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DATA NOTE



The genome sequence of a drosophilid fruit fly,

Hirtodrosophila cameraria (Haliday, 1833) [version 1; peer

review: awaiting peer review]

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Abstract

We present a genome assembly from an individual female *Hirtodrosophila cameraria* (a drosophilid fruit fly; Arthropoda; Insecta; Diptera; Drosophilidae). The genome sequence is 214.5 megabases in span. Most of the assembly is scaffolded into 4 chromosomal pseudomolecules. The mitochondrial genome has also been assembled and is 15.94 kilobases in length.

Keywords

Hirtodrosophila cameraria, a drosophilid fruit fly, genome sequence, chromosomal, Diptera



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Any reports and responses or comments on the article can be found at the end of the article.

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Species taxonomy

Eukaryota; Metazoa; Eumetazoa; Bilateria; Protostomia; Ecdysozoa; Panarthropoda; Arthropoda; Mandibulata; Pancrustacea; Hexapoda; Insecta; Dicondylia; Pterygota; Neoptera; Endopterygota; Diptera; Brachycera; Muscomorpha; Eremoneura; Cyclorrhapha; Schizophora; Acalyptratae; Ephydroidea; Drosophilidae; Drosophilinae; Drosophilini; *Hirtodrosophila*; *Hirtodrosophila cameraria* (Haliday, 1833) (NCBI:txid1262473).

Background

Hirtodrosophila cameraria (Haliday, 1833) is a medium sized (2.5-3.5 mm) pale greyish-brown drosophilid 'fruit fly' (Figure 1A and B), distantly related to the laboratory model Drosophila melanogaster. It is one of three British and Irish species currently classified in the genus Hirtodrosophila (Chandler, 2023). Originally placed in the genus Drosophila by Haliday, it was moved to the newly-elevated (sub-)genus Hirtodrosophila (Grimaldi, 1990) by Bächli et al. (2004). However, relationships between Drosophila and Hirtodrosophila remain unclear, with the genera being paraphyletic with respect to each other and Zygothrica and Mycodrosophila, and with no single diagnostic morphological character available to separate them (Bächli et al., 2004; Finet et al., 2021; Gautério et al., 2020; Grimaldi, 1990). Nevertheless, H. cameraria itself is easy to identify among other similar British and Irish drosophilids, having plumose aristae with a single ventral branch behind the terminal fork and lacking a pre-apical seta on the mesotibia (Bächli et al., 2004).

Like its close relatives, *H. cameraria* is a specialist fungus breeder and in the UK the adults are easily collected or reared from toadstools and bracket fungi, including *Phallus impudicus*, *Lactarius quietus*, *Amanita rubescens*, *Russula* cyanorantha and Paxillus (Charlesworth & Shorrocks, 1980; Gautério et al., 2020; Shorrocks & Charlesworth, 1980). Adults have also been reported in association with the violet helleborine orchid Epipactis purpurata (Roper, 2013), perhaps as a result of pheromones released by the plant to attract pollinators (Policha et al., 2019). Hirtodrosophila cameraria is broadly distributed in wooded areas across Europe, from the extreme north of Sweden, to Turkey in the east, and the Canary islands in the south west (Bächli, 2023). In the UK, breeding is likely to be focused of toadstool flushes in the late summer and early autumn (Charlesworth & Shorrocks, 1980), but adults can be caught at any time of year (Basden, 1954), and are quite commonly recorded in the months of May to November (GBIF Secretariat, 2022). It is not thought to be threatened and is in fact likely under-collected compared to its human-commensal relatives, as it rarely comes to fruit bait (Basden, 1954).

Here we present a chromosomally complete genome sequence for *Hirtodrosophila cameraria*, derived from the DNA of two female specimens that were collected from a bracket fungus in the Hermitage of Braid, Edinburgh, as part of the Darwin Tree of Life Project. This genome sequence will help to resolve relationships among the Drosophilidae and will further build on the value of this family as a model clade for comparative genomics and molecular evolution. This project is a collaborative effort to sequence all named eukaryotic species in the Atlantic Archipelago of Britain and Ireland.

