

## Various marginal marine environments in the Central Paratethys: Late Badenian and Sarmatian (middle Miocene) marine and non-marine microfossils from Pécs-Danitzpuszta, southern Hungary

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### *Késő badeni és szarmata (középső miocén) mikrofoszszíliák Pécs-Danitzpusztáról*

#### Összefoglalás

A Középső-Paratethys középső miocén foraminifera és kagylósrák együttese általában stabil normál tengeri viszonyokat tükröznek a badeniben, míg térben és időben változatosabb, mozaikos környezeteket a szarmatában. A pécs-danitzpusztai homokbányában kiásott kutatóárokban 17 méter vastagságban tárult fel a tektonikusan kibillentett középső miocén rétegsor, amely jelentős környezeti változásokról tanúskodik a késő badeni és a pannóniai között. A rétegsor alján normál tengeri, sekélyvízi, meleg, jól szellőzött, aránylag nagy energiájú, mikrobaszőnyeges aljzatú környezetre utalnak a mészkő-, márgarétegek mikrofoszszíliái. A foraminiferák alapján ezek a rétegek a késő badeniben (13,82 és 12,65 millió év között) rakódtak le. A szelvény középső szakaszán a tengeri rétegeket mikrofoszszília-mentes, gravitációsán áthalmozott durva homok-breccsa, aleurit váltakozásából álló sorozat követi, amelyben valószínűleg szárazföldi kitértiségre utaló gyökérbekéregzéseket is találtunk. A következő tengerelöntés éles közettani váltás mellett a szelvény felső részéből kinyert mikrofoszszíliák alapján a késő szarmatában történt, kb. 12 és 11,6 millió év között. Ezek a rétegek a felső szarmata *Porosonion granosum* zónát (foraminiferák) és *Aurila notata* zónát (kagylósrákok) képviselik. Az együttesek kizárólag tágtűrűsű fajokból állnak, és változó só-, oxigén- és tápanyagtartalmú, növényzettel rendelkező, brakkvízi tengeri környezetet jeleznek. Az 5 m vastag felső szarmata egységben néhány rétegből, amelyek együttesen egy métert képviselnek, édesvízi vagy legfeljebb oligohalin kagylósrákfauna és kivételesen tág sótűrűsű foraminiferák kerültek elő. Az együttesek megváltozását nem kíséri jelentős litológiai váltás, nincs jele megnövekedett szárazföldi eredetű behordásnak, amely egy közeli folyótorkolatot jelezne. Sem a késő szarmata geomorfológiai viszonyok, sem a kőzetminőség nem utal a tengertől részben elzárt környezet (lagúna, parti mocsár) kialakulására. A helyi viszonyokon túlmutató tényezők (pl. az éghajlat változása) nagyobb területen is megfigyelhető lenne, ilyen adatokkal azonban nem rendelkezünk. A tágtűrűsű foraminiferák és édesvízi-oligohalin kagylósrákok megjelenése mindenesetre helyben vagy a közelben élő közösségre utal, és így a helyi alacsony (5–10 ‰) sótartalmat jelzi. Ilyen közösséget más szarmata szelvényből a Középső-Paratethys területén eddig nem ismertünk. A szelvény tetején a foraminiferák hirtelen eltűnése és ezzel egy időben a kagylósrákfauna teljes kicserélődése a szarmata és pannóniai emeletek határát jelöli ki (11,6 millió év).

*Kulcsszavak: Középső-Paratethys, Mecsek, foraminifera, kagylósrák, taxonómia, biosztratigráfia, paleoökológia*

#### Abstract

The middle Miocene foraminifera and ostracod record of the Central Paratethys usually reflects stable normal marine depositional environments for the Badenian and more patchy, less stable restricted marine environments for the Sarmatian. A 17 m thick outcrop at Pécs-Danitzpuszta, Mecsek Mts, SW Hungary exposed an upper Badenian to Pannonian succession where foraminifers and ostracods document significant environmental changes. The basal layers of the section contain micro- and macrofossils indicating normal marine, shallow, warm, well-oxygenated habitat with relatively high-energy conditions and algal vegetation on the bottom, and represent the upper Badenian (13.82 to 12.65 Ma). The marine deposits are followed by coarse sandstone, breccia and siltstone layers barren of microfossils but containing rhizoliths. The sediments were probably subaerially exposed for some time. The following marine inundation, marked by the appearance of clays and limestones as well as fossils, was dated to the late Sarmatian (ca. 12 to 11.6 Ma) on the basis of the restricted marine microfossil assemblages from the upper part of the succession (*Porosonion granosum* Zone, *Aurila notata* Zone). This community is characterized by exclusively eurytopic forms indicating an unstable and vegetated marginal marine environment with fluctuations in salinity, as well as oxygen and food availability. Within the 5 m thick upper Sarmatian marine interval, a unique fresh- to oligohaline fauna characterizes a few layers in less than 1 m thickness. This fauna consists of highly euryhaline foraminifera and freshwater to oligohaline ostracod assemblages, indicating a

temporary salinity reduction to 5–10‰. No similar freshwater fauna has been reported from the Sarmatian of the Central Paratethys so far. The eventual disappearance of the foraminifera from the paleontological record coupled with a complete turnover in the ostracod fauna indicates the transition from the marginal marine Sarmatian Sea to the brackish Lake Pannon, marking the Sarmatian/Pannonian boundary (11.6 Ma).

**Keywords:** Central Paratethys, Mecsek Mts, Foraminifera, Ostracoda, taxonomy, biostratigraphy, paleoecology

## Introduction

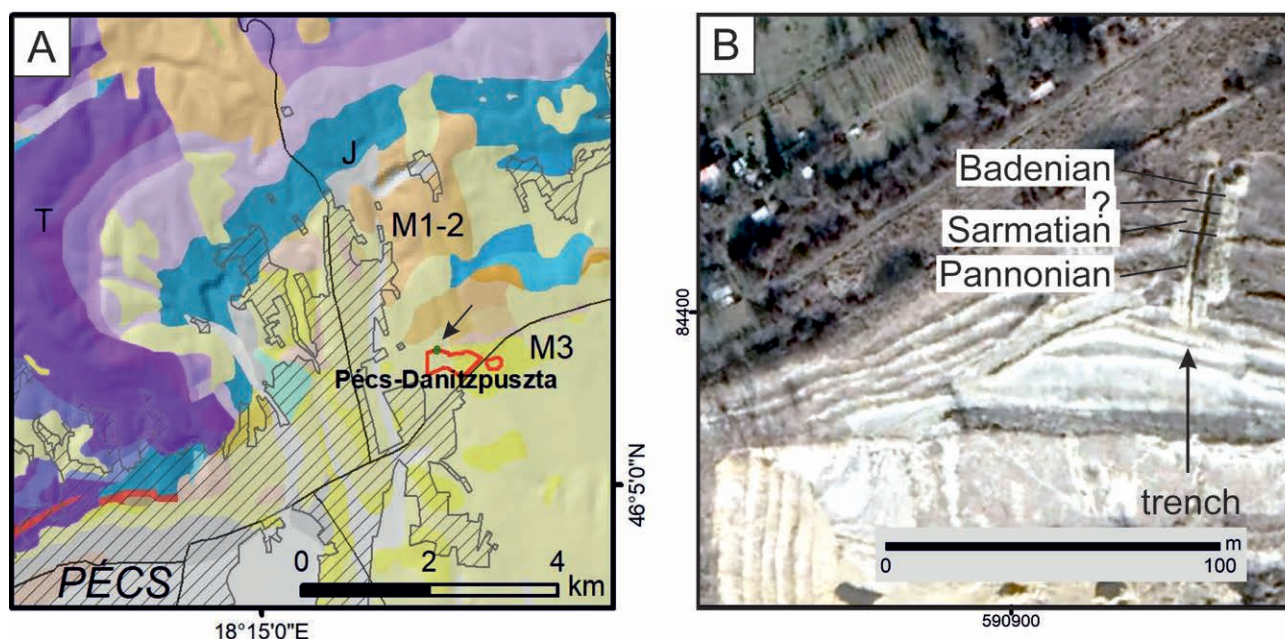
The distribution of marine microorganisms in an epicontinental sea is driven by the local and regional changes of environmental conditions such as salinity, water temperature, oxygen-level, food availability, substrates, and water depth. These environmental conditions and the evolution of the microfauna were controlled by the openings and closures of the seaways towards the adjacent seas and the world ocean in the Paratethys, an epicontinental sea of central and eastern Europe during the Oligocene and Miocene (RÖGL 1998, POPOV et al. 2004). The connection toward the Mediterranean Sea was terminated due to the uplift of the Dinarides at the Badenian/Sarmatian boundary, triggering an endemic evolution of the marine faunas in the Paratethys (e.g., PALCU et al. 2015). The seaway towards the Indopacific was closed in the late Sarmatian, eliminating the last Indo-Pacific planktonic elements that were detected in the Transylvanian Basin (FILIPESCU & SILYE 2008). All of these changes might have influenced the biota at the study area in SW Hungary.

The present study focuses on the taxonomy and paleoecological and biostratigraphical interpretation of foraminifer and ostracod communities from a middle Miocene succession exposed in an exploratory trench in the Pécs-Danitzpuszta sand pit, Mecsek Mts, SW Hungary. Earlier studies

of middle Miocene foraminifera in Hungary (BÁLDI 1999, 2006; BÁLDI et al. 2002; BÁLDI 2006; GÖRÖG 1992; KORECZ-LAKY 1964, 1965, 1968, 1973, 1982; KORECZ-LAKY & NAGY-GELLAI 1985; TÓTH & GÖRÖG 2008) showed the wide distribution of the normal marine Badenian and restricted marine (brackish and hypersaline) Sarmatian faunas, which are well-known in the entire Central Paratethys. The study of Sarmatian ostracods resulted in a biostratigraphic system for the entire Pannonian Basin (TÓTH 2004, 2008), whereas Badenian ostracods from Hungary have not been studied yet. By investigating the Pécs-Danitzpuszta micropaleontological record, we give the first documentation of Badenian ostracods from Hungary and also describe a so far unknown upper Sarmatian non-marine ostracod assemblage.

## Geological setting

The Pécs-Danitzpuszta sand pit lies in the eastern outskirts of Pécs, at the foot of the Mecsek Mts (Figure 1). The region north of the sand pit is built up of Mesozoic rocks, mostly Lower Jurassic marls and sandstones, overlain by a succession of lower to middle Miocene terrestrial clastics and middle Miocene marine clastics and carbonates (SEBE et al. 2015, 2019; SEBE et al. 2021). These are capped by upper



**Figure 1.** Location of the Pécs-Danitzpuszta sand pit (A) and the exploratory trench (B)

*I. ábra.* A pécs-danitzpusztai homokbánya (A) és a kutatóárok (B) elhelyezkedése

Miocene (Pannonian) marls and sands, exposed in many outcrops around the Mecsek. The boundary between Sarmatian and Pannonian deposits is continuous in (sub)basin centres, while they are separated by an unconformity with increasing hiatus towards the margins. Similar, but several km thick Neogene successions were reported from the Drava Basin to the south and southwest (SAFTIĆ et al. 2003; SEBE et al. 2020) reflecting the opening and evolution of the Pannonian Basin, flooding by the Paratethys sea and later by the brackish Lake Pannon.

## Material and methods

### *Studied section of Pécs-Danitzpuszta sand pit*

The sand pit exposes strongly tilted upper Miocene (Pannonian) calcareous marls and sands. In 2018, an exploratory trench was excavated in the northwestern part of the sand pit across the tilted beds that underlie the exposed Pannonian marl (Figure 1). The trench revealed the lowermost part of the Pannonian succession and the underlying Sarmatian and Badenian deposits. Due to tectonic deformation, most of the exposed succession is overturned, and the stratigraphically lowest (oldest) layers are located in the north (SEBE 2021). Overturned beds become steeper towards the south (upsection) and they are almost vertical close to the southern end of the trench. The oldest part of the studied section is represented by yellowish white calcareous marl (Layer D72) in the northern end of the trench (Figures 2, 3). It contains a typical Badenian mollusk fauna and belongs to the Lajta Formation (SEBE et al. 2021, DULAI et al. 2021). D71 also shows features typical of the Lajta Limestones: it is a sandy limestone with coralline algae, echinoids, abundant molluscs, and sporadic fish remains (DULAI et al. 2021, SEBE et al. 2021, SZABÓ et al. 2021). The following beds (D70 to D57) did not provide stratigraphically valuable fossils; thus, their age is uncertain (Figure 2). These are unconformably overlain by a ca. 5 m thick unit of alternating thin clay, marl and limestone beds (layers D56–D36), identified as the Sarmatian Kozárd Formation based on its fossil content and lithology (SEBE et al. 2021).

### *Micropaleontological samples and methods*

Fifteen middle Miocene samples from the trench were studied for their foraminiferal and ostracod content (Figures 2, 3). The samples derived from soft sediments (about 200 g of air-dried clayey, sandy and marly sediments) were processed with hydrogen-peroxide (10%). Hard limestones and calcareous marls were examined in thin sections, or the samples were treated by acetolysis following a protocol originally worked out by LETHIERS & CRASQUIN-SOLEAU (1988) to extract the isolated carbonate skeletal microfauna. The applied extraction methods and the frequency of the extracted fossil groups from the studied layers are summarized in Figure 4.

Thirteen samples yielded interpretable microfossil content; D57 and D69 were free of microfossils (Appendix). The microfossils were determined using a Zeiss SteREO Discovery.V12 modular binocular stereo microscope in the Laboratory of MOL Plc., Budapest. Thin sections were prepared in the Laboratory of MOL Plc., Budapest and they were investigated with a Zeiss Axio Imager.A1 polarizing microscope. Microscopic images were taken by a Zeiss AxioCam MRc 5 camera, mounted on the Zeiss microscope, using the AxioVision 40x64 v.4.9.1.0 software. The SEM images were taken at the Botanical Department of the Hungarian Natural History Museum in Budapest.

## Results

Relatively diverse and well-preserved benthic foraminiferal and ostracod assemblages were found in the studied middle Miocene beds. Altogether, 30 foraminifer and 32 ostracod taxa were identified (see Appendix and Digital annex). The foraminifera specimens are moderately to well-preserved, except for layers D70 and D71, where they were probably affected by transport of the tests and/or diagenetic processes. The ostracod specimens are disarticulated valves in most cases; however, a few carapaces also occur. The ostracod material is characterized by both adult and juvenile forms.

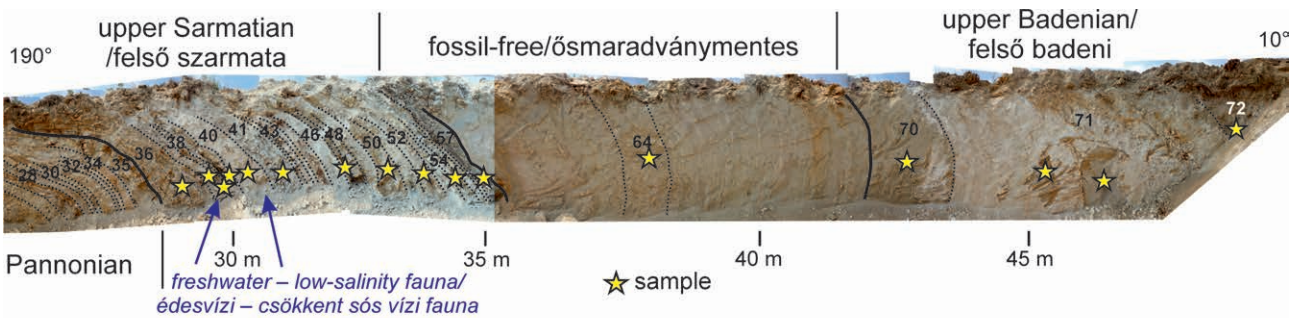
The oldest layer (D72) yielded the most diverse and abundant microfossil assemblage. Twenty-one foraminifera and 11 ostracod taxa were identified (Figure 2, Plate I). The foraminiferal assemblage was dominated by eurytopic taxa of keeled elphidiids (*Elphidium aculeatum*, *E. crispum*, and *E. macellum*) and miliolids (*Borelis* sp., *Cycloforina contorta*, *Affinetrina ucrainica*, *Miliolinella selene*, and *Quinqueloculina hauerina*). The ostracod fauna is characterized by the dominance of marine neritic taxa, such as *Aurila cicatricosa*, *Callistocythere canaliculata*, and *Phlyctenophora arcuata*. *Urocythereis kostelensis*, *Loxocochia punctatella*, *Loxocorniculina hastata*, *Xestoleberis dispar*, and *Polycope* sp. also occur in low abundance. Besides foraminifers and ostracods, sample D72 also yielded significant amounts of echinoderm skeletal and spike fragments.

