

TITLE:

Draft Genome Sequence of Thermolongibacillus altinsuensis Strain B1-1, a Novel Hydrogenogenic CO Oxidizer Isolated from Sediment from Lake Biwa in Japan.

AUTHOR(S):

Suzuki, Jota; Imaura, Yoshinari; Nishida, Shiho; Kamikawa, Ryoma; Yoshida, Takashi

CITATION:

Suzuki, Jota ...[et al]. Draft Genome Sequence of Thermolongibacillus altinsuensis Strain B1-1, a Novel Hydrogenogenic CO Oxidizer Isolated from Sediment from Lake Biwa in Japan.. Microbiology Resource Announcements 2023, 12(7): e00334-23.

ISSUE DATE:

2023-07-18

URL:

http://hdl.handle.net/2433/285069

RIGHT:

© 2023 Suzuki et al.; This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International license.











GENOME SEQUENCES



Draft Genome Sequence of *Thermolongibacillus altinsuensis* Strain B1-1, a Novel Hydrogenogenic CO Oxidizer Isolated from Sediment from Lake Biwa in Japan

Jota Suzuki,^a Yoshinari Imaura,^a Shiho Nishida,^a Ryoma Kamikawa,^a Takashi Yoshida^a

^aLaboratory of Marine Microbiology, Graduate School of Agriculture, Kyoto University, Kyoto, Japan

ABSTRACT A facultative anaerobic, thermophilic, hydrogenogenic CO-oxidizing bacterial strain, B1-1, was isolated from a sediment sample from Lake Biwa, a freshwater lake in Japan. B1-1, which is a novel strain of *Thermolongibacillus altinsuensis*, is capable of hydrogenogenic CO oxidation. Here, we report the draft genome sequence of B1-1 (2.92 Mbp, with a GC content of 41.3%).

Thermolongibacillus is an aerobic, thermophilic, endospore-forming, long rod bacterium (1). In the present study, we isolated a strain of *Thermolongibacillus altinsuensis* from freshwater sediment that is capable of hydrogenogenic CO oxidation.

The sediment was collected from Lake Biwa (35°13′13″N, 135°59′48″E), a freshwater lake in Japan, at a depth of 72.6 m using an HR-type core sampler (Rigo Co.). In 5 mL of modified B medium (2, 3), 2.5 g of the sediment was incubated at 65°C under 20% CO/80% N_2 . For isolation of the strain, 50 μ L of the liquid phase was spread on NBRC802 agar medium and incubated at 65°C under aerobic conditions. A single colony of B1-1 was selected and suspended in modified B medium. After 72 h of incubation under the aforementioned conditions, CO depletion (3.011 μ mol/mL) and H_2 and CO_2 production (2.975 and 1.031 μ mol/mL, respectively) were observed using gas chromatography (Shimadzu).

The partial sequence of the 16S rRNA gene of B1-1 was PCR amplified using the universal primers B27f (5'-AGAGAGTTTGATCCTGGCTCAG-3') (4) and U515r (5'-TTACCGCGGCKGCTGVCAC-3') (5). The PCR products were subjected to Sanger sequencing (Eurofins-MWG). A BLASTn search revealed that the sequence of the isolated strain showed the greatest sequence identity (99.13%) to that of *T. altinsuensis* (GenBank accession number NR_125531.1) (6), suggesting that this strain should be assigned to *T. altinsuensis*.

After isolation but prior to genome sequencing, the isolate was passaged 4 times in total in the modified B medium or the NBRC802 medium. The genomic DNA was extracted using a DNeasy blood and tissue kit (Qiagen, Valencia, CA, USA). A DNBSEQ-G400 sequencer (MGI Tech Co., Ltd.) was used for 200-bp, paired-end sequencing analysis (Bioengineering Lab. Co., Ltd.), which yielded 20,034,763 paired-end reads.

Default parameters were used for all software unless otherwise specified. Adapter trimming and quality filtering for the generated reads were conducted using Trimmomatic v0.39 (7), setting the SLIDINGWINDOW option as 4:30. The resulting 12,337,528 reads were assembled using SPAdes v3.15.4 (8), and the assembled scaffolds were annotated using the DFAST server v1.2.18 (9).

The draft genome of B1-1 comprised 352 scaffolds, with an average genome coverage of $634\times$, an N_{50} value of 260,943 bp, a total length of 2,923,574 bp, a GC content of 41.3%, and 2,933 coding sequences. The average nucleotide identity (ANI) between strain B1-1 and T. altinsuensis DSM 24979 (GenBank assembly accession number GCA_004341205.1) calculated using JSpeciesWS (10) was 99.08%, which exceeded the 95% threshold for species identity (11). Therefore, strain B1-1 belongs to T. altinsuensis.

Editor J. Cameron Thrash, University of Southern California

Copyright © 2023 Suzuki et al. This is an openaccess article distributed under the terms of the Creative Commons Attribution 4.0 International license.

Address correspondence to Takashi Yoshida, yoshida.takashi.7a@kyoto-u.ac.jp.

The authors declare no conflict of interest.

