Review of dohan eherenfest et al. (2009) on "classification of platelet concentrates: from pure platelet-rich plasma (p-prp) to leucocyte- and platelet-rich fibrin (l-prf)"

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REVIEW OF DOHAN EHERENFEST ET AL. (2009) ON "CLASSIFICATION OF PLATELET CONCENTRATES: FROM PURE PLATELET-RICH PLASMA (P-PRP) TO LEUCOCYTE- AND PLATELET-RICH FIBRIN (L-PRF)"

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1	REVIEW	OF	DOHAN	EHERENFEST	ET	AL.	(2009)	ON	"CLASSIFICATION	OF	PLATELET
2	CONCEN	TRAT	ES: FROM	PURE PLATEL	ET-R	ICH P	LASMA	(P-PRI	P) TO LEUCOCYTE-	AND	PLATELET-
3	RICH FIBI	RIN (I	L-PRF)"								

4 ABSTRACT

This classic discusses the original publication of Dohan Eherenfest et al. on "Classification of platelet 5 6 concentrates: from pure platelet-rich plasma (P-PRP) to leucocyte- and platelet-rich fibrin (L-PRF)", 7 in which the authors propose four categories of platelet concentrates depending on their leukocyte 8 and fibrin content (P-PRP, leucocyte- and platelet-rich plasma (L-PRP), pure platelet-rich fibrin (P-9 PRF), and L-PRF) to group a "jungle" of products in which the term platelet-rich plasma (PRP) was 10 used indistinctly. They were able to identify common factors such as: (1) the use of anticoagulant 11 and immediate centrifugation of the blood after its collection, (2) most preparation techniques 12 allowed platelet concentrate preparation within an hour, (3) the centrifugation aimed to separate 13 the blood in layers that would allow the extraction of specific fractions, and (4) the product was 14 activated with thrombin or calcium chloride. The reviewed manuscript has been listed among the 15 most cited PRP articles in regenerative medicine, with more than 800 citations, driving the current scientific research and clinical practice by categorizing L-PRP and P-PRP (now, leukocyte-poor PRP). 16 17 The classification has also opened the door to understanding intrinsic biological mechanisms 18 between the platelets, leukocytes, fibrin, and growth factors, later considered for studying the 19 proliferation and differentiation of cells in different tissues affected by PRP. Since the initial 20 classification of platelet concentrates, several other classification systems have been proposed and 21 published in the current literature, such as the PAW, Mishra, PLRA, DEPA, MARSPILL, etc. These 22 classifications have identified important aspects of PRP that affect the biological composition and, 23 ultimately, the indications and outcomes. To date, there is still a lack of standardization in sample

- 24 preparation, cohort heterogeneity, and incomplete reporting of sample preparation utilized, leading
- 25 to a lack of clarity and challenging researchers and clinicians.
- 26 Keywords: platelet-rich plasma, classification, platelet concentrates, orthobiologics, growth factors,
- 27 leukocytes.

Journal Prevention

28 ABBREVIATIONS

- 29 DEPA Dose of platelet, efficiency, purity, and activation
- 30 L-PRF Leucocyte- and platelet-rich fibrin
- 31 L-PRP Leucocyte- and platelet-rich plasma
- 32 MARSPILL Method, activation, red blood cells, spin, platelets, image guidance, leukocytes, and
- 33 light activation
- 34 MIBO Minimum information for studies evaluating biologics in orthopaedics
- 35 PAW Platelet, activation, white blood cell
- 36 PLRA Platelet, leukocyte, red blood cells, and activation
- 37 P-PRF Pure platelet-rich fibrin
- 38 P-PRP Pure platelet-rich plasma (now, leukocyte-poor platelet-rich plasma)
- 39 PRP Platelet-rich plasma

40 INTRODUCTION

Platelet-rich plasma (PRP) is a worldwide implemented regenerative medicine therapy. The clinical
applications and use of platelet-rich therapy in medicine and surgery have thrived over the past two
decades. It was described in 1970 as a plasma portion from autologous blood with increased platelet
concentration obtained by a centrifugation process [1].

PRP harnesses the signaling molecules and growth factors of platelets such as vascular endothelial growth factor (VEGF), fibroblast growth factor (FGF), platelet-derived growth factor (PDGF), epidermal growth factor, insulin-like growth factors (e.g., IGF-1, IFF-2), and others that enhance the natural healing potential of tissues, pain and inflammation modulation, and functional improvement [1].

50 PRP is currently used to treat multiple musculoskeletal conditions, including knee osteoarthritis, 51 cartilage injuries, patellar tendonitis, and tennis elbow [2, 3]. The media, celebrity athletes, the 52 desire for novel treatments, and its autologous nature have boosted its use in multiple sports-53 related injuries during the last decades [4-7]. As a result, its high demand has led to an industry-54 driven development of various platelet concentrate systems and products exceeding the pace of 55 evidence-based practice [1, 3, 7, 8].

