



King's Research Portal

DOI:

[10.1016/j.jinf.2017.04.009](https://doi.org/10.1016/j.jinf.2017.04.009)

Document Version

Peer reviewed version

[Link to publication record in King's Research Portal](#)

Citation for published version (APA):

Jeljeli, M., Guérin-El Khourouj, V., Pommelet, V., Hormi, M., Gressens, P., Faye, A., & Sterkers, G. (2017). Cytokine/chemokine secretion for detecting tuberculosis in quantiferon supernatants from HIV(+) and HIV(-) children. *The Journal of infection*, 75(1), 77-80. <https://doi.org/10.1016/j.jinf.2017.04.009>

Citing this paper

Please note that where the full-text provided on King's Research Portal is the Author Accepted Manuscript or Post-Print version this may differ from the final Published version. If citing, it is advised that you check and use the publisher's definitive version for pagination, volume/issue, and date of publication details. And where the final published version is provided on the Research Portal, if citing you are again advised to check the publisher's website for any subsequent corrections.

General rights

Copyright and moral rights for the publications made accessible in the Research Portal are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognize and abide by the legal requirements associated with these rights.

- Users may download and print one copy of any publication from the Research Portal for the purpose of private study or research.
- You may not further distribute the material or use it for any profit-making activity or commercial gain
- You may freely distribute the URL identifying the publication in the Research Portal

Take down policy

If you believe that this document breaches copyright please contact librarypure@kcl.ac.uk providing details, and we will remove access to the work immediately and investigate your claim.

Infection

Elsevier Editorial System(tm) for Journal of

Manuscript Draft

Manuscript Number:

Title: Cytokine/chemokine secretion for detecting tuberculosis in
quantiferon supernatants from HIV+ and HIV- children

Article Type: Letter to the Editor

Section/Category: Rest of the World Submissions

Corresponding Author: Pr. Ghislaine Sterkers, MD-PHD

Corresponding Author's Institution: Robert Debré Hospital

First Author: Mohamed Jeljeli

Order of Authors: Mohamed Jeljeli; Valérie Guérin-El Khourouj; Virginie
Pommelet; Myriam Hormi; Pierre Gressens; Albert Faye; Ghislaine Sterkers,
MD-PHD

Manuscript Region of Origin: FRANCE

Title: Cytokine/chemokine secretion for detecting tuberculosis in quantiferon supernatants from HIV⁺ and HIV⁻ children

Running title: Biomarkers of tuberculosis in HIV-infected children

Mohamed Jeljeli ^a, Valérie Guérin-El Khourouj ^a, Virginie Pommelet ^b, Myriam Hormi ^a, Pierre Gressens ^c, Albert Faye ^{b, d} and Ghislaine Sterkers ^{a,*}

Institutional/professional affiliations:

^a Laboratory of Immunology, Robert-Debré Hospital, Assistance Publique-Hôpitaux de Paris (AP-HP), Univ. Paris Diderot, Sorbonne Paris Cité, 75019 Paris, France.

^b Department of Pediatric Infectious Disease, Robert-Debré Hospital, Assistance Publique-Hôpitaux de Paris (AP-HP), Univ. Paris Diderot, Sorbonne Paris Cité, 75019 Paris, France.

^c INSERM U1141, Robert-Debré Hospital, Assistance Publique-Hôpitaux de Paris (AP-HP), Univ. Paris Diderot, Sorbonne Paris Cité, 75019 Paris, France.

^d INSERM 1123, ECEVE, Univ Paris Diderot, Sorbonne Paris Cité, France

***Corresponding author**

Pr. Ghislaine STERKERS

Robert Debré hospital – Laboratory of Immunology

48, Boulevard Sérurier, 75019 Paris, France

Tel: +33.1.40.03.53.05 - Fax: +33.1.40.03.47.76

E-mail: ghislaine.sterkers@aphp.fr

Keywords: HIV/Tuberculosis, co-infection, children, quantiFERON, biomarkers

Conflicts of interest: none

Financial support: Financial support was provided by Assistance Publique-Hôpitaux de Paris-France.

