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Surface electromyography to quantify neuro-respiratory drive and neuro-mechanical coupling in mechanically ventilated children

Alette A. Koopman^{1*}, Jefta van Dijk¹, Eline Oppersma², Robert G. T. Blokpoel¹ and Martin C. J. Kneyber^{1,3}

Abstract

Background The patient's neuro-respiratory drive, measured as electrical activity of the diaphragm (EAdi), quantifies the mechanical load on the respiratory muscles. It correlates with respiratory effort but requires a dedicated esophageal catheter. Transcutaneous (surface) monitoring of respiratory muscle electromyographic (sEMG) signals may be considered a suitable alternative to EAdi because of its non-invasive character, with the additional benefit that it allows for simultaneously monitoring of other respiratory muscles. We therefore sought to study the neuro-respiratory drive and timing of inspiratory muscles using sEMG in a cohort of children enrolled in a pediatric ventilation liberation trial. The neuro-mechanical coupling, relating the pressure generated by the inspiratory muscles to the sEMG signals of these muscles, was also calculated.

Methods This is a secondary analysis of data from a randomized cross-over trial in ventilated patients aged < 5 years. sEMG recordings of the diaphragm and parasternal intercostal muscles (ICM), esophageal pressure tracings and ventilator scalars were simultaneously recorded during continuous spontaneous ventilation and pressure controlled-intermittent mandatory ventilation, and at three levels of pressure support. Neuro-respiratory drive, timing of diaphragm and ICM relative to the mechanical ventilator's inspiration and neuro-mechanical coupling were quantified.

Results Twenty-nine patients were included (median age: 5.9 months). In response to decreasing pressure support, both amplitude of sEMG (diaphragm: p = 0.001 and ICM: p = 0.002) and neuro-mechanical efficiency indices increased (diaphragm: p = 0.05 and ICM: p < 0.001). Poor correlations between neuro-respiratory drive and respiratory effort were found, with R²: 0.088 [0.021–0.152].

Conclusions sEMG allows for the quantification of the electrical activity of the diaphragm and ICM in mechanically ventilated children. Both neuro-respiratory drive and neuro-mechanical efficiency increased in response to lower inspiratory assistance. There was poor correlation between neuro-respiratory drive and respiratory effort.

Trial registration Clinical Trials.gov ID NCT05254691. Registered 24 February 2022, registered retrospectively.

Keywords Children, Mechanical ventilation, Neuro-respiratory drive, Neuro-mechanical coupling, Surface electromyography, Pressure support

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Background

Acute respiratory failure necessitating mechanical ventilation (MV) is one of the main indications for pediatric intensive care unit (PICU) admissions. Although the technique of MV itself is lifesaving for patients with impaired gas exchange, it can also cause lung and diaphragm injury by a variety of interacting mechanisms [1, 2]. Whereas lung injury mainly comes from mechanical stress and strain caused by the ventilator (ventilatorinduced lung injury), there is also mounting evidence that spontaneous breathing during especially moderate-to-severe disease plays a role in the progression of lung injury, a paradigm known as patient self-inflected lung injury (P-SILI). Vigorous spontaneous breathing causes increased transpulmonary pressure swings, thereby amplifying lung damage [3-7]. It is commonly accepted to allow for spontaneous breathing in mechanically ventilated patients to protect the respiratory muscles. However, this must be balanced against the risk for P-SILI. Ventilator-induced diaphragmatic dysfunction refers to diaphragm atrophy and injury, occurring mainly through overassistance myotrauma (i.e., excessive unloading of the respiratory muscles), leading to a reduction or even suppression of the neuro-respiratory drive [8, 9]. Oppositely, insufficient ventilatory support failing to adequately unload respiratory muscles (i.e., excessive respiratory muscle loading), may cause inflammation and injury leading to underassistance myotrauma [8]. In critically ill adults, both types of myotrauma have been linked to prolonged duration of MV, prolonged ICU admission and a higher risk of complications including reintubation and need for tracheostomy [10].

Individualized monitoring of respiratory effort is necessary for the delivery of lung and diaphragm protective ventilation. Esophageal pressure (Pes) manometry is considered the gold standard for measuring patient inspiratory effort and transpulmonary driving pressure [11, 12]. However, accurate Pes monitoring is invasive, requires correct catheter positioning and an individualized balloon filling volume making it a cumbersome monitoring tool [11]. Monitoring the electrical activity of the diaphragm (EAdi) using a specifically designed esophageal catheter, represents the patient's neuro-respiratory drive and correlates strongly with the transpulmonary driving pressure [13–15]. However, monitoring EAdi is limited to only one ventilator brand in clinical practice. The invasive nature of esophageal pressure manometry and EAdi monitoring may be perceived as challenging especially in small children. We propose that transcutaneous (surface) monitoring of respiratory muscle electromyographic (sEMG) signals may be considered a good alternative to EAdi because of it is non-invasive design, plus that it also allows for simultaneously monitoring of other respiratory muscles. We therefore sought to study the neuro-respiratory drive, timing of inspiratory muscles and neuro-mechanical coupling using sEMG in a cohort of children enrolled in a pediatric ventilation liberation trial.

Materials and methods

Study population

This study is a secondary analysis of data from mechanically ventilated children enrolled in a randomized crossover trial investigating pediatric ventilation liberation. The study was performed in the 20-bed tertiary medical-surgical PICU of the Beatrix Children's Hospital, University Medical Center Groningen (Groningen, the Netherlands). Subjects < 5 years of age deemed ready for weaning from invasive MV by their treating clinician were eligible for inclusion. Those with congenital or acquired neuromuscular disorders or paralysis of the diaphragm, severe traumatic brain injury (i.e., Glasgow Coma Score < 8), uncorrected congenital heart disorder, chronic lung disease and severe pulmonary hypertension were excluded. The Institutional Review Board approved the study (NL38361.042.11), and written informed consent was obtained from the parents or legal caretakers.