Genome sequence report

The genome was sequenced from one female *Hirtodrosophila cameraria* (Figure 1C) collected from Hermitage of Braid, Edinburgh, Scotland (55.92, -3.20). A total of 45-fold



Figure 1. Photographs of Hirtodrosophila cameraria specimens and locale. A: Male (above) and female (below) *Hirtodrosophila cameraria* presented with a 3 mm scale bar. **B**: Tree stump from which the sequenced individuals were collected (Hermitage of Braid, Edinburgh, Scotland; 55.92, -3.20). **C**: The four unrelated wild-collected females provided to the Darwin Tree of Life project. Individual idHirCame1 (biospecimen SAMEA12110595) (left) was used for Hi-C sequencing, and individual idHirCame2 (biospecimen SAMEA12110596) (second left) was used for PacBio sequencing. **D**: Female *Hirtodrosophila cameraria* photographed above a toadstool in Perth & Kinross, Scotland.

coverage in Pacific Biosciences single-molecule HiFi long reads was generated. Primary assembly contigs were scaffolded with chromosome conformation Hi-C data. Manual assembly curation corrected 78 missing joins or misjoins and removed 5 haplotypic duplications, reducing the assembly length by 0.32% and the scaffold number by 52.38%, and increasing the scaffold N50 by 101.28%.

The final assembly has a total length of 214.5 Mb in 39 sequence scaffolds with a scaffold N50 of 82.8 Mb (Table 1). Most (98.38%) of the assembly sequence was assigned to 4 chromosomal-level scaffolds. Chromosome-scale scaffolds confirmed by the Hi-C data are named in order of size (Figure 2–Figure 5; Table 2). We did not identify the sex chromosome(s) as sequence data from the heterogametic sex

Project accession data					
Assembly identifier	idHirCame2.1				
Species	Hirtodrosophila cameraria				
Specimen	idHirCame2				
NCBI taxonomy ID	1262473				
BioProject	PRJEB56615				
BioSample ID	SAMEA12110596				
Isolate information	idHirCame2, female (DNA sequencing) idHirCame1 (Hi-C scaffolding)				
Assembly metrics*		Benchmark			
Consensus quality (QV)	57.9	≥ 50			
k-mer completeness	99.99%	≥95%			
BUSCO**	C:99.1%[S:97.8%,D:1.3%], F:0.3%,M:0.6%,n:3,285	C ≥ 95%			
Percentage of assembly mapped to chromosomes	98.38%	≥ 95%			
Sex chromosomes	Not assigned	localised homologous pairs			
Organelles	Mitochondrial genome assembled	complete single alleles			
Raw data accessions					
PacificBiosciences SEQUEL II	ERR10368987				
Hi-C Illumina	ERR10378038				
Genome assembly					
Assembly accession	GCA_949708635.1				
Accession of alternate haplotype	GCA_949708645.1				
Span (Mb)	214.5				
Number of contigs	419				
Contig N50 length (Mb)	1.0				
Number of scaffolds	39				
Scaffold N50 length (Mb)	82.8				
Longest scaffold (Mb)	93.4				

Table 1. Genome data for *Hirtodrosophila cameraria*, idHirCame2.1.

* Assembly metric benchmarks are adapted from column VGP-2020 of "Table 1: Proposed standards and metrics for defining genome assembly quality" from (Rhie *et al.*, 2021).

** BUSCO scores based on the diptera_odb10 BUSCO set using v5.3.2. C = complete [S = single copy, D = duplicated], F = fragmented, M = missing, n = number of orthologues in comparison. A full set of BUSCO scores is available at https://blobtoolkit.genomehubs.org/view/idHirCame2.1/dataset/CATIWZ01/busco.



Figure 2 Genome assembly of *Hirtodrosophila cameraria*, **idHirCame2.1: metrics.** The BlobToolKit Snailplot shows N50 metrics and BUSCO gene completeness. The main plot is divided into 1,000 size-ordered bins around the circumference with each bin representing 0.1% of the 214,511,801 bp assembly. The distribution of scaffold lengths is shown in dark grey with the plot radius scaled to the longest scaffold present in the assembly (93,358,477 bp, shown in red). Orange and pale-orange arcs show the N50 and N90 scaffold lengths (82,766,246 and 31,806,066 bp), respectively. The pale grey spiral shows the cumulative scaffold count on a log scale with white scale lines showing successive orders of magnitude. The blue and pale-blue area around the outside of the plot shows the distribution of GC, AT and N percentages in the same bins as the inner plot. A summary of complete, fragmented, duplicated and missing BUSCO genes in the diptera_odb10 set is shown in the top right. An interactive version of this figure is available at https://blobtoolkit.genomehubs.org/view/idHirCame2.1/dataset/CATIWZ01/snail.

was not available and homology is unreliable for sex chromosome identification in Diptera due to frequent sex chromosome turnover (Vicoso & Bachtrog, 2015). While not fully phased, the assembly deposited is of one haplotype. Contigs corresponding to the second haplotype have also been deposited. The mitochondrial genome was also assembled and can be found as a contig within the multifasta file of the genome submission.

