Patterns of inflammatory immune responses in patients with septic shock receiving vitamin C, hydrocortisone, and thiamine: clustering analysis in Korea

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Background: Sepsis is characterized by heterogeneous immune responses that may evolve during the course of illness. This study identified inflammatory immune responses in septic patients receiving vitamin C, hydrocortisone, and thiamine.
Methods: This was a single-center, post-hoc analysis of 95 patients with septic shock who received the vitamin C protocol. Blood samples were drawn on days 1–2, 3–4, and 6–8 after shock onset. Group-based multi-trajectory modeling was used to identify immune trajectory groups.
Results: The median age was 78 years (interquartile range, 70–84 years), and 56% were male. Clustering analysis identified group 1 (n=41), which was characterized by lower interleukin (IL)-6, tumor necrosis factor (TNF)-α, and IL-10 levels, and these levels remained stationary or mildly increased until day 7. Conversely, group 2 (n=54) expressed initially higher IL-6, TNF-α, and IL-10 levels that decreased rapidly by day 4. There was a nonsignificant increase in lymphocyte count and a decrease in C-reactive protein level until day 7 in group 2. The intensive care unit mortality rate was significantly lower in group 2 (39.0% vs. 18.5%, P=0.03). Group 2 also had a significantly higher decrease in the mean (standard deviation) vasopressor dose (norepinephrine equivalent: –0.09±0.16 µg/kg/min vs. –0.23±0.31 µg/kg/min, P<0.001) and Sequential Organ Failure Assessment score (0±5 vs. –4±3, P=0.002) between days 1 and 4.
Conclusions: There may be different subphenotypes in septic patients receiving the vitamin C protocol.

Key Words: ascorbic acid; hydrocortisone; immune system; septic shock; thiamine

INTRODUCTION

Sepsis is a dysregulation of host response to infection that results in organ dysfunction and frequently death [1]. Despite major advances in critical care management, sepsis-related morbidity and mortality remain markedly high [2]. Sepsis has a complex pathophysiology that involves proinflammatory and anti-inflammatory responses, oxidative burst, endothelial

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dysfunction, coagulation activation, and metabolic dysfunction [3]. In addition, immune responses to sepsis may be heterogeneous among patients and evolve during the course of illness. Incorporating biological heterogeneity with biomarkers could help in providing individualized treatment. A recent phase 2a trial investigating the safety and tolerability of an adrenomedullin antibody in septic shock showed that patient enrollment based on the adrenomedullin level was feasible [4]. It may also be useful to monitor the immune status of septic patients and identify those who would benefit from immunomodulatory agents.

Vitamin C has pleiotropic mechanisms of action that target multiple pathogenic pathways in sepsis. Several possible beneficial effects include antioxidant, anti-inflammatory, immunomodulatory, and antithrombotic activities [5]. A preclinical model showed that vitamin C and hydrocortisone synergistically reversed the lipopolysaccharide-induced barrier dysfunction [6]. Thiamine, the enzyme required for converting pyruvate to acetyl-CoA for entry into the Krebs cycle [7], has also been suggested as a treatment option for sepsis. However, the effects of the combination of vitamin C, hydrocortisone, and thiamine (the vitamin C protocol) on clinical outcomes have been inconsistent among previous randomized trials [8-10]. It is possible that these outcomes may have been affected by patient heterogeneity. In a recent study, a 33-mRNA classifier was able to distinguish host response endotypes with prognostic significance among patients from the Outcomes of Metabolic Resuscitation Using Ascorbic Acid, Thiamine, and Glucocorticoids in the Early Treatment of Sepsis trial (ORANGES) [11]. However, there was no difference in survival when the patients receiving the vitamin C protocol were stratified according to these endotypes. Thus, it has been a challenge to identify the optimal population of patients with sepsis who would benefit from the vitamin C protocol. The present study aimed to identify the subphenotypes of inflammatory immune responses in septic patients receiving the vitamin C protocol.

MATERIALS AND METHODS

Study Design and Population
This was a post-hoc analysis of a prospective cohort study conducted in an 835-bed university-affiliated tertiary care hospital (Chung-Ang University Hospital, Seoul, Korea) between September 2019 and July 2021. Written informed consent was obtained directly from participants who were deemed to have the capacity to make their own decisions. If a participant was

KEY MESSAGES

- Clustering analysis identified two subphenotypes of inflammatory immune responses in septic patients receiving vitamin C, hydrocortisone, and thiamine.
- Group 1 involved sustained levels of interleukin (IL)-6, tumor necrosis factor-α, and IL-10, while group 2 involved higher initial levels of these cytokines that rapidly declined thereafter.
- Group 2 demonstrated a better clinical course and lower intensive care unit mortality, and inclusion in group 2 was independently associated with a lower risk of 60-day mortality.

not determined to be capable of consent due to current mental or physical state, a legally authorized representative was asked to provide consent. The study protocol was approved by the Institutional Review Board of Chung-Ang University Hospital (No. 1820-006-353).