Study protocol

In the parent study, the effects of two different ventilator modes and down tapering the level of pressure support (PS) were studied [16]. Briefly, subjects were randomly assigned to first a 10 min period in a continuous spontaneous ventilation (CSV) mode (i.e., PS ventilation) or pressure controlled-intermittent mandatory ventilation (PC-IMV) (i.e., pressure controlled assist/control [PC/AC]) with a low mandatory breath rate (25% of the set rate prior to study enrollment). Subsequently, subjects were crossed over to the other randomization arm. Afterwards, subjects were switched to PS ventilation. Then the PS level was decreased on three consecutive steps by 2 cm H₂O. Positive end-expiratory pressure (PEEP) and fraction of inspired oxygen (FiO₂) remained constant throughout the study.

Data acquisition

sEMG recordings, Pes manometry and ventilator scalars were acquired at baseline and subsequently during each intervention according to the randomization outcome [16]. Data was recorded for 5 min during stable breathing.

Electrical activity of the diaphragm and parasternal intercostal muscles (ICM) was measured transcutaneous using pairs of single Ag/AgCl electrodes (EasyTrode TM Pre gelled Electrodes, Multi Bio Sensors Inc, El Paso, USA). Two electrodes were placed bilaterally at the costoabdominal margin in the nipple line, and an electrode

at both the left and right second intercostal space. A common electrode was placed at the sternal level [17]. sEMG was recorded using the Porti-16 data acquisition system (22 bits, TMSi; The Netherlands) with unipolar electrophysiological channels (71.5 nV/bit, gain: 20). An age-appropriate esophageal balloon catheter (Avea SmartCath 6 or 8 Fr, Vyaire, Mettawa, III, USA) was positioned in the lower 1/3 of the esophagus and connected to the Bicore II (Vyaire, Mettawa, III, USA). Correct position was verified by negative pressure deflections during spontaneous breathing and/or chest radiography that was done for other indications [18]. Esophageal balloon volume was titrated up to a maximum of 1.6 ml (pediatric catheter) or 2.6 ml (adult catheter). Optimal balloon volume was achieved by determining the volume with the Pes maximum amplitude.

A proximal flow sensor (VarFlex[™], Vyaire, Mettawa, III, USA) was used to measure flow and Vt near the Y-piece of the endotracheal tube. Ventilator scalars were acquired using the ventilator's analog output port. All signals were digitized with a sample frequency of 1024 Hz and stored offline (Polybench, Applied Biosignals GmbH, Weener, Germany).

Patient characteristics (gender, age, weight, 24-h Pediatric RISK of Mortality (PRISM) III score, admission diagnosis and endotracheal tube size) were obtained to characterize the study population [19].

Offline signal processing and parameter calculation

The recorded sEMG signals of both diaphragm and ICM were processed as described previously [20].

We visually selected a period of 30 consecutive breaths free of artifacts from each series of measurements. Onset, peak, and termination of inspiratory muscle activity were determined in sEMG signals as reported by us previously [20]. The neuro-respiratory drive was quantified by the following breath-by-breath parameters, normalized to muscle activity at baseline measurement: maximal electrical activity (peak activity, EMG_{peak}), the amplitude (the difference between the peak and tonic activity, EMG_{amp}), the integral of EMG signal over time during the inspiration multiplied with respiratory rate (area under curve, EMG_{AUC}/min), and mean electrical activity during a whole breath (mean activity, EMG_{mean}). To evaluate the timing of inspiratory muscles relative to the MV, trigger and cycle times were determined as described previously [20]. The tidal esophageal peak-to-through during inspiration ($\mathrm{Pes}_{\mathrm{amp}})$ was calculated. The esophageal pressure time product (PTP) was calculated by the integral of Pes over time during inspiration multiplied by respiratory rate. Expiratory tidal volume, respiratory rate and inspiratory time were calculated from the proximal flow signal. The neuro-mechanical coupling, i.e., the relationship between Pes and sEMG was analyzed by calculating indices for neuro-mechanical efficiency (NME) (i.e., Pes_{amp}/EMG_{amp} , and $PTP/EMG_{AUC}/min$).

Statistical analysis

Statistical analyses were performed using Prism 5 (Graphpad software, San Diego, CA, USA) and Matlab R2018a (Mathworks, Natick, MA, USA). The Shapiro-Wilk test was used to test data for normality. Descriptive data were expressed as median [interquartile range (IQR)] or percentage (%) of total. The neuro-respiratory drive, neuro-mechanical coupling, and timing of the inspiratory muscles at each measurement series were compared using the Wilcoxon signed rank test. To assess if the changes in PS level induced significant changes in neuro-respiratory drive, neuro-mechanical coupling, and timing of the inspiratory muscles, one-way analysis of variance for repeated measures was performed (Skillings-Mack test). The relationship between neuro-respiratory drive and Pes-derived data (effort) in each ventilatory condition was assessed. For both inspiratory muscles, the correlation between EMG_{amp} and Pes_{amp}, and between EMG_{AUC}/min and PTP were determined using the determination coefficient R^2 . A p value < 0.05 was considered statistically significant.

Results

Thirty-six subjects were enrolled in the parent study. sEMG recordings from seven subjects were excluded from analysis because of missing data (N=1) and inability to detect inspiratory sEMG activity in both diaphragm and ICM due to crosstalk—i.e. electrical muscle activity of adjacent muscles (N=4) or due to electrical interference (N=2). Thus, 29 subjects (19 boys and 10 girls) with a median age of 5.9 [1.3–14.1] months and weight 6.8 [4.9–10.0] kg were included (Table 1). The majority of the patients was admitted for primary respiratory failure (86.2%).

