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# Dose-finding study of valspodar (PSC 833) with daunorubicin and cytarabine to reverse multidrug resistance in elderly patients with previously untreated acute myeloid leukemia

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*Introduction*: This trial was designed to determine the maximum tolerated dose of intravenous daunorubicin (DNR) in combination with valspodar and to test the feasibility of P-glycoprotein modulation using valspodar in elderly patients with previously untreated acute myelogenous leukemia receiving standard induction chemotherapy.

*Methods*: Patients  $\geq 60$  years of age with previously untreated AML received valspodar (10 mg/kg/24 h by continuous intravenous infusion [CIV] on days 1-4 with a 2-mg/kg loading dose on day 1) in conjunction with two cycles of induction chemotherapy consisting of cytarabine (200 mg/m<sup>2</sup> CIV on days 1-7), and DNR (35 mg/m<sup>2</sup> [cohort 1] or 45 mg/m<sup>2</sup> [cohort 2] on days 1-3, intravenous bolus). Patients were assessed for dose-limiting toxicities (DLT), response rate, event-free and overall survival, and pharmacokinetics of valspodar and DNR.

**Results**: Valspodar was well tolerated at the lower DNR dose level (ie,  $35 \text{ mg/m}^2$ ) resulting in a 21% rate of DLT and only three toxic deaths. Treatment-related mortality was unacceptably high at the  $45 \text{ mg/m}^2$  DNR dose level. The complete response rate was 49% overall and similar in both cohorts. The median overall survival of patients was 333 days in cohort 1 compared to 98 days in cohort 2. At baseline, 70% of assessable patients were Pglycoprotein positive.

**Conclusion**: Substantial inhibition of P-glycoprotein activity can be achieved in this patient population at clinically tolerable doses of valspodar and DNR. The maximum tolerated dose of DNR was established as  $35 \text{ mg/m}^2$ . This regimen is being further evaluated in phase III trials. *The Hematology Journal* (2000) **1**, 411-421

*Keywords:* acute myeloid leukemia; multidrug resistance; reversal of resistance; daunorubicin; valspodar

#### Introduction

Intrinsic or acquired resistance to chemotherapy remains a major obstacle to achieving long-term survival in patients with acute myeloid leukemia (AML). Standard induction therapy produces complete remission (CR) in 52-72% of patients with *de novo* AML<sup>1-8</sup> depending on a variety of factors,

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including age of the patient, history of myelodysplastic syndrome (MDS), and cytogenetic and immunophenotypic profile. Lower response rates are observed in older patients<sup>9–13</sup> or in those with secondary AML or a history of MDS.<sup>14</sup> Overall median duration of remission is 8-12 months, and median survival is 9-16 months. Approximately 40% of patients who received intensive induction or consolidation therapy with high-dose cytarabine remain disease free at four years.<sup>8,15,16</sup> However, the benefits of intensive therapy have been confined primarily to younger patients (ie, <55 years of age).

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One hypothesized reason for failure to respond to chemotherapy is the presence of multidrug resistance (MDR), a mechanism of resistance to structurally unrelated cytostatic agents. In AML, MDR is most frequently mediated through enhanced drug efflux by P-glycoprotein (P-gp).<sup>17,18</sup> This 170-kd transmembrane glycoprotein is a member of the adenosine triphosphate-binding cassette (ABC) proteins<sup>19,20</sup> and is encoded by the multidrug resistance gene-1 (MDR-I).<sup>21–23</sup> In cancer cells, P-gp is capable of extruding a wide variety of structurally unrelated drugs taken up through passive diffusion, including daunorubicin (DNR), doxorubicin, idarubicin, mitoxantrone, etoposide, and vinca alkaloids that are frequently used for treating AML.<sup>24</sup> P-glycoprotein is also expressed in several normal tissues, such as CD34<sup>+</sup> hematopoietic progenitor cells, biliary canaliculi, kidney tubules, and the blood-brain barrier.18,25

Since 1989 several studies have reported the *MDR-1* phenotype to be expressed in about 30-50% of AML patients, depending on the technique and the threshold used,<sup>26-29</sup> with a higher percentage in AML patients relapsing after or refractory to chemotherapy.<sup>30</sup> More recent studies by the Southwest Oncology Group (SWOG) indicate that the *MDR-1* gene is expressed with extremely high frequency in elderly AML patients (ie, patients >55 years of age). As many as 80% of elderly patients with *de novo* AML present with a MDR phenotype as a result of P-gp expression; the frequency of P-gp expression is similar in both *de novo* and secondary disease.<sup>31</sup>

The correlation between MDR-1 gene expression and treatment outcome in AML is well documented.<sup>32</sup> In most studies, MDR-1 expression was an adverse prognostic factor for achieving a CR.<sup>33–50</sup> In elderly, newly diagnosed patients, the CR rate of strongly P-gp-positive as compared to P-gp-negative patients was recently found to be 31 vs 71%.<sup>31</sup> Finally, several recently published large studies have shown that MDR-1 expression, as well as secondary AML, CD34 positivity of leukemic blasts, unfavorable cytogenetics, and age were each, significantly and independently, associated with lower CR rates in elderly patients.<sup>36,49-52</sup>

Phase I/II studies of cyclosporine A (CsA), combined with DNR in patients with relapsed/ refractory AML or with VAD (vincristine, doxorubicin, dexamethasone) in patients with multiple myeloma, have demonstrated the feasibility of CsA-mediated MDR reversal in vivo.53-55 Valspodar (PSC 833) is an analog of cyclosporine D, which is approximately 5-30-fold more potent than CsA in vitro.<sup>56</sup> Unlike CsA, however, valspodar is neither immunosuppressive nor nephrotoxic.<sup>56-58</sup> In fact, unlike CsA, which is a substrate for P-gp and appears to exert its modulatory effects by competitive inhibition,59,60 valspodar is a high-affinity, noncompetitive inhibitor of P-gp and is not appreciably transported by it.<sup>61</sup> In phase I/II trials, it was established that valspodar blood levels high enough to reverse MDR in resistant cell lines in vitro (1000 to 2000 ng/ml)<sup>62-64</sup> could be achieved and were well tolerated. Finally, in hematological malignancies, MDR reversal by valspodar has been demonstrated in patients with VAD-resistant/refractory multiple myelo-ma.<sup>34,65</sup>

Because P-gp has a physiological role in tissues normally responsible for the transport and excretion of cytotoxic drugs, treatment with P-gp inhibitors delays the elimination of other anticancer drugs. In phase I studies the dose-normalized area under the curve (AUC) increased by 61-74% for doxorubicin and by 84% (mean values) for etoposide when these drugs were given in combination with valspodar.62,66 This pharmacokinetic interaction therefore requires appropriate dose reduction of the cytotoxic drugs that are P-gp substrates, when used in association with valspodar, to continue to provide effective treatment without increasing toxicity. Herein we report the results of a phase I study to establish the maximum tolerated dose (MTD) of DNR. combined with fixed doses of valspodar and cytarabine, in previously untreated, elderly AML patients.

#### Methods

The study was an open, multicenter, multinational trial. The study protocol was approved by the Institutional Review Boards of the participating hospitals. All patients gave prior written informed consent.