The estimated Quality Value (QV) of the final assembly is 57.9 with *k*-mer completeness of 99.99%, and the assembly has a BUSCO v5.3.2 completeness of 99.1% (single = 97.8%, duplicated = 1.3%), using the diptera_odb10 reference set (n = 3,285).

Metadata for specimens, spectral estimates, sequencing runs, contaminants and pre-curation assembly statistics can be found at https://links.tol.sanger.ac.uk/species/1262473.

Methods

Sample acquisition and nucleic acid extraction

The specimen used for genome sequencing was a female *Hirtodrosophila cameraria* (biospecimen ID SAMEA12110596, individual idHirCame2; Figure 1C). The specimen used for Hi-C scaffolding was a female *Hirtodrosophila cameraria* (biospecimen ID SAMEA12110595, individual idHirCame1; Figure 1C). Both specimens were collected from Hermitage of Braid, Edinburgh, Scotland, UK (latitude 55.92, longitude –3.20) on 2021-10-04. The specimens were aspirated



Figure 3. Genome assembly of *Hirtodrosophila cameraria*, idHirCame2.1: BlobToolKit GC-coverage plot. Scaffolds are coloured by phylum. Circles are sized in proportion to scaffold length. Histograms show the distribution of scaffold length sum along each axis. An interactive version of this figure is available at https://blobtoolkit.genomehubs.org/view/idHirCame2.1/dataset/CATIWZ01/blob.

from a bracket fungus on a deciduous tree stump in an urban woodland. The anaesthetised flies were placed directly into collection tube, and frozen from live at -80° C. The specimens were collected and identified by Darren Obbard (University of Edinburgh).

DNA was extracted at the Tree of Life laboratory, Wellcome Sanger Institute (WSI). The idHirCame2 sample was weighed and dissected on dry ice. Tissue from the whole organism was disrupted using a Nippi Powermasher fitted with a BioMasher pestle. High molecular weight (HMW) DNA was extracted using the Qiagen MagAttract HMW DNA extraction kit. HMW DNA was sheared into an average fragment size of 12–20 kb in a Megaruptor 3 system with speed setting 30. Sheared DNA was purified by solid-phase reversible immobilisation using AMPure PB beads with a 1.8X ratio of beads to sample to remove the shorter fragments and concentrate the DNA sample. The concentration of the sheared and purified DNA was assessed using a

Nanodrop spectrophotometer and Qubit Fluorometer and Qubit dsDNA High Sensitivity Assay kit. Fragment size distribution was evaluated by running the sample on the FemtoPulse system.

Sequencing

Pacific Biosciences HiFi circular consensus DNA sequencing libraries were constructed according to the manufacturers' instructions. DNA sequencing was performed by the Scientific Operations core at the WSI on a Pacific Biosciences SEQUEL II (HiFi) instrument. Hi-C data were also generated from whole organism tissue of idHirCame1 using the Arima2 kit and sequenced on the Illumina NovaSeq 6000 instrument.

Genome assembly, curation and evaluation

Assembly was carried out with Hifiasm (Cheng *et al.*, 2021) and haplotypic duplication was identified and removed with purge_dups (Guan *et al.*, 2020). The assembly was



Figure 4. Genome assembly of *Hirtodrosophila cameraria*, idHirCame2.1: BlobToolKit cumulative sequence plot. The grey line shows cumulative length for all scaffolds. Coloured lines show cumulative lengths of scaffolds assigned to each phylum using the buscogenes taxrule. An interactive version of this figure is available at https://blobtoolkit.genomehubs.org/view/idHirCame2.1/dataset/ CATIWZ01/cumulative.