Adult patients (age ≥19 years) who were admitted to the medical intensive care unit (ICU) within 48 hours of diagnosis of septic shock and received the vitamin C protocol were screened for inclusion. Patients were excluded if they were aged <19 years, did not have septic shock, did not receive the vitamin C protocol, had cardiac arrest, were moribund and not expected to survive 24 hours, had a do-not-resuscitate order, refused to participate, or did not have available blood samples. Septic shock was defined as sepsis with persisting hypotension requiring vasopressors to maintain a mean arterial pressure of ≥65 mm Hg and a serum lactate level of >2 mmol/L despite adequate fluid resuscitation [1]. However, patients with suspected infection who had serum lactate levels of <2 mmol/L but required high-dose vasopressors were also screened.

The vitamin C protocol involved a combination of intravenous vitamin C 1.5 g every 6 hours for 4 days, hydrocortisone 50 mg every 6 hours for 7 days, and thiamine 200 mg every 12 hours for 4 days [12]. Patient management was performed according to the recommendations from the 2016 Surviving Sepsis Campaign guidelines [13].

Data Collection and Variable Definition
Baseline data including age, sex, body mass index, incidence of comorbidities, nosocomial infection, bacteremia, and acute respiratory distress syndrome (ARDS), cause of sepsis, Acute Physiology and Chronic Health Evaluation (APACHE) II score [14], Sequential Organ Failure Assessment (SOFA)

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score [15], antibiotics, mechanical ventilation, and renal replacement therapy were collected. Clinical data of vital signs, laboratory tests, vasopressor dose, SOFA score, fluid intake, and output volumes within the first 4 days after ICU admission were also collected. The daily vasopressor dose was acquired at 07:00 and converted to an equivalent norepinephrine dose [16]. Moreover, the time of shock onset and of vitamin C protocol initiation and discontinuation were recorded. ARDS was diagnosed using the Berlin definition [17]. Vasopressor weaning was defined as not receiving vasopressors for ≥48 hours. Ventilator weaning was determined as being free from ventilator support for ≥48 hours. Superinfection was determined as a diagnosis of a new microbiological infection that occurred ≥48 hours after admission and required a new course of antibiotics.

Biomarker Measurement

Blood samples were prospectively collected at three time points after septic shock diagnosis: days 1–2, 3–4, and 6–8. The samples were evaluated for interleukin (IL)-6, tumor necrosis factor (TNF)-α, and IL-10 levels by blinded laboratory physicians, using the sandwich enzyme-linked immunosorbent assay kit (human IL-6/TNF-α/IL-10 Quantikine ELISA Kit, R&D Systems). In addition, lymphocyte counts from complete blood cell counts and C-reactive protein (CRP) levels at days 1–2, 3–4, and 6–8 after septic shock diagnosis were recorded.

Statistical Analysis

Continuous data were presented as the mean (standard deviation [SD]) or as the median (interquartile range [IQR]) and were compared using the Mann-Whitney U-test. Categorical data were presented as the number (percentage) and were compared using the chi-square test or Fisher's exact test, as appropriate. Missing values of lymphocyte count, CRP level, lactate level, norepinephrine equivalent dose, and inflammatory biomarker level were imputed using the last observation carried forward (LOCF) approach. For missing SOFA scores, a maximum SOFA score of 24 was assigned to deceased patients, and the LOCF approach was used for discharged patients [18].

To define the trajectories of serial biomarker patterns regardless of clinical information, group-based multi-trajectory modeling (GBMTM) was performed based on longitudinal measurements (days 1–2, 3–4, and 6–8) of lymphocyte count and CRP, IL-6, TNF-α, and IL-10 levels. Briefly, GBMTM is an application of finite mixture modeling designed to identify latent clusters of individuals following similar trajectories across

multiple variables of interest [19]. Maximum likelihood was used to determine both the trajectory shape for each group and the estimated probabilities of each individual belonging to each trajectory group. The Akaike information criterion and the Bayesian information criterion were used for measuring the adequacy and fit of trajectory model and final model selection. Standardization (centering and division by the sample SD) was performed to enable comparisons among the measurements of different biomarkers [20].

Baseline characteristics and inflammatory biomarkers, clinical course, and outcomes were compared between the identified trajectory groups. To minimize survivorship bias, a rank-based statistical analysis was performed by applying a worst-rank SOFA score to deceased patients. However, this may have introduced additional sources of bias. Thus, a sensitivity analysis was conducted to test the consistency of the main results, with the SOFA scores recalculated excluding those of the deceased patients. Multivariable Cox proportional hazard regression was performed to quantify the association of baseline characteristics and of the trajectory groups with 60-day mortality. Significant variables in the univariate analysis of <0.05 were included in the multivariate analysis with stepwise backward selection. Survival curves were plotted and compared between the trajectory groups using Cox regression. All statistical analyses were performed using SAS software ver. 9.4 (SAS Institute) and the corresponding implementation PROC TRAJ, freely available at https://www.andrew.cmu.edu/~bjones/index.htm. All tests were two tailed, and differences were considered statistically significant at a P-value of <0.05.

