



# Ultra-processed food consumption and the risk of non-alcoholic fatty liver disease—What are the proposed mechanisms?

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## Abstract

A high consumption of ultra-processed food (UPF) is a hallmark of Western diets that has been related to increased risk of non-communicable diseases. As an underlying mechanism, UPF may promote non-alcoholic fatty liver disease (NAFLD) which is a key driver of metabolic impairment with extra-hepatic manifestations like type 2 diabetes, cardiovascular disease, chronic kidney disease, and osteoporosis among others. The present review provides an overview of UPF properties that may promote NAFLD and are thus potential targets for reformulation of UPF. Such approaches should address improvements in the quality of carbohydrates and fat, changes in food texture that lower eating rate as well as ingredients that prevent excess caloric intake or avoid dysbiosis and leaky gut syndrome. Promising strategies are enrichment with fiber, prebiotics, phytochemicals, and protein with a concurrent reduction in glycemic load, energy density, saturated fatty acids (FA; SFA), emulsifiers, fructose, and non-caloric sweeteners. Future studies are needed to examine the interactive and protective effects of such modifications in the composition of UPF on prevention and treatment of NAFLD.

## Keywords

Ultra-processed foods, non-alcoholic fatty liver disease, appetite control, energy balance, saturated fatty acids, glycemic load

## Introduction

The development of non-alcoholic fatty liver disease (NAFLD) is closely linked to lifestyle factors that promote a positive energy balance. The increasing adoption of ultra-processed food (UPF) according to the NOVA-classification [1] is coincidental to the rising global prevalence rate of NAFLD, e.g., in Asia over the past 15 years (from +25.3% between 1999–2005 to +33.9% between 2012–2017) [2]. Whereas increasing evidence shows that the consumption of UPF promotes excess energy intake and may thus contribute to the development of NAFLD, there are also other proposed mechanisms that may explain the simultaneous

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increase in NAFLD and the consumption of UPF. According to the NOVA-classification, food products can be classified depending on processing as (1) unprocessed or minimally processed foods, (2) processed culinary ingredients, (3) processed foods, and (4) UPF [1]. Potential mechanisms that link the consumption of UPF to the development of NAFLD are typical characteristics of “Western” type foods like hyper-palatability, soft texture, high energy density, high saturated fat, and high sugar content. The present review addresses these mechanisms as potential targets for reformulation of UPF mainly by analyzing observational studies but also considering evidence from human interventions as well as animal studies.

## Consumption of UPF may lead to NAFLD by promoting overfeeding

NAFLD is a common comorbidity of obesity and the metabolic syndrome [3, 4]. Compared to persons with normal weight, individuals with obesity are at 3.5-fold higher risk for NAFLD [3]. Excess hepatic triglyceride accumulation originates from adipose tissue lipolysis (~60%), *de novo* lipogenesis in hepatocytes (~25%) and from diet (~15%) [5]. Since the majority of adipose tissue-derived free fatty acids (FA; FFA) passing through the liver originates from lipolysis of abdominal subcutaneous adipose tissue [6], central obesity with impaired lipid storage capacity rather than the total amount of fat mass is a major risk factor for accumulation of liver fat.

An increasing number of cross-sectional and prospective cohort trials have shown a positive association between UPF consumption according to NOVA classification and the risk for weight gain and obesity [7–12], for a meta-analysis see [13]. In prospective studies, a higher UPF consumption was associated with a greater accumulation of abdominal obesity or visceral fat [14, 15]. In a cross-sectional study in adults with or without NAFLD, a positive association between UPF consumption and metabolic syndrome has been observed [16]. Although no association between a higher UPF consumption and NAFLD was found, subjects with a higher UPF consumption had a higher total energy intake, a higher relative intake of carbohydrates and a lower relative protein intake. However, among subjects with NAFLD, a higher UPF consumption was associated with higher odds for presumed non-alcoholic steatohepatitis (NASH) [16]. In an analysis of the PREDIMED-Plus trial (5,867 subjects, one-year follow-up), a higher UPF consumption was associated with higher levels of the non-invasive fatty liver index and the hepatic steatosis index [17]. Similarly, in a prospective cohort of Chinese adults (16,168 subjects, 4.2 years follow-up) increasing UPF consumption by one standard deviation (62.7 g/1,000 kcal per day) increased the risk for NAFLD by 6%, even when adjusted for traditional risk factors as age, sex, body mass index (BMI), smoking, etc. [18].

Causality between UPF consumption and weight gain can, however, only be derived from intervention studies. In a highly regarded randomized controlled trial by Hall et al. [19] in 20 inpatient adults (10 women and 10 men, 31.2 years  $\pm$  1.6 years, weight-stable with BMI 27 kg/m<sup>2</sup>  $\pm$  1.5 kg/m<sup>2</sup>, as mean  $\pm$  SE), *ad libitum* energy intake was increased by ~500 kcal/day when UPF were provided compared to a largely unprocessed diet matched for energy content, macronutrients and fiber. During 14 days of each intervention, participants gained 0.9 kg body weight consuming the ultra-processed diet whereas they lost 0.9 kg body weight consuming the unprocessed diet. These findings suggest, that there is indeed a causal relationship between regular consumption of a diet high in UPF and an increased risk for weight gain and obesity.

## High energy density of UPF may lead to overfeeding

There are several mechanisms by which UPF consumption can trigger excess calorie intake. First of all, UPF usually have a higher energy density (i.e., energy content divided by weight of food served; kcal/g) compared to low processed foods (LPF). More energy dense diets tend to be associated with a higher daily energy intake and weight gain [20]. In an elegantly conducted intervention trial with normal-weight women, food consumption during a meal was more consistent in terms of weight of food compared to energy content of the consumed food when palatability and fat content were kept constant [21]. This insensitivity to food energy density was not only observed in women but also in men and in adults of different weight status as well as in 3- to 5-year-old children [22]. This insensitivity facilitates “passive overconsumption” [23] when an energy dense diet is consumed [21] and may partly explain the excess

caloric intake with a diet high in UPF. This effect has been confirmed by the study by Hall and colleagues [19] because the UPF diet had a higher non-beverage energy density compared to the unprocessed diet (1.95 kcal/g vs. 1.05 kcal/g). In line with this finding, a recent meta-analysis has shown that lowering the energy density of the diet (with or without macronutrient manipulation) was associated with a large decrease of daily energy intake. This effect was dose-dependent and only minimal compensation at the following meals was observed [24]. A recently published secondary analysis using controlled intervention data from Hall et al. [19] as well as free-living data from the UK National Diet and Nutrition Survey, however, suggests that passive overconsumption only occurs to some degree and that overcompensation for higher energy density of foods (e.g., by reducing portion size) can be seen [25].

## High eating rate of UPF consumption may lead to overfeeding

Eating more slowly may prevent overconsumption as it allows to register satiation before too much food is consumed [26, 27]. It has been shown that eating rates and thus also energy intake rates [i.e., eating rate (g/min) multiplied by the energy density (kcal/g) of the specific food; kcal/min] differ according to food processing defined by NOVA classification. Thus, UPF consumption leads to higher average energy intake rates (69 kcal/min  $\pm$  3 kcal/min) compared to processed (54 kcal/min  $\pm$  4 kcal/min) and unprocessed foods (36 kcal/min  $\pm$  4 kcal/min) [28]. This would provide an additional mechanism by which UPF may lead to overconsumption and weight gain. In line with this supposition, in the randomized controlled intervention by Hall and colleagues [19], the energy intake rate was increased by > 50% with UPF compared to the unprocessed diet. Energy intake rates showed, however, a wide range within each NOVA group demonstrating that the degree of processing is not the exclusive criterium for inducing a higher energy intake rate [28].