The microfossil assemblages of layers D70 and D71 were similar to, but significantly poorer than, that of D72. Poor preservation of the carbonate skeletons allowed only genus level determination in most cases (*Xestoleberis* sp., *Callistocythere* sp., *Polycope* sp., and *Elphidium* sp.). Echinoderm fragments were also more sporadic than in sample D72. The microfossils of layer D70 are probably reworked based on the scarcity and poor preservation of the specimens, although a diagenetic effect cannot be excluded either.

The soft sediments of layers D54 to D41 yielded a less diverse (5–10 taxa), well-preserved foraminifer and ostracoda fauna (Figure 2, Plates II–III). Among the foraminifera, exclusively eurytopic forms (taxa with wide environmental tolerance) were present. Keeled elphidiids with an acute periphery, sometimes equipped with spines, were the most common (e.g., *Elphidium aculeatum*, *E. macellum*, *E. obtu-*







**Figure 3.** The northern part of the exploratory trench exposes overturned middle Miocene (D72 to D36) and stratigraphically overlying Pannonian (D35 to D28) layers. Sampling locations are indicated by yellow stars

**3. ábra.** A kutatóárok északi része, mely az átkutatott középső miocén (D72 – D26) és a pannóniai rétegeket (D35 – D28) tárja fel. A sárga csillag jelzi a mintavételi helyeket

*sum*, and *E. crispum*). Among the non-keeled elphidiids, where the periphery of the tests is rounded or bluntly angular, specimens of *Porosonion granosum* were abundant. The ostracod fauna was characterized by different species of the genera *Aurila*, *Loxoconcha* and *Euxinocythere* (e.g., *Aurila notata*, *Loxoconcha kochi*, *L. porosa*, and *Euxinocythere [Euxinocythere] praebosqueti*). Specimens of *Xestoleberis tumida* are also present in the samples.

In layers D40 to D37, mainly specimens of the infaunal, non-keeled elphidiid *P. granosum* and *Ammonia* sp. were found (Figure 2). Beside the sporadic occurrence of marginal marine ostracods (e.g., *Loxoconcha porosa* and *Aurila* sp.), non-marine, freshwater to oligohaline ostracods, like *Fabaeformiscandona* sp., *Heterocypris salina*, *Darwinula stevensoni*, and *Vestalenula pagliolii* are present in the recovered assemblages.

Layer D36 is characterized by the dominance of eurytopic non-keeled elphidiids and nonionids and the representatives of leptocytherid *Euxinocythere* (*E. [E.] praebosqueti* and *E. [E.] naca*) (Plate II).

## Discussion

### Biostratigraphy

Benthic foraminifera are instrumental in the biostratigraphy of the middle Miocene sediments of the Central Paratethys, because the best index fossils, such as planktonic foraminifers and nannoplankton, are commonly missing from the fossil record, especially in the coastal regions (Figure 5).

← **Figure 2.** Sedimentary log of the middle Miocene part of the Pécs-Danitzpuszta succession, with sample locations, micropaleontological intervals and subintervals based on the stratigraphic distribution and ecological needs of the studied microfossil assemblages and the distribution of the paleoecologically important foraminifer and ostracod taxa and morphogroups in the samples  
Abbreviation: Pa= Pannonian

← **2. ábra.** A pécs-danitzpusztai homokbányában kiásított kutatóárok középső miocén szakaszának szelvénye a vizsgált minták feltüntetésével, a mikrofauna biostratigráfiai és paleoökológiai értékelése alapján elkülönített intervallumokkal, valamint a környezetjelzés szempontjából fontos foraminifera és ostracoda taxonok, illetve morfofocsoportok megoszlásával  
Rövidítés: Pa= pannóniai

A commonly used threefold subdivision of the Badenian in the Pannonian, Vienna and Danube basins is partly based on the composition of benthic foraminifers reflecting distinct paleoenvironmental changes (PAPP et al. 1978). The lower Badenian is represented by the “Lagenidae Zone,” the middle Badenian by the “*Spiroplectamina* Zone,” and the upper Badenian by the “*Bulimina/Bolivina* Zone” (GRILL 1943, PAPP et al. 1978). Sarmatian sediments of the Pannonian, Vienna and Danube basins can be divided to four benthic foraminiferal zones: *Anomalinoidea dividens*, *Elphidium reginum*, and *Elphidium hauerinum* Zones in the lower Sarmatian, and *Porosonion granosum* Zone in the upper Sarmatian (GRILL 1943, JIŘIČEK 1972, PAPP & SENEŠ 1974). For the Sarmatian of the Pannonian Basin, TÓTH (2009) proposed a two-fold ostracod zonation: *Cytheridea hungarica-Aurila mehesi* Zone for the lower Sarmatian and *Aurila notata* Zone for the upper Sarmatian.

Layer D72 belongs to the upper Badenian based on the co-eval occurrence of *Pyrgo subsphaerica* (upper Badenian to recent) and *Miliolinella selene* (Badenian) among the foraminifera (ŁUCZKOWSKA 1974). Some ostracods in these layers, such as *Urocythereis kostelensis* and *Phlyctenophora affinis*, are restricted to the Badenian (GROSS & PILLER 2006). Although the microfauna is dominated by eurytopic forms, normal marine taxa (e.g., *Callistocythere canaliculata* and *Heterolepa dutemplei*) also occur in these samples; they disappeared from the Central Paratethys at the end of the Badenian. Thus, the microfossil assemblages of layers D72 to D70 indicate late Badenian age, equivalent of the “*Bulimina/Bolivina* Zone” (13.82 to 12.65 Ma, according to HOHENEGGER et al. 2014 and RAFFI et al. 2020), which correlates with the standard nannoplankton Zone NN6 (RÖGL et al. 2008).

The presence of *Aurila notata* in layers D54 to D36 suggests correlation with the *Aurila notata* Zone (ca. 12 to 11.6 Ma). Several other taxa, such as *Euxinocythere* (*E.*) *praebosqueti*, *E. (E.) naca*, *Loxoconcha kochi* are also restricted to the upper Sarmatian in the Pannonian Basin (TÓTH 2009). The foraminiferal assemblages are characterized by a great abundance of *Porosonion granosum* in almost all samples, indicating the *Porosonion granosum* Zone. This cor-

Layers	72	71	70	64	57	54	52	50	47	41	40	39	38	37	36	
Methods	TS, AA, HP	HP	HP	HP	HP	HP	HP	HP	HP	HP	AA, TS	AA, TS	TS, HP	TS, AA	HP	
Microfossils	Root traces	—	—	+	+	+	—	—	—	—	—	—	—	—	—	
	Organic matter	—	—	—	—	+	—	—	—	—	—	—	—	—	—	
	Calci-sphaera	—	—	—	—	—	+	+	—	—	—	+	—	—	+	
	Red algae & Bryozoa (Bry)	+	—	—	—	—	+ <sup>Bry</sup>	+ <sup>Bry</sup>	—	—	—	—	—	—	—	
	Pteropoda & Echinodermata (Ech)	+	+ <sup>Ech</sup>	+ <sup>Ech</sup>	—	—	—	—	—	—	—	—	—	—	—	
	Fish remains	—	+	—	—	+	+	+	+	+	+	—	—	+	—	+
	Mollusca (G=Gastropoda)	+	+	+ <sup>G</sup>	—	—	+	+	+	—	+ <sup>G</sup>	+	+	—	—	+
	Ostracoda	+	+	+	—	—	+	+	+	—	+	+	+	+	+	+
Foraminifera	+	+	+	—	—	+	+	+	+	+	+	+	+	+	+	
Age	late Badenian		reworked	late Badenian? or early Sarmatian?		late Sarmatian										
Facies	Normal marine, high-energy conditions			Very shallow-water, paludal conditions	Brackish-water, littoral, well-ventilated conditions						Littoral conditions with freshwater influence			Brackish-water		
Interval	1a	1b		2	3a						3b			3c		

## Legend

## Test methods

TS: thin section  
 AA: acetic acid preparation  
 HP: hydrogen-peroxide preparation

## Frequency

+ few      + some  
 + several    — none

Figure 4. The extraction method of the studied layers and the frequency of the extracted fossil groups from the studied samples

4. ábra. Az egyes rétegek mintáinak mikropaleontológiai feltárási módszere, és a kinyert ősmaradványcsoportok gyakorisága a mintákban

relates with the younger part of the *Aurila notata* Zone (TÓTH 2009). This biostratigraphic interpretation is in accordance with the nannoplankton zonation of the same layers (NN6 or younger, according to ČORIĆ, 2021).

A sudden change in the microfossil assemblages can be observed between layers D36 and D35, indicating the Sarmatian/Pannonian boundary (11.6 Ma). Foraminifera are entirely missing from sample D35, and the ostracod faunas of the two samples are completely different, without any species in common. In D36, juvenile *Aurila notata* and *Cyprideis* sp. specimens, *Loxocorniculum hastatum*, *Euxinocythere* (*Euxinocythere*) *praeboqueti*, *E. (E.) naca*, and *Amnicythere tenuis* occur. In contrast, sample D35 is dominated by *Candona* and *Herpetocyrella* species. Calci-sphaera-like large algal cysts and mysid statoliths (ballast stones of the shrimplike mysids; following the interpretation of VOICU 1981) are present in sample D35 in low abundance. These are characteristic fossils in strata near the S/P boundary at several locations in Hungary where it was not possible to precisely assign the boundary

itself (e.g., KÓVÁRY 1974, BARDÓCZ et al. 1987). Mysids are very common in unanimously Sarmatian layers of the Transylvanian and Dacian Basins (e.g., POPESCU 1995).

Despite the sharp microfaunal change, no major shift can be observed in the lithofacies of the sediment. The mollusc assemblage of sample D35 contains abundant "Sarmatian-type" small-sized cardiids (BOTKA et al. 2021). This fauna, affected by the Lilliput Effect (HARRIES & KNORR 2009), is often related to environmental stress and has been published from the Sarmatian/Pannonian transition by several authors from different parts of the Pannonian Basin (e.g., Zsámbék Basin, Hungary, BOHN-HAVAS 1983; Lajoskomárom–1 well, Hungary, JÁMBOR et al. 1985; Medvednica Mts, Croatia, VRSALJKO 1999). Although the ostracod faunas of layers D36 and D35 are very different, and mollusks are missing from D36 while D35 shows the mass occurrence of tiny cardiid bivalves, it is not obvious if a short gap or continuous sedimentation occurred at the Sarmatian/Pannonian boundary.

Age (Ma)	Epoch/ Age	Polarity	Chron	Nanno Zones Martini 1971	Central Paratethys stages	Benthic Foram Zones Grill 1943, Papp et al. 1978	Defining events
10	Tortonian 11.63	[Black bar]	C5	NN9	Pannonian ~11.6		SPEE
11				NN8			
12				NN7			
12	Serravallian 13.82	[Black bar]	C5A	NN6	Sarmatian ~12.65	<i>P. granosum</i> <i>E. hauennum</i> <i>E. reginum</i> Zone	BSEE
13				C5AA	upper Badenian ~13.3	<i>Bulimimal</i> <i>Bolivina</i> Zone agg. f. zone	BSC
14				C5AB	~13.82	upper	
14	Langhian	[Black bar]	C5AC	NN5	~14.38		
15				C5AD	lower Badenian	lower	Lagenidae Zone
15			C5B	NN4			

**Figure 5.** Middle to upper Miocene geochronology, geomagnetic polarity chrons, biozonations of calcareous nannoplankton and benthic foraminifers correlated to regional chronostratigraphy of the Central Paratethys with defining events (modified after HOHENEGGER et al. 2014 and RAFFI et al. 2020).

Abbreviations: BSC= Badenian Salinity Crisis, SPEE = Sarmatian-Pannonian Extinction Event, BSEE=Badenian-Sarmatian Extinction Event (after HARZHAUSER & PILLER 2007)

**5. ábra.** Középső miocén rétegtani ábra radiometrikus koradatokkal, magnetosztratigráfiai és biosztratigráfiai (mészvázú nannoplankton, bentosz foraminifera) beosztással, illetve a Középső-Paratethysben lejátszódott meghatározó események feltüntetésével (HOHENEGGER et al. 2014 és RAFFI et al. 2020 után módosítva).

Rövidítések: BSC= Badeni Sókrízis, SPEE = Szarmata/pannóniai kihalási esemény, BSEE= Badeni/szarmata kihalási esemény (HARZHAUSER & PILLER 2007 után)

### Paleoecology

#### Ecological requirements of the extant relatives of the studied middle Miocene taxa

Extant representatives of keeled elphidiids live in temperate to warm, shallow marine (at water depths up to 50 m) environments (inner shelf) and hypersaline lagoons (MURRAY 1991, 2006). They are mostly epiphytic dwellers (live on plants) and prefer sandy sediment (LANGER 1993, MURRAY 2006). In the Mediterranean Sea, *E. aculeatum* and *E. macellum* live on arborescent algal vegetation (LANGER et al. 1998). They are chromatophore-bearing foraminifera and the “symbionts” may control the phototaxis and the depth distribution of the host organism. The chromatophores are pigment-containing cells that produce color. However, the nature of this symbiosis and the role of the chromatophores in phototaxis – the ability of organisms to move directionally in response to a light source – are poorly known. *E. macellum* is a common member of foraminiferal assemblages in the Black Sea living in the shallow sublittoral zone and coastal pools (down to 20 m depth) (TEMELKOV 2008). *Miliolinella* and *Quinqueloculina* are epiphytic or they cling on hard substrates in the inner shelf or in normal marine to hypersaline lagoons and marshes; they rarely can be found in deep-sea records (MURRAY 2006). Recent miliolids prefer waters rich in calcium carbonate (JORISSEN 1988). *Borelis* is a large, benthic foraminifera with photosynthetic diatom algal symbionts. The recent species are restricted to depths of

5–65 m in, for example, the Gulf of Aqaba, and to minimum sea-surface temperatures greater than 18 °C (REISS & HOTTINGER 1984, LANGER & HOTTINGER 2000). Non-keeled infaunal elphidiids are characteristic species of brackish to hypersaline marshes and lagoons; however, they can also be found in the inner shelf (water depth up to 50 m) (MURRAY 2006). *Ammonia* is widespread in marginal marine environments worldwide and is common in sediments with highly variable mud and organic matter contents, even at low oxygen levels in marsh environments (MURRAY 2006).

Among the ostracods, *Aurila* and *Urocythereis* recently live in great abundance in the infralittoral and uppermost circalittoral zone (water depth up to 40 m) of the Black Sea, the Mediterranean, the Eastern Atlantic, and the Indo-Pacific area (e.g., ATHERSUCH 1977, RUIZ et al. 1997, KILIÇ

2000, AIELLO et al. 2006, TANAKA 2008). Modern representatives of *Aurila*, *Xestoleberis*, and *Loxococoncha* species mainly live on algae or seagrasses (PURI et al. 1969). *Loxococoncha punctatella* and *Xestoleberis dispar* are found in neritic shallow sublittoral, littoral environments in the Mediterranean, Black and Marmara Seas (PERÇIN-PAÇAL et al. 2015). In the present-day Mediterranean Sea, *Xestoleberis dispar* is a phytal marine species, but it also occurs in hypersaline environments (SCIUTO et al. 2015, KOEHN-ZANINETTI & TÉTARD 1982). *Phlyctenophora* occurs in marginal marine estuarine, gulf and lagoonal environments in the Indo-Pacific Realm (WOUTERS 1999, HUSSAIN et al. 2004, MISHRA et al. 2019). Recent polycopids have a nektobenthic lifestyle and are found from abyssal ocean depths (KARANOVIC & BRANDÃO 2012, 2016) to less saline estuarine environments (TANAKA & TSUKAGOSHI 2010).

Recent *Euxinocythere*, similarly to *Aurila* and *Loxococoncha*, occur in shallow marine sublittoral and littoral environments in the Black Sea (PERÇIN-PAÇAL et al. 2015). The extant species *Heterocypris salina* and *Darwinula stevensoni* are cosmopolitan and are known from all continents. *Heterocypris salina* lives in saline coastal and inland water bodies coexisting with other halophilic ostracods and tolerate salinities up to 20‰ (MEISCH 2000). The modern species of *Darwinula* are mostly found in freshwater, although *D. stevensoni* also tolerates stable, brackish conditions in coastal waters (e.g., Baltic Sea) or saline lakes (NEALE 1988, VAN



DONINCK et al. 2003), and is reported to tolerate salinities as high as 15‰ (DE DECKKER 1981). Today, *Vestalenula pagliolii* occurs in Brazil, where it thrives in riverine pools and lakes, semiterrestrial and/or interstitial habitats and occurs in geographically restricted areas (MARTENS et al. 1997).