#### Patient population

The study population consisted of previously untreated patients  $\geq 60$  years of age, who had a cytopathologically confirmed diagnosis of primary or secondary AML (M0 to M7) according to the French-American-British (FAB) classification. Patients who progressed from myelodysplastic syndrome had (MDS) were eligible if they had received no previous chemotherapy. Patients with a prior history of polycythemia rubra vera or primary myelofibrosis, or in blast cell crisis of chronic myeloid leukemia, were excluded. Other eligibility criteria included World Health Organization (WHO) performance status  $\leq 2$ , life expectancy  $\geq 12$  weeks, and suitability for intensive chemotherapy in the opinion of the treating physician. Documented central nervous system involvement, left ventricular ejection fraction of < 50%as measured by cardiac radionuclide-gated blood pool scan, serum creatinine  $\geq 175 \, \mu \text{mol/l}$ , serum bilirubin  $\geq$  2.5-fold the institutional upper limit of normal, positive hepatitis B surface antigen or a known history of hepatitis C or human immunodeficiency virus infection, known hypersensitivity to CsA, or prior treatment with cytotoxic agents and/or radiotherapy were all causes for exclusion. All patients received a central venous access catheter and were hospitalized until their granulocytes had recovered to  $\ge 0.5 \times 10^9$ cells/l.

#### Schedule of chemotherapy and valspodar

All patients were planned to receive two identical induction cycles, irrespective of the response to the first cycle. In each induction cycle, valspodar (which was supplied in a cremophor EL-based solution and was diluted in saline) was administered via continuous intravenous infusion (CIV) at a dose of 10 mg/kg/24 h on days 1-4; on day 1, the infusion was started concomitantly with a loading dose of 2 mg/kg administered over 2 h. Cytarabine was administered CIV at a dose of 200 mg/m<sup>2</sup>/24 h on days 1-7. Daunorubicin was administered as an intravenous (IV) bolus injection on days 1-3: the starting dose (cohort 1) was 35 mg/m<sup>2</sup>. This dose was escalated in cohort 2 to 45 mg/m<sup>2</sup>. No further dose escalation of DNR was foreseen in the protocol. All patients who were in CR after the induction cycles received a single consolidation course consisting of etoposide at a dose of 80 mg/m<sup>2</sup>/day intravenous (IV), mitoxantrone at a dose of 6 mg/m<sup>2</sup>/day IV, and cytarabine (1.0 g/m<sup>2</sup>) as a 6-h infusion on days 1-4. Valspodar was not administered during the consolidation course.

## Determination of maximum tolerated dose and dose-limiting toxicities

The primary objective of the study was to determine the MTD of DNR combined with a fixed dose of valspodar and cytarabine. Two doses of DNR were evaluated:  $35 \text{ mg/m}^2/\text{day}$  (cohort 1) and  $45 \text{ mg/m}^2/\text{day}$ (cohort 2). Dose escalation was allowed if the observed dose-limiting toxicity (DLT) rate in cohort 1 was <25%; dose reduction was necessary if the observed DLT rate was >40%. The decision to escalate (or reduce) the DNR dose in the next cohort was based on the safety profile of the first 16 evaluable patients. However, accrual on the ongoing dose was continued until the next cohort was open, thus allowing for more than 16 patients to be enrolled in each cohort.

Dose-limiting toxicities were defined as: (1) neutropenia (absolute neutrophil count [ANC]  $< 0.5 \times 10^9$ cells/l or thrombocytopenia ( $< 50 \times 10^9$  platelets/l) in the absence of AML for >8 weeks after the start of treatment cycle 1; (2) death during or after treatment in cycle 1 without recovery of blood cell counts (ANC  $\ge 0.5 \times 10^9$  cells/l and platelets  $\ge 50 \times 10^9$ /l) or hypoplasia without evidence of persistent AML; and (3) any Common Toxicity Criteria (CTC) grade 3/4 nonhematologic toxicity except nausea, alopecia, cerebellar ataxia, hyperbilirubinemia lasting  $\le 20$  days, or infection responsive to antibiotic treatment.

#### Response criteria and treatment outcome

Complete remission was defined as: (1) a normal marrow cellularity (ie, >20%) with <5% blasts and no Auer rods; (2) no evidence of extramedullary leukemia; and (3) recovery of peripheral blood neutrophils to at least 1500 cells/ $\mu$ l and platelets to at least 100 000/ $\mu$ l (all persisting for at least 28 days).

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Treatment failure was defined as: (1) absolute drug resistance with failure to achieve a reduction of the leukemic infiltration in the marrow that would meet the criteria of CR, or persistent leukemia in the peripheral blood, or persistent extramedullary disease; (2) regeneration failure with persistent hypoplasia for more than 40 days after completion of induction therapy; or (3) aplastic death: death during hypoplasia without signs of persistent leukemia. The overall survival was calculated from start of treatment until the date of death. The frequencies of excessive toxicities, infections, and hemorrhages were evaluated separately.

#### Pharmacokinetic analyses

Measurements of valspodar blood levels were performed to determine that the concentration of valspodar was within the range of maximal *in vitro* P-gp inhibition. Blood samples (2 ml) were drawn before starting valspodar infusion on days 1-4. The samples were stored in EDTA-coated tubes at  $-20^{\circ}$ C. The valspodar concentration in whole blood was determined using a radioimmunoassay (RIA) as previously published.<sup>67</sup>

Samples for measurement of plasma concentrations of DNR and the hydroxylated metabolite DNR-ol were taken on day 3 of cycle 1, just before, just after, and at 4, 24, and 48 h after DNR administration. Blood samples (5 ml) were drawn into heparinized tubes and centrifuged as soon as possible, and the plasma was immediately transferred into uncoated polypropylene tubes and deep frozen at  $-20^{\circ}$ C. Plasma levels of DNR and DNR-ol were evaluated by high-performance liquid chromatography as previously described.68 Plasma AUC of DNR and DNR-ol up to the last measurable sampling time point (AUC<sub>last</sub>) was calculated by the log-linear trapezoidal method. The terminal half-life was assessed by linear regression to the log-concentration-time plot and AUC from time zero to infinity was determined by the formula  $AUC_{inf} = AUC_{last} + C (last)/(0.693/t_{\frac{1}{2}})$ . The AUC<sub>inf</sub> was corrected for carry-over from the two previous doses by AUC<sub>inf,corr</sub> = AUC<sub>inf</sub> - C(0h)/0.693/ $t_{\frac{1}{2}}$ ). This correction of AUC allowed for a comparison to literature values.

#### Determination of P-gp expression and function

At diagnosis, and after informed consent was given, bone marrow aspirates were obtained from the sternum or the posterior iliac crest. Marrow samples (3 ml) were collected into 10-ml tubes containing 5000 International units (IU) of heparin in 2 ml of DMEM (Gibco BRL, Paisley, UK), cooled to between 0°C and 4°C, and shipped on ice overnight to the central laboratory. Mononuclear bone marrow cells were collected by centrifugation over Lymphoprep (Nycomed, Oslo, Norway), and suspended at a concentration of  $4 \times 10^9$  cells/ml in DMEM supplemented with 10% fetal calf serum and gentamycin.

