



**Scientific Letter**

## Bacteraemia Among Patients with Sickle Cell Disease in Nigeria: Association with Spleen Size and Function

**Keywords:** Sickle cell disease; Spleen; Ultrasound.

**Published:** September 1, 2023

**Received:** July 23, 2023

**Accepted:** August 14, 2023

**Citation:** Ladu A.I., Kadaura M.U., Dauda M., Baba A.S., Jeffery C., Farate A., Adekile A., Bates I., Dacombe R. Bacteraemia among patients with sickle cell disease in Nigeria: association with spleen size and function. *Mediterr J Hematol Infect Dis* 2023, 15(1): e2023054, DOI: <http://dx.doi.org/10.4084/MJHID.2023.054>

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### To the editor.

In Sub-Saharan Africa, infections are a leading cause of morbidity among sickle cell disease (SCD) individuals. The causes of the increased risk of infection are poorly documented, but the loss of splenic function is important. Previous studies have documented increased susceptibility to bacterial infections among SCD patients, evidenced by increasing markers of splenic dysfunction;<sup>1,2</sup> however, there are no data on the association between bacterial infections and splenic function among the SCD population in Sub-Saharan Africa, partly because most of the techniques required to assess splenic function are not readily available.<sup>3</sup> We recently employed the presence of two red cell-containing inclusions - Howell-Jolly bodies (HJB) and argyrophilic (silver staining) inclusion (AI) red cells - to assess splenic dysfunction among our SCD patients.<sup>4</sup> In the present study, we aimed to determine the prevalence and pattern of organisms causing bacteraemia among our acutely-ill SCD patients and to describe any association between bacteraemia with splenic status on ultrasound and two markers of splenic dysfunction (i.e., HJB and AI red cells)

**Methods.** This cross-sectional study was conducted at the University of Maiduguri Teaching Hospital, North-Eastern Nigeria, from October 2020 to May 2021. All febrile and/or acutely ill SCD patients presenting to the adult or paediatric emergency unit during the study period were invited to participate. A case report form was used to obtain baseline clinical characteristics from the patients (or their carers). Under aseptic conditions, between 3 and 8 ml of venous blood for cultures were collected directly into appropriate BACTEC plus bottles and incubated manually at 37°C. Positive blood cultures were sub-cultured on standard media using routine microbiological techniques. Non-pathogenic organisms commonly associated with contaminated blood cultures, such as coagulase-negative *staphylococci*,

*Acinetobacter* species, *Bacillus* species, *Corynebacterium* species, *Micrococcus* species, and non-meningitidis *Neisseria* species were considered as contaminants.<sup>5</sup> The splenic function was assessed by manual estimation of Howell-Jolly bodies (HJB) and argyrophilic inclusion (AI) containing red cells from blood smears as previously described.<sup>4</sup> The splenic status was assessed using ultrasound. Data were analysed using Statistical Package for the Social Sciences (SPSS) (version 25; SPSS, Chicago, IL, USA). The data were summarized using descriptive statistics. The prevalence of bacteraemia was defined as the proportion of positive cultures in all the blood cultures. Clinical and laboratory features of patients with bacteraemia were compared to those without bacteraemia, using non-parametric analysis.

**Ethics statement.** Written informed consent was obtained from the adults and paediatric participants' parents/guardians. The study protocol was approved by the University of Maiduguri Teaching Hospital (UMTH/REC/ 20/606) and Liverpool School of Tropical Medicine (LSTM) (REC reference number: 20-010) Ethics Review Boards.

**Results.** Over the 7-month study period, 162 febrile episodes involving 140 SCD patients (median age 13.0 years; IQR 5.0 - 21.0) occurred during the visits to adult and paediatric emergency units. The predominant haemoglobin phenotype was HbSS (98.1%); only two patients (1.9%) had HbSC phenotype. A total of 113 (69.8%) blood cultures were obtained. Bacteraemia occurred in six (5.2%, 95% CI, 1% to 10%) of the culture samples. Two cultures from children aged 2 and 3 years grew *Salmonella* species. The remaining four positive cultures were among the adult patients (one culture each grew *Serratia Marcescens*, *Citrobacter* spp, *Enterobacter* spp, and an unidentified Gram-negative bacillus). Four other positive cultures were