RESULTS

Patient Characteristics

Among the 392 patients identified with sepsis, 95 had septic shock diagnosis and were analyzed (Figure 1). The median age was 78 years (IQR, 70–84 years), with a higher proportion of men (56%). In total, 26 patients (27.4%) died in the ICU. The baseline characteristics of the survivors and non-survivors are described in Supplementary Table 1. Non-survivors were more likely to be immunosuppressed, have pneumonia, have higher APACHE II and SOFA scores, and have higher requirements for mechanical ventilation and renal replacement therapy. The mean (SD) duration of vitamin C administration was 4.0±0.9 days for survivors and 4.0±1.8 days for non-survivors (P=0.12). The median time from shock onset to initiation of the vitamin C protocol was significantly shorter in survivors (1 hours [IQR,
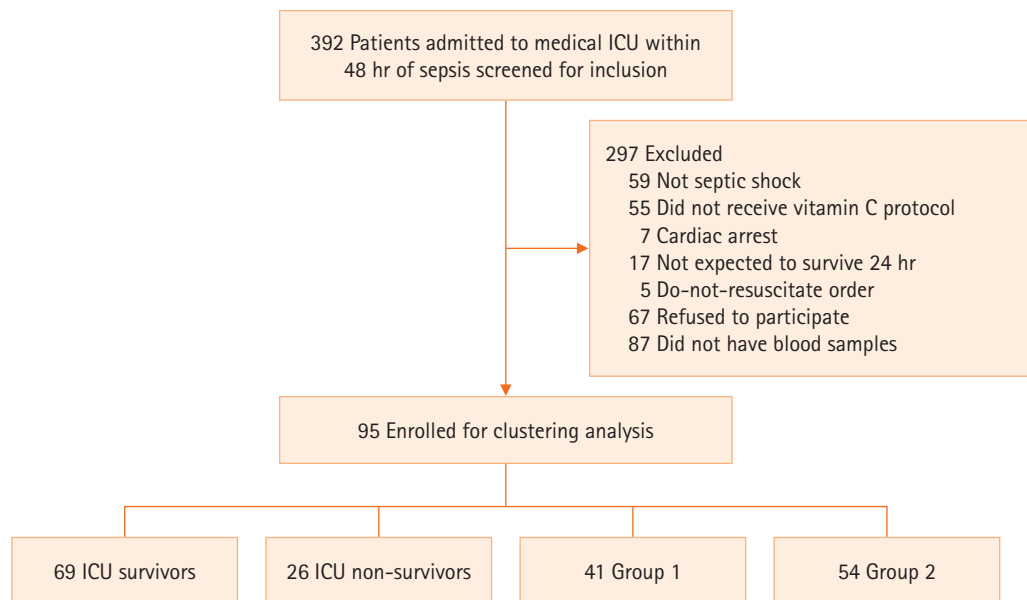
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Figure 1. Patient inclusion flowchart. ICU: intensive care unit.

Table 1. BIC and AIC values and predicted group proportions from GBMTM

Group	BIC	AIC	Patients in each predicted group (%)						
			1	2	3	4	5	6	7
2	−1,786.28	−1,731.37	43.2	56.8					
3	−1,787.18	−1,708.01	22.1	60.0	17.9				
4	−1,872.80	−1,769.37	0	43.2	56.8	0			
5	−1,916.07	−1,788.37	0	43.2	0	56.8	0		
6	−1,849.98	−1,698.02	0	8.4	29.5	48.4	13.7	0	
7	−1,893.24	−1,717.02	0	8.4	29.5	0	48.4	13.7	0

The optimal model (the two-group model suggested) was considered based on the pattern of results and the balancing of BIC values and subgroup sizes.
BIC: Bayesian information criterion; AIC: Akaike information criterion; GBMTM: group-based multi-trajectory modeling.

0–6] vs. 4 hours [IQR, 2–7], P=0.02). Violin plots showing the distribution of pooled time are presented in Supplementary Figure 1. Baseline (days 1–2) lymphocyte count and CRP, IL-6, TNF-α, and IL-10 levels were similar between the groups (Supplementary Tables 1 and 2).

The clinical parameters at days 4 and 7 in ICU survivors and non-survivors are shown in Supplementary Table 3. Compared to non-survivors, survivors had a significantly greater decrease in vasopressor dose and SOFA score between days 1 and 4 (Supplementary Table 3, Supplementary Figure 2). Inflammatory biomarkers at days 3–4 and 6–8 for ICU survivors and non-survivors are shown in Supplementary Table 4 and Supplementary Figure 3. Survivors had a significantly higher lymphocyte count and lower levels of CRP, IL-6, IL-10, and IL-10/TNF-α at days 3–4 and 6–8.