In 2009, Dohan Eherenfest et al. proposed the first attempt at PRP classification to categorize the platelet concentrates concerning their fibrin and leukocyte content and the degree of standardization of the procedure, providing an overview of the available systems [9]. This classification later inspired authors to investigate the role of the different PRP components and the development of new classifications and reporting guidelines, highlighting leukocyte properties in orthobiologics products. However, the goal of standardization still seems far. A systematic review by Magalon et al. [8] revealed a great heterogeneity among fifty platelet concentrate products from

forty companies, which may explain the inconsistent outcomes in the literature. Thus, as a scientific
orthopedic community, we should question ourselves: how far have we gotten from the first
attempt at orthobiologics classification? or has the panorama changed since then?

66 **CONSIDERATION**

67 Historical perspective

68 In 2009, Dohan Eherenfest et al. [9] faced a scenario where commercial interests were obscuring 69 real clinical benefits, a "jungle" in their own words, developing a plethora of preparation methods, 70 systems, and centrifuges, and multiple platelet-derived products were covered by the umbrella term 71 PRP which did not allow a distinction between them. However, the authors were able to identify 72 common factors among the available products, such as: (1) the use of anticoagulant and immediate 73 centrifugation of the blood after its collection, (2) most preparation techniques allowed platelet 74 concentrate preparation within an hour, (3) the centrifugation aimed to separate the blood in layers 75 that would allow the extraction of specific fractions, and (4) the product was activated with 76 thrombin or calcium chloride. The situation led his team to propose a classification to provide an 77 objective approach to the growth and advance of PRP therapy [9].

78 Back then, Dohan Ehrenfest and colleagues were implementing Choukroun's leukocyte- and 79 platelet-rich fibrin (L-PRF) protocol in oral and maxillofacial surgery (Box 1 and 2) [10]. The 80 technique's benefits included a high efficiency in retrieving and concentrating platelets and 81 leukocytes and its semisolid and three-dimensional fibrin matrix structure mimicking a natural blood 82 clot. However, it was technically demanding because its success depended on rapid blood collection 83 and centrifugation. After all, the lack of anticoagulant deemed almost instant coagulation of the 84 blood once in contact with the walls of the dry-glass tubes. Otherwise, the fibrin would polymerize 85 diffusely in the tube, failing to concentrate most of the available platelets.

86 Understanding the rise of the idea – The clinical implication

The proposed platelet concentrates classification of Dohan Eherenfest and colleagues included three main parameters (Table 1), allowing the characterization of platelet concentrates in pure platelet-rich plasma (P-PRP), leukocyte- and platelet-rich plasma (L-PRP), pure platelet-rich fibrin (P-PRF), and L-PRF [9].

91 This classification brought new insights into the relevance of other components of platelet 92 concentrates, the leukocytes, and fibrin. By then, anti-microbial and immunomodulatory properties 93 were already attributed to the leukocytes in platelet concentrates and their role in angiogenesis 94 with vascular endothelial growth factor synthesis [11-14]. The role of L-PRP in tendon healing was 95 starting its rise [15]. Similarly, they supported the theory that the fibrin matrix and its composition 96 (including cytokines) were crucial for the platelet concentrates clinical efficacy. In fact, the clotting 97 pattern they were inducing in Choukron's L-PRF technique enhanced platelet growth factor release. The author confirmed his claims with the publication of another article in 2012 [16], where his team 98 99 proved not only an increased release of growth factors but also a more extended release period 100 (seven days) due to the naturally formed dense fibrin network in contrast to the light fibrin network present in P-PRP, observed when artificial activation was triggered with bovine thrombin, calcium 101 102 chloride or other clotting agents [17-19].

103 Scientific and societal impact

In a recent publication by the European Society for Sports Traumatology, Knee Surgery, and Arthroscopy Orthobiologic Initiative, the paper by Dohan Ehrenfest et al. [9] was listed fourth among the most cited PRP articles in regenerative medicine with more than 800 citations and including two additional papers from the author in the top 100 [6]. The current scientific research and clinical practice, driven by the main categorization between L-PRP and P-PRP (now, leukocyte-poor PRP),

109 confirms the impact of this article. Indeed, recent studies have proposed specific roles of different 110 leukocytes in PRP clinical efficiency depending on the healing stage and the type of injury [20]. 111 Lymphocytes, for example, have an anti-inflammatory role by steering monocyte differentiation 112 from M1 to the M2 subtype. On the other hand, neutrophils lead the so-called "regenerative 113 inflammation" by secreting chemokines to recruit macrophages and promoting an inflammatory 114 process desired to trigger the healing process [20, 21].

Before the classification system proposed by Dohan Ehrenfest et al. [9], the characterization of 115 116 platelet concentrates was confusing and contradictory, and created a methodological bias in many 117 publications. The authors addressed this challenging problem by simply proposing a classification 118 framework. Additionally, the classification opened the door to understanding intrinsic biological 119 mechanisms between the platelets, leukocytes, fibrin, and growth factors later considered for 120 studying the proliferation and differentiation of cells in different tissues affected by PRP. In other 121 words, the authors proposed evaluating these products as living tissues instead of pharmaceutical 122 preparations with a simple and precise composition.