**Table 1.** Clinical characteristics of SCD patients with positive blood cultures.

Isolates	Sex	Age (years)	Temp	Ill looking	WBC	Prior antibiotics use	Immunization completed	% AI red cells (median)	% HJB red cells (median)	Spleen size on ultrasound
<i>Salmonella typhi</i>	Female	2	38.5	Yes	34.7	Yes	Yes	65.6%	1.3%	5.6 cm
<i>Salmonella typhi</i>	Female	3	37.8	Yes	30.0	Yes	Yes	45.0%	0.6%	8.2 cm
<i>Enterococci sp</i>	Female	16	37.2	Yes	33.5	No	No	61.1%	26.0%	6.8 cm
<i>Serratia marcescens</i>	Female	22	37.2	Yes	18.1	No	Yes	73.0%	3.5%	Autosplenectomy*
<i>Citrobacter</i>	Male	25	37.8	Yes	26.7	No	Not sure	74.6%	NR	Autosplenectomy
<i>GNB (unidentified)</i>	Male	17	36.0	Yes	22.8	Yes	Yes	53.2%	2.9%	Autosplenectomy

AI argyrophilic inclusion; HJB Howell-Jolly bodies. M: male; F: female. Temp: temperature. GNB: Gram-negative bacillus WBC: white blood cell count; NR not reported; \* spleen was not visualized on ultrasound.

considered contaminants and excluded during the analysis.

The clinical characteristics and splenic status on ultrasound of patients with bacteraemia are shown in **Table 1**. The spleen was present in three patients (2 children and one adult) and absent in the remaining three (i.e., autosplenectomy). Bacteraemia was not significantly different in SCD patients with spleen present or absent spleen on ultrasound ( $P = 0.87$ ). A comparison of clinical and laboratory parameters between patients with and without bacteraemia is shown in **Table 2**. The median HJB and AI red cell counts were 1.8% (IQR 0.5% - 7.3%) and 48.2% (IQR 27.2% - 63.5%) respectively among the study participants. There was a trend towards higher counts of red cells with HJB (median 2.4% vs. 1.9%) and AI (62.4% vs. 43.4%) in patients with bacteraemia compared to those without bacteraemia respectively; however, the result was not significant for either the HJB ( $P = 0.744$ ) or AI red cell counts ( $P = 0.075$ ) (**Table 2**). Patients with bacteraemia had significantly higher white blood cell counts (mean 34.9 vs. 21.6;  $P = 0.018$ ) and raised neutrophil counts (19.4 vs. 11.3;  $P = 0.006$ ) than those without bacteraemia. All other clinical and laboratory parameters were not significantly different between the two groups.

**Discussion.** In the present study, we determined the prevalence and pattern of bacteraemia among our patients with SCD.

Our study observed prevalence rate of 5.2% is comparable to previous studies across Africa, which ranged between 4.0% and 9.7%.<sup>6-8</sup> Our prevalence rate is also similar to studies beyond Africa, including 6.1% in Jamaica,<sup>9</sup> 5.2% in the USA,<sup>10</sup> and 3.4% in the United Kingdom.<sup>11</sup> Though our finding is comparable to other studies, it is not clear if the use of over-the-counter antibiotics may have contributed to the low rates observed in studies from Africa, where there is unrestricted access to over-the-counter antibiotics;<sup>6,12</sup>

more than a quarter of our patients admitted using antibiotics prior to presentation at the health facility. Most isolates cultured in this study were Gram-negative, similar to reports among SCD patients in Nigeria.<sup>13,14</sup> In contrast, some studies from Africa have reported Gram-positive organisms like *Staphylococcus aureus* as the predominant bacteria isolated,<sup>12,15</sup> and other studies from Africa<sup>8</sup> and Western countries have identified the Gram-positive organism, *Streptococcus pneumoniae*, as the major bacterial pathogen implicated in infection among their SCD patients.<sup>10,16</sup> Patients with SCD are susceptible to infection with encapsulated organisms (*S. pneumoniae*, *H. influenzae*) and *Salmonella* species due to their underlying splenic dysfunction.<sup>17</sup> The prevalence and pattern of pathogens implicated in SCD infections may differ in the various countries. This can be influenced by the local epidemiology of infections, available vaccines and population-specific vaccine efficacy, environment, health care systems, and cultural behaviors.<sup>18</sup> Furthermore, the frequency with which specific pathogens cause infections have been shown to follow an age-specific pattern. *Salmonella* bacteremia is common among younger patients with SCD, with a peak incidence between 2 and 10 years, and can be associated with an increased risk of osteomyelitis.<sup>19,10</sup> In the current study, *Salmonella sp* was isolated in two of the children less than five years old, one of whom had a previous history and management for osteomyelitis of both femurs. The expanded bone marrow in patients with SCD, with its sluggish blood flow, is vulnerable to thrombosis, infarction, and fibrosis; this can result in ischemic foci, allowing for salmonellae localization. The proliferation of previously dormant foci of infection can accompany a sickle crisis and, with local bone changes, can result in the passage of the organisms into the bloodstream.<sup>20</sup>

