Check for updates

OPEN ACCESS

EDITED BY Daiqing Liao, University of Florida, United States

REVIEWED BY Yajun Lin, Peking University, China Heiko Bruns, University Hospital Erlangen, Germany

*CORRESPONDENCE Wuli Zhao, ⊠ wenlyzh@imb.pumc.edu.cn Yanni Xu, ⊠ xuyanniwendeng@hotmail.com

RECEIVED 22 March 2023 ACCEPTED 07 August 2023 PUBLISHED 28 August 2023

CITATION

Zhou H, Zhao C, Shao R, Xu Y and Zhao W (2023), The functions and regulatory pathways of S100A8/A9 and its receptors in cancers. *Front. Pharmacol.* 14:1187741. doi: 10.3389/fphar.2023.1187741

COPYRIGHT

© 2023 Zhou, Zhao, Shao, Xu and Zhao. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

The functions and regulatory pathways of S100A8/A9 and its receptors in cancers

Huimin Zhou¹, Cong Zhao¹, Rongguang Shao¹, Yanni Xu^{2*} and Wuli Zhao^{1*}

¹State Key Laboratory of Respiratory Health and Multimorbidity, Key Laboratory of Antibiotic Bioengineering, Ministry of Health, Laboratory of Oncology, Institute of Medicinal Biotechnology, Chinese Academy of Medical Sciences and Peking Union Medical College, Beijing, China, ²NHC Key Laboratory of Biotechnology of Antibiotics, National Center for New Microbial Drug Screening, Institute of Medicinal Biotechnology, Chinese Academy of Medical Sciences and Peking Union Medical College, Beijing, China

Inflammation primarily influences the initiation, progression, and deterioration of many human diseases, and immune cells are the principal forces that modulate the balance of inflammation by generating cytokines and chemokines to maintain physiological homeostasis or accelerate disease development. S100A8/A9, a heterodimer protein mainly generated by neutrophils, triggers many signal transduction pathways to mediate microtubule constitution and pathogen defense, as well as intricate procedures of cancer growth, metastasis, drug resistance, and prognosis. Its paired receptors, such as receptor for advanced glycation ends (RAGEs) and toll-like receptor 4 (TLR4), also have roles and effects within tumor cells, mainly involved with mitogen-activated protein kinases (MAPKs), NF-κB, phosphoinositide 3-kinase (PI3K)/Akt, mammalian target of rapamycin (mTOR) and protein kinase C (PKC) activation. In the clinical setting, S100A8/A9 and its receptors can be used complementarily as efficient biomarkers for cancer diagnosis and treatment. This review comprehensively summarizes the biological functions of S100A8/A9 and its various receptors in tumor cells, in order to provide new insights and strategies targeting S100A8/A9 to promote novel diagnostic and therapeutic methods in cancers.

KEYWORDS

cancer, inflammation, RAGEs, S100A8/A9, TLR4

1 Introduction

Some adverse conditions, such as dietary imbalance, tobacco smoking, pollutants, radiation, as well as bacterial and viral infections, can cause inflammation in multiorgan systems, eventually resulting in tumorigenesis and even necrosis. Inflammation is closely related to all stages of tumor development including initiation, growth, invasion, and survival, by mainly boosting genomic mutations, angiogenesis, and immunomodulation (Grivennikov et al., 2010; Hou et al., 2021). Various growth factors, cytokines, and chemokines are produced in the tumor microenvironment, which can bind to multitudinous cell surface receptors, like receptor for advanced glycation end products (RAGEs), vascular endothelial growth factor receptor (VEGFR), epidermal growth factor receptor (EGFR), and CXC chemokine receptor (CXCR), thus exerting diverse and profound effects via complex transduction ways including NF- κ B, TGF- β , phosphoinositide 3-kinase (PI3K)/Akt, hypoxia-inducible factor (HIF) signaling, and reactive oxygen species (ROS)

accumulation, in order to support the balance situation of tumor inflammation (Quail and Joyce, 2013; Libby and Kobold, 2019; Wattenberg and Beatty, 2020).

The S100 protein family is the principal participant in inflammation and includes 25 known members with a high degree of structural and sequence similarity. They act as intracellular and extracellular signal transduction factors bearing complicated and extensive physiological responsibilities, including but not limited to Ca^{2+} sensing, cell proliferation and differentiation, tumor development, cell cycle modulation, proinflammatory cytokine production, adipogenesis, and fatty acid transport (Bresnick et al., 2015; Gonzalez et al., 2020).

S100A8 and S100A9, also known as myeloid-related protein (MRP)8 or Calgranulin A and MRP14 or Calgranulin B, respectively, as well as the S100A8/A9 hetero-complex (Calprotectin), belong to the Ca2+-binding S100 protein family and are abundant in neutrophils and monocytes (Pruenster et al., 2016). As a significant inflammatory mediator, S100A8/ A9 extensively influences granulocyte and monocyte migration, antibiosis, inflammatory propagation, macrophage activation, and cytostatic effect (Kerkhoff et al., 1998), playing prominent and complicated roles in myocardial infarction (Cai et al., 2020), cancer (Ghavami et al., 2009), coronavirus disease 2019 (Guo et al., 2021), tuberculosis (Scott et al., 2020), psoriasis (Matsunaga et al., 2021), and systemic lupus erythematosus (Lood et al., 2016). It is also a promising therapeutic target and clinical diagnostic biomarker for inflammatory bowel disease (Burri and Beglinger, 2014), rheumatoid arthritis (Hurnakova et al., 2015), type 2 diabetes (Ortega et al., 2012), acute Kawasaki disease (Hirono et al., 2006) and community-acquired pneumonia (Xie et al., 2023). Accordingly, we focused on the roles and mechanisms of \$100A8/ A9 in cancers to summarize the research advances, expecting to provide valid theoretical support for this study and future perspectives.

2 S100A8/A9 storage, structure, and effects

2.1 S100A8/A9 expression and release

S100A8/A9 is mainly stored in immune cells such as dendritic cells, neutrophils, monocytes, and macrophages, and is secreted to modulate inflammatory metabolic processes during infection, drug stress, heat, and other irritant conditions (Wang S. et al., 2018). Fibroblasts, keratinocytes, and vascular endothelial cells can also express S100A8/A9 upon activation (Schiopu and Cotoi, 2013). S100A8/A9 is predominantly localized in the cytoplasm and can translocate to the cytoskeleton and plasma membrane to increase intracellular calcium levels (Roth et al., 1993). S100A8/A9 constitutes approximately 40% cytosolic protein content in the main reserve source of neutrophils, and some S100A8 exists in granule membranes and euchromatin regions of the nucleus. S100A8/A9 isolated from neutrophils can further dose-dependently induce neutrophil activation and adhesion (Edgeworth et al., 1991; Sprenkeler et al., 2022).

At the same time, IL-17 (Bai et al., 2021), LPS (Fontaine et al., 2014), HIF-1 (Grebhardt et al., 2012), and tumor necrosis factor- α

(TNF- α) (Nicaise et al., 2017) can also stimulate S100A8/ A9 expression and release, while p38 MAPK signaling pathway (Mose et al., 2013), C/EBP α (Hayashi et al., 2007), protein kinase C (PKC) activation (Rammes et al., 1997), ROS production and K⁺ exchange (Tardif et al., 2015) are essential regulators for S100A8/ A9 synthesis and secretion.

2.2 S100A8/A9 composition

Human S100A8 and S100A9 comprise 93 and 113 amino acids, respectively, with 10.8 and 13.2 kDa molecular weight, respectively. S100A8 and S100A9 monomers preferentially form stable heterodimers as their basic form, and two heterodimers can assemble into an (S100A8/A9)₂ heterotetramer, which is indispensable for the basic biological functions of cells (Pruenster et al., 2016; Jukic et al., 2021).

Like other S100 proteins, S100A8/A9 is characterized by two EFhand (such as helix–loop–helix)-type Ca²⁺-binding domains with different affinities to calcium, which are located at the N- (EF-hand 1, low affinity) and C-terminal (EF-hand 2, high affinity) halves, and connected with an intermediate region usually called the hinge region. Each Ca²⁺-binding site is flanked by two helices: EF-hand 1 is flanked by helices I and II, helices III and IV flank EF-hand 2, and the hinge region links helices II and III (Donato, 1999, 2001; Leukert et al., 2006; Korndörfer et al., 2007). The specific structure of S100A8/A9 is also described in Figure 1.

2.3 The fundamental functions of S100A8/A9

S100A8/A9 plays complex roles in chemotaxis (Ryckman et al., 2003), cell migration (Gross et al., 2014), inflammation (Tong et al., 2014), phagocytosis (Basu et al., 2018), and oxidative metabolism (Sroussi et al., 2010), involving sophisticated functions and mechanisms. First, being an important inflammatory factor, S100A8/A9 is involved in inflammation caused by injury, microorganisms, and stress. The released S100A8/A9 can promote IL-1 β secretion in neutrophils in an autocrine or paracrine fashion to sustain the inflammatory response by stimulating myelopoiesis and other leukocyte production (Sreejit et al., 2020). The secreted S100A8 also stimulates myeloid cell chemotaxis by moving to inflammatory sites and sustaining leukocyte recruitment (Harrison et al., 1999).

Furthermore, calcium-induced S100A8/A9 co-localizes with microtubules in activated monocytes. It promotes microtubule formation to become more elongated and thread-like in resting phagocytes, and S100A8 is chiefly responsible for tubulin polymerization, whereas S100A9 plays important regulatory roles in this process. Together with p38 MAPK signaling pathway activation, calcium signaling-induced S100A8/A9 conformational changes can disrupt this stabilizing state of microtubules, thus facilitating the trans-endothelial migration of phagocytes (Vogl et al., 2004). Other studies have demonstrated that phosphorylation of S100A8/A9, superoxide production, and PKC modulation are essential for translocation from the cytosol to the membranes and cytoskeleton of S100A8/A9, which is of great



importance for Ca²⁺-dependent reorganization of cytoskeletal filaments and inflammatory activation (Lemarchand et al., 1992; Goebeler et al., 1995; Guignard et al., 1996; van den Bos et al., 1996).

Moreover, S100A8/A9 heterodimers bind to arachidonic acid (AA) in a Ca²⁺-dependent manner and transfer it to nicotinamide adenine dinucleotide 3-phosphate (NADPH) oxidase to trigger enzymatic activity and stabilize the activated enzyme in neutrophils (Kerkhoff et al., 1999; Bai et al., 2022). Simultaneously, S100A8/A9 can also transfer AA to neighboring cells such as leukocytes, vascular endothelium, and smooth muscle cells at inflammatory loci to adjust the physiological state of the cells and produce inflammatory mediators to exacerbate inflammation (Kerkhoff et al., 2001; Kannan, 2003; Kerkhoff et al., 2005).

In addition to these functions, S100A8/A9 also participates in procoagulant platelet formation (Colicchia et al., 2022), intestinal microbiota development (Willers et al., 2020), autophagy-dependent ferroptosis (Tao et al., 2022), anemia, and bone marrow failure (Giudice et al., 2019), most of which are closely related to immunity and inflammation, indicating that S100A8/A9 may serve as a biomarker and therapeutic target in this field.

3 S100A8/A9 has complicated influence on cancer cells

S100 proteins widely participate in cellular proliferation, metabolism, inflammation, and many other diverse physiological

activities by EF-hand binding to calcium, zinc, or copper to change the spatial conformation and interact with other target proteins to activate downstream signals (Oesterle and Bowman, 2015). In cancer cells, S100A8/A9 is the main modulator of apoptosis (Qin et al., 2010), migration, invasion (Yin et al., 2013), brain metastasis (Monteiro et al., 2022), drug resistance (Zavorka Thomas et al., 2021) and many other processes, acting as a dual and diverse sensor in different steps of tumor progression, some of which are described below.

3.1 S100A8/A9 is overexpressed in carcinoma and has negative outcomes in patients

According patient to tissue microarravs and immunohistochemical detection, both S100A8 mRNA and protein levels are highly elevated in many cancers, including anaplastic thyroid carcinoma (Reeb et al., 2015), breast cancer (Wang D. et al., 2018), lung cancer (Sumardika et al., 2019), acute myeloid leukemia (Mondet et al., 2021), and pancreatic cancer (Basso et al., 2014), and associated with less survival, poor outcome, and high probability of relapse; thus, S100A8 could act as a potential biomarker for different cancers. Simultaneously, S100A9 expression is also elevated in the tissues or sera of colorectal carcinoma (Huang et al., 2019), hepatocellular carcinoma (Zhong et al., 2022), gastric cancer (Fan et al., 2012),

nasopharyngeal carcinoma (Hu et al., 2021), and renal cell carcinoma (Zhang et al., 2015), consistent with low disease-free, overall, and recurrence-free survival periods, which can also be used as a powerful diagnostic indicator (Koh et al., 2019; Koh et al., 2021). The DNA methylation level of S100A8 decreases in hepatocellular carcinoma tissues compared to that in adjacent normal tissues, which is closely related to reduced overall and progression-free survival periods, indicating that S100A8 DNA methylation could also be a useful diagnostic and prognostic marker (Liu K. et al., 2016). Thus, incorporating S100A8/A9 as a supplementary biomarker will aid the early detection and timely treatment of cancers.