## Food texture and matrix of UPF may lead to overfeeding

Food texture and matrix are known to also play a role in eating rate because softer/thinner foods can be consumed more quickly compared to harder/more chewy foods [29]. The satiating value of foods (per kcal) that are eaten quickly is lower compared to foods usually ingested slower [30] leading to consumption of higher quantities of foods that can be ingested more rapidly [31]. In other words, harder foods (e.g., raw vegetables) reduce *ad libitum* energy intake compared to soft foods (e.g., boiled vegetables) [32]. The impact of food texture on satiety was also investigated in a recent meta-analysis confirming an enhanced effect of solid and higher viscous foods on reducing hunger and increasing fullness ratings compared to liquid and low viscous foods [33]. It can therefore be assumed that a softer food texture per se may accelerate eating rate with minor impact on satiation. The effect of food texture in combination with food processing level on eating rate was investigated in a cross-over intervention [34]. Participants with normal-weight consumed *ad libitum* lunch meals at four occasions consisting of “soft minimally processed”, “hard minimally processed”, “soft ultra-processed”, and “hard ultra-processed” components. The results revealed an effect of food texture on eating rate, as hard foods were consumed slower evoking a 26% reduction in energy intake while foods with a soft texture were consumed faster. On the other hand, also the food processing level had an impact on energy intake as the lowest intake was seen with the “hard minimally processed” meal and the most energy was consumed with the “soft ultra-processed” meal with a difference of ~300 kcal [34]. These findings show that UPF can lead to faster ingestion and thus facilitate overconsumption. Comparing the instrumental texture properties from textural profile analysis with eating behaviors of solid foods identified increased springiness and chewiness as the textural food characteristics that contribute to a reduced eating rate [35]. These texture properties could be used to design special “slow UPF” that prevent overconsumption.

The proposed mechanism behind the association between *ad libitum* energy intake and energy density, eating rate or food texture is the oro-sensory exposure (OSE) time during oral processing of food (i.e., mastication and lubrication of the food until swallowing [36]). Increasing the time a food spends in the mouth during oral processing as well as the number of chews required per bite (i.e., harder foods) directly slow energy intake rate and thus reduce *ad libitum* energy intake [31, 32, 37–39]. For example, adding

peach gel particles to yoghurt and thus increasing the OSE due to chewing decreased the eating rate by up to 60% compared to homogenous yoghurt in young and elderly healthy adults without differences in palatability ratings [37]. In another intervention increasing OSE by food composition (chocolate custard with caramel fudge, high OSE instead of chocolate custard with caramel sauce, low OSE), reduced *ad libitum* food intake [38]. The underlying mechanism is the direct effect of OSE to food on satiation by signaling to the brainstem and further to higher cortical regions as described in detail in a recent review [40]. Hence, the sensory stimulation by a prolonged OSE mediates earlier satiation by brain signaling leading to earlier meal termination and thus resulting in a reduced *ad libitum* food intake. On the other hand, peripheral mechanisms like gastrointestinal hormone responses or gastric signaling were less affected by OSE [40].

## High palatability of UPF may lead to overfeeding

In addition to the above-mentioned properties, UPF are also engineered to maximize palatability and therefore consumption [41]. Naturally occurring foods contain one primary nutrient inducing the palatability of the food. In contrast, UPF with maximized palatability, so called “hyper-palatable foods (HPF)”, contain several palatability-inducing nutrients in combinations that are not typically found in nature, e.g., simultaneous high amounts of fat and sugar [42]. HPF have been defined as rich in (i) fat and sodium, (ii) fat and simple sugars, or (iii) carbohydrates and sodium with specific contributions to the energy content of the food [43]. Applying this definition to the Food and Nutrient Database for Dietary Studies, revealed that 62% of foods in the US may be hyper-palatable. It was also shown that rather the food processing, than the food item category is crucial for hyper-palatability. For example, foods that would not intuitively be attributed as HPF, e.g., vegetables cooked in creams or sauces also fall within the definition of HPF [43]. What makes the consumption of HPF problematic is the ability to modulate feeding-related neural systems and food-seeking behavior by inducing reward and thus leading to overconsumption and ultimately to weight gain [44]. In a recent review, the reinforcing natures of HPF showed strong evidence that they may induce behaviors similar to that seen in drug abuse. The highly rewarding experience of HPF-consumption is suggested to be responsible for delayed satiety mechanism which in turn lead to excess energy intake with HPF [45]. HPF have been suggested as potential target substance of food addiction already a decade ago as they share several similarities with addictive drugs [46].

High sodium content of UPF not only effects hyper-palatability but may also contribute to NAFLD. This is supported by the Prevention of Renal and Vascular End-Stage Disease (PREVEND) study that found that a higher sodium intake assessed by 24h-urinary sodium excretion was associated with fatty liver index [47]. The author suggested that insulin resistance (IR) may represent a metabolic intermediate in explaining the relationship of higher sodium intake with NAFLD.

## Low protein content of UPF may lead to overfeeding

Another potential mechanism by which UPF may lead to excess energy intake is the lower protein content seen in UPF compared to less processed foods (9.5% vs. 25.3%) [48]. Calories from protein are more expensive than from fat or carbohydrates, thus the food industry has a financial incentive to replace calories from protein with cheaper alternatives and therefore dilute the protein content of UPF [49]. According to the protein-leverage hypothesis, individuals show a protein-specific appetite trying to maintain a fixed protein intake. If the protein density of the diet consumed is low, the amount of food ingested will be increased to maintain absolute protein intake regardless if this means to increase total energy intake above the needs due to higher intake of fat and carbohydrates [50]. Protein dilution in UPF could therefore lead to excess energy intake when consuming a high proportion of UPF in an attempt to maintain protein intake. In line with the protein-leverage hypothesis, analyzing data from the National Health and Nutrition Examination Survey 2009–2010, it has been shown that with a higher percentage of energy intake from UPF the percent of total energy intake from protein decreased and total energy intake increased while the absolute protein intake remained relatively constant [48]. In the study by Hall et al. [20], it was calculated that protein leverage could potentially have explained up to ~50% of the observed difference of ~500 kcal/day in energy intake between the ultra-processed and unprocessed diet.

In summary, UPF may lead to excess energy intake by a high energy density, a soft texture that promotes a higher eating rate, a low protein content and hyper-palatability [51]. This excess energy intake with UPF may lead to weight gain and ultimately to the development of overweight and obesity and thus also increases the risk for NAFLD. However, not only physiological mechanism may increase UPF consumption, but also the fact that UPF are easily accessible and highly convenient, mostly being ready-to-eat or ready-to-heat products that may promote snacking behavior. They also provide inexpensive calories compared to unprocessed foods and have a long shelf-life [52]. These factors contribute to a regular consumption and a higher proportion of UPF in the diet.

## **Consumption of UPF may lead to NAFLD by low quality of dietary fat (i.e., high intake of saturated FA and a low intake of polyunsaturated FA)**

There is evidence that excessive intake of saturated FA (SFA) with pervasive UPF consumption contributes to the explanation of adverse health outcomes [53, 54]. In population based case-control and prospective studies, Western dietary patterns with a high SFA content are associated with higher risk and prevalence of liver fat and NAFLD (for review see [55]) whereas a plant-based dietary pattern rich in poly-unsaturated FA (PUFA) was associated with lower NAFLD risk [56]. The adverse effect of high dietary intakes of SFA on hepatic fat deposition was independent of energy balance because it was not only observed in hypercaloric [57–59] but also in isocaloric dietary intervention studies [60–62].

Nutritional survey data from different countries revealed that contemporary diets with relatively higher contribution of UPF are characterized by higher contents of SFA [53, 63]. This relationship is not supported by a prospective study in British children in which UPF consumption was associated with lower calculated intake of fat, and particularly saturated fat [64]. UPF are, however, often characterized by a high quantity of fat and, by definition, also by a large proportion of highly processed dietary fats and oil [65, 66]. SFA and mono-unsaturated FA (MUFA) were predominant in fast food and ready-to-eat food items, while PUFA were abundant primarily in seafood UPF meals [65]. SFA was the second major content of FA of all UPF categories with 37.9%, just behind the MUFA (38.6%). SFA content increases stability and thus extends shelf life as compared to unsaturated FA [63].