### Paleoenvironments

Three main intervals were differentiated in the studied layers of Pécs-Danitzpuszta trench based on the stratigraphic distribution and ecological needs of the identified foraminifera and ostracod taxa, within which further subintervals were designated (Figure 2). The paleoecological interpretations are based on the ecology of extant relatives of the studied taxa.

Interval 1 (sample D72) represents the upper Badenian, and it is characterized with the most diverse fossil assemblage within the sedimentary record (Figure 2). The dominance of keeled elphidiids and miliolids among the foraminifera and marine neritic genera (*Aurila*, *Callistocythere*, *Loxoconcha*, *Urocythereis*, *Phlyctenophora*, and *Xestoleberis*) among the ostracods suggests shallow marine, calcium-carbonate rich littoral environment (inner shelf) with water depths up to 50 m. Although several of the identified forms can live today in hypersaline lagoons as well, the high diversity of the microfauna excludes such environmental interpretation. The presence of the large benthic foraminifera *Borelis* in the assemblage indicates warm seawater, with temperatures higher than 18 °C (REISS & HOTTINGER 1984; LANGER & HOTTINGER 2000). Based on the great abundance of epiphytic dweller foraminifera taxa such as *E. aculeatum* and *E. macellum* and phytal ostracods (*Aurila*, *Loxoconcha* and *Xestoleberis*), a rich arborescent algal vegetation is supposed to have been present on the sea bottom. The keeled elphidiids are chromatophore-bearing foraminifers that must have lived in the euphotic zone with well-ventilated conditions. The abundance of thick-shelled ostracods, often with worn valves, and the abundance of echinoderm fragments indicate high energy conditions in the sea bottom. The red algal and bryozoan fragments also support this environmental interpretation.

Interval 2 (samples D69 and D57) yielded only one fish tooth. Carbonate-cemented cylinders around holes were interpreted as rhizoliths (root traces; Figure 2). The Fe-Mn encrusted unconformity on top of bed D57 and the appearance of fossiliferous clays, marls and limestones with upper Sarmatian marine microfossils above the unconformity denote a sharp change in the depositional environment, probably from terrestrial to marine.

Interval 3 (layers D54 to D36) belongs to the upper Sarmatian, suggesting that the area was re-flooded by the sea only during the late Sarmatian.

Subinterval 3a (samples D54 to D41) is characterized by exclusively eurytopic forms and lower diversity than in Interval 1 (Figure 2). The impoverishment of the marine faunal elements is explained by the Badenian-Sarmatian Extinction event (BSEE) caused by the final isolation of

the Central Paratethys from the Mediterranean and coeval reconnection with the Eastern Paratethys (HARZHAUSER & PILLER 2007). Among the elphidiids, non-keeled forms (mainly the specimens of *Porosonion granosum*) appeared in great abundance due to the unstable environment, e.g., slight fluctuation in salinity or other factors such as food availability. The non-keeled infaunal elphidiids tolerate brackish to hypersaline conditions suggesting marginal marine depositional environments such as a lagoon or a hypersaline marsh. The disappearance of *Phlyctenophora* and *Urocythereis* and the dominance of *Euxinocythere* corroborate the marginal marine conditions. The abundance of the keeled elphidiids and phytal ostracods (*Aurila*, *Loxoconcha* and *Xestoleberis*) implies a rich vegetation on the substrate. The co-occurrence of shallow infaunal non-keeled and epiphytic keeled elphidiids suggests mixed assemblages indicating a very differentiated seafloor.

In Subinterval 3b (samples D40 to D37) the faunal composition radically changed (Figure 2). The abundance and diversity of foraminifera and ostracoda decreased. Beside the non-keeled infaunal *Porosonion*, the specimens of *Ammonia* cf. *confertitesta* became dominant. *Ammonia* cf. *confertitesta* tolerates a wide range of salinity (10–50‰) and also occurs in non-marine foraminifera faunas (MURRAY 2006). The ostracod fauna is characterized by non-marine, freshwater to oligohaline ostracods, such as *Darwinula stevensoni*, *Heterocypris salina*, *Vestalenula pagliolii*, *Cyprideis* cf. *torosa*, *Fabaeformiscandona* sp., and *Limnocythere* sp. This microfossil assemblage indicates a sudden decrease in salinity (which is also supported by the mollusk fauna represented by *Radix*, *Gyraulus* and *Theodoxus* occurring without the brackish *Congerina* and cardids). The interpretation of this phenomenon, however, remains a hard nut to crack. The lithology does not show any sign of increased terrestrial input that the proximity of a river mouth would cause, and the Sarmatian geomorphological position of the outcrop, reconstructed as a tip of a promontory protruding into a wide basin, does not support the idea of a freshened lagoon or coastal marsh either. A more regional cause of the salinity drop, such as a climate change, would have left its mark on the fossil record of a wider region, but we are not aware of such observations. Thus, what we can conclude is only that the euryhaline foraminifera and freshwater-oligohaline ostracods lived together in a brackish water (5–10‰ salinity) habitat.

In Subinterval 3c (sample D36), characteristic Sarmatian eurytopic taxa (non-keeled elphidiids, nonionids, and representatives of the leptocytherid *Euxinocythere*) replace the non-marine, freshwater-oligohaline species. The low diversity microfossil assemblage with the dominance of infaunal foraminifera (non-keeled elphidiids, nonionids, and bolivinids) and thin-shelled ostracods indicates low-oxygenated environment and/or higher organic content. The latter is supported by the nannoflora, suggesting increasing nutrient supply in this period (ĆORIĆ 2021).



## Conclusions

The microfossil record of the middle Miocene sedimentary succession of Pécs-Danitzpuszta indicates significant environmental changes through the late Badenian–early Pannonian. The lowermost part of the section belongs to the upper Badenian, with typical Badenian faunal elements indicating stable, normal marine, shallow (inner shelf), warm, well-ventilated environment with relatively high-energy conditions and algal vegetation on the bottom. The overlying layers are devoid of marine microfossils and may indicate terrestrial deposition and subaerial exposure. Following an unconformity, the upper part of the middle Miocene succession belongs to the upper Sarmatian with two distinct biofacies. The lower part and the uppermost layer of the upper Sarmatian are characterized by exclusively eurytopic forms, indicating an unstable and vegetated marginal marine environment with fluctuations in salinity, as well as oxygen and food availability. The middle part of the upper

Sarmatian, however, contains highly euryhaline forams and a unique freshwater to oligohaline ostracod fauna, indicating low salinity. Finally, the disappearance of foraminifera taxa and a complete turnover in the ostracod fauna indicates the boundary between the marginal marine Sarmatian and the brackish lacustrine Pannonian stages (11.6 Ma).

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## Plate I – I. tábla

Badenian microfossils from the studied exploratory trench in Pécs-Danitzpuszta: 1: *Callistocythere canaliculata* (REUSS), RV in lateral view, layer D72, scale bar: 250 µm; 2-3: *Aurila cicatricosa* (REUSS), 2: LV in lateral view, 3: C in right view, layer D72, scale bar: 250 µm; 4: *Urocythereis kostelensis* (REUSS), LV in lateral view, layer D72, scale bar: 250 µm; 5: *Loxococoncha punctatella* (REUSS), LV in lateral view layer D72, scale bar: 200 µm; 6: *Senesia cinctella* (REUSS), C in right view, layer D72, scale bar: 250 µm; 7: *Xestoleberis tumida* (REUSS), RV in lateral view, layer D72, scale bar: 250 µm; 8: *Xestoleberis dispar* MUELLER, C in left view, layer D72, scale bar: 250 µm; 9: *Borelis* sp., SV, layer D72, scale bar: 200 µm; 10: *Heterolepa dutemplei* (D'ORBIGNY), UV, layer D72, scale bar: 500 µm; 11: *Cycloforina contorta* (D'ORBIGNY), SV, layer D72, scale bar: 200 µm; 12: *Affinetrina ucrainica* (SEROVA), SV, layer D72, scale bar: 500 µm; 13: *Nonion commune* (D'ORBIGNY), SV, layer D72, scale bar: 500 µm; 14: *Elphidium crispum* (LINNÉ), SV, scale bar: 500 µm; 15: *Textularia* sp., layer D72; 16: *Pyrgo subsphaerica* (D'ORBIGNY), layer D72; 17: *Asterigerinata planorbis* (D'ORBIGNY), layer D72; 18: *Heterolepa dutemplei* (D'ORBIGNY), layer D72; 19: echinoid spine, layer D72; 20: sponge spicule, layer D72; 21: red algae fragment, layer D72; 22: serpulid worm burrow, layer D72

Abbreviations: LV= left valve, RV= right valve, C= carapace, SV= side view, UV= umbilical view

Badeni mikrofosztiliák a pécs-danitzpusztai homokbányában ásott kutatóárokából: 1: *Callistocythere canaliculata* (REUSS), RV oldalnézetben, D72 réteg, méretarány: 250 µm; 2-3: *Aurila cicatricosa* (REUSS), 2: LV oldalnézetben, 3: C jobb oldali nézetben, D72 réteg, méretarány: 250 µm; 4: *Urocythereis kostelensis* (REUSS), LV oldalnézetben, D72 réteg, méretarány: 250 µm; 5: *Loxococoncha punctatella* (REUSS), LV oldalnézetben, D72 réteg, méretarány: 200 µm; 6: *Senesia cinctella* (REUSS), C jobb oldali nézetben, D72 réteg, méretarány: 250 µm; 7: *Xestoleberis tumida* (REUSS), RV oldalnézetben, D72 réteg, méretarány: 250 µm; 8: *Xestoleberis dispar* MUELLER, C in bal oldali nézetben, D72 réteg, méretarány: 250 µm; 9: *Borelis* sp., SV, D72 réteg, méretarány: 200 µm; 10: *Heterolepa dutemplei* (D'ORBIGNY), UV, D72 réteg, méretarány: 500 µm; 11: *Cycloforina contorta* (D'ORBIGNY), SV, D72 réteg, méretarány: 200 µm; 12: *Affinetrina ucrainica* (SEROVA), SV, D72 réteg, méretarány: 500 µm; 13: *Nonion commune* (D'ORBIGNY), SV, D72 réteg, méretarány: 500 µm; 14: *Elphidium crispum* (LINNÉ), SV, méretarány: 500 µm; 15: *Textularia* sp., D72 réteg; 16: *Pyrgo subsphaerica* (D'ORBIGNY), D72 réteg; 17: *Asterigerinata planorbis* (D'ORBIGNY), D72 réteg; 18: *Heterolepa dutemplei* (D'ORBIGNY), D72 réteg; 19: tengerisüntüske, D72 réteg; 20: szivacsstű, D72 réteg; 21: vörösalga-töredék, D72 réteg; 22: féregjárát, D72 réteg

Rövidítések: LV= bal teknő, RV= jobb teknő, C= kettősteknő, SV= oldalnézet, UV= köldökoldali nézet

## Plate II – II. tábla

Sarmatian ostracods from the studied exploratory trench in Pécs-Danitzpuszta: 1: *Amnicocythere tenuis* (REUSS), RV in lateral view, layer D50, scale bar: 200 µm; 2-3: *Amnicocythere cernajseki* STANCHEVA, 2: LV in lateral view, 3: RV in lateral view, layer D38, scale bar: 200 µm; 4-6: *Euxinocythere (Euxinocythere) praebosqueti* (SUZIN), 4: ecophenotype, LV in lateral view, layer D36, scale bar: 250 µm; 5: RV in lateral view, layer D36, scale bar: 200 µm; 6: LV in lateral view, layer D50, scale bar: 200 µm; 7: *Euxinocythere (Euxinocythere) naca* (MÉHES), RV in lateral view, layer D50, scale bar: 200 µm; 8-9: *Cyprideis pokorny* JIŘÍČEK, 8: male, RV in lateral view, 9: female, RV in lateral view, layer D38, scale bar: 250 µm; 10: *Cyprideis* sp., layer D38, scale bar: 250 µm; 11: *Hemicocytheria omphalodes* (REUSS), juvenile, RV in lateral view, layer D36, scale bar: 200 µm; 12-13: *Aurila notata* (REUSS), 12: RV in lateral view, 13: LV in lateral view, layer D50, scale bar: 500 µm; 14-15: *Loxococoncha kochi* MÉHES, 14: LV in lateral view, 15: RV in lateral view, layer D50, scale bar: 250 µm; 16: *Loxococoncha laeta* STANCHEVA, LV in lateral view, layer D54, scale bar: 200 µm; 17: *Loxococoncha porosa* MÉHES, RV in lateral view, layer D54, scale bar: 200 µm; 18: *Loxocauda* sp., layer D38, scale bar: 250 µm; 19-20: *Loxocorniculum hastatum* (REUSS), 19: LV in lateral view, 20: RV in lateral view, layer D36, scale bar: 250 µm; 21: *Darwinula stevensoni* (BRADY & ROBERTSON), C in right view, layer D 40, scale bar: 250 µm; 22-23: *Vestalenula pagliolii* (PINTO & KOTZIAN), 22: RV in lateral view, 23: C in left view, layer D38, scale bar: 200 µm; 24: *Fabaeformiscandona* ? sp. juv., RV in lateral view, layer D38, scale bar: 200 µm; 25: *Limnocythere* sp., LV in lateral view, layer D38, scale bar: 200 µm; 26: *Heterocypris salina* (BRADY), C in left view, layer D40, scale bar: 500 µm

Abbreviations: LV= left valve, RV= right valve, C= carapace

Szarmata kagylósrakok a pécs-danitzpusztai homokbányában ásott kutatóárokából: 1: *Amnicocythere tenuis* (REUSS), RV oldalnézetben, D50 réteg, méretarány: 200 µm; 2-3: *Amnicocythere cernajseki* STANCHEVA, 2: LV oldalnézetben, 3: RV oldalnézetben, D38 réteg, méretarány: 200 µm; 4-6: *Euxinocythere (Euxinocythere) praebosqueti* (SUZIN), 4: ökojenotípus, LV oldalnézetben, D36 réteg, méretarány: 250 µm; 5: RV oldalnézetben, D36 réteg, méretarány: 200 µm; 6: LV oldalnézetben, D50 réteg, méretarány: 200 µm; 7: *Euxinocythere (Euxinocythere) naca* (MÉHES), RV oldalnézetben, D50 réteg, méretarány: 200 µm; 8-9: *Cyprideis pokorny* JIŘÍČEK, 8: hím, RV oldalnézetben, 9: nőstény, RV oldalnézetben, D38 réteg, méretarány: 250 µm; 10: *Cyprideis* sp., D38 réteg, méretarány: 250 µm; 11: *Hemicocytheria omphalodes* (REUSS), juvenilis, RV oldalnézetben, D36 réteg, méretarány: 200 µm; 12-13: *Aurila notata* (REUSS), 12: RV oldalnézetben, 13: LV oldalnézetben, D50 réteg, méretarány: 500 µm; 14-15: *Loxococoncha kochi* MÉHES, 14: LV oldalnézetben, 15: RV oldalnézetben, D50 réteg, méretarány: 250 µm; 16: *Loxococoncha laeta* STANCHEVA, LV oldalnézetben, D54 réteg, méretarány: 200 µm; 17: *Loxococoncha porosa* MÉHES, RV oldalnézetben, D54 réteg, méretarány: 200 µm; 18: *Loxocauda* sp., D38 réteg, méretarány: 250 µm; 19-20: *Loxocorniculum hastatum* (REUSS), 19: LV oldalnézetben, 20: RV oldalnézetben, D36 réteg, méretarány: 250 µm; 21: *Darwinula stevensoni* (BRADY & ROBERTSON), C jobb oldali, D40 réteg, méretarány: 250 µm; 22-23: *Vestalenula pagliolii* (PINTO & KOTZIAN), 22: RV oldalnézetben, 23: C baloldali nézetben, D38 réteg, méretarány: 200 µm; 24: *Fabaeformiscandona* ? sp. juv., RV oldalnézetben, D38 réteg, méretarány: 200 µm; 25: *Limnocythere* sp., LV oldalnézetben, D38 réteg, méretarány: 200 µm; 26: *Heterocypris salina* (BRADY), C bal oldali nézetben, D40 réteg, méretarány: 500 µm

Rövidítések: LV= bal teknő, RV= jobb teknő, C= carapace

## Plate III – III. tábla

Sarmatian foraminifers and other microfossils from the studied exploratory trench in Pécs-Danitzpuszta: 1: *Articulina* sp. indet., fragmented specimen, layer D41, scale bar: 200 µm; 2: *Bolivina sarmatica* DIDKOWSKI, SV, layer D41, scale bar: 250 µm; 3: *Buliminella elegantissima* (D'ORBIGNY), SV, layer D41, scale bar: 500 µm; 4: *Ammonia* cf. *confertitesta* ZHENG, UV, layer D41, scale bar: 200 µm; 5: *Porosonion granosum* (D'ORBIGNY), SV, layer D54, scale bar: 250 µm; 6: *Elphidium hauerinum* (D'ORBIGNY), SV, layer D54, scale bar: 200 µm; 7-8: *Elphidium aculeatum* (D'ORBIGNY), SV, layer D54, scale bar: 250 µm; 9: *Porosonion granosum* (D'ORBIGNY), layer D37; 10: *Vestalenula pagliolii* (PINTO & KOTZIAN), layer D38; 11-12: *Fabaeformiscandona* ? sp. juv., layer D38; 13: *Calcisphaera*-like large algal cyst, layer D35; 14: Mysid statolith, layer D35.