Neuro-respiratory drive

Figure 1A displays the neuro-respiratory drive (i.e., ${\rm EMG_{peak}}$, ${\rm EMG_{amp}}$, ${\rm EMG_{mean}}$ and ${\rm EMG_{auc}}/{\rm min}$) for the whole cohort and stratified by ventilator mode (i.e., PS ventilation or PC-IMV). For the whole cohort, the neuro-respiratory drive to the diaphragm was estimated by a median ${\rm EMG_{peak}}$ of 95.5% and 91.3%, median ${\rm EMG_{ampl}}$ of 97.4% and 98.5%, median ${\rm EMG_{mean}}$ of 92.3% and 92.9%, and median ${\rm EMG_{AUC}}/{\rm min}$ 92.0% and 89.3% during PS ventilation and PC-IMV, respectively. The neuro-respiratory drive to the ICM was described by a median ${\rm EMG_{peak}}$ of 101.9% and 95.8%, median ${\rm EMG_{ampl}}$ of 106.8% and 109.1%, median ${\rm EMG_{mean}}$ of 99.5% and 95.0%, and median ${\rm EMG_{AUC}}/{\rm min}$ 97.9% and 96.0% during PS

Table 1 Patient characteristics

	Subjects	
	(n = 29)	
Age (months)	5.9 [1.3 to 14.1]	
Sex male/female	19/10	
Weight (kg)	6.8 [4.9 to 10.0]	
PRISM III score	3.0 [1.0 to 4.25]	
PIM II score	-4.38 [-4.66 to 3.92]	
Admission diagnosis (n)		
Respiratory	25	
Postoperative	2	
Upper airway obstruction	2	
Respiratory disease (n)		
Healthy lungs	4	
Obstructive disease	1	
Restrictive disease	4	
Obstructive + restrictive	20	
Duration of MV (days)	5.0 [3.3 to 6.9]	
HFOV (n)	11	
Duration of HFOV (d)	2.2 [1.4 to 2.6]	
Length of PICU stay (d)	7.0 [5.1 to 9.5]	
Extubation failure (n)	3	

PRISM III 24-h Pediatric Risk of Mortality III, PIM II Pediatric Index of Mortality II, MV mechanical ventilation, HFOV high frequency oscillation ventilation

Data are shown in number or median [interquartile range]

ventilation and PC-IMV, respectively. The neuro-respiratory drive estimates were comparable between the two ventilation modes. $\mathrm{EMG}_{\mathrm{amp}}$ changed with decreasing PS in both the diaphragm (median EMG_{amp} 87.0, 98.2, 105.9, and 107.4% of baseline for PS base, -2, -4, and -6 cm H_2O , respectively; p = 0.001) and ICM (median EMG_{amp} 105.1, 112.1, 112.6, and 137.6% of baseline for PS base, -2, -4, and -6 cm H_2O , respectively; p = 0.002) (Fig. 1B). Similar patterns were observed for diaphragmatic EMG_{peak} and EMG_{auc}/min (median EMG_{peak} 91.4, 92.6, 100.9, and 96.8% of baseline for PS base, -2, -4, and -6 cm H₂O, respectively; p = 0.02, and median EMG_{auc}/min 91.0, 95.1, 102.1, and 101.4% of baseline for PS base, -2, -4, and -6 cm H_2O , respectively; p = 0.03). The other neurorespiratory drive estimates remained constant at various PS levels (p > 0.05). Absolute values of neuro-respiratory drive estimates are shown in Table S1 of Additional file 1.

Timing of respiratory muscles

For the whole cohort, median diaphragmatic trigger time was 0.429 s during PS ventilation and 0.428 s during PC-IMV, and median ICM trigger time was 0.449 s during PS ventilation and 0.508 s during PC-IMV. Trigger times were not significantly different between PS ventilation and PC-IMV (p > 0.05). An increased cycle time during

PC-IMV compared to PS ventilation was found for both diaphragm (median trigger time 0.017 and 0.157 s during PS ventilation and PC-IMV, respectively; $p\!=\!0.04$) and ICM (median trigger time 0.066 and 0.205 s during PS ventilation and PC-IMV, respectively; $p\!=\!0.06$). Trigger and cycle times of both inspiratory muscles are graphically depicted in Fig. 2. We did not observe any changes in trigger or cycle times when PS was reduced (Table 2). There were no timing differences between diaphragm and ICM within one ventilation condition.