Clustering Analysis

GBMTM categorized the 95 patients into two groups based on temporal measurements of lymphocyte count and CRP, IL-6, TNF-α, and IL-10 levels (Table 1, Figure 2). The levels of IL-6, TNF-α, and IL-10 at day 1 were significantly lower in group 1 (n=41, 43%) than in group 2 (n=54, 57%). In group 1, the levels remained stationary or mildly increased until day 7. The day 1 mean (SD) of IL-6, TNF-α, and IL-10 levels peaked at 558.6±1,664.5 pg/ml, 37.1±32.4 pg/ml, and 130.6±194.8 pg/ml, respectively, in group 2. However, the levels decreased rapidly by day 4. Inflammatory biomarkers at days 1–2, 3–4, and 6–8 in groups 1 and 2 are summarized in Supplementary Table 5. All measurements were fully captured without any missing values at baseline (days 1–2). During the follow-up, missing values were identified and imputed in 14/95 patients (15%). Details of
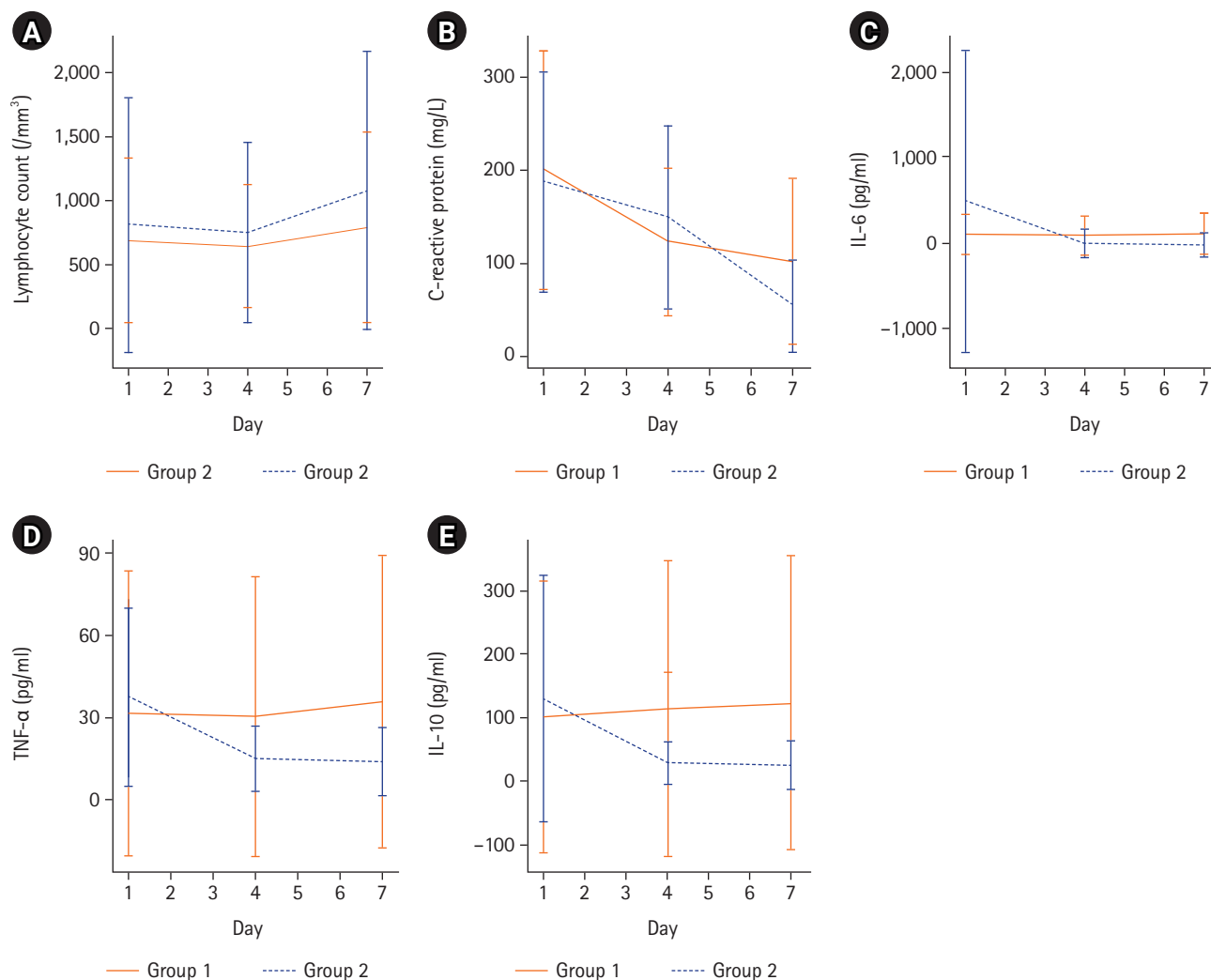
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Figure 2. Clustering of inflammatory responses in septic patients receiving the vitamin C protocol. Group-based multi-trajectory modeling reveals two subphenotypes of inflammatory immune responses (group 1, 41 patients; group 2, 54 patients). The mean changes in (A) lymphocyte counts, (B) C-reactive protein levels, (C) Interleukin (IL)-6 levels, (D) tumor necrosis factor (TNF)-α levels, and (E) IL-10 levels in groups 1 and 2 during the study period.

blood sampling including the reasons for missing samples are listed in Supplementary Table 6. Both groups showed low lymphocyte counts and high CRP levels at day 1. However, group 2 showed a greater increase in mean (SD) lymphocyte count (102 ± 754/mm^3 vs. $272\pm1,517$/mm^3, P=0.29) and a greater decrease in mean (SD) CRP level (-98 ± 126 mg/L vs. -134 ± 123 mg/L, P=0.13) between days 1 and 7, although these differences were not statistically significant (Figure 2, Supplementary Table 7).