Sir,

The QuantiFERON-Gold In Tube assay (QFT) relies on the detection of IFN-gamma secreted by immune T-cells after whole blood stimulation with ESAT-6, CFP-10 and TB7-7 mycobacterium tuberculosis (M.tb) antigens (Ag). Although QFT has proved more specific than the Tuberculin-Skin test (TST) for detecting tuberculosis (TB) in countries where TB incidence is low [1], its use in pediatric tuberculosis (TB) is limited by the lower sensitivity of the test in immunocompetent children younger than 5 years old and in HIV-infected children [2].

In this journal, we recently reported the potential added value of combining IFN-gamma, IP-10 and IL-13 releases for diagnosing latent TB in 0-5 years old immunocompetent children [3]. Here, we further explored the potential added value of these biomarkers in HIV-infected children. The study was conducted in Ile-de-France (France), a low TB burden area where the sensitivity of QFT to diagnose active TB is impaired in HIV-infected children (Hormi et al. *Ped Infect Dis J* 2017 *in press*).

Among 63 consecutive HIV⁺ children who were referred to our center (hospital Robert-Debré Paris-France) for QFT reactivity analysis between January 2011 and November 2015, residual plasma from QFT were available for 12 HIV⁺/TB⁺ co-infected children (stage III of HIV-infection, n=11/12). At QFT sampling, four HIV⁺/TB⁺ children had untreated active TB (culture-confirmed, n=3/4). Four had untreated latent TB. Four have had previous active TB. Two other groups of children not-infected with HIV (HIV⁻) that were evaluated for QFT reactivity during the same period as HIV⁺/TB⁺ children were included as reference; i) Twelve HIV⁻/TB⁺ children were used for comparison with the HIV⁺/TB⁺ co-infected group. Inclusion of HIV⁻/TB⁺ children was based, as far as possible, on a HIV⁺/TB⁺, HIV⁻/TB⁺ case-control design. ii) Eleven age-matched children who were presumed not-infected (HIV⁻/TB⁻) based on one-year follow-up QFT and/or TST negativity and normal chest x-ray in the

absence of anti-TB treatment (11/11) following a recent contact with a not-infectious (not-bacillary) index case (10/11) were included to identify biomarkers that discriminate TB⁺ from TB⁻ children. Informed consent was obtained from the parents and the study was approved by the Robert Debré hospital local ethic committee.

According to selection design, HIV⁺/TB⁺ and HIV⁻/TB⁺ clinical groups displayed similar demographic and clinical main characteristics (median age: 10y6m and 8y11m; M/F gender: 7/5 and 6/6; birth and/or a long stay in a country with a high TB burden 7/12 and 6/12; clear documentation of BCG-vaccination 10/10 and 9/12 in HIV⁺/TB⁺ and HIV⁻/TB⁺ children respectively). Also, TB status was similar in the two groups (ongoing active TB before anti-TB treatment, n=4/12 and 3/12, untreated latent TB or past active TB n=8/12 and 9/12 children in HIV⁺/TB⁺ and HIV⁻/TB⁻ respectively).

Lower M.tb-specific QFT value was observed in HIV⁺/TB⁺ children (median: 0.63 IU/ml) than among HIV⁻/TB⁺ children, median: 9.42 IU/ml, p=0.007). Overall, the sensitivity of QFT was 91% among HIV⁻/TB⁺ children (one negative result: 0.03 IU/ml) but 58% among HIV⁺/TB⁺ children (five negative results ranging from 0.00 to 0.01 IU/ml).