123 Current evidence

124 After highlighting the importance of classifying the platelet concentrates according to the presence 125 of leukocytes, several clinical and laboratory studies conducted during the last decade have 126 demonstrated clinical benefit, especially in tendinopathies. In a controlled laboratory study, Lin et 127 al. [22] revealed a higher induction of platelet growth factors and tenocyte proliferation with L-PRP 128 preparations than P-PRP. Furthermore, a network meta-analysis of 18 randomized controlled trials 129 by Fitzpatrick et al. [3] showed that the most significant positive outcomes were obtained from a 130 single ultrasound-guided L-PRP injection in tendinopathies such as rotator cuff, tennis elbow, 131 patellar tendon, and Achilles tendon.

On the other hand, the presence of leukocytes in PRP has been found to be chondrotoxic, while in the absence of leukocytes, PRP promotes chondrogenesis [23, 24]. However, clinical studies on knee osteoarthritis have shown conflicting results when comparing both PRP preparations [24-26]. Although L-PRP has offered comparable results to leukocyte-poor preparations, the latter is preferred due to a higher risk of swelling from the increased inflammatory response [27].

137 Lessons learned

138 As with many biologic therapies in medicine, particularly musculoskeletal medicine, additional 139 research has often led to more questions than answers. Quite possibly the most important lesson 140 learned with biologic treatments, and PRP specifically, is that there is a vast range of variability not 141 only in the PRP preparation (instruments, devices, spin rate and time, activators, among others) but 142 also in the quality of the product due to inter- and intra-human variability. Since PRP is an 143 autologous product, the quality of the sample, growth factors concentration, and activity of the 144 components within the specimen are likely affected by the health status of the individual, 145 medications, diet, and cortisol stress levels, among others. Furthermore, the heterogeneity of 146 clinical outcomes among the studies has also moved the spotlight to the key elements that allow 147 patients to benefit from PRP therapy. Researchers have now classified patients into responders and 148 non-responders and started phenotyping the ideal patient [28-31].

149 New developments – New classifications and research originated from the original study

Since the initial classification of platelet concentrates, several other classification systems have been proposed and published in the current literature. Rossi et al. [32] recently reviewed the available classification systems for PRP, examining the advantages and limitations of each (Figure 1). Rossi and colleagues acknowledge the Dohan Ehrenfest classification system and its basic component breakdown. 155 DeLong et al. [33] presented their classification system in 2012, known as the platelet, activation, 156 white blood cell (PAW) classification. The PAW classification system was based on the absolute 157 number of platelets (P1 to P4, depending on the number of platelets), the method of platelet 158 activation, and the presence (α) or absence (β) of white blood cells. Mishra et al. [34] presented a 159 similar classification system. However, they classified the variables differently, which resulted in 160 four separate categories of PRP: elevated platelets and leukocytes without an external activator, 161 elevated platelets and leukocytes with an external activator, elevated platelets without leukocytes 162 and no external activator, and elevated platelets without leukocytes but with an external activator.

163 The classification systems continued to evolve as Mautner et al. [35] noted the importance of red 164 blood cells and their potential detrimental effects in PRP; thus, they added red blood cell analysis to 165 the classification known as platelet, leukocyte, red blood cells, and activation (PLRA). Magalon et al. 166 [36] then proposed the dose of platelet, efficiency, purity, and activation (DEPA) classification. In 167 this classification system, the proportion of the platelets recovered from PRP and the purity of the 168 PRP sample was included as essential qualities of the sample preparation. Lana et al. [37] proposed 169 a classification in 2017 evaluating method, activation, red blood cells, spin, platelets, image 170 guidance, leukocytes, and light activation (MARSPILL). Finally, The Platelet Physiology 171 Subcommittee of the Scientific and Standardization Committee of the International Society on 172 Thrombosis and Haemostasis has recently proposed a classification system that includes the 173 presence of leukocytes, red blood cells, activation product, platelet concentration, and preparation 174 category [38]. Individually, these classification systems have identified important aspects of PRP that 175 affect the biological composition and, ultimately, the indications and outcomes. In any situation, 176 simple and elegant classification systems are often preferred due to their ease of use; however, the 177 bulkier classification systems are typically more comprehensive. Based on the current indications of 178 PRP, the presence or absence of leukocytes seems to be the main dividing point. Still, the complexity

of the makeup of PRP makes a simple and elegant classification system quite challenging to develop.
While there is no perfect classification system to rely entirely on when evaluating biologics, including
PRP, the field strives to characterize this biological product better. Until we determine exactly what
elements of PRP are most important in affecting outcomes, we may be stuck relying on bulky
systems or even combining multiple classification systems.

184 Future directions

185 Much of the challenge in practicing evidence-based medicine in the case of biological treatments is 186 the lack of standardization in sample preparation, the heterogeneity in cohorts, and the incomplete 187 reporting of sample preparation utilized. Thus, there has been a call for minimum reporting 188 standards for studies involving biologics in musculoskeletal care [39-42]. Specifically, Murray et al. [41] reported the 23-item checklist compiled by the PRP working group to report minimum 189 190 information for studies evaluating biologics in orthopaedics (MIBO). In an assessment of the 50 most 191 cited articles related to PRP in musculoskeletal medicine, Bugarin et al. [43] reported a high level of 192 evidence in approximately 50% of the studies. Still, most of the studies were of only fair 193 methodological quality. Systematic reviews by Chahla et al. [44], DeClercq et al. [45], and Marín 194 Fermín et al. [5] have revealed that less than 10% of the studies provided a clear description of the 195 implemented PRP preparation protocol, which significantly limits the study's reproducibility. A 196 paucity of accurate reporting of a highly variable product has led to a lack of clarity and continues 197 challenging researchers and clinicians. In this sense, journal editors can play an essential role in 198 evidence quality improvement by requesting mandatory adherence to acceptable orthobiologics 199 reporting guidelines in submission and review [46]. The goal of the present decade is to build a new 200 body of evidence with high-quality reporting and reproducibility that will serve as the foundation of 201 its so-longed-for standardization.