Neuro-mechanical coupling

Neuro-mechanical coupling of the diaphragm was estimated by a median Pes_{ampl}/EMG_{ampl} of 3.3 and 2.8 cmH₂O/ μ V, and median PTP/EMG_{AUC} × min⁻¹ 0.69 and 0.59 cmH₂O·s·min⁻¹/μV during PS ventilation and PC-IMV, respectively. Neuro-mechanical coupling of ICM was described by a median Pes_{ampl}/EMG_{ampl} of 5.4 and 4.9 cmH₂O/ μ V, and median PTP /EMG_{AUC} × min⁻¹ 0.70 and 0.81 cmH₂O·s·min⁻¹/μV during PS ventilation and PC-IMV, respectively. Neuro-mechanical coupling was comparable between PS ventilation and PC-IMV (Fig. 3). Estimates for neuro-mechanical coupling increased for diaphragm and ICM when PS was reduced, indicating that inspiratory muscles generated more pressure for 1 μ V of EMG, as shown in Fig. 3. Diaphragm neuro-mechanical coupling estimates were median Pes_{ampl}/EMG_{ampl} 3.1, 3.7, 4.8, and 5.2 cm $H_2O/$ μV for PS base, -2, -4, and -6 cm H₂O, respectively; p = 0.05; and median PTP /EMG_{AUC} × min⁻¹ 0.56, 0.63, 0.76, and 0.89 cm $H_2O \cdot s \cdot min^{-1}/\mu V$ for PS base, -2, -4, and -6 cm H_2O_2 , respectively; p=0.004. ICM neuromechanical coupling estimates were median Pes_{ampl}/ EMG_{ampl} 4.4, 6.3, 6.8, and 7.4 $cmH_2O/\mu V$ for PS base, -2, -4, and -6 cm H_2O , respectively; p < 0.001 and median PTP /EMG $_{AUC} \times min^{-1}$ 0.78, 0.96, 0.92, and 1.18 cm $H_2O\cdot s\cdot min^{-1}/\mu V$ for PS base, -2, -4, and -6 cm H_2O , respectively; p < 0.001. Diaphragmatic EMG_{amp} and EMG_{auc}/min were poorly correlated with Pes_{amp} $(R^2 < 0.15)$ and PTP $(R^2 < 0.26)$ (Fig. 4). For the parasternal intercostals, comparable weak relationships were found between EMG_{amp} and Pes_{amp} (R²<0.31), and EMG_{auc} /min and PTP ($R^2 < 0.06$) (Fig. 4).

Discussion

The main finding of this study is that it was possible to quantify the electrical activity of the diaphragm and parasternal intercostal muscles in mechanically ventilated children through transcutaneous recordings (sEMG). Breath-by-breath analysis showed a time-dependent relation between inspiratory sEMG and the ventilator pressurization reflected by a positive trigger time and almost neutral cycle time in PS ventilation. Furthermore, the

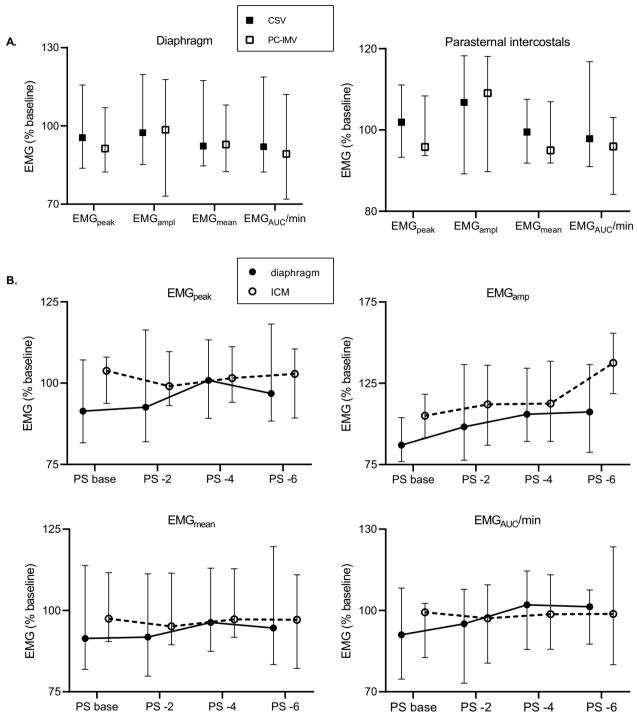


Fig. 1 Inspiratory muscle activity of diaphragm and parasternal intercostal muscle (ICM) during **A** two different weaning methods and **B** reduction of pressure support (PS) level. The electromyography (EMG) values are normalized to activity during baseline measurement. Values are depicted as median (interquartile range). The number of subjects in which EMG parameters could be determined differed per muscle and ventilation mode or PS level. Median number of subjects, for the diaphragm N = 29 and ICM N = 24. CSV continuous spontaneous ventilation, EMG_{peak} peak EMG activity, EMG_{amp} EMG activity amplitude, EMG_{mean} mean EMG activity level during one breath, EMG_{AUC}/min integral of EMG signal over time during the inspiration multiplied with the respiratory rate, PC-IMV pressure-controlled intermittent mandatory ventilation

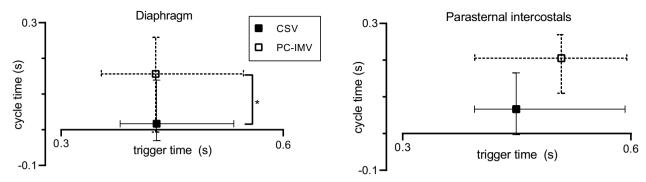


Fig. 2 Trigger and cycle times of diaphragm and parasternal intercostals in two weaning approaches. Values are depicted as median (interquartile range). *Significant difference between two weaning approaches (p < 0.05). CSV continuous spontaneous ventilation, PC-IMV pressure-controlled intermittent mandatory ventilation

Table 2 Trigger and cycle times of diaphragm and parasternal intercostal muscles at different pressure support levels

	PS base	PS-2	PS-4	PS-6
Diaphragm				
Trigger time (s)	0.42 [0.35 to 0.48]	0.43 [0.36 to 0.55]	0.38 [0.28 to 0.49]	0.41 [0.35 to 0.51]
Cycle time (s)	0.03 [-0.08 to 0.08]	0.06 [-0.01 to 0.12]	0.03 [-0.02 to 0.10]	0.03 [-0.04 to 0.14]
ICM				
Trigger time (s)	0.43 [0.38 to 0.56]	0.49 [0.39 to 0.52]	0.47 [0.37 to 0.55]	0.43 [0.38 to 0.50]
Cycle time (s)	0.08 [0.00 to 0.17]	0.06 [-0.01 to 0.13]	0.05 [0.02 to 0.15]	0.04 [0.00 to 0.12]

ICM parasternal intercostal muscles, *PS* pressure support level Data is shown as median [interquartile range]

electrical activity of both diaphragm and ICM increased in response to decreasing inspiratory level of assist on a cohort level.

