doi:10.1038/mp.2010.8
- Bradley D, Whelan R, Kimmich O, O'Riordan S, Mulrooney N, Brady P, et al. Temporal discrimination in adult-onset primary torsion dystonia: an analysis by task type and by dystonia phenotype. *J Neurol* (2012) **259**:77–82. doi:10.1007/s00415-011-6125-7
- Bradley D, Whelan R, Walsh R, Reilly RB, Hutchinson S, Molloy F, et al. Temporal discrimination threshold as an endophenotype in adult-onset primary torsion dystonia. *Brain* (2009) **132**:2327–35. doi:10.1093/brain/awp156
- Kimmich O, Bradley D, Whelan R, Mulrooney N, Reilly RB, Hutchinson S, et al. Sporadic adult onset primary torsion dystonia is a genetic disorder by the temporal discrimination test. *Brain* (2011) **134**:2656–63. doi:10.1093/brain/awr194
- Etgen T, Mührlau M, Gaser C, Sander D. Bilateral grey-matter increase in the putamen in primary blepharospasm. *J Neurol Neurosurg Psychiatry* (2006) **77**:1017–20. doi:10.1136/jnnp.2005.087148
- Granert O, Peller M, Jabusch HC, Altenmüller E, Siebner HR. Sensorimotor skills and focal dystonia are linked to putaminal grey-matter volume in pianists. *J Neurol Neurosurg Psychiatry* (2011) **82**:1225–31. doi:10.1136/jnnp.2011.245811
- Pastor MA, Macaluso E, Day BL, Frackowiak RSJ. Putaminal activity is related to perceptual certainty. *Neuroimage* (2008) **41**:123–9. doi:10.1016/j.neuroimage.2008.01.034
- Fecteau JH, Munoz DP. Saliency, relevance, and firing: a priority map for target selection. *Trends Cogn Sci* (2006) **10**:382–90. doi:10.1016/j.tics.2006.06.011
- Redgrave P, Coizet V, Comoli E, McHaffie JG, Leriche M, Vautrelle N, et al. Interactions between the midbrain superior colliculus and the basal ganglia. *Front Neuroanat* (2010) **4**:132. doi:10.3389/fnana.2010.00132
- White BJ, Munoz DP. The superior colliculus. In: Liversedge S, Gilchrist I, Everling S editors. *The Oxford Handbook of Eye Movements*. Oxford: Oxford University Press (2011). p. 195–213.
- Okada Y. Distribution of gamma-aminobutyric acid (GABA) in the layers of the superior colliculus of the rabbit. *Brain Res* (1974) **75**:362–6. doi:10.1016/0006-8993(74)90762-8
- Endo T, Yanagawa Y, Obata K, Isa T. Characteristics of GABAergic neurons in the superficial superior colliculus in mice. *Neurosci Lett* (2003) **346**:81–4. doi:10.1016/S0304-3940(03)00570-6
- Sooksawat T, Isa K, Behan M, Yanagawa Y, Isa T. Organization of GABAergic inhibition in the motor output layer of the superior colliculus. *Eur J Neurosci* (2011) **33**:421–32.
- Perry VH, Cowey A. Retinal ganglion cells that project to the superior colliculus and pretectum in the macaque monkey. *Neuroscience* (1984) **12**:1125–37. doi:10.1016/0306-4522(84)90006-X
- Sumner P, Adamjee T, Mollon JD. Signals invisible to the collicular and magnocellular pathways can capture visual attention. *Curr Biol* (2002) **12**:1312–6. doi:10.1016/S0960-9822(02)01020-5
- White BJ, Boehnke SE, Marino RA, Itti L, Munoz DP. Color-related signals in the primate superior colliculus. *J Neurosci* (2009) **29**:12159–66. doi:10.1523/JNEUROSCI.1986-09.2009
- Bompas A, Sumner P. Oculomotor distraction by signals invisible to the retinotectal and magnocellular pathways. *J Neurophysiol* (2009) **102**:2387–95. doi:10.1152/jn.00359.2009
- Isa T. Intrinsic processing in the mammalian superior colliculus. *Curr Opin Neurobiol* (2002) **12**:668–77. doi:10.1016/S0959-4388(02)00387-2
- Isa T, Endo T, Saito Y. The visuo-motor pathway in the local circuit of the rat superior colliculus. *J Neuroscience* (1998) **18**:8496–504.