Palm oil accounts for more than half of all vegetable fats and oils consumed [67]. It has technological and chemical properties that allow a broad use in food processes and it guarantees neutral taste, fragrance and defined texture in the food products [68–70]. Refined palm oil can be fractionated into a liquid olein fraction with better cold stability and a solid stearin fraction with better melting properties [71]. Palm stearin is preferentially utilized as a baking fat because of its stable  $\beta'$ -polymorphic form [72]. The use of palm oil by the food industry has markedly increased in the last few decades [73]. Palm oil and palm kernel oil contain 50% and 85% SFA, respectively [68]. In healthy persons, a single oral dosage of 80–92 g palm oil during a hyperinsulinemic-euglycemic clamp led to a rapid increase in liver fat (+35% 4 h after ingestion of the palm oil) as well as increases in hepatic and peripheral IR compared to vehicle (still water) [74].

Impaired insulin signaling or IR is discussed to be central in the pathogenesis of NAFLD and there is evidence that SFA adversely affect this process [75, 76]. In a cross-over dietary intervention in 16 males with overweight [47.9 years  $\pm$  1.1 years, BMI 27.7 kg/m<sup>2</sup>  $\pm$  0.4 kg/m<sup>2</sup>, as mean  $\pm$  standard error of the mean (SEM)] that compared a high SFA vs. high sugar diet for four weeks each, postprandial glucose and insulin responses were exaggerated in the SFA as compared to the sugar diet indicating impaired insulin metabolism [61]. This was paralleled by an increase in hepatic fat storage with the SFA but not with the sugar diet. Another dietary intervention study in 67 subjects with abdominal obesity (66% women, 50–64 years, 15% with diabetes, 31% using antihypertensive drugs, 16% using lipid-lowering drugs) comparing high SFA vs. high PUFA intakes found increased fasting insulin concentrations with the SFA diet indicating impaired insulin sensitivity and liver fat enrichment as compared with a diet rich in PUFA [60]. Three-week overfeeding with a SFA rich diet in 38 individuals with overweight (21 women and 17 men, 48 years  $\pm$  2 years, BMI 31 kg/m<sup>2</sup>  $\pm$  1 kg/m<sup>2</sup>, liver fat 4.7%  $\pm$  0.9%) increased hepatic fat content by stimulating adipose tissue lipolysis and inducing IR which may be mediated by plasma ceramides that were also increased [57]. Proposed mechanisms of ceramides increasing lipid accumulation are stimulation of

hepatic fat uptake as well as *de novo* lipogenesis [59] but also stimulation of adipose tissue lipolysis and induction of IR [57].

Other pathways contributing to liver fat accumulation with a diet high in SFA and low in unsaturated FA involve an imbalance between hepatic lipid uptake, *de-novo* lipogenesis and removal [77]. Interestingly, a diet high in SFA led to increased liver fat without increasing hepatic *de-novo* lipogenesis [61]. This was confirmed in a two-week cross-over dietary intervention with constant macronutrient relation that showed an increase in hepatic fat content with a high SFA and high glycemic index diet without affecting *de-novo* lipogenesis or FA oxidation [62]. By contrast, eight weeks of an isoenergetic high MUFA diet (40% carbohydrate, 42% fat whereof 28% MUFA) led to reduced liver fat in a controlled randomized study in type 2 diabetic patients [78] presumably due to high postprandial hepatic FA oxidation [79]. Similarly, eight week supplementation of omega-3 FA was shown to decrease liver fat by increasing dietary FA oxidation and downregulating hepatic *de-novo* lipogenesis [80]. Enrichment of food products such as bread, milk products, margarine and eggs with long-chain n-3 PUFA has, however, become less popular due to the largely disappointing results from studies investigating the use of n-3 FA for the primary and secondary prevention of cardiovascular disease [81].

### **Consumption of UPF may lead to NAFLD by low quality of carbohydrates (i.e., high glycemic load and high fructose content)**

Since insulin is known to stimulate sterol regulatory element binding protein 1c (SREBP1c) and thus lipogenesis [77], an insulinogenic diet can promote development of NAFLD. Minimally processed foods led to a lower postprandial glycemia and thus a lower insulin demand compared to UPF and an increased level of food processing was associated with higher glycemic responses [82, 83]. The underlying mechanism includes that food processing with high pressures and temperatures or intense refining leads to deconstruction of the food matrix structure, and thus to a higher degree of starch gelatinization that increases postprandial glycemia. In contrast, reformulation of UPF in a low-carbohydrate, high-protein version can produce a significantly lower glycemic response reducing the risk to develop NAFLD whilst maintaining the high palatability valued by consumers [84].

In line with this, in a cross-over study including 13 adults with overweight and obesity (67 years  $\pm$  6 years, BMI 29.5 kg/m<sup>2</sup>  $\pm$  1.9 kg/m<sup>2</sup>), simultaneous improvements in carbohydrate and fat quality by reducing glycemic index (60 vs. 36) and replacing SFA with PUFA (15% SFA vs. 5% SFA) for just two weeks led to drastic improvements on postprandial glycemia and liver fat content (-28%) even at unchanged energy content and percentage of total fat and carbohydrate [62].

Although low in glycemic index, excessive intake of fructose by sugar sweetened beverages (SSB) and possibly also by other processed foods has been linked to increased risk of developing NAFLD [85]. In a systematic review and meta-analysis of controlled trials on the effect of fructose-containing sugars from different food sources on NAFLD markers it was shown that total fructose-containing sugars from SSB but not from other food sources increased liver fat in addition trials and decreased aspartate transaminase (AST) in subtraction trials with no effect on any outcome in substitution or *ad libitum* trials [86]. When consumed in positive energy balance, fructose-sweetened beverages promote hepatic *de novo* lipogenesis, liver fat accumulation, dyslipidemia, and IR compared to glucose-sweetened beverages [87, 88].

### **Consumption of UPF may lead to NAFLD by dietary components promoting dysbiosis and leaky gut**

Dysfunction of the gut-liver axis is a generally accepted mechanism for the progression of NAFLD [89] that is substantially influenced by the diet. Intestinal factors of increased portal endotoxin levels include impaired barrier function (leaky gut syndrome) and dysbiosis. Exposure of the liver to endotoxins from intestinal bacteria, antigens, pathogens and other proinflammatory substances leads to an increase of inflammatory cytokines as observed in animal models and humans which are expected to aggravate the progression of NAFLD early on [89].

Food processing and formulation may contribute to dysbiosis and impaired barrier function through various mechanisms. Non-caloric sweeteners (e.g., saccharin, sucralose, aspartame) that improve the taste and texture of food as well as advanced glycation endproducts (AGE) generated by thermal processes from amino acids and sugars can alter gut microbial metabolism [90–93]. Emulsifiers (i.e., lecithin, mono- and diglycerides of FA, guar gum, xanthan gum, carrageenan, celluloses, and polysorbates) that are common in bakery, dairy, beverages, ice cream, sauces, chocolate, and other foods are suspected to impair the diversity of gut microbiota (decrease in anti-inflammatory genera like *Akkermansia* and promotion of proinflammatory genera like *Escherichia*, *Roseburia*, *Bradyrhizobium*, and *Turicibacter*) [94]. In addition, emulsifiers may stimulate inflammatory pathways [95], impair the function of the mucus layer and the tight junctions [96]. High consumption of UPF is therefore suspected to play a causal role in several diseases associated with dysbiosis and impaired intestinal permeability, such as cardiovascular and microvascular diseases like chronic kidney disease, obesity, type 2 diabetes, and cancer [97–99].

