



King's Research Portal

DOI:

[10.1093/cercor/bhab404](https://doi.org/10.1093/cercor/bhab404)

Document Version

Publisher's PDF, also known as Version of record

[Link to publication record in King's Research Portal](#)

Citation for published version (APA):

Zelenina, M., Kosilo, M., Da Cruz, J., Antunes, M., Figueiredo, P., Mehta, M. A., & Prata, D. (2022). Temporal dynamics of intranasal oxytocin in human brain electrophysiology. *Cerebral Cortex*, 32(14), 3110-3126. <https://doi.org/10.1093/cercor/bhab404>

Citing this paper

Please note that where the full-text provided on King's Research Portal is the Author Accepted Manuscript or Post-Print version this may differ from the final Published version. If citing, it is advised that you check and use the publisher's definitive version for pagination, volume/issue, and date of publication details. And where the final published version is provided on the Research Portal, if citing you are again advised to check the publisher's website for any subsequent corrections.

General rights

Copyright and moral rights for the publications made accessible in the Research Portal are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognize and abide by the legal requirements associated with these rights.

- Users may download and print one copy of any publication from the Research Portal for the purpose of private study or research.
- You may not further distribute the material or use it for any profit-making activity or commercial gain
- You may freely distribute the URL identifying the publication in the Research Portal

Take down policy

If you believe that this document breaches copyright please contact librarypure@kcl.ac.uk providing details, and we will remove access to the work immediately and investigate your claim.

Temporal dynamics of intranasal oxytocin in human brain electrophysiology

Marie Zelenina^{1,2}, Maciej Kosilo¹, Janir da Cruz^{3,4}, Marília Antunes⁵, Patrícia Figueiredo^{4,6}, Mitul A Mehta⁷, Diana Prata^{id}^{1,8,9,*}

¹Instituto de Biofísica e Engenharia Biomédica, Faculdade de Ciências, Universidade de Lisboa, Lisboa 1749-016, Portugal,

²Section on Clinical and Computational Psychiatry, NIMH, NIH, MD 20814, USA,

³Laboratory of Psychophysics, Brain Mind Institute, École Polytechnique Fédérale de Lausanne (EPFL), Lausanne 1015, Switzerland,

⁴Institute for Systems and Robotics–Lisbon (LARSyS) and Department of Bioengineering, Instituto Superior Técnico, Universidade de Lisboa, Lisboa 1049-001, Portugal,

⁵Centro de Estatística e Aplicações e Departamento de Estatística e Investigação Operacional, Faculdade de Ciências, Universidade de Lisboa, Lisboa 1749-016, Portugal,

⁶INESC-ID, Instituto Superior Técnico, 1749-016 Lisboa, Portugal,

⁷Department of Neuroimaging, Institute of Psychiatry, Psychology and Neuroscience, King's College London, London SE5 8AF, UK,

⁸Instituto Universitário de Lisboa (ISCTE-IUL), CIS-IUL, Lisboa 1649-026, Portugal,

⁹Department of Old Age Psychiatry, Institute of Psychiatry, Psychology and Neuroscience, King's College London, SE5 8AF London, UK

*Corresponding author: Dr. Diana Prata, Instituto de Biofísica e Engenharia Biomédica, Faculdade de Ciências, Universidade de Lisboa, Portugal.

Email: diana.prata@kcl.ac.uk.

†These authors gave equivalent contribution.

Oxytocin (OT) is a key modulator of human social cognition, popular in behavioral neuroscience. To adequately design and interpret intranasal OT (IN-OT) research, it is crucial to know for how long it affects human brain function once administered. However, this has been mostly deduced from peripheral body fluids studies, or uncommonly used dosages. We aimed to characterize IN-OT's effects on human brain function using resting-state EEG microstates across a typical experimental session duration. Nineteen healthy males participated in a double-blind, placebo-controlled, within-subject, cross-over design of 24 IU of IN-OT in 12-min windows 15 min-to-1 h 42min after administration. We observed IN-OT effects on all microstates, across the observation span. During eyes-closed, IN-OT increased duration and contribution of A and contribution and occurrence of D, decreased duration and contribution of B and C; and increased transition probability C-to-B and C-to-D. In eyes-open, it increased A-to-C and A-to-D. As microstates A and D have been related to phonological auditory and attentional networks, respectively, we posit IN-OT may tune the brain for reception of external stimuli, particularly of social nature—tentatively supporting current neurocognitive hypotheses of OT. Moreover, we contrast our overall results against a comprehensive literature review of IN-OT time-course effects in the brain, highlighting comparability issues.

Key words: electroencephalography; microstates; oxytocin; pharmacodynamics; resting-state.

Introduction

Intranasal oxytocin (IN-OT) has been used extensively to probe this neuropeptide's role in human social cognition and behavior as we and others have reviewed (Evans et al. 2014; Leng and Ludwig 2016; Torres et al. 2018), and reported on (Neto et al. 2020). The potential therapeutic value of IN-OT has been studied in neuropsychiatric disorders with profound social and affective deficits, such as autism (Anagnostou et al. 2014), schizophrenia (De Berardis et al. 2013), and anxiety and depressive disorders (De Cagna et al. 2019)—with a general pattern of mixed findings, and lack of clear results (Bradley and Woolley 2017). IN-OT psychopharmacological studies need to collect outcome data within a time window that captures the pharmacodynamic effects of IN-OT. Generally, the timing of assessments can be determined by the plasma pharmacokinetics of a drug, or measurement of brain outcomes. We have summarized the timings used in pre-

vious studies in Table 1, also discussed them in detail in Box 1.

Overall, studies demonstrate variability in the time windows tested and in the effects. Most of the findings used to inform subsequent decisions on psychopharmacological administration timings do not stem from brain activity measurements, but instead peripheral levels of OT in: saliva (Huffmeijer et al. 2012; van IJzendoorn et al. 2012; Weisman et al. 2012; Daughters et al. 2015), blood plasma (Gossen et al. 2012; Striepens et al. 2013), and cerebrospinal fluid (CSF) (Striepens et al. 2013). Out of 11, only three studies have used magnetic resonance imaging (MRI) to examine the effects of IN-OT on the human brain across time, two measuring cerebral blood flow (CBF) (Paloyelis et al. 2016; Martins et al. 2020b), and one using blood oxygenation-level-dependent imaging (BOLD) (Spengler et al. 2017). As such, findings that seem consistent (i.e., because they overlap in time) may not be in fact directly comparable given the different tissues in

Received: August 9, 2021. Revised: October 5, 2021. Accepted: October 21, 2021

© The Author(s) 2021. Published by Oxford University Press.

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<https://creativecommons.org/licenses/by/4.0/>), which permits unrestricted reuse, distribution, and reproduction in any medium, provided the original work is properly cited.

Table 1. Systematic summary of all previous human pharmacodynamics studies of intranasal oxytocin.

Study	N, gender	Design	Methods (OT extraction and quantification; or MRI technique)	IN-OT Dose	Post-administration time windows measured	Peak effect time window	Statistically significant effect time windows
SALIWA (Weisman et al. 2012)	10, m/f	Within-subj, double-blind PL	No extraction (lyophilization only for placebo); ELISA Extraction; ELISA	24 IU	15 min, 30 min, 45 min, 1 h, 1 h20min, 1 h40min, 2 h, 3 h, 4 h	15 min	15 min, 30 min, 45 min, 1 h, 1 h20min, 1 h40min, 2 h, 3 h, 4 h (all time windows)
(Huifmeijer et al. 2012)	57, f	Within-subj, double-blind PL	Extraction; ELISA	16 IU	Approx. 1 h15 min, approx. 2 h15 min	Approx. 1 h15 min	Approx. 1 h15 min, approx. 2 h15 min (all time windows)
(van Ijzendoorn et al. 2012)	47, f	Between-subj, double-blind PL	Extraction; ELISA	16/24 IU	1 h, 2 h, 3 h, 4 h, 5 h, 6 h, 7 h	1 h	1 h, 2 h, 3 h, 4 h, 5 h, 6 h, 7 h (all time windows)
(Daughters et al. 2015)	40, m	Within-subj, double-blind PL	No extraction (unclear if lyophilization); ELISA	24 IU	30 min, 1 h, 1 h30 min, 1 h45 min, 1 h48min	30 min	30 min, 1 h, 1 h30 min, 1 h45 min, 1 h48min (all time windows)
(Spengler et al. 2017)	116, m	Crossover, double-blind PL	Extraction; RIA	12/24/48 IU	12/48 IU: approx. 15 min, 40 min, 1 h20min, 1 h45min 24 IU: approx. 15 min, 45 min, 1 h25min, 1 h40min	NA	(all time windows)
BLOOD (Landgraf 1985)	3, m	No PL	Extraction; RIA	" about 65 µ," appr. 38 IU	10 min, 20 min, 30 min, 40 min, 50 min, 1 h, 1 h10min	NA	10 min, 20 min, 30 min
(Gossen et al. 2012)	8, m	No PL	Extraction; FEIA	26 IU	30 min, 1 h30 min, 2 h30 min, 3 h30 min	30 min	30 min, 1 h30 min (2 h30 min for some subj.)
(Striepens et al. 2013)	15, m	Between-subj, double-blind PL	Extraction; RIA	24 IU	15 min, 30 min, 45 min, 1 h, 1 h15 min, 1 h30 min	15 min	15 min, 30 min, 45 min, 1 h, 1 h15 min, 1 h30 min (all time windows)
(Quintana et al. 2015)*	16, m	Within-subj, double-blind, double-dummy, crossover	Extraction; ELISA	8/24 IU	10 min, 30 min, 1 h, 2 h	NA	10 min, 30 min, 1 h, 2 h (all time windows)
(Spengler et al. 2017)	116, m	Crossover, double-blind PL	Extraction; RIA	12/24/48 IU	24 IU: 45 min, 1 h25min, 1 h45min 12/48 IU: 1 h25min	NA	45 min, 1 h25min, 1 h45min (all time windows)
CSF (Striepens et al. 2013)	15, m	Between-subj, double-blind PL	Extraction; RIA	24 IU	45 min, 1 h, 1 h15 min	NA	only 1 h15 min
MRI (Paloyelis et al. 2016)	32, m	Between-subj design, single-blind PL	MRI: CBF	40 IU	25–38 min, 32–44 min, 39–51 min, 45–58 min, 52 min–1 h5min, 59 min–1 h11min, 1 h6min–1 h18min	39–51 min	39–51 min, 45–58 min, 52 min–1 h5 min
(Spengler et al. 2017)	116, m	Crossover, double-blind PL	MRI: BOLD	12/24/48 IU	24 IU: 15–38 min, 45 min–1 h8min, 1 h15 min–1 h38 min 12 IU: only 45 m–1 h8 48 IU: only 45 m–1 h8	NA	Only 45 min–1 h8min (only 24 IU, only in high-intensity fear condition)
(Martins et al. 2020b)	17, m	Crossover, double-blind PL	MRI: CBF	40 IU	15–23 min, 24–32 min, 35–43 min, 44–52 min, 1 h6min–1 h14min, 1 h15 min–1 h23min, 1 h27min–1 h35min, 1 h36min–1 h24 min	NA	15–23 min, 24–32 min, 35–43 min, 1 h27min–1 h35min

Approx. = approximately, BOLD = blood oxygenation level dependent, CBF = cerebral blood flow, CSF = cerebro-spinal fluid, RIA = radioimmunoassay, ELISA = enzyme linked immunosorbent assay, FEIA = fluorescent enzyme immunoassay, f = female, IU = international units, m = male, MRI = magnetic resonance imaging, NA = not available, PL = placebo, subj = subject. Search used Google Scholar with keywords "oxytocin" AND "human" AND "time" AND ("mri" OR "eeg" OR "blood" OR "saliva" OR "csf"), complemented with a further backward and forward reference search *This study used a breath powered closed-palate bi-directional device rather than the usual nasal spray.

Box 1

Systematic overview of previous findings addressing temporal pharmacodynamics of IN-OT in humans.