Despite the high morbidity and mortality among SCD patients attributed to loss of splenic function, no study has evaluate the presence of bacterial infection and markers of splenic dysfunction in SCD patients in

**Table 2.** Association of clinical and laboratory parameters with bacteraemia.

	<b>Bacteraemia absent [n = 107/113 (94.8)]</b>	<b>Bacteraemia present [n = 6/113 (5.2)]</b>	<b>P</b>
<b>Clinical parameters</b>			
Age, years, mean (SD)	12.8 (8.9)	17.1 (11.5)	0.565
Male, n/n (%)	56/107 (52.3)	2/6 (33.3)	0.611
Temperature, mean (SD)	37.5 (0.9)	37.5 (1.0)	0.938
Jaundice, n/n (%)	36/106 (34.0)	3/6 (50.0)	0.418
Pallor, n/n (%)	50/106 (47.2)	4/6 (66.7)	0.426
Immunization completed	58/107 (54.2)	3/6 (50)	0.340
<b>Spleen parameters</b>			
Spleen status on ultrasound: n/n (%)			
Spleen present	53/99 (53.5)	3/6 (50)	0.87
Spleen absent**	46/99 (46.5)	3/6 (50)	
% AI red cells, median (IQR)	43.4 (35.5)	62.4 (22)	0.075
% HJB red cells, median (IQR)	1.9 (6.7)	2.4 (19.7)	0.744
<b>Laboratory parameters</b>			
White blood cell count (10 <sup>6</sup> /l, mean (SD))	21.6 (17.3)	34.9 (20.0)	0.018*
Hb (g/dl) mean (SD)	6.3 (2.1)	7.3 (2.1)	0.209
Platelets (10 <sup>9</sup> /l), mean (SD)	418 (210)	516 (241)	0.274
ANC (10 <sup>9</sup> /l) mean (SD)	11.3 (8.8)	19.4 (6.0)	0.006*
Reticulocyte count (%), mean (SD)	5.7 (6.3)	10.2(7.2)	0.148
Bilirubin (total)(umol/l), mean (SD)	28.7 (27.7)	22.0 (10.4)	0.881
ASAT (iu/l) mean (SD)	17.5 (16.3)	30.8 (29.6)	0.218
Haemoglobin F, %, mean (SD)	8.3 (5.9)	6.9 (1.9)	0.172

ASAT: aspartate amino transferase; ANC: absolute neutrophil count; Hb: haemoglobin; SD: standard deviation; IQR interquartile range  
\*Significant P value by Mann Whitney U test. \*\*Autosplenectomy.

SSA.<sup>3</sup> We have recently used two markers of splenic dysfunction that required simple techniques and thus can easily be performed in most resource-poor settings; the proportion of both markers, HJB- and AI-containing red cells, were higher in patients with autosplenectomy than those with visible spleens.<sup>4</sup> In the current study, we noted that SCD patients with bacteraemia were more likely to have higher AI and HJB red cell counts. However, the difference for both markers failed to reach statistical significance. The small number of patients with bacteraemia (n=6) may have affected the power to detect any significant relationship, limiting our ability to make concrete conclusions on the relationship between these markers and the risk of bacteraemia.

Furthermore, although a high count of markers of splenic dysfunction is expected to be associated with an increased risk of bacterial infections, the ability of the spleen to filter the blood of pathogens depends on several other mechanisms, including complement

activation, of humoral and cellular immune responses.<sup>17</sup> Therefore, it is unclear whether the HJB or AI counts alone can accurately reflect the spleen-related risk of bacterial infection. A larger study will be useful in providing more insight into the relationship between bacteraemia and splenic parameters.

