Łopuszyńska Anna, Pawlicki Bartłomiej, Pawlicki Mateusz, Lorenc Karol, Kozioł Magdalena, Misztal Zofia. Diagnosis of cervical cancer by a urine test. Journal of Education, Health and Sport. 2019;9(8):823-828. eISSN 2391-8306. DOI http://dx.doi.org/10.5281/ zenodo.3408182

http://ojs.ukw.edu.pl/index.php/johs/article/view/7246

The journal has had 7 points in Ministry of Science and Higher Education parametric evaluation. Part B item 1223 (26/01/2017). 1223 Journal of Education, Health and Sport eISSN 2391-8306 7

© The Authors 2019:

© 1 fe Authors 2019; This article is published with open access at Licensec Open Journal Systems of Kazimierz Wielki University in Bydgoszcz, Poland Access. This article is distributed under the terms of the Creative Commons Attribution Noncommercial License which permits any noncommercial use, distribution, and reproduction in any medium, provided the original author (s) and source are credited. This is an open access article licensed under the terms of the Creative Commons Attribution Non commercial license there are credited. (http://creativecommons.org/licenses/by-nc-sa/4.0) which permits unrestricted, non commercial use, distribution and reproduction in any medium, provided the work is properly cited.

The authors declare that there is no conflict of interests regarding the publication of this paper.

Received: 05.07.2019. Revised: 25.07.2019. Accepted: 18.08.2019.

Diagnosis of cervical cancer by a urine test

Anna Łopuszyńska¹, Bartłomiej Pawlicki², Mateusz Pawlicki¹, KarolLorenc¹, Magdalena Kozioł¹, Zofia Misztal³

1 Student Scientific Association at Department of Epidemiology and Clinical Research Radziwiłłowska Methodology, Medical University of Lublin, ul. 11, Lublin 20-080, Poland;lopuszynskaania@gmail.com; 0000-0001-5133-4180; lorenckarol2@gmail.com; 0000-0002-6414-5984; magdalena.koziol@icloud.com; 0000-0002-8671- 5968; pawlak32@gmail.com; 0000-0001-8318-6573;

"Pod Łysicą", Ostrowiec Świętokrzyski; 2 Apteka bp.pawlak@gmail.com; 0000-0003-4278-4168;

3 Student Scientific Association at Department of Family Medicine, Medical University of Lodz, ul. Narutowicza 60, Lodz 90-136, Poland; zosia.misztal6@gmail.com; 0000-0003-2317-9667.

ABSTRACT:

Cervical cancer is the fourth most common malignant tumor among women in the world. However, 90% of deaths occur in developing countries. Tumor pathogenesis is associated with exposure to highrisk human papillomavirus (hrHPV), most often 16 and 18 strains. The sooner precancerous lesions or cancer are detected, the higher the chance of survival is. That is why prophylaxis is so important in this case. Due to the low turnout of women in cytology, new, alternative methods of prevention are needed. According to the research, women prefer tests in which samples are taken by themselves. Hence, more and more studies on the use of urine in the prevention of cervical cancer. Urine is a material that is easy to pick up. Patients feel comfortable because they can do it by themselves. Still, more research is needed to optimize its collection, transport, or tests used on samples.

KEY WORDS: human papillomavirus, uterine cervical neoplasms, urine

INTRODUCTION:

Cervical cancer is the primary malignant tumor of this organ. The pathogenesis of cancer and precancerous lesions, i.e. CIN (cervical intraepithelial neoplasia), is associated in most cases with exposure to high-pathogenic HPV (human papillomavirus) - 16 and 18 (account for 70% of cases of disease [1]), and strains 31, 33, 45, 52 and 58 [2]. Risk factors of this cancer are: early age of sexual life, numerous sexual partners, smoking, pregnancy and childbirth at an early age, and the occurrence of this cancer in the family [3]. HPV viruses are characterized by tropism to immature epithelial cells of the transitional cervix. Infected cells express oncoproteins. E6 and E7 oncoproteins bind and inactivate p53 and Rb suppressor genes, which promotes carcinogenesis [4]. The most common histological types of cervical cancer are squamous cell carcinoma and adenocarcinoma. The peak of illness occurs around the age of 45 and the symptoms reported by the patient are non-specific. These include unexpected bleedings, vaginal discharge, painful urination and sexual intercourse. The tumor infiltrates the surrounding tissues, and also passes through the lymphatic vessels to the lymph nodes or via the blood vessels to the lungs and bones. In order to assess the staging of this cancer, we use the four-stage FIGO scale, where:

- I is carcinoma limited to the cervix,
- II carcinoma extends beyond the cervix and/or succumbing, but does not infiltrate the lower third of the vagina or pelvic wall,
- III cancer infiltrates the lower third vaginal walls and pelvic wall,
- IV-cancer infiltrates the bladder, rectum and gives distant metastases [5].