3.2 S100A8/A9 plays important roles in the proliferation of cancer cells

Since proliferation is one of the most important tumor characteristics, many studies have focused on the function of S100A8/A9 in it. S100A8/A9 overexpression notably improves the viability of nasopharyngeal carcinoma cells, and S100A8/ A9 knockdown represses proliferation and colony formation by inhibiting the PI3K/Akt pathway (Wen et al., 2021). Exogenous S100A8/A9 can bind to RAGEs to induce squamous cell carcinoma proliferation, and direct interruption by a specific RAGE-blocking antibody can reduce cellular proliferation through MAPK phosphorylation alteration (Iotzova-Weiss et al., 2015). Moreover, S100A8/A9 induces the apoptosis of chronic eosinophilic leukemia cells by suppressing the downstream signal of FIP1L1-PDGFRa through decreasing its mRNA and proteins expression, as well as eliciting the activity of mitochondria-associated proteins such as caspase 3/9, AIF, and Bax (Lee et al., 2020). In an orthotopic lung cancer mouse model, intranasal S100A8 delivery effectively delayed tumor growth and prolonged survival by inducing the antioxidant activity of many key genes and activating immune cells to positively modify the immune microenvironment (Wong et al., 2022). The distinct roles of S100A8/A9 in tumor proliferation might be consistent with its concentration and the tumor cell environment. If S100A8/A9 cooperates with other immune factors to activate suppressive pathways in tumor cells, it would lead to cell death. Moreover, S100A8/A9 concentration lower or higher than a particular concentration might induce the proliferation or apoptosis of tumor cells, respectively. Therefore, the specific concentration and pathways involved must be further explored.

3.3 S100A8/A9 has a dual effect on the migration and invasion of cancer cells

Metastasis is a basic property of tumor cells. In colorectal cancer (CRC) cells, signaling factors TGF- β can induce S100A8 expression, thus promoting the movement of CRC cells. However, cell migration and invasion significantly decrease after S100A8 knockdown, which is not restored by TGF- β (Li et al., 2021). The addition of recombinant human S100A8/A9 protein promotes esophageal squamous cell carcinoma cell migration and invasion via the Akt and p38 MAPK signaling pathways, whereas S100A8/A9 silencing suppresses carcinoma cell migration and invasion, as well as Akt and

p38 MAPK activation (Tanigawa et al., 2022). Simultaneously, the exogenous S100A8/A9 complex promotes the migration of triplenegative breast cancer cells by activating the RAGE-FAK-Hippo/ Yes kinase-associated protein (YAP) transduction system (Rigiracciolo et al., 2022). In animal models, compared to controls, anaplastic thyroid carcinoma cells with S100A8 knockdown showed less lung metastasis and tumor load by tail vein injection, whereas S100A9 knockdown did not affect tumorigenesis and metastasis (Reeb et al., 2015). In addition, many studies have shown that Toll-like receptor 4 (TLR4), NF-KB, matrix metalloprotease-2 (MMP-2), and other proteins also participate in S100A8/A9-mediated cancer cell migration and invasion through different transduction pathways (Kwon et al., 2013; Silva et al., 2014; Herik Rodrigo et al., 2022). Moreover, intracellular S100A8 can suppress invasion and epithelial-mesenchymal transition (EMT) in breast cancer cells (Cormier et al., 2014). This might be related to the source of S100A8/A9 as extracellular could bind to specific receptors to activate certain downstream pathways, whereas intracellular S100A8/A9 might be regulated in completely different ways. At the same time, the inflammatory microenvironment also participates in this process, where cytokines could trigger crossing pathways to push forward to entirely different cell fates, along with positively or negatively modulating S100A8/A9 effects, and the detailed role of \$100A8/A9 for tumor metastasis need to be explored separately.

3.4 S100A8/A9 contributes to angiogenesis in cancer cells

Some studies have shown that low S100A8/A9 concentrations can not only stimulate the proliferation and migration of vascular endothelial cells in vitro, but also promote tumor cell vascularization to benefit their growth (Li et al., 2012; Zhong et al., 2020). Studies have shown that myeloid cell-derived S100A9 can increase blood supply and promote angiogenesis by inducing IL6 and IL10 to create a survival niche for subsequent tumor progression in multiple myeloma (De Veirman et al., 2017). Meantime, S100A9 activates angiogenesis via an autocrine effect on tumor cells and a paracrine effect on stromal cells in oral cancer, similarly accompanied by IL6 production (Fang et al., 2015). In addition, studies using a mouse model have found that the S100A8/ lung cancer S100A9-EMMPRIN-Vegfa axis promotes tumor angiogenesis and progression, which is associated with the prognosis of lung cancer patients (Nguyen et al., 2021). Although the promotional effect of S100A8/A9 on angiogenesis has been confirmed by many studies, whether S100A8/A9 could induce nutritional deprivation under some unique conditions and be targeted to prevent angiogenesis remains unclear, and its function in tumor angiogenesis needs to be further investigated.

3.5 S100A8/A9 promotes immunosuppression of cancer cells

In addition to the complex functions mentioned above, S100A8/ A9 plays a major role in tumor immunology and metabolic activity (Wu et al., 2022). Programmed death 1 (PD-1) is an important co-

inhibitory signal, and tumor or immune cells express PD-1 ligands (PD-L1) to evade antitumor immune responses (Lei et al., 2020). Studies have shown that S100A8 expression is positively correlated with PD-L1 levels in tumor samples and can induce PD-L1 in macrophages by activating PI3K/Akt, MAPKs, NF-KB, and STAT3 to increase PD-L1 transcription. Furthermore, S100A8treated macrophages cannot promote T cell proliferation in vitro and can suppress the cytotoxic function of tumor-primed T cells in vivo through PD-L1 (Li et al., 2020). Breast cancer-associated gene 1 (BRCA1) is a familiar tumor suppressor gene, and BRCA1 deficiency can contribute to various breast cancers. Moreover. BRCA1 mutations can activate the S100A9-CXCL12-pSTAT3 axis to induce myeloid-derived suppressor cells (MDSCs) accumulation, further establishing an immunosuppressive environment for tumorigenesis, and blockade of S100A9-CXCL10 could diminish immunosuppressive niches and inhibit cancer initiation and growth in the mouse model (Li et al., addition, S100A9-RAGEs-p38 2022). In MAPK and S100A9-TLR4-NF-KB also participate in MDSCs activation and chemotaxis in vitro, which are closely related to immunosuppression and neoplastic progression (Huang et al., 2019). In recent years, tumor immunotherapy has become an emerging research hotspot. Thus, it is inspiring to find that S100A8/A9 takes part in immunomodulatory processes and particularly has a positive correlation with PD-L1, which might help us find new molecular mechanisms to detect immune evasion; however, further elucidation is needed.

3.6 S100A8/A9 increases the chemoresistance of cancer cells

Drug resistance is one of the important problems in cancer therapy. Previous studies have shown that recombinant human S100A8/A9 can reduce the cytotoxicity of doxorubicin/ cyclophosphamide in breast cancer cells (Hsu et al., 2015). Moreover, S100A9 overexpression can lead to resistance to cisplatin-induced apoptosis in bladder cancer, whereas S100A9 inhibition can enhance tumor cell sensitization to cisplatin-induced apoptosis (Kim et al., 2014). Interleukin 1 receptor-associated kinase 1 (IRAK1) is a serine/threonineprotein kinase that mediates IL1-TLR-NF-KB signaling pathway; and IRAK1-S100A9 axis plays a critical role in the paclitaxel resistance in nasopharyngeal carcinoma, while S100A9 ablation could increase the sensitivity of cells to paclitaxel, and the addition of recombinant S100A9 could rescue the resistance to paclitaxel after S100A9 knockdown (Liu et al., 2021). IL6 is an essential driver of S100A8/A9 expression in acute myeloid leukemia cells via JAK/STAT3 signaling; and acute myeloid leukemia populations with elevated S100A8/A9 levels are highly resistant to doxorubicin, which can be efficiently reversed by blocking JAK/STAT3 signaling (Böttcher et al., 2022).

Autophagy is a catabolic process involving the formation of autophagosomes that engulf proteins or organelles and the translocation of autophagosomes to lysosomes for degradation, which also causes chemotherapeutic resistance (Levy et al., 2017). Previous studies have reported that S100A8 triggers autophagy by promoting BECN1–PI3KC3 complex expression and BECN1-BCL2 complex formation inhibition to maintain the strong resistance of B-cell lymphoma cells to adriamycin and vincristine (Zhang et al., 2021). Simultaneously, S100A8 knockdown decreases LC3 conversion, p62 degradation, and autophagosome maturation with ROS participation in leukemia cells, thus inhibiting the initiation of autophagy to increase chemotherapy sensitivity (Yang et al., 2012). In view of the facilitation of drug resistance by S100A8/A9, S100A8/ A9 antagonists might be developed to increase the vulnerability of tumor cells to chemotherapy, and the combined use of autophagy inhibitors might have excellent therapeutic effects. All concepts must be further tested and verified.

4 Main S100A8/A9-binding receptors and molecular pathways in cancer cells

Since S100A8/A9 functions like soil signal and the paired receptors act like soil sensor, S100A8/A9 receptors, including RAGEs, TLR4, extracellular matrix metalloproteinase inducer (EMMPRIN), melanoma cell adhesion molecule (MCAM), activated leukocyte cell adhesion molecule (ALCAM), and neuroplastin (NPTN), have a common name 'S100 soil sensor receptors (SSSRs)' (Kinoshita et al., 2019a; Tomonobu et al., 2020). These receptors are always used as biomarkers for the diagnosis and prediction of idiopathic pulmonary fibrosis (Machahua et al., 2018), proliferative diabetic retinopathy (Abu El-Asrar et al., 2021), lupus nephritis (Lei et al., 2022), obstructive sleep apnea (Sun et al., 2020), and subarachnoid hemorrhage (Heinz and Schneider, 2022). Simultaneously, the S100A8/A9-SSSRs axis participates in many aspects of tumor initiation and development, usually involving MAPKs, NF-KB, PI3K/Akt, and mTOR along with the assistance of VEGF, YAP, Nrf2, and HIF-1 (Jiang et al., 2012; Tang et al., 2015). The structure and regulatory pathways of SSSRs are introduced below, which are also summarized in Figures 1, 2 respectively.

4.1 RAGEs

RAGEs, mainly located on the surfaces of macrophages, endothelial cells, dendritic cells, and tumor cells, act as patternrecognition receptors and belong to the immunoglobulin (Ig) superfamily (Ott et al., 2014). They distinguish and bind to many different ligands, including high-mobility group box-1 (HMGB1), amyloid-peptide, S100 proteins, and advanced glycation end products (AGEs), triggering numerous downstream signal transductions (El-Far et al., 2020). The full-length RAGE is probably composed of a large extracellular domain, a single transmembrane-spanning helix, and a short cytoplasmic region essential for signal transduction (Matsumoto et al., 2008). The extracellular region consists of one N-terminal variable (V-type) domain and two constant (C-type) domains, and the ligand-binding site is located on the V-type domain and predominately folded into a two β -sheet and a conserved pair of cysteines with a disulfide bond (Dattilo et al., 2007).

S100 proteins can bind to RAGEs to modulate tumor cell growth, survival, and invasion, accompanied by the activation of



FIGURE 2

The main transduction pathways involving with S100A8/A9 and its receptors in cancers. Extracellular S100A8/A9 can bind to different receptors and activate signaling pathways to regulate the progression and development of tumor cells, including proliferation, metastasis, drug resistance, immune evasion, and angiogenesis. Intracellular S100A8/A9 also participates in the activation of MAPKs, NF-kB, PI3K/Akt, and other transduction pathways. Simultaneously, intracellular and extracellular cytokines and kinases assist the functions of S100A8/A9 and jointly regulate the eventual fate of tumor cells, depending on the comprehensive effects of physiological status, environmental conditions, and external incentives.

various signal transduction pathways, such as JNK, P38 MAPK, ERK, NF- κ B, and PI3K/Akt (Ahmad et al., 2018). Previous studies have reported that the P38 MAPK and JNK pathways can induce cell apoptosis through caspase activation and their inhibitors can alter caspase-3, -8, and -9 expression (Liu T. et al., 2016). Meanwhile, exogenous S100A8/A9-induced ERK1/2 phosphorylation can promote the proliferation and invasion of tumor cells (Iotzova-Weiss et al., 2015). I κ B α phosphorylation and NF- κ B activation due to low S100A8/A9 concentrations usually play indispensable roles in the development of inflammation-based tumors; namely, nuclear translocation of NF- κ B could promote the expression of many pro-tumorigenic genes, like apoptotic gene Fas, chemokines CXCL1, growth factor Plf2, and zinc transporter Slc39a10, actively participating in angiogenesis and recruitment (Ichikawa et al., 2011). Under hypoxic conditions, oxidative stress, and inflammation, RAGE activation sustains the KRAS signaling pathway to facilitate the stabilization and transcriptional activity of HIF1a, resulting in pancreatic tumor growth. RAGE knockdown and KRAS signaling inhibition can promote HIF1a degradation and increase hypoxia-induced tumor cell death (Kang et al., 2014). Since RAGEs are the most common S100A8/A9 receptors, when focusing on traditional targets to screen new drugs, we need to focus on innovative signaling pathways to find novel methods to meet emerging clinical needs.