Comparison of Baseline Characteristics between the Trajectory Groups

The baseline characteristics were not significantly different

between the groups, although patients in group 1 were more likely to be immunosuppressed (Table 2). Immunocompromised patients showed lower lymphocyte counts and higher TNF-α and IL-10 levels, whereas immunocompetent patients showed a greater decrease in IL-6 levels between days 1 and 4 (Supplementary Figure 4). All patients were treated with antibiotics, and the rates of mechanical ventilation and renal replacement therapy were similar between the groups. The lactate level and vasopressor dose were significantly higher for group 2. The mean (SD) duration of vitamin C administration was significantly longer for group 2 (3.6 ± 0.9 days vs. 4.4 ± 1.2 days, P=0.002), although the difference was not statistically significant after the exclusion of the deceased patients (3.9 ± 0.7

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Table 2. Baseline patient characteristics by group

Characteristics	Group 1 (n=41)	Group 2 (n=54)	P-value
Age (yr)	75 (70–84)	78 (68–84)	0.94
Sex			0.72
Male	22 (54)	31 (57)	
Female	19 (46)	23 (43)	
Body mass index (kg/m^2)	21.3 (18.5–23.5)	21.2 (18.1–24.2)	0.93
Comorbidity			
Diabetes	14 (34)	26 (48)	0.17
Hypertension	25 (61)	34 (63)	0.84
Chronic heart failure	5 (12)	4 (7)	0.49
Chronic lung disease	7 (17)	15 (28)	0.22
Liver cirrhosis	4 (10)	3 (6)	0.46
Chronic kidney disease	6 (15)	9 (17)	0.79
Immunosuppression[a]	15 (37)	8 (15)	0.01
Nosocomial infection	17 (42)	16 (30)	0.23
Cause of sepsis			
Pneumonia	25 (61)	29 (54)	0.48
Urosepsis	7 (17)	13 (24)	0.41
Gastrointestinal/biliary	8 (20)	7 (13)	0.39
Bacteremia	13 (32)	22 (41)	0.37
ARDS	9 (22)	12 (22)	0.98
APACHE II score	28 (20–35)	27 (20–33)	0.47
SOFA score	12±3	12±3	0.49
Antibiotics	41 (100)	54 (100)	-
Mechanical ventilation	29 (71)	37 (69)	0.82
Renal replacement therapy	13 (32)	22 (41)	0.37
Vital signs and laboratory data			
Body temperature (°C)	36.9 (36.5–37.6)	37.1 (36.6–37.6)	0.54
Mean arterial pressure (mm Hg)	59 (52–65)	62 (54–66)	0.26
Respiratory rate (breaths/min)	30 (26–32)	28 (25–32)	0.38
PaO$_2$/FiO$_2$	194 (113–281)	179 (113–248)	0.67
Bicarbonate (mEq/L)	21.0 (18.3–23.6)	21.7 (17.6–24.5)	0.51
Creatinine (mg/dl)	1.3 (0.9–1.9)	1.3 (0.9–2.3)	0.96
Lymphocyte count (/mm^3)	690±644	810±998	0.60
Total bilirubin (mg/dl)	0.6 (0.5–1.3)	0.9 (0.5–1.8)	0.41
C-reactive protein (mg/L)	200±128	188±119	0.67
Procalcitonin (ng/ml)	2.4 (0.7–23.4) (n=38)	5.8 (1.6–40.5) (n=52)	0.06
Lactate (mmol/L)	1.7 (1.3–3.0)	2.7 (1.9–4.9)	0.02
Norepinephrine equivalent dose (µg/kg/min)	0.19±0.21	0.29±0.24	0.006
Duration (day)			
Vitamin C	3.6±0.9	4.4±1.2	0.002
Hydrocortisone	4.2±2.6	5.0±1.9	0.004
Thiamine	3.6±0.9	4.4±1.2	0.002
Time from shock onset to vitamin C protocol (hr)	3 (1–6)	1 (0–7)	0.36

Values are presented as median (interquartile range), number (%), or mean±standard deviation.

ARDS: acute respiratory distress syndrome; APACHE: Acute Physiology and Chronic Health Evaluation; SOFA: Sequential Organ Failure Assessment; PaO$_2$: arterial partial pressure of oxygen; FiO$_2$: fraction of inspired oxygen.

a) Immunosuppression includes malignancies, human immunodeficiency virus infection, severe neutropenia, or administration of immunosuppressive therapy.

days vs. 4.4±1.3 days, P=0.08). The median (IQR) time from shock onset to initiation of the vitamin C protocol was 3 hours (1–6) in group 1 and 1 hours (0–7) in group 2 (P=0.36). Violin plots showing the distribution of pooled time are presented in Supplementary Figure 5.