This study has some limitations. The small number of patients with bacteraemia (5.2%) may have affected the power to identify an association with splenic parameters; the low prevalence of bacteraemia observed could be due to prior use of antibiotics by the patients. The use of one culture bottle per set rather than two for adults and the use of a manual incubator instead of the standard BAC/Alert system for our bacterial detection may have affected the identification of fastidious organisms and demonstrate the difficulties in performing clinical research in laboratories with limited resources characterizing the conditions in most Sub-Saharan African countries.

Adama I Ladu<sup>1,2</sup>, Mairo U Kadaura<sup>3</sup>, Mohammed Dauda<sup>3</sup>, Abubakar Sadiq Baba<sup>3</sup>, Caroline Jeffery<sup>1,4</sup>, Abubakar Farate<sup>5</sup>, Adekunle Adekile<sup>6</sup>, Imelda Bates<sup>1</sup> and Russell Dacombe<sup>1</sup>.

<sup>1</sup> Department of International Public Health, Liverpool School of Tropical Medicine, Liverpool, United Kingdom.

<sup>2</sup> Department of Haematology, Faculty of Basic Clinical Sciences, University of Maiduguri. Borno State, Nigeria.

<sup>3</sup> Department of Microbiology, Faculty of Basic Clinical Sciences, University of Maiduguri. Borno State, Nigeria.

<sup>4</sup>Department of Clinical Infection, Microbiology and Immunology, University of Liverpool, Liverpool, United Kingdom.

<sup>5</sup>Department of Radiology, Faculty of Clinical Sciences, University of Maiduguri. Borno State, Nigeria.

<sup>6</sup>Department of Paediatrics, Faculty of Medicine, Kuwait University, Kuwait.

**Competing interests:** The authors declare no conflict of Interest.

Correspondence to: Dr. Adama I Ladu. Department of International Public Health, Liverpool School of Tropical Medicine, Liverpool, United Kingdom; Department of Haematology, Faculty of Basic Clinical Sciences, University of Maiduguri. Borno State, Nigeria. E-mail: [adamaisahladu@gmail.com](mailto:adamaisahladu@gmail.com); [adama.ladu@lastmed.ac.uk](mailto:adama.ladu@lastmed.ac.uk).