4.2 TLR4

To date, 10 different and functional TLRs have been found in humans. Structurally, TLRs are transmembrane domain proteins

consisting of three core parts: an ectodomain of leucine-rich region (LRR), transmembrane, and a cytoplasmic domain of Toll/interleukin-1 receptor (TIR) domain that recruits MyD88 and TRAM adapter to activate downstream NF- κ B, AP-1, and MAPKs signaling axes (Mishra and Pathak, 2019; Kashani et al., 2021).

S100A8/A9 can bind to TLR4 and induce many various signal transduction pathways, including ERK, p38 MAPK, PKC, NF-ĸB, TNF-a, and IRAK1 (Ehrchen et al., 2009). S100A8 overexpression in cholangiocarcinoma could facilitate in vitro and in vivo cell invasion and migration by activating the NF-kB pathway as well as increasing VEGF expression, while S100A8 knockdown or TLR4 and NF-KB inhibition could significantly inhibit cell migration and metastasis (Pan et al., 2020). Meantime, after TLR4/Akt activation, the Keap1/ Nrf2 cascade can promote tumorigenesis, invasion, and migration of CRC cells by upregulating cytochrome P450 monooxygenase CYP2J2 expression (Kong et al., 2021). Moreover, TLR4 activation can trigger pyroptosis in non-small cell lung cancer in vivo and in vitro by promoting mitochondrial ROS generation and Ca2+ aggregation in an NLRP3/caspase-1/GSDMD-dependent manner (Yuan et al., 2021). The Apc $^{\rm Min/+}$ mouse model is a well-utilized intestinal cancer model. TLR4 can promote intestinal tumorigenesis by activating cytokine-cytokine receptor interactions, while TLR4 knockdown can significantly decrease the expression of S100A8/A9 and other factors to inhibit the formation of intestinal adenomas and prolong the life of Apc Min/+ mice (Shi et al., 2020). Although many studies have focused on the relationship and effects of TLR4 and S100A8/A9 in cancer, the mechanisms that could be developed as drug targets are not wellreported, and research in this area should be constantly strengthened.

4.3 EMMPRIN

EMMPRIN, also called CD147 or basigin, serves as a receptor for calprotectin in tumor cells and has a higher affinity for S100A9 than that for S100A8 (Hibino et al., 2013). EMMPRIN activation will mainly initiate PI3K, MAPKs, and NF-κB pathways, promoting the growth, angiogenesis, dissemination, and inflammation of tumor cells (Xin et al., 2016; Dana et al., 2021). It comprises 269 amino acid residues and can be divided into three core parts: an extracellular N-terminal domain containing two C2-type Ig regions responsible for activating matrix metalloproteinases and becoming glycosylated, a hydrophobic transmembrane domain acting as a signal peptide, and a C-terminal intracellular cytoplasmic domain responsible for association with integrins (Bai et al., 2014; Xiong et al., 2014).

EMMPRIN is overexpressed in penile carcinoma with a remarkable influx of S100A8⁺S100A9⁺ immune cells, resulting in intracellular signaling that enhances the expression of protumorigenic genes (Mohr et al., 2022). Simultaneously, EMMPRIN expression on CD8⁺ tumor-infiltrating lymphocytes is significantly elevated, which cooperates with other receptors to negatively regulate antitumor immune responses and facilitate tumor-immune escape, whereas EMMPRIN deficiency can inhibit tumor growth and enhance the effector function of T cells (Chen Y. et al., 2021). Protein methylation is an important post-translational modification; methylated EMMPRIN is overexpressed in non-small cell lung cancer and enhances glycolysis and lactate export to promote cancer progression *in vivo* and decrease the overall survival of patients (Wang et al., 2021). EMMPRIN also influences the drug resistance of tumor cells. Temozolomide is a common chemotherapeutic agent for glioma treatment. Overexpressed EMMPRIN can inhibit Akt/GSK3β-dependent Nrf2 degradation to promote Nrf2-mediated antioxidant gene expression, which inhibits temozolomide-triggered ROS production and cell death, ultimately resulting in the resistance of glioma cells to temozolomide (Bu et al., 2021). Reprogramming of lipid metabolism is also a representative characteristic of tumor cells. EMMPRIN could promote de novo lipogenesis via the upregulation of lipogenic enzymes by activating the Akt/mTOR signaling pathway as well as suppress fatty acid oxidation via downregulating the expression of fatty acid oxidative enzymes and PPARa by the p38 MAPK signaling pathway, thus increasing the growth and aggressiveness of hepatocellular carcinoma cells (Li et al., 2015). Unlike RAGEs and TLR4, EMMPRIN plays an important role in tumor immunity and metabolism, in which field can be used as a target to design clinical drugs. Combined with drug sensitizers such as autophagy inhibitors, it might have significant antitumor effects.

4.4 MCAM

MCAM, originally identified as CD146, is a cell adhesion molecule involved in various tumor physiological activities including angiogenesis, immune response, and migration (Wang and Yan, 2013; Wang et al., 2020). Structurally, CD146 is a 113 kDa transmembrane glycoprotein with three main parts: an N-terminal extracellular domain, a transmembrane domain, and a long (lgCD146) and short (shCD146) C-terminal cytoplasmic domain. The extracellular portion contains five Ig-like domains, consisting of two variable regions (V) and three constant regions (C2) (V-V-C2-C2-C2), including eight potential N-glycosylation sites. lgCD146 and shCD146 are generated by alternative splicing and have different functions: the longer one contains two phosphorylation sites for PKC and a dileucine motif for CD146 internalization and trafficking, and the shorter one has one PKC phosphorylation site and one PDZ-binding domain for cell signaling (Lei et al., 2015; Leroyer et al., 2019; Rapanotti et al., 2021).

S100A8/A9 can bind to MCAM, which is overexpressed in triple-negative malignant breast cancer, promoting cell EMT phenotype and mortality by activating the S100A8/ A9-MCAM-ETV4-ZEB1 axis, along with MCAM knockdown leading to a reduced response to S100A8/A9 and downregulation of migration ability (Chen et al., 2019a). In melanoma cells, activation of the S100A8/A9-MCAM-TPL2-ETV4-MMP25 axis promotes growth and lung metastasis in vivo (Chen et al., 2019b). Moreover, NF-KB activation and ROS formation usually play pivotal roles in MCAM-mediated adhesion and invasion capacity in cancer progression (Ruma et al., 2016). Glucosaminyl (N-acetyl) transferase 3 (GCNT3) is a glycosyltransferase that is overexpressed in various cancer cells. It promotes the maintenance of MCAM protein at high levels and elevates the stability of MCAM by glycosylation, thus maintaining the responsiveness of MCAM to S100A8/A9 to increase melanoma cellular migration and invasion (Sumardika et al., 2018). MCAM also mediates chemoresistance in small-cell lung cancer and is upregulated in chemo-resistant cells. MCAM knockdown can increase chemosensitivity, induce cell cycle arrest, and promote apoptosis by increasing p-BAD and caspase-3 expression. PI3K/ Akt/SOX2/CREB1 signaling and glycolytic shifts also participate in MCAM-guided chemoresistance in cancer cells (Tripathi et al., 2017). MCAM can also promote angiogenesis to supply nutrients for tumorigenesis, and MCAM inhibition can increase the penetration of chemotherapeutic drugs, indicating that MCAM may be used as a new therapeutic target (Lei et al., 2021). Owing to the widespread involvement of MCAM in various aspects of tumor development, drugs targeting MCAM are promising and meaningful, as they may act from multiple perspectives rather than on a specific aspect of tumors, potentially having strong antitumor effects.

4.5 ALCAM

ALCAM also called CD166, is a pivotal cell adhesion protein mediating migration, growth, and adhesiveness in cancer cells involving NF-κB, β-catenin, and Twist signaling, as well as has complicated clinical prognostic significance in different cancers (Yang et al., 2021). ALCAM is a transmembrane molecule divided into three parts: a large N-terminal extracellular domain (ED), a transmembrane region, and a short cytoplasmic domain. The ED has five Ig-like domains (V-V-C2-C2-C2) consisting of three membrane-proximal constant regions (C2) and two membrane-distal variable regions (V), containing 10 potential N-glycosylation sites. The Ig-like domains are mainly responsible for regulating the affinity of homophilic ALCAM-ALCAM and heterophilic ALCAM-CD6 interactions. Cell-cell adhesion is mediated by trans-ALCAM-ALCAM interactions, which touch the amino-terminal ligand-binding modules (V-domains) (von Lersner et al., 2019; Darvishi et al., 2020; Ferragut et al., 2021).

In non-small-cell lung cancer, ALCAM expression is significantly elevated and closely linked to short overall survival and poor prognosis. ALCAM contributes to brain metastasis by interacting with the brain vascular endothelium (Münsterberg et al., 2020). Simultaneously, ALCAM acts as a critical anti-apoptotic regulator by activating PI3K/Akt signaling, upregulating YAP protein expression, and downregulating Bcl-2 expression in liver cancer cells (Ma et al., 2014). ALCAM can also enhance the cancer stem cell-like (CSC) phenotype of nasopharyngeal carcinoma by activating the EGFR/ERK1/2 signaling pathway, and ALCAM downregulation decreases tumor sphere formation, thus inhibiting CSC-like characteristics and weakening tumorigenesis capacity in vivo (Chen X. et al., 2021). ALCAM is also associated with tamoxifen resistance in breast cancer cells, and ALCAM knockdown substantially enhances tamoxifen-induced cancer cell viability inhibition and apoptosis (Chen et al., 2017). Moreover, ALCAM mediates the resistance of prostate cancer to enzalutamide (ENZA), a second-generation androgen receptor inhibitor frequently used for treating metastatic castration-resistant prostate cancer and ALCAM gene silencing in drug-resistant cells could enhance ENZA sensitivity (Csizmarik et al., 2022). Drug research targeting ALCAM is relatively common, with most achieving ideal results; however, it is limited to laboratory studies and cannot be applied in clinical practice. Currently, there is an urgent need to use drugs with good therapeutic effects in clinics.

4.6 NPTN

NPTN or Np is a transmembrane glycoprotein belonging to the Ig superfamily of cell adhesion molecules and has three domains: extracellular Ig domains, transmembrane domains, and short cytoplasmic domains. NPTN has two splicing variants, NPTN α and NPTN β , also named Np55 and Np65 owing to their apparent molecular weights. In their extracellular structure, NPTN α has two Ig domains, while NPTN β has three, and the extra domain is useful to bind with S100A8 and Np65 homophilic interactions. Notably, S100A8 only binds to NPTN β , while S100A9 can bind to NPTN α (Owczarek and Berezin, 2012; Sakaguchi et al., 2016; Ilic et al., 2021).

In lung cancer cells, S100A8/A9 can bind to overexpressed NPTNB, activating transcription factors NFIA by TRAF2 and RAS signaling, and ultimately promotes cellular spheroidal growth, migration, and invasion through downstream molecules SAM pointed-domain containing E26 transformation-specific transcription factor (SPDEF) (Sumardika et al., 2019). SPDEF plays a vital role in NPTNB functions and the NPTNB-SPDEFinduced solute carrier family 22 member 18 antisense (SLC22A18AS) is essential for EMT and cellular motility in lung cancer, with a negative correlation with overall survival of patients (Bajkowska et al., 2020). Moreover, in breast cancer cells, overexpressed-NPTN contributes to tumor growth and angiogenesis in vivo, which is related to the increased VEGF production in microenvironmental hypoxia (Rodriguez-Pinto et al., 2009). Compared with that of other receptors, the functions and related mechanisms of NPTN in tumor cells are relatively less known, and further studies should aim to clarify the involvement of NPTN in the growth, migration, and angiogenesis of different cancers.

5 Potential S100A8/A9-targeting strategies to treat cancers

Since S100A8/A9 is a prominent mediator of cancers, targeting S100A8/A9 to explore novel strategies in tumor treatment is promising. S100A8/A9 can be used as an effective tool to detect early-phase inflammation and tissue damage by anticancer agents (Morikawa-Ichinose et al., 2022). Using specific antibodies, changes of S100A8/A9 in tumor-bearing mice can be monitored to evaluate the establishment of an immunosuppressive state and a metastasis-permissive microenvironment (Eisenblaetter et al., 2017). In addition, comparing the expression of S100A8 in tumor cells and immune cells at different stages can help determine the clinicopathological features and clinical outcomes of patients (Woo et al., 2021).