- Isa T, Hall WC. Exploring the superior colliculus in vitro. *J Neurophysiol* (2009) **102**:2581–93. doi:10.1152/jn.00498.2009
- McPeck RM, Keller EL. Deficits in saccade target selection after inactivation of superior colliculus. *Nat Neurosci* (2004) **7**:757–63. doi:10.1038/nn1269
- Zenon A, Krauzlis RJ. Attention deficit without cortical neuronal deficits. *Nature* (2012) **489**:434–7. doi:10.1038/nature11497
- Carello CD, Krauzlis RJ. Manipulating intent: evidence for a causal role of the superior colliculus in target selection. *Neuron* (2004) **43**:575–83. doi:10.1016/j.neuron.2004.07.026
- Cavanaugh J, Wurtz RH. Subcortical modulation of attention counters change blindness. *J Neurosci* (2004) **24**:11236–43. doi:10.1523/JNEUROSCI.3724-04.2004
- Müller JR, Philiastrides MG, Newsome WT. Microstimulation of the superior colliculus focuses attention without moving the eyes. *Proc Natl Acad Sci U S A* (2005) **102**:524–9. doi:10.1073/pnas.0408311101
- Sooksawat T, Isa K, Isa T. Cholinergic responses in crossed tecto-reticular neurons of rat superior colliculus. *J Neurophysiol* (2008) **100**:2702–11. doi:10.1152/jn.90723.2008
- Aizawa H, Kobayashi Y, Yamamoto M, Isa T. Injection of nicotine into the superior colliculus facilitates occurrence of express saccades in monkeys. *J Neurophysiol* (1999) **82**:1642–6.

33. Sooksawat T, Yanagawa Y, Isa T. Cholinergic responses in GABAergic and non-GABAergic neurons in the intermediate gray layer of mouse superior colliculus. *Eur J Neurosci* (2012) **36**:2440–51. doi:10.1111/j.1460-9568.2012.08169.x
34. Hikosaka O, Wurtz RH. Visual and oculomotor functions of monkey substantia nigra pars reticulata. IV. Relation of substantia nigra to superior colliculus. *J Neurophysiol* (1983) **49**:1285–301.
35. Chevalier G, Deniau JM. Disinhibition as a basic process in the expression of striatal functions. *Trends Neurosci* (1990) **13**:277–80. doi:10.1016/0166-2236(90)90109-N
36. Chevalier G, Vacher S, Deniau JM. Inhibitory nigral influence on tectospinal neurons, a possible implication of basal ganglia in orienting behavior. *Exp Brain Res* (1984) **53**:320–6. doi:10.1007/BF00238161
37. Chevalier G, Vacher S, Deniau JM, Desban M. Disinhibition as a basic process in the expression of striatal functions. I. The striato-nigral influence on tecto-spinal/tecto-diencephalic neurons. *Brain Res* (1985) **334**:215–26. doi:10.1016/0006-8993(85)90213-6
38. Lestienne FG, Thullier F. Performance of visually triggered wrist movements task in monkey: an application of information theory to evaluate deficits following unilateral substantia nigra pars reticulata lesion. *Neurosci Lett* (1998) **251**:177–80. doi:10.1016/S0304-3940(98)00522-9
39. Scudder CA, Kaneko CS, Fuchs AF. The brainstem burst generator for saccadic eye movements: a modern synthesis. *Exp Brain Res* (2002) **142**:439–62. doi:10.1007/s00221-001-0912-9
40. Munoz DP, Dorris MC, Paré M, Everling S. On your mark, get set: brainstem circuitry underlying saccadic initiation. *Can J Physiol Pharmacol* (2000) **78**:934–44. doi:10.1139/y00-062
