

Contents lists available at ScienceDirect

Comptes Rendus Biologies

www.sciencedirect.com



Evolution/Évolution

Evolutionary history of aphid-plant associations and their role in aphid diversification

Histoire évolutive des associations plantes-pucerons et leur rôle dans la diversification des pucerons

Jean Peccoud ^{a,*}, Jean-Christophe Simon ^b, Carol von Dohlen ^c, Armelle Coeur d'acier ^d, Manuel Plantegenest ^b, Flavie Vanlerberghe-Masutti ^d, Emmanuelle Jousselin ^d

ARTICLE INFO

Article history:

Available online 13 May 2010

Keywords:
Aphididae
Speciation
Host-plant association
Ecological speciation
Phylogenetic reconstruction

ABSTRACT

Aphids are intimately linked with their host plants that constitute their only food resource and habitat, and thus impose considerable selective pressure on their evolution. It is therefore commonly assumed that host plants have greatly influenced the diversification of aphids. Here, we review what is known about the role of host plant association on aphid speciation by examining both macroevolutionary and population-level studies. Phylogenetic studies conducted at different taxonomic levels show that, as in many phytophagous insect groups, the radiation of angiosperms has probably favoured the major Tertiary diversification of aphids. These studies also highlight many aphid lineages constrained to sets of related host plants, suggesting strong evolutionary commitment in aphids' host plant choice, but they fail to document cospeciation events between aphid and host lineages. Instead, phylogenies of several aphid genera reveal that divergence events are often accompanied by host shifts, and suggest, without constituting a formal demonstration, that aphid speciation could be a consequence of adaptation to new hosts. Experimental and field studies below the species level support reproductive isolation between host races as partly due to divergent selection by their host plants. Selected traits are mainly feeding performances and life cycle adaptations to plant phenology. Combined with behavioural preference for favourable host species, these divergent adaptations can induce pre- and post-zygotic barriers between host-specialized aphid populations. However, the hypothesis of host-driven speciation is seldom tested formally and must be weighed against overlooked explanations involving geographic isolation and nonecological reproductive barriers in the process of speciation.

 $\ensuremath{\texttt{©}}$ 2010 Académie des sciences. Published by Elsevier Masson SAS. All rights reserved.

RÉSUMÉ

Mots clés : Aphididé Spéciation Association plante hôte

Les pucerons entretiennent des relations intimes avec leurs plantes hôtes qui constituent leur seule nourriture et leur seul habitat, et qui ainsi imposent des contraintes sélectives considérables sur leur évolution. On pense donc que les plantes hôtes ont grandement influencé la diversification des pucerons. Ici nous synthétisons les connaissances

E-mail addresses: jean.peccoud@supagro.inra.fr (J. Peccoud), jousseli@supagro.inra.fr (E. Jousselin).

^a INRA, UMR BGPI, Campus international de Baillarguet, TA A-54/K, 34398 Montpellier cedex 5, France

^b INRA, UMR BiO3P, agrocampus Ouest, université Rennes 1, domaine de la Motte, 35653 le Rheu cedex, France

^c Department of Biology, Utah State University, 5305 Old Main Hill, Logan, Utah 84322, USA

d INRA, UMR CBGP, IRD, Cirad, Montpellier SupAgro, Campus international de Baillarguet, CS 30016, 34988 Montferrier-sur-Lez cedex, France

^{*} Corresponding author.

Spéciation écologique Reconstruction phylétique concernant le rôle des associations avec les plantes hôtes sur la spéciation des pucerons. en examinant des études menées au niveau macro-évolutif et au sein des populations. Les études phylogénétiques conduites à des niveaux taxonomiques variés montrent que, à l'instar de nombreux groupes d'insectes phytophages, la radiation des angiospermes a probablement favorisé la diversification majeure des pucerons au tertiaire. Ces études révèlent aussi l'existence de nombreuses lignées de pucerons strictement associées à des groupes de plantes apparentées et suggèrent de fortes contraintes évolutives sur le choix des plantes par les pucerons. Ces études ne mettent cependant pas en évidence des événements de co-spéciation entre lignées de pucerons et de plantes. Les phylogénies de plusieurs genres de pucerons montrent que la plupart des événements de divergence sont accompagnés de changements d'hôtes, et suggèrent donc, sans en constituer une démonstration formelle, que la spéciation chez les pucerons peut être une conséquence de l'adaptation à de nouveaux hôtes. Les études intraspécifiques expérimentales et de terrain montrent que l'isolement reproducteur entre races d'hôtes résulte en partie de sélection divergente imposée par les plantes hôtes. Les traits soumis à la sélection sont principalement la performance alimentaire et les adaptations du cycle de vie à la phénologie des plantes. Ces adaptations divergentes, combinées à la préférence comportementale pour les plantes favorables, induisent des barrières pré- et postzygotiques entre populations de pucerons spécialisées. Cependant, l'hypothèse de spéciation induite par les plantes hôtes est rarement testée formellement et mérite d'être opposée à des explications alternatives impliquant l'isolement géographique et des barrières non écologiques dans le processus de spéciation.

© 2010 Académie des sciences. Publié par Elsevier Masson SAS. Tous droits réservés.

1. Introduction

Most insect herbivores have very intimate associations with their host plants. Debates continue over the extent to which the diversification of phytophagous insects is driven by their association with plants [1]. Several hypothesized pathways of diversification recognize host plants as playing a major role in the process of speciation; two of these are:

- (i) cospeciation–herbivores diversify contemporaneously with their host plants, resulting in parallel phylogenetic trees between insects and their hosts [2,3];
- (ii) speciation through host shifts—a group of individuals of an insect species colonizes a new existing plant species and diverges from its population of origin [4].

Both processes rely on some degree of host plant specialization and suggest that host plant diversity, if not necessarily the sole cause of reproductive isolation and speciation, is a major driver of insect diversification [5].

There are approximately 4700 described aphid species (Aphididae, *sensu* Remaudière and Remaudière [6]). Forty percent of aphid species live wholly or partly on trees and 55% live on herbaceous plants or shrubs (the remaining 5% live on unknown hosts) [7]. Although the host breadth of aphids is variable, aphids are recognized as being intimately linked with their host plants [8]. This statement stems from several well-documented observations. First, aphids and their host plants are not haphazardly associated. Some plant families are far overrepresented among aphid hosts, such as Asteraceae (>600 aphid species), Apiaceae (~600), Rosaceae (~300) and Coniferae (~350) [7]. Conversely, some large plant families (e.g. Cactaceae, Euphorbiaceae, Orchidaceae, Lauraceae, Moraceae, Myrtaceae, [9]) host very few aphid species. Second, among all

phytophagous insects, aphids are generally recognized as being near the end of the continuum from generalists to specialists. Specialization occurs at various taxonomic levels. Some aphid species are specific to a single plant species. At a higher taxonomic level, some aphid genera/ families are strictly associated with a single plant genus/ family. However, several highly polyphagous aphid species are also known (less than 1% of all species), including some of the most injurious agricultural pests. For instance, Aphis gossypii Glover can feed on 912 species plants belonging to 116 families [10]. However, apparently polyphagous species frequently encompass host races or host-specialized populations with a narrow host range [11-15]. The detection of several differentiated host races within apparently polyphagous species suggest an ongoing process of speciation by adaptation to distinct host plants in several aphid groups.

Altogether, these patterns of host use prelude the idea that host-plant associations have influenced aphid diversification. Here, we evaluate this hypothesis by documenting the history of host-plant association and specialization through phylogenetic reconstructions, and by reporting on how host plants can promote reproductive isolation in natural populations.

2. Macroevolutionary patterns of aphid-plant associations

2.1. Major evolutionary transitions in the history of aphidplant associations

Aphididae are divided into several subfamilies, tribes and subtribes (we use here the most recent comprehensive classification of [6]). Among subfamilies and tribes, few are strictly associated with gymnosperms, some use herbaceous monocotyledons and most use various angiosperm

families. Few molecular phylogenies have investigated the origin of these tribes and their relationships; we have attempted here to summarize these studies and what they tell us about the evolution of host plant association (Fig. 1).

Phylogenetic reconstruction places the coniferous feeding family of Adelgidae as a sister group of Aphididae [16]. Several lineages of Aphididae also use coniferous hosts, and these may represent ancient associations [17]; one such lineage is the subfamily Neophyllaphidinae, which has a Gondwanan distribution and is associated with two gymnosperm genera, *Podocarpus* and *Araucaria*. It has been suggested that early Aphididae ancestors lived on coniferous hosts, a hypothesis consistent with fossil records [18,19]. For example, Neophyllaphidinae look very similar to the Cretaceous fossil *Aniferella*, which by inference were probably associated with Podocarpaceae or Araucariaceae [20]. These two gymnosperm families were likely ancestral aphid hosts in the Early Cretaceous before angiosperms arose, along with several other

abundant gymnosperm groups [21]. Fossils have allowed the calibration of molecular phylogenetic reconstructions: the age of the common ancestor of extant subfamilies in the Aphididae has been estimated at 84-99 mva [21]. which corresponds to the Late Cretaceous epoch and roughly the diversification of woody angiosperms. The lack of resolution of molecular phylogenetic relationships among Aphididae subfamilies and tribes and the short branches found in the deep nodes of the phylogenetic tree suggest a rapid radiation of aphids at this period [21,22]. A more recent study found tentative support for three main lineages, with Lachninae as sister group to all other aphids; as one of the few subfamilies with conifer feeders, the position of Lachninae also supports the hypothesis of an ancestral gymnosperm feeding in aphids [23] with the caveat that conifer feeding in lachnines is thought to be a derived state by some systematists [24,25]. That most of present-day aphid diversity likely arose after one or more shifts from gymnosperms to angiosperms strongly suggests that the radiation of aphids has been driven by the