202 ADDITIONAL EXPERT OPINION

203 Importantly, when approaching a novel treatment option, we must do our best to practice evidence-204 based medicine. This first requires research-driven processes to identify the crucial components of 205 the product so that it can be appropriately characterized and reliably recreated. In the case of 206 biologics such as PRP, where countless variables may ultimately impact the preparation and final 207 product, it becomes vital to identify the key elements and differences in the preparation method. 208 Classification systems can play a key role in driving this standardization process. As mentioned 209 above, the available classification systems have identified essential aspects of PRP that affect the 210 biological composition and, ultimately, the indications and outcomes. The simple and elegant 211 system proposed by Dohan Ehrenfest et al. [9] identified what seems to be the main dividing point 212 based on the current indications for PRP in musculoskeletal medicine: the presence or absence of leukocytes. While there is no perfect classification system to rely entirely on when evaluating PRP, 213 214 there is no question that the Dohan Ehrenfest classification system began the conversation, leading 215 to many more comprehensive classification systems.

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219 **REFERENCES**

- Obana KK, Schallmo MS, Hong IS, Ahmad CS, Moorman CT 3rd, Trofa DP, Saltzman BM.
 Current Trends in Orthobiologics: An 11-Year Review of the Orthopaedic Literature. Am J
 Sports Med. 2022 Sep;50(11):3121-3129. https://doi.org/10.1177/03635465211037343.
- Bansal H, Leon J, Pont JL, Wilson DA, Bansal A, Agarwal D, Preoteasa I. Platelet-rich plasma
 (PRP) in osteoarthritis (OA) knee: Correct dose critical for long term clinical efficacy. Sci Rep.
 2021 Feb 17;11(1):3971. https://doi.org/10.1038/s41598-021-83025-2.
- Fitzpatrick J, Bulsara M, Zheng MH. The Effectiveness of Platelet-Rich Plasma in the
 Treatment of Tendinopathy: A Meta-analysis of Randomized Controlled Clinical Trials. Am J
 Sports Med. 2017 Jan;45(1):226-233. https://doi.org/10.1177/0363546516643716.
- Makaram NS, Murray IR, Rodeo SA, Sherman SL, Murray AD, Haddad FS, McAdams TR,
 Abrams GD. The use of biologics in professional and Olympic sport: a scoping review
 protocol. Bone Jt Open. 2020 Nov 17;1(11):715-719. https://doi.org/10.1302/2633 1462.111.BJO-2020-0159.
- Marín Fermín T, Papakostas E, Macchiarola L, Zampeli F, Kalifis G, De Girolamo L, Zikria BA,
 Khoury M, D'Hooghe P. Injectable orthobiologics in professional football (Soccer) players: A
 systematic review. The Journal of Cartilage & Joint Preservation. 2022;2(2):100050.
 https://doi.org/10.1016/j.jcjp.2022.100050
- Coulange Zavarro A, De Girolamo L, Laver L, Sánchez M, Tischer T, Filardo G, Sabatier F,
 Magalon J. The Top 100 Most Cited Articles on Platelet-Rich Plasma Use in Regenerative
 Medicine-A Bibliometric Analysis-From the ESSKA Orthobiologic Initiative. Bioengineering
 (Basel). 2022 Oct 19;9(10):580. https://doi.org/10.3390/bioengineering9100580.

 Frank RM, Sherman SL, Chahla J, Dragoo JL, Mandelbaum B; Members of the Biologic Association, Anz AW, Bradley JP, Chu CR, Cole BJ, Farr J, Flanigan DC, Gomoll AH, Halbrecht J, Horsch K, Lattermann C, Leucht P, Maloney WJ, McIntyre LF, Murray I, Muschler GF, Nakamura N, Piuzzi NS, Rodeo SA, Saris DBF, Shaffer WO, Shapiro SA, Spindler KP, Steinwachs M, Tokish JM, Vangsness CT, Watson JT, Yanke AB, Zaslav KR. Biologic Association Annual Summit: 2020 Report. Orthop J Sports Med. 2021 Jun 7;9(6):23259671211015667. https://doi.org/10.1177/23259671211015667.