The accuracy of 18 cytokines/chemokines for diagnosing TB were evaluated using the human 17-Plex and IP-10 kits (Bio-Rad, Laboratories Inc., Marnes-La-Coquette, France) as previously described [4]. Cytokine/chemokine concentrations were measured in residual plasmas (cryopreserved at -80°C until use) from QFT (QuantiFERON GIT, Cellestis). Plasmas from the 3 QFT tubes (not-stimulated, M.tb Ag-stimulated and PHA-stimulated) were analyzed for each child.

Five biomarkers showing either concentrations lying outside an interpretable range (IL-8, MCP-1, MIP-1beta) or similar levels in all culture conditions (IL-7, IL-12) were excluded from analysis.

As shown in table 1, stimulation with M.tb Ag was associated with increased expression levels of IL-2, IL-5, IL-13, IFN-gamma and IP-10 in plasmas from HIV⁻/TB⁺ but not in plasmas from HIV⁻/TB⁻ children. ROC analysis of M.tb-specific cytokine/chemokine releases (value in the M.tb Ag-stimulated tube minus value in the corresponding not-stimulated tube) showed that IL-2, IL-5, IL-13 and IP-10, in addition to IFN-gamma could discriminate HIV⁻/TB⁺ from HIV⁻/TB⁻ children (table 2).

In the entire panel of cytokines/chemokines we examined, only IP-10 expression displayed a higher median level in the M.tb Ag-stimulated tube than in the not-stimulated tube among HIV⁺/TB⁺ children (table 1). ROC analysis confirmed IP-10 as a potential biomarker to discriminate HIV⁺/TB⁺ children from HIV⁻/TB⁻ children (table 2). The IP-10 threshold-value that best discriminated TB⁺ from TB⁻ children was however less elevated among HIV⁺/TB⁺ (912.9pg/ml) than among HIV⁻/TB⁺ children (3644pg/ml). Using the 912.9pg/ml value as the cut-off of positivity 3/5 HIV⁺/TB⁺ children with QFT negative results displayed a positive IP-10 release (active TB, n=1; past active TB, n=2) and the sensitivity of IP-10 (83%) was somewhat better than IFN- γ release in the multiplex assay (50%) or QFT (58%).

This study is the first that used excess QFT supernatants to compare M.tb-specific cytokine/chemokine releases in HIV⁺/TB⁺ and HIV⁻/TB⁻ children. Our results showing defective IL-2, IL-5 and IL-13 responses to M.tb in co-infected children ($p \leq 0.002$) make a contribution to our knowledge in the immunopathology of pediatric HIV/TB co-infection. IP-10 response to M.tb appeared less affected by HIV and showed promise in improving QFT sensitivity. A recent meta-analysis concluded that the IP-10 test performs slightly better or comparable to QFT in HIV⁺/TB⁺ adults [5]. To our knowledge, only two previous pediatric studies regarding QFT and IP-10 performance included HIV⁺ children. These two studies, both conducted in low/middle income countries and high TB burden area, concluded that both QFT and IP-10 release exhibited an equally poor performance in diagnosing TB [6, 7]. We

have no clear explanations for the discrepancies between these results and our findings. The impact of the TB burden in the different places where the studies were conducted should be investigated.

In conclusion, although combining IL-13 and IP-10 may improve QFT sensitivity for diagnosing TB in immunocompetent young children [3], only IP-10 showed promise for improving TB diagnosis in HIV⁺ children. Larger pediatric cohorts are warranted to investigate whether measuring IP-10 in combination with IFN- γ may improve QFT sensitivity without compromising specificity in HIV⁺/TB⁺ children and invite to identify HIV⁺ pediatric subgroups that could best benefit from IP-10 release implementation.