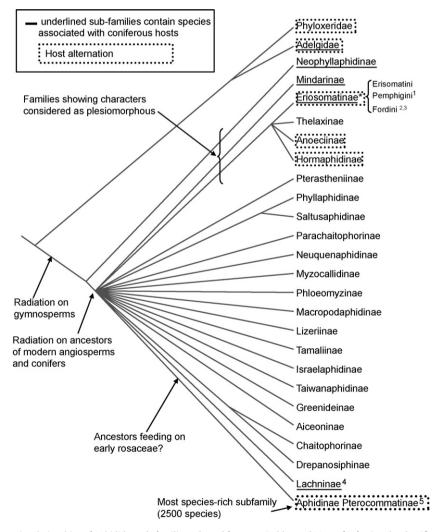


Fig. 1. Putative phylogenetic relationships of Aphididae sub-families, adapted from von Dohlen and Moran [21] using the classification of Remaudière and Remaudière [6] and results from references [22,23].* indicates groups in which monophyly can be contested based on molecular phylogenies. References are indicated for subfamilies for which molecular phylogenies have been conducted: ¹[53] ²[52], ³[51] ⁴ [141], ⁵[21].

diversification of flowering plants [26]. Fewer than 30 extant species remain in two subfamilies thought to be primitively conifer feeding (Neophyllaphidinae and Mindarinae), compared to more than 4000 angiosperm-feeding species. These observations suggest that either aphids could have speciated in parallel with particular groups of angiosperms (i.e., have partly cospeciated) [27] or that their diversification has rapidly followed the radiation of angiosperm families, favoured by the diversity of new ecological niches offered by these plants.

Compared to other phytophagous insect groups of similar age, aphid biodiversity is rather low. Furthermore, aphids are relatively rare in the tropics, where plant diversity is the highest, and underrepresented in the southern hemisphere. The plant families cited by Eastop [24] and Dixon [9] as having no specific aphids are mostly diversified in tropical or hot regions and/or in the southern hemisphere. Thus, absence of diversification on certain plant families may have little to do with aphid feeding preferences. Several hypotheses might explain the paucity of aphids in the tropics and southern hemisphere. One of them suggests that aphids have been less successful in tropical regions due to the difficulty of finding their specific host plants in the highly diversified tropical ecosystems; aphids seem to locate their hosts somewhat randomly and may depend on high plant apparency to do so [9,28]. Another hypothesis suggests that cyclical parthenogenesis, the ancestral life cycle of all sexually reproducing species, is maladapted to the tropics [29]. Low diversity in the southern hemisphere compared to the northern one has also been explained in a historical biogeographic context, predicated on the idea that aphids originated in the northern hemisphere and underwent extensive species radiations there during the Tertiary Period; subsequently the tropics acted as a barrier to southward migrations [30]. Aphid geographical distribution and plant association is most certainly the result of both ecological and historical constraints. Comparative biogeographical studies of aphids and their host plants integrating advances in molecular phylogenetics are still quite rare [31,32], and more are needed to elucidate several of the major transitions in aphid diversification.

Host alternation, a characteristic trait of certain aphid life cycles, may have played an important role in favouring the colonization of new plants and speciation in aphids. About 10% of aphid species alternates seasonally, and mostly obligately, between two unrelated plant species [7,33,34]; these aphids are said to be host alternating or heteroecious, in contrast to the vast majority of species that complete their entire life cycle on the same host (known as non-host-alternating or monoecious species). The evolution of host alternation has occurred in several aphid families, in which the cycle follows a roughly similar pattern of host use: after egg hatching followed by parthenogenetic development of colonies on their primary/winter host (most generally a woody host), aphids alternate to secondary/summer hosts (most generally herbaceous plants) on which they reproduce parthenogenetically; later generations return to their primary host for sexual production of eggs and usually overwintering. The two host plants of host alternating species are almost never closely related, and some consist of rather surprising combinations. For instance, the Nearctic species *Melaphis rhois* (Fordini) and the related eastern-Asian species of *Nurudea*, *Kaburagia* and *Schlechtendalia* (Fordini) alternate between *Rhus* (Anacardiaceae) and mosses [35]. The exact number of evolutionary gains and/or losses of this life cycle throughout aphid history is not known, but almost certainly both gains and losses have occurred multiple times, and some proportion of extant aphid-host associations probably have been shaped by such transitions [35]. The evolution of host alternation has two intricately linked consequences:

- (i) the colonization of new ecological niches via a new life cycle:
- (ii) the broadening of host plant range.

Host-alternating species often have a much larger range of secondary hosts than primary hosts (see section 2.2 for possible causes). Hence, host alternation seems to allow for an escape from host plant specialization. This transition in aphid diet might be followed by a shift of all or part of the population to permanent existence on some of the new secondary hosts, and eventually leading to speciation. These sequences of evolutionary changes could have fuelled the diversification of certain aphid lineages [34].

Such a scenario entailing acquisition of secondary hosts and broadening of host range, followed by specialization and speciation on newly acquired hosts, might be exemplified by the diversification of Aphidinae. With about 57% of all described aphid species [24], Aphidinae constitutes the most species-rich subfamily. It encompasses many genera that live exclusively or partially on Rosaceae and are also associated with modern grasses. It also includes many highly polyphagous species among which most of the agricultural aphid pests are found. The scarcity of Eocene and Oligocene fossils suggests that this lineage probably emerged and diversified in the latter half of the Tertiary (Miocene and later), when plant communities dominated by grasses became abundant [19,26]. Molecular phylogenetic reconstruction implies several evolutionary transitions in life cycles in this subfamily [36]. Further, phylogenies suggest that several shifts to and from Rosaceae have occurred. One plausible evolutionary scenario is that ancestors of Aphidinae fed on a woody host, possibly an early Rosaceae species [27]. This lineage then radiated following the acquisition of host alternation to herbaceous hosts; in some cases descendent lineages lost their primary rosaceous hosts to become monoecious on more recently acquired secondary hosts (also called secondary monoecy) [8,19,36,37]. Several genera in Aphidinae exhibit all the life cycle states in this hypothetical process (primary monoecy, host alternation, and secondary monoecy), including Dysaphis, Cryptomyzus, Myzus, and Brachycaudus. In contrast, no other aphid lineages with host alternation (Eriosomatinae, Hormaphidinae, Anoeciinae) exhibit complete secondary monoecy including a sexual generation. Given the disparity in species diversity between Aphidinae and other lineages, it seems possible that acquisition followed by loss of heteroecy may have accelerated the diversification process in this lineage [34].

A true comparative analysis of the diversification rate is difficult to conduct based on the current phylogenetic reconstructions, but it would be interesting to test whether increased diversification rates are correlated with acquisition and loss of host alternation. Such comparative tests could be conducted at several taxonomic levels: among subfamilies, and among genera within Aphidinae. These analyses could yield insights into the historical role of evolutionary transitions in life cycle and host plant range in aphid speciation.

2.2. Patterns of host plant specialization: are aphids actually good botanists and why?

As mentioned above, aphids are specialized at several taxonomic levels. More than half of all aphid species feed on a single plant species, and many aphid genera are restricted to a single plant genus or several closely related plant genera [24]. Among the 445 non-host-alternating European Aphidinae, 407 (91%) feed on host plant species confined to a single genus [24]. Such host specificity extends across all aphid lineages: for example, all primary hosts of *Pemphigus* (Eriosomatinae) belong to *Populus* [8]; all species of Chaitophorus (Chaitophorinae) are confined to the genera Salix and Populus (both in Salicaceae). Some tribes are restricted to one or more genera within a single plant family: all primary hosts of Eriosomatini belong to two genera of Ulmaceae; Hormaphidini use primary hosts in Hamamelis (Hamamelidaceae); Siphini (Chaitophorinae) live on numerous genera of Poaceae, etc. Some aphid genera are confined to one or two plant families, e.g., Uroleucon feed only on Asteraceae and Campanulaceae [8,38]. Of course, such a pattern could result from the fact that many aphid taxa were circumscribed based on host association. However, these affinities of closely related aphids for particular host plant groups are confirmed by phylogenetic studies, and in some cases, aphid phylogenetic relationships are better explained by their host plant similarity than by morphological criteria. The molecular phylogenetic reconstruction of the Aphidinae clusters some aphid genera that are morphologically divergent but use related plants [36]: Myzus and Brachycaudus were thought unrelated based on their morphology [39], but nevertheless were found closely related in molecular studies, and both contain many species using Prunoideae as a host plant (as sole hosts or primary hosts). Similarly, pterocommatines (Pterocomma, Plocamaphis, etc.), feeding on Populus and Salix (Salicaceae), are so morphologically different from any other aphids as to be placed in a separate subfamily (Pterocommatinae) by a recent classification [6]; however, molecular phylogenetics placed pterocommatines firmly within Aphidinae-Macrosiphini as a sister group to Cavariella, the only other lineage of Aphidinae that has also diversified on Salicaceae [36].