Promising strategies to prevent or treat dysbiosis and leaky gut syndrome are enrichment of processed foods with phytochemicals like polyphenols and prebiotics/dietary fibers, like inulin, fructo-oligosaccharides, galacto-oligosaccharides, isomalto-oligosaccharides, xylo-oligosaccharides, arabino-xylo-oligosaccharides, pectic-oligosaccharides, or resistant starch. Per definition, prebiotics are “selective fermented ingredients that allow specific changes, both in the composition and/or activity in the gastrointestinal microflora that confer benefits upon host well-being and health” [100]. It is expected that prebiotics promote the abundance of beneficial bacteria, like *Bifidobacterium*, *Lactobacillus*, and *Faecalibacterium prausnitzii* [101–103] that improve barrier function and gut health by increasing the production of short-chain FAs and decreasing endotoxins levels [104].

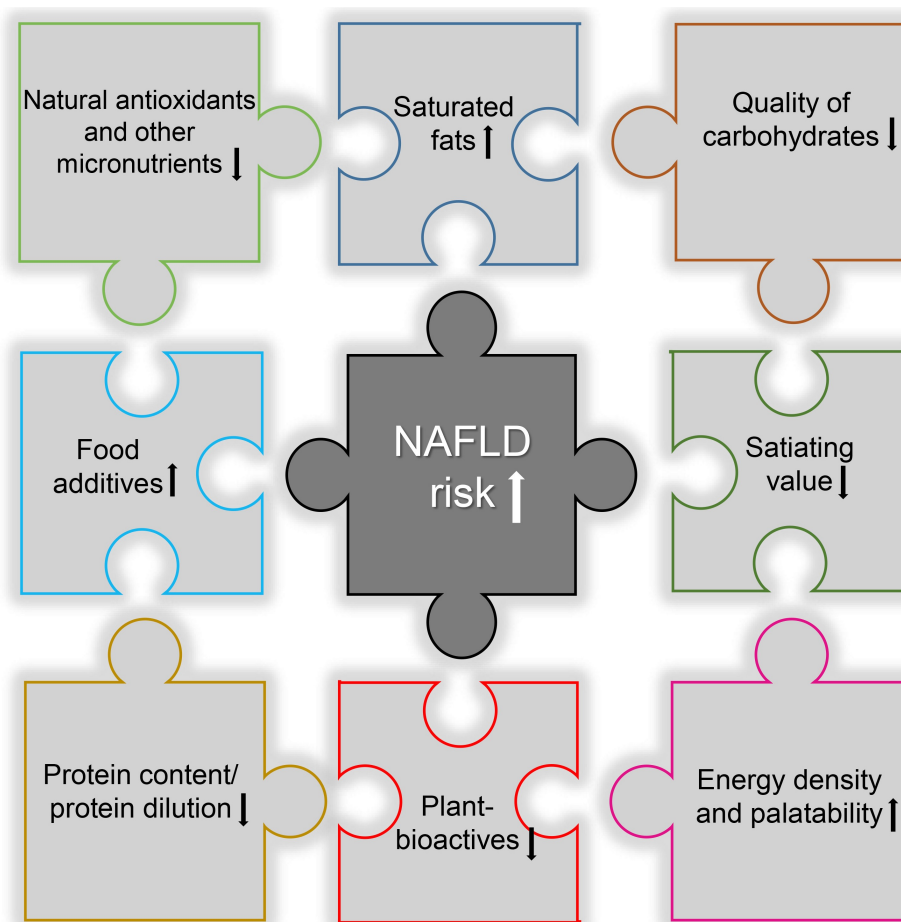
UPF are generally low in phytochemicals and enrichment of these food products with polyphenols like quercetin, epigallocatechin gallate, curcumin as well as carotenoids, phytosterols, and others may help to increase the abundance of beneficial bacteria and reduce intestinal permeability or exhibit antioxidant and anti-inflammatory properties (for a review see [105]) that contribute to the prevention and treatment of NAFLD.

In addition, enrichment of UPF with so called plant lipotropes like choline, betaine, myo-inositol, methionine and carnitine could facilitate the removal of fat from the liver [i.e., by promoting phospholipid synthesis for very low density lipoprotein (VLDL) secretion], reduce hepatic lipid synthesis and increase FA oxidation [106, 107]. However, evidence from human intervention studies that confirm the capacity of these components to prevent or treat individuals with NAFLD is needed.

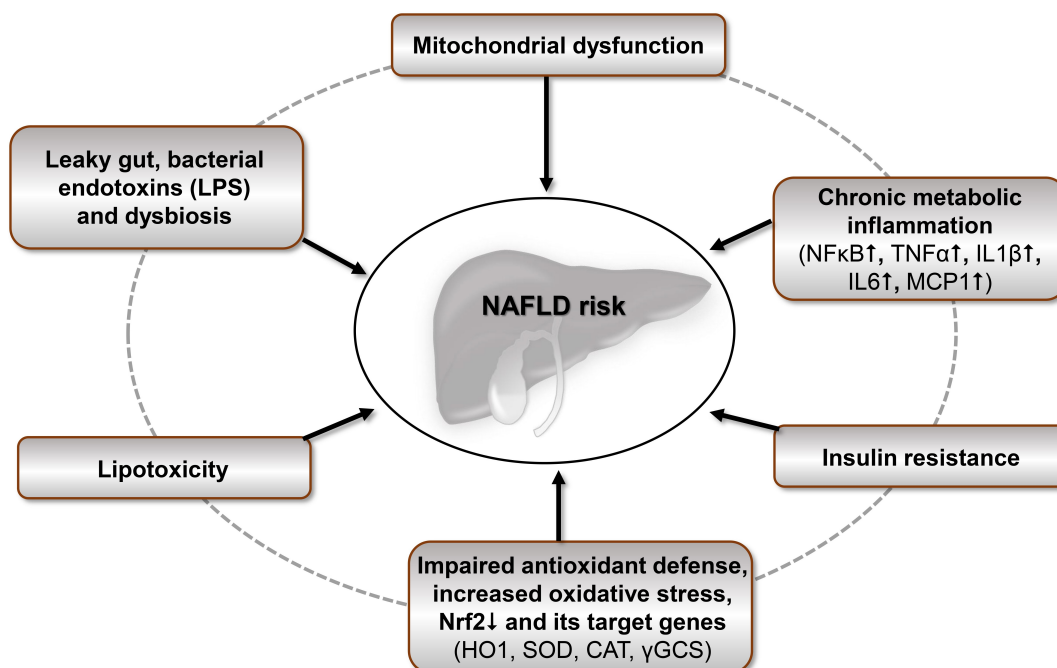
## Proposed molecular mechanisms

UPF are often high in sugar, low in dietary fiber, natural antioxidants, plant bioactives and micronutrients. At the same time UPF may contain substantial amounts of saturated fats (Figure 1). Collectively this may result in chronic low-grade metabolic inflammation [108] as observed by an increased nuclear factor  $\kappa$ -light-chain-enhancer of activated B cells (NF $\kappa$ B) activity in laboratory rodents [109]. The transcription factor NF $\kappa$ B regulates the production of proinflammatory cytokines such as tumor necrosis factor alpha (TNF- $\alpha$ ), interleukin 1 beta (IL1 $\beta$ ), interleukin 6 (IL6) and chemoattractants including monocyte chemoattractant protein 1 (MCP1) amongst others (Figure 2). Similarly, a positive association between UPF consumption and C-reactive protein levels (as a proxy of chronic inflammation) as well as other biomarkers of inflammation (e.g., IL1 $\beta$ , IL6, MCP1) has been observed in several studies in children and adolescents [110–112].

Furthermore, fiber-poor UPF as part of the so-called Western type diet can result in dysbiosis and impaired gut integrity. A leaky gut increases the concentration of bacterial endotoxins (e.g., LPS) known to trigger the above mentioned proinflammatory signal transduction pathways [113] of the gut-liver-axis partly via toll like receptor signaling [114].