Saliva studies

Out of the five saliva studies (which given the OT administration was intranasal, warrant healthy skepticism—see below) three used 24 IU of IN-OT (Weisman et al. 2012; Daughters et al. 2015), one used 16 IU (Huffmeijer et al. 2012), and two compared the effect of different dosages: 24 IU versus 16 IU (van IJzendoorn et al. 2012), 24 IU versus 12 IU versus 48 IU (Spengler et al. 2017). The time span of measurements ranged from 15 min (Weisman et al. 2012; Spengler et al. 2017) to 7 h post-administration (van IJzendoorn et al. 2012). In all studies, saliva OT levels were significantly elevated during the whole observation period. The peak of the effect varied between saliva studies but was always at the earliest measurement point in all five studies, at 15 min (Weisman et al. 2012; Spengler et al. 2017), at 30 min (Daughters et al. 2015), at 1 h (van IJzendoorn et al. 2012), and at 1 h 15 min (Huffmeijer et al. 2012). However, caution interpreting the detected effects of IN-OT is needed, since the OT levels measured in the saliva may be partially originating from the nasal cavity to the back of the throat, rather than a consequence of the effect of OT on the brain (Daughters et al. 2015; Quintana et al. 2018). This argument is the main limitation of the saliva method—which seems difficult to control and thus we recommend that IN-OT studies refrain from measuring OT in saliva. Another limitation is the high rate of non-specific binding between non-OT compounds to OT antibodies in the preferred commercial saliva ELISA (enzyme linked immunosorbent assay) kit, which can lead to artificially elevated concentrations of OT in the sample (McCullough et al. 2013). The last issue was addressed in only two studies by using an improved (Daughters et al. 2015) or a different (Spengler et al. 2017) kit. Furthermore, three of the saliva studies (Huffmeijer et al. 2012; van IJzendoorn et al. 2012; Weisman et al. 2012) included female participants, and only van IJzendoorn et al. accounted for the menstrual phase. One of those (Weisman et al. 2012) mixed male and female participants and include sex in the analysis model. We argue that both sex and menstrual cycle phase should be taken into account because of is evidence that OT baseline levels are 3x higher in women than in men, as measured in plasma (Marazziti et al. 2019), that menstrual cycle impacts OT levels (Mitchell et al. 1981; Stock et al. 1991; Salonia et al. 2005), as well as several reports on sex-related differences in functional effects of OT (Evans et al. 2014).

Blood plasma studies

Three studies measured OT concentration in blood plasma (Gossen et al. 2012; Striepens et al. 2013; Spengler et al. 2017). All used only male participants and 24 IU (Striepens et al. 2013), 26 IU (Gossen et al. 2012), or 12/24/48 IU (Spengler et al. 2017) of IN-OT. In agreement with saliva studies, plasma studies have showed peak effects at the earliest measurement point of 15 min (Striepens et al. 2013) and 30 min (Gossen et al. 2012) post-administration. One study did not report the peak time window (Spengler et al. 2017). As for the duration of the effect, two studies observed significant differences from baseline at the latest measurement point: at 1 h45min and 1 h30 min, respectively (Striepens et al.

2013; Spengler et al. 2017). Gossen et al. had OT concentrations return to baseline levels at 1 h30 min for some participants and at 2 h30 min for all participants, even when the last measurement was at 3 h (Gossen et al. 2012). This highlights the previously noted (Quintana et al. 2018) tissue-dependent nature of OT pharmacodynamics, since saliva studies showed, instead, a significant effect up to 7 h post-administration. There was also another study (Quintana et al. 2015), which found a significant increase of OT in blood plasma at 10, 30, 60, and 120 min post-administration with 8 IU as well as with 24 IU (using a novel Breath Powered device, rather than the usual nasal spray).

Blood plasma studies have similar methodological drawbacks as saliva studies in terms of the inference of IN-OT's temporal dynamics in the brain. Nevertheless, even though plasma OT concentration does not necessarily reflect basal OT levels in the brain (McCullough et al. 2013; Leng and Ludwig 2016; Martins et al. 2020a), these have shown positive correlation after IN-OT and after stressor state measures (Valstad et al. 2017) and in some measurement methods (Lefevre et al. 2017). Second, some studies used non-specific antibodies, which may make the measurements less sensitive to the drug (Haraya et al. 2017). Third, some studies (Weisman et al. 2012; Daughters et al. 2015) did not use the OT extraction step prior to its quantification via enzyme immunoassay (EIA), which some authors suspect may render results invalid as extra immunoreactive products with similar structure to OT may also be inadvertently measured along with OT (McCullough et al. 2013). Commercially available EIA assays without extraction output values two orders of magnitude higher than those obtained using conventional RIA methods with extraction; and these extra immunoreactive products' levels do vary across individuals, and physiological states (McCullough et al. 2013). (Lyophilization has sometimes been used as a replacement of the extraction step in the ELISA—this may have been the case of one study (Daughters et al. 2015) although the references they used cast doubt; and was the case of another (Weisman et al. 2012) but the use was only in placebo samples, which could be an important confounding factor for their reported effects of drug.) Adding to this, two of the three studies used a small number of participants (Gossen et al. 2012; Striepens et al. 2013). Finally, authors pointed out (Gossen et al. 2012), plasma measurements are less indicative of OT availability in brain than CSF measurements (Mens et al. 1983) and thus IN-OT administration may affect neural function even after OT plasma levels have returned to baseline. To address this issue, one of those three studies (Striepens et al. 2013) added a measurement of OT in the CSF, to be compared with that in plasma, at half of the time windows (see below).

Cerebrospinal fluid studies

OT concentrations in the CSF were significantly increased only at 1 h15 min post-administration, i.e., the latest measurement point (Striepens et al. 2013). This is in contrast to OT plasma levels, which were elevated from 15 min to 1 h45 min. Also, there was no correlation between concentrations of OT in plasma and CSF samples, which raises an additional concern when it comes to comparing blood and brain findings; although a later study found levels of endogenous plasma OT to significantly and positively predict endogenous OT levels in CSF (Carson et al. 2015). One limitation of

Striepens et al. (2013)'s study was that due to the underlying invasive and complicated procedure, the CSF measurements could not be taken earlier than 45 min post-administration; however, as the authors state (Striepens et al. 2013), it is unlikely OT levels in CSF had risen and fallen before this time window. A methodological explanation the authors provide for the late onset of the OT increase in the CSF, compared to blood or saliva, is that the CSF measurement is taken via a lumbar puncture, which is down the spinal cord, and it might have taken that amount of time for OT, either exogenous (IN-OT) or exogenously stimulated (hypothalamically released due to stimulation by intranasal administration), to travel down the spinal cord (Striepens et al. 2013). In that case, an effect of IN-OT in the brain could have been achieved earlier.

Magnetic resonance imaging studies

The three studies that we are aware of which measured the functional influence of IN-OT administration on the human brain directly and across time (Paloyelis et al. 2016; Spengler et al. 2017; Martins et al. 2020b), all used MRI. Spengler et al. (Spengler et al. 2017) administered 24 IU and tracked the BOLD contrast in response to fearful faces images, from 15 min to 100 min post-administration. As expected from previous studies (Gamer et al. 2010), they observed the expected left amygdala inhibition response to fear to be most effective at the 45–70 min interval post-administration. This pattern was not observed with either lower (12 IU) or higher doses (48 IU), the latter dose showing in fact an opposite trend. This suggests a possible reversal of the functional effect of IN-OT for this higher dose and calls for extra care when interpreting studies that used different OT doses. This is in agreement with the inverted U-shaped dose-effect dependency of OT, whereby the observable effects of IN-OT diminish with high doses, which was put forward based on diverse evidence at both the neurophysiological (Wynn et al. 2019) and behavioral (Zhong et al. 2012); (Borland et al. 2019) levels. Overall, while this study adds an important insight on the temporal and dose effect of IN-OT, its focus on the amygdala leaves the question of IN-OT's effects on the rest of the brain open.

Another study (Paloyelis et al. 2016) recorded changes in CBF across the whole brain in a resting-state functional MRI (fMRI) session, after 40 IU of IN-OT/PL administration, and applied pattern recognition to predict probabilities of the nasal spray effect. After comparing the predicted to the administered nasal spray effect, they observed 80% classification accuracies in the IN-OT group across the whole post-administration period (25–78 min), which was significantly different from chance. In the group, they noticed 38%–81% accuracies, which was not different from chance except, surprisingly, for 32 to 44 min post-administration. They also localized the IN-OT across the whole post-administration period with univariate analysis and found significant results in four clusters spanning the midbrain, basal ganglia, limbic system, and cingulate, frontal, parietal temporal cortices and the cerebellum. This study (Paloyelis et al. 2016) provides an important insight into the effect on IN-OT at the whole brain level, but given the results of Gamer et al. (2010) (Gamer et al. 2010) it is not clear whether this result is representative of the lower and most commonly used (for research) dose of 24 IU or limited to the higher dose where the effect was not so clear (40 IU). Finally, although the authors found a significant

difference in CBF between the IN-OT and placebo groups at the pre-administration baseline, the classification accuracies in the IN-OT and in the groups were compared against their respective baselines.

Recently, the same research group (Martins et al. 2020b) compared the CBF resulting from the usual intranasal nose spray, intranasal nebulizer, and intravenous administration of, again, 40 IU of OT, in a time span from 15 min to 1 h24min post-administration. They report a significant effect of IN-OT, or a significant interaction between treatment and time, on separate brain clusters, in time windows 15–23 min, 24–32 min, 35–43 min, and 1 h27min–1 h35min post-administration—thus with a hiatus of 44 min to 1 h27min where no effect was detected. Results of Paloyelis et al. (2016) and Martins et al. (2020b) are not fully comparable or consistent (the first reports no effect from 25 to 44 min, it starting from then on to 1 h05min, while the later study showed a non-overlapping effect that starts as early as 15 min and last only up to 43 min), even though a similar experiment design and measure (CBF), experimental site, and a matching IN-OT dose (40 IU) was used. The second effect detected in the later study (starting as late as 1 h27 min) was in a timing not tested in the former study, so the replication would have not been possible. Location wise, both studies found largely overlapping and widespread regional effect, although only in the later study could they directly be related to the timeline results, since the former reports the analyses of time and location in separate, and different types of, models.

which OT has been measured. Plasma and saliva OT measurements seem not to represent well those in the brain, especially at basal or trait level (McCullough et al. 2013; Leng and Ludwig 2016; Martins et al. 2020a), even though there is evidence that plasma OT is positively associated with CSF OT: after IN-OT administration, after an experimental stressor (via meta-analysis (Valstad et al. 2017)), and when measurement methodology is carefully considered (Lefevre et al. 2017). Other limitations stemming from the current study pool include: 1) limited amount of subjects and lack of gender control; 2) non-specific binding agents in OT quantification in saliva and blood; 3) use of a range of OT doses between studies, whose effects cannot thus be inter-extrapolatable given evidence of an inverted-U shaped dose-effect relationship (Wynn et al. 2019); and 4) the problematic use of unextracted samples in fluid studies (McCullough et al. 2013).

Surprisingly, given its ability to record time-varying changes in neural activity (traceable to post-synaptic potentials of the pyramidal neurons (Kirschstein and Köhling 2009)) electroencephalography (EEG) has not yet been used to track pharmacodynamics of IN-OT; i.e., IN-OT induced changes in brain activity at different times following drug administration. This lack is even more relevant for the design and interpretation of IN-OT EEG studies, which have been steeply accumulating (Perry et al. 2010; Mu et al. 2016; Singh et al. 2016; de Bruijn et al. 2017; Luo et al. 2017; Rutherford et al. 2018; Schiller et al. 2019), and have assessed, for example, its effect on EEG time-frequency changes during videos of biological

motion (Perry et al. 2010) and images of sadistic context (Luo et al. 2017). These studies have mostly used 24 IUs, and healthy, predominantly male participants, with exceptions (Singh et al. 2016; Rutherford et al. 2018), and two used a resting-state paradigm (Rutherford et al. 2018; Schiller et al. 2019). For example, cross-frequency coupling between slow and fast waves has been found to be decreased under IN-OT, suggesting it may modulate integration of motivational and cognitive processes (Rutherford et al. 2018). This effect was apparent even at the latest point amongst EEG studies (70 min after drug administration); which mostly started at 40 min (Mu et al. 2016; de Bruijn et al. 2017; Luo et al. 2017) or 45 min (Perry et al. 2010; Singh et al. 2016; Schiller et al. 2019).