The degree of host plant specialization in phytophagous insects is difficult to define and often subjective. Depending on the breadth of their host range, aphids have been classified as monophagous, oligophagous, polyphagous and even pantophagous [24]. An aphid species that uses

several plant species from the same genus probably exhibits a higher level of host specialization than a species that uses only two hosts, which are drawn from distantly related plant families. The former suggests stronger phylogenetic constraints in the colonization of new host plants and is more indicative of specialization from an evolutionary point of view. Blackman and Eastop [7.29.33] compiled an impressive body of data on host-plant association for all aphids. However, the level of host specialization in aphids has still not been extensively compared between subfamilies and genera. To date, only a few generalities have been drawn, such as all aphid species feeding on conifers are monophagous or oligophagous; aphids associated with monocotyledons are less specific than aphids associated with dicotyledons [18,40]; galling aphids (e.g., Hormaphidinae) are often very specialized [41]. Furthermore, levels of specialization in aphids have rarely been compared with other groups of phytophagous insects. Compared to other hemipterans, such as psyllids [24], aphids might not be as strictly monophagous, but they exhibit greater phylogenetic structure in their host plant ranges.

Several types of selective pressures could favour a specialized diet in aphids. Here we briefly review behavioural and life-history traits of aphids that are considered as important in shaping host-plant association ([9,42] for more comprehensive discussions). Plant chemical defences (secondary compounds), which are usually involved in plant-insect coevolution [43-45], have rarely been implicated in shaping long-term patterns of aphidplant associations. As phloem feeders, aphids probably circumvent many of those defences ([9] pp. 32-38 regarding the occurrence of chemical defences in phloem). One hypothesis to explain their specialist diet is simply that particular aphid species just perform better on particular plants that are more suitable as a resource for them; specificity would thus be the result of metabolic adaptation (see section 3) [42]. Mechanisms (behavioural, chemical) of host-plant location and recognition in aphids may also play an important role in either favouring or discouraging specialization [9]. However, the fact that some aphids host alternate between very different plant species argues against host specificity being solely a consequence of adaptation for finding and using the resources offered by a specific plant. An alternative hypothesis has been proposed by Ward [46], who suggested that host specificity was favoured because host plants serve not only as a source of nutrients but also as a mating site (termed the "rendezvous" hypothesis [9]); hence, host-range limitation is probably strongly linked to mating success. This hypothesis is clearly in agreement with the observation that heteroecious species exhibit a wider diversity of secondary hosts and a narrower range of primary hosts, where the sexual phase of the cycle is produced and mating occurs. Of the 141 heteroecious European Aphidinae, only nine have their secondary host plants confined to one genus [24]. Another strongly debated hypothesis is that of "fundatrix specialization" [37,47,48]. Under this hypothesis, host specialization in aphids is due largely to the phylogenetic constraints on the morph that hatches from overwintering eggs in spring, the

fundatrix, and perhaps also the sexual morphs. These morphs would be strongly adapted to the primary, often ancestral hosts, and less able to switch hosts than would other morphs. This hypothesis is also consistent with the general pattern of a narrower primary host-plant range than secondary host plant range in heteroecious species.

2.3. Phylogenetic studies of the role of host plants on speciation modes in aphids

There are no studies on the evolution of host association at the level of Aphididae as a whole; however, several case studies have focused on the historical association of certain aphid tribes and genera and their host plants in order to investigate how specialization, cospeciation and host shifts have influenced diversification. These studies are summarized in Table 1 and reviewed below.

Through a phylogenetic reconstruction of North American *Cinara*, Favret and Voetglin [49] proposed that speciation was accompanied by host shifts. Closely related species in the *Cinara* phylogenetic tree are often found associated with different *Pinus* species, while they maintain similar feeding sites (they are either branch or shoot feeders). This suggests that these aphids are better at switching hosts than at colonizing different ecological niches on the same host plant. Although the *Pinus* species serving as hosts of closely related *Cinara* are sympatric, no information exists on whether these host shifts and specialization occurred in sympatry or whether they were facilitated by geographical separation of populations.

Several phylogenetic and ecological studies of galling aphids in Fordini have been published [50–52]. Fordines are often host specific and repeated host shifts between related hosts have been inferred [51]. However, several occurrences of aphid sister species sharing the same host

plant species suggest that host shifts followed by host specialization and speciation are not the sole mechanism promoting diversification in this group. Study of two sympatric *Pemphigus* species sharing the same host (*P. populi* and *P. obesinymphae* on *Populus deltoides*) suggests that allochronic speciation (temporal isolation) has occurred. Hence, variation in plant phenology could be an important factor in generating aphid diversity [53].

Containing more than 197 valid species, Uroleucon constitutes one of the many genera that probably arose during the diversification of aphids on herbaceous plants in the Tertiary. Most *Uroleucon* species are generally restricted to a single host plant species or to several closely related species within Asteraceae or Campanulaceae. In some cases, species complexes are associated with groups of closely related plants; thus, Uroleucon is a seemingly good candidate for an example of aphid-host parallel diversification. However, a molecular phylogenetic reconstruction of the genus, based on only 17 species, clearly does not support a cospeciation hypothesis [38]: Uroleucon species have shifted repeatedly between Asteraceae tribes. Furthermore, the phylogeny implies that *Uroleucon* ancestors were not associated with a common ancestor of Campulanaceae and Asteraceae, as would be expected under a cospeciation scenario; rather, the association with Campulanaceae is a derived condition. Estimation of the age of the radiation suggests that Uroleucon species have diversified after Asteraceae became abundant in the fossil record, and speciation linked to host specialization has more likely originated through the acquisition of existing host taxa. Again, it is difficult to conclude whether host specialization followed by speciation could have occurred in sympatry based on molecular phylogenies alone. The authors of this study mentioned that most host plants of Uroleucon overlap over large parts of their geographic range; therefore,

Table 1Summary of phylogenetic studies within aphid genera or tribes investigating speciation scenario in relation to host plant association.

Taxon	Sampling	Host plants	Patterns of host plant association	Putative speciation mode
Cinara (Lachninae)	25 sp./150 sp. in the genus [49]	Mainly Pinus, but also Picea, Abies, Juniperus, Pseudotaxifoliae, Pseudotsuga	All species specialized on a genus of Conifers, 16 species specialized on one or two species, 9 more generalists. The same conifer species often hosts several <i>Cinara</i> sp.	Host switch
Tribe Fordini	16 sp./70 in the tribe [51] 12 species [52]	Pistacia and various secondary hosts	Specialized on one or two species of <i>Pistacia</i> . No specialization on secondary hosts. The same <i>Pistacia</i> sp. often hosts several Fordini	Host switch
Pemphigus (Eriosomatinae)	Phylogeography of 2 sp. and phylogeny of 8 sp./70 sp. [53]	Populus	Most species are specialized on one or several <i>Populus</i> sp. The same <i>Populus</i> sp. can host several <i>Pemphigus</i> sp.	Allochronic speciation
Uroleucon (Aphidinae)	15 sp./180 sp. [38]	Asteraceae, Campanulaceae	Most species are specialized on one or several plant species within the same genus, several polyphagous species	Host switch
Cryptomyzus (Aphidinae)	9 sp./17 sp. [54]	Ribes and/or Labiatae	Specialized on one or several species of <i>Ribes</i> and/or one or several plant species within Labiatae	Loss of host alternation Host switch
Brachycaudus (Aphidinae)	27 sp./50 sp. [56] Jousselin et al. in preparation	Prunus and or/various families of herbaceous plants	Specialized on one or several species of <i>Prunus</i> and/or specific to various species of herbaceous hosts, several polyphagous species	Loss and gains of host alternation Host switch

because mating does occur on the specific host plant, reproductive isolation could be favoured even in sympatry. Rapid diversification (as indicated by the lack of resolution in the deep nodes of the phylogeny) is also in favour of a sympatric mode of speciation [38].

A thorough study of speciation mode in *Cryptomyzus* aphids was conducted using phylogenetic studies and experimental work [54,55]. There are 17 valid species in this genus, with some species alternating between *Ribes* and several herbaceous species of the Lamiaceae family, and other species being monoecious on Lamiaceae. Two possible pathways of speciation were proposed in *Cryptomyzus*: host shift of a monoecious species onto a new herbaceous host followed by specialization and reproductive isolation (similar to the scenario inferred for *Uroleucon* [38]), and speciation via loss of the heteroecious life cycle and shift of all primary-host generations onto some of the secondary hosts of the original life cycle [54,55].

The genus *Brachycaudus* presents a pattern of life cycle evolution resembling the situation in *Cryptomyzus*. It includes heteroecious species (often alternating from a *Prunus* tree to herbaceous hosts from very diverse angiosperm families) and monoecious species that are often found on the secondary hosts of closely related heteroecious species [56]. *Brachycaudus* species also vary greatly in their host plant repertoire, ranging from strictly monophagous (most monoecious species) to highly polyphagous on secondary hosts when heteroecious; the latter condition might favour the capture of new hosts. This genus potentially illustrates speciation via acquisition of new, sometimes distantly related, hosts during the "summer" phase of the heteroecious life cycle, and through changes in life cycle (Jousselin et al. in prep).

Overall, the few studies of the history of host association within aphid genera all demonstrate that speciation is often associated with a host shift. However, these macroevolutionary patterns do not firmly demonstrate the role of plant use on reproductive isolation in aphids. At the most, they provide hints that host plant specialization, together with life cycle evolution, has probably played central roles in aphid diversification. Assessing the role of host plant association as a promoter of speciation necessitates intraspecific studies within natural populations, which are reviewed below.

3. Host specialization and divergence in natural populations: How can host plants drive aphid speciation?

3.1. Ecological speciation and patterns of host association

Host plants exert considerable selective pressures on their associated aphids, constituting their only food resource and habitat (including mating and oviposition sites). Because the vast majority of aphid species are host-specific (see section 2), it is tempting to infer their origin and divergence as a direct consequence of adaptation to different host plants, a hypothesis that was proposed long ago for phytophagous insects [57]. However, distinct host specificities may just result from local adaptation and competition between species that have previously diverged in geographical isolation (allopatry) [58]. Under

such processes, host specialization would merely allow the coexistence of host-adapted insects as a consequence of reproductive isolation, rather than causing it. This hypothesis does not appear to be considered in the scientific literature on aphid ecology [9].