Figure 5. Genome assembly of *Hirtodrosophila cameraria*, idHirCame2.1: Hi-C contact map of the idHirCame2.1 assembly, visualised using HiGlass. Chromosomes are shown in order of size from left to right and top to bottom. An interactive version of this figure may be viewed at https://genome-note-higlass.tol.sanger.ac.uk/l/?d=WPeKseKJS1uSeK0NS7ZsOA.

Table 2. Chromosomal pseudomolecules in the genome assembly of *Hirtodrosophila cameraria*, idHirCame2.

INSDC accession	Chromosome	Length (Mb)	GC%
OX453089.1	1	93.36	37.0
OX453090.1	2	82.77	36.5
OX453091.1	3	31.81	38.0
OX453092.1	4	3.12	35.0
OX453093.1	MT	0.02	21.5

then scaffolded with Hi-C data (Rao *et al.*, 2014) using YaHS (Zhou *et al.*, 2023). The assembly was checked for contamination and corrected as described previously (Howe *et al.*, 2021). Manual curation was performed using HiGlass (Kerpedjiev *et al.*, 2018) and Pretext (Harry, 2022). The mitochondrial genome was assembled using MitoHiFi (Uliano-Silva *et al.*, 2023), which runs MitoFinder (Allio *et al.*, 2020) or MITOS (Bernt *et al.*, 2013) and uses these annotations to select the final mitochondrial contig and to ensure the general quality of the sequence.

A Hi-C map for the final assembly was produced using bwa-mem2 (Vasimuddin *et al.*, 2019) in the Cooler file format (Abdennur & Mirny, 2020). To assess the assembly metrics, the *k*-mer completeness and QV consensus quality values were calculated in Merqury (Rhie *et al.*, 2020). This work was done using Nextflow (Di Tommaso *et al.*, 2017) DSL2 pipelines "sanger-tol/readmapping" (Surana *et al.*, 2023a) and "sanger-tol/genomenote" (Surana *et al.*, 2023b). The genome was analysed within the BlobToolKit environment (Challis *et al.*, 2020) and BUSCO scores (Manni *et al.*, 2021; Simão *et al.*, 2015) were calculated.

Table 3 contains a list of relevant software tool versions and sources.

Wellcome Sanger Institute - Legal and Governance

The materials that have contributed to this genome note have been supplied by a Tree of Life collaborator. The Wellcome Sanger Institute employs a process whereby due diligence is carried out proportionate to the nature of the materials themselves, and the circumstances under which they have been/are to be collected and provided for use. The purpose of this is to address and mitigate any potential legal and/or ethical implications of receipt and use of the materials as part of the research project, and to ensure that in doing so we align with best practice wherever possible. The overarching areas of consideration are:

- · Ethical review of provenance and sourcing of the material
- Legality of collection, transfer and use (national and international)

Each transfer of samples is undertaken according to a Research Collaboration Agreement or Material Transfer Agreement

Table 3. Software	tools:	versions	and	sources.
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Software tool	Version	Source
BlobToolKit	4.1.7	https://github.com/blobtoolkit/ blobtoolkit
BUSCO	5.3.2	https://gitlab.com/ezlab/busco
Hifiasm	0.16.1-r375	https://github.com/chhylp123/ hifiasm
HiGlass	1.11.6	https://github.com/higlass/ higlass
Merqury	MerquryFK	https://github.com/ thegenemyers/MERQURY.FK
MitoHiFi	2	https://github.com/ marcelauliano/MitoHiFi
PretextView	0.2	https://github.com/wtsi-hpag/ PretextView
purge_dups	1.2.3	https://github.com/dfguan/ purge_dups
sanger-tol/ genomenote	v1.0	https://github.com/sanger-tol/ genomenote
sanger-tol/ readmapping	1.1.0	https://github.com/sanger-tol/ readmapping/tree/1.1.0
YaHS	yahs- 1.1.91eebc2	https://github.com/c-zhou/yahs

entered into by the Tree of Life collaborator, Genome Research Limited (operating as the Wellcome Sanger Institute) and in some circumstances other Tree of Life collaborators.

Data availability

European Nucleotide Archive: *Hirtodrosophila cameraria*. Accession number PRJEB56615; https://identifiers.org/ena.embl/ PRJEB56615. (Wellcome Sanger Institute, 2023)

The genome sequence is released openly for reuse. The *Hir*todrosophila cameraria genome sequencing initiative is part of the Darwin Tree of Life (DToL) project. All raw sequence data and the assembly have been deposited in INSDC databases. The genome will be annotated using available RNA-Seq data and presented through the Ensembl pipeline at the European Bioinformatics Institute. Raw data and assembly accession identifiers are reported in Table 1.