A novel and promising EEG analysis approach is based on the observation that the scalp voltage distribution, although dynamically changing over time, displays a limited number of characteristic topography patterns, so-called “microstates” (Lehmann et al. 1987). Four canonical microstate classes (labeled A, B, C, and D) have been identified and consistently reported across studies (Michel and Koenig 2018). Each microstate remains in a quasi-stable state for 60–120 ms and then rapidly transitions to a different microstate. Although the discreteness of individual microstates has also been questioned (Mishra et al. 2020), this approach has an important advantage over other EEG measures. First, since signals are simultaneously considered from across the entire scalp, large-scale brain networks can be assessed. Second, the four canonical resting-state microstates seem to correspond to well-known fMRI resting-state networks (Britz et al. 2010; Musso et al. 2010; Yuan et al. 2012; Milz et al. 2016; Custo et al. 2017). We have recently confirmed this by showing microstates to predict fMRI dynamic functional connectivity states with 90% accuracy, using simultaneous EEG and fMRI recording (Abreu et al. 2021). Furthermore, we have also highlighted the utility of microstates in identifying potential endophenotypes of psychiatric disorders, such as schizophrenia (da Cruz et al. 2020), for which OT has been proposed by others as a potential intervention to alleviate social impairments (De Berardis et al. 2013; Shilling and Feifel 2016). In sum, a microstates approach can be revealing of broad effects of IN-OT across the brain, and across post-administration time, with the potential of to bridge resting state findings across neuroimaging techniques, as well as to inform its therapeutic use. Despite that, so far only one resting-state EEG study (Schiller et al. 2019) measured the effect of IN-OT on human resting-state EEG using a microstates approach. They showed that IN-OT significantly increased duration of all microstates, decreased occurrence of microstates B and C, increased the contribution of microstate D, and decreased transitions from microstate B to C (see section Materials and Methods, for microstates features’ definitions). Nevertheless, no conclusions regarding the time course of IN-OT effects on the brain can be derived from this study,

as it uses only a single, 5 min-long (40–45 min post-administration), recording time window.

In the present study, we aimed to characterize the temporal dynamics of IN-OT using microstate analysis of EEG data. We tracked IN-OT-induced changes across the whole brain, and across almost 2 h of experimental time, on microstates function during eyes-open and eyes-closed resting-state. We used 24 IU with a throughout double-blind randomized placebo-controlled cross-over design (in 6 time windows: 15–27, 30–42, 45–57, 60–72, 75–87, and 90–102 min, post-administration), in a young male sample, at the same time of day. The use of a dose commonly employed in neuroscience research brings the advantage of aiding interpretation of previous both positive and negative behavioral and brain phenotype findings. We collected data from both eyes open (EO) and eyes closed (EC) conditions, as there is evidence of different microstate patterns between the conditions (Seitzman et al. 2017), as well as across-scalp means of EEG frequency waves and their topographies (Barry et al. 2007), and because IN-OT effects could not be assumed to be the same across eye statuses. In addition, we briefly discuss the role of OT on the specific microstates in relation to the cognitive network function they may reflect. We hope our findings can inform comparability between past studies, and the design of future IN-OT studies, especially those aiming to employ psychological tasks and thus naturally concerned with the start, duration, and end of task presentation and psychophysiological recording times; as well as to entice hypothesis-generating reflections on the role of OT in human cognition.

Materials and methods

Participants

We recruited 20 young healthy, male, Portuguese adults, one of which was excluded due to EEG recording problems, through mailouts and pamphlets in the university community and online social networks. The participants used for analysis ($n = 19$) were on average 27 years old (mean = 27.1, SD = 4.02, range = 18–35 y). We applied the following exclusion criteria: history of endocrinological, cardiovascular, or neurological disorders, substance abuse, blocked nose; use of cannabis 2 weeks prior, and alcohol consumption, drugs or medication 24 h prior, and smoking 2 h prior to the experimental session; caffeine consumption or heavy physical exercise or sexual activity on that day. Screening for exclusion and inclusion criteria was performed via self-report during an initial phone interview and in person, immediately prior to the recording. All participants gave their written informed consent, received financial compensation for their time. The study was approved by the Ethics Committee of the Lisbon Medical Academic Center (Centro Académico Médico de Lisboa, CAML) and complies with national and EU legislation for clinical research.

Experimental procedure

The experimental session took place at a quiet room of the CAML's Clinical Research Centre (Centro para Investigação Clínica) in the Hospital de Santa Maria, Lisbon, Portugal. We used a double-blind (through-out data collection up to statistical analysis, inclusive), randomized placebo-controlled, cross-over design, whereby each participant took part in two sessions: one for IN-OT and once for placebo administration, in a counterbalanced order. The IN-OT administration of 24 IU was via 3 puffs of 0.1 mL each, in each nostril, from a 40 IU 5 mL Syntocinon bottle (using the Novartis formula—batch H5148 produced by Huningue Production, France) or an identical placebo bottle (with the same ingredients, except OT—batch 170317.01 produced by VolksApotheke Schffhausen, Switzerland), both supplied by Victoria Apotheke Zürich, Switzerland. Drug acquisition, storage, and randomization of drug administration was performed and controlled by the hospital's pharmacy. IN-OT/placebo administration was at 2:22 pm (SD=29 min) for all participants, and EEG recording started 12 min prior to the drug administration, to restrict the impact of the circadian rhythm on baseline endogenous OT levels (Perlow et al. 1982; Reppert et al. 1984; Devarajan and Rusak 2004). Upon arrival, participants confirmed all eligibility criteria via questionnaire and their health state was assessed via medical examination, which included heart rate, blood pressure, and electrocardiogram measurements. In the second session, only the eligibility questionnaire was administered. OT and placebo sessions were seven days apart (mean = 7.3, SD = 1.95 days).

During the 7 resting-state EEG recording windows (−12–0 min pre-administration and 15–27, 30–42, 45–57 min, 1–1 h 12min, 1 h 15 min–1 h 27min, 1 h 30 min–1 h 42min after administration), participants were instructed to stay still, not engage in any cognitive process, and to practice meditation or mind-wandering (more details in [Supplementary Figure S1](#)). Each recording window (“time window”) included 5-min recording with eyes closed, followed by 5-min recording with eyes open. Participants compliance with eye-status instructions was confirmed via an eye tracker (EyeLink 1000 plus, SR Research, Canada). At the end of each time window, participants filled three Likert scales: for alertness (1—alert, 5—sleepy), excitement (1—excited, 5—calm), and desire to socialize (1—desire to socialize, 5—desire to be left alone).

EEG data acquisition and processing

We recorded the EEG signal at a 1000 Hz sampling rate using a 64-channel Brain Vision actiCHamp system (Brain products, München, Germany). Electrodes were placed according to the 10/20 system, with the ground electrode at the central front location on the cap. To ensure optimal signal quality, we aimed to calibrate all electrodes below 5 k Ω , and kept all electrodes below 44 k Ω for all participants. During recording, electrodes

on the left and right mastoid served as a reference. We placed horizontal electrooculogram and vertical electrooculogram electrodes to record eye-movements and blinks; however, due to excessive noise, these data were discarded and ocular artifacts were removed using Independent Component Analysis (ICA; see next section).

Offline EEG data were preprocessed in Matlab, version R2019b, using an automatic pre-processing pipeline (APP; da Cruz et al. 2018) based on EEGLAB toolbox (Delorme and Makeig 2004) v12. We designed this pipeline specifically for, and tested it on, resting-state data; and it was shown to perform similarly to supervised pre-processing done by EEG experts, and to outperform existing alternatives (Bigdely-Shamlo et al. 2015; Hatz et al. 2015) in terms of the amount of data lost. The pre-processing consisted of the following steps: 1 to 30 Hz bandpass filtering, powerline noise removal, down-sampling to 250 Hz, re-referencing to the bi-weight estimate of the mean of all channels, removal and 3D spline interpolation of bad channels; removal of bad epochs; independent component analysis (ICA) to remove eye movement, muscular and bad channel related artifacts; re-referencing to common average reference. After automatic pre-processing, data were visually inspected for any additional artifacts, which were removed if needed. We decided, posterior to data collection and preprocessing but prior to data analysis, to limit our analysis to the first artifact-free 176 s, of each condition, in each time window, in order to: 1) minimize effects of drowsiness or sleepiness (which may take place after 3 min of non-stop recording (Tagliazucchi and Laufs 2014)), and 2) to be directly comparable to the (only) previous IN-OT EEG microstates study (Schiller et al. 2019), which used these data average length.

Microstate extraction and quantification

We extracted microstates of each time window and each condition (eyes-closed/eyes-open) separately, using the Microstates toolbox (Koenig 2017) for EEGLAB. First, we further bandpass-filtered the data from 2 to 20 Hz, as commonly done in the literature (Koenig et al. 2002; Lehmann et al. 2005; Pascual-Marqui et al. 2014; Milz et al. 2016), including in the only existing study that employed microstates in IN-OT research (Schiller et al. 2019). Next, for each of the seven time windows, we concatenated each subject's epochs into one and calculated the Global Field Power (GFP) across all electrodes over this observation period (that equaled 176 s), for eyes closed and open separately. GFP is calculated as the standard deviation across electrodes at a given time window and it quantifies the overall potential strength across the given set of electrodes. Local maxima of GFP represent the highest signal-to-noise ratio, and are associated with a stable EEG topography (Lehmann et al. 1987).

Next, in moments of time that corresponded to local GFP maxima in each time window, we extracted scalp topographies and clustered them into four classes, A, B, C, and D (following the previous literature, which converged on four optimal microstate classes; see 37,52,53),

using the atomize-agglomerate hierarchical clustering algorithm (AAHC; (Murray et al. 2008; Tagliazucchi and Laufs 2014)). The clustering was done on an individual level, and then across all participants in both recording sessions (but separately for each time window). Next, we assigned the microstate labels A, B, C, and D to the grand-grand average of both sessions based on visual inspection of the grand-grand average maps, and on their similarity with the 4 canonical microstates found in the literature (Michel and Koenig 2018), and back-propagated the sorting to the individual maps (i.e., participants' topographies at the GFP peaks were labeled as microstates A, B, C, and D based on spatial correlation with the grand-grand average). In both sessions, the similar (and thus comparable) four microstates resembled the four class model maps consistently identified in the literature (Koenig et al. 2002); (Michel and Koenig 2018)—see [Supplementary Figure S2](#). Using spatial correlation, we confirmed that the microstates in IN-OT and placebo sessions were (see [Supplementary Material B](#) for detailed analysis and results). The spatiotemporal dynamics of the four microstates categories were quantified as standard (Michel and Koenig 2018) by measuring their: 1) duration (s), i.e., the mean time the microstate was present; 2) occurrence (times/s); i.e., the mean frequency it was present; 3) contribution (%); i.e., the proportion of total time it was present; and (4) delta transition probability from one microstate to another (%); i.e., the relative amount of times microstate X was followed by microstate Y, calculated as the difference between the observed and the predicted value given previously observed occurrences.