41. Fischer B, Weber H, Biscaldi M, Aiple F, Otto P, Stuhr V. Separate populations of visually guided saccades in humans: reaction times and amplitudes. *Exp Brain Res* (1993) **92**:528–41. doi:10.1007/BF00229043
42. Pélissier D, Goffart L, Guillaume A, Catz N, Raboyeau G. Early head movements elicited by visual stimuli or collicular electrical stimulation in the cat. *Vision Res* (2001) **41**:3283–94. doi:10.1016/S0042-6989(01)00224-3
43. Corneil BD, Olivier E, Munoz DP. Visual responses on neck muscles reveal selective gating that prevents express saccades. *Neuron* (2004) **42**:831–41. doi:10.1016/S0896-6273(04)00267-3
44. Boehnke SE, Munoz DP. On the importance of the transient visual response in the superior colliculus. *Curr Opin Neurobiol* (2008) **18**:544–51. doi:10.1016/j.conb.2008.11.004
45. Corneil BD, Munoz DP, Olivier E. Priming of head premotor circuits during oculomotor preparation. *J Neurophysiol* (2007) **97**:701–14. doi:10.1152/jn.00670.2006
46. Rezvani S, Corneil BD. Recruitment of a head-turning synergy by low-frequency activity in the primate superior colliculus. *J Neurophysiol* (2008) **100**:397–411. doi:10.1152/jn.90223.2008
47. Corneil BD, Olivier E, Munoz DP. Neck muscle responses to stimulation of monkey superior colliculus. I. Topography and manipulation of stimulation parameters. *J Neurophysiol* (2002) **88**:1980–99.
48. Corneil BD, Olivier E, Munoz DP. Neck muscle responses to stimulation of monkey superior colliculus. II. Gaze shift initiation and volitional head movements. *J Neurophysiol* (2002) **88**:2000–18.
49. Isa T, Sasaki S. Brainstem control of head movements during orienting: organization of the premotor circuits. *Prog Neurobiol* (2002) **66**:205–41. doi:10.1016/S0301-0082(02)00006-0
50. Corneil BD, Munoz DP, Chapman BB, Admans T, Cushing SL. Neuromuscular consequences of reflexive covert orienting. *Nat Neurosci* (2008) **11**:13–5. doi:10.1038/nn2023
51. Guitton D, Munoz DP, Galiana HL. Gaze control in the cat: studies and modeling of the coupling between orienting eye and head movements in different behavioral tasks. *J Neurophysiol* (1990) **64**:509–31.
52. Gandhi NJ, Sparks DL. Dissociation of eye and head components of gaze shifts by stimulation of the omnipause neuron region. *J Neurophysiol* (2007) **98**:360–73. doi:10.1152/jn.00252.2007
53. Grantyn A, Kuze B, Brandt AM, Thomas MA, Quenech' du N. Direct projections of omnipause neurons to reticulospinal neurons: a double-labeling light microscopic study in the cat. *J Comp Neurol* (2010) **518**:4792–812. doi:10.1002/cne.22488
54. Walton MM, Bechara B, Gandhi NJ. Role of the primate superior colliculus in the control of head movements. *J Neurophysiol* (2007) **98**:2022–37. doi:10.1152/jn.00258.2007
55. Nagy B, Corneil BD. Representation of horizontal head-on-body position in the primate superior colliculus. *J Neurophysiol* (2010) **103**:858–74. doi:10.1152/jn.00099.2009
56. Redgrave P, Gurney K. The short-latency dopamine signal: a role in discovering novel actions? *Nat Rev Neurosci* (2006) **7**:967–75. doi:10.1038/nrn2022
57. Stuphorn V, Bauswein E, Hoffmann KP. Neurons in the primate superior colliculus coding for arm movements in gaze-related coordinates. *J Neurophysiol* (2000) **83**:1283–99.