Considering the increasing plasma levels of S100A8/A9 in malignancies, blocking the interaction between S100A8/A9 and its receptors seems likely to be an efficacious therapeutic method (Figure 3). Oxyclozanide, a chemical probe and inhibitor of S100A9 and RAGEs, can inhibit tumor growth *in vivo* (Björk et al., 2013). Ab45, an S100A8/A9 heterodimer-specific neutralizing antibody, efficiently blocks melanoma lung metastasis (Kinoshita et al., 2019b). Meanwhile, quinoline-3-carboxamide tasquinimod, a strong S100A9 binder, can inhibit the establishment of primary tumors and hinder the formation of



neutralizing S100A8/A9 and blocking its binding with receptors. Current studies have focused on the most common receptors RAGEs and TLR4. These targets are promising diagnostic and prognostic biomarkers for various cancer types.

distant metastatic niches in prostate cancer, even breaking the angiogenic balance by downregulating VEGF and HIF-1 α (Jennbacken et al., 2012). The therapeutic peptides Pep-H6 and Pep-G3, which recognize extracellular S100A8/A9 on MDSCs surface, could completely deplete blood and intra-tumoral MDSCs, thus enhancing the antitumor immunity of tumorbearing mice and inhibiting tumor growth *in vivo*, indicating an emerging treatment direction for S100A8/A9 together with immune evasion (Qin et al., 2014).

Meantime, different S100A8/A9 receptors may serve as potential therapeutic targets (Figure 3). Baicalein directly binding to TLR4 could inhibit HIF-1a/VEGF signaling pathway to reduce the growth, angiogenesis, and metastasis of CRC cells (Chen M. et al., 2021). Meanwhile, paclitaxel can promote tumor regression via TLR4/NF-KB signaling in melanoma by inducing M1 macrophage polarization (Wanderley et al., 2018). Quercetin induces autophagy, apoptosis, and chemosensitivity by RAGE/PI3K/Akt/mTOR pathways in pancreatic cancer cells (Lan et al., 2019). Antibody-drug conjugates (ADCs) against RAGEs kill endometrial tumor cells both in vitro and in vivo (Healey et al., 2019). Since MCAM has a positive effect on cancer metastasis, studies have demonstrated that MCAM knockdown

can reduce lytic bone metastasis in preclinical mouse models of prostate cancer, and MCAM-targeted mAbs can effectively postpone intraosseous metastatic lesions (Zoni et al., 2019). Recently, chimeric antigen receptor (CAR)-T cells with remarkable clinical responses and high specificity have become a new research hotspot for cancer treatment (Sterner and Sterner, 2021). In osteosarcoma cells, modified ALCAM CAR-T cells exhibit specific and potent cytotoxicity, induce regression of orthotopic osteosarcoma *in vivo*, and have no cytotoxic activity against healthy tissues (Wang et al., 2019). Moreover, ALCAMspecific CAR-T cells can efficiently lyse CRC cells and show powerful cytotoxicity against CRC stem cells, demonstrating a feasible approach for reconstructing CAR-T cells with S100A8/ A9 and its receptors for cancer therapy (He et al., 2023).

Despite the endless emergence of drugs targeting S100A8/ A9 and its receptors, which substantially inhibit different characteristics of tumor cells, they are only at the level of theoretical research, regrettably, there are no drugs that can be applied in practical life. The main problem is that the functional explanation of S100A8/A9 and its receptors is incomplete, very few studies have focused on the relevant mechanisms to support its clinical application. Thus, there is an urgent need to deeper study S100A8/A9 and its receptors to identify new strategies.

6 Summary

S100 proteins are involved in various human physiological activities, such as energy metabolism, cellular composition, metal ion transportation, and signal transmission. S100A8/A9 is an inflammatory mediator that broadly participates in many diseases, and can act as a useful biomarker for diagnosis and prediction. S100A8/A9 can regulate the initiation, proliferation, invasion, metastasis, angiogenesis, chemoresistance, and apoptosis of tumor cells by binding with its receptors scattered in tumor cell surface, including RAGEs, TLR4, EMMPRIN, MCAM, ALCAM, and NPTN, mostly by activating the JNK, ERK, P38 MAPK, PI3K/ Akt, NF- κ B, TGF β , and STAT3 signal transduction pathways; thus, they can be potential targets for cancer treatments.

Currently, some tactics aimed at S100A8/A9 and its receptors in cancer treatments have been studied, such as chemical drugs, specific antibodies, antagonists, and peptides. However, further studies should aim to clarify the relevance and function of S100A8/A9 and its receptors in cancers or other serious disorders, such as the balance of promotion and inhibition to cancers, new effects on oncogenesis, crosstalk of different receptors and concrete mechanism, correlation with tumor microenvironment, synergistic or antagonism cooperation with inflammatory factors, as well as discovery of feasible drug targets, which could open a new door leading to a brand new era when cancer may be disappearing.

Author contributions

WZ and YX proposed the idea for this review; HZ wrote the first draft of this manuscript; WZ and YX critically reviewed the

References

Abu El-Asrar, A. M., Nawaz, M. I., Ahmad, A., Siddiquei, M. M., Allegaert, E., Gikandi, P. W., et al. (2021). CD146/Soluble CD146 pathway is a novel biomarker of angiogenesis and inflammation in proliferative diabetic retinopathy. *Invest. Ophthalmol. Vis. Sci.* 62 (9), 32. doi:10.1167/iovs.62.9.32

Ahmad, S., Khan, H., Siddiqui, Z., Khan, M. Y., Rehman, S., Shahab, U., et al. (2018). AGEs, RAGEs and s-RAGE; friend or foe for cancer. *Semin. Cancer Biol.* 49, 44–55. doi:10.1016/j.semcancer.2017.07.001

Bai, B., Xu, Y., and Chen, H. (2022). Pathogenic roles of neutrophil-derived alarmins (S100A8/A9) in heart failure: from molecular mechanisms to therapeutic insights. *Br. J. Pharmacol.* 180, 573–588. doi:10.1111/bph.15998

Bai, S., Wang, W., Ye, L., Fang, L., Dong, T., Zhang, R., et al. (2021). IL-17 stimulates neutrophils to release S100A8/A9 to promote lung epithelial cell apoptosis in Mycoplasma pneumoniae-induced pneumonia in children. *Biomed. Pharmacother*. 143, 112184. doi:10.1016/j.biopha.2021.112184

Bai, Y., Huang, W., Ma, L. T., Jiang, J. L., and Chen, Z. N. (2014). Importance of N-glycosylation on CD147 for its biological functions. *Int. J. Mol. Sci.* 15 (4), 6356–6377. doi:10.3390/ijms15046356

Bajkowska, K., Sumardika, I. W., Tomonobu, N., Chen, Y., Yamamoto, K. I., Kinoshita, R., et al. (2020). Neuroplastinf-mediated upregulation of solute carrier family 22 member 18 antisense (SLC22A18AS) plays a crucial role in the epithelialmesenchymal transition, leading to lung cancer cells' enhanced motility. *Biochem. Biophys. Rep.* 22, 100768. doi:10.1016/j.bbrep.2020.100768

Basso, D., Bozzato, D., Padoan, A., Moz, S., Zambon, C. F., Fogar, P., et al. (2014). Inflammation and pancreatic cancer: molecular and functional interactions between S100A8, S100A9, NT-S100A8 and TGF β 1. *Cell Commun. Signal* 12, 20. doi:10.1186/1478-811x-12-20

Basu, A., Munir, S., Mulaw, M. A., Singh, K., Crisan, D., Sindrilaru, A., et al. (2018). A novel S100A8/A9 induced fingerprint of mesenchymal stem cells associated with enhanced wound healing. *Sci. Rep.* 8 (1), 6205. doi:10.1038/s41598-018-24425-9

manuscript and added relevant information; RS and CZ contributed to the reference collection. All the authors have read and approved the final version of the manuscript.

Funding

This study was funded by the National Key Research and Development Program of China (2016YFA0201504), National Natural Science Foundation of China (Nos. 81473249 and 81102464), National Megaproject for Innovative Drugs (2014ZX09201042), CAMS Innovation Fund for Medical Sciences (CIFMS) (2021-I2M-1-030), and Drug Innovation Major Project of China (2018ZX09711001-007-002).

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Björk, P., Källberg, E., Wellmar, U., Riva, M., Olsson, A., He, Z., et al. (2013). Common interactions between S100A4 and S100A9 defined by a novel chemical probe. *PLoS One* 8 (5), e63012. doi:10.1371/journal.pone.0063012

Böttcher, M., Panagiotidis, K., Bruns, H., Stumpf, M., Völkl, S., Geyh, S., et al. (2022). Bone marrow stroma cells promote induction of a chemoresistant and prognostic unfavorable S100A8/A9high AML cell subset. *Blood Adv.* 6 (21), 5685–5697. doi:10. 1182/bloodadvances.2021005938

Bresnick, A. R., Weber, D. J., and Zimmer, D. B. (2015). S100 proteins in cancer. Nat. Rev. Cancer 15 (2), 96–109. doi:10.1038/nrc3893

Bu, X., Qu, X., Guo, K., Meng, X., Yang, X., Huang, Q., et al. (2021). CD147 confers temozolomide resistance of glioma cells via the regulation of β -TrCP/Nrf2 pathway. *Int. J. Biol. Sci.* 17 (12), 3013–3023. doi:10.7150/ijbs.60894

Burri, E., and Beglinger, C. (2014). The use of fecal calprotectin as a biomarker in gastrointestinal disease. *Expert Rev. Gastroenterol. Hepatol.* 8 (2), 197–210. doi:10.1586/17474124.2014.869476

Cai, Z., Xie, Q., Hu, T., Yao, Q., Zhao, J., Wu, Q., et al. (2020). S100A8/A9 in myocardial infarction: a promising biomarker and therapeutic target. *Front. Cell Dev. Biol.* 8, 603902. doi:10.3389/fcell.2020.603902

Chen, M. J., Cheng, Y. M., Chen, C. C., Chen, Y. C., and Shen, C. J. (2017). MiR-148a and miR-152 reduce tamoxifen resistance in ER+ breast cancer via downregulating ALCAM. *Biochem. Biophys. Res. Commun.* 483 (2), 840–846. doi:10.1016/j.bbrc.2017. 01.012

Chen, M., Zhong, K., Tan, J., Meng, M., Liu, C. M., Chen, B., et al. (2021a). Baicalein is a novel TLR4-targeting therapeutics agent that inhibits TLR4/HIF-1 α /VEGF signaling pathway in colorectal cancer. *Clin. Transl. Med.* 11 (11), e564. doi:10.1002/ctm2.564

Chen, X., Liang, R., Lin, H., Chen, K., Chen, L., Tian, G., et al. (2021b). CD166 promotes cancer stem cell-like phenotype via the EGFR/ERK1/2 pathway in the nasopharyngeal carcinoma cell line CNE-2R. *Life Sci.* 267, 118983. doi:10.1016/j.lfs. 2020.118983

Chen, Y., Sumardika, I. W., Tomonobu, N., Kinoshita, R., Inoue, Y., Iioka, H., et al. (2019a). Critical role of the MCAM-ETV4 axis triggered by extracellular S100A8/A9 in breast cancer aggressiveness. *Neoplasia* 21 (7), 627–640. doi:10.1016/j.neo.2019.04.006

Chen, Y., Sumardika, I. W., Tomonobu, N., Winarsa Ruma, I. M., Kinoshita, R., Kondo, E., et al. (2019b). Melanoma cell adhesion molecule is the driving force behind the dissemination of melanoma upon S100A8/A9 binding in the original skin lesion. *Cancer Lett.* 452, 178–190. doi:10.1016/j.canlet.2019.03.023

Chen, Y., Xu, J., Wu, X., Yao, H., Yan, Z., Guo, T., et al. (2021c). CD147 regulates antitumor CD8(+) T-cell responses to facilitate tumor-immune escape. *Cell Mol. Immunol.* 18 (8), 1995–2009. doi:10.1038/s41423-020-00570-y

Colicchia, M., Schrottmaier, W. C., Perrella, G., Reyat, J. S., Begum, J., Slater, A., et al. (2022). S100A8/A9 drives the formation of procoagulant platelets through GPIba. *Blood* 140 (24), 2626–2643. doi:10.1182/blood.2021014966

Cormier, K., Harquail, J., Ouellette, R. J., Tessier, P. A., Guerrette, R., and Robichaud, G. A. (2014). Intracellular expression of inflammatory proteins S100A8 and S100A9 leads to epithelial-mesenchymal transition and attenuated aggressivity of breast cancer cells. *Anticancer Agents Med. Chem.* 14 (1), 35–45. doi:10.2174/18715206113136660333

Csizmarik, A., Keresztes, D., Nagy, N., Bracht, T., Sitek, B., Witzke, K., et al. (2022). Proteome profiling of enzalutamide-resistant cell lines and serum analysis identified ALCAM as marker of resistance in castration-resistant prostate cancer. *Int. J. Cancer* 151 (8), 1405–1419. doi:10.1002/ijc.34159