**Figure 1.** Determinants of UPFs in the context of NAFLD risk. ↑: High; ↓: low



**Figure 2.** Underlying mechanisms, by which UPF increase NAFLD risk. CAT: catalase; HO1: heme oxygenase 1; LPS: lipopolysaccharide; Nrf2: nuclear factor erythroid 2-related factor 2; SOD: superoxide dismutase; γGCS: γ-glutamylcysteine-synthetase. ↑: High/increase; ↓: low/decrease

The low dietary supply of natural antioxidants (e.g., vitamin C, vitamin E, carotenoids), plant bioactives (e.g., polyphenols, isothiocyanate derived glucosinolates), micronutrients (e.g., zinc, selenium) via UPF affects the cellular oxidant/antioxidant homeostasis. This in turn may induce cellular redox stress which interferes with Nrf2 signaling. The transcription factor Nrf2 orchestrates the expression of genes encoding



important antioxidant and stress proteins [115–117] such as HO1, SOD, CAT and  $\gamma$ GCS—the latter is the rate regulating enzyme of endogenous glutathione (GSH) synthesis a potent cytosolic antioxidant.

Redox stress as well as an overflow of FFA due to excessive UPF consumption leads to hepatic lipotoxicity, mitochondrial dysfunction, beta cell death and ultimately to IR [118] which contribute to the pathogenesis of hepatic injury and NAFLD as summarized in Figure 2. Furthermore, it is plausible to assume that UPF affects autophagy, proteasomal activity and endoplasmic reticulum stress which warrants further systematic investigations.

## Conclusion

Overall, literature data clearly indicate that UPF increase NAFLD risk. The nutritional quality of processed food could be improved due to enrichment with fiber, prebiotics, phytochemicals and protein with a concurrent reduction in glycemic load, energy density, SFA, fructose and food additives. Future studies are needed to examine the interactive and protective effects of such modifications in the composition of UPF on prevention and treatment of NAFLD. Furthermore, complimentary studies in cultured cells and laboratory rodents are needed to systematically elucidate the underlying molecular, cellular and physiological mechanisms by which UPF and its constituents affect NAFLD risk. In addition, prospective as well as sufficiently powered long-term human intervention studies addressing the role of UPF in the context of NAFLD and other chronic diseases should be conducted in the future. Finally, the identification of robust biomarkers which are indicative for UPF consumption in humans is crucial to better pinpoint UPF consumption in the population.

## Abbreviations

BMI: body mass index

FA: fatty acids

HPF: hyper-palatable foods

IL1 $\beta$ : interleukin 1 beta

IL6: interleukin 6

IR: insulin resistance

MCP1: monocyte chemoattractant protein 1

MUFA: mono-unsaturated fatty acid

NAFLD: non-alcoholic fatty liver disease

Nrf2: nuclear factor erythroid 2-related factor 2

OSE: oro-sensory exposure

PUFA: poly-unsaturated fatty acids

SFA: saturated fatty acids

UPF: ultra-processed food

## Declarations

### Author contributions

FAH and JE: Writing—original draft, Writing—review & editing. GR: Writing—original draft, Writing—review & editing, Visualization. ABW: Writing—original draft, Writing—review & editing, Conceptualization, Supervision. All authors read and approved the submitted version.

### Conflicts of interest

The authors declare that they have no conflicts of interest.

### **Ethical approval**

Not applicable.

### **Consent to participate**

Not applicable.

### **Consent to publication**

Not applicable.

### **Availability of data and materials**

Not applicable.

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## **References**

1. Monteiro CA, Cannon G, Moubarac JC, Levy RB, Louzada MLC, Jaime PC. The UN Decade of Nutrition, the NOVA food classification and the trouble with ultra-processing. *Public Health Nutr.* 2018;21:5–17.
2. Li J, Zou B, Yeo YH, Feng Y, Xie X, Lee DH, et al. Prevalence, incidence, and outcome of non-alcoholic fatty liver disease in Asia, 1999–2019: a systematic review and meta-analysis. *Lancet Gastroenterol Hepatol.* 2019;4:389–98.
3. Li L, Liu DW, Yan HY, Wang ZY, Zhao SH, Wang B. Obesity is an independent risk factor for non-alcoholic fatty liver disease: evidence from a meta-analysis of 21 cohort studies. *Obes Rev.* 2016;17:510–9.
4. The Global BMI Mortality Collaboration, Di Angelantonio E, Bhupathiraju SN, Wormser D, Gao P, Kaptoge S, de Gonzalez AB, et al. Body-mass index and all-cause mortality: individual-participant-data meta-analysis of 239 prospective studies in four continents. *Lancet.* 2016;388:776–86.
5. Donnelly KL, Smith CI, Schwarzenberg SJ, Jessurun J, Boldt MD, Parks EJ. Sources of fatty acids stored in liver and secreted via lipoproteins in patients with nonalcoholic fatty liver disease. *J Clin Invest.* 2005;115:1343–51.
6. Nielsen S, Guo Z, Johnson CM, Hensrud DD, Jensen MD. Splanchnic lipolysis in human obesity. *J Clin Invest.* 2004;113:1582–8.
7. Beslay M, Srouf B, Méjean C, Allès B, Fiolet T, Debras C, et al. Ultra-processed food intake in association with BMI change and risk of overweight and obesity: a prospective analysis of the French NutriNet-Santé cohort. *PLOS Med.* 2020;17:e1003256.
8. Canhada SL, Luft VC, Giatti L, Duncan BB, Chor D, Fonseca MJMD, et al. Ultra-processed foods, incident overweight and obesity, and longitudinal changes in weight and waist circumference: the Brazilian Longitudinal Study of Adult Health (ELSA-Brasil). *Public Health Nutr.* 2020;23:1076–86.
9. Cordova R, Kliemann N, Huybrechts I, Rauber F, Vamos EP, Levy RB, et al. Consumption of ultra-processed foods associated with weight gain and obesity in adults: a multi-national cohort study. *Clin Nutr.* 2021;40:5079–88.
10. Mendonça RD, Pimenta AM, Gea A, de la Fuente-Arrillaga C, Martinez-Gonzalez MA, Lopes ACS, et al. Ultraprocessed food consumption and risk of overweight and obesity: the University of Navarra Follow-Up (SUN) cohort study. *Am J Clin Nutr.* 2016;104:1433–40.