Statistical analysis

We designed a series of linear mixed models (LMM) using R (*lme4* package and the *lmer* function (Bates et al. 2015)); see [Supplementary Material C](#) for syntax and full results). These models allow for the presence of missing data, as well as to specify drug-varying covariates (such as the baseline values, which differed in each drug session per participant), and take into account inter-individual random differences. Models were run separately for each microstate feature as dependent variable (duration, occurrence, contribution) of each microstate (A-D), and for microstate-to-microstate transition probability (in 12 models, i.e., one per every possible transition pair). Each model estimated the fixed effects of the categorical factors Drug session (IN-OT, placebo) and Time (post-administration time windows: 1, 2, 3, 4, 5, 6), and their interaction. For baseline correction, the microstate feature at time 0 (i.e., before administration) in the placebo and the IN-OT session was included as a covariate of no interest in each model. We used an identical LLM to analyze behavioral scales (one model per scale). The number of degrees of freedom, and consequently *P*-values, were calculated using Type III analysis of variance with the Satterthwaite's method. If main effects of drug and/or

interactions between drug and time were statistically significant (uncorrected *P*-value < 0.05), we performed pairwise comparisons on estimated marginal means (degrees of freedom estimated using the Kenward–Roger method) using EMMEANS package for R. The main effects of time were not interpreted, as it was not relevant to our hypothesis—but is reported as [Supplementary Material C](#). Finally, although the drug inhalation procedure was standardized, we further ensured that there was no confounding effect due to differences in the amount of inhaled IN-OT (details in [Supplementary Material B](#)).

Results

All main effects and interactions, for both the eyes-closed and eyes-open conditions, for each microstate feature and for each microstate (A, B, C, and D), are reported in [Supplementary Material C](#) and summarized in [Table 2](#). For eyes-closed, the results for contribution, duration, and occurrence are plotted in [Figure 1](#), and for both eyes-closed and eyes open status, transition probabilities are plotted in [Figure 2](#) (no other effects being statistically significant in this condition). Below, we describe all statistically significant effects (or notable trends) of IN-OT (main or interactions), there being none on mood scales.

Eyes-closed data

Microstate A

Duration of microstate A was longer in the IN-OT ($M = 0.087$, $SE = 0.002$) than the placebo session ($M = 0.085$, $SE = 0.002$; $F(1, 190.4) = 5.05$, $P = 0.026$) irrespective of time. Pairwise comparisons on estimated marginal means indicated that microstate A duration was longer for IN-OT than placebo, particularly in time window 5 ($t(189) = 1.98$, $P = 0.049$). The occurrence of microstate A appeared higher under IN-OT ($M = 2.320$, $SE = 0.061$) compared to placebo ($M = 2.250$, $SE = 0.060$), irrespective of time, but this difference was not statistically significant ($F(1, 194) = 3.83$, $P = 0.052$). Contribution of microstate A was larger for IN-OT ($M = 0.21\%$, $SE = 1\%$) than placebo ($M = 19\%$, $SE = 1\%$; $F(1, 190) = 8.57$, $P = 0.004$). Pairwise comparisons between IN-OT and placebo at separate time windows did not reveal significant effects.

Microstate B

Duration of microstate B was shorter under IN-OT ($M = 0.085$, $SE = 0.002$) than placebo ($M = 0.088$, $SE = 0.001$; $F(1, 194.3) = 5.82$, $P = 0.017$) irrespective of time. Duration was shorter under IN-OT than placebo particularly in time window 4 as revealed by pairwise comparisons (1–1 h12min post-administration; $t(190) = -2.31$, $P = 0.022$). Contribution of microstate B was smaller in IN-OT ($M = 19\%$, $SE = 1\%$) than placebo ($M = 20\%$, $SE = 1\%$; $F(1, 193.5) = 10$, $P = 0.002$) also particularly in time window 4 (1 h–1 12 min; $t(190) = -2.49$, $P = 0.013$).

Table 2. Summary of selected microstate features and transition probabilities' main effects of drug (intranasal oxytocin vs. placebo), or interaction between drug and time, and the corresponding pairwise comparisons between IN-OT and placebo in time windows where the comparison was significant ($P < 0.05$, with the two borderline-significant trends also described). Time windows: 1 = 15–27 min after drug (IN-OT/PL) administration, 2 = 30–42 min, 3 = 45–57 min, 4 = 1 h–1 h12min, 5 = 1 h15 min–1 h27min, 6 = 1 h30 min–1 h42 min; MS = microstate. D = drug, T = time, IN-OT = intranasal oxytocin.

Microstate label	Microstate measure	IN-OT effects	Time windows (pairwise comparisons)	Direction
Closed eyes				
MS A	Duration	D: $F(1, 190) = 5.05, P = 0.026$	T5: $t(189) = 1.98, P = 0.049$	IN-OT ↑
	Occurrence	n.s. [$F(1, 194) = 3.83, P = 0.052$]	n.s.	IN-OT ↑
MS B	Contribution	D: $F(1, 190) = 8.57, P = 0.004$	n.s.	IN-OT ↑
	Duration	D: $F(1, 194) = 5.82, P = 0.017$	T4: $t(190) = -2.31, P = 0.022$	IN-OT ↓
	Occurrence	n.s.	n.s.	-
MS C	Contribution	D: $F(1, 193) = 10, P = 0.002$	T4: $t(190) = -2.49, P = 0.013$	IN-OT ↓
	Duration	D: $F(1, 188) = 6.68, P = 0.011$	T2: $t(188) = -3.04, P = 0.003$	IN-OT ↓
	Occurrence	n.s.	n.s.	-
MS D	Contribution	$F(1, 189) = 10.59, P = 0.001$	T1: $t(188) = -2.34, P = 0.020$	IN-OT ↓
	Duration	n.s. [D: $F(1, 191) = 3.60, P = 0.059$]	T2: $t(188) = -2.53, P = 0.012$	IN-OT ↓
	Occurrence	D: $F(1, 186) = 4.36, P = 0.038$	n.s.	IN-OT ↑
Transitions	Contribution	D: $F(1, 191) = 5.65, P = 0.018$	n.s.	IN-OT ↑
	C → B	D: $F(1, 190) = 4.58, P = 0.034$	T5: $t(189) = -2.82, P = 0.005$	IN-OT ↓
	C → D	D: $F(1, 191) = 4.06, P = 0.045$	T6: $t(189) = 2.34, P = 0.021$	IN-OT ↑
Open eyes				
Transitions	A → C	D*T: $F(5, 188) = 2.95, P = 0.014$	T6: $t(189) = 2.34, P = 0.021$	IN-OT ↑
	A → D	D*T: $F(5, 188) = 2.30, P = 0.046$	n.s.	N/A

Microstate C

Duration of microstate C was shorter under IN-OT ($M = 0.106, SE = 0.004$) than placebo ($M = 0.110, SE = 0.004$; $F(1, 188) = 6.68, P = 0.011$), particularly in time window 2 (30–42 min; $t(188) = -3.04, P = 0.003$). Contribution of microstate C was smaller in the IN-OT ($M = 32\%, SE = 1\%$) session than placebo ($M = 34\%, SE = 1\%$; $F(1, 189) = 10.59, P = 0.001$), particularly in time windows 1 (15–27 min; $t(188) = -2.34, P = 0.0203$) and 2 (30–42 min; $t(188) = -2.53, P = 0.012$).

Microstate D

The duration of microstate D was longer for IN-OT ($M = 0.099, SE = 0.002$) than placebo ($M = 0.096, SE = 0.002$), irrespective of time, but this did not reach statistical significance ($F(1, 191.3) = 3.6, P = 0.059$). Occurrence of microstate D was increased in IN-OT session ($M = 2.88, SE = 0.059$) compared to placebo ($M = 2.8, SE = 0.059$; $F(1, 186) = 4.36, P = 0.038$), particularly in time window 2 (30–42 min; $t(188) = 2.52, P = 0.013$). Contribution of microstate D was larger for IN-OT ($M = 28\%, SE = 1\%$) than placebo ($M = 27\%, SE = 1\%$; $F(1, 191.2) = 5.65, P = 0.018$); pairwise comparisons did not point to significant time windows.

Transition probabilities

Transition C → B was decreased for the IN-OT ($M = -0.006, SE = 0.001$) compared to placebo ($M = -0.005, SE = 0.001$; $F(1, 189.85) = 4.58, P = 0.034$), particularly in

time window 5 (1 h15 min–1 h27 min; $t(189) = -2.82, P = 0.005$). Transition C → D was increased for IN-OT ($M = 0.002, SE = 0.001$) compared to placebo ($M = -0.0002, SE = 0.001$; $F(1, 191.18) = 4.063, P = 0.045$), particularly in time window 6 (1 h30 min–1 h42 min; $t(189) = 2.34, P = 0.021$).

Eyes-open

Transition probabilities

Transition A → C showed a drug by time interaction ($F(5, 188) = 2.95, P = 0.014$), such that IN-OT elicited increased transition probability ($M = -0.003, SE = 0.002$) relative to placebo ($M = -0.007, SE = 0.001$), in time window 6 (1 h30 min–1 h42 min; $t(192) = 2.15, P = 0.033$) whilst the opposite (non-significant) trend was seen in time windows 2, 4, and 5, from inspection of Figure 2. Transition A → D also showed a drug by time interaction ($F(5, 187.76) = 2.30, P = 0.046$). Drug effect was not significant at any individual time window, but, from inspection of Figure 2, time window 5 is the only showing a trend, whereby IN-OT shows a greater transition probability ($M = -0.002, SE = 0.002$) vs. placebo ($M = -0.004, SE = 0.002$).

Discussion

We used EEG to characterize the temporal dynamics of IN-OT in humans for the first time, to our knowledge. We tracked IN-OT-induced microstates dynamic changes