For measurement of P-gp expression, cells were incubated at room temperature with a monoclonal antibody against P-gp, either MRK16 (Kamiya Biomedical Company, Tukwila, USA) at a concentration of 10 µg/ml or UIC-2 (Immunotech, Marseille, France) at a concentration of 12.5  $\mu$ g/ml, or a matching aspecific isotype control antibody mouse IgG2a (Sigma) at a concentration of 10  $\mu$ g/ml. Cellbound antibodies were detected by isothiocyanate (FITC)-labeled rabbit antimouse immunoglobulin antibodies (DAKO, Glostrup, Denmark). Cells were incubated with 7-aminoactinomycin (7-AAD, Sigma) to exclude nonviable cells. Fluorescence was measured using a FACScalibur (Becton and Dickinson, San Jose, USA). The results are expressed as the ratio of cellassociated fluorescence of cells incubated with the anti-P-gp antibody to the mean of cell-associated fluorescence of cells incubated with the matching aspecific control antibody.40

For measurement of P-gp function, cells were incubated for 1 h at 37°C at 5% CO<sub>2</sub> in the absence or presence of 2  $\mu$ M valspodar. After this incubation, 2 ng/ml rhodamin-123 (Rho) obtained from Sigma (St Louis, USA) was added to the cells. A sample was taken at time 0 to correct for background fluorescence and after 75 min to measure intracellular Rho accumulation. Cells were incubated with 7-AAD to exclude nonviable cells. Fluorescence was measured using a FACScalibur. Results are given as the ratio of the mean intracellular Rho fluorescence of cells exposed to valspodar to the mean intracellular fluorescence of cells not exposed.

The drug-sensitive cell line RMPI 8226 S and the drug-resistant variant 8226 D6, kindly provided by Dr W Dalton (Lee Moffit Cancer Centre, Tampa, USA), were used as controls in each experiment. Based on all control tests run to measure P-gp function, the mean+s.d. ratio of Rho fluorescence of the 8226 S was 0.90+0.04 and of the 8226 D6 13.80+7.48. For Pgp expression, the mean MRK16/ IgG2a ratio or UIC-2/IgG2a ratio of the negative controls, of ie the 8226 S cell line, was  $1.25 \pm 0.29$  or  $1.24 \pm 0.26$  respectively; the mean fluorescence ratio of the positive controls, ie the 8226 D6 cell line, with MRK16 or UIC-2 were  $34.05 \pm 4.00$  and  $36.18 \pm 4.00$  respectively. Patient bone marrow samples were considered positive for P-gp function when the ratio of Rho fluorescence was >1.05, this value representing a significantly higher ratio than that observed with 8226 S (P < 0.05, 2-sided t-test). Patient samples were considered positive for P-gp expression if the ratio of P-gp fluorescence was >1.50 (P<0.05, 2-sided t-test).

#### Statistical methods

The association between response to treatment and P-gp expression was described by Kendall's correlation coefficient. This coefficient was also used to assess a possible correlation between the AUC of DNR and the occurrence of a DLT. Overall survival distributions were analysed descriptively by cohort using the Kaplan-Meier estimation method, and median survival times were derived.

#### Results

From March 1996 to March 1999, 39 patients with de novo AML were accrued into the study at 15 centers in five countries. The median age was 67 years (range, 60-83 years); other baseline characteristics are summarized in Table 1. The majority of patients (80%) had newly diagnosed AML, and 20% of patients had a prior MDS. The most common subtype was FAB M2 disease (33%), and 41% of patients had M4 disease or higher. The majority of patients (74%) had WHO performance status  $\leq 1$ . Among 34 patients with viable bone marrow samples at baseline who were evaluable for P-gp expression and function, 24 (70%) patients were P-gp positive by either immunohistochemical reactivity with the MRK16 antibody or by measurement of P-gp activity as measured by the Rho retention assay. Ten evaluable patients were clearly P-gp negative at baseline by both assays.

The first 19 patients were treated with  $35 \text{ mg/m}^2$  DNR (cohort 1), and in the next 20 patients, the dose of DNR was escalated to  $45 \text{ mg/m}^2$  (cohort 2). A total of 27 (69%) patients discontinued during the study, 16 due to death, three due to adverse events, seven due to treatment failure, and one due to inability to tolerate further treatment.

Table 1         Demographics and baseline characteristics of patier
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	Number of patients (%)		
	<i>Cohort 1</i> (n = 19)	<i>Cohort 2</i> (n = 20)	Total (n=39)
Median age, year (range)	67 (60-73)	66 (60-83)	67 (60-83)
Gender, male/female	13/6	10/10	23/16
WHO performance index	,	,	,
0	8 (42)	4 (20)	12 (31)
1	6 (32)	11 (55)	17 (44)
2	5 (26)	5 (25)	10 (26)
AML FAB			
M0	0 (0)	1 (5)	1 (3)
M1	2 (11)	5 (25)	7 (18)
M2	9 (47)	4 (20)	13 (33)
M4	5 (26)	2 (10)	7 (18)
M5A	2 (11)	2 (10)	4 (10)
M5B	1 (5)	3 (15)	4 (10)
M6	0 (0)	1 (5)	1 (3)
unknown	0 (0)	2 (10)	2 (5)
De novo AML	14 (74)	17 (85)	31 (80)
Prior MDS	5 (26)	3 (15)	8 (20)
$WBC > 20 \times 10^{6}/L$	5 (26)	10 (50)	15 (38)
Karyotype*		~ /	
NN	11 (58)	10 (50)	21 (54)
Unfavorable	9 (47)	8 (40)	17 (44)
Not done	0 (0)	1 (5)	1 (3)
P-gp expression or function <sup>†</sup>	14 (74)	10 (50)	24 (70)

FAB = French-American-British Classification; MDS = myelodysplastic syndrome; WBC=white blood cell count; P-gp=P glycoprotein. \*NN:46(xy) or 46(xx); Unfavorable: all abnormal exceptt(8,21),t(15,17),inv(16). †Positive for either MRK16 staining orP-gp activity measured by rhodamine intracellular retention assay. Safety

The incidence of DLT was <25% in cohort 1, therefore, the planned DNR dose escalation was carried out. In cohort 2, a high rate of DLTs and toxic deaths suggested that 35 mg/m<sup>2</sup> DNR was the MTD in combination with valspodar.

In cohort 1, four of 19 (21%) patients experienced a nonhematologic protocol-specified DLT during the first induction cycle (Table 2): CTC grade 3 mucositis, grade 3 hypotension, cardiac arrhythmia, and myocardial infarction (one patient each). None of these was fatal. The myocardial infarction occurred in a patient with pre-existing active coronary heart disease. There were seven deaths in cohort 1 (six deaths during cycle 1 and one death during cycle 2); six deaths occurred within 30 days of the start of treatment. Three patients died while in aplasia, and three patients died with leukemic re-growth. The cause of death was one of the following: gastrointestinal bleeding, septic shock, pneumonia, pulmonary embolism, or concomitant bronchopneumopathy. Ten patients were treated on the second induction cycle: one of these patients died with persistent leukemia before day 30 of the second cycle. One patient developed grade 3 mucositis during cycle 2.

In cohort 2, six of 20 (30%) patients experienced a protocol-specified DLT during the first cycle (Table 2). Three patients developed grade 3 mucositis, one patient each had a fatal cardiac arrhythmia and cardiac arrest, and one patient had a grade 3 rash. In addition, one patient developed during cycle 1 a grade 4 hyperbilirubinemia that persisted >20 days and thus fulfilled the protocol criteria for a DLT, yet was deemed secondary to persisting AML documented at day 22. A total of five patients died during cycle 1, four in the aplastic phase before day 30 of the cycle from either cerebral hemorrhage, pulmonary hemorrhage, septic shock, or rapid disease progression. Twelve patients in cohort 2 received a second induction cycle: five of these patients died either from sepsis (n=3), pneumonia (n=1), or cerebral hemorrhage (n=1). Four of the patients who died during the second induction cycle had achieved CR after the first cycle, the fifth patient had persistent leukemia and died on day 37. One patient had grade 4 hyperbilirubinemia, and four patients had grade 3 mucositis following cytotoxic treatment in cycle 2.