References

1. Laurenti P, Raponi M, de Waure C, Marino M, Ricciardi W, Damiani G. Performance of interferon-gamma release assays in the diagnosis of confirmed active tuberculosis in immunocompetent children: a new systematic review and meta-analysis. *BMC Infect Dis* 2016,**16**:131.
2. Starke JR. Interferon-gamma release assays for diagnosis of tuberculosis infection and disease in children. *Pediatrics* 2014,**134**:e1763-1773.
3. Jeljeli M, Guerin-El Khourouj V, Hormi M, Sterkers G, Pommelet V, Faye A, *et al.* Immune response to Mycobacterium tuberculosis in young contacts with discordant immunological test results. *J Infect* 2016.
4. Armand M, Chhor V, de Lauzanne A, Guerin-El Khourouj V, Pedron B, Jeljeli M, *et al.* Cytokine responses to quantiferon peptides in pediatric tuberculosis: a pilot study. *J Infect* 2014,**68**:62-70.
5. Ruhwald M, Aabye MG, Ravn P. IP-10 release assays in the diagnosis of tuberculosis infection: current status and future directions. *Expert Rev Mol Diagn* 2012,**12**:175-187.
6. Holm LL, Rose MV, Kimaro G, Bygbjerg IC, Mfinanga SG, Ravn P, *et al.* A comparison of interferon-gamma and IP-10 for the diagnosis of tuberculosis. *Pediatrics* 2014,**134**:e1568-1575.
7. Chegou NN, Detjen AK, Thiart L, Walters E, Mandalakas AM, Hesselning AC, *et al.* Utility of host markers detected in Quantiferon supernatants for the diagnosis of tuberculosis in children in a high-burden setting. *PLoS One* 2013,**8**:e64226.

Table 1

Table 1: Biomarker concentrations (pg/ml) in QFT plasmas from the not-stimulated and the M.tb-Ag stimulated tubes

Cytokine	HIV ⁻ TB ⁻ children (n=11)			HIV ⁻ TB ⁺ children (n=12)			HIV ⁺ TB ⁺ children (n=12)		
	Not-stimulated	M.tb-Ag stimulated	P	Not-stimulated	M.tb-Ag stimulated	P	Not-stimulated	M.tb-Ag stimulated	P
IL-1β	2603 (146 - 4297)	4434 (1196 - 6015)	0.2145	2532 (952 - 12172)	6301 (1453 - 13064)	0.0304	1908 (247 - 12583)	3469 (384 - 14078)	0.1572
IL-2	120 (42 - 306)	123 (53 - 566)	0.9320	104 (42 - 260)	2025 (110 - 9414)	0.0001	94 (39 - 233)	161 (65 - 7276)	0.0351
IL-4	123 (54 - 185)	115 (67 - 236)	0.6350	103 (56 - 258)	168 (72 - 263)	0.0531	109 (29 - 213)	130 (46 - 222)	0.5067
IL-5	17 (10 - 178)	21 (1 - 334)	0.6860	13 (8 - 23)	70 (13 - 175)	0.0002	14 (12 - 15)	14 (11 - 115)	0.2024
IL-6	6328 (318 - 10641)	4920 (168 - 14831)	0.9770	5476 (441 - 19763)	8063 (3513 - 23279)	0.2145	5181 (109 - 15827)	5782 (315 - 18514)	0.5834
IL-10	73 (17 - 179)	85 (28 - 195)	0.7950	55 (21 - 387)	75 (25 - 152)	0.8173	58 (46 - 222)	63 (49 - 160)	0.5440
IL-13	46 (23 - 188)	47 (23 - 408)	0.8173	26 (14 - 59)	524 (25 - 2005)	0.0002	26 (16 - 58)	41 (15 - 1865)	0.1332
IL-17	433 (172 - 642)	348 (197 - 736)	0.5444	427 (139 - 767)	541 (211 - 959)	0.1939	506 (73 - 728)	524 (208 - 783)	0.5834
IFN-γ	86 (31 - 138)	80 (41 - 185)	0.7508	70 (29 - 213)	893 (205 - 4038)	<0.0001	66 (31 - 180)	99 (38 - 1159)	0.1410
TNF-α	465 (84 - 887)	552 (74 - 991)	0.6350	383 (160 - 1711)	1009 (208 - 17154)	0.0266	235 (29 - 1812)	240 (54 - 1763)	0.9770
IP-10	972 (174 - 13286)	1144 (277 - 13910)	0.8852	970 (348 - 1686)	18149 (1812 - 20299)	<0.0001	772 (198 - 3264)	3962 (654 - 24756)	0.0036
G-CSF	129 (40 - 298)	100 (39 - 155)	0.1939	106 (25 - 1046)	147 (27 - 376)	0.4357	126 (25 - 785)	159 (29 - 710)	0.9370
GM-CSF	274 (178 - 679)	293 (138 - 394)	0.8173	203 (93 - 381)	324 (110 - 385)	0.0120	262 (87 - 339)	289(93 - 471)	0.7073