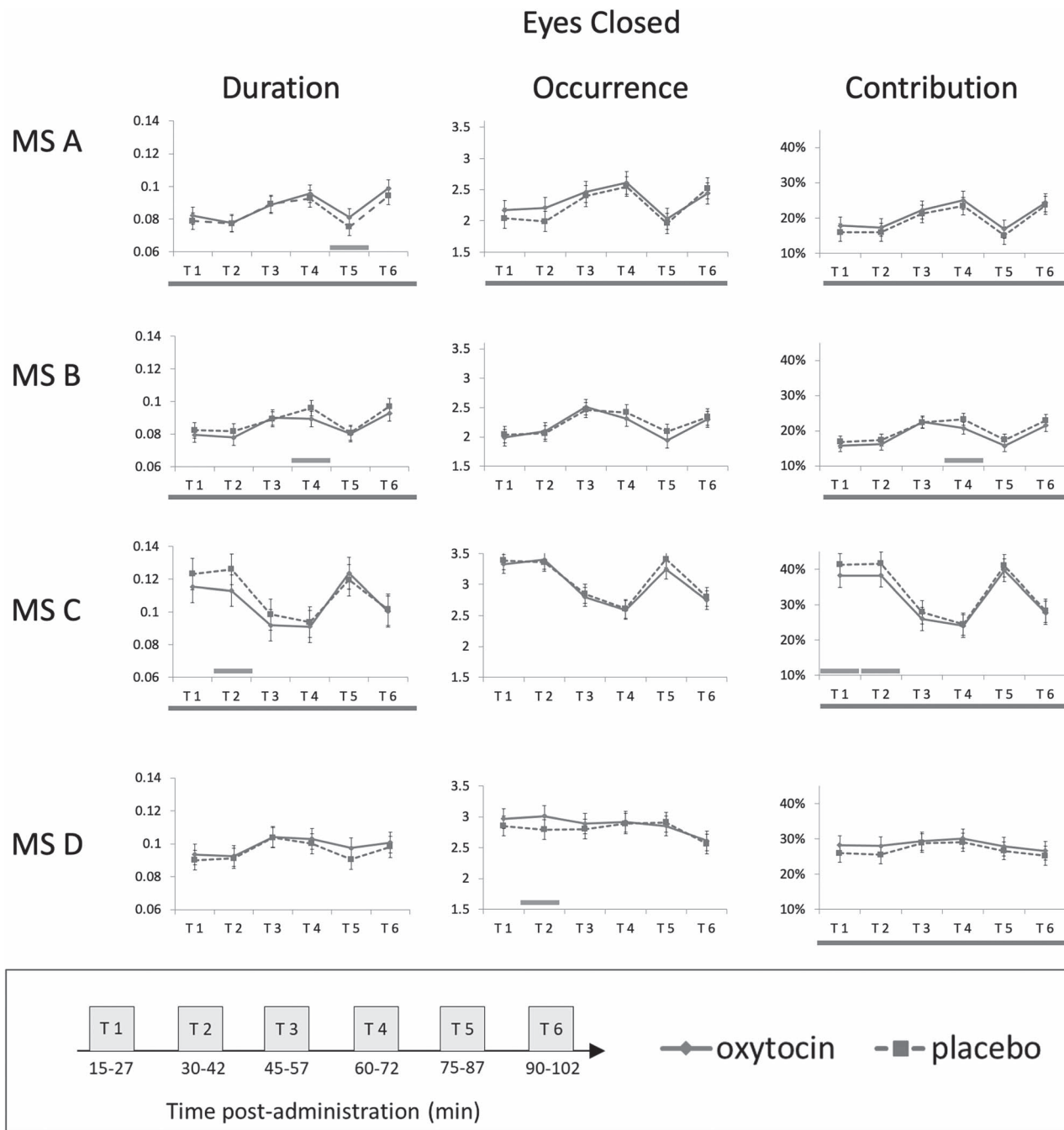


Fig. 1. IN-OT- and PL-induced changes in the microstate features for the eyes-closed condition over the post-administration observation period. Dashed line corresponds to placebo and continuous line to oxytocin. For each subplot, bottom line across all time windows signals a main effect of drug. Significant pairwise comparisons (oxytocin vs. placebo) at specific time windows are signaled with a green bar above the x axis. Error bars: 95% CI. Time windows: 1 = 15–27 min after drug (IN-OT/PL) administration, 2 = 30–42 min, 3 = 45–57 min, 4 = 1 h–1 h12min, 5 = 1 h15 min–1 h27min, 6 = 1 h30 min–1 h42 min. MS = microstate.

across the whole male brain cortex (using the dose of 24 IU, most commonly used in neuroscience research, so findings can be of relevance to most past and expected future studies), and across time, during resting-state, from 15 min to 1 h42min (in 6 time windows) post-administration. We found the effects were widespread but diverse, meaning that they were observed on different features of different microstates, between time

windows. Next, we first consider a “global” (or any) difference between IN-OT and placebo across time—and compare those results against earlier saliva, blood and MRI pharmacodynamic studies (illustrated in Fig. 3). Second, we discuss the “specific” microstate effects found and their direction and suggest how they can generate or consolidate previous hypotheses for a role of OT in human neurobiology.

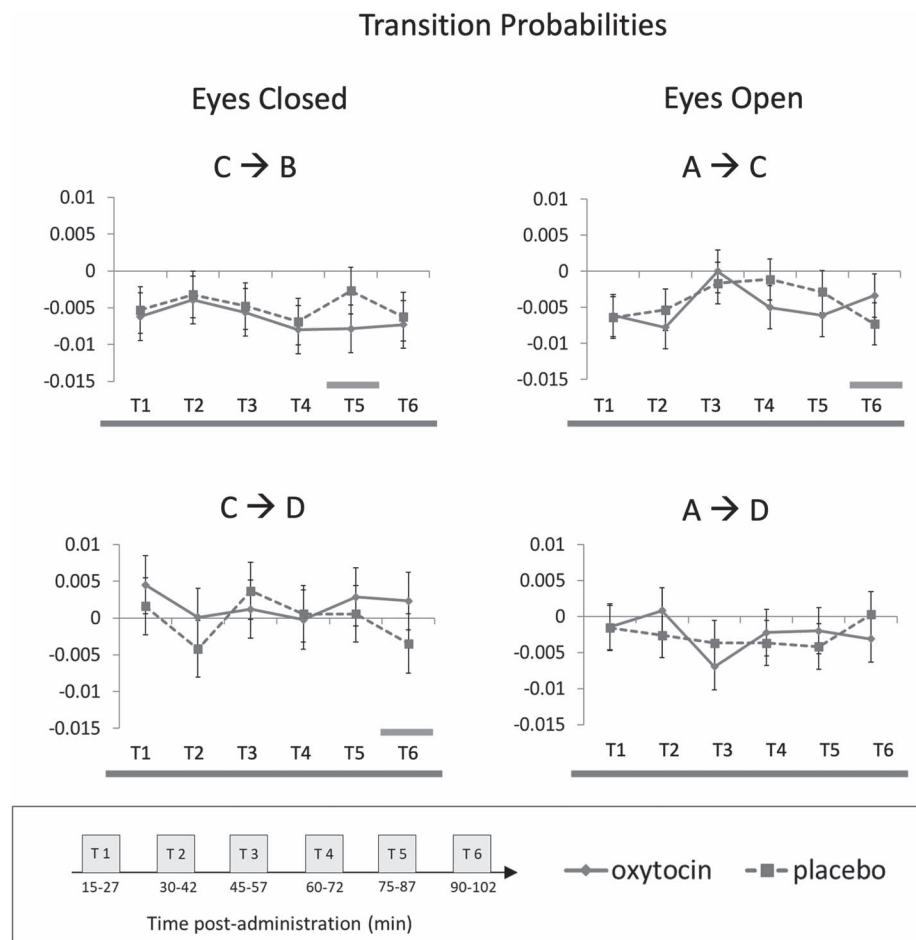


Fig. 2. IN-OT- and PL-induced changes in transition probabilities for eyes-closed and eyes-closed recording, where main effects of drug or drug \times time interaction was detected. Dashed line corresponds to placebo and continuous line to oxytocin. For each subplot, bottom line across all time windows signals a main effect of drug or interaction. Significant pairwise comparisons (oxytocin vs. placebo) at specific time windows are signaled with a bar above the x axis. Error bars: 95% CI. Time windows: 1 = 15–27 min after drug (IN-OT/PL) administration, 2 = 30–42 min, 3 = 45–57 min, 4 = 1 h–1 h12min, 5 = 1 h15 min–1 h27min, 6 = 1 h30 min–1 h42 min. MS = microstate.

IN-OT altered microstate dynamics across most of 15 min to 1 h 42min post-administration

Overall, we found IN-OT delivery to have functional effects at the resting state brain that started at the same time as reported OT detection in saliva (Huffmeijer et al. 2012; van IJendoorn et al. 2012; Weisman et al. 2012; Daughters et al. 2015; Spengler et al. 2017), blood plasma (Landgraf 1985; Gossen et al. 2012; Striepens et al. 2013; Quintana et al. 2015; Spengler et al. 2017) (15 min) and the most recent MRI study (as discussed below in detail) (Martins et al. 2020b). This onset time is difficult to compare with that of the (only) CSF study as the latter started measuring OT only at 45 min (and only detected at 75 min—with the limitation of the collection being at the “brain-distant” lumbar region) (Striepens et al. 2013). This effect continued at least until 1 h42min (the end of our recording), with an interruption (45–57 min after IN-OT administration). Thus, although there is some alignment between pharmacodynamics studies across-tissues, in terms of onset, care is advised when inferring OT’s availability, or effect, across tissues (see Fig. 3 for study comparison). In more detail, we

found a statistically significant main effect of IN-OT on: contributions of all four canonical microstate classes, duration of microstates A, B, and C, and occurrence of microstate A and D, and transition probabilities (C \rightarrow B, and C \rightarrow D), during eyes-closed resting state. During eyes-open, only transition probabilities were significantly different between IN-OT and placebo (A \rightarrow C and A \rightarrow D). When looking in time windows separately, our results showed statistically significant effects of IN-OT during eyes-closed in all time windows except time window 3 (45–57 min); and for eyes-open, in time windows 4 (1 h–1 h12 min) and 6 (1 h30 min–1 h42) (see Fig. 3 for a summary). This may suggest a possible decreased (“dip”) effect of IN-OT in the middle time window (45–57 min); however, this was not statistically supported given that no drug by time interactions were statistically significant (further discussed ahead). In terms of main effects of time on microstates dynamics, these were statistically significant, which was expected given the session’s long duration and thus likelihood to induce changes in arousal, drowsiness, etc.—over time. Since our focus was on the effects of drug, we

Overview of research on temporal dynamics of intranasal oxytocin (IN-OT) in humans

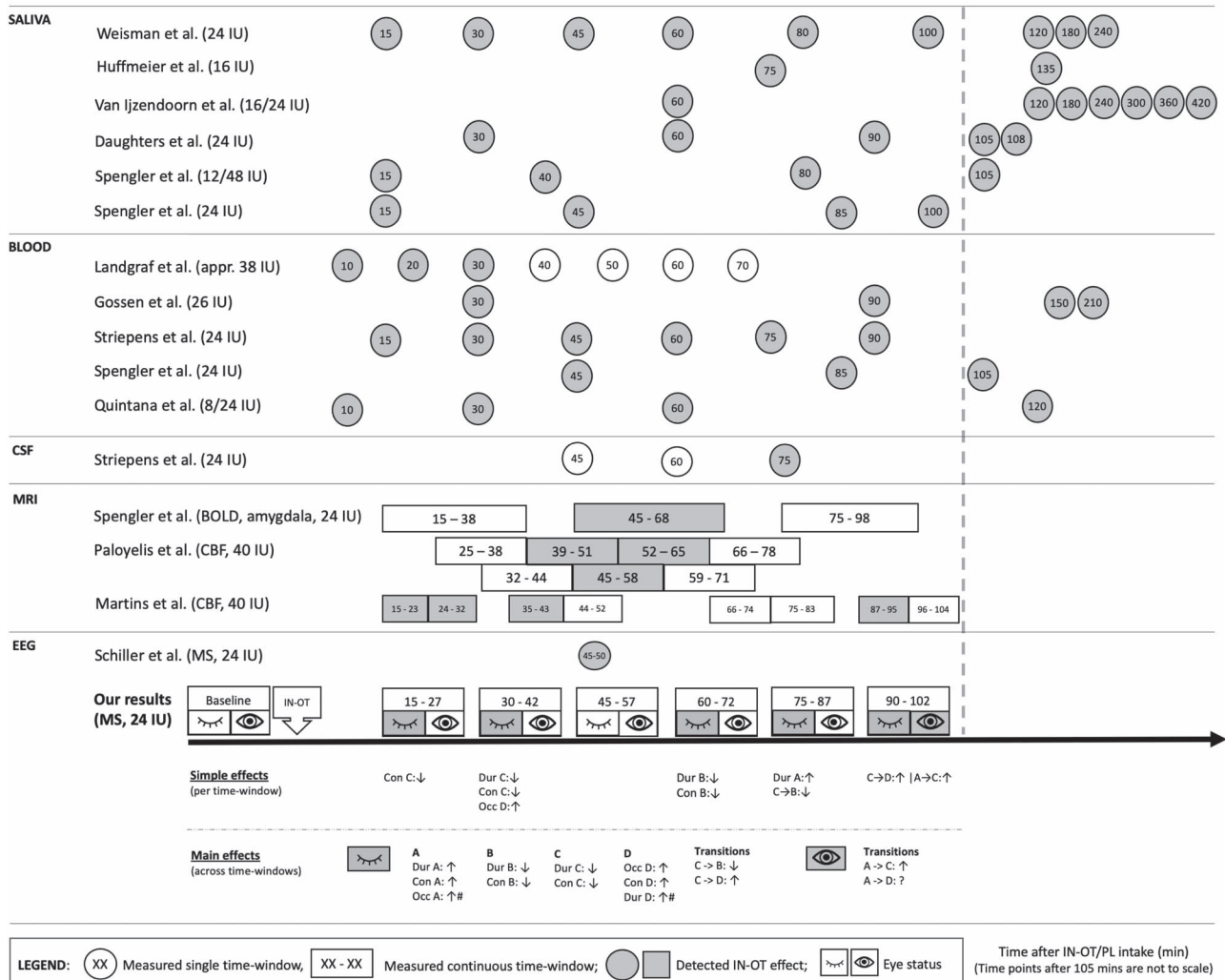


Fig. 3. An overview of our experiment’s time frame (time windows with a significant, $P < 0.05$, effect of drug are highlighted in grey, including two borderline-significant trends distinguished with #) in comparison with that of all previous pharmacodynamics studies and the only microstates EEG using IN-OT (time windows with a significant effect of drug are highlighted in grey). Appr. = approximately, BOLD = blood oxygenation level dependent, CBF = cerebral blood flow, CSF = cerebrospinal fluid, EEG = electroencephalography, IN-OT = intranasal oxytocin, IU = international units, MRI = magnetic resonance imaging, MS = microstates, PL = placebo, Dur = duration, Con = contribution, Occ = occurrence, Tran = transition.

refrained from discussing these results but provide them as supplementary material, so they can be used in future research pertaining to the temporal stability of microstates.