 Table 2
 Protocol-specified dose-limiting toxicities occurring in cycle 1

Number of patients	Times of onset (Day)	Description
Cohort 1		
1	3	Grade 3 cardiac arrhythmia
1	2	Grade 4 myocardial infarction
1	2	Grade 3 hypotension
1	24	Grade 3 stomatitis
Cohort 2		
3	4, 9, 10	Grade 3 mucositis
1	4	Cardiac arrhythmia (death)
1	31	Cardiac arrest (death)
1	4	Grade 3 skin rash

Fever, nausea, hypokalemia, and peripheral edema and were the most frequently observed nonhematologic toxicities in both cohorts (Table 3). The DLT of valspodar is cerebellar ataxia, which is transient and fully reversible. In this trial, 10 (26%) patients experiencing grade 1/2 ataxia, and three (8%) patients experienced grade 3 ataxia or cerebellar syndrome, which usually occurred between days 3 and 4 of valspodar administration. No patient experienced grade 4 ataxia. One patient had a dose reduction of valspodar to 7.5 mg/kg/day as a result of grade 3 cerebellar syndrome.

The incidence of hematologic toxicity was as expected with conventional anthracycline-based remission induction schedules in elderly patients. Only 46% of patients were assessable for neutrophil recovery. The median time to neutrophil recovery (ANC  $\ge 0.5 \times 10^{\circ}$  cells/l) in assessable patients was approximately 31 days (range, 21–48 days) and was similar in both cohorts following cycles 1, 2, and 3. During cycle 1, all assessable patients recovered by day 56, the maximum number of days allowed per protocol. Similarly, 92% of patients developed grade 4 thrombocytopenia; however, recovery was complete by day 56 in all assessable patients. The incidence of grade 3/4 bleeding events was higher in cohort 2 (20%) than in cohort 1 (5%).

#### Response and survival

Eight of 19 (42%) patients in cohort 1 and 11 of 20 (55%) patients in cohort 2 achieved a CR for an overall response rate of 49% (Table 4). The majority of responding patients achieved a CR after 1 cycle of therapy. There were 13 treatment failures as a result of absolute or relative resistance: eight in cohort 1, and five in cohort 2. In cohort 1, early death (ie, within 30 days of the beginning of chemotherapy of either induction cycle) occurred in seven patients, compared with 10 patients in cohort 2, four of whom were in CR after cycle 1.

Table 3 Proportion of patients with the most common CTC grade1/2 and grade 3/4 nonhematologic toxicities

	Cohort 1 (n=19; 30  cycles)		Cohort 2 (n=20; 28  cycles)	
Toxicity	Grade 1/2 (%)	Grade 3/4 (%)	Grade 1/2 (%)	Grade 3/4 (%)
Fever	89	11	60	15
Nausea	58	0	55	5
Edema	53	5	35	0
Skin	26	0	0	5
Mucositis	27	5	25	25
Hyperbilirubinemia	6	5	0	25
Hypokalemia	53	0	55	5
Arrhythmia	25	5	20	5
Sepsis	21	16	15	20
Ataxia	21	5	30	5
Headache	37	0	20	0
Hemorrhage	15	5	25	20
Hypertension	21	11	15	0

The median overall survival was 333 days in cohort 1 compared with 98 days in cohort 2 (Figure 1). The majority of patients died of progressive disease; however, the incidence of toxic deaths, primarily as a result of sepsis, was greater in cohort 2. The 6-, 9-, and 12-month survival rates were consistently higher in cohort 1 compared with cohort 2.

#### P-gp Expression

Among 34 patients evaluable for P-gp expression and function, 24 (70%) patients were positive for either Pgp expression or function (ie, bone marrow blasts were either positive for P-gp expression by flow cytometry using the MRK16 antibody or demonstrated P-gp activity by the Rho retention assay). Of these 24 P-gppositive patients, 11 (46%) achieved a CR (Table 5). In

Table 4 Clinical outcomes

	<i>Cohort 1</i> (n = 19)	<i>Cohort 2</i> (n=20)	Total (n=39)
Complete remission	8 (42)	11 (55)	19 (49)
Treatment failure	8 (42)	5 (25)	13 (33)
Alive in CR after cycle 2	9 (47)	6 (30)	15 (38)
Total deaths during study	7 (37)	10 (50)	17 (43)
Early deaths in CR during cycle 2	0 (0)	4 (20)	4 (10)
Death while in aplasia	3 (16)	4 (20)	7 (18)
Death from persistent leukemia	4 (21)	2 (10)	6 (15)



Figure 1 Kaplan-Meier estimate of overall survival time by cohort for patients treated with daunorubicin, cytarabine, and valspodar.

10 of these 24 patients, MRK16 reactivity and P-gp functional assays were discordant (ie, four patients were MRK16<sup>+</sup>/Rho<sup>-</sup> and six patients were MRK16<sup>-</sup>/Rho<sup>+</sup>). Ten patients were negative for P-gp expression and function: of these, six (60%) achieved a CR. The CR rate was not significantly different in P-gp-positive compared with P-gp-negative patients. Furthermore, analysis based on Kendall's correlation coefficient demonstrated no correlation between response to treatment and P-gp expression or function. Among the 13 patients with absolute resistance (ie, treatment failures), three patients were P-gp positive, three patients were P-gp negative, and four were MRK16<sup>+</sup>/Rho<sup>-</sup>.

#### **Pharmacokinetics**

Repeated daily measurements of blood concentrations of valspodar were taken in 34 patients. One patient, however, was excluded from the pharmacodynamic analysis because he only received one-tenth of the planned valspodar dose. Daily blood concentrations of valspodar are shown in Figure 2. The median whole blood concentration during the continuous infusion of valspodar was approximately 2200 ng/ml (range, 778 –

 Table 5
 P-glycoprotein expression and function and clinical response in relation to P-gp expression

	No. Patients	CR	No CR
P-glycoprotein positive (MRK16 <sup>+</sup> or Rho <sup>+</sup> )	24	11	13
MRK16 <sup>+</sup> , Rho <sup>+</sup>	12	8	4
MRK16 <sup>+</sup> , Rho <sup>-</sup>	4	1	3
MRK16 <sup>-</sup> , Rho <sup>+</sup>	6	1	5
MRK16 <sup>+</sup> , Rho not assessable	2	1	1
P-glycoprotein negative (MRK16 <sup>-</sup> and Rho <sup>-</sup> )	10	6	4
Unassessable	5	2	3
Total	39	19	20

 $MRK16^+$  indicates positive for staining with the MRK16 antibody;  $Rho^+$  indicates a positive P-gp functional rhodamine 123 cellular retention assay.



Figure 2 Daily valspodar blood concentrations during and immediately after the period of valspodar administration.

9394 ng/ml). With very few exceptions, all patients showed valspodar blood levels >1000 ng/ml at all time points. The four patients showing grade 2/3 cerebellar toxicity had mean peak valspodar blood concentrations similar to those with grade 1 or no cerebellar toxicity ( $4680 \pm 1316$  ng/ml vs  $5254 \pm 3733$  ng/ml, n = 26).

Mean plasma concentrations of DNR and DNR-ol were nearly the same in cohort 1 and cohort 2 despite a 29% dose increase in cohort 2. The mean plasma 48-h AUCs for DNR was  $2.26\pm1.67 \ \mu$ M·h in cohort 1 (*n*=15) compared with  $2.02\pm1.00 \ \mu$ M·h in cohort 2 (*n*=16). The mean plasma 48-h AUC for the hydroxyl metabolite DNR-ol was  $17.7\pm6.4 \ \mu$ M·h in cohort 1 vs 19.9 $\pm6.2 \ \mu$ M·h in cohort 2. When extrapolated to infinity and corrected for carry-over from previous doses the mean AUCs for DNR in cohorts 1 and 2 were  $2.57\pm1.85$  and  $2.29\pm1.13 \ \mu$ M·h, respectively. The terminal plasma half-life of DNR was  $51.3\pm25.9$  h in cohort 1 and  $68.6\pm47.7$  h in cohort 2, thus considerably longer than the 15-47 h reported in the literature when DNR is given without valspodar.<sup>69-74</sup>

No significant correlation was found between the plasma AUC of DNR and DLT. Kendall's correlation coefficient was 0.12. Further, no significant correlation between the observed response to treatment and the DNR AUC was apparent (Figure 3).