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Members of the Wellcome Sanger Institute Tree of Life programme are listed here: https://doi.org/10.5281/zenodo.4783585.

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Members of the Tree of Life Core Informatics collective are listed here: https://doi.org/10.5281/zenodo.5013541.

Members of the Darwin Tree of Life Consortium are listed here: https://doi.org/10.5281/zenodo.4783558.

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References

Abdennur N, Mirny LA: Cooler: Scalable storage for Hi-C data and other genomically labeled arrays. *Bioinformatics*. 2020; 36(1): 311–316. PubMed Abstract | Publisher Full Text | Free Full Text

Allio R, Schomaker-Bastos A, Romiguier J, et al.: MitoFinder: Efficient automated large-scale extraction of mitogenomic data in target enrichment phylogenomics. Mol Ecol Resour. 2020; 20(4): 892–905. PubMed Abstract | Publisher Full Text | Free Full Text

Bächli G: TaxoDros: The Database on Taxonomy of Drosophilidae. 2023; Accessed: 29 May 2023. **Reference Source**

Bächli G, Vilela CR, Escher SA, *et al.*: **The Drosophilidae (Diptera) of Fennoscandia and Denmark.** Leiden: Brill Academic Publishers, 2004. **Reference Source**

Basden EB: **The distribution and biology of Drosophilidae (Diptera) in Scotland, including a new species of Drosophila**. Transactions of the Royal Society of Edinburgh. 1954; **62**(4): 603–654.

Bernt M, Donath A, Jühling F, et al.: MITOS: Improved de novo metazoan mitochondrial genome annotation. Mol Phylogenet Evol. 2013; 69(2): 313–319. PubMed Abstract | Publisher Full Text

Challis R, Richards E, Rajan J, et al.: BlobToolKit - interactive quality assessment of genome assemblies. G3 (Bethesda). 2020; **10**(4): 1361–1374. PubMed Abstract | Publisher Full Text | Free Full Text

Chandler P: Checklist of Diptera of the British Isles. 2023. Accessed: 15 May 2023.

Reference Source

Charlesworth P, Shorrocks B: The reproductive biology and diapause of the British fungal-breeding Drosophila. Ecol Entomol. 1980; 5(4): 315-326 **Publisher Full Text**

Cheng H, Concepcion GT, Feng X, et al.: Haplotype-resolved de novo assembly using phased assembly graphs with hifiasm. Nat Methods. 2021; 18(2): 170-175

PubMed Abstract | Publisher Full Text | Free Full Text

Di Tommaso P, Chatzou M, Floden EW, et al.: Nextflow enables reproducible computational workflows. Nat Biotechnol. 2017; 35(4): 316-319. PubMed Abstract | Publisher Full Text

Finet C, Kassner VA, Carvalho AB, et al.: DrosoPhyla: Resources for Drosophilid Phylogeny and Systematics. Genome Biol Evol. 2021; 13(8): evab179. PubMed Abstract | Publisher Full Text | Free Full Text

Gautério TB, Machado S, da Silva Loreto EL, et al.: Phylogenetic relationships between fungus-associated Neotropical species of the genera Hirtodrosophila, Mycodrosophila and Zygothrica (Diptera, Drosophilidae), with insights into the evolution of breeding sites usage. Mol Phylogenet Evol. 2020; 145(8): 106733 PubMed Abstract | Publisher Full Text

GBIF Secretariat: Hirtodrosophila cameraria (Haliday, 1833). GBIF Backbone Taxonomy. Checklist dataset. 2022; Accessed: 29 May 2023. **Reference Source**

Grimaldi DA: A phylogenetic revised classification of genera in the Drosophilidae (Diptera). Bulletin of the American Museum of Natural History. 1990; **197**: 1–139.

Reference Source

Guan D, McCarthy SA, Wood J, et al.: Identifying and removing haplotypic duplication in primary genome assemblies. Bioinformatics. 2020; 36(9): 2896-2898

PubMed Abstract | Publisher Full Text | Free Full Text

Haliday AH: Catalogue of Diptera occurring about Holywood in Downshire. Entomological Magazine. 1833; 1: 147-180. **Reference Source**

Harry E: PretextView (Paired REad TEXTure Viewer): A desktop application for viewing pretext contact maps. 2022; [Accessed 19 October 2022]. **Reference Source**

Howe K, Chow W, Collins J, et al.: Significantly improving the quality of genome assemblies through curation. GigaScience. Oxford University Press, 2021; 10(1): giaa153.