- Magalon J, Brandin T, Francois P, Degioanni C, De Maria L, Grimaud F, Veran J, Dignat George F, Sabatier F. Technical and biological review of authorized medical devices for
 platelets-rich plasma preparation in the field of regenerative medicine. Platelets. 2021 Feb
 17;32(2):200-208. https://doi.org/10.1080/09537104.2020.1832653.
- Dohan Ehrenfest DM, Rasmusson L, Albrektsson T. Classification of platelet concentrates:
 from pure platelet-rich plasma (P-PRP) to leucocyte- and platelet-rich fibrin (L-PRF). Trends
 Biotechnol. 2009 Mar;27(3):158-67. https://doi.org/10.1016/j.tibtech.2008.11.009.
- 255 10. Dohan DM, Choukroun J, Diss A, Dohan SL, Dohan AJ, Mouhyi J, Gogly B. Platelet-rich fibrin
 256 (PRF): a second-generation platelet concentrate. Part I: technological concepts and
 257 evolution. Oral Surg Oral Med Oral Pathol Oral Radiol Endod. 2006 Mar;101(3):e37-44.
 258 https://doi.org/10.1016/j.tripleo.2005.07.008.
- 259 11. Cieslik-Bielecka A, Gazdzik TS, Bielecki TM, Cieslik T. Why the platelet-rich gel has anti 260 microbial activity? Oral Surg Oral Med Oral Pathol Oral Radiol Endod. 2007 Mar;103(3):303 261 5; author reply 305-6. https://doi.org/10.1016/j.tripleo.2006.08.034.

- Moojen DJ, Everts PA, Schure RM, Overdevest EP, van Zundert A, Knape JT, Castelein RM,
 Creemers LB, Dhert WJ. Anti-microbial activity of platelet-leukocyte gel against
 Staphylococcus aureus. J Orthop Res. 2008 Mar;26(3):404-10.
- 265 13. El-Sharkawy H, Kantarci A, Deady J, Hasturk H, Liu H, Alshahat M, Van Dyke TE. Platelet-rich
 266 plasma: growth factors and pro- and anti-inflammatory properties. J Periodontol. 2007
 267 Apr;78(4):661-9. https://doi.org/10.1902/jop.2007.060302.
- Werther K, Christensen IJ, Nielsen HJ. Determination of vascular endothelial growth factor
 (VEGF) in circulating blood: significance of VEGF in various leucocytes and platelets. Scand J
 Clin Lab Invest. 2002;62(5):343-50. https://doi.org/10.1080/00365510260296492.
- 271 15. Schnabel LV, Mohammed HO, Miller BJ, McDermott WG, Jacobson MS, Santangelo KS,
 272 Fortier LA. Platelet rich plasma (PRP) enhances anabolic gene expression patterns in flexor
 273 digitorum superficialis tendons. J Orthop Res. 2007 Feb;25(2):230-40.
 274 https://doi.org/10.1002/jor.20278.
- 275 16. Dohan Ehrenfest DM, Bielecki T, Jimbo R, Barbé G, Del Corso M, Inchingolo F, Sammartino
 276 G. Do the fibrin architecture and leukocyte content influence the growth factor release of
 277 platelet concentrates? An evidence-based answer comparing a pure platelet-rich plasma (P278 PRP) gel and a leukocyte- and platelet-rich fibrin (L-PRF). Curr Pharm Biotechnol. 2012
 279 Jun;13(7):1145-52. https://doi.org/10.2174/138920112800624382.
- 280 17. Dohan Ehrenfest DM, Del Corso M, Inchingolo F, Sammartino G, Charrier JB. Platelet-rich
 281 plasma (PRP) and platelet-rich fibrin (PRF) in human cell cultures: growth factor release and
 282 contradictory results. Oral Surg Oral Med Oral Pathol Oral Radiol Endod. 2010
 283 Oct;110(4):418-21; author reply 421-2. https://doi.org/10.1016/j.tripleo.2010.05.059.

284	18. Fern	ández-l	Barbero JE,	Galin	do-Mor	eno P, Avila-Ortiz G	, Caba O, Sánchez-Fernández E,
285	Wan	g HL. F	low cytome	etric an	id morp	hological characteriz	ation of platelet-rich plasma gel.
286	Clin	Oral	Implants	Res.	2006	Dec;17(6):687-93.	https://doi.org/10.1111/j.1600-
287	0501	2006.0	01179.x.				

- Mazzucco L, Balbo V, Cattana E, Guaschino R, Borzini P. Not every PRP-gel is born equal.
 Evaluation of growth factor availability for tissues through four PRP-gel preparations:
 Fibrinet, RegenPRP-Kit, Plateltex and one manual procedure. Vox Sang. 2009 Aug;97(2):1108. https://doi.org/10.1111/j.1423-0410.2009.01188.x.
- 20. Lana JF, Huber SC, Purita J, Tambeli CH, Santos GS, Paulus C, Annichino-Bizzacchi JM.
 Leukocyte-rich PRP versus leukocyte-poor PRP The role of monocyte/macrophage function
 in the healing cascade. J Clin Orthop Trauma. 2019 Oct;10(Suppl 1):S7-S12.
 https://doi.org/10.1016/j.jcot.2019.05.008.
- 296 21. Marathe A, Patel SJ, Song B, Sliepka JM, Shybut TS, Lee BH, Jayaram P. Double-Spin
 297 Leukocyte-Rich Platelet-Rich Plasma Is Predominantly Lymphocyte Rich With Notable
 298 Concentrations of Other White Blood Cell Subtypes. Arthrosc Sports Med Rehabil. 2021 Dec
 299 10;4(2):e335-e341. https://doi.org/10.1016/j.asmr.2021.10.004.
- Lin KY, Chen P, Chen AC, Chan YS, Lei KF, Chiu CH. Leukocyte-Rich Platelet-Rich Plasma Has
 Better Stimulating Effects on Tenocyte Proliferation Compared With Leukocyte-Poor
 Platelet-Rich Plasma. Orthop J Sports Med. 2022 Mar 15;10(3):23259671221084706.
 https://doi.org/10.1177/23259671221084706.
- Xu Z, Yin W, Zhang Y, Qi X, Chen Y, Xie X, Zhang C. Comparative evaluation of leukocyte- and
 platelet-rich plasma and pure platelet-rich plasma for cartilage regeneration. Sci Rep. 2017
 Mar 7;7:43301. https://doi.org/10.1038/srep43301.