58. Reyes-Puerta V, Philipp R, Lindner W, Hoffmann KP. Role of the rostral superior colliculus in gaze anchoring during reach movements. *J Neurophysiol* (2010) **103**:3153–66. doi:10.1152/jn.00989.2009
59. Reyes-Puerta V, Philipp R, Lindner W, Hoffmann KP. Neuronal activity in the superior colliculus related to saccade initiation during coordinated gaze-reach movements. *Eur J Neurosci* (2011) **34**:1966–82. doi:10.1111/j.1460-9568.2011.07911.x
60. Nagy A, Kruse W, Rottmann S, Dannenberg S, Hoffmann KP. Somatosensory-motor neuronal activity in the superior colliculus of the primate. *Neuron* (2006) **52**:525–34. doi:10.1016/j.neuron.2006.08.010
61. Pruszynski JA, King GL, Boisse L, Scott SH, Flanagan JR, Munoz DP. Stimulus-locked responses on human arm muscles reveal a rapid neural pathway linking visual input to arm motor output. *Eur J Neurosci* (2010) **32**:1049–57. doi:10.1111/j.1460-9568.2010.07380.x
62. Dommert E, Coizet V, Blaha CD, Martindale J, Lefebvre V, Walton N, et al. How visual stimuli activate dopaminergic neurons at short latency. *Science* (2005) **307**:1476–9. doi:10.1126/science.1107026
63. May PJ, Porter JD. The laminar distribution of macaque tectobulbar and tectospinal neurons. *Vis Neurosci* (1992) **8**:257–76. doi:10.1017/S0952523800002911
64. Cowie RJ, Robinson DL. Subcortical contributions to head movements in macaques. I. Contrasting effects of electrical stimulation of a medial pontomedullary region and the superior colliculus. *J Neurophysiol* (1994) **72**:2648–64.
65. Nudo RJ, Masterton RB. Descending pathways to the spinal cord: a comparative study of 22 mammals. *J Comp Neurol* (1988) **277**:53–79. doi:10.1002/cne.902770105
66. Nudo RJ, Masterton RB. Descending pathways to the spinal cord: II. Quantitative study of the tectospinal tract in 23 mammals. *J Comp Neurol* (1989) **286**:96–119. doi:10.1002/cne.902860107
67. Muto N, Kakei S, Shinoda Y. Morphology of single axons of tectospinal neurons in the upper cervical spinal cord. *J Comp Neurol* (1996) **372**:9–26. doi:10.1002/(SICI)1096-9861(19960812)372:1<9::AID-CNE2>3.3.CO;2-Q
68. Meredith MA, Miller LK, Ramoa AS, Clemo HR, Behan M. Organization of the neurons of origin of the descending pathways from the ferret superior colliculus. *Neurosci Res* (2001) **40**:301–13. doi:10.1016/S0168-0102(01)00240-1
69. Alstermark B, Isa T. Circuits for skilled reaching and grasping. *Annu Rev Neurosci* (2012) **35**:559–78. doi:10.1146/annurev-neuro-062111-150527
70. Grantyn A, Berthoz A. Reticulo-spinal neurons participating in the control of synergic eye and head movements during orienting in the cat. I. Behavioral-properties. *Exp Brain Res* (1987) **66**:339–54. doi:10.1007/BF00243310
71. Keay K, Westby GW, Frankland P, Dean P, Redgrave P. Organization of the crossed tecto-reticulo-spinal projection in rat – II. Electrophysiological evidence for separate output channels to the periaqueductus area and caudal medulla. *Neuroscience* (1990) **37**:585–601. doi:10.1016/0306-4522(90)90093-J
72. Redgrave P, Dean P, Westby GW. Organization of the crossed tecto-reticulo-spinal projection in rat – I. Anatomical evidence for separate output channels to the periaqueductus area and caudal medulla. *Neuroscience* (1990) **37**:571–84. doi:10.1016/0306-4522(90)90092-I
73. Yasui Y, Ono K, Tsumori T, Yokota S, Kishi T. Tectal projections to the parvocellular reticular formation and the upper cervical spinal cord in the rat, with special reference to axon collateral innervation. *Brain Res* (1998) **804**:149–54. doi:10.1016/S0006-8993(98)00664-7
74. Udo M, Mano N. Discrimination of different spinal monosynaptic pathways converging into reticular neurons. *J Neurophysiol* (1970) **33**:227–38.