Abbreviations: SV= side view, UV= umbilical view

Szarmata foraminiferák és egyéb mikrofosztiliák a pécs-danitzpusztai homokbányában ásott kutatóárokából: 1: *Articulina* sp. indet., töredékes példány, D41 réteg, méretarány: 200 µm; 2: *Bolivina sarmatica* DIDKOWSKI, SV, D41 réteg, méretarány: 250 µm; 3: *Buliminella elegantissima* (D'ORBIGNY), SV, D41 réteg, méretarány: 500 µm; 4: *Ammonia* cf. *confertitesta* ZHENG, UV, D41 réteg, méretarány: 200 µm; 5: *Porosonion granosum* (D'ORBIGNY), SV, D54 réteg, méretarány: 250 µm; 6: *Elphidium hauerinum* (D'ORBIGNY), SV, D54 réteg, méretarány: 200 µm; 7-8: *Elphidium aculeatum* (D'ORBIGNY), SV, D54 réteg, méretarány: 250 µm; 9: *Porosonion granosum* (D'ORBIGNY), D37 réteg; 10: *Vestalenula pagliolii* (PINTO & KOTZIAN), D38 réteg; 11-12: *Fabaeformiscandona* ? sp. juv., D38 réteg; 13: *Calcisphaera*-hoz hasonló nagyméretű alga ciszta, D35 réteg; 14: *Misida* statolith, D35 réteg.

Rövidítések: SV= oldalnézet, UV= köldökoldali nézet



Plate I – I. tábla

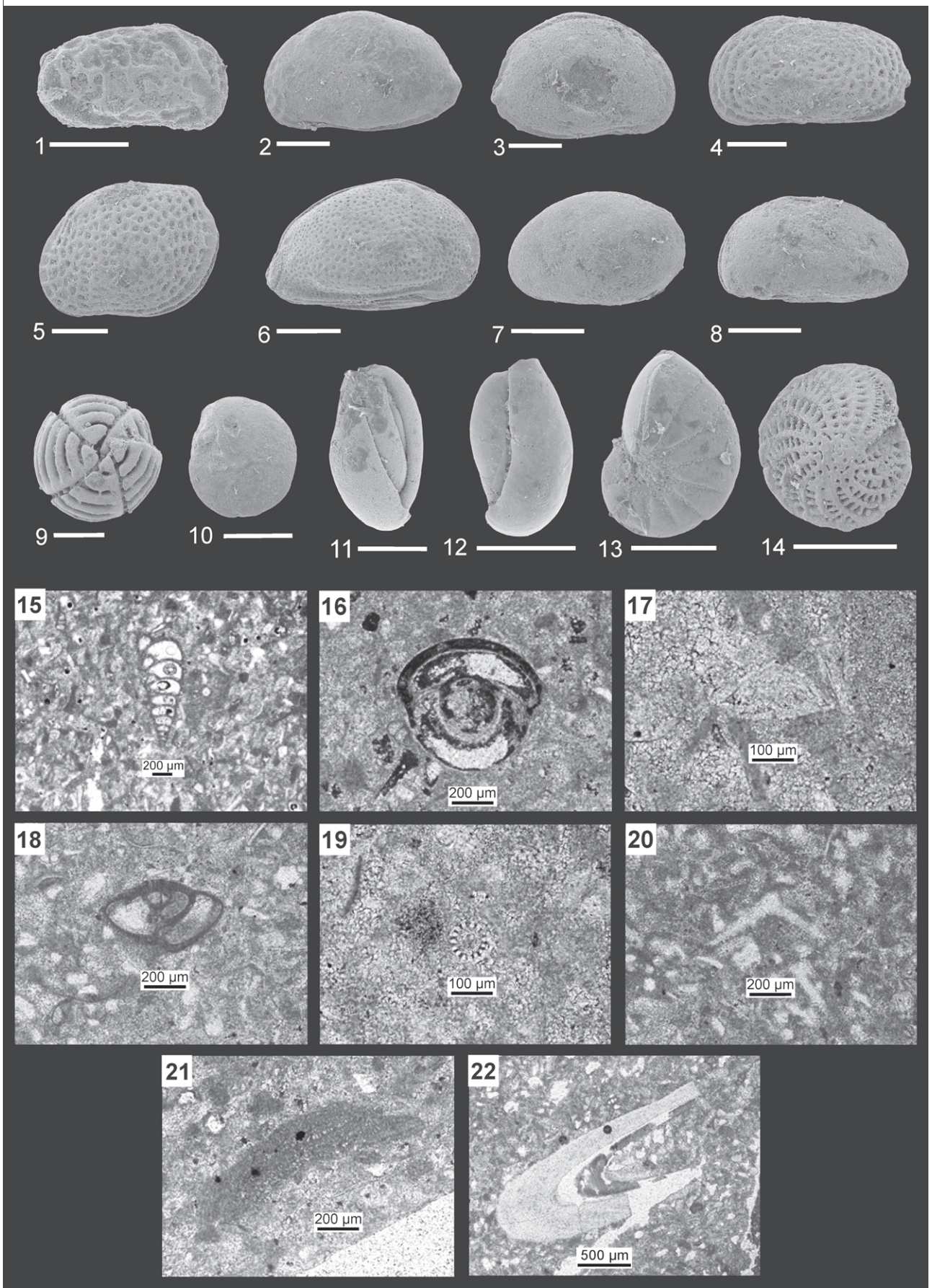
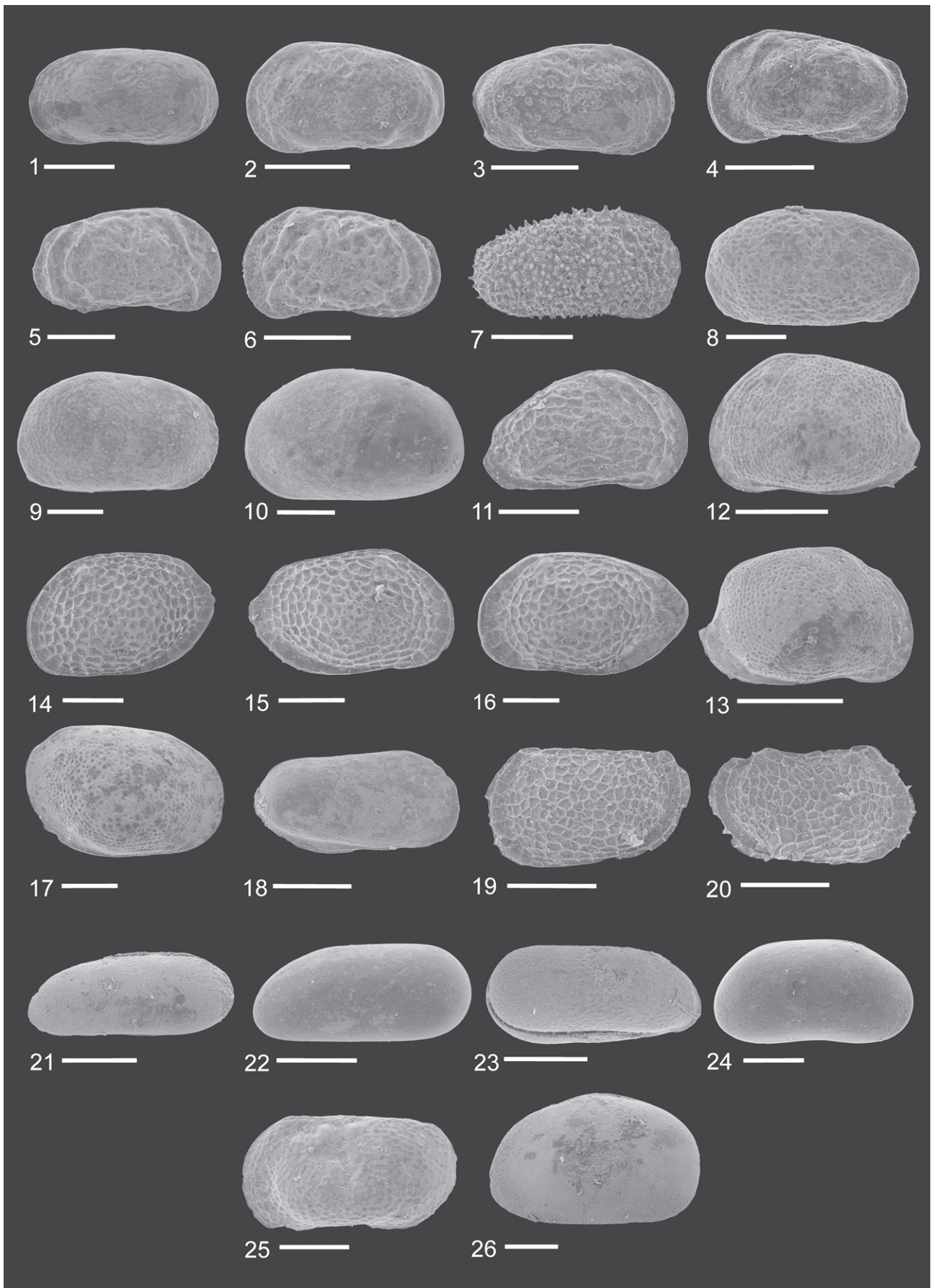
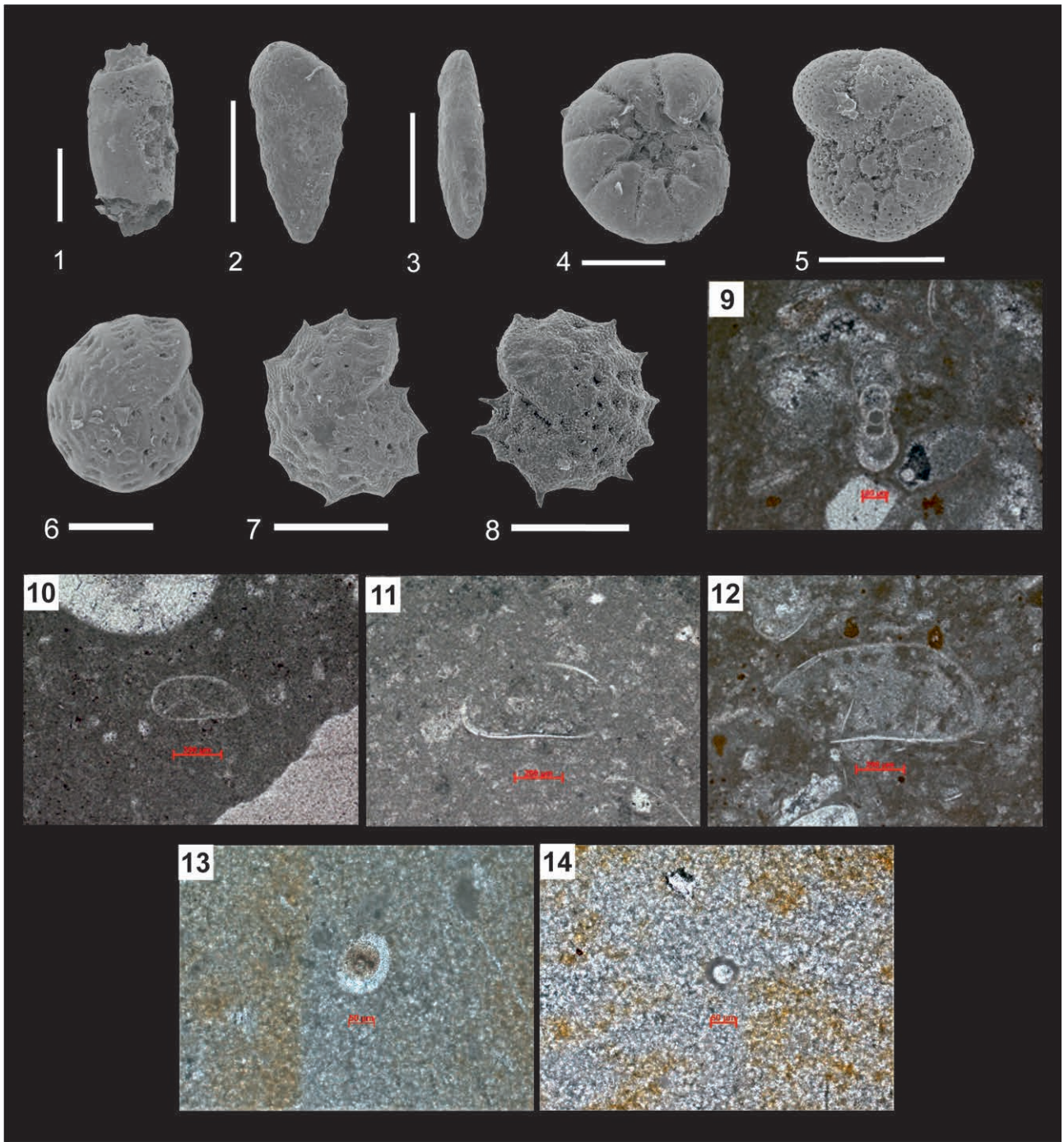




Plate II – II. tábla



## Plate III – III. tábla





## Appendix

### Systematic Palaeontology

The specimens of foraminifers and ostracods are deposited in the Laboratory of MOL Plc., Exploration and Production Division (Budapest, Hungary).

#### Foraminifera

Higher classification of the foraminifera follows that of LOEBLICH & TAPPAN (1992). Abbreviations: L: length, B: breadth, D: diameter and Th: thickness.

Phylum Protista

Subphylum Sarcodina SCHMARDT, 1871

Class Foraminifera J.J. LEE, 1990

Order Miliolida LANKESTER, 1885

Suborder Miliolina DELAGE & HERUARD, 1896

Superfamily Milioloidea EHRENBERG, 1839

Family Hauerinidae SCHWAGER, 1876

Subfamily Hauerininae SCHWAGER, 1876

Genus *Cycloforina* LUCZKOWSKA, 1972

*Cycloforina contorta* (D'ORBIGNY, 1846)

Plate I, fig. 11

1846 *Quinqueloculina contorta* n. sp. D'ORBIGNY, p. 298, pl. 20, figs 4–6.

2008 *Cycloforina contorta* (D'ORBIGNY) – TÓTH & GÖRÖG, p. 196, pl. 1, fig. 1. (cum syn.)

2012 *Cycloforina contorta* (D'ORBIGNY) – GONERA, fig. 2/M.

2012 *Cycloforina contorta* (D'ORBIGNY) – MILKER & SCHMIEDL, pp. 53–54, fig. 14/6.

2014 *Cycloforina contorta* (D'ORBIGNY) – YOKES et al., fig. 8/2.

2016 *Cycloforina contorta* (D'ORBIGNY) – LEI & LI, pp. 98–99, fig. 6.

2016 *Cycloforina contorta* (D'ORBIGNY) – KIRCI-ELMAS & MERIC, fig. 3/8.

*Dimensions:* L= 530–540 µm, B=400–410 µm, Th= 220–240 µm

*Stratigraphic range and geographic distribution:* Miocene: Carpathian Foredeep and Transcarpathian Basin, Ukraine (BOGDANOWICH 1952, DIDKOWSKY & SATANOVSKAJA 1970); Badenian: Carpathian Foredeep, Poland (LUCZKOWSKA 1974; GONERA 2012), Vienna Basin, Austria (D'ORBIGNY 1846); Badenian to Sarmatian: Mecsek Mts, Hungary (KORCZ-LAKY 1968); Sarmatian: Zsámbék Basin and Budapest, Hungary (GÖRÖG 1992, TÓTH & GÖRÖG 2008). Recently widely distributed over the world.