The study design did not allow for a comparison of the anthracycline PK with and without valspodar. During the first 4 h after administration, the pharmacokinetic profile of DNR in combination with valspodar in this trial is comparable to that observed with the 45-mg/m<sup>2</sup> dose of DNR alone in elderly AML patients in a previous trial<sup>75</sup> (Figure 4) (ie, peak levels were comparable). However, due to the prolonged terminal half-life, DNR persisted in plasma for a longer period of time and hence resulted in an increase in AUC<sub>inf</sub> when compared to literature values for DNR given without valspodar.<sup>68,70–74</sup> For doses of 45–75 mg/m<sup>2</sup>/d, mean AUCs in the range of 0.57–1.97  $\mu$ M·h have

### Plasma Daunorubicin AUC<sub>24hours</sub> and Response to Treatment



Figure 3 Relationship between plasma daunorubicin  $AUC_{48 h}$  and patient response to treatment. No difference was observed between cohorts.

Daunorubicin Plasma Levels

Figure 4 Pharmacokinetic profile of daunorubicin in combination with valspodar. No difference was observed between cohorts.

been reported. The AUC<sub>inf</sub> values of the present study were higher than all the reported values and it is estimated that a DNR dose of  $90-120 \text{ mg/m}^2/\text{d}$  given without valspodar would be needed in order to achieve similar AUCs.

#### Discussion

Resistance to combination chemotherapy is a major obstacle to improving remission rates and survival in AML. Unlike many other tumors, in which MDR is often acquired during therapy, many untreated AML patients exhibit P-gp-related MDR at diagnosis. Several independent studies have found that P-gp expression is an unfavorable prognostic factor for response and/or survival. Reversal of the MDR phenotype by blockade of P-gp offers a possible route to overcome MDR. Several trials have attempted to demonstrate MDR reversal in AML patients using a Pgp modulator. However, most of these trials have been inconclusive because they failed to achieve blood levels of the P-gp modulator high enough to reverse drug resistance and/or they included patients whose tumor cells possessed other mechanisms of resistance in addition to P-gp expression. Therefore, previously untreated, elderly AML patients in whom only P-gp expression has been shown to be predictive of clinical outcome<sup>50</sup> provide a good model for investigating the clinical utility of P-gp modulators.

Because inhibition of P-pg alters the PK of concurrently administered cytostatic agents that are substrates for P-gp, including DNR, the dose of these chemotherapy agents must be reduced to avoid dose-limiting toxicity. The primary goal of this study was to establish the optimal dose of DNR when administered concomitantly with a dose of valspodar that can fully reverse MDR. Cytarabine is not transported by P-gp; therefore, the dose of this drug was left unchanged. The dose of DNR used in protocols for elderly AML patients has varied greatly; however, 45 mg/m<sup>2</sup> is a fairly typical standard dose.<sup>12,76,77</sup> In the present study a reduced dose of 35 mg/m<sup>2</sup> was chosen as the starting

dose based on the schedule used in a large cooperative group study conducted by the European Organization for the Research and Treatment of Cancer, Leukemia Cooperative Group (EORTC-LCG) and the Dutch Hematology-Oncology Cooperative Group (HO-VON).<sup>77</sup> In that study, the median time to hematologic recovery was 25 days in responding patients. The median time to recovery observed in the present study (ie, 31 days) was slightly longer compared with the previous experience with this regimen. The longer hematologic recovery time may have been due to the effect of valspodar on undifferentiated CD34<sup>+</sup> hematopoietic cells, which express functional P-gp in the cell membrane, thus making them more sensitive to the cytotoxic effects of DNR. However, these data have to be interpreted with caution because only 46% of patients were assessable for ANC recovery.

The results of this trial established  $35 \text{ mg/m}^2 \text{ DNR}$ as the MTD when administered concurrently with 10 mg/kg/day valspodar. This dose of valspodar yielded blood concentrations that have been shown to effectively reverse MDR (ie, median blood level  $\geqslant\!2000~ng/ml)^{62-64}$  over four days and was associated with a low incidence of grade 3/4 toxicities. Severe ataxia and hyperbilirubinemia occurred in  $\leq 10\%$  of patients. In the absence of a dose relationship, the specific contributions of valspodar and chemotherapy to the occurrence of cardiac arrhythmias remain unclear. In general, the toxicity profile observed when valspodar was added to 35  $mg/m^2$  DNR did not differ from that normally observed in elderly AML patients treated with an anthracycline-based induction chemotherapy regimen. Although the toxicity profiles of the two cohorts did not differ dramatically, cohort 1 had a consistently more favorable profile than cohort 2. In cohort 1 (35 mg/m<sup>2</sup> DNR), 10 of 19 (53%) patients completed a second induction cycle, and a total of seven deaths occurred over 30 cycles, mostly due to disease progression. Only three deaths in cohort 1 were a result of fatal infections or hemorrhage precipitated by hematologic toxicity. In contrast, at the higher dose level of DNR (ie,  $45 \text{ mg/m}^2$ ), 10 deaths occurred over 31 cycles, and seven deaths could be attributed to treatment-related toxicity, including the death of four patients who had achieved a CR after the first cycle. This increase in treatment-related mortality at the higher DNR dose level may have been related to increased DNR exposure as a result of the interaction with valspodar. Furthermore, there was a higher incidence of grade 3/4 mucositis and bleeding events in cohort 2.

The CR rates were 42% in cohort 1 and 55% in cohort 2, and the majority (14 of 19) of patients entered CR after only one treatment cycle, which may be an important benefit for elderly patients. Although the number of patients included in this study is relatively small, the overall survival analysis clearly shows an advantage for the patients treated with the lower dose of DNR. Most encouraging was the observation that the CR rate did not differ in P-gppositive compared with P-gp-negative patients, suggesting that valspodar effectively reversed MDR in the group of P-gp-positive patients who would ordinarily be expected to have a substantially lower CR rate compared with comparable patients with a P-gp-negative phenotype. Interestingly, four patients had negative functional tests although P-gp was present in the cell membrane as demonstrated by MRK16 staining. Nevertheless, only one of these patients achieved CR, suggesting the possible involvement of other functional mechanisms of drug resistance. Despite the relatively small number of patients included in this study, these results suggest that the adverse impact of P-gp on treatment outcome in patients with P-gp-positive AML blasts may be overcome by treating these patients with induction chemotherapy plus valspodar.

No relationship was found between the plasma concentration of the DNR or DNR-ol and the response to treatment. This is in agreement with the previous observation that no plasma DNR or DNR-ol PK parameter is predictive of the clinical outcome of therapy in *de novo* AML and ALL.<sup>75,78</sup> Galletis *et al.* found that intracellular DNR concentrations may be a better predictor for clinical outcome.<sup>78</sup>

Although the dose of DNR was increased by 29% in cohort 2, there was no difference in the AUC between cohorts. The AUC of DNR in this study was consistent with a theoretical DNR dose of  $90-120 \text{ mg/m}^2$  without valspodar, but this high AUC did not translate into increased toxicity. The interpretation of these findings is unclear, but it is difficult to draw definitive conclusions because of the large inter-patient variability in the DNR AUC.