307	24. Riboh JC, Saltzman BM, Yanke AB, Fortier L, Cole BJ. Effect of Leukocyte Concentration on
308	the Efficacy of Platelet-Rich Plasma in the Treatment of Knee Osteoarthritis. Am J Sports
309	Med. 2016 Mar;44(3):792-800. https://doi.org/10.1177/0363546515580787.

- 25. Di Martino A, Boffa A, Andriolo L, Romandini I, Altamura SA, Cenacchi A, Roverini V,
 Zaffagnini S, Filardo G. Leukocyte-Rich versus Leukocyte-Poor Platelet-Rich Plasma for the
 Treatment of Knee Osteoarthritis: A Double-Blind Randomized Trial. Am J Sports Med. 2022
 Mar;50(3):609-617. https://doi.org/10.1177/03635465211064303.
- 26. Abbas A, Du JT, Dhotar HS. The Effect of Leukocyte Concentration on Platelet-Rich Plasma
 Injections for Knee Osteoarthritis: A Network Meta-Analysis. J Bone Joint Surg Am. 2022
 Mar 16;104(6):559-570. https://doi.org/10.2106/JBJS.20.02258.
- 27. Kim JH, Park YB, Ha CW, Roh YJ, Park JG. Adverse Reactions and Clinical Outcomes for
 Leukocyte-Poor Versus Leukocyte-Rich Platelet-Rich Plasma in Knee Osteoarthritis: A
 Systematic Review and Meta-analysis. Orthop J Sports Med. 2021 Jun
 30;9(6):23259671211011948. https://doi.org/10.1177/23259671211011948.
- 28. Mantripragada VP, Csorba A, Bova W, Boehm C, Piuzzi NS, Bullen J, Midura RJ, Muschler GF.
 Assessment of Clinical, Tissue, and Cell-Level Metrics Identify Four Biologically Distinct Knee
 Osteoarthritis Patient Phenotypes. Cartilage. 2022 Jan-Mar;13(1):19476035221074003.
 https://doi.org/10.1177/19476035221074003.
- 29. Ganguly P, Fiz N, Beitia M, Owston HE, Delgado D, Jones E, Sánchez M. Effect of Combined
 Intraosseous and Intraarticular Infiltrations of Autologous Platelet-Rich Plasma on
 Subchondral Bone Marrow Mesenchymal Stromal Cells from Patients with Hip
 Osteoarthritis. J Clin Med. 2022 Jul 4;11(13):3891. https://doi.org/10.3390/jcm11133891.

329	30. Zahir H, Dehghani B, Yuan X, Chinenov Y, Kim C, Burge A, Bandhari R, Nemirov D, Fava P,							
330	Moley P, Potter H, Nguyen J, Halpern B, Donlin L, Ivashkiv L, Rodeo S, Otero M. In vitro							
331	responses to platelet-rich-plasma are associated with variable clinical outcomes in patients							
332	with knee osteoarthritis. Sci Rep. 2021 Jun 1;11(1):11493. https://doi.org/10.1038/s41598-							
333	021-90174-x.							
334	31. Saita Y, Kobayashi Y, Nishio H, Wakayama T, Fukusato S, Uchino S, Momoi Y, Ikeda H, Kaneko							
335	K. Predictors of Effectiveness of Platelet-Rich Plasma Therapy for Knee Osteoarthritis: A							
336	Retrospective Cohort Study. J Clin Med. 2021 Sep 29;10(19):4514.							
337	https://doi.org/10.3390/jcm10194514.							
338	32. Rossi LA, Murray IR, Chu CR, Muschler GF, Rodeo SA, Piuzzi NS. Classification systems for							
339	platelet-rich plasma. Bone Joint J. 2019 Aug;101-B(8):891-896.							
340	https://doi.org/10.1302/0301-620X.101B8.BJJ-2019-0037.R1.							
341	33. DeLong JM, Russell RP, Mazzocca AD. Platelet-rich plasma: the PAW classification system.							
342	Arthroscopy. 2012 Jul;28(7):998-1009. https://doi.org/10.1016/j.arthro.2012.04.148.							
343	34. Mishra A, Harmon K, Woodall J, Vieira A. Sports medicine applications of platelet rich							
344	plasma. Curr Pharm Biotechnol. 2012 Jun;13(7):1185-95.							
345	https://doi.org/10.2174/138920112800624283.							
346	35. Mautner K, Malanga GA, Smith J, Shiple B, Ibrahim V, Sampson S, Bowen JE. A call for a							
347	standard classification system for future biologic research: the rationale for new PRP							
348	nomenclature. PM R. 2015 Apr;7(4 Suppl):S53-S59.							
349	https://doi.org/10.1016/j.pmrj.2015.02.005.							