75. Grillner S, Lund S. The origin of a descending pathway with monosynaptic action on flexor motoneurons. *Acta Physiol Scand* (1968) **74**:274–84. doi:10.1111/j.1748-1716.1968.tb04236.x
76. Perkins E, Warren S, May PJ. The mesencephalic reticular formation as a conduit for primate collicular gaze control: tectal inputs to neurons targeting

- the spinal cord and medulla. *Anat Rec (Hoboken)* (2009) **292**:1162–81. doi:10.1002/ar.20935
77. Rodgers CK, Munoz DP, Scott SH, Paré M. Discharge properties of monkey tectoreticular neurons. *J Neurophysiol* (2006) **95**:3502–11. doi:10.1152/jn.00908.2005
 78. Guitton D, Munoz DP. Control of orienting gaze shifts by the tectoreticulospinal system in the head-free cat. I. Identification, localization, and effects of behavior on sensory responses. *J Neurophysiol* (1991) **66**:1605–23.
 79. Sooksawate T, Saito Y, Isa T. Electrophysiological and morphological properties of identified crossed tecto-reticular neurons in the rat superior colliculus. *Neurosci Res* (2005) **52**:174–84. doi:10.1016/j.neures.2005.03.007
 80. Kaneda K, Isa T. GABAergic mechanisms for shaping transient visual responses in the mouse superior colliculus. *Neuroscience* (2013) **235**:129–40. doi:10.1016/j.neuroscience.2012.12.061
 81. Kaneda K, Phongphanphanee P, Katoh T, Isa K, Yanagawa Y, Obata K, et al. Regulation of burst activity through pre- and postsynaptic GABA_B receptors in mouse superior colliculus. *J Neurosci* (2008) **28**:816–27. doi:10.1523/JNEUROSCI.4666-07.2008
 82. Kaneda K, Isa K, Yanagawa Y, Isa T. Nigral inhibition of GABAergic neurons in mouse superior colliculus. *J Neurosci* (2008) **28**:11071–8. doi:10.1523/JNEUROSCI.3263-08.2008
 83. Lee PH, Helms MC, Augustine GJ, Hall WC. Role of intrinsic synaptic circuitry in collicular sensorimotor integration. *Proc Natl Acad Sci U S A* (1997) **94**:13299–304. doi:10.1073/pnas.94.24.13299
 84. Edwards MD, White AM, Platt B. Characterisation of rat superficial superior colliculus neurone: firing properties and sensitivity to GABA. *Neuroscience* (2002) **110**:93–104. doi:10.1016/S0306-4522(01)00558-9
 85. Endo T, Yanagawa Y, Obata K, Isa T. Nicotinic acetylcholine receptor subtypes involved in facilitation of GABAergic inhibition in mouse superficial superior colliculus. *J Neurophysiol* (2005) **94**:3893–902. doi:10.1152/jn.00211.2005
 86. Katsuta H, Isa T. Release from GABA_A receptor-mediated inhibition unmasks interlaminar connection within superior colliculus in anesthetized adult rats. *Neurosci Res* (2003) **46**:73–83. doi:10.1016/S0168-0102(03)00029-4
 87. Burbaud P, Bonnet B, Guehl D, Lagueny A, Bioulac B. Movement disorders induced by gamma-aminobutyric agonist and antagonist injections into the internal globus pallidus and substantia nigra pars reticulata of the monkey. *Brain Res* (1998) **780**:102–7. doi:10.1016/S0006-8993(97)01158-X
 88. Dybdal D, Forcelli PA, Dubach M, Oppedisano M, Holmes A, Malkova L. Topography of dyskinesias and torticollis evoked by inhibition of substantia nigra pars reticulata. *Mov Disord* (2013) **28**:460–8. doi:10.1002/mds.25215