Cytological examination is an example of cervical cancer prevention. It involves scraping the epithelial cells of the cervical transition zone, and then evaluating the smear according to the Bethesda scale. This allows for on early detection of pre-cancer lesions. If an abnormal cytological result is obtained, the changes or their absence may be verified by histopathological examination. However, the consistency of samples varies from 40% to 89% depending on the literature, which indicates the possible wrong results of this method [6]. Another method is the detection of high-pathogenic HPV strains cervical in scrapings. According to the WHO, cervical cancer is the fourth most common malignant tumor in women. In 2018, about 570000 new cases were diagnosed, which accounted for 6.6% of malignant tumors of women. 90% of the deaths caused by this cancer have been reported in developing countries. The earlier the cancer is detected, the higher the chance of being cured. The mortality rate may be reduced appropriate education, prevention early diagnosis [7]. bv or The aim of this work is to present new, effective, non-invasive methods to diagnose cervical cancer by means of a urine test. Urine test can increase the comfort of women undergoing testing. It can allow them to gather samples for the test alone, which can affect a larger number of women undergoing prevention of this cancer and thus may reduce the mortality rate.

A REVIEW OF AVAILABLE RESEARCH:

Research conducted in the Federated States of Micronesia showed that out of 217 women, up to 95% were satisfied with the urine test. In the case of cytology it was 82% of the patients, but only 42% of women would prefer that an experienced clinician doing it. These studies suggest that self-sampling is preferred among [8]. women In the case of a study conducted in Korea, 732 women aged 20 to 69 showed that overall satisfaction was significantly higher for both vaginal sampling and urine sampling compared to the cytology performed by the clinician (odds ratio [OR] = 2.01, 95% confidence interval [CI] = 1.48-3.00 and OR = 2.47, 95% CI = 1.75-3.48, respectively). This suggests that the possibility of self-sampling may increase the number of women undergoing cervical cancer prevention [9]. Research in Thailand has shown that attendance in screening tests for cervical cancer detection is very low (25-38% of women aged 30-35 have had a cytology performed once in life). In a study of 164 women, cervical swabs and urine samples were compared using HPV test (HPV Geno Array Diagnostic Kits). The overall agreement between paired samples was 62.5%. Analysis of urine samples and a second analysis of cervical smear samples showed that differences in the overall rate of HPV detection between women with normal and abnormal cytology were not significant (p> 0.05). This result suggests that urine is a feasible and possible substitute for cervical smears. Urine test to detect infection with high-pathogenic HPV strain may be an alternative to cytology [10]. 240 women took a urine sample themselves, and the clinician additionally performed a cytology. Among all examined samples, the incidence of HPV was 42.9% among urine samples. The compatibility between the two types of samples was 98.4%, k = 0.792. Incompatible results were observed in 27 cases; 5 were positive only in urine samples, and 22 were positive only in swab specimens. The sensitivity and specificity for total HPV DNA in the urine fraction using cervical samples as reference was 68.4% and 99.9%. The results of these studies also suggest that urine may method be а non-invasive, alternative for detecting HPV infections [11]. Studies in Spain consisted of comparing samples of urine collected in the morning, urine collected later, material taken independently from the cervix, and material collected by the clinician. Samples from 91 patients were analyzed. All 6 cases of CIN3 showed a positive hrHPV test in each type of sample, in both HPV tests (SPF10-DEIA-LiPA25 and GP5+/6+-EIA-LMNX). The sensitivity for CIN2+ in the SPF10 system was 95% for the urine sample collected in the morning and 100% for all other samples. In the GP5+/6+ test, the sensitivity was 95% in all types of samples. The sensitivity and specificity for both tests on each type of samples did not differ significantly. There was a 10-14% inconsistency in the hrHPV genotype. Similar sensitivity of CIN2+ was shown for HPV testing in the first-void urine, a smear taken by a doctor and a cervical sample taken alone [12]. Studies in Belgium on 110 women from 25 to 64 years of age directed to colposcopy suggest that first-void urine samples may be an alternative to cervical specimens to detect HPV DNA. In the case of high-risk HPV strains, the compliance of paired urine samples and cervical smear was very high (Kappa Cohen 0.688 (95% CI: 0.542-0.835)). In addition, women have been shown to prefer selfcollection of urine to the study than the cytology performed by the clinician [13]. In North Thailand, studies have been carried out on HPV+ women. Urine and cervical samples were obtained from 168 women. Out of 123 correctly collected paired samples, compliance in high-risk HPV DNA detection was present in 106 cases (86.2%), with kappa statistics of 0.65 (significant compliance). Using the HPV results from the cervix as a reference, the sensitivity of HPV tests in the urine was 68.6% (24/35) and the specificity was 93.2% (82/88). In order to detect HSIL+, the sensitivity of HPV in the urine was 80.0% (4/5) and the specificity was 78.0% (92/118). HPV in urine had a high specificity in HPV detection, as well as high sensitivity in histological detection of HSIL+ [14].