Dana, P., Kariya, R., Lert-Itthiporn, W., Seubwai, W., Saisomboon, S., Wongkham, C., et al. (2021). Homophilic interaction of CD147 promotes IL-6-mediated cholangiocarcinoma invasion via the NF-kB-Dependent pathway. *Int. J. Mol. Sci.* 22 (24), 13496. doi:10.3390/ijms222413496

Darvishi, B., Boroumandieh, S., Majidzadeh, A. K., Salehi, M., Jafari, F., and Farahmand, L. (2020). The role of activated leukocyte cell adhesion molecule (ALCAM) in cancer progression, invasion, metastasis and recurrence: a novel cancer stem cell marker and tumor-specific prognostic marker. *Exp. Mol. Pathol.* 115, 104443. doi:10.1016/j.yexmp.2020.104443

Dattilo, B. M., Fritz, G., Leclerc, E., Kooi, C. W., Heizmann, C. W., and Chazin, W. J. (2007). The extracellular region of the receptor for advanced glycation end products is composed of two independent structural units. *Biochemistry* 46 (23), 6957–6970. doi:10. 1021/bi7003735

De Veirman, K., De Beule, N., Maes, K., Menu, E., De Bruyne, E., De Raeve, H., et al. (2017). Extracellular \$100A9 protein in bone marrow supports multiple myeloma survival by stimulating angiogenesis and cytokine secretion. *Cancer Immunol. Res.* 5 (10), 839–846. doi:10.1158/2326-6066.cir-17-0192

Donato, R. (1999). Functional roles of S100 proteins, calcium-binding proteins of the EFhand type. *Biochim. Biophys. Acta* 1450 (3), 191–231. doi:10.1016/s0167-4889(99)00058-0

Donato, R. (2001). S100: a multigenic family of calcium-modulated proteins of the EF-hand type with intracellular and extracellular functional roles. *Int. J. Biochem. Cell Biol.* 33 (7), 637–668. doi:10.1016/s1357-2725(01)00046-2

Edgeworth, J., Gorman, M., Bennett, R., Freemont, P., and Hogg, N. (1991). Identification of p8,14 as a highly abundant heterodimeric calcium binding protein complex of myeloid cells. *J. Biol. Chem.* 266 (12), 7706–7713. doi:10.1016/s0021-9258(20)89506-4

Ehrchen, J. M., Sunderkötter, C., Foell, D., Vogl, T., and Roth, J. (2009). The endogenous Toll-like receptor 4 agonist S100A8/S100A9 (calprotectin) as innate amplifier of infection, autoimmunity, and cancer. J. Leukoc. Biol. 86 (3), 557–566. doi:10.1189/jlb.1008647

Eisenblaetter, M., Flores-Borja, F., Lee, J. J., Wefers, C., Smith, H., Hueting, R., et al. (2017). Visualization of tumor-immune interaction - target-specific imaging of S100a8/ A9 reveals pre-metastatic niche establishment. *Theranostics* 7 (9), 2392–2401. doi:10. 7150/thno.17138

El-Far, A. H., Sroga, G., Jaouni, S. K. A., Mousa, S. A., Elewa, Y. H. A., Elfeky, M., et al. (2020). Quercetin attenuates pancreatic and renal D-galactose-induced aging-related oxidative alterations in rats. *Int. J. Mol. Sci.* 21 (10), 4348. doi:10.3390/ijms21124348

Fan, B., Zhang, L. H., Jia, Y. N., Zhong, X. Y., Liu, Y. Q., Cheng, X. J., et al. (2012). Presence of \$100A9-positive inflammatory cells in cancer tissues correlates with an early stage cancer and a better prognosis in patients with gastric cancer. *BMC Cancer* 12, 316. doi:10.1186/1471-2407-12-316

Fang, W. Y., Chen, Y. W., Hsiao, J. R., Liu, C. S., Kuo, Y. Z., Wang, Y. C., et al. (2015). Elevated S100A9 expression in tumor stroma functions as an early recurrence marker for early-stage oral cancer patients through increased tumor cell invasion, angiogenesis, macrophage recruitment and interleukin-6 production. *Oncotarget 6* (29), 28401–28424. doi:10.18632/oncotarget.4951

Ferragut, F., Vachetta, V. S., Troncoso, M. F., Rabinovich, G. A., and Elola, M. T. (2021). ALCAM/CD166: a pleiotropic mediator of cell adhesion, stemness and cancer progression. *Cytokine Growth Factor Rev.* 61, 27–37. doi:10.1016/j.cytogfr.2021.07.001

Fontaine, M., Planel, S., Peronnet, E., Turrel-Davin, F., Piriou, V., Pachot, A., et al. (2014). S100A8/A9 mRNA induction in an *ex vivo* model of endotoxin tolerance: roles of IL-10 and IFNγ. *PLoS One* 9 (6), e100909. doi:10.1371/journal.pone.0100909

Ghavami, S., Chitayat, S., Hashemi, M., Eshraghi, M., Chazin, W. J., Halayko, A. J., et al. (2009). S100A8/A9: a janus-faced molecule in cancer therapy and tumorgenesis. *Eur. J. Pharmacol.* 625 (1-3), 73–83. doi:10.1016/j.ejphar.2009.08.044 Giudice, V., Wu, Z., Kajigaya, S., Fernandez Ibanez, M. D. P., Rios, O., Cheung, F., et al. (2019). Circulating S100A8 and S100A9 protein levels in plasma of patients with acquired aplastic anemia and myelodysplastic syndromes. *Cytokine* 113, 462–465. doi:10.1016/j.cyto.2018.06.025

Goebeler, M., Roth, J., van den Bos, C., Ader, G., and Sorg, C. (1995). Increase of calcium levels in epithelial cells induces translocation of calcium-binding proteins migration inhibitory factor-related protein 8 (MRP8) and MRP14 to keratin intermediate filaments. *Biochem. J.* 309 (2), 419–424. doi:10.1042/bj3090419

Gonzalez, L. L., Garrie, K., and Turner, M. D. (2020). Role of S100 proteins in health and disease. *Biochim. Biophys. Acta Mol. Cell Res.* 1867 (6), 118677. doi:10.1016/j. bbamcr.2020.118677

Grebhardt, S., Veltkamp, C., Ströbel, P., and Mayer, D. (2012). Hypoxia and HIF-1 increase S100A8 and S100A9 expression in prostate cancer. *Int. J. Cancer* 131 (12), 2785–2794. doi:10.1002/ijc.27591

Grivennikov, S. I., Greten, F. R., and Karin, M. (2010). Immunity, inflammation, and cancer. *Cell* 140 (6), 883–899. doi:10.1016/j.cell.2010.01.025

Gross, S. R., Sin, C. G., Barraclough, R., and Rudland, P. S. (2014). Joining S100 proteins and migration: for better or for worse, in sickness and in health. *Cell Mol. Life Sci.* 71 (9), 1551–1579. doi:10.1007/s00018-013-1400-7

Guignard, F., Mauel, J., and Markert, M. (1996). Phosphorylation of myeloid-related proteins MRP-14 and MRP-8 during human neutrophil activation. *Eur. J. Biochem.* 241 (1), 265–271. doi:10.1111/j.1432-1033.1996.0265t.x

Guo, Q., Zhao, Y., Li, J., Liu, J., Yang, X., Guo, X., et al. (2021). Induction of alarmin S100A8/A9 mediates activation of aberrant neutrophils in the pathogenesis of COVID-19. *Cell Host Microbe* 29 (2), 222–235.e4. doi:10.1016/j.chom.2020.12.016

Harrison, C. A., Raftery, M. J., Walsh, J., Alewood, P., Iismaa, S. E., Thliveris, S., et al. (1999). Oxidation regulates the inflammatory properties of the murine \$100 protein \$100A8. *J. Biol. Chem.* 274 (13), 8561–8569. doi:10.1074/jbc.274.13.8561

Hayashi, N., Kido, J., Kido, R., Wada, C., Kataoka, M., Shinohara, Y., et al. (2007). Regulation of calprotectin expression by interleukin-1alpha and transforming growth factor-beta in human gingival keratinocytes. *J. Periodontal Res.* 42 (1), 1–7. doi:10.1111/j.1600-0765.2005.00857.x

He, S., Li, S., Guo, J., Zeng, X., Liang, D., Zhu, Y., et al. (2023). CD166-specific CAR-T cells potently target colorectal cancer cells. *Transl. Oncol.* 27, 101575. doi:10.1016/j. tranon.2022.101575

Healey, G. D., Pan-Castillo, B., Garcia-Parra, J., Davies, J., Roberts, S., Jones, E., et al. (2019). Antibody drug conjugates against the receptor for advanced glycation end products (RAGE), a novel therapeutic target in endometrial cancer. *J. Immunother. Cancer* 7 (1), 280. doi:10.1186/s40425-019-0765-z

Heinz, R., and Schneider, U. C. (2022). TLR4-Pathway-Associated biomarkers in subarachnoid hemorrhage (SAH): potential targets for future anti-inflammatory therapies. *Int. J. Mol. Sci.* 23 (20), 12618. doi:10.3390/ijms232012618

Herik Rodrigo, A. G., Tomonobu, N., Yoneda, H., Kinoshita, R., Mitsui, Y., Sadahira, T., et al. (2022). Toll-like receptor 4 promotes bladder cancer progression upon S100.88/ A9 binding, which requires TIRAP-mediated TPL2 activation. *Biochem. Biophys. Res. Commun.* 634, 83–91. doi:10.1016/j.bbrc.2022.09.116

Hibino, T., Sakaguchi, M., Miyamoto, S., Yamamoto, M., Motoyama, A., Hosoi, J., et al. (2013). S100A9 is a novel ligand of EMMPRIN that promotes melanoma metastasis. *Cancer Res.* 73 (1), 172–183. doi:10.1158/0008-5472.can-11-3843

Hirono, K., Foell, D., Xing, Y., Miyagawa-Tomita, S., Ye, F., Ahlmann, M., et al. (2006). Expression of myeloid-related protein-8 and -14 in patients with acute Kawasaki disease. *J. Am. Coll. Cardiol.* 48 (6), 1257–1264. doi:10.1016/j.jacc.2006.02.077

Hou, J., Karin, M., and Sun, B. (2021). Targeting cancer-promoting inflammation - have anti-inflammatory therapies come of age? *Nat. Rev. Clin. Oncol.* 18 (5), 261–279. doi:10.1038/s41571-020-00459-9

Hsu, Y. L., Hung, J. Y., Tsai, E. M., Wu, C. Y., Ho, Y. W., Jian, S. F., et al. (2015). Benzyl butyl phthalate increases the chemoresistance to doxorubicin/cyclophosphamide by increasing breast cancer-associated dendritic cell-derived CXCL1/GRO α and S100A8/ A9. Oncol. Rep. 34 (6), 2889–2900. doi:10.3892/or.2015.4307

Hu, W., Tao, Z., Zhou, Q., Zhao, D., Gu, L., Zhu, S., et al. (2021). Effects of S100 calcium-binding protein A8 (S100A8) and S100 calcium-binding protein A9 (S100A9) on matrix metalloproteinase (MMP) expression in nasopharyngeal carcinoma CNE-2 cells. *Transl. Cancer Res.* 10 (4), 1874–1884. doi:10.21037/tcr-21-441

Huang, M., Wu, R., Chen, L., Peng, Q., Li, S., Zhang, Y., et al. (2019). S100A9 regulates MDSCs-mediated immune suppression via the RAGE and TLR4 signaling pathways in colorectal carcinoma. *Front. Immunol.* 10, 2243. doi:10.3389/fimmu.2019.02243

Hurnakova, J., Zavada, J., Hanova, P., Hulejova, H., Klein, M., Mann, H., et al. (2015). Serum calprotectin (S100a8/9): an independent predictor of ultrasound synovitis in patients with rheumatoid arthritis. *Arthritis Res. Ther.* 17 (1), 252. doi:10.1186/s13075-015-0764-5

Ichikawa, M., Williams, R., Wang, L., Vogl, T., and Srikrishna, G. (2011). S100A8/ A9 activate key genes and pathways in colon tumor progression. *Mol. Cancer Res.* 9 (2), 133–148. doi:10.1158/1541-7786.mcr-10-0394

Ilic, K., Mlinac-Jerkovic, K., Sedmak, G., Rosenzweig, I., and Kalanj-Bognar, S. (2021). Neuroplastin in human cognition: review of literature and future perspectives. *Transl. Psychiatry* 11 (1), 394. doi:10.1038/s41398-021-01509-1 Iotzova-Weiss, G., Dziunycz, P. J., Freiberger, S. N., Läuchli, S., Hafner, J., Vogl, T., et al. (2015). S100A8/A9 stimulates keratinocyte proliferation in the development of squamous cell carcinoma of the skin via the receptor for advanced glycation-end products. *PLoS One* 10 (3), e0120971. doi:10.1371/journal.pone.0120971

Jennbacken, K., Welén, K., Olsson, A., Axelsson, B., Törngren, M., Damber, J. E., et al. (2012). Inhibition of metastasis in a castration resistant prostate cancer model by the quinoline-3-carboxamide tasquinimod (ABR-215050). *Prostate* 72 (8), 913–924. doi:10. 1002/pros.21495