Cognitive interpretation of IN-OT effects on specific microstates

To procure a hypothesis-generating cognitive meaning for our resting-state microstates findings, and because their employment in neuroscience has been recent, it may be useful to look into resting-state MRI findings—with the caveat that inferences should remain speculative given the still small body of evidence relating both data modalities. Previous studies using both EEG resting-state microstates and MRI resting-state networks, recorded during closed eyes, via spatial correlation (Khanna et al. 2008; Britz et al. 2010; Michel

and Koenig 2018) have suggested that: 1) Microstate A is correlated with the auditory and phonological processing network; 2) microstate B with the visual network; and 3) microstate C (albeit by far the least well understood (Michel and Koenig 2018; Krylova et al. 2021)) tentatively with subjective representation of one’s own body (Taylor et al. 2009; Britz et al. 2010), but also with the part of the default mode network linked to self-referential thoughts (Xu et al. 2016) and internally guided cognition (Andrews-Hanna 2012) thus possibly processes directed at one’s self, rather than the external world (Schiller et al. 2019) (but also see (Pipinis et al. 2017) for evidence of a negative link somatic awareness and positive with vigilance levels (Krylova et al. 2021)); and 4) microstate D with the attention network, in particular the reflexive detection of relevant external stimuli, attention-switching and reorientation (Corbetta

and Shulman 2002; Britz et al. 2010), and vigilance level (Krylova et al. 2021).

Our data suggest that IN-OT effects on specific microstates may not hold the same profile throughout a 2 h experimental session. Nevertheless, overall (i.e., irrespective of time window), our eyes-closed data, showed IN-OT to induce a greater involvement of microstate A and D, and diminished involvement of microstate B and C (see Fig. 3). The first pattern was reflected in increased duration and greater contribution of microstate A, and longer duration, more frequent occurrence, and larger contribution of D. In light of the microstates and fMRI network findings above-summarized, this would suggest a role of OT in enticing greater resource to language brain resources (linked to A) and to vigilance and external attention-orienting (linked to D)—in order to tune it for the reception of external stimuli, particularly those of the social kind, which are those requiring phonological awareness. As such, our data support both the social saliency (Shamay-Tsoory and Abu-Akel 2016) and the social approach hypotheses of OT (Cohen and Shamay-Tsoory 2018). These cognitive models posit that OT promotes saliency and approach to social stimuli, respectively. Regarding eyes-open, we (only) found increased transition frequency from A to C and to D, suggesting, rather, an IN-OT-provoked *diminished* involvement of A in this eye status, possibly because attention to phonological signs (being mainly auditory in normal-sighted people) is less relevant than in a closed-eyes state (albeit eye-states were not statistically contrasted). The second IN-OT effect pattern we observed, again in eyes-closed, was of a decreased duration and smaller contribution of B and C (and concurrently, a tendency for C to transition more frequent to D, and less to B). This may reflect a lower engagement of visual processing resources (linked to B), which would represent higher resource allocation efficiency given subjects have their eyes closed; and of self-oriented attention (linked to C), but we remind the reader that microstate C is particularly poorly understood.

Notably, our results partially replicate, albeit not at the same time window, the so far single IN-OT microstates study (Schiller et al. 2019). Within the same (and only) time window of Schiller et al.'s (2019) study (45–50 min), we found no effects for either microstate. However, our results' overall pattern appears to partially agree with that study's, in that IN-OT also showed: 1) an increased duration of microstate A (albeit they also report it for B, C, and D); 2) a lower contribution of C; and 3) a higher contribution of D. The authors suggested that the observed increased temporal stability (i.e., duration) of microstates reflects the well-known IN-OT anxiolytic effects (Neumann and Slattery 2016) and a shift from internally- to externally-oriented processing modes at rest (as reflected by a decreased presence of microstate C—linked to self-oriented cognition—in favor of external attention-related microstate D), which our data supports. Some differences between studies exist, as Schiller et al.

(2019) (Schiller et al. 2019) observed IN-OT to induce lower occurrences of B and C, while we observed lower contributions of both, and increased durations of B, C, and D; and we observed higher transition probability from C to A. These disparities may or may not be due to methodological differences: besides using a single time window, that study did not adjust measurements for their pre-administration recording, and used ANOVA; whilst we collected data continuously, and used LMM and baseline correction by entering a pre-administration measurement as a covariate of no interest in our model, specific to each session.

Lastly, our recent report of an increased contribution of microstate C and decreased of D in schizophrenia patients (da Cruz et al. 2020) may suggest a promising therapeutic role for IN-OT in the illness, given that we herein find it to have an exactly reverse, thus potentially compensatory, effect. A potential of IN-OT to therapeutically target abnormal microstate dynamics in this disorder is in line with others' findings (De Berardis et al. 2013; Shilling and Feifel 2016).

Is there an IN-OT effect “dip” at 45–57 min?

Our particular finding of a lack of statistically significant time window-specific (i.e., pair-wise) effects of IN-OT on microstates in the middle time window (45–57 min), both during closed and open eyes, could not be supported by drug by time interactions (as none was statistically significant)—which could have been due to a lack of power in our design. Nevertheless, the possibility of such an effect “interruption or sharp decrease” merits, some discussion, given some previous (and recent) evidence that seems to support it. Although most studies, in a range of different tissues, do not report such a “dip” (Fig. 3), one blood study (Landgraf 1985), the CSF study (Striepens et al. 2013) and, most interestingly, the most recent fMRI study (Martins et al. 2020b) have also not detected OT's availability/effect in the 45–57 min approximate middle-window of time (with statistically significant drug by time interaction). Using a very similar timeline design (but a higher dose of 40 IU), Martins et al. (2020b) reported an effect at 15–43 min and at 1 h27min—1 h35min after administration, and, also similarly to us, not at 44 min—1 h23 min. This means that the abovementioned IN-OT effect hiatus started in both studies at exactly the same time, and encompassed the 44–45 min time window. (Their effect also lasted longer (49 vs. 18 min in our study), possibly due to the higher dose they used.) A possible explanation, also put forward by those authors, is based on two complementary effects of the IN-OT on the brain. The earlier effect of IN-OT (at 15–42 min herein, and 15–43 min in (Martins et al. 2020b)), could be due to the exogenous, inhaled, IN-OT. After the subsequent “dip,” the later window of effect (at 1 h–1 h42min herein, and at 1 h27min—1 h35min in (Martins et al. 2020b)) could be due to the “feed forward” release of endogenous OT, a mechanism previously suggested (Churchland

and Winkelman 2012), whereby administration of the exogenous IN-OT stimulates the release of the endogenous OT (as social-triggered visceral reactions do, via the vagus nerve (Churchland and Winkelman 2012)). Alternatively, but not mutually exclusively, perhaps such “dip” is explained by the inverted U-shape IN-OT concentration-effect model, which posits that when IN-OT accumulates the highest in the synaptic cleft, OT receptor (OTR) activity diminishes temporarily—which could have happened in the middle time window. As previously suggested (Mustoe et al. 2019), a higher IN-OT concentration tends to activate the similarly structured vasopressin receptors, which may reduce OT receptor (OTR) signaling (Borland et al. 2019); or can also lead to OTR desensitization, due to: 1) causing it to couple to different G-protein subtypes (Borland et al. 2019) (leading to desensitization from as early as 5 min of OT stimulation in human embryonic kidney cells (Conti et al. 2009) to 6 h in human myometrium cells (Phaneuf et al. 1998; Robinson et al. 2003)); 2) OTR internalization (Berrada et al. 2000; Conti et al. 2009); 3) OTR mRNA down-regulation (Phaneuf et al. 1997, 1998); or 4) a phosphorylation-dependent mechanism (Gimpl and Fahrenholz 2001; Plested and Bernal 2001).

The only other CBF study (Paloyelis et al. 2016) is more difficult to reconcile with ours and Martins et al.'s (2020b), regarding the “dip” issue. Although those two studies used the same design, OT dose, MRI scanner/site and data modality, they report an opposite progression of the IN-OT effect; i.e., one reports a peak effect of IN-OT in the middle of the observation period (Paloyelis et al. 2016), exactly where the other (and we) reports an interruption (Martins et al. 2020b). This may be because Paloyelis et al. (2016) performed a between-subjects comparison (IN-OT vs. placebo), without accounting for the (existing) pre-administration baseline differences, and made different statistical modeling choices. The fMRI study from Spengler et al. (2017) showed the highest effect of IN-OT (24 IU) in an extended middle window (45 min—1 h 08 min), which overlaps with both our “dip” windows and most of our time window 4 where we also observed an effect. (Martins et al. 2020b) did not report any effect within that extended time window. Nevertheless, our studies are not directly reconcilable with Spengler et al. (2017)'s task-based fMRI (Spengler et al. 2017) given that their effects were exclusive to high-intensity fear stimuli (with null effects in low-intensity) and only reported on the amygdala (moreover, a region EEG may not fully access). Lastly, even if this “dip” effect of IN-OT is true, it may not affect task-based studies that record over an extended period, as they would have enough power to reach statistical significance when averaging their measurements in each condition, across the task.

Strengths and limitations

To examine IN-OT effects on the human brain, we recorded brain activity directly, with high temporal

resolution; and used EEG microstates, which allowed an inter-modal bridge with fMRI resting-state networks cognitive proxies (although their sensitivity to sub-cortical function is not yet clear). We excluded potential effects of gender and age, and thus further stratified studies are needed to increase generalizability. We used a semi-automatic pre-processing pipeline validated on resting-state EEG data (da Cruz et al. 2018). LMM statistics allowed us to take into account the pre-administration baseline in each drug session, account for subject-level random effects, and is more appropriate than ANOVA due to the non-independence of microstate data, especially in transition probabilities. We also took into account the amount of IN-OT inhaled.

Limitation-wise, we start with an important disclaimer highlighting that microstates dynamics do not represent all aspects of cortical activity, just as EEG cannot also capture all aspects of brain activity. As such, any absence of findings (in certain time windows) cannot be considered a finding of absence of an effect of IN-OT on brain function as a whole, but rather specifically on microstates dynamics. Second, the data collection was performed in hot weather conditions, which made a few participants impatient. Also, participants' sweating added noise to our signal in the form of drifts, which may have remained in our data after preprocessing to an extent, which is hard to ascertain. Third, we acknowledge that assuming a single pharmacodynamic model of IN-OT across different brain areas, with a single time course, is overly simplistic. Previous research (Paloyelis et al. 2016; Schiller et al. 2019; Martins et al. 2020b), as well as our own results, suggest a widespread, and time dependent, effect of IN-OT across the brain. Furthermore, as we restricted our study to 1 h 42 min and we observed IN-OT effects in our last time window we could not locate their end in time. Fourth, we used grand-grand average of both sessions to back-propagate the sorting to individual maps, separately for each time window—instead of averaging across all time windows, which has also been done (Michel and Koenig 2018). Our approach ensured that microstate maps were similar between the sessions at each time window, whilst the latter would have prioritized comparison between time windows—which was not our statistical goal. Spatial correlation between time windows was high, suggesting that our approach was valid. Fifth, we used four microstate classes (A, B, C, and D), which have been previously found to be optimal (Koenig et al. 2002); (Michel and Koenig 2018) and are still the literature standard, although more recent evidence (Custo et al. 2017) suggested a larger number of microstates. We kept the number of microstates at four to ensure better comparability with other microstates studies, in turn allowing us increased interpretability. Sixth, in regard to power, as our data collection took place before any IN-OT's effects on microstates were published, we could not get reliable effect sizes for an a priori power analysis. However, our sample size is similar to other within-subject and resting state In-OT studies ranging from $N = 17$ for CBF

(Martins et al. 2020b) to $N=24$ for EEG time-frequency analysis (Perry et al. 2010; de Bruijn et al. 2017) which have detected statistically significant effects. Finally, we again highlight the limitation, not regarding methodology but rather data interpretation, that although we have discussed recent literature on inferences relating EEG microstates to fMRI resting state networks—in the hopes it may help interpret our reported IN-OT effects and generate new and better hypotheses for this neuropeptide's role in cognition—this bridge between modalities should remain speculative given the still small body of evidence associating them.