Subfamily Miliollinellinae VELLA, 1957

Genus *Affinetrina* LUCZKOWSKA, 1972

*Affinetrina ucrainica* (SEROVA, 1952)

Plate I, fig. 12

1952 *Miliolina ucrainica* n. sp. SEROVA in BOGDANOWICH, p. 104, pl. 8, fig. 2.

1992 *Affinetrina ucrainica* (SEROVA) – GÖRÖG, pp. 79–80, pl. 6, figs 1–3. (cum syn.)

2007 *Affinetrina ucrainica* (SEROVA) – SCHÜTZ et al., p. 453, pl. 2, fig. 2.

2012 *Affinetrina ucrainica* (SEROVA) – MILKER & SCHMIEDL, p. 61, fig. 16/11–13.

2015. *Affinetrina ucrainica* (SEROVA) – SILYE, p. 111, pl. 1, figs 4–5.

*Dimensions:* L= 500–530 µm, B=200–300 µm, Th= 160–220 µm

*Stratigraphic range and geographic distribution:* Upper Badenian: Carpathian Foredeep, Poland (LUCZKOWSKA 1974); Upper Badenian – Sarmatian: Transcarpathian Basin and Carpathian Foredeep, Ukraine (DIDKOWSKY & SATANOVSKAJA 1970); Lower Sarmatian: Vienna Basin, Austria (SCHÜTZ et al. 2007); Sarmatian: Moesian Platform, Bulgaria (STANCHEVA 1960), Transylvanian Basin, Romania (SILYE 2015); Zsámbék Basin, Hungary (GÖRÖG 1992). Recently widely distributed in the Mediterranean Sea.

Genus *Pyrgo* DEFRANCE, 1824

*Pyrgo subsphaerica* (D'ORBIGNY, 1839)

Plate I, fig. 16 (thin section)

1839 *Biloculina subsphaerica* n. sp. D'ORBIGNY, p. 162, pl. 8, figs 25–27.

1974 *Pyrgo subsphaerica* (D'ORBIGNY) – LUCZKOWSKA, pp. 118–119, pl. 22, figs 4a, b.

2008 *Pyrgo subsphaerica* (D'ORBIGNY) – DE ARAÚJO & MACHADO, pl. 1, fig. 3.

*Dimensions:* B= 660 µm (other dimensions are not examined)

*Stratigraphic range and geographic distribution:* Upper Badenian: Carpathian Foredeep, Poland (LUCZKOWSKA 1974). Recently widely distributed in the Mediterranean Sea, Caribbean Sea and Atlantic Ocean.

Order Buliminida FURSENKO, 1958  
 Superfamily Bolivinoidea GLAESSNER, 1937  
 Family Bolivinidae GLAESSNER, 1937  
 Genus *Bolivina* D'ORBIGNY, 1839

*Bolivina sarmatica* DIDKOWSKY, 1959

Plate III, fig. 2

- 1970 *Bolivina sarmatica* DIDKOWSKY – DIDKOWSKY & SATANOVSKAJA, p. 144, pl. 82, fig. 9. (holotype)  
 2008 *Bolivina sarmatica* DIDKOWSKY – TÓTH & GÖRÖG, p. 198, pl. 1, fig. 12. (cum syn.)  
 2011 *Bolivina sarmatica* DIDKOWSKY – GARECKA & OLSZEWSZKA, fig. 6/e.  
 2011 *Bolivina sarmatica* DIDKOWSKY – FILIPESCU et al., fig. 5/3.  
 2014 *Bolivina sarmatica* DIDKOWSKY – FILIPESCU et al., fig. 5/19.  
 2015 *Bolivina sarmatica* DIDKOWSKY – SILYE, p. 129, pl. 4, fig. 17.  
 2017 *Bolivina sarmatica* DIDKOWSKY – DUMITRIU et al., fig. 13/p.  
 2018 *Bolivina sarmatica* DIDKOWSKY – HARZHAUSER et al., fig. 5/10.

*Dimensions:* L= 150–160 µm, B= 90–95 µm

*Stratigraphic range and geographic distribution:* Sarmatian: Moldavian Plateau (DIDKOWSKY & SATANOVSKAJA 1970), Transcarpathian Basin, Carpathian Foredeep, Volhynian-Podolian Plateau, Ukraine (VENGLINSKY 1975), Western Carpathians (CICHA & ZAPLETALOVÁ 1961), easternmost Pannonian and Transylvanian Basins, Romania (FILIPESCU 1996; FILIPESCU et al. 2011, 2014), Zsámbék Basin and Budapest, Hungary (GÖRÖG 1992, TÓTH & GÖRÖG 2008), Carpathian Foredeep, Poland and Romania (GARECKA & OLSZEWSZKA 2011, DUMITRIU et al. 2017), Vienna Basin, Austria (HARZHAUSER et al. 2018).

Superfamily Buliminoidea JONES, 1875  
 Family Buliminellidae HOFKER, 1951  
 Genus *Buliminella* CUSHMAN, 1911

*Buliminella elegantissima* (D'ORBIGNY, 1839)

Plate III, fig. 3

- 1839 *Bulimina elegantissima* n. sp. D'ORBIGNY, p. 51, pl. 7, figs 13–14.  
 2004 *Buliminella elegantissima* (D'ORBIGNY) – VILELA et al., fig. 4/4.  
 2008 *Buliminella elegantissima* (D'ORBIGNY) – TÓTH & GÖRÖG, pp. 198–199, pl. 2, figs 2–4. (cum syn.)  
 2011 *Buliminella elegantissima* (D'ORBIGNY) – FILIPESCU et al., fig. 5/10.  
 2014 *Buliminella elegantissima* (D'ORBIGNY) – FILIPESCU et al., fig. 6/13.

*Dimensions:* L= 230–320 µm, D= 90–100 µm

*Stratigraphic range and geographic distribution:* Sarmatian: Black Sea Depression, Ukraine, Moldavian Plateau (DIDKOWSKY & SATANOVSKAJA 1970), Zsámbék Basin and Budapest, Hungary (GÖRÖG 1992, TÓTH & GÖRÖG 2008), easternmost Pannonian and Transylvanian Basins, Romania (FILIPESCU et al. 2011, 2014). Recently widely distributed over the world.

Superfamily Asterigerinoidea D'ORBIGNY, 1839  
 Family Asterigerinatidae REISS, 1963  
 Genus *Asterigerinata* REISS, 1963

*Asterigerinata planorbis* (D'ORBIGNY, 1846)

Plate I, fig. 17 (thin-section)

- 1846 *Asterigerina planorbis* n. sp. D'ORBIGNY, p. 225, pl. 11, figs 1–3.  
 1985 *Asterigerina planorbis* D'ORBIGNY – PAPP & SCHMID, pl. 66, figs 9–14.  
 1985 *Asterigerina planorbis* D'ORBIGNY – KORECZ-LAKY & NAGY-GELLAI, pl. 158, figs 1–4.  
 1998 *Asterigerinata planorbis* (D'ORBIGNY) – CICHA et al., pl. 64, figs 8–10.  
 1998 *Asterigerinata planorbis* (D'ORBIGNY) – ZLINSKÁ, pl. 8, figs 10–11.  
 2007 *Asterigerinata planorbis* (D'ORBIGNY) – SCHÜTZ et al., p. 457, pl. 4, fig. 6.  
 2010 *Asterigerina planorbis* D'ORBIGNY – ISMAIL et al., pl. 4, figs 4–5.  
 2012 *Asterigerinata planorbis* (D'ORBIGNY) – GONERA, fig. 4/c.  
 2013 *Asterigerinata planorbis* (D'ORBIGNY) – PEZELJ et al., fig. 6/17.  
 2016 *Asterigerinata planorbis* (D'ORBIGNY) – PEZELJ et al., fig. 5/A–H.  
 2014 *Biasterigerina planorbis* (D'ORBIGNY) – TER BORGH et al., fig. 5/31–32.  
 2019 *Asterigerinata planorbis* (D'ORBIGNY) – JOVANOVIĆ et al., pl. 1, figs f/6, g/5, h/6.  
 2020 *Asterigerinata planorbis* (D'ORBIGNY) – PERYT et al., fig. 4/h.

*Dimensions:* D= 250–350 µm

*Stratigraphic range and geographic distribution:* Kiscellian: Börzsöny Mts, Hungary (KORECZ-LAKY & NAGY-GELLAI 1985); Badenian: Vienna Basin, Austria (D'ORBIGNY 1846), East-Slovakian Basin (ZLINSKÁ 1998), Dacian Basin, Romania (TER BORGH et al. 2014), Mt Majevica, Bosnia and Herzegovina (PEZELJ et al. 2013, 2016); Koceljjeva area, Western Serbia (JOVANOVIĆ et al. 2019); Carpathian Foredeep, Poland (GONERA 2012, PERYT et al. 2020), North-Croatian Basin, Croatia (PEZELJ et al. 2016); Lower Sarmatian: Vienna Basin, Austria (SCHÜTZ et al. 2007); Pliocene: Nile Delta, Egypt (ISMAIL et al. 2010).

Superfamily Nonionoidea SCHULTZE, 1854  
 Family Nonionidae SCHULTZE, 1854  
 Subfamily Nonioninae SCHULTZE, 1854  
 Genus *Nonion* MONTFORT, 1808

*Nonion commune* (D'ORBIGNY, 1846)

Plate I, fig. 13

- 1798 *Nautilus scapha* n. sp. FICHTEL & MOLL, p. 105, pl. 19, figs d–f.  
 1846 *Nonionina communis* D'ORBIGNY – D'ORBIGNY, p. 106, pl. 5, figs 7–8.  
 2008 *Nonion commune* (D'ORBIGNY) – TÓTH & GÖRÖG, pp. 22–203, pl. 2, figs 14–18. (cum syn.)  
 2009 *Nonion commune* (D'ORBIGNY) – GEBHARDT et al., pl. 2, fig. 39.  
 2010 *Nonion commune* (D'ORBIGNY) – KOUBOVÁ & HUDÁČKOVÁ, pl. 1, fig. 15.  
 2012 *Nonion commune* (D'ORBIGNY) – FERRER GARCÍA & BLÁZQUEZ MORILLA, fig. 4/6.

- 2012 *Nonion commune* (D'ORBIGNY) – GONERA, fig. 4/e.  
 2013 *Nonion commune* (D'ORBIGNY) – PERYT, fig. 4/F.  
 2013 *Nonion commune* (D'ORBIGNY) – PEZELJ et al., fig. 6/18.  
 2014 *Nonion commune* (D'ORBIGNY) – FILIPESCU et al., fig. 6/7.  
 2019 *Nonion commune* (D'ORBIGNY) – JOVANOVIĆ et al., pl. 1, figs f/14, g/6, h/3.  
 2019 *Nonion commune* (D'ORBIGNY) – ROSLIM et al., fig. 4/33–36.

*Dimensions:* D= 350–370 µm, Th= 130–160 µm

*Stratigraphic range and geographic distribution:* Karpatian: Molasse Basin, Austria (RÖGL 1969); Badenian: Vienna Basin, Austria and Slovakia (D'ORBIGNY 1846, KOVÁČOVÁ & HUDAČKOVÁ 2005), Carpathian Foredeep, Poland (SZCZUCHURA 1982, PERYT 2013); Volhynian-Podolian Plateau, Carpathian Foredeep, Transcarpathian Basin, Crimea-Caucasus region and Kuban Lowland, Ukraine and Russia (VOLOSHINOVA 1952, DIDKOWSKY & SATANOVSKAJA 1970), Slovenia (OBLAK 2007), Mt Majejica, Bosnia and Herzegovina (PEZELJ et al. 2013), Koceljeva area, Western Serbia (JOVANOVIĆ et al. 2019); Badenian to Sarmatian: Mecsek Mts, Tokaj Mts and SW-Hungary, Budapest (KORECZ-LAKY 1968, 1973, 1982; BÁLDI 1999; TÓTH & GÖRÖG 2008), Appenines, Italy (DIECI 1959); Sarmatian: E-Slovakian Basin, Slovakia (ZLINSKÁ 1997, KOUBOVÁ & HUDAČKOVÁ, 2010), Vienna Basin, Austria (SCHÜTZ et al. 2007, GEBHARDT et al. 2009), easternmost Pannonian Basin, Romania (FILIPESCU et al. 2014); Upper Miocene: Ambug Hill, Borneo (ROSLIM et al. 2019). Recently widely distributed over the world.

Superfamily Rotalioidea EHRENBERG, 1839

Family Rotaliidae EHRENBERG, 1839

Subfamily Ammoniinae SAIDOVA, 1981

Genus *Ammonia* BRÜNNICH, 1772

*Ammonia* cf. *confertitesta* ZHENG, 1978

Plate III, fig. 4

*Dimensions:* D= 250–450 µm

*Remarks:* The studied specimens are very similar (mainly the spiral side of the test) to the holotype described by ZHENG (1978) however the last chamber of the studied specimens in most cases is missing.

Family Elphidiidae GALLOWAY, 1933

Subfamily Elphidiinae GALLOWAY, 1933

Genus *Elphidium* MONTFORT, 1808

*Elphidium aculeatum* (D'ORBIGNY, 1846)

Plate III, figs 7–8

- 1846 *Polystomella josephina* n. sp. D'ORBIGNY, p. 130, pl. 6, figs 25–26.  
 1846 *Polystomella aculeata* n. sp. – D'ORBIGNY, p. 131, pl. 6, figs 27–28.  
 1995. *Elphidium aculeatum* (D'ORBIGNY) – POPESCU, p. 94, pl. 7, figs 4–7.  
 2004 *Elphidium aculeatum* (D'ORBIGNY) – BRÂNZILĂ, pl. 4, fig. 5.  
 2004 *Elphidium aculeatum* (D'ORBIGNY) – MERIÇ et al., pl. 32, figs 5–8.  
 2005 *Elphidium aculeatum* (D'ORBIGNY) – GOLDBECK et al., pl. 1, fig. 12.  
 2008 *Elphidium aculeatum* (D'ORBIGNY) – TÓTH & GÖRÖG, pp. 204–205, pl. 3, figs 5–6. (cum syn.)

- 2010 *Elphidium josephinum* (D'ORBIGNY) – KOUBOVA & HUDAČKOVÁ, pl. 1, fig. 26.  
 2011 *Elphidium aculeatum* (D'ORBIGNY) – GEDL & PERYT, pl. 1, fig. 9/F, I–K.  
 2012 *Elphidium aculeatum* (D'ORBIGNY) – ALOULOU et al., pl. 1, fig. 13.  
 2012 *Elphidium aculeatum* (D'ORBIGNY) – MILKER & SCHMIEDL, p. 119, fig. 27/5–6.  
 2012 *Elphidium aculeatum* (D'ORBIGNY) – GONERA et al., fig. 4/K.  
 2012 *Elphidium aculeatum* (D'ORBIGNY) – MELIS et al., pl. 1, fig. 1.  
 2012 *Elphidium aculeatum* (D'ORBIGNY) – PERYT & JASIONOWSKI, fig. 4/C, D, L, M.  
 2012 *Elphidium aculeatum* (D'ORBIGNY) – JASIONOWSKI et al., fig. 12/A, B, D, E.  
 2013 *Elphidium aculeatum* (D'ORBIGNY) – TER BORGH et al., fig. 6, 8–9.  
 2014 *Elphidium josephinum* (D'ORBIGNY) – FILIPESCU et al., fig. 7/10.  
 2014 *Elphidium aculeatum* (D'ORBIGNY) – YOKES et al., fig. 11/8.  
 2015 *Elphidium aculeatum* (D'ORBIGNY) – SILYE, p. 150, pl. 8, figs 1–2, 4.  
 2017 *Elphidium aculeatum* (D'ORBIGNY) – DUMITRIU et al., fig. 11/Q, R.  
 2020 *Elphidium aculeatum* (D'ORBIGNY) – PERYT et al., fig. 3/h.

*Dimensions:* D= 450–600 µm, Th= 200–350 µm.