 89. Holmes AL, Forcelli PA, Desjardin JT, Decker AL, Teferra M, West EA, et al. Superior colliculus mediates cervical dystonia evoked by inhibition of the substantia nigra pars reticulata. *J Neurosci* (2012) **32**:13326–32. doi:10.1523/JNEUROSCI.2295-12.2012
 90. Hallett M. Neurophysiology of dystonia: the role of inhibition. *Neurobiol Dis* (2011) **42**:177–84. doi:10.1016/j.nbd.2010.08.025
 91. Khooshnoodi MA, Factor SA, Jinnah HA. Secondary blepharospasm associated with structural lesions of the brain. *J Neurol Sci* (2013) **331**:98–101. doi:10.1016/j.jns.2013.05.022
 92. Sadnicka A, Hoffland BS, Bhatia KP, van de Warrenburg BP, Edwards MJ. The cerebellum in dystonia – help or hindrance? *Clin Neurophysiol* (2012) **123**:65–70. doi:10.1016/j.clinph.2011.04.027
 93. Morgante F, Peretti A, Mariotti C, Panzeri M, Fiorio M, Fasano A. Impaired temporal processing of tactile and proprioceptive stimuli in cerebellar degeneration. *PLoS One* (2013) **8**(11):e78628.
 94. Anastasopoulos D, Ziavra N, Pearce R, Bronstein AM. Trunk bradykinesia and foveation delays during whole-body turns in spasmodic torticollis. *J Neurol* (2013) **260**:2057–65. doi:10.1007/s00415-013-6937-8
 95. Klier EM, Wang H, Constantin AG, Crawford JD. Midbrain control of three-dimensional head orientation. *Science* (2002) **295**:1314–6. doi:10.1126/science.1067300
 96. Shaikh AG, Wong AL, Zee DS, Jinnah HA. Keeping your head on target. *J Neurosci* (2013) **33**:11281–95. doi:10.1523/JNEUROSCI.3415-12.2013
 97. Defazio G, Berardelli A, Hallett M. Do primary adult-onset focal dystonias share aetiological factors? *Brain* (2007) **130**:1183–93. doi:10.1093/brain/awl355
 98. Riddle CN, Baker SN. Convergence of pyramidal and medial brain stem descending pathways onto macaque cervical spinal interneurons. *J Neurophysiol* (2010) **103**:2821–32. doi:10.1152/jn.00491.2009
 99. Baker SN. The primate reticulospinal tract, hand function and functional recovery. *J Physiol* (2011) **589**:5603–12. doi:10.1113/jphysiol.2011.215160
 100. Soteropoulos DS, Williams ER, Baker SN. Cells in the monkey ponto-medullary reticular formation modulate their activity with slow finger movements. *J Physiol* (2012) **590**:4011–27. doi:10.1113/jphysiol.2011.225169
 101. Honeycutt CF, Kharouta M, Perreault EJ. Evidence for reticulospinal contributions to coordinated finger movements in humans. *J Neurophysiol* (2013) **110**:1476–83. doi:10.1152/jn.00866.2012
 102. Fiorio M, Tinazzi M, Bertolasi L, Aglioti SM. Temporal processing of visuotactile and tactile stimuli in writer's cramp. *Ann Neurol* (2003) **53**:630–5. doi:10.1002/ana.10525
 103. Fiorio M, Tinazzi M, Scontrini A, Stanzani C, Gambarin M, Fiaschi A, et al. Tactile temporal discrimination in patients with blepharospasm. *J Neurol Neurosurg Psychiatry* (2008) **79**:796–8. doi:10.1136/jnnp.2007.131524
 104. Goodchild RE, Grundmann K, Pisani A. New genetic insights highlight “old” ideas on motor dysfunction in dystonia. *Trends Neurosci* (2013) **36**:717–25. doi:10.1016/j.tins.2013.09.003

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