Neuro-respiratory drive

In clinical practice, monitoring of the electrical activity of inspiratory muscles may facilitate in identifying patient-ventilator asynchrony [20–22], diaphragm paresis [23], and central hypoventilation and apneas [24], titrating ventilatory support [25, 26], and assessing the use of (accessory) inspiratory and expiratory muscles in spontaneously breathing children [26–30]. Although absolute [30, 31] as well as normalized sEMG [25–28] values have been published in previous investigations, normal sEMG values of inspiratory muscles are still lacking. Therefore, it is difficult to interpret the sEMG values observed in this study. Theoretically, sEMG values can be interpreted in relation to the maximal sEMG obtained during a voluntary maximal effort, equal to EAdi [32]. However, voluntary maximal effort cannot reliably be obtained in critically ill children. In this study, we therefore normalized the inspiratory muscle activity to activity at baseline measurement for the interpretation of the neuro-respiratory drive.

To our best knowledge, this is the first study examining several neuro-respiratory drive parameters of the diaphragm and ICM in MV pediatric patients. We found that ${\rm EMG_{amp}}$ of both diaphragm and ICM, and diaphragmatic ${\rm EMG_{peak}}$ and ${\rm EMG_{AUC}}/{\rm min}$ increased in response to decreasing pressure support. In contrast, MacBean and colleagues only assessed ${\rm EMG_{peak}}$ of ICM and reported also higher values during less ventilatory support [29]. Others reported that (pre-term) infants and children failing extubation had a higher diaphragmatic ${\rm EMG_{peak}}$ and tonic EMG, both pre and post extubation [30]. In ventilated adults, it was shown that ${\rm EMG_{peak}}$ and ${\rm EMG_{AUC}}$ of both diaphragm and extradiaphragmatic inspiratory muscles increased in response to lower inspiratory support levels [25, 26].

Based on our results, $\mathrm{EMG}_{\mathrm{amp}}$ is the most accurate neuro-respiratory drive parameter. For both inspiratory muscles, $\mathrm{EMG}_{\mathrm{amp}}$ responded to increasing respiratory load on a group level. When reducing the level of inspiratory support, first diaphragmatic $\mathrm{EMG}_{\mathrm{amp}}$ increased linearly whereas $\mathrm{EMG}_{\mathrm{amp}}$ of ICM remained constant. Subsequently, diaphragmatic $\mathrm{EMG}_{\mathrm{amp}}$ reached a plateau and $\mathrm{EMG}_{\mathrm{amp}}$ of ICM increased exponentially at the lowest level of support. Therefore, our data suggests that when the maximum diaphragm capacity is reached,

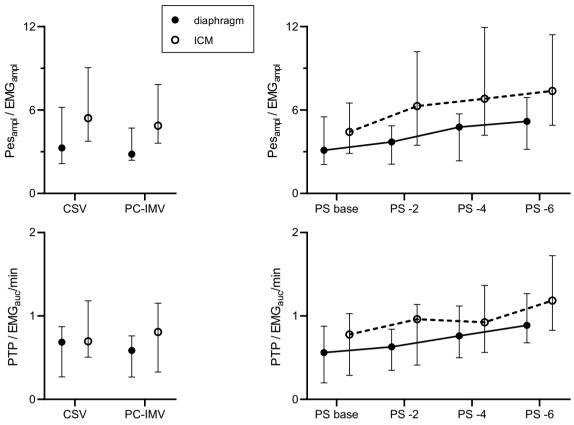


Fig. 3 Neuro-mechanical coupling (NMC) of diaphragm and parasternal intercostals (ICM) during two different weaning methods (*left*) and reduction of pressure support (PS) level (*right*). Values are depicted as median (interquartile range). The number of subjects in which EMG and Pes parameters could be determined differed per muscle and ventilation mode or PS level. For the diaphragm N = 20 (median) and ICM N = 18 (median). *CSV* continuous spontaneous ventilation, *PC-IMV* pressure-controlled intermittent mandatory ventilation, *Pes_{amp}/EMG_{amp}* esophageal pressure amplitude divided by EMG activity amplitude and PTP/*EMG_{AUC}/min* pressure time product of esophageal pressure multiplied with the respiratory rate divided by integral of EMG signal over time during the inspiration multiplied with the respiratory rate

ICM will be increasingly recruited in case of a further PS reduction.

The neural respiratory drive is best represented by measured electrical activity of the diaphragm [33]. Recruitment of the accessory muscles is a well-known clinical sign of an increase in respiratory load. In both healthy subjects and ventilated patients, there is a hierarchy with respect to respiratory muscle recruitment [25]. In case of an increase in respiratory load, the diaphragm is immediately activated, followed by the chest wall muscles and subsequently by expiratory muscles [27, 28]. In our pediatric patients, we observed a similar recruitment pattern of the diaphragm and ICM represented by EMG_{amp}.

Remarkably, in our study diaphragmatic EMG parameters showed values less than 100% of baseline. The higher levels of diaphragmatic electrical activity during baseline measurements might be explained by patients being agitated from instrumenting them for the data acquisition, since the diaphragm responds to an acute

increase of respiratory load. However, accessory inspiratory muscles including ICM are recruited by a prolonged increased work of breathing, explaining possibly that EMG levels of ICM were not lower than baseline levels.