Tests carried out in Korea consisted of taking vaginal and urine samples by the patient herself. Highrisk HPV strains were detected in 6.7% of urine samples and in 9.6% of vaginal smear specimens. HPV 16/18 was detected in 1.5% (other hrHPV strains 5.2%) of urine samples and 2.0% (other hrHPVstrains 7.6%) of vaginal smear samples. Although a statistically significant difference in the frequency of hrHPV detection between urine samples and vaginal smears was observed (p <0.001), the compliance for HPV 16/18 was relatively high (99.1%, 95% CI 98.1 \sim 99.6%), from kappa 0.75. In addition, satisfaction with self-collection of both urine samples (91.4%) and vaginal swabs (92.7%) was higher than in the case of the clinician (88.1%). The study suggests that self-sampling may be an clinician's performance alternative the to [15]. In the case of detection of hrHPV infection in a woman, this result can be confirmed by the presence of DNA methylation markers in the cervical material. It was decided to check whether these markers are also detectable in urine samples. 43 urine samples and 38 paired cervical scrapings were collected from patients with cervical cancer, aged from 27 to 86 years. It has been shown that both native urine (24/28-86%) and sediment (25/28-89%) are suitable for the detection of high-risk HPV strains as well as DNA methylation markers. A strong relationship was found, both between native urine and sediment and all methylation markers tested (FAM19A4, GHSR, PHACTR3, PRDM14, SST, ZIC1). The results of the test for the presence of HPV and methylation markers in the urine and in the cervical scrapings were compared. In paired samples, hrHPV infection was detected in 31 (82%) urine sediments and 34 (89%) of cervical scratches, what led to almost perfect compliance (with kappa value of 0.85, which gives 95% confidence interval) а [16] In the case of CIN2+ detection studies, the preservative fixed urine showed good compatibility with the vaginal samples to detect hrHPV. The detection sensitivity of CIN2+ was 15/18 (83%) for urine and 16/18 (89%) for cervical and vaginal samples according to ART (Abbott RealTime), and 15/17 (88%) for all samples by RC (Roche Cobas 4800). Urine tests have been shown to be widely accepted by women. It suggests that research should be continued to develop an alternative method of

prophylaxis [17]. For detection of HPV DNA in the urine was accepted as a screening test for cervical cancer, there is research on the most effective method. Innovative studies were conducted involving the use of polypyrrole polyolefin polypeptides (PEI-mPpy NW) coupled to polyethyleneimine for the extraction, identification and detection of colorimetric strains of HPV DNA in urine samples of patients with cervical cancer. A 100% compliance rate was obtained between urine samples and cervical smears, even with a small amount of urine (300 μ L). This method gives high hopes and may be a future in the prevention of this cancer [18]. Studies conducted on 43 patients in Puerto Rico showed that there are three metabolites in the urine of women infected with high-pathogenic HPV strain: 5-oxoprolinate, erytronic acid and N-acetylaspartic acid. All of them differentiate samples from negative samples as well as samples infected simultaneously with high and low-oxygen HPV strain. However, it is necessary to study a larger of patients prove this finding group to [19]. Another study shows that during cervical uterine cancer, the concentration of 60 different proteins increased, including leucine-rich α -2-glycoprotein (LRG1) and isoform-1 multimerin-1 (MMRN1). In contrast, the concentration of 73 proteins decreased, such as the S100 A8 (S100A8) calcium binding protein, serpin B3 (SERPINB3) and the differential antigen-44 cluster (CD44). ROC analysis showed that LRG1 and SERPINB3 can be used individually to detect cervical cancer. It was also shown that these 5 proteins together can be used in the diagnosis of this cancer [20].