By contrast, speciation promoted by divergent selection in contrasting environments, a process termed "ecological speciation" [59], is actively studied in a variety of organisms, including fishes, birds, fungi and plants [59,60]. Premating isolation (reduction of hybridization rate) may arise from mating occurring within the preferred habitat. In addition, the contrasting parental environments may not be suitable for hybrids, thereby causing post-zygotic isolation (hybrid unfitness). While the contribution of divergent natural selection to particular components of reproductive isolation has been quantified only in a handful of biological models [61–63], phytophagous insects, including aphids, offer good systems for studying these ecological reproductive barriers [64–66]. Ecological speciation can theoretically occur under any geographical context [67], but its study requires populations in partial reproductive isolation, in which ecological and non-ecological reproductive barriers can be distinguished (this would not be possible if no hybrid is produced). In phytophagous insects, these populations would correspond to host races [64].

Few insect species have been shown to encompass hostspecialized populations undergoing appreciable hybridization (typically at a rate greater than 1%) while maintaining significant genetic differentiation [64,68], such as the wellknown races of the apple maggot fly, Rhagoletis pomonella [69–71]. Within this scarcity of host race systems, there is still a non-negligible proportion of aphids, comprising attested or serious candidate models, most belonging to the most diverse subfamily, Aphidinae. The pea aphid, Acyrthosiphon pisum that essentially comprises monoecious populations (reviewed in reference [72]) encompasses at least eight conspecific host races specialized on different legume species [73], embracing the much-studied "clover" (Trifolium sp) and "alfalfa" (Medicago sativa) races in Europe [74,75] and North America [76,77]. In Hyalopterus pruni, two populations using either plum (Prunus domestica) or almond (P. dulcis) as primary hosts are highly genetically distinct in Europe and North America (a zone of introduction), but they hybridize on a shared host, apricot (P. armeniaca), at a significant rate (approximately 4%) [78]. The polyphagous aphid Myzus persicae comprises a specialized form on tobacco (Nicotiana tabacum) as a summer host [79] that is formally designated as the subspecies M. persicae nicotianae [7]. In Greece, sexual populations in tobacco-growing areas are genetically differentiated from those in other regions [80]. Distance alone cannot explain this differentiation, as the tobacco-feeding population is genetically uniform in most of the country. Reproductive isolation is likely ongoing, as suggested by behavioral studies of mating between tobacco lineages and those feeding on other crops, which occurs on the shared primary host, peach (Prunus persica) [80]. Given the moderate genetic differentiation between host-associated populations (populations analyzed at many microsatellites do not form genetic clusters as distinct as in the host race models presented above), biotypes of M. persicae probably have not reached species status, and may constitute host races in areas of sexuality and sympatry.

In several other aphid species, host races are suspected but not vet confirmed due to unknown hybridization rates. or even to a lack of any population genetic analysis. Here, the distinction of "biotypes", often named as subspecies, denotes any degree of population divergence below or at the species level [64] and the presence of minute morphological variations (body color or the presence of wings in specific morphs), sometimes confirmed on the basis of performance and preference assays on different host species. Müller [11,81] reviewed complexes of biotypes belonging to monoecious species, such as Aulacorthum solani, Acyrthosiphon malvae and Macrosiphum euphorbiae. Each complex presents various subspecies of unclear status, with various degrees of sympatry, host specificity and ability to hybridize. The Aphis fabae complex encompasses at least three subspecies alternating between spindle (Euonymus europaeus) and various herbaceous plants, and a fourth monoecious subspecies that remains on the primary host (spindle) all year round (case reviewed in reference [82]). These subspecies show distinct summer host preference and some degree of behavioral isolation [83,84], but most of them can be hybridized in the laboratory [82]. A comparable system is found in Pemphigus bursarius (Pemphigideae), whose specialized populations on distinct Asteraceae are highly genetically differentiated despite reproducing on a shared primary host, poplar [85]. The case of Cryptomyzus galeopsidis is opposite, in that two proposed host races (within a larger complex) colonize different primary hosts, redcurrant (Ribes rubrum) and blackcurrant (Ribes nigrum), but share their summer host (case reviewed in reference [64]). The absence of mating preference in the laboratory and the identification of possible hybrid clones [86] suggest host race status, but hybridization in the field is limited by preference to different primary hosts [87].

In addition to typical aphid populations that alternate asexual and sexual generations on a regular basis (see Simon et al., this issue), we may mention a number of host-associated biotypes comprising asexual lineages, and sometimes single clones. These occur in *Schizaphis graminum* on cereals [88], *Therioaphis trifolii* on clover and alfalfa in Australia [89] and *Aphis gossypii*, which presents specialized clones on unrelated host families (cotton, potato and Cucurbitaceae, amongst others) [14]. In the following, we do not consider asexual biotypes, as host plants cannot promote their reproductive isolation.

Hence, intraspecific differentiation onto different host plants is not unusual in aphids, at least in the Aphidinae. Moreover, many host-associated biotypes that do not differ morphologically might have gone unnoticed.

3.2. Host plants as catalysts of reproductive isolation in aphids

3.2.1. Sources of divergent selection and post-zygotic isolation

Feeding performance. In many complexes of biotypes, host specialization is measured through reciprocal transplants between host plants under laboratory conditions.

Under such a controlled setup, the cause of pronounced differential performance most likely entails nutrition in the broad sense: sap intake and assimilation, and reaction to associated plant defenses. Among the physiological and biochemical factors known to affect aphid nutrition [90,91], none has been unambiguously linked to food specialization in related biotypes. For instance, amino-acid requirements differ among pea aphid races [92], but performance on artificial diets designed to mimic the amino acid composition of the phloem sap from the different hosts does not reflect performance on the plants themselves [93].

Perhaps more relevant to speciation are the genetic components of differential performance, which few studies were able to distinguish from the effect of conditioning, since lineages of different biotypes are generally reared on their respective native plant before fitness experiments. Conditioning (including maternal effects) can markedly improve feeding performance on alternate hosts, without totally explaining difference observed between lineages from different plant origins ([94,95] for tests on *Sitobion avenae* and *Aphis gossypii*, respectively). In pea aphids, performance on alternate hosts is only marginally altered by conditioning [96] and strong specialization to the native host is still observed after rearing on a shared host plant prior to tests [73,97].

Genetically based food specialization may induce reproductive isolation if intermediate genotypes, typically hybrids, present mitigated performance, as the expression of a fitness tradeoff preventing the optimal use of both parental environments. Such ecological hybrid unfitness is difficult to distinguish from non-ecological causes of poor performance, mainly intrinsic genetic incompatibilities affecting hybrids independently of their environment [98]. A rare example comes from a cross between pea aphid lines from red clover and common broom (Cytisus scoparius) where resulting hybrids performed very poorly on the parental plants, but reportedly grew well on the universally favorable broad bean (Vicia faba) [99]. The quantitative genetics of performance in hybrids between races on red clover and alfalfa revealed genomic regions conferring a higher fecundity on a parental host species, but a lower performance on the other [76]. This genetic fitness trade-off prevents the evolution of a generalist genotype on both red clover and alfalfa and explains the intermediate hybrid performances on these plants. Mackenzie [100,101] also demonstrated a trade-off between the optimal use of broad bean (Vicia faba) and nasturtium (Tropaeolum majus) by different genotypes of Aphis fabae and F2 crosses. In other models, crosses between hostspecialized biotypes gave disparate results (reviewed in references [11,81]). Some hybrid lines can complete their cycle normally on at least one parental host, suggesting the absence of strong intrinsic genetic incompatibility. Others show various deficiencies in the laboratory, such as poor egg hatching success and defects of sexual morph production [102], which are unlikely to be related to nutrition or host adaptation in general. The strength of post-zygotic isolation often corroborates the perceived degree of divergence between biotypes, considered as races or good species, but it does not constitute evidence for a contribution of host plants in this reproductive barrier. However, it seems reasonable to propose that feeding on several host plants is subject to tradeoffs and that it may be a common source of hybrid unfitness in aphids.

Phenology and life cycle. An insect's life cycle may not be optimal on several plant species with contrasting phenologies (reviewed in reference [66]). In aphids, trade-offs might prevent the optimal timing of some crucial phases of the life cycle on hosts that are suitable for different periods. Egg-hatch is genetically based and synchronized with spring budburst in some species [103,104]. In Aphis spireacola, two biotypes on mikan (Citrus unshiu) and Spiraea thunbergii primary hosts differ by approximately a month in the dates of fundatrix emergence [105]. Hybrid lineages emerging before bud break on mikan starve to death [106], providing a possible case of ecological selection against gene flow through maladapted duration of egg dormancy, in addition to probable food specialization [107]. Similar divergent selection on adult emergence time may act on biotypes of Tetraneura yezoensis producing galls on *Ulmus davidiana* or *U. laciniata* [103], but selection against hybrids has not vet been investigated. Overall. divergent selection by host phenology in aphids may not be as universal as selection by food resource. Perennial herbaceous hosts of monoecious biotypes, like most of those constituting the pea aphid complex [108], may be favorable all year round and would not strongly select for different parasite life cycles.