Timing of respiratory muscles

In mechanically ventilated children, the patient ventilator interaction is often asynchronous [34–36]. We showed an increased cycle time during PC-IMV compared to CSV in both diaphragm and ICM, indicating a more asynchronous patient-ventilator interaction during PC-IMV. In PS ventilation, the timing of expiration is indirectly determined by the patient, i.e. the expiration trigger setting, instead of a set inspiration time as in PC-IMV, underlying a more synchronous interaction. In contrast to our finding, other research did not report a difference in cycle time between PC-IMV and CSV but we cannot easily explain this [36].

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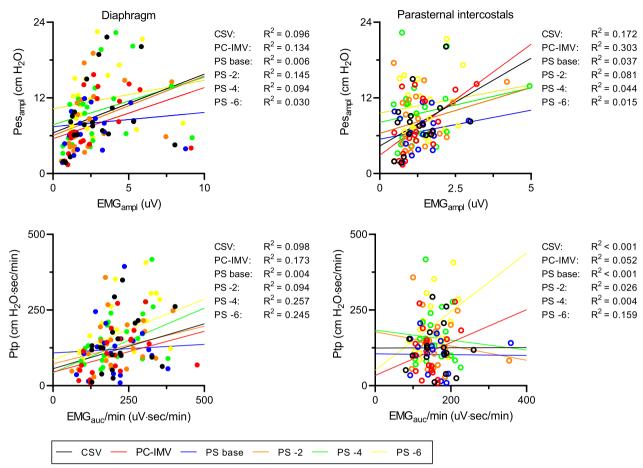


Fig. 4 Correlation for the global population between inspiratory muscle activity of diaphragm (left) and parasternal intercostals (right), and inspiratory work of breathing in each condition of ventilator assistance. On the right side of each graph, the respective determination coefficient R^2 is shown for each condition of ventilator assistance. CSV continuous spontaneous ventilation, EMG_{amp} EMG activity amplitude, EMG_{AUC}/min integral of EMG signal over time during the inspiration multiplied with the respiratory rate, PC-IMV pressure-controlled intermittent mandatory ventilation, Pes_{amp} esophageal pressure amplitude, PS pressure support, and PTP pressure time product of esophageal pressure multiplied with the respiratory rate

Neuro-mechanical coupling

We found that NME indices for both diaphragm and ICM were affected by the level of assistance. Such a relation has also been described by the level by Essouri et al. [13] in pediatric patients when comparing the NME during mechanical ventilation and post extubation, using EAdi instead of surface EMG. Of note, Mortamet et al. [37] did not find significant changes in $Pes_{ampl}/EAdi_{ampl}$ before, during and after a spontaneous breathing trial. However, they also reported no increase in neuro-respiratory drive and respiratory effort, indicating that patients did not develop respiratory distress or fatigue during their spontaneous breathing trial. The dependency of NME indices on the level of muscle loading might be caused by the recruitment of accessory muscles [26], the increased efficiency of the diaphragm and ICM together or a combination of both. In addition to the diaphragm and ICM, other accessory inspiratory and expiratory muscles should be recorded to model the interaction between neurorespiratory drive and respiratory effort accurately, in future research.

In each condition of ventilator assistance, a poor correlation was observed between neuro-respiratory drive and respiratory effort in the global population, suggesting muscle electrical activity is not synonymous with muscle contraction and force generation. In contrast, a strong linear correlation was previously described between EAdi and respiratory effort in different ventilator conditions in pediatric patients [13]. However, in that particular study measurement noise was reduced trough aggregation of similar breaths in their analysis. The NME index is highly variable between different patients but it is quite stable within a respiratory

stable patient, regardless of whether the electrical activity of the diaphragm is recorded transcutaneous or transesophageal [14, 31, 38]. However, in case of respiratory distress or muscle fatigue, this relationship can change as indicated by our data.

Limitations

Several limitations of our study need to be addressed. First, this study was designed a single center study potentially limiting the generalizability of our findings, although our unit is comparable to most large PICUs globally. Second, patients could be enrolled in our study when the attending physician deemed the patient eligible for weaning. Nevertheless, weaning is often not considered early enough in the course of MV in pediatric patients [39]. This may have led to a selection bias with the respiratory less stable patients not being included. Also, not all patients tolerated the lowest PS level in our study because of respiratory distress. Third, the study was not blinded but all signals were analyzed offline. From each enrolled subject, a single time period to be analyzed was manually selected. Subsequently those time periods were analyzed automatically using a customwritten Matlab script, making it unlikely that the results are affected by the unblinded nature of the study. Fourth, we found that in several patients no muscle activity could be detected from parasternal intercostals during particular ventilatory conditions or the whole study period. This could be caused by inactivity of the muscles possibly due to overassistance or oversedation, or low signal-to-noise ratio. Finally, we were not able to perform synchronized breath-by-breath analysis of EMG- and Pes-derived data as these physiological data was simultaneously recorded on two different non-synchronized devices.

Conclusions

In summary, monitoring sEMG of parasternal intercostal muscles and diaphragm in the weaning phase of ventilated children is feasible and it might be helpful in a better understanding of the pediatric ventilation liberation process. We demonstrated that both neuro-respiratory drive and neuro-mechanical efficiency increase in response to lower inspiratory assistance.