11. Nardocci M, Leclerc BS, Louzada ML, Monteiro CA, Batal M, Moubarac JC. Consumption of ultra-processed foods and obesity in Canada. *Can J Public Heal.* 2019;110:4–14.
12. Rauber F, Chang K, Vamos EP, da Costa Louzada ML, Monteiro CA, Millett C, et al. Ultra-processed food consumption and risk of obesity: a prospective cohort study of UK Biobank. *Eur J Nutr.* 2021;60:2169–80.
13. Askari M, Heshmati J, Shahinfar H, Tripathi N, Daneshzad E. Ultra-processed food and the risk of overweight and obesity: a systematic review and meta-analysis of observational studies. *Int J Obes.* 2020;44:2080–91.
14. Konieczna J, Morey M, Abete I, Bes-Rastrollo M, Ruiz-Canela M, Vioque J, et al. Contribution of ultra-processed foods in visceral fat deposition and other adiposity indicators: prospective analysis nested in the PREDIMED-Plus trial. *Clin Nutr.* 2021;40:4290–300.
15. Sandoval-Insausti H, Jiménez-Onsurbe M, Donat-Vargas C, Rey-García J, Banegas JR, Rodríguez-Artalejo F, et al. Ultra-processed food consumption is associated with abdominal obesity: a prospective cohort study in older adults. *Nutrients.* 2020;12:2368.
16. Ivancovsky-Wajcman D, Fliss-Isakov N, Webb M, Bentov I, Shibolet O, Kariv R, et al. Ultra-processed food is associated with features of metabolic syndrome and non-alcoholic fatty liver disease. *Liver Int.* 2021;41:2635–45.
17. Konieczna J, Fiol M, Colom A, Martínez-González MÁ, Salas-Salvadó J, Corella D, et al. Does consumption of ultra-processed foods matter for liver health? Prospective analysis among older adults with metabolic syndrome. *Nutrients.* 2022;14:4142.
18. Zhang S, Gan S, Zhang Q, Liu L, Meng G, Yao Z, et al. Ultra-processed food consumption and the risk of non-alcoholic fatty liver disease in the Tianjin Chronic Low-grade Systemic Inflammation and Health Cohort Study. *Int J Epidemiol.* 2022;51:237–49.
19. Hall KD, Ayuketah A, Brychta R, Cai H, Cassimatis T, Chen KY, et al. Ultra-processed diets cause excess calorie intake and weight gain: an inpatient randomized controlled trial of Ad libitum food intake. *Cell Metab.* 2019;30:67–77.e3.
20. Rouhani MH, Haghghatdoost F, Surkan PJ, Azadbakht L. Associations between dietary energy density and obesity: a systematic review and meta-analysis of observational studies. *Nutrition.* 2016;32:1037–47.
21. Bell EA, Castellanos VH, Pelkman CL, Thorwart ML, Rolls BJ. Energy density of foods affects energy intake in normal-weight women. *Am J Clin Nutr.* 1998;67:412–20.
22. Rolls BJ. The relationship between dietary energy density and energy intake. *Physiol Behav.* 2009;97:609–15.
23. Blundell JE, Macdiarmid JI. Passive overconsumption fat intake and short-term energy balance. *Ann N Y Acad Sci.* 1997;827:392–407.
24. Robinson E, Khuttan M, McFarland-Lesser I, Patel Z, Jones A. Calorie reformulation: a systematic review and meta-analysis examining the effect of manipulating food energy density on daily energy intake. *Int J Behav Nutr Phys Act.* 2022;19:48.
25. Flynn AN, Hall KD, Courville AB, Rogers PJ, Brunstrom JM. Time to revisit the passive overconsumption hypothesis? Humans show sensitivity to calories in energy-rich meals. *Am J Clin Nutr.* 2022;116:581–8.
26. Andrade AM, Kresge DL, Teixeira PJ, Baptista F, Melanson KJ. Does eating slowly influence appetite and energy intake when water intake is controlled? *Int J Behav Nutr Phys Act.* 2012;9:135.
27. Melanson KJ. Food intake regulation in body weight management: a primer. *Nutr Today.* 2004;39:203–13.
28. Forde CG, Mars M, de Graaf K. Ultra-processing or oral processing? A role for energy density and eating rate in moderating energy intake from processed foods. *Curr Dev Nutr.* 2020;4:nzaa019.

29. Forde CG, Bolhuis D. Interrelations between food form, texture, and matrix influence energy intake and metabolic responses. *Curr Nutr Rep.* 2022;11:124–32.
30. de Graaf C, Kok FJ. Slow food, fast food and the control of food intake. *Nat Rev Endocrinol.* 2010;6:290–3.
31. Zijlstra N, de Wijk RA, Mars M, Stafleu A, de Graaf C. Effect of bite size and oral processing time of a semisolid food on satiety. *Am J Clin Nutr.* 2009;90:269–75.
32. Bolhuis DP, Forde CG, Cheng Y, Xu H, Martin N, de Graaf C. Slow food: sustained impact of harder foods on the reduction in energy intake over the course of the day. *PLoS One.* 2014;9:e93370.
33. Stribițcaia E, Evans CEL, Gibbons C, Blundell J, Sarkar A. Food texture influences on satiety: systematic review and meta-analysis. *Sci Rep.* 2020;10:12929.
34. Teo PS, Lim AJ, Goh AT, Janani R, Michelle Choy JY, McCrickerd K, et al. Texture-based differences in eating rate influence energy intake for minimally processed and ultra-processed meals. *Am J Clin Nutr.* 2022;116:244–54.
35. Wee MSM, Goh AT, Stieger M, Forde CG. Correlation of instrumental texture properties from textural profile analysis (TPA) with eating behaviours and macronutrient composition for a wide range of solid foods. *Food Funct.* 2018;9:5301–12.
36. de Graaf C. Texture and satiety: the role of oro-sensory exposure time. *Physiol Behav.* 2012;107:496–501.
37. Aguayo-Mendoza M, Santagiuliana M, Ong X, Piqueras-Fiszman B, Scholten E, Stieger M. How addition of peach gel particles to yogurt affects oral behavior, sensory perception and liking of consumers differing in age. *Food Res Int.* 2020;134:109213.
38. Lasschuijt MP, Mars M, de Graaf C, Smeets PAM. How oro-sensory exposure and eating rate affect satiety and associated endocrine responses—a randomized trial. *Am J Clin Nutr.* 2020;111:1137–49.
39. McCrickerd K, Lim CM, Leong C, Chia EM, Forde CG. Texture-based differences in eating rate reduce the impact of increased energy density and large portions on meal size in adults. *J Nutr.* 2017;147:1208–17.
40. Lasschuijt MP, de Graaf K, Mars M. Effects of oro-sensory exposure on satiety and underlying neurophysiological mechanisms—what do we know so far? *Nutrients.* 2021;13:1391.
41. Moss M. *Salt sugar fat: How the food giants hooked us.* 1st ed. New York: Random House; 2013.
42. Balter V, Braga J, Télouk P, Thackeray JF. Evidence for dietary change but not landscape use in South African early hominins. *Nature.* 2012;489:558–60.
43. Fazzino TL, Rohde K, Sullivan DK. Hyper-palatable foods: development of a quantitative definition and application to the US food system database. *Obesity (Silver Spring).* 2019;27:1761–8.
44. Leigh SJ, Lee F, Morris MJ. Hyperpalatability and the generation of obesity: roles of environment, stress exposure and individual difference. *Curr Obes Rep.* 2018;7:6–18.
45. Fazzino TL. The reinforcing natures of hyper-palatable foods: behavioral evidence for their reinforcing properties and the role of the US food industry in promoting their availability. *Curr Addict Rep.* 2022;9:298–306.
46. Gearhardt AN, Davis C, Kuschner R, Brownell KD. The addiction potential of hyperpalatable foods. *Curr Drug Abuse Rev.* 2011;4:140–5.
47. van den Berg EH, Gruppen EG, Blokzijl H, Bakker SJL, Dullaart RPF. Higher sodium intake assessed by 24 hour urinary sodium excretion is associated with non-alcoholic fatty liver disease: the PREVENT cohort study. *J Clin Med.* 2019;8:2157.
48. Martínez Steele E, Raubenheimer D, Simpson SJ, Baraldi LG, Monteiro CA. Ultra-processed foods, protein leverage and energy intake in the USA. *Public Health Nutr.* 2018;21:114–24.
49. Simpson SJ, Raubenheimer D. Perspective: tricks of the trade. *Nature.* 2014;508:S66.
50. Simpson SJ, Raubenheimer D. Obesity: the protein leverage hypothesis. *Obes Rev.* 2005;6:133–42.