Aphids may be under divergent selection by other features of host plants and their associated habitat, such as morphology, host-associated enemies and mutualists (reviewed in reference [9]), but few of these have been investigated in relation to aphid speciation. Notably, in aphids as in many sap-sucking insects, bacterial symbionts might influence nutrition [109,110]. The obligate endosymbiont of Aphididae, Buchnera aphidicola supplements the imbalanced phloem sap by supplying aphids with essential amino acids [111]. Specific symbiont strains are equivalent to cytoplasmic loci affecting feeding performance (see above). However, some facultative bacterial symbionts, which are frequent in aphids, may be acquired by lateral transfer [112], and may be partly seen as elements of the aphid ecological niche affecting host adaptation. So far, there is little evidence for the influence of secondary symbionts on host use by aphids. In one pea aphid clone, the suppression of the bacterium Regiella insecticola, prevailing in pea aphids from clovers (Trifolium), critically reduced performance on Trifolium repens while performance on Vicia sativa remained unchanged [113]. This result has not been reproduced in other clones or in other symbionts that are associated with different host plants of the complex [114-116]. The association of facultative symbionts with specialized populations or clones is not general [117] and it is not clear how the presence of symbionts may act against hybrids, the same way a genetic tradeoff in host performance can.

3.2.2. Host plants as a source of premating isolation

Many aphid biotypes feed on different plant species that grow within their dispersal range. Mating on these different hosts translates into *habitat isolation* (reduction of encounter rate between populations mating within distinct habitats). The relevance of habitat isolation to speciation assumes the incompleteness of other reproductive barriers. In *Hyalopterus pruni* and *Acyrthosiphon pisum*, occasional hybridizations between races meeting on the same host plants have been observed [73,78], suggesting that premating isolation in the field can largely be enhanced by specialization to distinct hosts.

The first premating barrier expressed in the aphid life cycle, during plant colonization, is preference to distinct host species. The stages and influencing factors of host plant selection in aphids are well described [91], but those explaining differential preference among related biotypes are rarely identified. The tobacco biotype of Myzus persicae responds to volatile cues emitted by tobacco while the generalist biotype does not [118], indicating distinct differences in olfactory perception between them (M. persicae subspecies still prefer the same primary host, peach). More often, gustatory reception appears involved in differential host preference, as colonizers frequently reject alternate hosts only after probing with their stylets before reaching the phloem, under the influence of chemical stimulants or repellents [81,91]. In pea aphid races [119], host acceptance can be triggered by epidermal extract from the home plant [120], suggesting that colonizers can sense host-specific stimulants. In this complex, differences in host preference appear genetically based and mildly affected by conditioning [81], although this observation may not apply for other specialized populations (e.g. in *Aphis fabae* [121]). Linkage between loci controlling host choice and host adaptation may further facilitate premating isolation in the face of gene flow between pea aphid races [76].

In several species, winged colonizers often show strong preference to host plants that they may not have even experienced before dispersal [122]. In heteroecious species, returning morphs of different biotypes will prefer different primary hosts, as shown in males of Crytpomyzus galeopsidis mating on redcurrant or blackcurrant [87]. Because male aphids may use plant volatiles to locate mates [122] they may show particularly pronounced host discrimination. Returning females (gynoparae) also show stronger host preference than subsequent generations using the same primary host in Aphis fabae on spindle [123] and in Rhopalosiphum padi on bird cherry, Prunus padus [124]. These female morphs may therefore particularly enhance spatial isolation between biotypes, but their degree of preference has not been investigated in a complex of biotypes on different primary hosts.

Because plant colonization occurs before mating, host preference may actually account for most of the reproductive isolation between biotypes [125]. However, a significant proportion of colonizers choose less favorable hosts in preference assays [118,126] or in the field [73,127]. In this case, selection against poorly adapted immigrants may strengthen habitat isolation. In monoecious lineages, where host colonization may long precede the mating season [9], host plants select not only against poorly adapted migrants, but also against their parthenogenetic progeny. Such selection has been evidenced in pea aphid races on red clover and alfalfa, whose genetic differentiation increases throughout the summer [12,128].

Allochronic isolation (populations mating at different periods) is another premating barrier that could evolve between host-associated biotypes. The biotypes of Aphis spiraecola on mikan and Spiraea, which differ in emergence time (see above) also differ in the period of production of sexuals and are therefore in strong allochronic isolation [106]. The pea aphid race using the pea as a summer host is also described as producing sexuals later than the races using perennial crops [129]. Allochrony in sexual morph production could match plant phenologies, but no causal link has been strictly demonstrated. In all likelihood, the contribution of allochrony to premating isolation in hostassociated aphids is probably weaker than the contribution of host preference and selection against immigrants, which affect virtually all sympatric biotypes mating on different host species.

In biotypes sharing the same primary host, specialization to different secondary hosts cannot directly promote premating isolation. However, summer host plants may indirectly promote premating isolation between biotypes on the same primary host if they select against hybrids (see section 3.2.1). The avoidance of maladaptive hybridization, termed *reinforcement* [130], may explain behavioral isolation in several complexes of biotypes [80,131]. Females emit pheromones at different times of the day corresponding to periods of male activity in *Myzus persicae* [80] and *Aphis fabae* [83] on the peach and spindle, respectively, and between *Cryptomyzus galeopsidis* cryptic species on redcurrant [131,132]. However, the hypothesis

that these premating barriers have evolved by reinforcement remains to be tested in these models.

3.3. Evidence and context of speciation through host shifts

The mechanisms by which host plants may induce reproductive isolation in aphids (Table 2) constitute various aspects of host specialization. However, widespread host specialization in aphids may actually have evolved as a consequence of divergence and does not constitute evidence for a direct role of host plants in speciation. In only few models have some specific predictions of ecological speciation been verified (Table 2, bottom). An experiment by Shaposhnikov on Dysaphis [133-135] (Table 2) might still constitute the most remarkable demonstration of hostinduced speciation. In Dysaphis anthrisci (feeding on Anthriscus), Shaposhnikov successfully transferred one clone to Chaerophyllum, a normally unsuitable plant that hosts the related species D. chaerophyllina. In about 20 parthenogenetic generations, the D. anthrisci clone apparently adapted to its new host, gained some morphological resemblance and reproductive compatibility with D. chaerophyllina, while almost losing all ability to produce viable eggs with its parent species! This unique case of experimentally repeated, or reversed, ecological speciation (depending on the original direction of the host shift in nature) should however be taken with great caution, as the possibility of contamination of the reared D. anthrisci clone by some *D. chaerophyllina* line has not been excluded.

Table 2Observations relevant to host-induced speciation in aphids.

Category	Observation	Biological models (non exhaustive list)
Features favourable to host-induced sp	eciation	
Ecological specialization	Range restricted to few related plant species or genera Intraspecific specialization to distinct host species	Most Aphididae [24] Many Aphidinae (reviewed in references [11,64,81,82])
Life cycle Feeding resources (selective agents) are mating sites All stages can feed on the same host plant/species		All aphids All non host-alternating biotypes
Host-induced post-zygotic isolation	Trade-offs in performance on the parental hosts affect hybrids Maladapted life cycle (timing of egg hatch) in hybrids	Acyrthosiphon pisum [76], Aphis fabae [100] Aphis spireacola [105,106]
Host-induced premating isolation	Preference to favourable mating sites (hosts)	Acyrthosiphon pisum [119,126], Cryptomyzus spp. [87]
	Selection against immigrants from other host species Timing of production of sexual morphs may match the host phenology (possible allochronic isolation)	Acyrthosiphon pisum, [12] Aphis spiraecola [106], A. pisum? [129]
	Behavioural isolation on shared primary hosts (possible reinforcement)	Myzus persicae [80], Aphis fabae [83], Cryptomyzus galeopsidis [131]
Tests of host-induced reproductive isol	• •	4 4 1 1 1 1001
Is RI lower in the absence of selection by host plants?	Hybrids perform better on non-selective medium	Acyrthosiphon pisum [99]
-, ·····	Substantial hybridization is observed on shared hosts or despite limited migration	Acyrthosiphon pisum [73], Hyalopterus pruni [78]
Does the strength of RI positively correlate with that of divergent selection?	Differences in host adaptation <i>do not</i> correlate with levels of genetic differentiation between biotypes	Acyrthosiphon pisum [73]
Does adaptation to different hosts induce genetic differentiation?	Loci controlling host use induce widespread genomic differentiation between biotypes	Acyrthosiphon pisum [140]
Can RI be induced by selection on different host plants?	Adaptation to a new host induce RI with lineages maintained on the ancestral host	Dysaphys anthrisci? (see text) [133–135]

The geographical context of divergence may also indirectly suggest an active role of host adaptation, if prolonged allopatry is excluded. Sympatric speciation in animals is a hotly debated topic [125,136] that has long been proposed in phytophagous insects, especially after the work on Rhagoletis pomonella host races [70]. This mode of ecological speciation assumes that host-associated populations remain within each other's dispersal range and that gene flow, starting at 50% (panmixia), is maintained until speciation is over. In practice, allopatric divergence cannot be excluded in aphids, as known species complexes have wide continental ranges, and because geographical distributions are not thoroughly known. For instance, certain European countries do not seem to harbor some subspecies of Aphis fabae [11], suggesting some possible geographic separation. Biotypes of Hyalopterus pruni occuring in sympatry across their widely studied range are weakly geographically differentiated [15], but their origin of several million years ago leaves ample time for earlier allopatry. On the other hand, the Myzus persicae nicotianae subspecies may present a rare case of divergence onto a recently introduced crop, tobacco (cultivated in Eurasia since the sixteenth century), similar to the emergence of the Rhagoletis pomonella race after apple was introduced in North America. Given the extreme polyphagy and wide distribution of Myzus persicae s.s. [7], strict allopatric divergence of the subspecies nicotianae appears improbable. Similarly, the very recent divergence of pea aphid races and species may have been promoted by the increasing availability of some of their legume hosts through agriculture and climate warming in the Holocene [137]. It is not clear how such a context would favor repeated allopatric divergences on plants that are now broadly sympatric, given that the radiation occurred after the last glacial episode. Furthermore, the eleven pea aphid biotypes show variable rates of hybridization and genetic differentiation that may illustrate the gradual evolution of host races into good species [73].