Jiang, T., Zhuang, J., Duan, H., Luo, Y., Zeng, Q., Fan, K., et al. (2012). CD146 is a coreceptor for VEGFR-2 in tumor angiogenesis. *Blood* 120 (11), 2330–2339. doi:10. 1182/blood-2012-01-406108

Jukic, A., Bakiri, L., Wagner, E. F., Tilg, H., and Adolph, T. E. (2021). Calprotectin: from biomarker to biological function. *Gut* 70 (10), 1978–1988. doi:10.1136/gutjnl-2021-324855

Kang, R., Hou, W., Zhang, Q., Chen, R., Lee, Y. J., Bartlett, D. L., et al. (2014). RAGE is essential for oncogenic KRAS-mediated hypoxic signaling in pancreatic cancer. *Cell Death Dis.* 5 (10), e1480. doi:10.1038/cddis.2014.445

Kannan, S. (2003). Inflammation: a novel mechanism for the transport of extracellular nucleotide-induced arachidonic acid by S100a8/A9 for transcellular metabolism. *Cell Biol. Int.* 27 (7), 593–595. doi:10.1016/s1065-6995(03)00082-9

Kashani, B., Zandi, Z., Pourbagheri-Sigaroodi, A., Bashash, D., and Ghaffari, S. H. (2021). The role of toll-like receptor 4 (TLR4) in cancer progression: a possible therapeutic target? *J. Cell Physiol.* 236 (6), 4121–4137. doi:10.1002/jcp.30166

Kerkhoff, C., Klempt, M., Kaever, V., and Sorg, C. (1999). The two calcium-binding proteins, S100A8 and S100A9, are involved in the metabolism of arachidonic acid in human neutrophils. *J. Biol. Chem.* 274 (46), 32672–32679. doi:10.1074/jbc.274.46.32672

Kerkhoff, C., Klempt, M., and Sorg, C. (1998). Novel insights into structure and function of MRP8 (S100A8) and MRP14 (S100A9). *Biochim. Biophys. Acta* 1448 (2), 200–211. doi:10.1016/s0167-4889(98)00144-x

Kerkhoff, C., Nacken, W., Benedyk, M., Dagher, M. C., Sopalla, C., and Doussiere, J. (2005). The arachidonic acid-binding protein S100A8/A9 promotes NADPH oxidase activation by interaction with p67phox and Rac-2. *Faseb J*. 19 (3), 467–469. doi:10.1096/fj.04-2377fje

Kerkhoff, C., Sorg, C., Tandon, N. N., and Nacken, W. (2001). Interaction of S100A8/ S100A9-arachidonic acid complexes with the scavenger receptor CD36 may facilitate fatty acid uptake by endothelial cells. *Biochemistry* 40 (1), 241–248. doi:10.1021/ bi001791k

Kim, W. T., Kim, J., Yan, C., Jeong, P., Choi, S. Y., Lee, O. J., et al. (2014). S100A9 and EGFR gene signatures predict disease progression in muscle invasive bladder cancer patients after chemotherapy. *Ann. Oncol.* 25 (5), 974–979. doi:10.1093/annonc/mdu037

Kinoshita, R., Sato, H., Yamauchi, A., Takahashi, Y., Inoue, Y., Sumardika, I. W., et al. (2019a). exSSSRs (extracellular S100 soil sensor receptors)-Fc fusion proteins work as prominent decoys to S100A8/A9-induced lung tropic cancer metastasis. *Int. J. Cancer* 144 (12), 3138–3145. doi:10.1002/ijc.31945

Kinoshita, R., Sato, H., Yamauchi, A., Takahashi, Y., Inoue, Y., Sumardika, I. W., et al. (2019b). Newly developed anti-S100A8/A9 monoclonal antibody efficiently prevents lung tropic cancer metastasis. *Int. J. Cancer* 145 (2), 569–575. doi:10.1002/ijc.31982

Koh, H. M., An, H. J., Ko, G. H., Lee, J. H., Lee, J. S., Kim, D. C., et al. (2019). Prognostic role of S100A9 expression in patients with clear cell renal cell carcinoma. *Med. Baltim.* 98 (40), e17188. doi:10.1097/md.000000000017188

Koh, H. M., Lee, H. J., and Kim, D. C. (2021). High expression of S100A8 and S100A9 is associated with poor disease-free survival in patients with cancer: a systematic review and meta-analysis. *Transl. Cancer Res.* 10 (7), 3225–3235. doi:10.21037/tcr-21-519

Kong, C., Yan, X., Zhu, Y., Zhu, H., Luo, Y., Liu, P., et al. (2021). Fusobacterium nucleatum promotes the development of colorectal cancer by activating a cytochrome P450/epoxyoctadecenoic acid Axis via TLR4/keap1/NRF2 signaling. *Cancer Res.* 81 (17), 4485–4498. doi:10.1158/0008-5472.can-21-0453

Korndörfer, I. P., Brueckner, F., and Skerra, A. (2007). The crystal structure of the human (S100A8/S100A9)2 heterotetramer, calprotectin, illustrates how conformational changes of interacting alpha-helices can determine specific association of two EF-hand proteins. *J. Mol. Biol.* 370 (5), 887–898. doi:10.1016/j.jmb.2007.04.065

Kwon, C. H., Moon, H. J., Park, H. J., Choi, J. H., and Park, D. Y. (2013). S100A8 and S100A9 promotes invasion and migration through p38 mitogen-activated protein kinase-dependent NF-κB activation in gastric cancer cells. *Mol. Cells* 35 (3), 226–234. doi:10.1007/s10059-013-2269-x

Lan, C. Y., Chen, S. Y., Kuo, C. W., Lu, C. C., and Yen, G. C. (2019). Quercetin facilitates cell death and chemosensitivity through RAGE/PI3K/AKT/mTOR axis in human pancreatic cancer cells. *J. Food Drug Anal.* 27 (4), 887–896. doi:10.1016/j.jfda. 2019.07.001

Lee, J. S., Lee, N. R., Kashif, A., Yang, S. J., Nam, A. R., Song, I. C., et al. (2020). S100A8 and S100A9 promote apoptosis of chronic eosinophilic leukemia cells. *Front. Immunol.* 11, 1258. doi:10.3389/fimmu.2020.01258

Lei, Q., Wang, D., Sun, K., Wang, L., and Zhang, Y. (2020). Resistance mechanisms of anti-PD1/PDL1 therapy in solid tumors. *Front. Cell Dev. Biol.* 8, 672. doi:10.3389/fcell. 2020.00672

Lei, R., Vu, B., Kourentzi, K., Soomro, S., Danthanarayana, A. N., Brgoch, J., et al. (2022). A novel technology for home monitoring of lupus nephritis that tracks the pathogenic urine biomarker ALCAM. *Front. Immunol.* 13, 1044743. doi:10.3389/fimmu.2022.1044743

Lei, X., Guan, C. W., Song, Y., and Wang, H. (2015). The multifaceted role of CD146/ MCAM in the promotion of melanoma progression. *Cancer Cell Int.* 15 (1), 3. doi:10. 1186/s12935-014-0147-z

Lei, X., Wang, K., Wang, W., Jin, H., Gu, W., Chen, Z., et al. (2021). Recognize the role of CD146/MCAM in the osteosarcoma progression: an *in vitro* study. *Cancer Cell Int.* 21 (1), 300. doi:10.1186/s12935-021-02006-7

Lemarchand, P., Vaglio, M., Mauël, J., and Markert, M. (1992). Translocation of a small cytosolic calcium-binding protein (MRP-8) to plasma membrane correlates with human neutrophil activation. *J. Biol. Chem.* 267 (27), 19379–19382. doi:10.1016/s0021-9258(18)41786-3

Leroyer, A. S., Blin, M. G., Bachelier, R., Bardin, N., Blot-Chabaud, M., and Dignat-George, F. (2019). CD146 (cluster of differentiation 146). *Arterioscler. Thromb. Vasc. Biol.* 39 (6), 1026–1033. doi:10.1161/atvbaha.119.312653

Leukert, N., Vogl, T., Strupat, K., Reichelt, R., Sorg, C., and Roth, J. (2006). Calciumdependent tetramer formation of \$100A8 and \$100A9 is essential for biological activity. *J. Mol. Biol.* 359 (4), 961–972. doi:10.1016/j.jmb.2006.04.009

Levy, J. M. M., Towers, C. G., and Thorburn, A. (2017). Targeting autophagy in cancer. Nat. Rev. Cancer 17 (9), 528-542. doi:10.1038/nrc.2017.53

Li, C., Li, S., Jia, C., Yang, L., Song, Z., and Wang, Y. (2012). Low concentration of S100a8/9 promotes angiogenesis-related activity of vascular endothelial cells: bridges among inflammation, angiogenesis, and tumorigenesis? *Mediat. Inflamm.* 2012, 248574. doi:10.1155/2012/248574

Li, J., Huang, Q., Long, X., Zhang, J., Huang, X., Aa, J., et al. (2015). CD147 reprograms fatty acid metabolism in hepatocellular carcinoma cells through Akt/mTOR/SREBP1c and P38/PPARa pathways. *J. Hepatol.* 63 (6), 1378–1389. doi:10. 1016/j.jhep.2015.07.039

Li, J., Shu, X., Xu, J., Su, S. M., Chan, U. I., Mo, L., et al. (2022). S100A9-CXCL12 activation in BRCA1-mutant breast cancer promotes an immunosuppressive microenvironment associated with resistance to immunotherapy. *Nat. Commun.* 13 (1), 1481. doi:10.1038/s41467-022-29151-5

Li, S., Zhang, J., Qian, S., Wu, X., Sun, L., Ling, T., et al. (2021). S100A8 promotes epithelial-mesenchymal transition and metastasis under TGF- β /USF2 axis in colorectal cancer. *Cancer Commun. (Lond)* 41 (2), 154–170. doi:10.1002/cac2. 12130

Li, Z., Wang, J., Zhang, X., Liu, P., Zhang, X., Wang, J., et al. (2020). Proinflammatory S100A8 induces PD-L1 expression in macrophages, mediating tumor immune escape. *J. Immunol.* 204 (9), 2589–2599. doi:10.4049/jimmunol.1900753

Libby, P., and Kobold, S. (2019). Inflammation: a common contributor to cancer, aging, and cardiovascular diseases-expanding the concept of cardio-oncology. *Cardiovasc Res.* 115 (5), 824–829. doi:10.1093/cvr/cvz058

Liu, K., Zhang, Y., Zhang, C., Zhang, Q., Li, J., Xiao, F., et al. (2016a). Methylation of S100A8 is a promising diagnosis and prognostic marker in hepatocellular carcinoma. *Oncotarget* 7 (35), 56798–56810. doi:10.18632/oncotarget.10792

Liu, L., Liu, S., Deng, P., Liang, Y., Xiao, R., Tang, L. Q., et al. (2021). Targeting the IRAK1-S100A9 Axis overcomes resistance to paclitaxel in nasopharyngeal carcinoma. *Cancer Res.* 81 (5), 1413–1425. doi:10.1158/0008-5472.can-20-2125

Liu, T., Wang, G., Tao, H., Yang, Z., Wang, Y., Meng, Z., et al. (2016b). Capsaicin mediates caspases activation and induces apoptosis through P38 and JNK MAPK pathways in human renal carcinoma. *BMC Cancer* 16 (1), 790. doi:10.1186/s12885-016-2831-v

Lood, C., Tydén, H., Gullstrand, B., Jönsen, A., Källberg, E., Mörgelin, M., et al. (2016). Platelet-derived \$100a8/A9 and cardiovascular disease in systemic lupus erythematosus. *Arthritis Rheumatol.* 68 (8), 1970–1980. doi:10.1002/art.39656

Ma, L., Wang, J., Lin, J., Pan, Q., Yu, Y., and Sun, F. (2014). Cluster of differentiation 166 (CD166) regulated by phosphatidylinositide 3-Kinase (PI3K)/AKT signaling to exert its anti-apoptotic role via yes-associated protein (YAP) in liver cancer. J. Biol. Chem. 289 (10), 6921-6933. doi:10.1074/jbc.M113.524819

Machahua, C., Montes-Worboys, A., Planas-Cerezales, L., Buendia-Flores, R., Molina-Molina, M., and Vicens-Zygmunt, V. (2018). Serum AGE/RAGEs as potential biomarker in idiopathic pulmonary fibrosis. *Respir. Res.* 19 (1), 215. doi:10.1186/s12931-018-0924-7

Matsumoto, S., Yoshida, T., Murata, H., Harada, S., Fujita, N., Nakamura, S., et al. (2008). Solution structure of the variable-type domain of the receptor for advanced glycation end products: new insight into AGE-RAGE interaction. *Biochemistry* 47 (47), 12299–12311. doi:10.1021/bi800910v

Matsunaga, Y., Hashimoto, Y., and Ishiko, A. (2021). Stratum corneum levels of calprotectin proteins S100A8/A9 correlate with disease activity in psoriasis patients. *J. Dermatol* 48 (10), 1518–1525. doi:10.1111/1346-8138.16032