Received 20 April 2023 Accepted 17 May 2023 Published 5 June 2023 Announcement



The B1-1 strain has one *codh* gene cluster containing *cooS*, which encodes the anaerobic CO dehydrogenase catalytic subunit CooS, the amino acid sequence of which is conserved for the residues constituting the active site (12). The *codh* gene cluster is located adjacent to a gene cluster encoding the membrane-bound hydrogenase (*ech*), implying the contribution of these genes to hydrogenogenic CO oxidation (13). The *codh-ech* gene cluster was not detected in the genome of *T. altinsuensis* DSM 24979 (GenBank assembly accession number GCA 004341205.1).

Data availability. The genome sequence of *Thermolongibacillus altinsuensis* strain B1-1 has been deposited in the DNA Data Bank of Japan (DDBJ) under the accession number BSVG01000000. Sequence data have been deposited in the DDBJ Sequence Read Archive (DRA) under the accession number DRX440543.

ACKNOWLEDGMENTS

We are grateful to Yukiko Goda, Tetsushi Akatsuka, and the crew of the sampling vessel *HASU* for providing technical assistance during sample collection. Furthermore, we thank Satori Imataka for valuable assistance during sampling. We thank Editage (www.editage.jp) for English language editing.

This work was funded by the Institute for Fermentation (Osaka, Japan) (grant L-2021-1-002 awarded to T.Y.) and supported in part by a Grant-in-Aid for Scientific Research (S) (grant 21H05057 awarded to T.Y.) from the Japan Society for the Promotion of Science.

REFERENCES

- Cihan AC, Koc M, Ozcan B, Tekin N, Cokmus C. 2014. Thermolongibacillus altinsuensis gen. nov., sp. nov. and Thermolongibacillus kozakliensis sp. nov., aerobic, thermophilic, long bacilli isolated from hot springs. Int J Syst Evol Microbiol 64:187–197. https://doi.org/10.1099/ijs.0.053280-0.
- Yoneda Y, Yoshida T, Yasuda H, Imada C, Sako Y. 2013. A thermophilic, hydrogenogenic and carboxydotrophic bacterium, *Calderihabitans maritimus* gen. nov., sp. nov., from a marine sediment core of an undersea caldera. Int J Syst Evol Microbiol 63:3602–3608. https://doi.org/10.1099/ijs.0.050468-0.
- Adachi Y, Inoue M, Yoshida T, Sako Y. 2020. Genetic engineering of carbon monoxide-dependent hydrogen-producing machinery in *Parageobacillus* thermoglucosidasius. Microbes Environ 35:ME20101. https://doi.org/10 .1264/jsme2.ME20101.
- Lane DJ, Pace B, Olsen GJ, Stahl DA, Sogin ML, Pace NR. 1985. Rapid determination of 16S ribosomal RNA sequences for phylogenetic analyses. Proc Natl Acad Sci U S A 82:6955–6959. https://doi.org/10.1073/pnas.82.20.6955.
- Gumerov VM, Mardanov AV, Beletsky AV, Bonch-Osmolovskaya EA, Ravin NV. 2011. Molecular analysis of microbial diversity in the Zavarzin Spring, Uzon Caldera, Kamchatka. Microbiology 80:244–251. https://doi.org/10 .1134/S002626171102007X.
- Altschul SF, Gish W, Miller W, Myers EW, Lipman DJ. 1990. Basic local alignment search tool. J Mol Biol 215:403–410. https://doi.org/10.1016/ S0022-2836(05)80360-2.
- 7. Bolger AM, Lohse M, Usadel B. 2014. Trimmomatic: a flexible trimmer for Illumina sequence data. Bioinformatics 30:2114–2120. https://doi.org/10.1093/bioinformatics/btu170.

- 8. Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. J Comput Biol 19:455–477. https://doi.org/10.1089/cmb.2012.0021.
- 9. Tanizawa Y, Fujisawa T, Nakamura Y. 2018. DFAST: a flexible prokaryotic genome annotation pipeline for faster genome publication. Bioinformatics 34: 1037–1039. https://doi.org/10.1093/bioinformatics/btx713.
- Richter M, Rosselló-Móra R, Oliver Glöckner F, Peplies J. 2016. JSpeciesWS: a web server for prokaryotic species circumscription based on pairwise genome comparison. Bioinformatics 32:929–931. https://doi.org/10.1093/ bioinformatics/btv681.
- Chun J, Oren A, Ventosa A, Christensen H, Arahal DR, da Costa MS, Rooney AP, Yi H, Xu X-W, De Meyer S, Trujillo MEY. 2018. Proposed minimal standards for the use of genome data for the taxonomy of prokaryotes. Int J Syst Evol Microbiol 68:461–466. https://doi.org/10.1099/ijsem.0.002516.
- Svetlitchnyi V, Dobbek H, Meyer-Klaucke W, Meins T, Thiele B, Römer P, Huber R, Meyer O. 2004. A functional Ni-Ni-[4Fe-4S] cluster in the monomeric acetyl-CoA synthase from Carboxydothermus hydrogenoformans. Proc Natl Acad Sci U S A 101:446–451. https://doi.org/10.1073/pnas.0304262101.
- Inoue M, Nakamoto I, Omae K, Oguro T, Ogata H, Yoshida T, Sako Y. 2019.
 Structural and phylogenetic diversity of anaerobic carbon-monoxide dehydrogenases. Front Microbiol 9:3353. https://doi.org/10.3389/fmicb.2018.03353.