In conclusion, the results of this study suggest that substantial inhibition of P-gp can be achieved *in vivo* at clinically tolerable doses of both valspodar and DNR. The feasibility of administering valspodar to elderly patients with untreated AML at a safe and welltolerated dose of DNR has been established. The higher than expected response rate of P-gp-positive patients suggests that valspodar, at the dose used in this study, may reverse MDR. However, phase III randomized trials are needed to determine if this approach will ultimately benefit patients. The 35-mg/  $m^2$  dose of DNR has been recommended for further study. A series of randomized phase III trials of valspodar in different subsets of AML are ongoing, and the results will be available in the near future.

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#### References

- 1 Rai, KR, Holland JF, Glidwell OJ, Weinberg V, Brummer K, Obrecht JP, Preisler HD, Nawabi IW, Prager D, Carey RW, Cooper MR, Haurani F, Hutchison JL, Silver RT, Falkson G, Wiernik P, Hoagland HC, Bloomfield CD, James GW, Gottlieb A, Ramanan SV, Blom J, Nissen NI, Bank A, Ellison RR, Kung F, Henry P, McIntyre OR, Kaan SK. Treatment of acute myeloid leukemia: A study by Cancer and Leukemia Group B. *Blood* 58: 1203, 1981.
- 2 Yates J, Glidwell O, Wiernik P, Cooper MR, Steinberg D, Dosik H, Levy R, Hoagland C, Henry P, Gottlieb A, Cornell C, Berenberg J. Hutchison JL, Raich P, Nissen N, Ellison RR, Frelick R, James GW, Falkson G, Silver RT, Haurani F, Green M, Henderson E, Leone L, Holland JF. Cytosine arabinoside with daunorubicin or adriamycin for therapy of acute myeloid leukemia: a CALGB study. *Blood* 60: 453, 1982.
- 3 Omura GA, Vogler WR, Lefante J, Silberman H, Knospe W, Gordon D, Jarrell R. Treatment of acute myelogenous leukemia: Influence of three induction regimens and maintenance with chemotherapy or BCG immunotherapy. *Cancer* 49: 1530, 1982.
- 4 Sauter C, Berchtold W, Fopp M, Gratwohl A, Imbach P, Maurice P, Tschopp L, von Fliedner V, Cavalli F. Acute myelogenous leukemia: Maintenance chemotherapy after early consolidation treatment does not prolong survival. *Lancet* 1: 379, 1984.
- 5 Vogler WR, Winton EE, Gordon DS, Raney MR, Go B, Meyer L. A randomized comparison of post remission therapy in acute myelogenous leukemia: A Southwestern Cancer Study Group trial. *Blood* **63**: 1039, 1984.
- 6 Preisler H, David RB, Kirshner J, Dupre E, Richards F 3d, Hoagland HC, Kopel S, Levy RN, Carey R, Schulman P, Gottlieb AJ, McIntyre OR. Comparison of 3 remission induction regimens and two postinduction regimens for the treatment of acute nonlymphocytic leukemia. *Blood* **69**: 1441, 1987.
- 7 Bishop JF, Lowenthal RM, Joshua D, Matthews JP, Todd D, Cobcroft R, Whiteside MG, Kronenberg H, Ma D, Dodds A, Herrman R, Szer J, Wolf MM, Young G. Etoposide in acute non-lymphocytic leukemia. *Blood* 75: 27, 1990.
- 8 Mayer RJ, Davis RB, Schiffer CA, Berg DT, Powell BL, Schulman P, Omura GA, Moore JO, McIntyre OR, Frei E 3rd. Intensive post-remission chemotherapy in adults with acute myeloid leukemia. *New England Journal of Medicine* 3331: 896, 1994.
- 9 Stone and Mayer. *Hematology Clinics of North America*, 7: 47, 1993.
- 10 Dombret H, Chastang C, Fenaux P, Reiffers J, Bordessoule D, Bouabdallah R, Mandelli F, Ferrant A, Auzanneau G, Tilly H. A controlled study of recombinant human granulocyte colony-stimulating factor in elderly patients after treatment for acute myelogenous leukemia. New England Journal of Medicine 332: 1678, 1995.
- 11 Stone RM, Berg DT, George SL, Dodge RK, Paciucci PA, Schulman P, Lee EJ, Moore JO, Powell BL, Schiffer CA. Granulocyte-macrophage colony-stimulating factor after initial chemotherapy for elderly patients with primary acute myelogenous leukemia. *New England Journal of Medicine* **332**: 1671, 1995.

- 12 Löwenberg B, Zittoun R, Kerkhofs H, Jehn U, Abels J, Debusscher L, Cauchie CH, Peetermans M, Solbu G, Suciu S, Stryckmans P. On the value of intensive remission-induction chemotherapy in elderly patients of 65+ years with acute myeloid leukemia: a randomized phase III study of the EORTC Leukemia Group. Journal of Clinical Oncology 1: 1268, 1993.
- 13 Liu Yin JA. Acute myeloid leukaemia in the elderly: biology and treatment. *British Journal of Haematology* **83:** 1, 1993.
- 14 Keating MJ, Estey E, Katarjian H. Acute Leukemia. In: DeVita Jr VT, Hellman S, and Rosemberg SA eds. *Cancer: Principles & Practice of Oncology*. JB Lippincott, Philadelphia, 1993. p 1938.
- 15 Bishop JF, Matthews JP, Young GA, Szer J, Gilett A, Joshua D, Bradstock K, Enno A, Wolf MM, Fox R, Cobcroft R, Herrmann R, Van Der Weyden M, Lowenthal RM, Page F, Garson OM, Juneja S. A randomized trial of high-dose cytarabine in induction in acute myeloid leukemia. *Blood* 87: 1710, 1996.
- 16 Weick JK, Kopecky TJ, Appelbaum FR, Head DR, Kingsbury LL, Balcerzak SP, Bickers JN, Hynes HE, Welborn JL, Simon SR, Grever M. A randomized investigation of high-dose versus standard-dose cytosine arabinoside with daunorubicin in patients with previously untreated acute myeloid leukemia: A Southwest Oncology Group study. *Blood* 88: 2841, 1996.
- 17 Beck WT. Multidrug resistance and its circumvention. *European Journal of Cancer* **26:** 513, 1990.
- 18 Borst P. Genetic mechanisms of drug resistance. A review. Acta Oncologica 30: 87, 1991.
- 19 Juliano RL, Ling V. A surface glycoprotein modulating drug permeability in Chinese hamster ovary cell mutants. *Biochemica Biophysica Acta* 455: 152, 1976.
- 20 Kartner N, Riordan JR, Ling V. Cell surface Pglycoprotein associated with multidrug resistance in mammalian cell lines. *Science* **221**: 1285, 1983.
- 21 Fojo AT, Whang-Peng J, Gottesmann MM. Amplification of DNA sequences in human multidrug-resistance KB carcinoma cells. *Proceedings of the National Academy* of Sciences USA 82: 7661, 1985.
- 22 Gros P, Neriah BY, Croop JM, Housman DE. Isolation and expression of a complementary DNA that confers multidrug resistance. *Nature* **323**: 728, 1986.
- 23 Goldstein LJ, Galski H, Fojo A, Willingham M, Lai SL, Gazdar A, Pirker R, Green A, Crist W, Brodeur GM. Expression of a multidrug resistance gene in human cancers. *Journal of the National Cancer Institute* **2:** 116, 1989.
- 24 Gottesman MM, Pastan I. Biochemistry of multidrug resistance mediated by the multidrug transporter. *Annual Reviews Biochemistry* **62:** 385, 1993.
- 25 Chaudhary PM, Roninson EB. Expression and activity of P-glycoprotein, a multidrug efflux pump, in human hematopoietic stem cells. *Cell* **66**: 85, 1991.
- 26 Pastan P, Schouten H. Multidrug resistance mediated by P-glycoprotein in haematological malignancies. *Netherlands Journal of Medicine* 42: 218, 1993.
- 27 Licht T, Pastan I, Gottesman M, Herrmann F. Pglycoprotein mediated multidrug resistance in normal and neoplastic haemopoietic cells. *Annals Hematology* **69:** 159, 1994.