51. Fazzino TL, Courville AB, Guo J, Hall KD. Ad libitum meal energy intake is positively influenced by energy density, eating rate and hyper-palatable food across four dietary patterns. *Nat Food*. 2023;4:144–7.
52. Weaver CM, Dwyer J, Fulgoni VL, King JC, Leveille GA, MacDonald RS, et al. Processed foods: contributions to nutrition. *Am J Clin Nutr*. 2014;99:1525–42.
53. Martini D, Godos J, Bonaccio M, Vitaglione P, Grosso G. Ultra-processed foods and nutritional dietary profile: a meta-analysis of nationally representative samples. *Nutrients*. 2021;13:3390.
54. Monteiro CA, Moubarac JC, Levy RB, Canella DS, Louzada MLDC, Cannon G. Household availability of ultra-processed foods and obesity in nineteen European countries. *Public Health Nutr*. 2018;21:18–26.
55. Berná G, Romero-Gomez M. The role of nutrition in non-alcoholic fatty liver disease: pathophysiology and management. *Liver Int*. 2020;40:102–8.
56. Tian A, Sun Z, Zhang M, Li J, Pan X, Chen P. Associations between dietary fatty acid patterns and non-alcoholic fatty liver disease in typical dietary population: a UK biobank study. *Front Nutr*. 2023;10:1117626.
57. Luukkonen PK, Sädevirta S, Zhou Y, Kayser B, Ali A, Ahonen L, et al. Saturated fat is more metabolically harmful for the human liver than unsaturated fat or simple sugars. *Diabetes Care*. 2018;41:1732–9.
58. Rosqvist F, Iggman D, Kullberg J, Cedernaes J, Johansson HE, Larsson A, et al. Overfeeding polyunsaturated and saturated fat causes distinct effects on liver and visceral fat accumulation in humans. *Diabetes*. 2014;63:2356–68.
59. Rosqvist F, Kullberg J, Ståhlman M, Cedernaes J, Heurling K, Johansson HE, et al. Overeating saturated fat promotes fatty liver and ceramides compared with polyunsaturated fat: a randomized trial. *J Clin Endocrinol Metab*. 2019;104:6207–19.
60. Bjermo H, Iggman D, Kullberg J, Dahlman I, Johansson L, Persson L, et al. Effects of n-6 PUFAs compared with SFAs on liver fat, lipoproteins, and inflammation in abdominal obesity: a randomized controlled trial. *Am J Clin Nutr*. 2012;95:1003–12.
61. Parry SA, Rosqvist F, Mozes FE, Cornfield T, Hutchinson M, Piche ME, et al. Intrahepatic fat and postprandial glycemia increase after consumption of a diet enriched in saturated fat compared with free sugars. *Diabetes Care*. 2020;43:1134–41.
62. Basset-Sagarminaga J, Roumans KHM, Havekes B, Mensink RP, Peters HPF, Zock PL, et al. Replacing foods with a high-glycemic index and high in saturated fat by alternatives with a low glycemic index and low saturated fat reduces hepatic fat, even in isocaloric and macronutrient matched conditions. *Nutrients*. 2023;15:735.
63. Martínez Steele E, Batis C, Cediel G, Louzada MLDC, Khandpur N, Machado P, et al. The burden of excessive saturated fatty acid intake attributed to ultra-processed food consumption: a study conducted with nationally representative cross-sectional studies from eight countries. *J Nutr Sci*. 2021;10:e43.
64. Handakas E, Chang K, Khandpur N, Vamos EP, Millett C, Sassi F, et al. Metabolic profiles of ultra-processed food consumption and their role in obesity risk in British children. *Clin Nutr*. 2022;41:2537–48.
65. Maldonado-Pereira L, Barnaba C, de Los Campos G, Medina-Meza IG. Evaluation of the nutritional quality of ultra-processed foods (ready to eat + fast food): fatty acids, sugar, and sodium. *J Food Sci*. 2022;87:3659–76.
66. Monteiro CA, Cannon G, Levy RB, Moubarac JC, Louzada ML, Rauber F, et al. Ultra-processed foods: What they are and how to identify them. *Public Health Nutr*. 2019;22:936–41.

67. Gonzalez-Diaz A, Pataquiva-Mateus A, García-Núñez JA. Recovery of palm phytonutrients as a potential market for the by-products generated by palm oil mills and refineries—a review. *Food Biosci.* 2021;41:100916.
68. Mancini A, Imperlini E, Nigro E, Montagnese C, Daniele A, Orrù S, et al. Biological and nutritional properties of palm oil and palmitic acid: effects on health. *Molecules.* 2015;20:17339–61.
69. Mba OI, Dumont MJ, Ngadi M. Palm oil: processing, characterization and utilization in the food industry – a review. *Food Biosci.* 2015;10:26–41.
70. Sulaiman NS, Sintang MD, Mantihal S, Zaini HM, Munsu E, Mamat H, et al. Balancing functional and health benefits of food products formulated with palm oil as oil sources. *Heliyon.* 2022;8:e11041.
71. Hishamuddin E, Nagy ZK, Stapley AGF. Thermodynamic analysis of the isothermal fractionation of palm oil using a novel method for entrainment correction. *J Food Eng.* 2020;273:109806.
72. Dian NLHM, Hamid RA, Kanagaratnam S, Isa WRA, Hassim NAM, Ismail NH, et al. Palm oil and palm kernel oil: versatile ingredients for food applications. *J Oil Palm Res.* 2017;29:487–511.
73. Sehgal S, Sharma V. Palm/palm kernel (*Elaeis guineensis*). In: Tanwar B, Goyal A, editors. *Oilseeds: health attributes and food applications.* Singapore: Springer; 2021. pp. 145–61.
74. Hernández EÁ, Kahl S, Seelig A, Begovatz P, Irmeler M, Kupriyanova Y, et al. Acute dietary fat intake initiates alterations in energy metabolism and insulin resistance. *J Clin Invest.* 2017;127:695–708.
75. Delarue J, Magnan C. Free fatty acids and insulin resistance. *Curr Opin Clin Nutr Metab Care.* 2007;10:142–8.
76. Chen Z, Tian R, She Z, Cai J, Li H. Role of oxidative stress in the pathogenesis of nonalcoholic fatty liver disease. *Free Radic Biol Med.* 2020;152:116–41.
77. Kawano Y, Cohen DE. Mechanisms of hepatic triglyceride accumulation in non-alcoholic fatty liver disease. *J Gastroenterol.* 2013;48:434–41.
78. Bozzetto L, Prinster A, Annuzzi G, Costagliola L, Mangione A, Vitelli A, et al. Liver fat is reduced by an isoenergetic MUFA diet in a controlled randomized study in type 2 diabetic patients. *Diabetes Care.* 2012;35:1429–35.
79. Bozzetto L, Costabile G, Luongo D, Naviglio D, Cicala V, Piantadosi C, et al. Reduction in liver fat by dietary MUFA in type 2 diabetes is helped by enhanced hepatic fat oxidation. *Diabetologia.* 2016;59:2697–701.
80. Green CJ, Pramfalk C, Charlton CA, Gunn PJ, Cornfield T, Pavlides M, et al. Hepatic *de novo* lipogenesis is suppressed and fat oxidation is increased by omega-3 fatty acids at the expense of glucose metabolism. *BMJ Open Diabetes Res Care.* 2020;8:e000871.
81. Abdelhamid AS, Brown TJ, Brainard JS, Biswas P, Thorpe GC, Moore HJ, et al. Omega-3 fatty acids for the primary and secondary prevention of cardiovascular disease. *Cochrane Database Syst Rev.* 2020;3:CD003177.
82. Fardet A. Minimally processed foods are more satiating and less hyperglycemic than ultra-processed foods: a preliminary study with 98 ready-to-eat foods. *Food Funct.* 2016;7:2338–46.
83. Fardet A, Méjean C, Labouré H, Andreeva VA, Feron G. The degree of processing of foods which are most widely consumed by the French elderly population is associated with satiety and glycemic potentials and nutrient profiles. *Food Funct.* 2017;8:651–8.
84. Lodi A, Karsten B, Bosco G, Gómez-López M, Brandão PP, Bianco A, et al. The effects of different high-protein low-carbohydrates proprietary foods on blood sugar in healthy subjects. *J Med Food.* 2016;19:1085–95.
85. Basaranoglu M, Basaranoglu G, Bugianesi E. Carbohydrate intake and nonalcoholic fatty liver disease: fructose as a weapon of mass destruction. *Hepatobiliary Surg Nutr.* 2015;4:109–16.
86. Lee D, Chiavaroli L, Ayoub-Charette S, Khan TA, Zurbau A, Au-Yeung F, et al. Important food sources of fructose-containing sugars and non-alcoholic fatty liver disease: a systematic review and meta-analysis of controlled trials. *Nutrients.* 2022;14:2846.