M.tb-Ag: *Mycobacterium-antigen*s contained in QFT (ESAT-6, CFP-10 and TB7.7)

Results are given as median, (range) concentrations (pg/ml).

p-values below 0.0038 thresholds (Mann-Whitney test comparing not-stimulated with M.tb-Ag stimulated values after Bonferroni adjustment for 13 tests) are highlighted in bold.

Table 2: Accuracies of IL-2, IL-5, IL-13, IFN- γ and IP-10 releases in the diagnosis of TB in HIV⁻/TB⁺ (A) and HIV⁺/TB⁺ (B) children

(A)

HIV ⁻ TB ⁺ children (n=12)								
Biomarker	AUC	p	Cut-off value(pg/l)	Sensitivity (%)	95% CI	Specificity (%)	95%CI	Incidence Positivity n (%)
IL-2	0.962	0.0001	175.7	91.67	61.52 to 99.79	90.91	71.51 to 100	11 (91.67)
IL-5	0.886	0.0010	9.6	91.67	61.52 to 99.79	90.91	58.72 to 99.77	11 (91.67)
IL-13	0.947	0.0002	60.8	91.67	61.52 to 99.79	90.91	58.72 to 99.77	11 (91.67)
IFN- γ	1.000	<0.0001	111.4	100	73.54 to 100	100	71.51 to 100	12 (100)
IP-10	0.977	0.0001	3644.0	91.67	61.52 to 99.79	90.91	58.72 to 99.77	11 (91.67)

(B)

HIV ⁺ TB ⁺ children (n=12)								
Biomarker	AUC	p	Cut-off value (pg/l)	Sensitivity (%)	95% CI	Specificity (%)	95%CI	Incidence Positivity n (%)
IL-2	0.651	0.218	115.5	41.67	15.17 to 72.33	90.91	58.72 to 99.77	5 (41.66)
IL-5	0.594	0.441	7.4	58.33	27.67 to 84.83	54.55	23.38 to 83.25	1 (8.33)
IL-13	0.678	0.148	32.5	33.33	9.25 to 65.11	81.82	48.22 to 97.72	4 (33.3)
IFN- γ	0.704	0.096	47.85	50	21.09 to 78.91	81.82	48.22 to 97.72	6 (50)
IP-10	0.825	0.008	912.9	83.33	51.59 to 97.91	81.82	48.22 to 97.72	10 (83.33)

AUC: Area under curve. 95% CI: 95% Confidence Interval.

Receiver operating characteristics (ROC) curves were generated by plotting biomarker release values observed in HIV⁻/TB⁻ children (specificity) against the corresponding values in HIV⁻/TB⁺ (A) or HIV⁺/TB⁺ (B) children (sensitivity). Only those analytes with significant ROC curves for HIV⁻/TB⁺ or HIV⁺/TB⁺ group are shown.

Specific release (value in the *M.tb* Ag tube minus value in the corresponding not-stimulated tube) was used in ROC curve analyses in agreement with the current literature.

Cut-off values were determined on the basis of the highest likelihood index (Youden's Index).

Sensitivity and specificity are expressed as a percentage.

Groups with AUC \geq 0.70 and p<0.05 after ROC analysis are highlighted in bold.