- 28 Nooter K, Sonneveld P. Clinical relevance of Pglycoprotein expression in haematological malignancies. *Leukemia Research* 18: 223, 1994.
- 29 Marie JP. P-glycoprotein in adult hematological malignancies. *Hematology Oncology Clinics of North America* 9: 239, 1995.
- 30 Zhou D, Marie JP, Suberville A, Zittoun R. Relevance of MDR-1 gene expression in acute myeloid leukemia and comparison of different diagnostic methods. *Leukemia* 6: 879, 1992.
- 31 Leith CP, Kopecky KJ, Godwin J, McConnell T, Slovak ML, Chen IM, Head DR, Appelbaum FR, Willman CL. Acute myeloid leukemia in the elderly: assessment of multidrug resistance (MDR1) and cytogenetics distinguishes biologic subgroups with remarkably distinct responses to standard chemotherapy. A Southwest Oncology Group study. *Blood* 89: 3323, 1997.
- 32 Holmes J, West R. The effect of MDR1 gene expression on outcome in acute myeloid leukaemia. *British Journal* of Cancer **69**: 382, 1994.
- 33 Herweijer H, Sonneveld P, Baas F, Nooter K. Expression of MDR1 and mdr3 multidrug-resistance genes in human acute and chronic leukemias and association with stimulation of drug accumulation by cyclosporin. *Journal of the National Cancer Institute* **28**: 1133, 1990.
- 34 Kuwazuru Y, Yoshimura A, Hanada S, Utsunomiya A, Makino T, Ishibashi K, Kodama M, Iwahashi M, Arima T, Akiyama S. Expression of the multidrug transporter Pglycoprotein in acute leukemia cells and correlation to clinical drug resistance. *Cancer* 66: 868, 1990.
- 35 Pirker R, Wallner J, Geissler K, Linkesch W, Haas OA, Bettelheim P, Hopfner M, Scherrer R, Valent P, Havelec L, Ludwig H, Lechner K. MDR1 gene expression and treatment outcome in acute myeloid leukemia. *Journal of the National Cancer Institute* 83: 708, 1991.
- 36 Marie JP, Zittoun R, Sikic BI. Multidrug resistance (MDR1) gene expression in adult acute leukemias: correlations with treatment outcome and in vitro drug sensitivity. *Blood* **78**: 586, 1991.
- 37 Campos L, Guyotat D, Archimbaud E, Calmard-Oriol P, Tsuruo T, Trancy J, Treille D, Fiere D. Clinical significance of multidrug resistance P-glycoprotein expression on acute nonlymphoblastic leukemia cells at diagnosis. *Blood* **79**: 473, 1992.
- 38 Michieli M, M. Damiani D, Geromin A, Michelutti A, Fanin R, Raspadori D, Russo D, Visani G, Dinota A, Pileri S. Overexpression of multidrug resistance-associated p-170 glycoprotein in acute non-lymphocytic leukemia. *European Journal of Haematology* 48: 87, 1992.
- 39 Arceci RJ. Clinical significance of P-glycoprotein in multidrug resistance malignancies. *Blood* 81: 2215, 1993.
- 40 Te Boekhorst PAW, Wittebol S, Hagemeijer A, Lowenberg B, Nooter K, Sonneveld P. Predominance of functional multidrug resistance (MDR-1) phenotype in CD34+ leukemia cells. *Blood* 82: 3157, 1993.
- 41 Tirikainen M, Elonen E, Ruutu T, Jansson S, Krusius T. Clinical significance of P-glycoprotein expression in acute leukemia as analysed by immunocytochemistry. *European Journal of Haematology* **50:** 279, 1993.
- 42 Ino T, Miyazaki H, Tsogai M, Nomura T, Tsuzuki M, Tsuruo T, Ezaki K, Hirano M. Expression of Pglycoprotein in de novo acute myelogenous leukemia at initial diagnosis: results of molecular and functional assays, and correlation with treatment outcome. *Leukemia* **8**: 1492, 1994.

- 43 Lamy T, Goasguen D, Mordeletb E, Grulois I, Dauriac C, Drenou B, Chaperon J, Fauchet R, le Prise PY. P-glycoprotein (P-170) and CD34 expression in adult acute leukemia (AML). *Leukemia* 8: 1879, 1994.
- 44 Zoechbauerm S, Gsur A, Brunner R, Kyrle P, Lechner K, Pirker R. P-glycoprotein expression as unfavorable prognostic factor in acute myeloid leukemia. *Leukemia* 8: 975, 1994.
- 45 Wood P, Burgess R, McGregor A, Liu Yin J. Pglycoprotein expression on acute myeloid leukaemia blasts cells at diagnosis predicts response to chemotherapy and survival. *British Journal of Haematology* **87:** 509, 1994.
- 46 Basara N, Radosevic-Radojkovic N, Colovic M, Boskovic D, Rolovic Z. *In vitro* drug sensitivity of leukemic pro-genitors and P-glycoprotein expression in adult myeloid leukemia: correlation with induction treatment outcome. *European Journal of Haematology* 55: 83, 1995.
- 47 Guerci A, Merlin HL, Missoun N, Feldmann L, Marchal S, Guerci O. Predictive value for treatment outcome in acute myeloid leukemia of cellular daunorubicin accumulation and P-glycoprotein expression simultaneously deter-mined by flow cytometry. *Blood* 85: 2147, 1995.
- 48 Schuurhuis GJ, Broxterman HJ, Ossenkoppele GJ, Baak JP, Eekman CA, Kuiper CM, Feller N, van Heijningen TH, Klumper E, Pieters R, Lankelma J, Pinedo HM. Functional multidrug resistance phenotype associated with combined overexpression of PgP/MDR1 and MRP together with 1- $\beta$ -D-arabinofuranosylcytosine sensitivity may predict clinical response in acute myeloid leukemia. *Clinical Cancer Research* **1**: 81, 1995.
- 49 Del Poeta G, Stasi R, Aronica G, Venditti A, Cox MC, Bruno A, Buccisano F, Masi M, Tribalto M, Amadori S, Papa G. Clinical relevance of P-glycoprotein expression in de novo acute myeloid leukemia. *Blood* 87: 1997, 1996.
- 50 Willman C. The prognostic significance of the expression and function of multidrug resistance transporter proteins in acute myeloid leukemia: studies of the Southwest Oncology Group Leukemia Research Program. *Seminars in Hematology* **34** (Suppl 5): 25, 1997.
- 51 Van den Heuvel-Eibrink MM, Van der Holt B, Te Boekhorst PAW, Pieters R, Schoester M, Löwenberg B, Sonneveld P. MDR1 expression is an independent prognostic factor for response and survival in de novo acute myeloid leukaemia. *British Journal of Haematology* 99: 76, 1997.
- 52 Broxterman HJ, Sonneveld P, Feller N. Ossenkoppele GJ, Wahrer DCR, Eekman CA, Schoester M, Lankelma J, Pinedo HM, Lowenberg B, Schuurhuis GJ. Quality control of multidrug resistance assays in adult myeloid leukemia:correlation between assays for P-glycoprotein expression and activity. *Blood* 87: 4809, 1996.
- 53 List AF, Spier C, Greer J, Wollf S, Hutter J, Dorr R, Salmon SE, Futscher B, Baier M, Dalton W. Phase I/II trial of cyclosporine as a chemotherapy resistance modifier in acute leukemia. *Journal of Clinical Oncology* 11: 1652, 1994.
- 54 Sonneveld P, Durie BGM, Lokhorst HM, Marie JP, Solbu G, Suciu S, Zittoun R, Lowenberg B, Nooter K. Modulation of multidrug resistance multiple myeloma by cyclosporin. *Lancet* 340: 255–259, 1992.
- 55 Fisher GA, Lum BL, Hausdorff J, Sikic BI. Pharmacological considerations in the modulation of multidrug resistance. *European Journal of Cancer* **32A**: 1082, 1996.