*Stratigraphic range and geographic distribution:*

Badenian: Carpathian Foredeep, Poland and Ukraine (GEDL & PERYT 2011, GONERA et al. 2012, PERYT et al. 2020); Late Badenian to Sarmatian: Volhynian-Podolian Plateau, Moldavian Plateau, Moldavia and Carpathian Foredeep, Ukraine (VENGLINSKY 1958; DIDKOWSKY & SATANOVSKAJA 1970, BRÂNZILĂ 2004), Crimea-Caucasus region, South-Caspian Depression, Russia and Azerbaijan (VOLOSHINOVA 1952); Sarmatian: Carpathian Foredeep, Poland, Romania and Ukraine (SZCZUCHURA 1982, 2000; JASIONOWSKI et al. 2012; DUMITRIU et al. 2017), Vienna Basin, Austria and Slovakia (MARKS 1951, PAPP 1963; SCHÜTZ et al. 2007; KOUBOVA & HUDAČKOVÁ 2010), Danube Basin and East-Slovakian Basin, Slovakia (BRESTENSKÁ 1974; ZLINSKÁ 1997), Tokaj Mts, Mecsek Mts, Zsámberk Basin and Budapest, Hungary (KORECZ-LAKY 1973, 1968, 1964, 1965, 1982; GÖRÖG 1992; TÓTH & GÖRÖG 2008), easternmost Pannonian and Transylvanian basins, Romania (KOVÁCS 2001, SUCIU 2005, FILIPESCU et al. 2014, SILYE 2015); Romanian Plain, Romania (POPESCU 1995), Moesian Platform, Bulgaria (STANCHEVA 1960); Pannonian Basin, Serbia (TER BORGH et al. 2013); Holocene: Mediterranean Sea, Italy (MELIS et al. 2012, YOKES et al. 2014). Recently widely distributed over the world.

*Remarks:* The number and size of spines are variable, it seems to be intraspecific variability. Making the species *Elphidium josephinum* described by D'ORBIGNY the junior synonym of *E. aculeatum*, thus an invalid name.

*Elphidium crispum* (LINNE, 1758)

Plate I, fig. 14

- 1758 *Nautilus crispus* n. sp. LINNAEUS, p. 709, pl. 1, figs 2d–e.  
 1988 *Elphidium crispum* (LINNÉ) – JORISSEN, p. 120, pl. 3, figs 8–9, pl. 24, figs 1–2.



- 2004 *Elphidium crispum* (LINNÉ) – MERIÇ et al., pl. 1, figs 16–18.  
 2004 *Elphidium crispum* (LINNÉ) – MENDES et al., pl. 1, fig. 6.  
 2004 *Elphidium crispum* (LINNÉ) – BRÂNZILĂ, pl. 4, fig. 11.  
 2005 *Elphidium crispum* (LINNÉ) – MORIGI et al., pl. 2, fig. 9a–c.  
 2008 *Elphidium crispum* (LINNÉ) – TÓTH & GÖRÖG, pp. 205–206, pl. 3, figs 7–8. (cum syn.)  
 2009 *Elphidium crispum* (LINNÉ) – FREZZA & CARBONI, pl. 1, fig. 16.  
 2010 *Elphidium crispum* (LINNÉ) – KOUBOVÁ & HUDAČKOVÁ, pl. 1, fig. 24.  
 2011 *Elphidium crispum* (LINNÉ) – GEDL & PERYT, fig. 9/C, R.  
 2012 *Elphidium crispum* (LINNÉ) – FERRER GARCÍA & BLÁZQUEZ MORILLA, pl. 4, fig. 12.  
 2012 *Elphidium crispum* (LINNÉ) – GONERA, fig. 4/j.  
 2012 *Elphidium crispum* (LINNÉ) – MILKER & SCHMIEDL, p. 120, fig. 27/13–14.  
 2012 *Elphidium crispum* (LINNÉ) – MELIS et al., pl. 1, fig. 4.  
 2012 *Elphidium crispum* (LINNÉ) – ALOULOU et al., pl. 1, fig. 15.  
 2014 *Elphidium crispum* (LINNÉ) – FILIPESCU et al., fig. 7/3.  
 2014 *Elphidium crispum* (LINNÉ) – YOKES et al., fig. 11/10–11.  
 2014 *Elphidium crispum* (LINNÉ) – TER BORGH et al., fig. 6/8.  
 2016 *Elphidium crispum* (LINNÉ) – LEI & LI, p. 361, fig. 84.  
 2016 *Elphidium crispum* (LINNÉ) – DIMIZA et al., pl. 4, fig. 20.  
 2016 *Elphidium crispum* (LINNÉ) – PEZELJ et al., fig. 5/D, I.  
 2019 *Elphidium crispum* (LINNÉ) – JOVANOVIĆ et al., pl. 1, figs f/3, g/7.  
 2019 *Elphidium crispum* (LINNÉ) – ROSLIM et al., fig. 4/25.

*Dimensions:* D= 450–1200 µm, Th=330–350 µm

*Stratigraphic range and geographic distribution:* Langhian: Aquitaine Basin, France (CAHUZAC & POIGNANT 2000); Karpatian-Badenian: East-Mecsek Mts, Hungary (KORECZ-LAKY 1968); Badenian: Dacian Basin, Romania and Serbia (TER BORGH et al. 2014), Carpathian Foredeep, Poland and Ukraine (GEDL & PERYT 2011, GONERA 2012), Vienna Basin, Austria (PAPP 1963), Apennines, Italy (DIECI 1959), Koceljjeva area, Western Serbia (JOVANOVIĆ et al. 2019); Badenian: North-Croatian Basin, Croatia (PEZELJ et al. 2016), Karpatian–Sarmatian: Transcarpathian Basin, Volhynian-Podolian Plateau and Caucasus, Ukraine and Russia (VENGLINSKY 1958, DIDKOWSKY & SATANOVSKAJA 1970); Sarmatian: Carpathian Foredeep, Poland (SZCZUCHURA 1982), Mecsek Mts, Zsámbék Basin and Budapest, Hungary (KORECZ-LAKY 1964, 1968; GÖRÖG 1992; TÓTH & GÖRÖG 2008), Vienna Basin, Slovakia (KOUBOVÁ & HUDAČKOVÁ 2010); Moldavian Plateau, Moldavia (BRÂNZILĂ 2004); Lower Sarmatian: easternmost Pannonian Basin, Romania (FILIPESCU et al. 2014); Upper Miocene: Ambug Hill, Borneo (ROSLIM et al. 2019); Pliocene: Toscana, Italy (FICHEL & MOLL 1798); Holocene: Mediterranean Sea, Italy (MORIGI et al. 2005, MELIS et al. 2012). Recently widely distributed over the world.

*Elphidium hauerinum* (D'ORBIGNY, 1846)

Plate III, fig. 6

- 1846 *Polystomella Hauerina* n. sp. D'ORBIGNY, p. 122, pl. 6, figs 5–10.  
 1995 *Elphidium hauerinum* (D'ORBIGNY) – POPESCU, p. 95, pl. 8, fig. 10.  
 2005 *Elphidium hauerinum* (D'ORBIGNY) – FILIPESCU et al., pl. 2, figs 4–5.  
 2008 *Elphidium hauerinum* (D'ORBIGNY) – TÓTH & GÖRÖG, pl. 3, figs 10–12. (cum syn.)  
 2010 *Elphidium hauerinum* (D'ORBIGNY) – KOUBOVÁ & HUDAČKOVÁ, pl. 1, fig. 18.  
 2011 *Elphidium hauerinum* (D'ORBIGNY) – FILIPESCU et al., fig. 4/3.  
 2011 *Elphidium hauerinum* (D'ORBIGNY) – IONESI & PASCARIU, pl. 1, fig. 29.  
 2012 *Elphidium hauerinum* (D'ORBIGNY) – JASIONOWSKI et al., fig. 14/E, H, I, M.  
 2014 *Elphidium hauerinum* (D'ORBIGNY) – FILIPESCU et al., figs 7–9.  
 2015 *Elphidium hauerinum* (D'ORBIGNY) – SILYE, p. 152, pl. 8, figs 5–7.  
 2017 *Elphidium hauerinum* (D'ORBIGNY) – DUMITRIU et al., fig. 12/G, H.

*Dimensions:* D=240–430 µm, Th= 100–150 µm

*Stratigraphic range and geographic distribution:* Badenian?: Vienna Basin, Austria (D'ORBIGNY 1846); Badenian-Sarmatian: Transcarpathian Basin, Carpathian Foredeep, Ukraine (VENGLINSKY 1958, DIDKOWSKY & SATANOVSKAJA 1970); Sarmatian: Carpathian Foredeep, Ukraine, Poland and Romania (JASIONOWSKI et al. 2012, DUMITRIU et al. 2017), Moldavian Plateau, Romania (IONESI & PASCARIU 2011), N-Caucasus, Russia (VOLOSHINOVA 1952); Moesian Platform, Bulgaria (STANCHEVA 1960), easternmost Pannonian and Transylvanian basins, Romania (FILIPESCU 1996; KOVÁCS 2001; SUCIU 2005; FILIPESCU et al. 2005, 2011, 2014), Romanian Plain, Romania (POPESCU 1995), Vienna Basin, Austria (D'ORBIGNY 1846, PAPP 1963, SCHÜTZ et al. 2007), Tokaj Mts, Zsámbék Basin, Mecsek Mts and Budapest, Hungary (KORECZ-LAKY 1964, 1965, 1968, 1973, 1982; GÖRÖG 1992; TÓTH & GÖRÖG 2008); Danube Basin and East-Slovakian Basin, Slovakia (BRESTENSKÁ 1974, ZLINSKÁ 1997, KOUBOVÁ & HUDAČKOVÁ 2010), Carpathian Foredeep, Poland (SZCZUCHURA 1982, 2000).

Genus *Porosonion* PUTRYA in VOLOSHINOVA, 1958

*Porosonion granosum* (D'ORBIGNY, 1846)

Plate III, figs 5, 9 (thin section)

- 1846 *Nonionina granosa* n. sp. D'ORBIGNY, p. 110, pl. 5, figs 19–20.  
 1988 *Elphidium granosum* (D'ORBIGNY) – JORISSEN, p. 104, pl. 2, figs 1–3, pl. 16–19.  
 1992 *Porosonion granosum* (D'ORBIGNY) – GÖRÖG, pp. 112–113, pl. 11, fig. 5. (cum syn.)  
 2000 *Porosonion granosum* (D'ORBIGNY) – POIGNANT et al., pp. 400–401, pl. 1, figs 13–14. (cum syn.)  
 2000 *Porosonion granosum* (D'ORBIGNY) – SZCZUCHURA, pl. 5, figs 3, 6.  
 2000 *Elphidium granosum* (D'ORBIGNY) – CARBONI et al., fig. 10.  
 2001 *Porosonion granosum* (D'ORBIGNY) – FILIPESCU et al., pl. 3, fig. 11.  
 2004 *Porosonion subgranosus monogranulata* GERKE – BRÂNZILĂ, pl. 2, figs 7–9.  
 2007 *Porosonion ex gr. granosum* (D'ORBIGNY) – SCHÜTZ et al., pl. 6, fig. 6.  
 2007 *Porosonion granosum* (D'ORBIGNY) – GROSS et al., pp. 210–211, fig. 4 a–e, h–i.  
 2008 *Criboelphidium ex gr. granosum* (D'ORBIGNY) – TÓTH & GÖRÖG, p. 204, pl. 3., figs 3–4.  
 non 2010 *Porosonion granosum* (D'ORBIGNY) – KOUBOVÁ & HUDAČKOVÁ, pl. 1, fig. 20.

- 2011 *Porosononion granosum* (D'ORBIGNY) – FILIPESCU et al., fig. 4/9.
- 2012 *Elphidium granosum* (D'ORBIGNY) – MILKER & SCHMIEDL, p. 121, fig. 27/17–18.
- 2013 *Porosononion granosum* (D'ORBIGNY) – TER BORGH et al., fig. 6/4–5.
- 2015 *Porosononion granosum* (D'ORBIGNY) – SILYE, p. 147, pl. 7, figs 4–5.
- 2018 *Porosononion granosum* (D'ORBIGNY) – HARZHAUSER et al., fig. 5/1–2.
- 2019 *Porosononion granosum* (D'ORBIGNY) – NÁÑEZ & MALUMIÁN, pp. 197–201, figs 5–6.
- 2014 *Heterolepa dutemplei* (D'ORBIGNY) – STOJANOVA & PETROV, pl. 1, fig. 11.
- 2016 *Heterolepa dutemplei* (D'ORBIGNY) – VALCHEV & STOJANOVA, pl. 2, figs 3–4.
- 2016 *Heterolepa dutemplei* (D'ORBIGNY) – PEZELJ et al., fig. 5/M
- 2017 *Heterolepa dutemplei* (D'ORBIGNY) – HARZHAUSER et al., pl. 2, fig. 13.
- 2017 *Heterolepa dutemplei* (D'ORBIGNY) – DUMITRIU et al., fig. 9/I, J.
- 2019 *Heterolepa dutemplei* (D'ORBIGNY) – JOVANOVIĆ et al., pl. 1, fig. h/1.
- 2019 *Heterolepa dutemplei* (D'ORBIGNY) – ROSLIM et al., fig. 4/8–13.

*Dimensions:* D=200–500 µm

*Stratigraphic range and geographic distribution:* Middle Miocene: Atlantic Ocean, Argentina (NÁÑEZ & MALUMIÁN 2019); Badenian: Transylvanian Basin, Romania (FILIPESCU 2001); Badenian–Sarmatian: Vienna Basin, Austria (D'ORBIGNY 1846); Sarmatian: Vienna Basin and Styrian Basin, Austria (GROSS et al. 2007, SCHÜTZ et al. 2007, HARZHAUSER et al. 2018), Zsámbék Basin and Budapest, Hungary (GÖRÖG 1992, TÓTH & GÖRÖG 2008), Transcarpathian Basin, Ukraine (VOLOSHINOVA 1952, VENGLINSKY 1958), Carpathian Foredeep, Poland (SZCZECURA 2000), Transylvanian Basin, Romania (FILIPESCU et al. 2011, SILYE 2015), Moldavian Plateau (BRÂNZILĂ 2004); Pliocene: Mediterranean Sea, Spain (CARBONNEL & MAGNÉ 1977) and Greece (HAGEMAN 1979). Recently widely distributed over the world.

*Remarks:* The umbilical region is very variable in this group. Due to the large morphological variation, the taxonomic status of fossil specimens is uncertain. The studied specimen is identical (including umbilical region) to the holotype described by D'ORBIGNY (1846).

Family Cibicididae CUSHMAN, 1927  
Subfamily Cibicidinae CUSHMAN, 1927  
Genus *Heterolepa* FRANZENAU, 1884

*Heterolepa dutemplei* (D'ORBIGNY, 1846)

Plate I, figs 10, 18 (thin section)

- 1846 *Rotalia dutemplei* n. sp. D'ORBIGNY, p. 157, pl. 8, figs 19–21.
- 1982 *Heterolepa dutemplei* (D'ORBIGNY) – SZCZECURA, pl. 16, figs 8–9.
- 1985 *Heterolepa dutemplei* (D'ORBIGNY) – PAPP & SCHMID, p. 59, pl. 50, figs 1–3.
- 1985 *Heterolepa dutemplei* (D'ORBIGNY) – KORECZ-LAKY & NAGY-GELLAI, pl. 20, fig. 4a–b.
- 1998 *Heterolepa dutemplei* (D'ORBIGNY) – CICHA et al., pp. 107–108, pl. 71, figs 1–3.
- 1999 *Heterolepa dutemplei* (D'ORBIGNY) – BÁLDI, pp. 209–210, pl. 9, figs 1–6, pl. 10, figs 1–2.
- 2000 *Heterolepa dutemplei* (D'ORBIGNY) – SZCZECURA, pl. 1, figs 6, 13.
- 2001 *Heterolepa dutemplei* (D'ORBIGNY) – FILIPESCU, pl. 3, figs 12–13.
- 2007 *Heterolepa dutemplei* (D'ORBIGNY) – OZSVÁRT, pp. 84–85, pl. 11, figs 11–13. (cum syn.)
- 2013 *Heterolepa dutemplei* (D'ORBIGNY) – PERYT, figs 4/V, W, 7/Y
- 2013 *Heterolepa dutemplei* (D'ORBIGNY) – PEZELJ et al., fig. 6/20.
- 2014 *Heterolepa dutemplei* (D'ORBIGNY) – TER BORGH et al., fig. 5/41–42.

*Dimensions:* D=450–600 µm, Th=200–350 µm

*Stratigraphic range and geographic distribution:* Middle to Upper Eocene: Paleogene Basin, Hungary (OZSVÁRT 2007); Upper Eocene – Lower Oligocene: Valandovo-Gevgelia Basin, Republic of Macedonia (STOJANOVA & PETROV 2014; VALCHEV & STOJANOVA 2016); Kiscellian to Badenian: Börzsöny Mts, Hungary (KORECZ-LAKY & NAGY-GELLAI 1985); SW-Hungary (BÁLDI 1999); Ottományian: Austria, Vienna Basin (HARZHAUSER et al. 2017); Badenian: Koceljjeva area, Western Serbia (JOVANOVIĆ et al. 2019); Mt Majejica, Bosnia and Herzegovina (PEZELJ et al. 2013); North-Croatian Basin, Croatia (PEZELJ et al. 2016), Austria, Vienna Basin (D'ORBIGNY 1846), Dacian and Transylvanian basins, Serbia and Romania (FILIPESCU 2001, TER BORGH et al. 2014); Carpathian Foredeep, Poland (SZCZECURA 1982, 2000; PERYT 2013; DUMITRIU et al. 2017), Upper Miocene: Ambug Hill, Borneo (ROSLIM et al. 2019).