87. Stanhope KL, Schwarz JM, Keim NL, Griffen SC, Bremer AA, Graham JL, et al. Consuming fructose-sweetened, not glucose-sweetened, beverages increases visceral adiposity and lipids and decreases insulin sensitivity in overweight/obese humans. *J Clin Invest*. 2009;119:1322–34.
88. Schwarz JM, Noworolski SM, Wen MJ, Dyachenko A, Prior JL, Weinberg ME, et al. Effect of a high-fructose weight-maintaining diet on lipogenesis and liver fat. *J Clin Endocrinol Metab*. 2015;100:2434–42.
89. Kessoku T, Kobayashi T, Tanaka K, Yamamoto A, Takahashi K, Iwaki M, et al. The role of leaky gut in nonalcoholic fatty liver disease: a novel therapeutic target. *Int J Mol Sci*. 2021;22:8161.
90. Suez J, Cohen Y, Valdés-Mas R, Mor U, Dori-Bachash M, Federici S, et al. Personalized microbiome-driven effects of non-nutritive sweeteners on human glucose tolerance. *Cell*. 2022;185:3307–28.e19.
91. Spencer M, Gupta A, Dam LV, Shannon C, Menees S, Chey WD. Artificial sweeteners: a systematic review and primer for gastroenterologists. *J Neurogastroenterol Motil*. 2016;22:168–80.
92. ALjahdali N, Carbonero F. Impact of Maillard reaction products on nutrition and health: current knowledge and need to understand their fate in the human digestive system. *Crit Rev Food Sci Nutr*. 2019;59:474–87.
93. Hellwig M, Henle T. Baking, ageing, diabetes: a short history of the Maillard reaction. *Angew Chem Int Ed Engl*. 2014;53:10316–29.
94. Jiang Z, Zhao M, Zhang H, Li Y, Liu M, Feng F. Antimicrobial emulsifier–glycerol monolaurate induces metabolic syndrome, gut microbiota dysbiosis, and systemic low-grade inflammation in low-fat diet fed mice. *Mol Nutr Food Res*. 2018;62:1700547.
95. Bancil AS, Sandall AM, Rossi M, Chassaing B, Lindsay JO, Whelan K. Food additive emulsifiers and their impact on gut microbiome, permeability, and inflammation: mechanistic insights in inflammatory bowel disease. *J Crohn’s Colitis*. 2021;15:1068–79.
96. Csáki KF. Synthetic surfactant food additives can cause intestinal barrier dysfunction. *Med Hypotheses*. 2011;76:676–81.
97. Fiolet T, Srour B, Sellem L, Kesse-Guyot E, Allès B, Méjean C, et al. Consumption of ultra-processed foods and cancer risk: results from NutriNet-Santé prospective cohort. *BMJ*. 2018;360:k322.
98. Rico-Campà A, Martínez-González MA, Alvarez-Alvarez I, Mendonça RD, de la Fuente-Arrillaga C, Gómez-Donoso C, et al. Association between consumption of ultra-processed foods and all cause mortality: SUN prospective cohort study. *BMJ*. 2019;365:l1949.
99. Snelson M, Tan SM, Clarke RE, de Pasquale C, Thallas-Bonke V, Nguyen TV, et al. Processed foods drive intestinal barrier permeability and microvascular diseases. *Sci Adv*. 2021;7:eabe4841.
100. Gibson GR, Probert HM, Loo JV, Rastall RA, Roberfroid MB. Dietary modulation of the human colonic microbiota: updating the concept of prebiotics. *Nutr Res Rev*. 2004;17:259–75.
101. Marasco G, Cirotta GG, Rossini B, Lungaro L, Di Biase AR, Colecchia A, et al. Probiotics, prebiotics and other dietary supplements for gut microbiota modulation in celiac disease patients. *Nutrients*. 2020;12:2674.
102. Maukonen J, Saarela M. Human gut microbiota: Does diet matter? *Proc Nutr Soc*. 2015;74:23–36.
103. Scott KP, Gratz SW, Sheridan PO, Flint HJ, Duncan SH. The influence of diet on the gut microbiota. *Pharmacol Res*. 2013;69:52–60.
104. Singh RK, Chang HW, Yan D, Lee KM, Ucmak D, Wong K, et al. Influence of diet on the gut microbiome and implications for human health. *J Transl Med*. 2017;15:73.
105. Liang L, Saunders C, Sanossian N. Food, gut barrier dysfunction, and related diseases: a new target for future individualized disease prevention and management. *Food Sci Nutr*. 2023;11:1671–704.
106. Fardet A, Martin JF, Chardigny JM. Lipotropic capacity of raw plant-based foods: a new index that reflects their lipotrope density profile. *J Food Compos Anal*. 2011;24:895–915.
107. Fardet A, Chardigny JM. Plant-based foods as a source of lipotropes for human nutrition: a survey of *in vivo* studies. *Crit Rev Food Sci Nutr*. ;535–90.

108. Tristan Asensi M, Napoletano A, Sofi F, Dinu M. Low-grade inflammation and ultra-processed foods consumption: a review. *Nutrients*. 2023;15:1546.
109. Al-Wakeel DE, El-Kashef DH, Nader MA. Renoprotective effect of empagliflozin in cafeteria diet-induced insulin resistance in rats: modulation of HMGB-1/TLR-4/NF- $\kappa$ B axis. *Life Sci*. 2022;301:120633.
110. Lopes AEDSC, Araújo LF, Levy RB, Barreto SM, Giatti L. Association between consumption of ultra-processed foods and serum C-reactive protein levels: cross-sectional results from the ELSA-Brasil study. *Sao Paulo Med J*. 2019;137:169–76.
111. Bujtor M, Turner AI, Torres SJ, Esteban-Gonzalo L, Pariante CM, Borsini A. Associations of dietary intake on biological markers of inflammation in children and adolescents: a systematic review. *Nutrients*. 2021;13:356.
112. Vivi ACP, Azevedo-Silva TR, Neri D, Strufaldi MWL, Lebrão CW, Fonseca FLA, et al. Association between ultraprocessed food intake and C-reactive protein levels in preterm and term infants. *Nutrition*. 2022;99–100:111649.
113. Anto L, Blesso CN. Interplay between diet, the gut microbiome, and atherosclerosis: role of dysbiosis and microbial metabolites on inflammation and disordered lipid metabolism. *J Nutr Biochem*. 2022;105:108991.
114. Ilan Y. Leaky gut and the liver: a role for bacterial translocation in nonalcoholic steatohepatitis. *World J Gastroenterol*. 2012;18:2609–18.
115. Stefanson AL, Bakovic M. Dietary regulation of Keap1/Nrf2/ARE pathway: focus on plant-derived compounds and trace minerals. *Nutrients*. 2014;6:3777–801.
116. Wagner AE, Terschluesen AM, Rimbach G. Health promoting effects of brassica-derived phytochemicals: from chemopreventive and anti-inflammatory activities to epigenetic regulation. *Oxid Med Cell Longev*. 2013;2013:964539.
117. Rahmani S, Naraki K, Roohbakhsh A, Hayes AW, Karimi G. The protective effects of rutin on the liver, kidneys, and heart by counteracting organ toxicity caused by synthetic and natural compounds. *Food Sci Nutr*. 2023;11:39–56.
118. Chakravarthy MV, Neuschwander-Tetri BA. The metabolic basis of nonalcoholic steatohepatitis. *Endocrinol Diabetes Metab*. 2020;3:e00112.