Abbreviations

CSV Continuous spontaneous ventilation
EAdi Electrical activity of the diaphragm
EMG_{amp} Amplitude electrical muscle activity
EMG_{mean} Mean electrical muscle activity
EMG_{neak} Maximal electrical muscle activity

EMG_{AUC}/min Integral of EMG signal over time during inspiration

multiplied per minute
FiO₂ Fraction of inspired oxygen

 ICM
 Parasternal intercostal muscles

 IQR
 Interquartile range

 MV
 Mechanical ventilation

 NME
 Neuro-mechanical efficiency

 PC/AC
 Pressure controlled assist/control

PC-IMV Pressure controlled-intermittent mandatory

ventilation

PEEP Positive end-expiratory pressure

'es Esophageal pressure

Pes_{amp} Tidal esophageal peak-to-trough during inspiration

PICU Pediatric intensive care unit
PS Pressure support

P-SILI Patient self-inflected lung injury
PRISM Pediatric RISK of Mortality
PTP Pressure time product
sEMG Surface electromyography

Supplementary Information

The online version contains supplementary material available at https://doi.org/10.1186/s12931-023-02374-w.

Additional file 1: Table S1. Absolute values electrical muscle activity of diaphragm and parasternal intercostals in two ventilation modes and at reducing PS levels.

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Not applicable.

Author contributions

AAK Conceptualization and design of the study, data analysis and interpretation, and writing the manuscript. JvD Conceptualization and design of the study, data collection and analysis, and writing the manuscript. RGTB Conceptualization and design of the study, data interpretation, and provided intellectual content to the manuscript. EO Data interpretation and provided intellectual content to the manuscript. MCJK Principal investigator, supervision of the project, conceptualization and design of the study, data interpretation and provided intellectual content to the manuscript. All authors edited and approved the final manuscript.

Fundina

Not applicable.

Availability of data and materials

Data sharing requests will be considered by the research group upon written request to the corresponding author.

Declarations

Ethics approval and consent to participate

The trial protocol was approved by the local Institutional Review Board (Medische Ethische Toetsingscommissie UMC Groningen, Groningen, the Netherlands), NL38361.042.11. Written informed consent was obtained from the parents or legal caretakers. The current study was performed in accordance with Dutch a wand the Declaration of Helsinki.

Consent for publication

Not applicable.

Competing interests

Martin C.J. Kneyber received lecture fees from Vyaire, Mettawa, III, USA and has received technical support from Vyaire, Mettawa, III, USA and Applied Biosignals, Weener, Germany. The remaining authors declare that they have no competing interests.

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References

- Slutsky AS, Ranieri VM. Ventilator-induced lung injury. N Engl J Med. 2013;369:2126–36.
- Brochard L, Slutsky A, Pesenti A. Mechanical ventilation to minimize progression of lung injury in acute respiratory failure. Am J Respir Crit Care Med. 2017;195:438–42.
- Yoshida T, Fujino Y, Amato MB, Kavanagh BP. Fifty years of research in ARDS. Spontaneous breathing during mechanical ventilation. Risks, mechanisms, and management. Am J Respir Crit Care Med. 2017;195:985–92.
- Yoshida T, Papazian L. When to promote spontaneous respiratory activity in acute respiratory distress patients? Anesthesiology. 2014;120:1313–5.
- Yoshida T, Uchiyama A, Fujino Y. The role of spontaneous effort during mechanical ventilation: normal lung versus injured lung. J Intensive Care. 2015;3:18.
- Yoshida T, Uchiyama A, Matsuura N, Mashimo T, Fujino Y. The comparison of spontaneous breathing and muscle paralysis in two different severities of experimental lung injury. Crit Care Med. 2013;41:536–45.
- Yoshida T, Uchiyama A, Matsuura N, Mashimo T, Fujino Y. Spontaneous breathing during lung-protective ventilation in an experimental acute lung injury model: high transpulmonary pressure associated with strong spontaneous breathing effort may worsen lung injury. Crit Care Med. 2012;40:1578–85.
- Goligher EC, Fan E, Herridge MS, Murray A, Vorona S, Brace D, et al. Evolution of diaphragm thickness during mechanical ventilation. Impact of inspiratory effort. Am J Respir Crit Care Med. 2015;192:1080–8.
- Levine S, Nguyen T, Taylor N, Friscia ME, Budak MT, Rothenberg P, et al. Rapid disuse atrophy of diaphragm fibers in mechanically ventilated humans. N Engl J Med. 2008;358:1327–35.
- Goligher EC, Dres M, Fan E, Rubenfeld GD, Scales DC, Herridge MS, et al. Mechanical ventilation-induced diaphragm atrophy strongly impacts clinical outcomes. Am J Respir Crit Care Med. 2018;197:204–13.
- Akoumianaki E, Maggiore SM, Valenza F, Bellani G, Jubran A, Loring SH, et al. The application of esophageal pressure measurement in patients with respiratory failure. Am J Respir Crit Care Med. 2014;189:520–31.
- Mauri T, Yoshida T, Bellani G, Goligher EC, Carteaux G, Rittayamai N, et al. Esophageal and transpulmonary pressure in the clinical setting: meaning, usefulness and perspectives. Intensive Care Med. 2016;42:1360–73.
- Essouri S, Baudin F, Mortamet G, Beck J, Jouvet P, Emeriaud G. Relationship between diaphragmatic electrical activity and esophageal pressure monitoring in children. Pediatr Crit Care Med. 2019;20:e319–25.
- Bellani G, Mauri T, Coppadoro A, Grasselli G, Patroniti N, Spadaro S, et al. Estimation of patient's inspiratory effort from the electrical activity of the diaphragm. Crit Care Med. 2013;41:1483–91.
- Carteaux G, Cordoba-Izquierdo A, Lyazidi A, Heunks L, Thille AW, Brochard L. Comparison between neurally adjusted ventilatory assist and pressure support ventilation levels in terms of respiratory effort. Crit Care Med. 2016;44:503–11.
- van Dijk J, Koopman AA, de Langen LB, Dijkstra S, Burgerhof JGM, Blokpoel RGT, et al. Effect of pediatric ventilation weaning technique on work of breathing. Respir Res. 2022;23:184.
- Maarsingh EJ, van Eykern LA, Sprikkelman AB, Hoekstra MO, van Aalderen WM. Respiratory muscle activity measured with a noninvasive EMG technique: technical aspects and reproducibility. J Appl Physiol. 1985;2000(88):1955–61.
- 18. Yoshida T, Brochard L. Ten tips to facilitate understanding and clinical use of esophageal pressure manometry. Intensive Care Med. 2018;44:220–2.
- Pollack MM, Patel KM, Ruttimann UE. PRISM III: an updated Pediatric Risk of Mortality score. Crit Care Med. 1996;24:743–52.
- Koopman AA, Blokpoel RGT, van Eykern LA, de Jongh FHC, Burgerhof JGM, Kneyber MCJ. Transcutaneous electromyographic respiratory muscle recordings to quantify patient-ventilator interaction in mechanically ventilated children. Ann Intensive Care. 2018;8:12.
- 21. Blokpoel RGT, Wolthuis DW, Koopman AA, Kneyber MCJ. Reverse triggering: a novel type of patient-ventilator asynchrony in mechanically ventilated children. Am J Respir Crit Care Med. 2019;200:e4–5.
- de Waal CG, van Leuteren RW, de Jongh FH, van Kaam AH, Hutten GJ. Patient-ventilator asynchrony in preterm infants on nasal intermittent positive pressure ventilation. Arch Dis Child Fetal Neonatal Ed. 2019;104:F280–4.