## Ostracoda

Classification of the ostracods follows that of HARTMANN & PURI (1974) and HORNE et al. (2002). Abbreviations: L: length, H: height.

Phylum Arthropoda SIEBOLD, STANNIUS, 1845  
Subphylum Crustacea PENNANT, 1777  
Class Ostracoda LATREILLE, 1802  
Order Podocopida MÜLLER, 1894  
Suborder Cytherocopina BAIRD, 1850  
Superfamily Cytheroidea BAIRD, 1850  
Family Cytheroidea SARS, 1925  
Subfamily Cytheroidea SARS, 1925  
Genus *Cyprideis* JONES, 1857

*Cyprideis pokorny* JIŘIČEK, 1974

Plate II, figs 8–9

- 1974 *Cyprideis pokorny* n. sp. JIŘIČEK, p. 439, pl. 2, figs 1–4.
- 2009 *Cyprideis pokorny* JIŘIČEK – TÓTH, p. 87, pl. 4, figs 3, 6.

*Dimensions:* L=660–720 µm, H=350–410 µm, L/H=1.6–1.8.

*Stratigraphic range and geographic distribution:* Upper Sarmatian: Vienna Basin, Slovakia (JIŘIČEK 1974); Vértes Hill, Hungary (TÓTH 2009).

Family Hemicytheridae PURI, 1953  
 Subfamily Hemicytherinae PURI, 1953  
 Genus *Aurila* POKORNÝ, 1955

*Aurila cicatricosa* (REUSS, 1850)  
 Plate I, figs 2–3

- 1850 *Cypridina cicatricosa* n. sp. REUSS, pp. 67–68, pl. 9, fig. 21.  
 1962 *Mutilus (Aurila) cicatricosa* (REUSS) – STANCHEVA, p. 32, pl. 4, fig. 8.  
 1971 *Aurila cicatricosa* (REUSS) – CERNAJSEK, pp. 65–69, pl. 6, figs 7–14, pl. 14, fig. 7, pl. 17, fig. 4 a–b. [partim, pl. 14, fig. 8]  
 1978 *Aurila cicatricosa* (REUSS) – BRESTENSKÁ & JIŘIČEK, p. 409, 432, pl. 6, fig. 1.  
 2008 *Aurila cicatricosa* (REUSS) – FARANDA et al., pl. 2, figs 4–5.  
 2004 *Aurila cicatricosa* (REUSS) – AIELLO & SZCZUCHURA, pp. 28–30, pl. 5, fig. 2.  
 2006 *Aurila cicatricosa* (REUSS) – GROSS & PILLER, pp. 47–48, text-fig. 6/1, pl. 21, figs 1–12, pl. 22, figs 8–10.  
 2006 *Aurila cicatricosa* (REUSS) – SZCZUCHURA, fig. 9/9–10.  
 2012 *Aurila cicatricosa* (REUSS) – SEKO et al., fig. 8/P.  
 2014 *Aurila (Aurila) cicatricosa* (REUSS) – TER BORGH et al., fig. 7/16.

*Dimensions:* L= 900–950 µm, H= 530–580 µm, L/H= 1.6–1.7.

*Stratigraphic range and geographic distribution:* Badenian: Vienna Basin, Austria (CERNAJSEK 1971, GROSS & PILLER 2006); Carpathian Foredeep, Czech Republic, Poland (BRESTENSKÁ & JIŘIČEK 1978, AIELLO & SZCZUCHURA 2004, SZCZUCHURA 2006, SEKO et al. 2012); Dacian Basin, Romania (TER BORGH et al. 2014); Late Miocene: Mediterranean, Greece (FARANDA et al. 2008).

*Aurila notata* (REUSS, 1850)  
 Plate II, figs 12–13.

- 1850 *Cypridina notata* n. sp. REUSS, p. 66, pl. 9, fig. 16.  
 2006 *Aurila (Euaurila?) notata* (REUSS) – GROSS & PILLER, p. 83–84, pl. 29, figs 1–9.  
 2008 *Aurila notata* (REUSS) – TÓTH, pp. 122–123, pl. 8, figs 3–7. (cum syn.)  
 2017 *Aurila notata* (REUSS) – DUMITRIU et al., fig. 12/Q.  
 2018 *Aurila notata* (REUSS) – HARZHAUSER et al., fig. 7/10.

*Dimensions:* L= 900–950 µm, H= 530–580 µm, L/H= 1.6–1.7.

*Stratigraphic range and geographic distribution:* Upper Sarmatian: Vienna Basin, Austria and Slovakia (CERNAJSEK 1974, JIŘIČEK 1983, ZELENKA 1990, JANZ & VENNEMANN 2005, GROSS & PILLER 2006, HARZHAUSER et al. 2018); Zsámbék Basin, Hungary (TÓTH 2008); Caucasus, Russia (SUZIN 1956); Lower Sarmatian: Moldovian Plateau, Romania (DUMITRIU et al. 2017).

Genus *Hemicytheria* POKORNÝ, 1952

*Hemicytheria omphalodes* (REUSS, 1850)  
 Plate II, fig. 11

- 1850 *Cypridina omphalodes* n. sp. REUSS, p. 75, pl. 10, fig. 7.  
 2008 *Hemicytheria omphalodes* (REUSS) – TÓTH, pl. 6, figs 2–6. (cum syn.)

- 2011 *Hemicytheria omphalodes* (REUSS) – OLTEANU, pl. 18, fig. 8.  
 2014 *Hemicytheria omphalodes* (REUSS) – FILIPESCU et al., fig. 8/10.

*Dimensions:* L= 810–820 µm, H= 470–480 µm, L/H= 1.7–1.75.

*Stratigraphic range and geographic distribution:* Upper Badenian: Transylvanian Basin, Romania (OLTEANU 2001); Sarmatian: Vienna Basin, Slovakia (JIŘIČEK 1974, ZELENKA 1990); Zsámbék Basin, Hungary (TÓTH 2008); Lower Sarmatian: Danube Basin and the eastern region, Slovakia (FORDINÁL et al. 2006, FORDINÁL & ZLINSKÁ 1994); Upper Sarmatian: Vienna Basin, Austria (CERNAJSEK 1974); Pannonian: easternmost Pannonian Basin, Transylvanian Basin, Romania (OLTEANU 2001, 2011; FILIPESCU et al. 2014), Pannonian Basin, Croatia (SOKAČ 1972).

Genus *Senesia* JIŘIČEK, 1974

*Senesia cinctella* (REUSS, 1850)  
 Plate I, fig. 6

- 1850 *Cypridina cinctella* n. sp. REUSS, p. 67, pl. 9, fig. 19.  
 1962 *Mutilus (Aurila) cinctella* (REUSS) – STANCHEVA, p. 35, pl. 4, fig. 9.  
 1979 *Aurila (Aurila) cinctella* n. ssp. – BASSIOUNI, pp. 118–119, pl. 19, figs 7–8.  
 2006 *Senesia cinctella* (REUSS) – GROSS & PILLER, pp. 57–58, pl. 31, figs 1–5.

*Dimensions:* L= 750–760 µm, H= 410–420 µm, L/H= 1.8–1.82

*Stratigraphic range and geographic distribution:* Lower Miocene: Black Sea Depression, Turkey (BASSIOUNI 1979); Badenian: Vienna Basin, Austria and Slovakia (REUSS 1850, CERNAJSEK 1971, BRESTENSKÁ & JIŘIČEK 1978, GROSS & PILLER 2006); Moesian Plateau, Bulgaria (STANCHEVA 1962).

Subfamily Urocythereidinae HARTMANN & PURI, 1974  
 Genus *Urocythereis* RUGGIERI, 1950

*Urocythereis kostelensis* (REUSS, 1850)  
 Plate I, fig. 4

- 1850 *Cypridina kostelensis* n. sp. REUSS, p. 68, pl. 9, fig. 22.  
 1978 *Urocythereis kostelensis* (REUSS) – BRESTENSKÁ & JIŘIČEK, p. 410, 432, pl. 6, fig. 12.  
 1985 *Urocythereis kostelensis* (REUSS) – ZELENKA, p. 246, pl. 3, fig. 2.  
 2004 *Urocythereis kostelensis* (REUSS) – ZORN, p. 180, fig. 4/10–11.  
 2006 *Urocythereis kostelensis* (REUSS) – GROSS & PILLER, pp. 106–108, pl. 38, figs 1–5, 9, 11–12.

*Dimensions:* L= 820–835 µm, H= 410–420 µm, L/H= 1.9–2.

*Stratigraphic range and geographic distribution:* Badenian: Carpathian Foredeep, Poland (REUSS 1850), Vienna and Molasse basins, Austria and Slovakia (REUSS 1850, BRESTENSKÁ & JIŘIČEK 1978, ZELENKA 1985, ZORN 2004; GROSS & PILLER 2006).



Family Leptocytheridae HANAI, 1957  
 Subfamily Leptocytherinae HANAI, 1957  
 Genus *Amnicythere* DEVOTO, 1965

*Amnicythere cernajseki* STANCHEVA, 1984  
 Plate II, figs 2–3

1963 *Leptocythere modesta* n. sp. STANCHEVA, p. 22, pl. 3, fig. 8.  
 1974 *Leptocythere* sp. – CERNAJSEK, p. 476, pl. 2, fig. 7.  
 1984 *Amnicythere cernajseki* nom. nov. – STANCHEVA, p. 39, pl. 1, fig. 5.  
 1998 *Amnicyther* aff. *plana* (SCHNEIDER) – OLTEANU, p. 153, pl. 8, fig. 7.  
 2008 *Amnicythere* (?) sp. – TÓTH, p. 110, pl. 2, figs 5–6.  
 2011 *Amnicythere cernajseki* STANCHEVA – FILIPESCU et al., fig. 5/20.

*Dimensions:* L= 570–600 µm, H= 260–300 µm, L/H= 2–2.19.

*Stratigraphic range and geographic distribution:* Sarmatian: Vienna Basin, Austria (CERNAJSEK 1974); Lower Sarmatian: Transylvanian Basin, Romania (OLTEANU 1998); Upper Sarmatian: Zsámbék Basin, Hungary (TÓTH 2008); Transylvanian Basin, Romania (FILIPESCU et al. 2011).

*Amnicythere tenuis* (REUSS, 1850)  
 Plate II, fig. 1

1850 *Cytherina tenuis* n. sp. REUSS, p. 53, pl. 8, fig. 14.  
 2008 *Amnicythere tenuis* (REUSS) – TÓTH, p. 109–110, pl. 2, figs 1–3, 5. (cum syn.)  
 2013 *Amnicythere tenuis* (REUSS) – TER BORGH et al., fig. 6/14–15.  
 2014 *Amnicythere tenuis* (REUSS) – TER BORGH et al., fig. 8/27–28.  
 2015 *Amnicythere tenuis* (REUSS) – SILYE, pl. 10, figs 1–3.  
 2018 *Amnicythere tenuis* (REUSS) – HARZHAUSER et al., fig. 7/3.

*Dimensions:* L= 510–550 µm, H= 250–290 µm, L/H= 1.96–2.3.

*Stratigraphic range and geographic distribution:* Sarmatian: Vienna Basin, Austria (CERNAJSEK 1974, HARZHAUSER et al. 2018); Carpathian Foredeep, Poland (SZCZUCHURA 2000); Zsámbék Basin and Budapest, Hungary (TÓTH 2004, 2008); Lower Sarmatian: East-Slovakian Basin, Slovakia (ZLINSKÁ & FORDINÁL 1995); Transylvanian Basin, Romania (OLTEANU 1998, SILYE 2015); Pannonian and Dacian basins, Serbia and Romania (TER BORGH et al. 2013, 2014); Bessarabian: Moesian Plate, Bulgaria (STANCHEVA 1963, 1990); Pannonian: Pannonian Basin, Hungary (MÉHES 1908); Pontian: Dacian Basin, Romania (HANGANU 1974).

Genus *Callistocythere* RUGGIERI, 1953

*Callistocythere canaliculata* (REUSS, 1850)  
 Plate I, fig. 1

1850 *Cypridina canaliculata* n. sp. REUSS, p. 76, pl. 9, fig. 12.  
 2006 *Callistocythere canaliculata* (REUSS) – GROSS & PILLER, pp. 25–26, pl. 8, figs 1–4, 8–9, pl. 10, figs 1–2. (cum syn.)  
 2011 *Callistocythere* aff. *canaliculata* (REUSS) – HAJEK-TADESSE & PRTOLOJAN, figs 4, 9.  
 2019 *Callistocythere canaliculata* (REUSS) – BRINKMANN et al., fig. 4/P.

*Dimensions:* L= 570–600 µm, H= 260–300 µm, L/H= 2–2.19.

*Stratigraphic range and geographic distribution:* Ottangian: North Alpine Foreland Basin, Germany (BRINKMANN et al. 2019); Karpatian: Molasse Basin, Austria (ZORN 2003, 2004); Badenian: Vienna Basin and Danube Basin, Slovakia (BRESTENSKÁ & JIŘIČEK 1978, GROSS & PILLER 2006); Transylvanian Basin, Romania (OLTEANU 1998); Carpathian Foredeep, Poland (PARUCH-KULCZYCKA 1992; PARUCH-KULCZYCKA & SZCZUCHURA 1996, AIELLO & SZCZUCHURA 2004); Sarmatian: Tokaj Mts, Hungary (PIETRZENIUK 1973); North-Croatian Basin, Croatia (HAJEK-TADESSE & PRTOLOJAN 2011).

Genus *Euxinocythere* STANCHEVA, 1968

*Euxinocythere (Euxinocythere) naca* (MÉHES, 1908)  
 Plate II, fig. 7

1908 *Cythere naca* n. sp. MÉHES, p. 548–549, pl. 10, figs 8–12.  
 1989 *Leptocythere naca* (MÉHES) – SOKAČ, p. 687, pl. 8, fig. 10.  
 1989 *Leptocythere (Amnicythere) naca* (MÉHES) – OLTEANU, pl. 8, fig. 6.  
 1989 *Euxinocythere (Euxinocythere)* cf. *naca* (MÉHES) – KRSTIĆ & STANCHEVA, p. 778, pl. 11, fig. 3.  
 2008 *Euxinocythere (Euxinocythere) naca* (MÉHES) – TÓTH, pp. 112–113, pl. 1, fig. 7. (cum syn.)  
 2009 *Euxinocythere (Euxinocythere) naca* (MÉHES) – TÓTH, p. 84, pl. 3, fig. 3.  
 2011 *Leptocythere (Euxinocythere) naca* (MÉHES) – OLTEANU, pl. 19, fig. 1.  
 2013 *Euxinocythere naca* (MÉHES) – TER BORGH et al., fig. 8/10.

*Dimensions:* L= 470–510 µm, H= 235–260 µm, L/H= 1.88–1.95.

*Stratigraphic range and geographic distribution:* Sarmatian: Vienna and Danube basins, Austria and Slovakia (CERNAJSEK 1974, ZELENKA 1990); Moldavian Plateau, Romania (IONESI & CHINTĂUAN 1975, 1985); Carpathian Foredeep, Poland (SZCZUCHURA 2000); Volhynian: Moesian Plate, Northern Bulgaria (STANCHEVA 1990); Zsámbék Basin, Hungary (TÓTH 2008, 2009); Pannonian-Pontian: Pannonian Basin, Hungary and Serbia (KRSTIĆ 1973, MÉHES 1908, SZÉLES 1982, KRSTIĆ & STANCHEVA 1989; TER BORGH et al. 2013); North-Croatian Basin, Croatia (SOKAČ 1967, 1972, 1989); Transylvanian Basin, Romania (OLTEANU 2011); Pontian: Dacian Basin, Romania (OLTEANU 1989); South Caspian Basin, Azerbaijan (AGALAROVA 1967).

*Euxinocythere (Euxinocythere) praebosqueti* (SUZIN, 1956)  
 Plate II, figs 4–6

1956 *Leptocythere praebosqueti* n. sp. SUZIN, p. 83, pl. 3, figs 2–4.  
 2008 *Euxinocythere (Euxinocythere) praebosqueti* (SUZIN) – TÓTH, p. 114, pl. 3, figs 2–5. (cum syn.)  
 2013 *Euxinocythere (Euxinocythere) praebosqueti* (SUZIN) – VAN BAAK et al., fig. 4/13.