Mishra, V., and Pathak, C. (2019). Human Toll-Like Receptor 4 (hTLR4): structural and functional dynamics in cancer. *Int. J. Biol. Macromol.* 122, 425–451. doi:10.1016/j. ijbiomac.2018.10.142

Mohr, T., Zwick, A., Hans, M. C., Bley, I. A., Braun, F. L., Khalmurzaev, O., et al. (2022). The prominent role of the S100A8/S100A9-CD147 axis in the progression of penile cancer. *Front. Oncol.* 12, 891511. doi:10.3389/fonc.2022.891511

Mondet, J., Chevalier, S., and Mossuz, P. (2021). Pathogenic roles of S100A8 and S100A9 proteins in acute myeloid and lymphoid leukemia: clinical and therapeutic impacts. *Molecules* 26 (5), 1323. doi:10.3390/molecules26051323

Monteiro, C., Miarka, L., Perea-García, M., Priego, N., García-Gómez, P., Álvaro-Espinosa, L., et al. (2022). Stratification of radiosensitive brain metastases based on an actionable S100A9/RAGE resistance mechanism. *Nat. Med.* 28 (4), 752–765. doi:10. 1038/s41591-022-01749-8

Morikawa-Ichinose, T., Fujimura, Y., Kumazoe, M., Onda, H., Miura, D., and Tachibana, H. (2022). Inflammatory markers S100A8/A9 and metabolic alteration for evaluating signs of early phase toxicity of anticancer agent treatment. *Food Chem. Toxicol.* 169, 113421. doi:10.1016/j.fct.2022.113421

Mose, M., Kang, Z., Raaby, L., Iversen, L., and Johansen, C. (2013). TNFa- and IL-17A-mediated S100A8 expression is regulated by p38 MAPK. *Exp. Dermatol* 22 (7), 476–481. doi:10.1111/exd.12187

Münsterberg, J., Loreth, D., Brylka, L., Werner, S., Karbanova, J., Gandrass, M., et al. (2020). ALCAM contributes to brain metastasis formation in non-small-cell lung cancer through interaction with the vascular endothelium. *Neuro Oncol.* 22 (7), 955–966. doi:10.1093/neuonc/noaa028

Nguyen, Y. T. M., Fujisawa, M., Nguyen, T. B., Suehara, Y., Sakamoto, T., Matsuoka, R., et al. (2021). Tet2 deficiency in immune cells exacerbates tumor progression by increasing angiogenesis in a lung cancer model. *Cancer Sci.* 112 (12), 4931–4943. doi:10.1111/cas.15165

Nicaise, C., Weichselbaum, L., Schandene, L., Gangji, V., Dehavay, F., Bouchat, J., et al. (2017). Phagocyte-specific S100A8/A9 is upregulated in primary Sjögren's syndrome and triggers the secretion of pro-inflammatory cytokines *in vitro*. *Clin. Exp. Rheumatol.* 35 (1), 129–136.

Oesterle, A., and Bowman, M. A. (2015). S100A12 and the S100/calgranulins: emerging biomarkers for atherosclerosis and possibly therapeutic targets. *Arterioscler. Thromb. Vasc. Biol.* 35 (12), 2496–2507. doi:10.1161/atvbaha.115.302072

Ortega, F. J., Sabater, M., Moreno-Navarrete, J. M., Pueyo, N., Botas, P., Delgado, E., et al. (2012). Serum and urinary concentrations of calprotectin as markers of insulin resistance and type 2 diabetes. *Eur. J. Endocrinol.* 167 (4), 569–578. doi:10.1530/eje-12-0374

Ott, C., Jacobs, K., Haucke, E., Navarrete Santos, A., Grune, T., and Simm, A. (2014). Role of advanced glycation end products in cellular signaling. *Redox Biol.* 2, 411–429. doi:10.1016/j.redox.2013.12.016

Owczarek, S., and Berezin, V. (2012). Neuroplastin: cell adhesion molecule and signaling receptor. *Int. J. Biochem. Cell Biol.* 44 (1), 1–5. doi:10.1016/j.biocel.2011.10.006

Pan, S., Hu, Y., Hu, M., Xu, Y., Chen, M., Du, C., et al. (2020). S100A8 facilitates cholangiocarcinoma metastasis via upregulation of VEGF through TLR4/NF- κ B pathway activation. *Int. J. Oncol.* 56 (1), 101–112. doi:10.3892/ijo.2019.4907

Pruenster, M., Vogl, T., Roth, J., and Sperandio, M. (2016). S100A8/A9: from basic science to clinical application. *Pharmacol. Ther.* 167, 120–131. doi:10.1016/j. pharmthera.2016.07.015

Qin, F., Song, Y., Li, Z., Zhao, L., Zhang, Y., and Geng, L. (2010). S100A8/A9 induces apoptosis and inhibits metastasis of CasKi human cervical cancer cells. *Pathol. Oncol. Res.* 16 (3), 353–360. doi:10.1007/s12253-009-9225-2

Qin, H., Lerman, B., Sakamaki, I., Wei, G., Cha, S. C., Rao, S. S., et al. (2014). Generation of a new therapeutic peptide that depletes myeloid-derived suppressor cells in tumor-bearing mice. *Nat. Med.* 20 (6), 676–681. doi:10.1038/nm.3560

Quail, D. F., and Joyce, J. A. (2013). Microenvironmental regulation of tumor progression and metastasis. *Nat. Med.* 19 (11), 1423–1437. doi:10.1038/nm.3394

Rammes, A., Roth, J., Goebeler, M., Klempt, M., Hartmann, M., and Sorg, C. (1997). Myeloid-related protein (MRP) 8 and MRP14, calcium-binding proteins of the S100 family, are secreted by activated monocytes via a novel, tubulin-dependent pathway. J. Biol. Chem. 272 (14), 9496–9502. doi:10.1074/jbc.272.14.9496

Rapanotti, M. C., Cugini, E., Nuccetelli, M., Terrinoni, A., Di Raimondo, C., Lombardo, P., et al. (2021). MCAM/MUC18/CD146 as a multifaceted warning marker of melanoma progression in liquid biopsy. *Int. J. Mol. Sci.* 22 (22), 12416. doi:10.3390/ijms222212416

Reeb, A. N., Li, W., Sewell, W., Marlow, L. A., Tun, H. W., Smallridge, R. C., et al. (2015). S100A8 is a novel therapeutic target for anaplastic thyroid carcinoma. *J. Clin. Endocrinol. Metab.* 100 (2), E232–E242. doi:10.1210/jc.2014-2988

Rigiracciolo, D. C., Nohata, N., Lappano, R., Cirillo, F., Talia, M., Adame-Garcia, S. R., et al. (2022). Focal Adhesion Kinase (FAK)-Hippo/YAP transduction signaling mediates the stimulatory effects exerted by S100A8/A9-RAGE system in triple-negative breast cancer (TNBC). J. Exp. Clin. Cancer Res. 41 (1), 193. doi:10.1186/s13046-022-02396-0

Rodriguez-Pinto, D., Sparkowski, J., Keough, M. P., Phoenix, K. N., Vumbaca, F., Han, D. K., et al. (2009). Identification of novel tumor antigens with patient-derived immune-selected antibodies. *Cancer Immunol. Immunother.* 58 (2), 221–234. doi:10. 1007/s00262-008-0543-0

Roth, J., Burwinkel, F., van den Bos, C., Goebeler, M., Vollmer, E., and Sorg, C. (1993). MRP8 and MRP14, S-100-like proteins associated with myeloid differentiation, are translocated to plasma membrane and intermediate filaments in a calcium-dependent manner. *Blood* 82 (6), 1875–1883. doi:10.1182/blood.v82.6.1875.1875 Ruma, I. M., Putranto, E. W., Kondo, E., Murata, H., Watanabe, M., Huang, P., et al. (2016). MCAM, as a novel receptor for S100A8/A9, mediates progression of malignant melanoma through prominent activation of NF-κB and ROS formation upon ligand binding. *Clin. Exp. Metastasis* 33 (6), 609–627. doi:10.1007/s10585-016-9801-2

Ryckman, C., Vandal, K., Rouleau, P., Talbot, M., and Tessier, P. A. (2003). Proinflammatory activities of S100: proteins S100A8, S100A9, and S100a8/ A9 induce neutrophil chemotaxis and adhesion. *J. Immunol.* 170 (6), 3233–3242. doi:10.4049/jimmunol.170.6.3233

Sakaguchi, M., Yamamoto, M., Miyai, M., Maeda, T., Hiruma, J., Murata, H., et al. (2016). Identification of an S100A8 receptor neuroplastin- β and its heterodimer formation with EMMPRIN. *J. Invest. Dermatol* 136 (11), 2240–2250. doi:10.1016/j. jid.2016.06.617

Schiopu, A., and Cotoi, O. S. (2013). S100A8 and S100A9: dAMPs at the crossroads between innate immunity, traditional risk factors, and cardiovascular disease. *Mediat. Inflamm.* 2013, 828354. doi:10.1155/2013/828354

Scott, N. R., Swanson, R. V., Al-Hammadi, N., Domingo-Gonzalez, R., Rangel-Moreno, J., Kriel, B. A., et al. (2020). S100A8/A9 regulates CD11b expression and neutrophil recruitment during chronic tuberculosis. *J. Clin. Invest.* 130 (6), 3098–3112. doi:10.1172/jci130546

Shi, Y. J., Zhao, Q. Q., Liu, X. S., Dong, S. H., E, J. F., Li, X., et al. (2020). Toll-like receptor 4 regulates spontaneous intestinal tumorigenesis by up-regulating IL-6 and GM-CSF. J. Cell Mol. Med. 24 (1), 385–397. doi:10.1111/jcmm.14742

Silva, E. J., Argyris, P. P., Zou, X., Ross, K. F., and Herzberg, M. C. (2014). S100A8/ A9 regulates MMP-2 expression and invasion and migration by carcinoma cells. *Int. J. Biochem. Cell Biol.* 55, 279–287. doi:10.1016/j.biocel.2014.09.007

Sprenkeler, E. G. G., Zandstra, J., van Kleef, N. D., Goetschalckx, I., Verstegen, B., Aarts, C. E. M., et al. (2022). S100A8/A9 is a marker for the release of neutrophil extracellular traps and induces neutrophil activation. *Cells* 11 (2), 236. doi:10.3390/ cells11020236

Sreejit, G., Abdel Latif, A., Murphy, A. J., and Nagareddy, P. R. (2020). Emerging roles of neutrophil-borne S100A8/A9 in cardiovascular inflammation. *Pharmacol. Res.* 161, 105212. doi:10.1016/j.phrs.2020.105212

Sroussi, H. Y., Lu, Y., Zhang, Q. L., Villines, D., and Marucha, P. T. (2010). S100A8 and S100A9 inhibit neutrophil oxidative metabolism *in-vitro*: involvement of adenosine metabolites. *Free Radic. Res.* 44 (4), 389–396. doi:10.3109/10715760903431434

Sterner, R. C., and Sterner, R. M. (2021). CAR-T cell therapy: current limitations and potential strategies. *Blood Cancer J.* 11 (4), 69. doi:10.1038/s41408-021-00459-7

Sumardika, I. W., Chen, Y., Tomonobu, N., Kinoshita, R., Ruma, I. M. W., Sato, H., et al. (2019). Neuroplastin- β mediates S100A8/A9-induced lung cancer disseminative progression. *Mol. Carcinog.* 58 (6), 980–995. doi:10.1002/mc.22987

Sun, H., Wen, W., Zhao, M., Yan, X., Zhang, L., Jiao, X., et al. (2020). Emmprin: a potential biomarker for predicting the presence of obstructive sleep apnea. *Clin. Chim. Acta* 510, 317–322. doi:10.1016/j.cca.2020.07.011

Tang, X., Chen, X., Xu, Y., Qiao, Y., Zhang, X., Wang, Y., et al. (2015). CD166 positively regulates MCAM via inhibition to ubiquitin E3 ligases Smurf1 and β TrCP through PI3K/AKT and c-Raf/MEK/ERK signaling in Bel-7402 hepatocellular carcinoma cells. *Cell Signal* 27 (9), 1694–1702. doi:10.1016/j.cellsig.2015.05.006

Tanigawa, K., Tsukamoto, S., Koma, Y. I., Kitamura, Y., Urakami, S., Shimizu, M., et al. (2022). S100A8/A9 induced by interaction with macrophages in esophageal squamous cell carcinoma promotes the migration and invasion of cancer cells via Akt and p38 MAPK pathways. *Am. J. Pathol.* 192 (3), 536–552. doi:10.1016/j.ajpath.2021. 12.002

Tao, Q., Qiu, X., Li, C., Zhou, J., Gu, L., Zhang, L., et al. (2022). S100A8 regulates autophagy-dependent ferroptosis in microglia after experimental subarachnoid hemorrhage. *Exp. Neurol.* 357, 114171. doi:10.1016/j.expneurol.2022.114171