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- 56 Boesch D, Muller K, Poutier-Manzanedo A, Loor F. Restoration of daunomycin retention in multidrug resistant P388 cells by submicromolar concentrations of SDZ PSC 833. *Experimental Cell Research* **196**: 26, 1991.
- 57 Twentyman PR. Modification of cytotoxic drug resistance by non-immunosuppressive cyclosporins. *British Journal of Cancer* 57: 254, 1988.
- 58 Twentyman PR, Bleehen NM. Resistance modification by PSC-833, a novel non-immunosuppressive cyclosporin. *European Journal of Cancer* 27: 1539, 1991.
- 59 Foxwell BM, Mackie A, Ling V, et al. Identification of the multidrug resistance-related P-glycoprotein as a cyclosporine binding protein. *Molecular Pharmacology* 36: 534, 1989.
- 60 Twentyman PR. Cyclosporins as drug resistance modifiers. *Biochemical Pharmacology* **43**: 109, 1992.
- 61 Archinal Mattheis A, Rzepka RW, Watanabe T, Kokubu N, Itoh Y, Combates NJ, Bair KW, Cohen D. Analysis of the interactions of SDZ PSC 833 ([3'keto-BMT1]-Val2]cyclosporine), a multidrug resistance modulator, with P-glycoprotein. *Oncology Research* 7: 603, 1995.
- 62 Boote DJ, Dennis IF, Twentyman PR, Osborne RJ, Laburte C, Hensel S, Smyth JF, Brampton MH, Bleehen NM. Phase I study of etoposide with SDZ PSC 833 as a modulator of multidrug resistance in patients with cancer. *Journal of Clinical Oncology* 14: 610, 1996.
- 63 Visani G, Milligan D, Leoni F, et al. A phase I dosefinding study of PSC 833, a novel MDR reversing agent, with mitoxantrone, etoposide and cytarabine (PSC-MEC) in poor prognosis acute leukemia (AML). Blood 90 (Suppl 1) Abstract 2518, 1997.
- 64 Advani R, Saba H, Tallman M, Rowe JM, Wiernik PH, Ramek J, Dugan K, Lum B, Villena J, Davis E, Paietta E, Litchman M, Covelli A, Sikic B, Greenberg P. Treatment of poor prognosis AML patients with PSC 833 plus mitoxantrone, etoposide and cytarabine (PSC-MEC). *Blood* 90 (Suppl 1) Abstract 2260, 1997.
- 65 Sonneveld P, Marie JP, Huisman C, Vekhoff A, Schoester M, Faussat AM, Van Kapel J, Groenewegen A, Charnick S, Zittoun R, Löwenberg B. Reversal of multidrug resistance by SDZ PSC 833, combined with VAD (vincristine, doxorubicin, dexamethasone) in refractory multiple myeloma. A phase I study. *Leukemia* 10: 1741, 1996.
- 66 Sikic BJ. Pharmacologic approaches to reversing multidrug resistance. *Seminars in Hematology* 34 (Suppl 5): 40, 1997.
- 67 Ehrlichman C, Moore MJ, De Angelis C, Goodman P, Manzo J. The pharmacokinetics and bioavailability of a new chemosensitizer, SDZ PSC 833, in patients with advanced cancer. *Anticancer Drugs* 5: 44, 1994.

- 68 Paul C, Liliemark J, Tidefel U, Gahrton G, Peterson C. Pharmacokinetics of daunorubicin and doxorubicin in plasma and leukemic cells from patients with acute nonlymphoblastic leukemia. *Therapeutic Drug Monitoring* **11**: 140, 1989.
- 69 Robert J, Rigal-Hueguet F, Hurteloup P. Comparative pharmacokinetic study of idarubicin and daunorubicin in leukemia patients. *Hematol Oncol* **10**: 111, 1992.
- 70 Speth PA, Linssen PCM, Boezemn JBM, Wessels HMC, Haanen C. Leukemic cell and plasma daunomycin concentrations after bolus injection and 72 h infusion. *Cancer Chemotherapy Pharmacology* **20:** 311, 1987.
- 71 Rahman A, Goodman A, Foo W, Harvey J, Smith FP, Schein PS. Clinical pharmacology of daunorubicin in Phase I patients with solid tumors: development of an analytical methodology for daunorubicin and its metabolites. *Seminars in Oncology* **11** (Suppl 3): 36, 1984.
- 72 Riggs CE. Clinical pharmacology of daunorubicin in patients with acute leukemia. *Seminars in Oncology* **11** (Suppl 3): 2, 1984.
- 73 Hullhoven R, Sokal G, Harvengt C. Human pharmacokinetics of daunorubicin-DNA complex. *Cancer Chemotherapy Pharmacology* **3**: 243, 1979.
- 74 Huffman DH, Benjamin RS, Bachur NR. Daunorubicin metabolism in acute non-lymphocytic leukemia. *Clinical Pharmacology Therapy* 13: 895, 1972.
- 75 Kokenberg E, Sonneveld P, Sizoo W, Hagenbeek A, Löwenberg B. Cellular pharmacokinetics of daunorubicin:relationships with the response to treatment in patients with acute myeloid leukemia. *Journal of Clinical Oncology* **6**: 802, 1988.
- 76 Kahn SB, Begg CB, Mazza JJ, Bennett JM, Bonner H, Glick JH. Full dose versus attenuated dose daunomycin, cytosine, arabinoside and 6-thioguanine in the treatment of acute nonlymphocytic leukemia in the elderly. *Journal* of Clinical Oncology 2: 865, 1981.
- 77 Löwenberg B, Suciu S, Archimbaud E, Ossenkoppele G, Verhoef GEG, Vellenga E, Wijermans P, Berneman Z, Dekker AW, Stryckmans P, Shouten H, Jehn U, Muus P, Sonneveld P, Dardenne M, Zittoun R. Use of recombinant Granulocyte-Macrophage Colony-Stimulating Factor during and after remission induction chemotherapy in patients aged 61 years and older with acute myeloid leukemia (AML):Final report of AML-11, a phase III randomized study of the Leukemia Cooperative Group of European Organisation for the Research and Treatment of Cancer (EORTC-LCG) and the Dutch Hemato-Oncology Cooperative Group. *Blood* **90**: 2952, 1997.
- 78 Galettis P, Boutagy J, Ma DDF. Daunorubicin pharmacokinetics and the correlation with P-glycoprotein and response in patients with acute leukemia. *British Journal of Cancer* **70**: 324, 1994.