- 36. Magalon J, Chateau AL, Bertrand B, Louis ML, Silvestre A, Giraudo L, Veran J, Sabatier F.
 DEPA classification: a proposal for standardising PRP use and a retrospective application of
 available devices. BMJ Open Sport Exerc Med. 2016 Feb 4;2(1):e000060.
 https://doi.org/10.1136/bmjsem-2015-000060.
- 37. Lana JFSD, Purita J, Paulus C, Huber SC, Rodrigues BL, Rodrigues AA, Santana MH, Madureira
 JL Jr, Malheiros Luzo ÂC, Belangero WD, Annichino-Bizzacchi JM. Contributions for
 classification of platelet rich plasma proposal of a new classification: MARSPILL. Regen
 Med. 2017 Jul;12(5):565-574. https://doi.org/10.2217/rme-2017-0042.
- 358 38. Harrison P; Subcommittee on Platelet Physiology. The use of platelets in regenerative
 medicine and proposal for a new classification system: guidance from the SSC of the ISTH. J
 Thromb Haemost. 2018 Sep;16(9):1895-1900. https://doi.org/10.1111/jth.14223.
- 361 39. Rodeo S. The Need for Minimum Reporting Standards for Studies of "Biologics" in Sports
 362 Medicine. Am J Sports Med. 2019 Sep;47(11):2531-2532.
 363 https://doi.org/10.1177/0363546519872219.
- 40. LaPrade RF, Graden NR, Kahat DH. Editorial Commentary: Platelet-Rich Plasma: The Devil Is
 in the Details, and the Details Need to Be Better Reported. Arthroscopy. 2019
 Nov;35(11):3114-3116. https://doi.org/10.1016/j.arthro.2019.08.016.
- Murray IR, Geeslin AG, Goudie EB, Petrigliano FA, LaPrade RF. Minimum Information for
 Studies Evaluating Biologics in Orthopaedics (MIBO): Platelet-Rich Plasma and
 Mesenchymal Stem Cells. J Bone Joint Surg Am. 2017 May 17;99(10):809-819.
 https://doi.org/10.2106/JBJS.16.00793.

- 42. Murray IR, Murray AD, Geeslin AG, Goudie EB, White TO, Petrigliano FA, LaPrade RF.
 Infographic: we need minimum reporting standards for biologics. Br J Sports Med. 2019
 Aug;53(15):974-975. https://doi.org/10.1136/bjsports-2017-098122.
- 43. Bugarin A, Schroeder G, Shi BY, Jones KJ, Kremen TJ Jr. Assessment of Characteristics and
 Methodological Quality of the Top 50 Most Cited Articles on Platelet-Rich Plasma in
 Musculoskeletal Medicine. Orthop J Sports Med. 2022 May 26;10(5):23259671221093074.
 https://doi.org/10.1177/23259671221093074.
- 44. Chahla J, Cinque ME, Piuzzi NS, Mannava S, Geeslin AG, Murray IR, Dornan GJ, Muschler GF,
 LaPrade RF. A Call for Standardization in Platelet-Rich Plasma Preparation Protocols and
 Composition Reporting: A Systematic Review of the Clinical Orthopaedic Literature. J Bone
 Joint Surg Am. 2017 Oct 18;99(20):1769-1779. https://doi.org/10.2106/JBJS.16.01374.
- 45. DeClercq MG, Fiorentino AM, Lengel HA, Ruzbarsky JJ, Robinson SK, Oberlohr VT, Whitney
 KE, Millett PJ, Huard J. Systematic Review of Platelet-Rich Plasma for Rotator Cuff Repair:
 Are We Adhering to the Minimum Information for Studies Evaluating Biologics in
 Orthopaedics? Orthop J Sports Med. 2021 Dec 7;9(12):23259671211041971.
 https://doi.org/10.1177/23259671211041971.
- Marín Fermín T, Scarlat MM, Laupheimer MW. Would you have an injection without
 knowing its formula? New challenges in platelet-rich plasma therapy. Int Orthop. 2022
 Oct;46(10):2179-2180. https://doi.org/10.1007/s00264-022-05566-z.

TABLES

PAR	AMETERS	SUBPA	SUBPARAMETER			
		1.	Size of the centrifuge			
		2.	Duration of the procedure			
Α	Preparation kits and	3.	Cost of the device and kits			
	centrifuges	4.	Ergonomics of the kit and the complexity of th			
			procedure			
		1.	Final volume of usable concentrate			
в	Content of the concentrate	2.	Efficiency in collecting platelets			
Б	content of the concentrate	3.	Leucocytes			
		4.	Preservation of the components			
с	Fibrin network	1.	Concentration and density			
	FIDITI NELWORK	2.	Polymerization process			

391 Table 1. Dohan Eherenfest et al. (2009) platelet concentrate classification.

393 BOXES

BOX 1. Choukroun's L-PRF protocol [10]

This novel method was characterized by its simplicity, reproducibility, and low cost. In this protocol, developed in Nice (France), venous blood was collected in 10 ml tubes and instantly centrifuged without anticoagulant. The lack of anticoagulant allowed the natural formation of a clot (platelet activation) that would facilitate the manipulation of L-PRF, avoiding the implementation of any additive. Thus, after centrifugation at 3000 rpm (400g) for 10 minutes, three distinct layers are visualized: the red blood cell layer in the bottom of the tube, a top acellular plasma layer, and a L-PRF clot in between, containing most of the platelets.