Comparison of Clinical Courses and Outcomes between the Trajectory Groups

The clinical outcomes in groups 1 and 2 are summarized in Table 3. The ICU mortality rates were 39.0% (16/41 patients) for group 1 and 18.5% (10/54 patients) for group 2 (P=0.03). Compared with group 1, group 2 had a significantly higher decrease in mean (SD) vasopressor dose (norepinephrine equivalent, –0.09±0.16 µg/kg/min vs. –0.23±0.31 µg/kg/min, P<0.001) and SOFA score (0±5 vs. –4±3, P=0.002) between days 1 and 4 (Table 3, Figure 3). Meanwhile, no significant differences were observed between the groups regarding net fluid retention, vasopressor weaning rate, vasopressor-free days at day 28, ventilator weaning rate, ventilator-free days at day 28, and hospital length of stay. With respect to mortality, the rates were also lower in group 2, but only the 28-day mortality was significantly different from that in group 1. Meanwhile, the rates of superinfection were similar between the groups. The

clinical parameters on days 4 and 7 in groups 1 and 2 are listed in Supplementary Table 7. No significant differences were observed between the groups regarding SOFA scores on days 4 and 7 after the exclusion of deceased patients (Supplementary Table 8).

Association between Trajectory Group Inclusion and 60-Day Mortality

After adjustment for confounding factors, the independent risk factors of mortality were higher APACHE II score (adjusted hazard ratio [HR], 1.09; 95% confidence interval [CI], 1.04–1.14; P<0.001) and longer hydrocortisone treatment (adjusted HR, 1.14; 95% CI, 1.01–1.30; P=0.04) (Table 4). Meanwhile, inclusion in group 2 was significantly associated with decreased mortality (adjusted HR, 0.32; 95% CI, 0.16–0.64; P=0.001) (Table 4, Supplementary Figure 6).

DISCUSSION

The present study based on clustering analysis of lymphocyte count and CRP, IL-6, TNF-α, and IL-10 levels identified two subphenotypes of inflammatory immune responses in septic patients receiving the vitamin C protocol. One type involved

Table 3. Clinical outcomes by group

Outcomes	Group 1 (n=41)	Group 2 (n=54)	P-value
Mortality			
ICU	16 (39.0)	10 (18.5)	0.03
28 Day	20 (48.8)	12 (22.2)	0.007
60 Day	20 (48.8)	17 (31.5)	0.09
Change in norepinephrine equivalent dose at day 4 (µg/kg/min)	–0.09±0.16	–0.23±0.31	<0.001
Change in SOFA score at day 4	0±5	–4±3	0.002
Net fluid retention (ml)[a]			
Day 1	1,280 (696 to 2,791)	1,118 (240 to 2,261)	0.28
Day 4	180 (–338 to 1,053) (n=31)	315 (–392 to 785) (n=54)	0.62
Day 7	279 (–341 to 725) (n=28)	160 (–769 to 850) (n=49)	0.73
Vasopressor weaning	31 (76)	46 (85)	0.24
Vasopressor-free day at day 28	26 (17 to 27)	26 (22 to 27)	0.55
Ventilator weaning	14 (48) (n=29)	24 (65) (n=37)	0.18
Ventilator-free day at day 28	10±11 (n=29)	13±11 (n=37)	0.19
Length of stay (day)			
ICU	6 (2 to 14)	9 (5 to 18)	0.03
Hospital	22 (16 to 49)	31 (17 to 50)	0.07
Superinfection	8 (20)	14 (26)	0.46

Values are presented as number (%), mean±standard deviation, or median (interquartile range).
ICU: intensive care unit; SOFA: Sequential Organ Failure Assessment.
a) Net fluid retention is calculated as the difference between intake and output of all fluids (urine volume, dialysis volume, drainage volume, and stool weight).
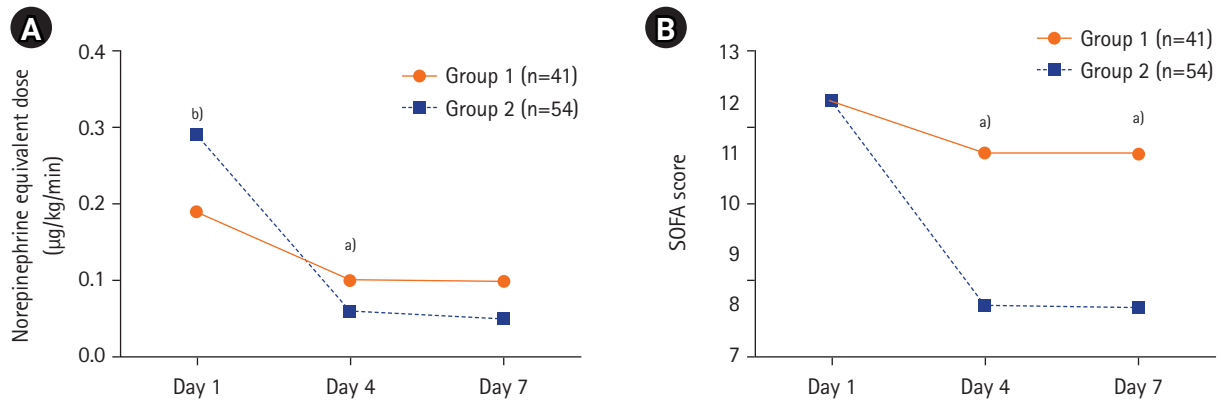
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Figure 3. Mean changes in (A) vasopressor dose (norepinephrine equivalent) and (B) Sequential Organ Failure Assessment (SOFA) score by group during the study period. $^{a)}$P<0.05, $^{b)}$P<0.01 when group 1 and group 2 are compared using the Mann–Whitney U-test.