PubMed Abstract | Publisher Full Text | Free Full Text

Kerpedjiev P, Abdennur N, Lekschas F, et al.: HiGlass: web-based visual exploration and analysis of genome interaction maps. Genome Biol. 2018; 19(1): 125

PubMed Abstract | Publisher Full Text | Free Full Text

Manni M, Berkeley MR, Seppey M, et al.: BUSCO update: Novel and streamlined workflows along with broader and deeper phylogenetic coverage for scoring of eukaryotic, prokaryotic, and viral genomes. *Mol Biol Evol.* 2021; **38**(10): 4647–4654.

PubMed Abstract | Publisher Full Text | Free Full Text

Policha T, Grimaldi DA, Manobanda R, et al.: Dracula orchids exploit guilds of fungus visiting flies: new perspectives on a mushroom mimic. Ecol Entomol. 2019; 44(4): 457-470. **Publisher Full Text**

Rao SSP, Huntley MH, Durand NC, et al.: A 3D map of the human genome at kilobase resolution reveals principles of chromatin looping. Cell. 2014; 159(7): 1665–1680. PubMed Abstract | Publisher Full Text | Free Full Text

Rhie A, McCarthy SA, Fedrigo O, *et al.*: Towards complete and error-free genome assemblies of all vertebrate species. *Nature*. 2021; **592**(7856): 737-746

PubMed Abstract | Publisher Full Text | Free Full Text

Rhie A, Walenz BP, Koren S, et al.: Merqury: Reference-free quality, completeness, and phasing assessment for genome assemblies. Genome Biol. 2020; 21(1): 245

PubMed Abstract | Publisher Full Text | Free Full Text

Roper P: Fruit Flies (Diptera, Drosophildiae) and orchids. Dipterists Digest. 2013: 20: 62.

Shorrocks B, Charlesworth P: The distribution and abundance of the British fungal-breeding Drosophila. Ecological Entomology. 1980; 5(1): 61-78. **Publisher Full Text**

Simão FA, Waterhouse RM, Ioannidis P, et al.: BUSCO: assessing genome assembly and annotation completeness with single-copy orthologs. Bioinformatics. 2015; 31(19): 3210–2. PubMed Abstract | Publisher Full Text

Surana P, Muffato M, Qi G: sanger-tol/readmapping: sanger-tol/ readmapping v1.1.0 - Hebridean Black (1.1.0). Zenodo. 2023a; [Accessed 21 July 2023]. Publisher Full Text

Surana P, Muffato M, Sadasivan Baby C: sanger-tol/genomenote (v1.0.dev). Zenodo. 2023b; Accessed: 21 July 2023. **Reference Source**

Uliano-Silva M, Ferreira JGRN, Krasheninnikova K, et al.: MitoHiFi: a python pipeline for mitochondrial genome assembly from PacBio high fidelity reads. BMC Bioinformatics. 2023; 24(1): 288.

PubMed Abstract | Publisher Full Text | Free Full Text Vasimuddin M, Misra S, Li H, et al.: Efficient Architecture-Aware Acceleration

of BWA-MEM for Multicore Systems. In: 2019 IEEE International Parallel and Distributed Processing Symposium (IPDPS). IEEE, 2019; 314–324. Publisher Full Text

Vicoso B, Bachtrog D: Numerous Transitions of Sex Chromosomes in Diptera. *PLoS Biol.* Edited by H.S. Malik, 2015; **13**(4): e1002078. PubMed Abstract | Publisher Full Text | Free Full Text

Wellcome Sanger Institute: **The genome sequence of a drosophilid fruit fly**, *Hirtodrosophila cameraria* (Haliday, 1833). European Nucleotide Archive. [dataset], accession number PRJEB56615, 2023.

Zhou C, McCarthy SA, Durbin R: **YaHS: yet another Hi-C scaffolding tool.** *Bioinformatics*. Edited by C. Alkan, 2023; **39**(1): btac808. PubMed Abstract | Publisher Full Text | Free Full Text