4. Conclusion and future prospects

Patterns of aphid-plant associations at recent or deeper evolutionary timescales are compatible with widespread plant-driven diversification in the aphid family, with the major radiation of Aphididae following the diversification of angiosperms. Future developments in molecular phylogenetics should focus on obtaining reliable evolutionary relationships of Aphididae subfamilies as well as investigating their monophyly in order to confirm major transitions in aphid-host plant associations. Current taxonomic and phylogenetic studies within subfamilies and genera reveal that divergence events typically involve shifts to new plant species that are often related to the ancestral host, and occasional changes in life cycle from host alternation to monoecy and vice versa. More studies investigating the history of host plant association within aphid genera should allow a better understanding of these patterns. Due to their pest status, aphids have generated an impressive amount of data on host plant association and thorough taxonomic studies. Global analyses such as comparative studies of levels of host plant specialization

and diversification rates in aphids represent promising perspectives for elucidating the role of host plants in aphid diversification.

Divergent natural selection imposed by several components of the host-plant niche, in particular food resource and phenology, may promote pre- and post-zygotic reproductive barriers, permitting speciation in the absence of complete geographical isolation. However, the hypothesis of diversification actively driven by host plants has been seldom tested with specific predictions (Table 2), and must be weighed against scenarios where adaptation to different host plants would be a consequence rather than a cause of reproductive isolation. This overlooked hypothesis suffers from the use of host plant information to delineate related aphid species or biotypes. Studies conducted above the species level cannot discriminate between non-ecological factors and the influence of host plant on reproductive isolation. However, population studies may allow comparisons between levels of reproductive isolation among populations with different degree of ecological divergence (same host plant vs. different host plants), but with similar divergence time [62]. Ecological speciation predicts higher reproductive isolation in ecologically different populations. Under a similar approach, correlating the strength of reproductive isolation with the degree of ecological divergence between populations [138,139] may reveal whether host adaptation generally contributes to divergence in aphids. Experimental studies may also focus on reproductive isolation between biotypes under various setups, in order to compare its strength in the presence and absence of selection by host plants. Finally, the production of discrete morphs under different environmental conditions (termed as polyphenism) is an intrinsic characteristic of aphids that could be investigated in the framework of speciation. The timing of production of sexual and winged migrating morphs, for instance, shapes life cycles in connection with host phenology and host alternation. Loci controlling this trait may be a target of divergent selection and contribute to reproductive isolation between aphid populations on different host ranges.

Acknowledgments

We thank J.-L. Bonnemain for giving us the opportunity to present this review, and two anonymous referees for their valuable comments on the manuscript.

References

- K. Tilmon, Specialization, speciation, and radiation: the evolutionary biology of herbivorous insects, University of California Press, Los Angeles, 2008.
- [2] C. Mitter, B. Farrell, D.J. Futuyma, Phylogenetic studies of insect-plant interactions: insights into the genesis of diversity, Trends in Ecology and Evolution 6 (1991) 290–293.
- [3] B. Farrell, C. Mitter, D.J. Futuyma, Diversification at the Insect-Plant Interface: insights from phylogenetics, Bioscience 42 (1992) 34–42.
- [4] T. Jermy, Evolution of insect/plant relationship, The American Naturalist 124 (1984) 609–630.
- [5] A.S. Winkler, C. Mitter, The phylogenetic dimension of insect-plant interactions: a review of the evidence, in: K.J. Tllmon (Ed.), Specialization, Speciation, and Radiation, The Evolutionary Biology of Herbivorous Insects, University of California Press, Berkeley, 2008, pp. 240– 263.

- [6] G. Remaudière, M. Remaudière, Catalogue des Aphididae du monde-Catalogue of the world's Aphididae, Homoptera, Aphidoidea, Paris, 1997
- [7] R. Blackman, V.F. Eastop, Aphids on the World's Herbaceous Plants and Shrubs, Wiley and Sons, Hoboken, USA, 2006.
- [8] D. Hille Ris Lambers, Aphids as Botanists? Symbolae Botanicae Uppsalienses. 1979 pp. 114–119.
- [9] A.F.G. Dixon, Aphid Ecology-An optimization approach, Kluwer Academic Pub. Norwell. 1998.
- [10] M. Inaizumi, Studies on the life cycle and polymorphism of *Aphis Gossypii* Glover (Homptera, Aphididae), Special Bulletin of the College of Agriculture, Utsunomiya University, 1980, pp. 132.
- [11] F.P. Müller, Biotype formation and sympatric speciation in aphids (homoptera, aphidinea), Entomologia Generalis 10 (1985) 161–181.
- [12] S. Via, A.C. Bouck, S. Skillman, Reproductive isolation between divergent races of pea aphids on two hosts. II, Selection against migrants and hybrids in the parental environments, Evolution 54 (2000) 1626–1637.
- [13] J.T. Margaritopoulos, C. Tsourapas, M. Tzortzi, O.M. Kanavaki, J.A. Tsitsipis, Host selection by winged colonisers within the *Myzus persicae* group: a contribution towards understanding host specialisation, Ecological Entomology 30 (2005) 406–418.
- [14] J. Carletto, E. Lombaert, P. Chavigny, T. Brevault, L. Lapchin, F. Vanlerberghe-Masutti, Ecological specialization of the aphid *Aphis gossypii* Glover on cultivated host plants, Molecular Ecology 18 (2009) 2198– 2212.
- [15] J.D. Lozier, R.G. Foottit, G.L. Miller, N.J. Mills, G.K. Roderick, Molecular and morphological evaluation of the aphid genus *Hyalopterus* Koch (Insecta: Hemiptera: Aphididae), with a description of a new species, Zootaxa (2008) 1–19.
- [16] C.D. von Dohlen, N.A. Moran, Molecular Phylogeny of the Homopteraa Paraphyletic Taxon, Journal of Molecular Evolution 41 (1995) 211– 223.
- [17] O.E. Heie, in: E.J. Brill (Ed.), The Aphidoidea (Hemiptera) of Fennoscandia and Denmark. IV. Family Aphididae: part 1 of tribe Macrosiphini of subfamily Aphidinae, Scandinavian Science Press Ltd, Leiden, New York, Kobenhavn, Köln, 1992.
- [18] G.K. Shaposhnikov, Evolution of aphids in relation to evolution of plants, in: A.K. Minks, P. Harrewin (Eds.), Aphids. Their biology, Natural Enemies and Control, Elsevier, Amsterdam, 1987, pp. 409– 413.
- [19] O.E. Heie, The evolutionary history of aphids and a hypothesis on the coevolution of aphids and plants, Bolletino di Zoologia agraria e di Bachicoltura Series II (1996) 149–155.
- [20] W.R. Richards, Systematics of fossil aphids from Canadian amber (Homoptera: Aphidinae), The Canadian Entomologist 98 (1966) 746–760.
- [21] C.D. von Dohlen, N.A. Moran, Molecular data support a rapid radiation of Aphids in the Cretaceous and multiple origins of host alternation, Biological Journal of the Linnean Society 71 (2000) 689–717.
- [22] D. Martinez-Torres, C. Buades, A. Latorre, A. Moya, Molecular systematics of aphids and their primary endosymbionts, Molecular Phylogenetics and Evolution 20 (2001) 437–449.
- [23] B. Ortiz-Rivas, A. Moya, D. Martinez-Torres, Molecular systematics of aphids (Homoptera: Aphididae): new insights from the long-wavelength opsin gene, Molecular Phylogenetics and Evolution 30 (2004) 24-37.
- [24] V.F. Eastop, Deductions from the present day host plants of aphids and related insects, in: H.F. van Emden (Ed.), Insect/Plant relationships., Symp. Roy. Entomol. Soc. Lond., John Wiley and Sons, New York, 1973 pp. 157–178.
- [25] O.E. Heie, Paleontology and Phylogeny, in: A.K. Minks, P. Harrewin (Eds.), Aphids, Their biology, Natural Enemies and Control, Elsevier, Amsterdam, 1987, pp. 367–391.
- [26] O.E. Heie, Studies on fossil aphids (Homoptera: Aphidoidea), Spolia Zoologica Musei Hauniensis 26 (1967) 1–273.
- [27] D. Hille Ris Lambers, Host plants and aphid classification, in: 8th International congress of Entomology, Stockholm, (1950), pp. 141– 144
- [28] A.F.G. Dixon, P. Kindlmann, J. Leps, J. Holman, Why there are so few species of aphids, especially in the tropics, The American Naturalist 129 (1987) 580–592.
- [29] B.K. Blackman, V.F. Eastop, Aphids on the World's Trees: an Identification and Information Guide, CABI, Wallingford, 1994.
- [30] O.E. Heie, Why are there so few aphid species in the temperate areas of the southern-hemisphere? European Journal of Entomology 91 (1994) 127–133.
- [31] C.D. von Dohlen, D.A.J. Teulon, Phylogeny and historical biogeography of New Zealand indigenous aphidini aphids (Hemiptera, Aphididae): An hypothesis, Annals of the Entomological Society of America 96 (2003) 107–116.