Tardif, M. R., Chapeton-Montes, J. A., Posvandzic, A., Pagé, N., Gilbert, C., and Tessier, P. A. (2015). Secretion of S100A8, S100A9, and S100A12 by neutrophils involves reactive oxygen species and potassium efflux. *J. Immunol. Res.* 2015, 296149. doi:10.1155/2015/296149

Tomonobu, N., Kinoshita, R., and Sakaguchi, M. (2020). S100 soil sensor receptors and molecular targeting therapy against them in cancer metastasis. *Transl. Oncol.* 13 (4), 100753. doi:10.1016/j.tranon.2020.100753

Tong, L., Lan, W., Lim, R. R., and Chaurasia, S. S. (2014). S100A proteins as molecular targets in the ocular surface inflammatory diseases. *Ocul. Surf.* 12 (1), 23–31. doi:10. 1016/j.jtos.2013.10.001

Tripathi, S. C., Fahrmann, J. F., Celiktas, M., Aguilar, M., Marini, K. D., Jolly, M. K., et al. (2017). MCAM mediates chemoresistance in small-cell lung cancer via the PI3K/AKT/ SOX2 signaling pathway. *Cancer Res.* 77 (16), 4414–4425. doi:10.1158/0008-5472.can-16-2874

van den Bos, C., Roth, J., Koch, H. G., Hartmann, M., and Sorg, C. (1996). Phosphorylation of MRP14, an S100 protein expressed during monocytic differentiation, modulates Ca(2+)-

dependent translocation from cytoplasm to membranes and cytoskeleton. J. Immunol. 156 (3), 1247-1254. doi:10.4049/jimmunol.156.3.1247

Vogl, T., Ludwig, S., Goebeler, M., Strey, A., Thorey, I. S., Reichelt, R., et al. (2004). MRP8 and MRP14 control microtubule reorganization during transendothelial migration of phagocytes. *Blood* 104 (13), 4260–4268. doi:10.1182/blood-2004-02-0446

von Lersner, A., Droesen, L., and Zijlstra, A. (2019). Modulation of cell adhesion and migration through regulation of the immunoglobulin superfamily member ALCAM/ CD166. *Clin. Exp. Metastasis* 36 (2), 87–95. doi:10.1007/s10585-019-09957-2

Wanderley, C. W., Colón, D. F., Luiz, J. P. M., Oliveira, F. F., Viacava, P. R., Leite, C. A., et al. (2018). Paclitaxel reduces tumor growth by reprogramming tumor-associated macrophages to an M1 profile in a TLR4-dependent manner. *Cancer Res.* 78 (20), 5891–5900. doi:10.1158/0008-5472.can-17-3480

Wang, D., Liu, G., Wu, B., Chen, L., Zeng, L., and Pan, Y. (2018a). Clinical significance of elevated S100A8 expression in breast cancer patients. *Front. Oncol.* 8, 496. doi:10. 3389/fonc.2018.00496

Wang, K., Huang, W., Chen, R., Lin, P., Zhang, T., Ni, Y. F., et al. (2021). Dimethylation of CD147-K234 promotes the progression of NSCLC by enhancing lactate export. *Cell Metab.* 33 (1), 160–173.e6. doi:10.1016/j.cmet.2020.12.010

Wang, S., Song, R., Wang, Z., Jing, Z., Wang, S., and Ma, J. (2018b). S100A8/A9 in inflammation. *Front. Immunol.* 9, 1298. doi:10.3389/fimmu.2018.01298

Wang, Y., Yu, W., Zhu, J., Wang, J., Xia, K., Liang, C., et al. (2019). Anti-CD166/4-1BB chimeric antigen receptor T cell therapy for the treatment of osteosarcoma. *J. Exp. Clin. Cancer Res.* 38 (1), 168. doi:10.1186/s13046-019-1147-6

Wang, Z., Xu, Q., Zhang, N., Du, X., Xu, G., and Yan, X. (2020). CD146, from a melanoma cell adhesion molecule to a signaling receptor. *Signal Transduct. Target Ther.* 5 (1), 148. doi:10.1038/s41392-020-00259-8

Wang, Z., and Yan, X. (2013). CD146, a multi-functional molecule beyond adhesion. *Cancer Lett.* 330 (2), 150–162. doi:10.1016/j.canlet.2012.11.049

Wattenberg, M. M., and Beatty, G. L. (2020). Overcoming immunotherapeutic resistance by targeting the cancer inflammation cycle. *Semin. Cancer Biol.* 65, 38–50. doi:10.1016/j.semcancer.2020.01.002

Wen, L., Ding, Y., Chen, X., Tian, K., Li, D., Liang, K., et al. (2021). Influences of S100A8 and S100A9 on proliferation of nasopharyngeal carcinoma cells through PI3K/Akt signaling pathway. *Biomed. Res. Int.* 2021, 9917365. doi:10.1155/2021/9917365

Willers, M., Ulas, T., Völlger, L., Vogl, T., Heinemann, A. S., Pirr, S., et al. (2020). S100A8 and S100A9 are important for postnatal development of gut microbiota and immune system in mice and infants. *Gastroenterology* 159 (6), 2130–2145. doi:10.1053/j.gastro.2020.08.019

Wong, S. W., McCarroll, J., Hsu, K., Geczy, C. L., and Tedla, N. (2022). Intranasal delivery of recombinant S100A8 protein delays lung cancer growth by remodeling the lung immune microenvironment. *Front. Immunol.* 13, 826391. doi:10.3389/fimmu.2022.826391

Woo, J. W., Chung, Y. R., Kim, M., Choi, H. Y., Ahn, S., and Park, S. Y. (2021). Prognostic significance of S100A8-positive immune cells in relation to other immune cell infiltration in pre-invasive and invasive breast cancers. *Cancer Immunol. Immunother.* 70 (5), 1365–1378. doi:10.1007/s00262-020-02776-5 Wu, Z., Jiang, D., Huang, X., Cai, M., Yuan, K., and Huang, P. (2022). S100A8 as a promising biomarker and oncogenic immune protein in the tumor microenvironment: an integrative pancancer analysis. *J. Oncol.* 2022, 6947652. doi:10.1155/2022/6947652

Xie, S., Wang, J., Tuo, W., Zhuang, S., Cai, Q., Yao, C., et al. (2023). Serum level of S100A8/A9 as a biomarker for establishing the diagnosis and severity of community-acquired pneumonia in children. *Front. Cell Infect. Microbiol.* 13, 1139556. doi:10.3389/fcimb.2023.1139556

Xin, X., Zeng, X., Gu, H., Li, M., Tan, H., Jin, Z., et al. (2016). CD147/EMMPRIN overexpression and prognosis in cancer: a systematic review and meta-analysis. *Sci. Rep.* 6, 32804. doi:10.1038/srep32804

Xiong, L., Edwards, C. K., 3rd, and Zhou, L. (2014). The biological function and clinical utilization of CD147 in human diseases: a review of the current scientific literature. *Int. J. Mol. Sci.* 15 (10), 17411–17441. doi:10.3390/ijms151017411

Yang, L., Yang, M., Zhang, H., Wang, Z., Yu, Y., Xie, M., et al. (2012). S100A8targeting siRNA enhances arsenic trioxide-induced myeloid leukemia cell death by down-regulating autophagy. *Int. J. Mol. Med.* 29 (1), 65–72. doi:10.3892/ijmm.2011.806

Yang, Y., Sanders, A. J., Dou, Q. P., Jiang, D. G., Li, A. X., and Jiang, W. G. (2021). The clinical and theranostic values of activated leukocyte cell adhesion molecule (ALCAM)/ CD166 in human solid cancers. *Cancers (Basel)* 13 (20), 5187. doi:10.3390/cancers13205187

Yin, C., Li, H., Zhang, B., Liu, Y., Lu, G., Lu, S., et al. (2013). RAGE-binding S100A8/ A9 promotes the migration and invasion of human breast cancer cells through actin polymerization and epithelial-mesenchymal transition. *Breast Cancer Res. Treat.* 142 (2), 297–309. doi:10.1007/s10549-013-2737-1

Yuan, R., Zhao, W., Wang, Q. Q., He, J., Han, S., Gao, H., et al. (2021). Cucurbitacin B inhibits non-small cell lung cancer *in vivo* and *in vitro* by triggering TLR4/NLRP3/GSDMD-dependent pyroptosis. *Pharmacol. Res.* 170, 105748. doi:10.1016/j.phrs.2021.105748

Zavorka Thomas, M. E., Jeon, J. Y., Talebi, Z., Buelow, D. R., Silvaroli, J., Campbell, M. J., et al. (2021). Gilteritinib-induced upregulation of S100A9 is mediated through BCL6 in acute myeloid leukemia. *Blood Adv.* 5 (23), 5041–5046. doi:10.1182/ bloodadvances.2021005614

Zhang, L., Jiang, H., Xu, G., Wen, H., Gu, B., Liu, J., et al. (2015). Proteins S100A8 and S100A9 are potential biomarkers for renal cell carcinoma in the early stages: results from a proteomic study integrated with bioinformatics analysis. *Mol. Med. Rep.* 11 (6), 4093–4100. doi:10.3892/mmr.2015.3321

Zhang, L., Zhou, S., Zhou, T., Yuan, K., Li, X., and Tang, J. (2021). S100A8 promotes chemoresistance via augmenting autophagy in B-cell lymphoma cells. *Oncol. Rep.* 45 (1), 151–158. doi:10.3892/or.2020.7841

Zhong, C., Niu, Y., Liu, W., Yuan, Y., Li, K., Shi, Y., et al. (2022). S100A9 derived from chemoembolization-induced hypoxia governs mitochondrial function in hepatocellular carcinoma progression. *Adv. Sci. (Weinh)* 9 (30), e2202206. doi:10.1002/advs.202202206

Zhong, X., Xie, F., Chen, L., Liu, Z., and Wang, Q. (2020). S100A8 and S100A9 promote endothelial cell activation through the RAGE-mediated mammalian target of rapamycin complex 2 pathway. *Mol. Med. Rep.* 22 (6), 5293–5303. doi:10.3892/mmr.2020.11595

Zoni, E., Astrologo, L., Ng, C. K. Y., Piscuoglio, S., Melsen, J., Grosjean, J., et al. (2019). Therapeutic targeting of CD146/MCAM reduces bone metastasis in prostate cancer. *Mol. Cancer Res.* 17 (5), 1049–1062. doi:10.1158/1541-7786.mcr-18-1220

Glossary

AA	Arachidonic acid
ADCs	Antibody-drug conjugates
AGEs	Advanced glycation end-products
AIF	Apoptosis-inducing factor
ALCAM	Activated leukocyte cell adhesion molecule
BRCA1	Breast cancer-associated gene 1
CAR	Chimeric antigen receptor
CRC	Colorectal cancer
CREB	CAMP-response-element-binding protein
CSC	Cancer stem cell
CXCL12	CXC chemokine ligand 12
CXCR	CXC chemokine receptor
ED	Extracellular domain
EGFR	Epidermal growth factor receptor
EMT	Epithelial-mesenchymal transition
EMMPRIN	Extracellular matrix metalloproteinase inducer
ENZA	Enzalutamide
ETV4	E26 transformation specific (ETS) variant transcription factor 4
FAK	Focal adhesion kinase
GCNT3	Glucosaminyl (N-acetyl) transferase 3
GSK3β	Glycogen synthase kinase 3β
HIF	Hypoxia-inducible factor
HMGB1	High mobility group box-1
Ig	Immunoglobulin
11.6	Interleukin-6
IRAK1	Interleukin 1 receptor associated kinase 1
IkB	Inhibitor kB
JAK	Janus kinase
LRR	leucine-rich region
МАРК	Mitogen-activated protein kinases
MCAM	Melanoma cell adhesion molecule
MDSCs	Myeloid-derived suppressor cells
ММР	Matrix metalloprotease
MRP8/14	Myeloid related protein 8/14
mTOR	Mammalian target of rapamycin
NADPH	Nicotinamide adenine dinucleotide 3-phosphate
NPTN	Neuroplastin
Nrf2	Nuclear factor E2-related factor 2
PD-1	Programmed death 1
PD-L1	PD-1 ligands

PDZ	Postsynaptic density-95/Discs large/Zonula occludens 1
РІЗК	Phosphoinositide 3-kinase
РКС	Protein kinase C
RAGEs	Receptor for advanced glycation end products
RAS	Rat sarcoma
ROS	Reactive oxygen species
SLC22A18AS	Solute carrier family 22 member 18 antisense
SPDEF	SAM pointed-domain containing E26 transformation-specific transcription factor
SSSRs	S100 soil sensor receptors
STAT3	Signal transducer and activator of transcription 3
TIR	Toll/interleukin-1 receptor
TLR4	Toll-like receptor 4
TLRs	Toll-like receptors
TNF-α	Tumor necrosis factor-a
TPL2	Tumor progression locus 2
TRAF2	Tumor necrosis factor receptor-associated factor 2
VEGFR	Vascular endothelial growth factor receptor
ҮАР	Yes kinase-associated protein
ZEB1	Zinc finger E-box binding homeobox 1