*Dimensions:* L= 490–510 µm, H= 200–260 µm, L/H= 1.9–2.1.

*Stratigraphic range and geographic distribution:* Sarmatian: Moesian Plate, Northern Bulgaria (STANCHEVA 1972, 1990); Upper Sarmatian: Zsámbék Basin, Hungary (TÓTH 2008); Bessarabian: Caucasus, Russia (SUZIN 1956); Plio-Pleistocene: South Caspian Basin, Azerbaijan (VAN BAAK et al. 2013).

Family Loxoconchidae Sars, 1925

Subfamily Loxoconchinae Sars, 1925

Genus *Loxoconcha* Sars, 1866

*Loxoconcha kochi* MÉHES, 1908

Plate II, figs 14–15

- 1908 *Loxoconcha kochi* n. sp. MÉHES, pp. 543–544, pl. 9, figs 5–9.  
 2005 *Loxoconcha kochi* MÉHES – FILIPESCU et al., pl. 3, fig. 6.  
 2006 *Loxoconcha kochi?* MÉHES – GROSS & PILLER, pp. 112–113, pl. 40, figs 1–7,9.  
 2008 *Loxoconcha kochi* MÉHES – TÓTH, p. 124, pl. 9, fig. 6. (cum syn.)  
 2013 *Loxoconcha kochi* MÉHES – TER BORGH et al., fig. 8/24–25.  
 2014 *Loxoconcha kochi* MÉHES – TER BORGH et al., fig. 7/23.  
 2014 *Loxoconcha kochi* MÉHES – FILIPESCU et al., fig. 8/15.  
 2018 *Loxoconcha kochi* MÉHES – HARZHAUSER et al., fig. 7/12.

*Dimensions:* L= 640–835 µm, H= 400–520 µm, L/H= 1.6–1.75.

*Stratigraphic range and geographic distribution:* Upper Badenian: Vienna Basin, Austria (GROSS & PILLER 2006); Dacian Basin, Romania (TER BORGH et al. 2014); Sarmatian: Vienna Basin, Austria (CERNAJSEK 1974, GROSS & PILLER 2006, HARZHAUSER et al. 2018); easternmost Pannonian and Transylvanian basins, Black Sea Depression, Romania (IONESI & CHINTĂUAN 1985; FILIPESCU et al. 2005, 2014); Upper Sarmatian: Zsámbék Basin, Hungary (TÓTH 2008); Pannonian Basin, Serbia (TER BORGH et al. 2013); Lower Pannonian (?): Pannonian Basin, Hungary (MÉHES 1908); Messinian and Pliocene (?): Rhône Valley, France (CARBONNEL 1978).

*Loxoconcha laeta* STANCHEVA, 1963

Plate II, fig. 16

- 1963 *Loxoconcha laeta* n.sp. STANCHEVA, pp. 34–35, pl. 6, fig. 9.  
 1990 *Loxoconcha laeta* STANCHEVA – STANCHEVA, pp. 88–89, pl. 31, figs 5–6.  
 2009 *Loxoconcha laeta* STANCHEVA – TÓTH, pp. 91–92, pl. 7, fig. 12.

*Dimensions:* L= 720–750 µm, H= 390–410 µm, L/H= 1.8–1.83.

*Stratigraphic range and geographic distribution:* Lower Sarmatian: Moesian Plate, Bulgaria (STANCHEVA 1963, 1990); Upper Sarmatian: Zsámbék Basin, Hungary (TÓTH 2009).

*Loxoconcha porosa* MÉHES, 1908

Plate II, fig. 17

- 1908 *Loxoconcha porosa* n. sp. MÉHES, pp. 542–543, pl. 8, figs 10–14.  
 2008 *Loxoconcha porosa* MÉHES – TÓTH, pp. 124–125, pl. 9, figs 3–5. (cum syn.)

*Dimensions:* L= 620–700 µm, H= 420–470 µm, L/H= 1.45–1.55.

*Stratigraphic range and geographic distribution:* Sarmatian: Pannonian Basin, Serbia (KRSTIĆ 1972); Black Sea Depression, Romania (IONESI & CHINTĂUAN 1985); Upper Sarmatian: Vienna Basin, Slovakia (ZELENKA 1990); Zsámbék Basin, Hungary (TÓTH 2008); Pannonian: Pannonian Basin, Hungary and Croatia (MÉHES 1908, SOKAČ 1972).

*Loxoconcha punctatella* (REUSS, 1850)

Plate I, fig. 5

- 1850 *Cypridina punctatella* n. sp. REUSS, pp. 65–66, pl. 9, fig. 15 a–b.  
 1978 *Loxoconcha punctatella* (REUSS) – BRESTENSKÁ & JIŘIČEK, pl. 2, figs 12–13.  
 1985 *Loxoconcha punctatella* (REUSS) – ZELENKA, pl. 3, figs 10–11.  
 2004 *Loxoconcha* ex. gr. *punctatella* (REUSS) – TÓTH, pp. 140–141, pl. 6, figs 1–2.  
 2006 *Loxoconcha punctatella* (REUSS) – GROSS & PILLER, pp. 73–74, pl. 40, figs 8,11, pl. 41, figs 1–10. (cum syn.)  
 2006 *Loxocorniculum* cf. *punctatella* (REUSS) – SZCZECZURA, fig. 10/3.  
 2008 *Loxoconcha* ex. gr. *punctatella* (REUSS) – TÓTH, p. 125, pl. 10, figs 1–2.  
 2011 *Loxoconcha punctatella* (REUSS) – HAJEK-TADESSE & PRTOJAN, fig. 4/16.  
 2012 *Loxoconcha punctatella* (REUSS) – SEKO et al., fig. 8/D.  
 2013 *Loxoconcha punctatella* (REUSS) – TER BORGH et al., fig. 6/28.  
 2019 *Loxoconcha punctatella* (REUSS) – BRINKMANN et al., fig. 8/N–O.

*Dimensions:* L= 540–670 µm, H= 400–450 µm, L/H= 1.4–1.54.

*Stratigraphic range and geographic distribution:* Burdigalian: Molasse Basin, Austria (BRINKMANN et al. 2019); Karpatian: Molasse Basin, Austria (ZORN 1998); Badenian: Danube Basin and Vienna Basin, Slovakia (BRESTENSKÁ & JIŘIČEK 1978, ZELENKA 1985); Molasse Basin, Austria (ZORN 2004); Carpathian Foredeep, Czech Republic and Poland (PARUCH-KULCZYCKA 1992, SZCZECZURA 2006, SEKO et al. 2012); North-Croatian Basin, Croatia (HAJEK-TADESSE & PRTOJAN 2011); Badenian to Sarmatian: Vienna Basin, Austria (GROSS & PILLER 2006); Lower Sarmatian: Zsámbék Basin, Hungary (TÓTH 2004, 2008); Pannonian Basin, Serbia (TER BORGH et al. 2013).

Genus *Loxocorniculum* BENSON & COLEMAN, 1963

*Loxocorniculum hastatum* (REUSS, 1850)

Plate II, figs 19–20

- 1850 *Cytherina hastata* REUSS sensu CERNAJSEK – REUSS, pl. 9, fig. 26.  
 2008 *Loxocorniculum hastatum* (REUSS) – TÓTH, pp. 125–126, pl. 9, figs 1–2. (cum syn.)  
 2012 *Loxocorniculum hastatum* (REUSS) – SEKO et al., fig. 8/F.  
 2014 *Loxocorniculum hastatum* (REUSS) – TER BORGH et al., fig. 7/22.  
 2017 *Loxocorniculum hastatum* (REUSS) – DUMITRIU et al., fig. 13/I–J.  
 2019 *Loxocorniculum hastatum* (REUSS) – BRINKMANN et al., p. 84, fig. 8/M.

*Dimensions:* L= 620–630 µm, H= 390–410 µm, L/H= 1.5–1.6.

*Stratigraphic range and geographic distribution:* Oligocene to Miocene (Aquitainian, Burdigalian, Langhian): Aquitaine Basin, France (DUCASSE et al. 1991, BEKAERT et al. 1991, DUCASSE & CAHUZAC 1996); Burdigalian: Molasse Basin, Austria (BRINKMANN et al. 2019); Rhône Basin, France (CARBONNEL 1969); Eggenburgian: Molasse Basin, Austria (KOLLMANN 1971); Karpatian: Vienna Basin, Czech Republic (KHEIL 1967); Molasse Basin, Austria (ZORN 1998, 2003, 2004); Badenian: Molasse Basin, Austria (ZORN 1998, 2004); Carpathian Foredeep, Poland and Czech Republic (PARUCH-KULCZYCKA 1992, SZCZECURA 2006, SEKO et al. 2012); Vienna Basin, Austria and Czech Republic (CERNAJSEK 1974, BRESTENSKÁ & JIŘÍČEK 1978, JANZ & VENNEMANN 2005, ZELENKA 1985); Moesian Platform, Bulgaria (STANCHEVA 1962); Dacian Basin, Romania (TER BORGH et al. 2014); Carpathian Foredeep, Poland (AIELLO & SZCZECURA 2004); Sarmatian: Mecsek Mts and Zsámbék Basin, Hungary (SZUROMI-KORECZ & SZEGŐ 2001, TÓTH 2008); Carpathian Foredeep, Poland (DUMITRIU et al. 2017).

Family Xestoleberididae SARS, 1928

Genus *Xestoleberis* SARS, 1866

*Xestoleberis dispar* MUELLER, 1894

Plate I, fig. 8

- 1894 *Xestoleberis dispar* n. sp. MÜLLER, p. 334, pl. 25, figs 2, 3, 9, 35.  
 1982 *Xestoleberis dispar* MÜLLER – FARANDA et al., pl. 2, figs 16–17.  
 1986 *Xestoleberis* sp. – MOSTAFAWI, pl. 3, fig. 33.  
 2006 *Xestoleberis* aff. *dispar* MÜLLER – GROSS & PILLER, pp. 137–138, pl. 2, fig. 4.  
 2008 *Xestoleberis dispar* MÜLLER – KOEHN-ZANINETTI & TÉTARD, fig. 4/10.  
 2014 *Xestoleberis dispar* (MUELLER) – TER BORGH et al., fig. 7/26–27.  
 2015 *Xestoleberis dispar* MUELLER – SCIUTO et al., pl. 2, fig. 6.  
 2016 *Xestoleberis dispar* MUELLER – PARLAK & NAZIK, pl. 3, fig. 14.  
 2017 *Xestoleberis fuscata* SCHNEIDER – DUMITRIU et al., fig. 13/H.

*Dimensions:* L= 660–665 µm, H= 350–370 µm, L/H= 1.80–1.88.

*Stratigraphic range and geographic distribution:* Badenian: Dacian Basin, Romania (TER BORGH et al. 2014); upper Badenian to lower Sarmatian: Vienna Basin, Austria (GROSS & PILLER 2006); lower Sarmatian: Carpathian Foredeep, Poland (DUMITRIU et al. 2017); Tortonian, Pleistocene: Mediterranean Sea, Greece (FARANDA et al. 2008, MOSTAFAWI 1986); Recently widely distributed in the Mediterranean Sea.

*Xestoleberis tumida* (REUSS, 1850)

Plate I, fig. 7

- 1850 *Cytherina tumida* n. sp. REUSS, pp. 57–58, pl. 8, fig. 29.  
 2006 *Xestoleberis tumida* (REUSS) – GROSS & PILLER, pp. 134–137, pl. 48, figs 1–10, pl. 49, figs 1–5, pl. 51, fig. 7. (cum syn.)  
 2006 *Xestoleberis* cf. *tumida* (REUSS) – SZCZECURA, fig. 10/2,4.

*Dimensions:* L= 510–540 µm, H= 320–330 µm, L/H= 1.6–1.8.

*Stratigraphic range and geographic distribution:* Karpatian: Molasse Basin, Austria (ZORN 1998); Badenian: Carpathian Foredeep, Poland (SZCZECURA 2006); Austria (ZORN 1998; GROSS & PILLER 2006).

Suborder Cypridocopina BAIRD, 1845

Superfamily Cypridoidea BAIRD, 1845

Family Cyprididae BAIRD, 1845

Subfamily Cyprinotinae BRONSHTEIN, 1947

Genus *Heterocypris* CLAUS, 1892

*Heterocypris salina* (BRADY, 1868)

Plate II, fig. 26

1868 *Cypris salina* n. sp. BRADY, p. 368; pl. 28, figs 8–13.

1980 *Heterocypris salina salina* (BRADY) – FREELS, p. 28, pl. 3, figs 1–6. cum syn.

2000 *Heterocypris salina* (BRADY) – MEISCH, pp. 349–352, fig. 135.

2003 *Heterocypris salina* (BRADY) – MISCHKE et al., fig. 1/7.

2004 *Heterocypris salina* (BRADY) – PÍPIK, p. 227, pl. 1, figs 6–7.

2005 *Heterocypris salina* (BRADY) – MATZKE-KARASZ, p. 126, pl. 3, fig. 4.

2005 *Heterocypris salina* (BRADY) – SCHARF et al., pl. 2, figs 17–20.

2008 *Heterocypris salina* (BRADY) – NAZIK et al., pl. 1, fig. 15.

2008 *Heterocypris salina* (BRADY) – POQUET et al., fig. 6/I.

2012 *Heterocypris salina* (BRADY) – MISCHKE et al., pl. 1, figs 7–10, 18.

2014 *Heterocypris salina* (BRADY) – SCHARF & MEISCH, fig. 3/I–K.

2014 *Heterocypris salina* (BRADY) – MISCHKE et al., fig. 7/2.

2016 *Heterocypris salina* (BRADY) – SALEL et al., pl. 4, figs 4–6.

2019 *Heterocypris salina* (BRADY) – TUNCER et al., pl. 1, figs 1–3.

*Dimensions:* L= 945–955 µm, H= 565–590 µm, L/H= 1.61–1.67.

*Stratigraphic range and geographic distribution:* Widely distributed in upper Miocene to Holocene freshwater to saline habitats (riverine pools and lakes) in Europe (MEISCH 2000) and recently over the world.

Suborder Darwinulocopina BRADY & NORMAN, 1889

Superfamily Darwinuloidea BRADY & NORMAN, 1889

Family Darwinulidae BRADY & NORMAN, 1889

Genus *Darwinula* BRADY & NORMAN, 1889

*Darwinula stevensoni* (BRADY & ROBERTSON, 1870)

Plate II, fig. 21

1870 *Polycheles stevensoni* m. BRADY & ROBERTSON, pp. 25–26, pl. 7, figs 1–7, pl. 10, figs 4–14.

2000 *Darwinula stevensoni* (BRADY & ROBERTSON) – MEISCH, p. 49, fig. 16/A–E.

2004 *Darwinula stevensoni* (BRADY & ROBERTSON) – PÍPIK et al., pl. 1, fig. 10.

2005 *Darwinula stevensoni* (BRADY & ROBERTSON) – CABRAL et al., pp. 53–55, pl. 1, figs 1–6. (cum syn.)

2012 *Darwinula stevensoni* (BRADY & ROBERTSON) – FUHRMANN, pl. 1, figs 1 a–f.

*Dimensions:* L= 670–680 µm, H= 420–425 µm, L/H= 1.59–1.6.

*Stratigraphic range and geographic distribution:* Wide-



ly distributed in Oligocene to Holocene lacustrine environments in Europe (MEISCH 2000) and recently over the world.

Genus *Vestalenula* ROSSETTI & MARTENS, 1998

*Vestalenula pagliolii* (PINTO & KOTZIAN, 1961)

Plate II, figs 22–23; Plate III, fig. 10 (thin-section)

1961 *Darwinula pagliolii* n. sp. PINTO & KOTZIAN, p. 27, pl. 1, figs 1–5, pl. 3, figs 1–4, pl. 5, figs 1–9, pl. 6, figs 1–9, pl. 9, figs 1–9.

2003 *Vestalenula pagliolii* (PINTO & KOTZIAN) – PIPÍK & BODERGAT, p. 348, pl. 1, figs 5–10, fig. 24. (cum syn.)

2004 *Vestalenula pagliolii* (PINTO & KOTZIAN) – PIPÍK et al., pl. 1, fig. 11.

2005 *Vestalenula pagliolii* (PINTO & KOTZIAN) – CABRAL et al., pp. 59–60, pl. 3, figs 5–16.

*Dimensions:* L= 455–470  $\mu\text{m}$ , H= 210–220  $\mu\text{m}$ , L/H= 2.16–2.18.

*Stratigraphic range and geographic distribution:* Widely distributed in Oligocene to Holocene freshwater to oligohaline habitats (riverine pools and lakes) in Europe (MEISCH 2000) and recently in Brazil (MARTENS et al. 1997).