Conclusions

We present the first study to our knowledge to examine IN-OT effects on the human brain across a typical neuroscience experimental session time length, using EEG. We observed effects of IN-OT on microstates dynamics from 15 min to 1 h42min post-administration. At the cognitive level, our eyes-closed data suggest IN-OT may heighten at-rest preparatory recruitment of attentional networks tuned for reception of external, and socially relevant stimuli, which, albeit being in line with social salience and approach hypotheses of OT, merits further confirmatory research, given the prior existence of a single microstates IN-OT study. By also providing a comprehensive summary of previous pharmacodynamics IN-OT studies, we hope to help inform study design and interpretability in past and future studies, regarding testing timings and duration, and provide a broader understanding of the role of OT in human cognition.

Supplementary material

Supplementary material can be found at *Cerebral Cortex* online.

Acknowledgements

The corresponding author is Diana Prata, diana.prata@kcl.ac.uk Instituto de Biofísica e Engenharia Biomédica, Faculdade de Ciências da Universidade de Lisboa, Campo Grande 016, 1749-016 Lisboa, Portugal. We thank the Direction of the CAML's CIC research centre, represented by Prof. Luís Costa, for making its facilities available for this data collection. We want to thank Andreia Santiago, Rafael Esteves and Meghan Beck for their help in data processing and manuscript revision, Katja Brodmann for help in study design and data collection, and Bastian Schiller for helpful discussion and suggestions for data processing. None of the authors declare a conflict of interest. M.Z. performed data collection, data preprocessing, microstate and statistical analyses and drafted the manuscript. M.K. contributed to data preprocessing, performed microstate and statistical analyses, revised, and co-drafted the manuscript. JdC provided scripts for, and

supervised, data preprocessing. M.A. supervised the statistical analysis. P.F. provided advice on data processing. M.M. contributed to study design and results interpretation. D.P. supervised and was responsible for the overall work, from study design to write-up. All authors revised and agreed with the final version of the manuscript.

Funding

The European Commission Seventh Framework Programme Marie Curie Career Integration to D.P. Grant FP7-PEOPLE-2013-CIG-631952, the 2016 Bial Foundation Psychophysiology Grant grant—Ref. 292/16, and the Fundação para a Ciência e Tecnologia (FCT) IF/00787/2014, LISBOA-01-0145-FEDER-030907 and DSAIPA/DS/0065/2018 grants, and the iMM Lisboa Director's Fund Breakthrough Idea Grant 2016; and is a co-founder and shareholder of the neuroimaging research services company NeuroPsyAI, Ltd MZ and M.K. were employed on FCT IF/00787/2014 and LISBOA-01-0145-FEDER-030907 grants. MA's research is partially funded by Fundação para a Ciência e Tecnologia (FCT), under the project UID/MAT/00006/2019.

References

- Abreu R, Jorge J, Leal A, Koenig T, Figueiredo P. 2021. EEG microstates predict concurrent fMRI dynamic functional connectivity states. *Brain Topogr.* 34:41–55.
- Anagnostou E, Soorya L, Brian J, Dupuis A, Mankad D, Smile S, Jacob S. 2014. Intranasal oxytocin in the treatment of autism spectrum disorders: a review of literature and early safety and efficacy data in youth. *Brain Res.* 1580:188–198.
- Andrews-Hanna JR. 2012. The brain's default network and its adaptive role in internal mentation. *Neuroscientist.* 18: 251–270.
- Barry RJ, Clarke AR, Johnstone SJ, Magee CA, Rushby JA. 2007. EEG differences between eyes-closed and eyes-open resting conditions. *Clin Neurophysiol.* 118:2765–2773.
- Bates D, Mächler M, Bolker BM, Walker SC. 2015. Fitting linear mixed-effects models using lme4. *J Stat Softw.* 67:1–48.
- Berrada K, Plesnicher CL, Luo X, Thibonnier M. 2000. Dynamic interaction of human vasopressin/oxytocin receptor subtypes with G protein-coupled receptor kinases and protein kinase C after agonist stimulation. *J Biol Chem.* 275: 27229–27237.
- Bigdely-Shamlo N, Mullen T, Kothe C, Su KM, Robbins KA. 2015. The PREP pipeline: standardized preprocessing for large-scale EEG analysis. *Front Neuroinform.* 9:1–19.
- Borland JM, Rilling JK, Frantz KJ, Albers HE. 2019. Sex-dependent regulation of social reward by oxytocin: an inverted U hypothesis. *Neuropsychopharmacology.* 44:97–110.
- Bradley ER, Woolley JD. 2017. Oxytocin effects in schizophrenia: reconciling mixed findings and moving forward. *Neurosci Biobehav Rev.* 80:36–56.
- Britz J, Van De Ville D, Michel CM. 2010. BOLD correlates of EEG topography reveal rapid resting-state network dynamics. *Neuroimage.* 52:1162–1170.
- Carson DS, Berquist SW, Trujillo TH, Garner JP, Hannah SL, Hyde SA, Sumiyoshi RD, Jackson LP, Moss JK, Strehlow MC, et al. 2015.

- Cerebrospinal fluid and plasma oxytocin concentrations are positively correlated and negatively predict anxiety in children. *Mol Psychiatry*. 20:1085–1090.
- Churchland P, Winkelman P. 2012. Modulating social behavior with oxytocin: how does it work? What does it mean? *Horm Behav*. 61: 392–399.
- Cohen D, Shamay-Tsoory SG. 2018. Oxytocin regulates social approach. *Soc Neurosci*. 13:680–687.
- Conti F, Sertic S, Reversi A, Chini B. 2009. Intracellular trafficking of the human oxytocin receptor: evidence of receptor recycling via a Rab4/Rab5 “short cycle”. *Am J Physiol Endocrinol Metab*. 296: E532–542.
- Corbetta M, Shulman GL. 2002. Control of goal-directed and stimulus-driven attention in the brain. *Nat Rev Neurosci*. 3: 201–215.
- Custo A, Van De Ville D, Wells WM, Tomescu MI, Brunet D, Michel CM. 2017. Electroencephalographic resting-state networks: source localization of microstates. *Brain Connect*. 7:671–682.
- da Cruz JR, Chicherov V, Herzog MH, Figueiredo P. 2018. An automatic pre-processing pipeline for EEG analysis (APP) based on robust statistics. *Clin Neurophysiol*. 129:1427–1437.
- da Cruz JR, Favrod O, Roinishvili M, Chkonia E, Brand A, Mohr C, Figueiredo P, Herzog MH. 2020. EEG microstates are a candidate endophenotype for schizophrenia. *Nat Commun*. 11:1–11.
- Daughters K, Manstead ASR, Hubble K, Rees A, Thapar A, Van Goozen SHM. 2015. Salivary oxytocin concentrations in males following intranasal administration of oxytocin: a double-blind, cross-over study. *PLoS One*. 10:1–11.
- De Berardis D, Marini S, Iasevoli F, Tomasetti C, de Bartolomeis A, Mazza M, Valchera A, Fornaro M, Cavuto M, Srinivasan V, et al. 2013. The role of intranasal oxytocin in the treatment of patients with schizophrenia: a systematic review. *CNS Neurol Disord Drug Targets*. 12:252–264.
- de Buijn ERA, Ruissen MI, Radke S. 2017. Electrophysiological correlates of oxytocin-induced enhancement of social performance monitoring. *Soc Cogn Affect Neurosci*. 12:1668–1677.
- De Cagna F, Fusar-Poli L, Damiani S, Rocchetti M, Giovanna G, Mori A, Politi P, Brondino N. 2019. The role of intranasal oxytocin in anxiety and depressive disorders: a systematic review of randomized controlled trials. *Clin Psychopharmacol Neurosci*. 17:1–11.
- Delorme A, Makeig S. 2004. EEGLAB: an open source toolbox for analysis of single-trial EEG dynamics. *J Neurosci Methods*. 13:9–21.
- Devarajan K, Rusak B. 2004. Oxytocin levels in the plasma and cerebrospinal fluid of male rats: effects of circadian phase, light and stress. *Neurosci Lett*. 367:144–147.
- Evans SL, Monte OD, Noble P, Averbach BB. 2014. Intranasal oxytocin effects on social cognition: a critique. *Brain Res*. 1580:69–77.
- Gamer M, Zurowski B, Büchel C. 2010. Different amygdala subregions mediate valence-related and attentional effects of oxytocin in humans. *Proc Natl Acad Sci U S A*. 107:9400–9405.
- Gimpl G, Fahrenholz F. 2001. The oxytocin receptor system: structure, function, and regulation. *Physiol Rev*. 81:629–683.
- Gossen A, Hahn A, Westphal L, Prinz S, Schultz RT, Gründer G, Spreckelmeyer KN. 2012. Oxytocin plasma concentrations after single intranasal oxytocin administration - a study in healthy men. *Neuropeptides*. 46:211–215.
- Haraya S, Karasawa K, Sano Y, Ozawa K, Kato N, Arakawa H. 2017. Development of a highly specific enzyme immunoassay for oxytocin and its use in plasma samples. *Ann Clin Biochem*. 54: 101–106.
- Hatz F, Hardmeier M, Bousleiman H, Rüegg S, Schindler C, Fuhr P. 2015. Reliability of fully automated versus visually controlled pre- and post-processing of resting-state EEG. *Clin Neurophysiol*. 126:268–274.
- Huffmeijer R, Alink LRA, Tops M, Grewen KM, Light KC, Bakermans-Kranenburg MJ, van Ijzendoorn MH. 2012. Salivary levels of oxytocin remain elevated for more than two hours after intranasal oxytocin administration. *Neuro Endocrinol Lett*. 33:21–25.
- Hyvarinen A, Oja E. 2013. Independent component analysis: algorithms and applications. *Trans ASABE*. 56:963–976.
- Khanna A, Pascual-Leone A, Michel CM, Farzan F. 2008. Microstates in resting-state EEG: current status and future directions. *North*. 29:1883–1889.
- Kirschstein T, Köhling R. 2009. What is the source of the EEG? *Clin EEG Neurosci*. 40:146–149.
- Koenig T. 2017. *The EEGLAB plugin for Microstates [WWW Document]*. Thomas Koenig's website.
- Koenig T, Prichep L, Lehmann D, Sosa PV, Braeker E, Kleinlogel H, Isenhardt R, John ER. 2002. Millisecond by millisecond, year by year: normative EEG microstates and developmental stages. *Neuroimage*. 16:41–48.
- Krylova M, Alizadeh S, Izyurov I, Teckentrup V, Chang C, van der Meer J, Erb M, Kroemer N, Koenig T, Walter M, et al. 2021. Evidence for modulation of EEG microstate sequence by vigilance level. *Neuroimage*. 224:117393.
- Landgraf R. 1985. Plasma oxytocin concentrations in man after different routes of administration of synthetic oxytocin. *Exp Clin Endocrinol*. 85:245–248.
- Lefevre A, Mottolose R, Dirheimer M, Mottolose C, Duhamel J-R, Sirigu A. 2017. A comparison of methods to measure central and peripheral oxytocin concentrations in human and non-human primates. *Sci Rep*. 7:17222.
- Lehmann D, Faber PL, Galderisi S, Herrmann WM, Kinoshita T, Koukkou M, Mucci A, Pascual-Marqui RD, Saito N, Wackermann J, et al. 2005. EEG microstate duration and syntax in acute, medication-naïve, first-episode schizophrenia: a multi-center study. *Psychiatry Res - Neuroimaging*. 138:141–156.
- Lehmann D, Ozaki H, Pal I. 1987. EEG alpha map series: brain micro-states by space-oriented adaptive segmentation. *Electroencephalogr Clin Neurophysiol*. 67:271–288.
- Leng G, Ludwig M. 2016. Intranasal oxytocin: myths and delusions. *Biol Psychiatry*. 79:243–250.
- Luo S, Zhu Y, Xu Y, Kong Q. 2017. The oxytocinergic system modulates sadistic context-dependent empathy in humans. *Sci Rep*. 7:12463.
- Marazziti D, Baroni S, Mucci F, Piccinni A, Moroni I, Giannaccini G, Carmassi C, Massimetti E, Dell'Osso L. 2019. Sex-related differences in plasma oxytocin levels in humans. *Clin Pract Epidemiol Ment Health*. 15:58–63.
- Martins D, Gabay AS, Mehta M, Paloyelis Y. 2020a. Salivary and plasma oxytocin are not reliable trait markers of the physiology of the oxytocin system in humans. *Elife*. 9:1–19.
- Martins D, Mazibuko N, Zelaya F, Vasilakopoulou S, Loveridge J, Oates A, Maltezos S, Mehta M, Wastling S, Howard M, et al. 2020b. Effects of route of administration on oxytocin-induced changes in regional cerebral blood flow in humans. *Nat Commun*. 11:1160.
- McCullough ME, Churchland PS, Mendez AJ. 2013. Problems with measuring peripheral oxytocin: can the data on oxytocin and human behavior be trusted? *Neurosci Biobehav Rev*. 37: 1485–1492.
- Mens WBJ, Witter A, Van Wimersma Greidanus TB. 1983. Penetration of neurohypophyseal hormones from plasma into cerebrospinal fluid (CSF): half-times of disappearance of these neuropeptides from CSF. *Brain Res*. 262:143–149.