sustained levels of IL-6, TNF-α, and IL-10 (group 1), while the other involved higher initial levels of these cytokines that rapidly declined thereafter (group 2). The baseline characteristics were similar between the trajectory groups, but group 2 demonstrated a better clinical course and lower ICU mortality. Lastly, inclusion in group 2 was independently associated with a lower risk of 60-day mortality.

One of several barriers to the effective treatment of critical illness has been discovering the optimal population of patients who would benefit from specific agents. Several studies suggest that more personalized treatment modalities can be considered via the identification of subphenotypes [21-23]. Interestingly, among patients from the Vasopressin vs. Norepinephrine as Initial Therapy in Septic Shock (VANISH) trial with the immunocompetent endotype, the mortality rate was higher among those treated with corticosteroids than among those treated with placebo [23]. In the current study, two trajectory groups were found to have unique cytokine signatures: group 1 involved persistent immune dysregulation, while group 2 involved a recovered well-balanced immune system after an initial adaptive response. Notably, the patients in these groups had markedly different clinical courses and outcomes despite having similar baseline characteristics.

However, it was difficult to establish a direct effect of the vitamin C protocol on improvement of the immune status owing to the lack of a control group. However, in a phase 1 trial, patients administered with high-dose vitamin C (200 mg/kg/day) showed significantly lower inflammatory biomarker levels than those who received the placebo [24]. In a recent randomized trial of vitamin C for critically ill patients with coronavirus disease 2019, the IL-6 level on day 7 was lower in the treatment

group than in the placebo group [25]. Interestingly, the survival benefit was evident in those with a higher baseline SOFA score, and the current study found consistent results that patients with initially higher lactate level, vasopressor dose, and inflammatory biomarker level had better outcomes.

Dynamic measures of variables may be more important than static measures. In this study, single values of baseline lymphocyte count and CRP, IL-6, TNF-α, or IL-10 levels were not useful in predicting outcomes, whereas the measurements at later time points provided prognostic information. Bhavani et al. reported a novel method to identify four sepsis subphenotypes based on longitudinal temperature trajectories [26]. Among these, the "hyperthermic, fast resolvers" showed the lowest mortality rate. There was no immunological basis for the proposed temperature-based subphenotypes, although the "hyperthermic, fast resolvers" may be similar to that of group 2 (high IL-6, TNF-α, and IL-10 levels at day 1 but rapidly decreased at day 4) in the present study. These findings support that quicker reversal of the pathophysiological process of sepsis should lead to better survival [27].

IL-6 acts as a crucial cytokine in the systemic inflammatory response, especially during tissue injury and organ dysfunction [28]. Meanwhile, IL-10 has a key role in modulating anti-inflammatory and immunosuppressive responses [29]. Previous studies have shown that low levels of IL-6 and IL-10 during the initial phase of septic shock were significantly associated with disease severity and survival [30,31]. The decreased IL-6, TNF-α, and IL-10 levels in group 2 and their value in predicting mortality are consistent with the results of preclinical studies showing that vitamin C administration to human monocytes decreased the levels of proinflammatory cytokines

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Table 4. Association between baseline characteristics and 60-day mortality

Characteristics	Unadjusted HR (95% CI)	P-value	Adjusted HR[a] (95% CI)	P-value
Age	1.02 (0.99–1.05)	0.28		
Male	0.91 (0.47–1.73)	0.91		
Body mass index	0.97 (0.90–1.04)	0.43		
Diabetes	0.93 (0.48–1.78)	0.82		
Hypertension	1.02 (0.52–1.97)	0.97		
Chronic heart failure	1.38 (0.49–3.89)	0.55		
Chronic lung disease	1.10 (0.52–2.34)	0.80		
Liver cirrhosis	1.11 (0.34–3.61)	0.87		
Chronic kidney disease	0.73 (0.28–1.86)	0.50		
Immunosuppression[b]	2.28 (1.16–4.49)	0.02	1.03 (0.43–2.45)	0.95
Nosocomial infection	2.17 (1.14–4.15)	0.02	1.51 (0.76–3.03)	0.24
Pneumonia	1.75 (0.88–3.48)	0.11		
Urosepsis	0.53 (0.21–1.37)	0.19		
Gastrointestinal/biliary	0.63 (0.22–1.77)	0.38		
Bacteremia	1.22 (0.63–2.36)	0.55		
ARDS	1.78 (0.89–3.54)	0.10		
APACHE II score	1.11 (1.06–1.16)	<0.001	1.09 (1.04–1.14)	<0.001
SOFA score	1.23 (1.09–1.39)	0.001	1.06 (0.90–1.25)	0.51
Mechanical ventilation	4.52 (1.60–12.79)	0.004	1.30 (0.35–4.78)	0.69
Renal replacement therapy	2.73 (1.42–5.22)	0.002	2.02 (0.96–4.24)	0.06
Lymphocyte count	1.00 (1.00–1.00)	0.55		
C-reactive protein	1.00 (1.00–1.00)	0.85		
Lactate	1.08 (0.96–1.21)	0.19		
Norepinephrine equivalent dose	0.34 (0.06–1.88)	0.22		
Vitamin C duration	0.83 (0.60–1.14)	0.24		
Thiamine duration	0.83 (0.60–1.14)	0.24		
Hydrocortisone duration	1.17 (1.03–1.34)	0.02	1.14 (1.01–1.30)	0.04
Time from shock onset to vitamin C protocol	1.01 (0.96–1.05)	0.77		
IL-6	1.00 (1.00–1.00)	0.73		
TNF-α	1.01 (0.00–1.01)	0.16		
IL-10	1.00 (1.00–1.00)	0.04	1.00 (1.00–1.00)	0.17
Group 2	0.48 (0.25–0.92)	0.03	0.32 (0.16–0.64)	0.001