CONCLUSIONS:

Urine testing may be the future and an alternative to other screening methods in the diagnosis of cervical cancer. This may increase the number of women undergoing screening due to the greater comfort, ease and ability to perform the test by themselves. Faster detection of pre-cancerous and cancerous lesions will increase the chance of recovery and also the survival rate. However, further research is needed to optimize this method such as how to retrieve material for test, how to store it, and how to research it.

REFERENCES:

- 1. Douglas R. Lowy and John T. Schiller. *Prophylactic human papillomavirus vaccines*. J Clin Invest. 2006 May 1; 116(5): 1167–1173. doi: 10.1172/JCI28607. PMCID: PMC1451224. PMID: 16670757.
- 2. Hooi DJ, Lissenberg-Witte BI, deKoning MNC, Pinedo HM, Kenter GG, Meijer CJ, Quint WG.*High prevalence of high-risk HPV genotypes other than 16 and 18 in cervical cancers of Curaçao: implications for choice of prophylactic HPV vaccine*. Sex Transm Infect. 2018 Jun;94(4):263-267. doi: 10.1136/sextrans-2017-053109. Epub 2017 Oct11.
- 3. Grzegorz Bręborowicz, *Położnictwo i ginekologia, tom 2,* Wydawnictwo Lekarskie PZWL, Warszawa 2007, wyd. 1, ISBN 83-200-3082-4 p. 819-832
- 4. Vinay Kumar, Abul K. Abbas, Jon C. Aster. *Robbins PATOLOGIA*. (red.) Włodzimierz Olszewski. Wyd. 2. Wrocław 2014. Elsevier Urban&Partner. P. 737-741.
- 5. Tsikouras P, Zervoudis S, Manav B, Tomara E, Iatrakis G, Romanidis C, Bothou A, Galazios G. *Cervical cancer: screening, diagnosis and staging.* J BUON. 2016 Mar-Apr;21(2):320-5.
- 6. Adrianna Skrajna, Aleksandra Zielińska, Małgorzata Kania, Krzysztof Cendrowski, Włodzimierz Sawicki. Assessment of the efficiency of cervical cytology in detection of cervical premalignant conditions and cervical carcinoma. Postępy Nauk Medycznych, t. XXVI, nr 7, 2013. Borgis. P. 457-460.
- 7. https://www.who.int/cancer/prevention/diagnosis-screening/cervical-cancer/en/____(accessed July16, 2019)
- 8. Angela U. Sy, Brenda Y. Hernandez, Aileen Tareg, Martina Reichhardt, and Lee Buenconsejo-Luma Acceptability and feasibility of acommunity based participatory research

project comparing cytology and urine HPV DNA testing for cervical cancer screening in Yap, Federated States of Micronesia Cancer Epidemiol. Author manuscript; available in PMC 2017 Dec 21. Published in final editedform as: Cancer Epidemiol. 2017 Oct; 50(Pt B): 283–288. doi: 10.1016/j.canep.2017.07.008 PMCID: PMC5739880 NIHMSID: NIHMS922469 PMID: 29120838.