- Michel CM, Koenig T. 2018. EEG microstates as a tool for studying the temporal dynamics of whole-brain neuronal networks: a review. *Neuroimage*. 180:577–593.
- Milz P, Faber PL, Lehmann D, Koenig T, Kochi K, Pascual-Marqui RD. 2016. The functional significance of EEG microstates-associations with modalities of thinking. *Neuroimage*. 125: 643–656.
- Mishra A, Englitz B, Cohen MX. 2020. EEG microstates as a continuous phenomenon. *Neuroimage*. 208:116454.
- Mitchell MD, Haynes PJ, Anderson ABM, Turnbull AC. 1981. Plasma oxytocin concentrations during the menstrual cycle. *Eur J Obstet Gynecol Reprod Biol*. 12:195–200.
- Mu Y, Guo C, Han S. 2016. Oxytocin enhances inter-brain synchrony during social coordination in male adults. *Soc Cogn Affect Neurosci*. 11:1882–1893.
- Murray MM, Brunet D, Michel CM. 2008. Topographic ERP analyses: a step-by-step tutorial review. *Brain Topogr*. 20:249–264.
- Musso F, Brinkmeyer J, Mobascher A, Warbrick T, Winterer G. 2010. Spontaneous brain activity and EEG microstates. A novel EEG/fMRI analysis approach to explore resting-state networks. *Neuroimage*. 52:1149–1161.
- Mustoe A, Schulte NA, Taylor JH, French JA, Toews ML. 2019. Leu8 and Pro8 oxytocin agonism differs across human, macaque, and marmoset vasopressin 1a receptors. *Sci Rep*. 9:1–10.
- Neto ML, Antunes M, Lopes M, Ferreira D, Rilling J, Prata D. 2020. Oxytocin and vasopressin modulation of prisoner's dilemma strategies. *J Psychopharmacol*. 34:891–900.
- Neumann ID, Slattery DA. 2016. Oxytocin in general anxiety and social fear: a translational approach. *Biol Psychiatry*. 79:213–221.
- Paloyelis Y, Doyle OM, Zelaya FO, Maltezos S, Williams SC, Fotopoulou A, Howard MA. 2016. A spatiotemporal profile of in vivo cerebral blood flow changes following intranasal oxytocin in humans. *Biol Psychiatry*. 79:693–705.
- Pascual-Marqui RD, Lehmann D, Faber P, Milz P, Kochi K, Yoshimura M, Nishida K, Isotani T, Kinoshita T. 2014. *The resting microstate networks (RMN): cortical distributions, dynamics, and frequency specific information flow*. arXiv:1411.1949v2.
- Perlow MJ, Reppert SM, Artman HA, Fisher DA, Seif SM. 1982. Oxytocin, vasopressin, and Estrogen-stimulated Neurophysin: daily patterns of concentration in cerebrospinal fluid. *Science*. 216: 1416–1418.
- Perry A, Bentin S, Shalev I, Israel S, Uzefovsky F, Bar-On D, Ebstein RP. 2010. Intranasal oxytocin modulates EEG mu/alpha and beta rhythms during perception of biological motion. *Psychoneuroendocrinology*. 35:1446–1453.
- Phaneuf S, Asbóth G, Carrasco MP, Europe-Finner GN, Saji F, Kimura T, Harris A, López BA. 1997. The desensitization of oxytocin receptors in human myometrial cells is accompanied by down-regulation of oxytocin receptor messenger RNA. *J Endocrinol*. 154: 7–18.
- Phaneuf S, Asbóth G, Carrasco MP, Liñares BR, Kimura T, Harris A, Bernal AL. 1998. Desensitization of oxytocin receptors in human myometrium. *Hum Reprod Update*. 4:625–633.
- Pipinis E, Melynyte S, Koenig T, Jarutyte L, Linkenkaer-Hansen K, Rukšenas O, Griskova-Bulanova I. 2017. Association between resting-state microstates and ratings on the Amsterdam resting-state questionnaire. *Brain Topogr*. 30:245–248.
- Plested CP, Bernal AL. 2001. Desensitisation of the oxytocin receptor and other G-protein coupled receptors in the human myometrium. *Exp Physiol*. 86(2):303–312.
- Quintana DS, Westlye LT, Rustan OG, Tesli N, Poppy CL, Smevik H, Tesli M, Røine M, Mahmoud RA, Smerud KT, et al. 2015. Low-dose oxytocin delivered intranasally with breath powered device affects social-cognitive behavior: a randomized fourway crossover trial with nasal cavity dimension assessment. *Transl Psychiatry*. 5:e602.
- Quintana DS, Westlye LT, Smerud KT, Mahmoud RA, Andreassen OA, Djupesland PG. 2018. Saliva oxytocin measures do not reflect peripheral plasma concentrations after intranasal oxytocin administration in men. *Horm Behav*. 102: 85–92.
- Reppert SM, Perlow MJ, Artman HG, Ungerleider LG, Fisher DA, Klein DC. 1984. The circadian rhythm of oxytocin in primate cerebrospinal fluid: effects of destruction of the suprachiasmatic nuclei. *Brain Res*. 307:384–387.
- Robinson C, Schumann R, Zhang P, Young RC. 2003. Oxytocin-induced desensitization of the oxytocin receptor. *Am J Obstet Gynecol*. 188:497–502.
- Rutherford HJV, Guo XM, Wu J, Graber KM, Hayes NJ, Pelphrey KA, Mayes LC. 2018. Intranasal oxytocin decreases cross-frequency coupling of neural oscillations at rest. 123:143–151.
- Salonia A, Nappi RE, Pontillo M, D'Averio R, Smeraldi A, Briganti A, Fabbri F, Zanni G, Rigatti P, Montorsi F, et al. 2005. Plasma levels of oxytocin during the menstrual cycle, pregnancy and following treatment with HMG. *Hum Reprod*. 47: 1056–1062.
- Schiller B, Koenig T, Heinrichs M. 2019. Oxytocin modulates the temporal dynamics of resting EEG networks. *Sci Rep*. 9:13418.
- Seitzman BA, Abell M, Bartley SC, Erickson MA, Bolbecker AR, Hetrick WP. 2017. Cognitive manipulation of brain electric microstates. *Neuroimage*. 146:533–543.
- Shamay-Tsoory SG, Abu-Akel A. 2016. The social salience hypothesis of oxytocin. *Biol Psychiatry*. 79:194–202.
- Shilling PD, Feifel D. 2016. Potential of oxytocin in the treatment of schizophrenia. *CNS Drugs*. 30:193–208.
- Singh F, Nunag J, Muldoon G, Cadenhead KS, Pineda JA, Feifel D. 2016. Effects of intranasal oxytocin on neural processing within a socially relevant neural circuit. *Eur Neuropsychopharmacol*. 26: 626–630.
- Spengler FB, Schultz J, Scheele D, Essel M, Maier W, Heinrichs M, Hurlmann R. 2017. Kinetics and dose dependency of intranasal oxytocin effects on amygdala reactivity. *Biol Psychiatry*. 82: 885–894.
- Stock S, Bremme K, Uvnäs-moberg K. 1991. Plasma levels of oxytocin during the menstrual cycle, pregnancy and following treatment with HMG. *Hum Reprod*. 6:1056–1062.
- Striepens N, Kendrick KM, Hanking V, Landgraf R, Wüllner U, Maier W, Hurlmann R. 2013. Elevated cerebrospinal fluid and blood concentrations of oxytocin following its intranasal administration in humans. *Sci Rep*. 3:3440.
- Tagliazucchi E, Laufs H. 2014. Decoding wakefulness levels from typical fMRI resting-state data reveals reliable drifts between wakefulness and sleep. *Neuron*. 82:695–708.
- Taylor KS, Seminowicz DA, Davis KD. 2009. Two systems of resting state connectivity between the insula and cingulate cortex. *Hum Brain Mapp*. 30:2731–2745.
- Torres N, Martins D, Santos AJ, Prata D, Veríssimo M. 2018. How do hypothalamic nonapeptides shape youth's sociality? A systematic review on oxytocin, vasopressin and human socio-emotional development. *Neurosci Biobehav Rev*. 90: 309–331.
- Valstad M, Alvares GA, Egknud M, Matziorinis AM, Andreassen OA, Westlye LT, Quintana DS. 2017. The correlation between central and peripheral oxytocin concentrations: a systematic review and meta-analysis. *Neurosci Biobehav Rev*. 78: 117–124.

- van Ijzendoorn MH, Bhandari R, van der Veen R, Grewen KM, Bakermans-Kranenburg MJ. 2012. Elevated salivary levels of oxytocin persist more than 7 h after intranasal administration. *Front Neurosci.* 6:1–6.
- Weisman O, Zagoory-Sharon O, Feldman R. 2012. Intranasal oxytocin administration is reflected in human saliva. *Psychoneuroendocrinology.* 37:1582–1586.
- Wynn JK, Green MF, Helleman G, Reavis EA, Marder SR. 2019. A dose-finding study of oxytocin using neurophysiological measures of social processing. *Neuropsychopharmacology.* 44:289–294.
- Xu X, Yuan H, Lei X. 2016. Activation and connectivity within the default mode network contribute independently to future-oriented thought. *Sci Rep.* 6:1–10.
- Yuan H, Zotev V, Phillips R, Drevets WC, Bodurka J. 2012. Spatiotemporal dynamics of the brain at rest - exploring EEG microstates as electrophysiological signatures of BOLD resting state networks. *Neuroimage.* 60:2062–2072.
- Zhong S, Monakhov M, Mok HP, Tong T, Lai PS, Chew SH, Ebstein RP. 2012. U-shaped relation between plasma oxytocin levels and behavior in the trust game. *PLoS One.* 7:e51095.