- [32] C.D. von Dohlen, U. Kurosu, S. Aoki, Phylogenetics and evolution of the eastern Asian-eastern North american disjunct aphid tribe, Hormaphidini (Hemiptera: Aphididae), Molecular Phylogenetics and Evolution 23 (2002) 257-267.
- [33] R.L. Blackman, V.F. Eastop, Aphids on the world's crops: an identification and information guide, Willey and sons, New York, 2000.
- [34] N.A. Moran, The Evolution of Aphid Life Cycles, Annual Review of Entomology 37 (1992) 321–348.
- [35] N.A. Moran, A 48-Million-Year-Old Aphid Host Plant-Association and Complex Life-Cycle-Biogeographic Evidence, Science 245 (1989) 173– 175.
- [36] C.D. von Dohlen, C.A. Rowe, O.E. Heie, A test of morphological hypotheses for tribal and subtribal relationships of Aphidinae (Insecta: Hemiptera: Aphididae) using DNA sequences, Molecular Phylogenetics and Evolution 38 (2006) 316–329.
- [37] N.A. Moran, Aphid life cycles: two evolutionary steps, The American Naturalist 136 (1990) 135–138.
- [38] N.A. Moran, M.E. Kaplan, M.J. Gelsey, T.G. Murphy, E.A. Scholes, Phylogenetics and evolution of the aphid genus *Uroleucon* based on mitochondrial and nuclear DNA sequences, Systematic Biology 24 (1999) 85–93.
- [39] C.V. Börner, K. Heinze, Aphidina-Aphidoidea, in: P. Sorauer (Ed.), Hanbuch der Pflanzenkrankheiten, 2. Teil, 4. Lief. Homoptera II, Verlag Paul Barey, Berlin, 1957, pp. 1–355.
- [40] G.K. Shaposhnikov, The main features of the evolution of aphids, in: International Aphidological Symposium at Jablonna, polska Academia Nauk, Warsaw, Jablonna, (1981), pp. 19–99.
- [41] D.L. Stern, Phylogenetic evidence that aphids, rather than plants, determine gall morphology, in: Proceedings of the Royal Society of London–B 260, 1995, pp. 85–89.
- [42] N.A. Moran, Evolutionary determinants of host specificity in *Uroleu-con*, in: J. Holman, J. Pelikan, A.F.G. Dixon, L. Weisman (Eds.), Population structure, genetics and Taxonomy of aphids and Thysanoptera, Smolenice, Czechoslovakia, SPB Academic Publishing, 1985, pp. 29–38.
- [43] P.R. Ehrlich, P.H. Raven, Butterflies and plants: a study in coevolution, Evolution 18 (1964) 586–608.
- [44] J. Jaenike, Host specialization in phytophagous insects, Annual Review of Ecology and Systematics (1990).
- [45] J.X. Becerra, Insects on plants: macroevolutionary chemical trends in host use. Science 276 (1997) 253–256.
- [46] S.A. Ward, Reproduction and host selection by aphids: the importance of "rendezvous" hosts, in: W.J. Bailey, J. Ridshill-Smith (Eds.), Reproductve Behaviour of Insects, Chapman and Hall, New york, 1991, pp. 202–226.
- [47] N.A. Moran, Adaptation and Constraint in the Complex Life-Cycles of Animals, Annual Review of Ecology and Systematics 25 (1994) 573– 600
- [48] N.A. Moran, The evolution of host plant alternation in aphids: evidence for specialization as a dead end, The American Naturalist 132 (1988) 681–706.
- [49] C. Favret, D.J. Voetglin, Speciation by host-switching in pinyon Cinara (Insecta: Hemiptera: Aphididae), Molecular Phylogenetics and Evolution 32 (2004) 139–151.
- [50] D. Wool, Galling aphids: specialization, biological complexity and variation, Annual Review of Entomology 49 (2004) 175–192.
- [51] M. Inbar, M. Wink, D. Wool, The evolution of host plant manipulation by insects: molecular and ecological evidence from gall-forming aphids on *Pistacia*, Molecular Phylogenetics and Evolution 32 (2004) 504–511.
- [52] B. Ortiz-Rivas, D. Martinez-Torres, N.P. Hidalgo, Molecular phylogeny of Iberian Fordini (Aphididae: Eriosomatinae): implications for the taxonomy of genera Forda and Paracletus, Systematic Entomology 34 (2009) 293–306.
- [53] P. Abbot, J.H. Withgott, Phylogenetic and molecular evidence for allochronic speciation in gall-forming aphids (*Pemphigus*), Evolution 58 (2004) 539–553.
- [54] J.A. Guldemond, On aphid, their host plants and speciation: a biosystematic study of the genus *Cryptomyzus*, Wagenigen (1990) 157.
- [55] J.A. Guldemond, Host Plant Shift, Host Race Formation and Speciation in Cryptomyzus (Homoptera, Aphididae), Acta Phytopathologica Et Entomologica Hungarica 25 (1990) 89–96.
- [56] A. Coeur d'Acier, G. Cocuzza, E. Jousselin, V. Cavalieri, S. Barbagallo, Molecular phylogeny and systematic in the genus Brachycaudus (Homoptera: Aphididae): insights from a combined analysis of nuclear and mitochondrial genes, Zoologica Scripta 37 (2008) 175–193.
- [57] B. Walsh, On phytophagic varieties and phytophagic species, Proceedings of the Entomological Society, Philadelphia 3 (1864) 403–430.
- [58] D.J. Futuyma, G. Moreno, The Evolution of Ecological Specialization, Annual Review of Ecology and Systematics 19 (1988) 207–233.

- [59] D. Schluter, Ecology and the origin of species, Trends in Ecology and Evolution 16 (2001) 372–380.
- [60] H.D. Rundle, P. Nosil, Ecological speciation, Ecology Letters 8 (2005) 336–352
- [61] J.S. McKinnon, S. Mori, B.K. Blackman, L. David, D.M. Kingsley, L. Jamieson, J. Chou, D. Schluter, Evidence for ecology's role in speciation, Nature 429 (2004) 294–298.
- [62] P. Nosil, B.J. Crespi, C.P. Sandoval, Host-plant adaptation drives the parallel evolution of reproductive isolation, Nature 417 (2002) 440–443.
- [63] J.R. Dettman, C. Sirjusingh, L.M. Kohn, J.B. Anderson, Incipient speciation by divergent adaptation and antagonistic epistasis in yeast, Nature 447 (2007), 585-+.
- [64] M. Drès, J. Mallet, Host races in plant-feeding insects and their importance in sympatric speciation, Philosophical Transactions of the Royal Society of London, Series B: Biological Sciences 357 (2002) 471–492.
- [65] S.H. Berlocher, J.L. Feder, Sympatric speciation in phytophagous insects: moving beyond controversy? Annual Review of Entomology 47 (2002) 773–815.
- [66] K.W. Matsubayashi, I. Ohshima, P. Nosil, Ecological speciation in phytophagous insects, Entomologia Experimentalis Et Applicata 134 (2009) 1–27.
- [67] D.I. Bolnick, B.M. Fitzpatrick, Sympatric Speciation: models and Empirical Evidence, Annual Review of Ecology, Evolution, and Systematics 38 (2007) 459–487.
- [68] J. Mallet, Hybridization, ecological races and the nature of species: empirical evidence for the ease of speciation, Philosophical Transactions of the Royal Society of London, Series B: Biological Sciences 363 (2008) 2971–2986.
- [69] B. Walsh, The apple-worm and the apple maggot, Journal of Horticulture 2 (1867) 338–343.
- [70] G.L. Bush, Sympatric host race formation and speciation in frugivorous flies of the genus *Rhagoletis* (Diptera: Tephritidae), Evolution 23 (1969) 237–251.
- [71] J.L. Feder, S.H. Berlocher, J.B. Roethele, H. Dambroski, J.J. Smith, W.L. Perry, V. Gavrilovic, K.E. Filchak, J. Rull, M. Aluja, Allopatric genetic origins for sympatric host-plant shifts and race formation in *Rhagoletis*, Proceedings of the National Academy of Sciences, USA 100 (2003) 10314–10319.
- [72] J. Peccoud, J.-C. Simon, The pea aphid complex as a model of ecological speciation, Ecological Entomology 35 (2010) 119–130.
- [73] J. Peccoud, A. Ollivier, M. Plantegenest, J.-C. Simon, A continuum of genetic divergence from sympatric host races to species in the pea aphid complex, in: Proceedings of the National Academy of Sciences, USA, 106, (2009), pp. 7495–7500.
- [74] J.-C. Simon, S. Carré, M. Boutin, N. Prunier-Leterme, B. Sabater-Munoz, A. Latorre, R. Bournoville, Host-based divergence in populations of the pea aphid: insights from nuclear markers and the prevalence of facultative symbionts, Proceedings of the Royal Society of London, Series B: Biological Sciences 270 (2003) 1703-1712.
- [75] A. Frantz, M. Plantegenest, L. Mieuzet, J.-C. Simon, Ecological specialization correlates with genotypic differentiation in sympatric hostpopulations of the pea aphid, Journal of Evolutionary Biology 19 (2006) 392–401.
- [76] D.J. Hawthorne, S. Via, Genetic linkage of ecological specialization and reproductive isolation in pea aphids, Nature 412 (2001) 904–907.
- [77] S. Via, The genetic structure of host plant adaptation in a spatial patchwork: demographic variability among reciprocally transplanted pea aphid clones, Evolution 45 (1991) 827–852.
- [78] J.D. Lozier, G.K. Roderick, N.J. Mills, Genetic evidence from mitochondrial, nuclear, and endosymbiont markers for the evolution of host plant associated species in the aphid genus Hyalopterus (hemiptera: Aphididae), Evolution 61 (2007) 1353–1367.
- [79] N.N. Nikolakakis, J.T. Margaritopoulos, J.A. Tsitsipis, Performance of Myzus persicae (Hemiptera: Aphididae) clones on different host-plants and their host preference, Bulletin of Entomological Research 93 (2003) 235–242.
- [80] J.T. Margaritopoulos, G. Malarky, J.A. Tsitsipis, R.L. Blackman, Microsatellite DNA and behavioural studies provide evidence of host-mediated speciation in *Myzus persicae* (Hemiptera: Aphididae), Biological Journal of the Linnean Society 91 (2007) 687–702.
- [81] F.P. Müller, Genetic and evolutionary aspects of host choice in phytophagous insects, especially aphids, Biologisches Zentralblatt 104 (1985) 225–237.
- [82] F.P Müller, The role of subspecies in aphids for affairs of applied entomology, Zeitschrift Fur Angewandte Entomologie 101 (1986) 295–303.
- [83] T. Thieme, A.F.G. Dixon, Mate recognition in the Aphis fabae complex: daily rhythm of release and specificity of sex pheromones, Entomologia Experimentalis Et Applicata 79 (1996) 85–89.