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BOX 2. Short interview with Pr. Lars Rasmusson – Co-author of the classic paper on "Classification of platelet concentrates: from pure platelet-rich plasma (P-PRP) to leucocyteand platelet-rich fibrin (L-PRF)"

Head of Department, Department of Maxillofacial Surgery, Sahlgrenska Academy, University of Gothenburg

Q1: What motivated the development of the classification system?

A-LR: The motivation to develop a classification system was the somewhat conflicting PRP and PRF handling methodology at the time.

Q2: Who were the researchers involved in the development of the classification and their contribution to it?

A-LR: It was Pr. Tomas Albrektsson, Head of the Department of Biomaterials, University of Gothenburg, and I at the time together with our postdoc, David Dohan Ehrenfest.

Q3: What were the clinical uses of PRP in your Institution?

A-LR: PRP and PRF were used both clinically in maxillofacial reconstruction and in experimental work at our lab. In clinical practice here at our unit, L-PRF has replaced PRP since it is easier to use and possible to manufacture in different consistency/preparations, for example, injectable and as membranes.

Q4: What are the current challenges of PRP therapy?

A-LR: The challenge has been (and still is) to prove long-term efficacy and superiority in bone healing.

Q5: What is the future of PRP therapy?

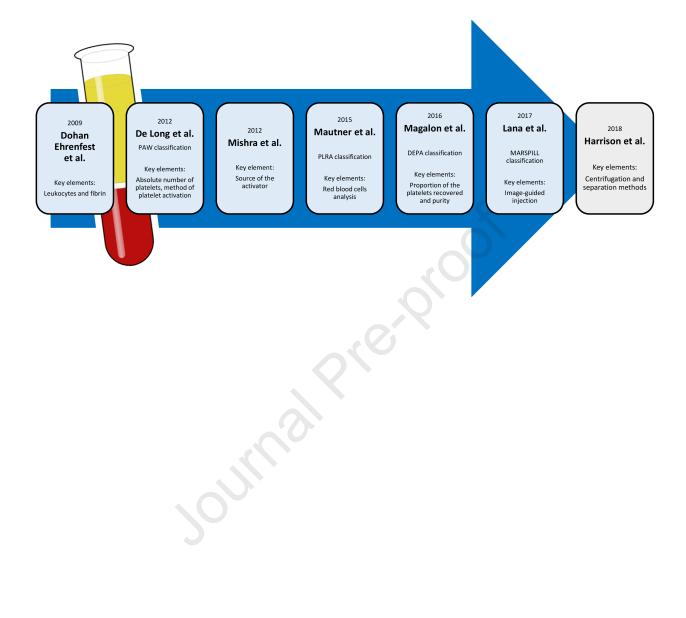
A-LR: I strongly believe that more indications for use will be discovered and evaluated in cartilage repair via injectable platelets.

BOX 3. Summary of The Classic
PRP is currently used to treat multiple musculoskeletal conditions, including knee osteoarthritis,
cartilage injuries, patellar tendonitis, and tennis elbow. In recent decades, the media, celebrity
athletes, the desire for novel treatments, and its autologous nature have boosted its use in
multiple sports-related injuries. As a result, its high demand has led to an industry-driven
development of various platelet concentrate systems and products exceeding the pace of
evidence-based practice.
In 2009, Dohan Eherenfest et al. proposed the first attempt at PRP classification to categorize the
platelet concentrates concerning their fibrin and leukocyte content and the degree of
standardization of the procedure, providing an overview of the available systems. The authors
proposed four categories of platelet concentrates (P-PRP, L-PRP, P-PRF, and L-PRF) to group a
"jungle" of products in which the term PRP was used indistinctly.
The classification opened the door to understanding intrinsic biological mechanisms between the
PRP components. Since the initial classification, several other classification systems have been
proposed and published in the current literature, identifying important aspects of PRP. To date,
there is still a lack of standardization in sample preparation, cohort heterogeneity, and
incomplete reporting of sample preparation utilized, leading to a lack of clarity and challenging
researchers and clinicians. A paucity of accurate reporting of a highly variable product has led to
a lack of clarity and continues challenging researchers and clinicians. In this sense, journal editors
can play an essential role in evidence quality improvement by requesting mandatory adherence
to acceptable orthobiologics reporting guidelines in submission and review. The goal of the
present decade is to build a new body of evidence with high-quality reporting and reproducibility
that will serve as the foundation of its so-longed-for standardization.

397 FIGURE LEGENDS

398 Figure 1. Historical landmarks of platelet-rich plasma classifications.

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Declaration of interests

☑ The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

□ The authors declare the following financial interests/personal relationships which may be considered as potential competing interests:

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