HR: hazard ratio; CI: confidence interval; ARDS: acute respiratory distress syndrome; APACHE: Acute Physiology and Chronic Health Evaluation; SOFA: Sequential Organ Failure Assessment; IL: interleukin; TNF: tumor necrosis factor.
a) Adjusted for immunosuppression, nosocomial infection, APACHE II score, SOFA score, mechanical ventilation, renal replacement therapy, hydrocortisone duration, IL-10, and inclusion in group 2; b) Immunosuppression includes malignancies, human immunodeficiency virus infection, severe neutropenia, or administration of immunosuppressive therapy.

[32]. Reduced lymphocyte count and impaired lymphocyte function are common features of sepsis-related immunosuppression [33]. Lastly, CRP exerts both proinflammatory and anti-inflammatory effects, and its elevation can be considered as a contributing factor to persistent inflammation/ immunosuppression and catabolism syndrome [34,35]. In the present study, the lymphocyte count was higher and the CRP level was lower in group 2 than in group 1 by day 7. Taken together, these findings support the relationship between immune trajecto-

ries and clinical outcomes in patients with sepsis receiving the vitamin C protocol and will also be helpful in identifying patients who would better respond to treatment.

The main strength of the current study was the use of clustering analysis of inflammatory immune responses. Unlike previous randomized trials, the vitamin C protocol in this study was initiated earlier within an average of 4 hours after shock onset, and the interval between shock onset and vitamin C administration was significantly shorter in survivors.

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Furthermore, the duration of vitamin C administration was significantly longer in group 2. These are consistent with previous reports suggesting that early or long-term vitamin C administration can improve patient outcomes [36,37].

However, this study has several limitations. First, the generalizability of the results are limited by the single-center design, relatively small number of patients, and the use of random set of biomarkers to define the trajectories. However, the small study population was a result of strict patient selection to ensure standardization and reproducibility of biomarker measurements. Second, there was no control group, and the effect of other sepsis management strategies on the improvement in the clinical outcomes of group 2 could not be distinguished. In addition, vitamin C levels were not closely monitored; thus, it was unclear whether the vitamin C protocol had a direct effect against septic shock. Third, a considerable proportion of patients were excluded because of their refusal to participate or the unavailability of blood samples. It is possible that the inclusion of these patients might have influenced the modeling. Some of the included patients also had missing data owing to death or hospital discharge, and the LOCF approach may be inadequate to minimize this bias. Fourth, subgroup analysis among different sepsis subpopulations and validation of clustering analysis results were not feasible owing to the small sample size. Fifth, the current trajectory model requires serial biomarker data, and this may limit its immediate clinical use. However, survival prediction in sepsis is also currently challenging owing to the lack of available tools. Sixth, the patients were not tested for relative adrenal insufficiency. In the presence of relative adrenal insufficiency, there may be a high likelihood of favorable response to hydrocortisone, which may also explain the outcomes of group 2. Seventh, the associations between the study groups and SOFA scores were attenuated after the exclusion of deceased patients. Lastly, this study included many older patients, reflecting the high rate of ICU admission among older patients in Korea. Our results should be validated in a large placebo-controlled trial.

In conclusion, there may be different subphenotypes in septic patients receiving the vitamin C protocol. Adequate initial adaptive responses with elevated IL-6, TNF-α, and IL-10 levels followed by a restoration of a well-balanced immune system with a rapid decline in the levels of these biomarkers are associated with better clinical course and outcomes. The ability to define the immune status of patients with sepsis may contribute to the success of future clinical trials of immunomodulatory therapies through the identification of the proper patient subset. The present results also suggest that the dynamic nature of immune responses should be considered in survival prediction in sepsis. Finally, further studies are required to assess the efficacy of early and long-term vitamin C in selected patients with sepsis, using a well-defined control group.

CONFLICT OF INTEREST

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SUPPLEMENTARY MATERIALS

Supplementary materials can be found via https://doi.org/10.4266/acc.2023.00507.

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