## References:

1. Pearson HA, McIntosh S, Ritchey AK, Lobel JS, Rooks Y, Johnston D. Developmental aspects of splenic function in sickle cell diseases. *Blood*. 1979;53(3):358-65. <https://doi.org/10.1182/blood.V53.3.358.358> PMID:760858
2. Rogers D, Serjeant B, Serjeant G. Early rise in the "pitted" red cell count as a guide to susceptibility to infection in childhood sickle cell anaemia. *Archives of Disease in Childhood*. 1982;57(5):338-42. <https://doi.org/10.1136/adc.57.5.338> PMID:7092288 PMCID:PMC1627578
3. Ladu AI, Aiyenigba AO, Adekile A, Bates I. The spectrum of splenic complications in patients with sickle cell disease in Africa: a systematic review. *British Journal of Haematology*. 2021;193(1):26-42. <https://doi.org/10.1111/bjh.17179> PMID:33161568
4. Ladu A, Satumari N, Abba A, Abulfathi F, Jeffery C, Adekile A, et al. Evaluation of two red cell inclusion staining methods for assessing spleen function among sickle cell disease patients in North-East Nigeria. medRxiv; 2023. <https://doi.org/10.1101/2023.01.12.23284472>
5. Doern GV, Carroll KC, Diekema DJ, Garey KW, Rupp ME, Weinstein MP, et al. A comprehensive update on the problem of blood culture contamination and a discussion of methods for addressing the problem. *Clin Microbiol Rev*. 2019;33(1):e00009-19. <https://doi.org/10.1128/CMR.00009-19> PMID:31666280 PMCID:PMC6822992
6. Alima Yanda AN, Nansseu JR, Mbassi Awa HD, Tatah SA, Seungue J, Eposse C, et al. Burden and spectrum of bacterial infections among sickle cell disease children living in Cameroon. *BMC Infect Dis*. 2017;17(1):211. <https://doi.org/10.1186/s12879-017-2317-9> PMID:28298206 PMCID:PMC5353947
7. Makani J, Mgaya J, Balandya E, Msami K, Soka D, Cox SE, et al. Bacteraemia in sickle cell anaemia is associated with low haemoglobin: a report of 890 admissions to a tertiary hospital in Tanzania. 2015. p. 273-6. <https://doi.org/10.1111/bjh.13553> PMID:26084722 PMCID:PMC4744759
8. Williams TN, Uyoga S, Macharia A, Ndila C, McAuley CF, Opi DH, et al. Bacteraemia in Kenyan children with sickle-cell anaemia: a retrospective cohort and case-control study. 2009. p. 1364-70. [https://doi.org/10.1016/S0140-6736\(09\)61374-X](https://doi.org/10.1016/S0140-6736(09)61374-X) PMID:19747721
9. Wierenga KJ, Hambleton IR, Wilson R, Alexander H, Serjeant BE, Serjeant GR. Significance of fever in Jamaican patients with homozygous sickle cell disease. *Archives of disease in childhood*. 2001;84(2):156-9. <https://doi.org/10.1136/adc.84.2.156> PMID:11159294 PMCID:PMC1718651
10. Zarkowsky HS, Gallagher D, Gill FM, Wang WC, Falletta JM, Lande WM, et al. Bacteremia in sickle hemoglobinopathies. *The Journal of pediatrics*. 1986;109(4):579-85. [https://doi.org/10.1016/S0022-3476\(86\)80216-5](https://doi.org/10.1016/S0022-3476(86)80216-5) PMID:3531449
11. Morrissey BJ, Bycroft TP, Almossawi O, Wilkey OB, Daniels JG. Incidence and predictors of bacterial infection in febrile children with sickle cell disease. *Hemoglobin*. 2015;39(5):316-9.
12. Kizito ME, Mworozzi E, Ndugwa C, Serjeant GR. Bacteraemia in homozygous sickle cell disease in Africa: is pneumococcal prophylaxis justified? *Arch Dis Child*. 2007;92(1):21-3. <https://doi.org/10.1136/adc.2005.088807> PMID:16531454 PMCID:PMC2083172
13. Brown B, Dada-Adegbola H, Trippe C, Olopade O. Prevalence and Etiology of Bacteremia in Febrile Children with Sickle Cell Disease at a Nigerian Tertiary Hospital. *Mediterr J Hematol Infect Dis*. 2017;9(1):e2017039. <https://doi.org/10.4084/mjhid.2017.039> PMID:28698782 PMCID:PMC5499496
14. Akinyanju O, Johnson A. Acute illness in Nigerian children with sickle cell anaemia. *Annals of tropical paediatrics*. 1987;7(3):181-6. <https://doi.org/10.1080/02724936.1987.11748503> PMID:2445266
15. Akuse RM. Variation in the pattern of bacterial infection in patients with sickle cell disease requiring admission. *Journal of tropical pediatrics*. 1996;42(6):318-23. <https://doi.org/10.1093/tropej/42.6.318> PMID:9009554
16. Ellison AM, Ota KV, McGowan KL, Smith-Whitley K. Epidemiology of bloodstream infections in children with sickle cell disease. *Pediatr Infect Dis J*. 2013;32(5):560-3. <https://doi.org/10.1097/INF.0b013e318286c75b> PMID:23340560
17. Booth C, Inusa B, Obaro SK. Infection in sickle cell disease: a review. *International Journal Of Infectious Diseases*. 2010;14(1):e2-e12. <https://doi.org/10.1016/j.ijid.2009.03.010> PMID:19497774
18. Obaro SK, Tam PYI. Preventing Infections in Sickle Cell Disease: The Unfinished Business. *Pediatr Blood Cancer*. 2016 May;63(5):781-5. doi: 10.1002/pbc.25911. <https://doi.org/10.1002/pbc.25911> PMID:26840500
19. Alsaiif M.A., Abdulbaqi M., Al Noaim K., Aghbari M., Alabdulqader M., Robinson J.L. Prevalence of serious bacterial infections in children with sickle cell disease at King Abdulaziz Hospital, Al Ahsa. *Mediterr J Hematol Infect Dis* 2021, 13(1): e2021002 <https://doi.org/10.4084/mjhid.2021.002> PMID:33489041 PMCID:PMC7813273
20. Anand AJ, Glatt AE, editors. *Salmonella osteomyelitis and arthritis in sickle cell disease. Seminars in arthritis and rheumatism*; 1994: Elsevier. [https://doi.org/10.1016/0049-0172\(94\)90076-0](https://doi.org/10.1016/0049-0172(94)90076-0) PMID:7899877