- 23. Kraaijenga JV, Hutten GJ, de Jongh FH, van Kaam AH. Diagnosis of hemidiaphragmatic paresis in a preterm infant with transcutaneous electromyography: a case report. Neonatology. 2015;108:38–41.
- Kraaijenga JV, Hutten GJ, de Waal CG, de Jongh FH, Onland W, van Kaam AH. Classifying apnea of prematurity by transcutaneous electromyography of the diaphragm. Neonatology. 2018;113:140–5.
- Schmidt M, Kindler F, Gottfried SB, Raux M, Hug F, Similowski T, et al. Dyspnea and surface inspiratory electromyograms in mechanically ventilated patients. Intensive Care Med. 2013;39:1368–76.
- Roesthuis LH, van der Hoeven JG, van Hees HWH, Schellekens WM, Doorduin J, Heunks LMA. Recruitment pattern of the diaphragm and extradiaphragmatic inspiratory muscles in response to different levels of pressure support. Ann Intensive Care. 2020;10:67.
- Pozzi M, Rezoagli E, Bronco A, Rabboni F, Grasselli G, Foti G, et al. Accessory and expiratory muscles activation during spontaneous breathing trial: a physiological study by surface electromyography. Front Med (Lausanne). 2022;9: 814219.
- Parthasarathy S, Jubran A, Laghi F, Tobin MJ. Sternomastoid, rib cage, and expiratory muscle activity during weaning failure. J Appl Physiol. 1985;2007(103):140–7.
- MacBean V, Jolley CJ, Sutton TG, Deep A, Greenough A, Moxham J, et al. Parasternal intercostal electromyography: a novel tool to assess respiratory load in children. Pediatr Res. 2016;80:407–14.
- van Leuteren RW, de Waal CG, de Jongh FH, Bem RA, van Kaam AH, Hutten G. Diaphragm activity pre and post extubation in ventilated critically ill infants and children measured with transcutaneous electromyography. Pediatr Crit Care Med. 2021;22:950–9.
- Bellani G, Bronco A, Arrigoni Marocco S, Pozzi M, Sala V, Eronia N, et al. Measurement of diaphragmatic electrical activity by surface electromyography in intubated subjects and its relationship with inspiratory effort. Respir Care. 2018;63:1341–9.
- Sinderby C, Beck J, Spahija J, Weinberg J, Grassino A. Voluntary activation of the human diaphragm in health and disease. J Appl Physiol. 1985:1998(85):2146–58.
- Sinderby C, Navalesi P, Beck J, Skrobik Y, Comtois N, Friberg S, et al. Neural control of mechanical ventilation in respiratory failure. Nat Med. 1999;5:1433

 6
- Blokpoel RG, Burgerhof JG, Markhorst DG, Kneyber MC. Patient-ventilator asynchrony during assisted ventilation in children. Pediatr Crit Care Med. 2016;17:e204-211.
- de la Oliva P, Schuffelmann C, Gomez-Zamora A, Villar J, Kacmarek RM. Asynchrony, neural drive, ventilatory variability and COMFORT: NAVA versus pressure support in pediatric patients. A non-randomized cross-over trial. Intensive Care Med. 2012;38:838–46.
- 36. Bordessoule A, Emeriaud G, Morneau S, Jouvet P, Beck J. Neurally adjusted ventilatory assist improves patient-ventilator interaction in infants as compared with conventional ventilation. Pediatr Res. 2012;72:194–202.
- 37. Mortamet G, Nardi N, Groleau V, Essouri S, Fauroux B, Jouvet P, et al. Impact of spontaneous breathing trial on work of breathing indices derived from esophageal pressure, electrical activity of the diaphragm, and oxygen consumption in children. Respir Care. 2019;64:509–18.
- van Leuteren RW, de Waal CG, Hutten GJ, de Jongh FH, van Kaam AH.
 Transcutaneous monitoring of diaphragm activity as a measure of work of breathing in preterm infants. Pediatr Pulmonol. 2021;56:1593–600.
- Newth CJ, Hotz JC, Khemani RG. Ventilator liberation in the pediatric ICU. Respir Care. 2020;65:1601–10.

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