- 9. Shin H, Lee B, Hwang SH, Lee DO, SungNY, Park JY, Jun JK. *Evaluation of satisfactionwith three different cervical cancer screening modalities: clinician-collected Pap test vs. HPV test by self-sampling vs. HPV test by urine sampling.* J Gynecol Oncol. 2019 Sep;30(5):e76. doi: 10.3802/jgo.2019.30.e76.
- Pornjarim Nilyanimit, Jira Chansaenroj, Anant Karalak, Piyawat Laowahutanont, Pairoj Junyangdikul, andYong Poovorawan Academic Editor: Salvatore Andrea Mastrolia. *Comparison of human papillomavirus (HPV) detection inurine and cervical swab samples using the HPV Geno Array Diagnostic assay* PeerJ. 2017; 5: e3910. Published online 2017 Oct 9. doi: 10.7717/peerj.3910 PMCID: PMC5637711 PMID: 29038761
- 11. Hagihara M, Yamagishi Y, Izumi K, Miyazaki N, Suzuki T, Kato H, Nishiyama N, Koizumi Y, Suematsu H, Mikamo H. *Comparison of initial stream urine samples and cervical samples for detection of human papillomavirus*. J Infect Chemother. 2016 Aug;22(8):559-62. doi: 10.1016/j.jiac.2016.05.009. Epub 2016 Jun21.
- 12. Leeman A, Del Pino M, Molijn A, Rodriguez A, Torné A, de Koning M, Ordi J, van Kemenade F, Jenkins D, Quint W. *HPV testing in first-void urine provides sensitivity for CIN2+ detection comparable with a smear taken by a clinician or a brush-based self-sample: cross-sectional data from a triage population.* BJOG. 2017 Aug;124(9):1356-1363. doi: 10.1111/1471-0528.14682.
- Severien Van Keer, Wiebren A. A. Tjalma, Jade Pattyn, Samantha Biesmans, Zoë Pieters, Xaveer Van Ostade, Margareta Ieven, Pierre Van Damme, and AlexVorsters. *Human papillomavirus genotype and viral load agreement between paired first-void urine and clinician-collected cervical samples*. Eur J Clin Microbiol Infect Dis. 2018; 37(5): 859– 869. Published online 2018 Feb7. doi: 10.1007/s10096-017-3179-1 PMCID: PMC5916996 PMID: 29417310
- 14. Surapan Khunamornpong, Jongkolnee Settakorn, Kornkanok Sukpan, Suree Lekawanvijit, Narisara Katruang, and Sumalee Siriaunkgul. *Comparison of Human Papillomavirus Detection in Urine and Cervical Samples Using High-Risk HPV DNA Testing in Northern Thailand* Obstet Gynecol Int. 2016; 2016: 6801491. Published online 2016 Dec 22. doi: 10.1155/2016/6801491 PMCID: PMC5215104 PMID: 28101107
- Sang-HyunHwang, Hye Young Shin, Dong Ock Lee, Na Young Sung, Bomyee Lee, Do-Hoon Lee, and JaeKwan Jun. A prospective pilot evaluation of vaginal and urine selfsampling for the Roche cobas 4800 HPV test for cervical cancer screening. Sci Rep. 2018; 8: 9015. Published online 2018 Jun 13. doi: 10.1038/s41598-018-27390-5PMCID: PMC5998027 PMID: 29899531
- 16. Barbara C. Snoek, Annina P. van Splunter, Maaike C. G. Bleeker, Maartje C. vanRuiten, Daniëlle A. M. Heideman, W. Frederik Rurup, Wina Verlaat, Hans Schotman, Mignon vanGent, Nienke E. van Trommel, and Renske D. M. Steenbergen. *Cervical cancer detection by DNA methylation analysis inurine*. Sci Rep. 2019; 9: 3088. Published online 2019 Feb 28. doi: 10.1038/s41598-019-39275-2 PMCID: PMC6395822 PMID: 30816167
- Alex Sargent, Samantha Fletcher, Katarina Bray, Henry C Kitchener, and Emma J Crosbie. Cross-sectional study of HPV testing in self-sampled urine and comparison with matched vaginal and cervical samples in women attendingcolposcopy for the management of abnormalcervical screening. BMJ Open. 2019; 9(4): e025388.mPublished online 2019 Apr 29. doi: 10.1136/bmjopen-2018-025388 PMCID: PMC6502061 PMID: 31036707
- Hyung Jae Lee, Mihye Choi, Sang-Hyun Hwang, and Youngnam Cho. A Versatile Nanowire Platform for Highly EfficientIsolation and Direct PCR-free Colorimetric Detection of Human Papillomavirus DNA from Unprocessed Urine Theranostics. 2018; 8(2): 399–409. Published online 2018 Jan 1. doi: 10.7150/thno.21696 PMCID: PMC5743556 PMID: 29290816

- Filipa Godoy, Gilmary Ortiz-Morales, Josefina Romaguera, Maria M. Sanchez, Magaly Martinez-Ferrer, and Natalyia Chorna, Kalimuthusamy Natarajaseenivasan. Discriminatinghigh-risk cervical Human Papilloma Virus infections with urinary biomarkers via non-targetedGC-MS-based metabolomics. PLoS One. 2018; 13(12): e0209936. Published online 2018 Dec 28. doi: 10.1371/journal.pone.0209936 PMCID: PMC6310238 PMID: 30592768
- 20. Daranee Chokchaichamnankit, Kamolwan Watcharatanyatip, Pantipa Subhasitanont, Churat Weeraphan, Siriporn Keeratichamroen, Narongrit Sritana, Nuttavut Kantathavorn, Penchatr Diskul-Na-Ayudthaya, Kittirat Saharat, JuthamardChantaraamporn, Chris Verathamjamras, Natacha Phoolcharoen, Kriangpol Wiriyaukaradecha, Nilubol Monique Paricharttanakul, Wandee Udomchaiprasertkul, Thaniya Sricharunrat, Chirayu Auewarakul, Jisnuson Svasti, and Chantragan Srisomsap. Oncol Lett. 2019 Jun; 17(6): 5453–5468. Published online 2019 Apr8. doi: 10.3892/ol.2019.10227 PMCID: PMC650743 PMID: 31186765