- [84] B. Raymond, J.B. Searle, A.E. Douglas, On the processes shaping reproductive isolation in aphids of the *Aphis fabae* (Scop.) complex (Aphididae: Homoptera), Biological Journal of the Linnean Society 74 (2001) 205, 215
- [85] N.J. Miller, N.B. Kift, G.M. Tatchell, Host-associated populations in the lettuce root aphid, Pemphigus bursarius (L.), Heredity 94 (2005) 556– 564.
- [86] J.A. Guldemond, A.F.G. Dixon, J.A. Pickett, L.J. Wadhams, C.M. Wood-cock, Specificity of sex-pheromones, the role of host-plant odor in the olfactory attraction of males, and mate recognition in the aphid *Cryptomyzus*, Physiological Entomology 18 (1993) 137–143.
- [87] J.A Guldemond, Choice of Host Plant as a Factor in Reproductive Isolation of the Aphid Genus Cryptomyzus (Homoptera, Aphididae), Ecological Entomology 15 (1990) 43–51.
- [88] K.A. Shufran, J.D. Burd, J.A. Anstead, G. Lushai, Mitochondrial DNA sequence divergence among greenbug (Homoptera: Aphididae) biotypes: evidence for host-adapted races, Insect Molecular Biology 9 (2000) 179–184.
- [89] P. Sunnucks, F. Driver, W.V. Brown, M. Carver, D.F. Hales, W.M. Milne, Biological and genetic characterization of morphologically similar Therioaphis trifolii (Hemiptera: Aphididae) with different host utilization, Bulletin of Entomological Research 87 (1997) 425–436.
- [90] A.E. Douglas, The nutritional physiology of aphids, Advances in Insect Physiology 31 (31) (2003) 73–140.
- [91] G. Powell, C.R. Tosh, J. Hardie, Host plant selection by aphids: behavioral, evolutionary, and applied perspectives, Annual Review of Entomology 51 (2006) 309–330.
- [92] P.N. Srivastava, Y. Gao, J. Levesque, J.L. Auclair, Differences in aminoacid requirements between 2 biotypes of the pea aphid, *Acyrthosiphon pisum*, Canadian Journal of Zoology 63 (1985) 603–606.
- [93] J. Sandström, J. Pettersson, Amino-Acid-Composition of Phloem Sap and the Relation to Intraspecific Variation in Pea Aphid (Acyrthosiphon pisum) Performance, Journal of Insect Physiology 40 (1994) 947-955.
- [94] P.J. Debarro, T.N. Sherratt, O. David, N. Maclean, An investigation of the differential performance of clones of the aphid Sitobion avenae on 2 host species, Oecologia 104 (1995) 379–385.
- [95] X.D. Liu, B.P. Zhai, X.X. Zhang, Specialized host-plant performance of the cotton aphid is altered by experience, Ecological Research 23 (2008) 919–925.
- [96] S. Via, Specialized Host Plant Performance of Pea Aphid Clones is not Altered by Experience, Ecology 72 (1991) 1420–1427.
- [97] J. Ferrari, S. Via, H.C.J. Godfray, Population differentiation and genetic variation in performance on eight hosts in the pea aphid complex, Evolution 62 (2008) 2508–2524.
- [98] D. Schluter, G.L. Conte, Genetics and ecological speciation, in: Proceedings of the National Academy of Sciences of the United States of America, 106, 2009, pp. 9955–9962.
- [99] F.P. Müller, Isolationmechanismen zwischen sympatrischen bionomischen Rassen am Beispiel der Erbsenblattlaus Acyrthosiphon pisum (Harris) (Homoptera, Aphididae), Zoologisches Jahrbuch für Systematik 98 (1971) 131–152.
- [100] A. Mackenzie, A trade-off for host plant utilization in the black bean aphid, Aphis fabae, Evolution 50 (1996) 155–162.
- [101] P.A. Johnson, F.C. Hoppensteadt, J.J. Smith, G.L. Bush, Conditions for sympatric speciation: a diploid model incorporating habitat fidelity and non-habitat assortative mating, Evolutionary Ecology 10 (1996) 187, 205
- [102] R. Rakauskas, Hybridisation between Aphis grossulariae and Aphis schneideri (Sternorrhyncha: Aphididae): an experimental approach, European Journal of Entomology 96 (1999) 401–408.
- [103] S. Akimoto, Local adaptation and host race formation of a gall-forming aphid in relation to environmental heterogeneity, Oecologia 83 (1990) 162–170.
- [104] T. Komatsu, S. Akimoto, Genetic differentiation as a result of adaptation to the phenologies of individual host trees in the galling aphid Kaltenbachiella japonica, Ecological Entomology 20 (1995) 33– 42.
- [105] S. Komazaki, Difference of egg diapause in two host races of the spirea aphid, *Aphis spiraecola*, Entomologia Experimentalis et Applicata 89 (1998) 201–205.
- [106] S. Komazaki, The inheritance of egg hatching time of the spirea aphid Aphis citricola homoptera aphididae on the two winter hosts, Kontyu 54 (1986) 48–53.
- [107] S. Komazaki, Studies on the biology of the spirea aphid, *Aphis spiraecola* Patch, with special reference to biotypic differences, Bulletin of the Fruit Tree Research Station (1991), iii + 60 pp.
- [108] V.F. Eastop, Keys for the identification of Acyrthosiphon (Hemiptera: Aphididae), Bulletin of the British Museum (Natural History) Entomology 26 (1971).

- [109] N.A. Moran, Symbiosis as an adaptive process and source of phenotypic complexity, in: Proceedings of the National Academy of Sciences of the United States of America, 104, 2007, pp. 8627–8633.
- [110] E.M. Janson, J.O. Stireman, M.S. Singer, P. Abbot, Phytophagous insectmicrobe mutualisms and adaptive evolutionary diversification, Evolution 62 (2008) 997–1012.
- [111] A.E. Douglas, Nutritional interactions in insect-microbial symbioses: aphids and their symbiotic bacteria *Buchnera*, Annual Review of Entomology 43 (1998) 17–37.
- [112] J.P. Sandstrom, J.A. Russell, J.P. White, N.A. Moran, Independent origins and horizontal transfer of bacterial symbionts of aphids, Molecular Ecology 10 (2001) 217–228.
- [113] T. Tsuchida, R. Koga, T. Fukatsu, Host plant specialization governed by facultative symbiont, Science 303 (2004) 1989.
- [114] J. Ferrari, C.L. Scarborough, H.C.J. Godfray, Genetic variation in the effect of a facultative symbiont on host-plant use by pea aphids, Oecologia 153 (2007) 323–329.
- [115] T.E. Leonardo, Removal of a specialization-associated symbiont does not affect aphid fitness, Ecology Letters 7 (2004) 461–468.
- [116] A.E. Douglas, C.L.M.J. François, L.B. Minto, Facultative "secondary" bacterial symbionts and the nutrition of the pea aphid, Acyrthosiphon pisum, Physiological Entomology 31 (2006) 262–269.
- [117] A. Frantz, V. Calcagno, L. Mieuzet, M. Plantégenest, J.-C. Simon, Complex trait differentiation between host-populations of the pea aphid Acyrthosiphon pisum (Harris): implications for the evolution of ecological specialisation, Biological Journal of the Linnean Society 97 (2009) 718–727.
- [118] R.R. Vargas, A.J. Troncoso, D.H. Tapia, R. Olivares-Donoso, H.M. Niemeyer, Behavioural differences during host selection between alate virginoparae of generalist and tobacco-specialist *Myzus persicae*, Entomologia Experimentalis et Applicata 116 (2005) 43–53.
- [119] M.C. Caillaud, S. Via, Specialized feeding behavior influences both ecological specialization and assortative mating in sympatric host races of pea aphids, American Naturalist 156 (2000) 606–621.
- [120] M.L. Del Campo, S. Via, M.C. Caillaud, Recognition of host-specific chemical stimulants in two sympatric host races of the pea aphid Acyrthosiphon pisum, Ecological Entomology 28 (2003) 405–412.
- [121] G. Gorur, C. Lomonaco, A. Mackenzie, Phenotypic plasticity in hostplant specialisation in *Aphis fabae*, Ecological Entomology 30 (2005) 657–664.
- [122] G. Powell, J. Hardie, The chemical ecology of aphid host alternation: how do return migrants find the primary host plant? Applied Entomology and Zoology 36 (2001) 259–267.
- [123] F.P. Müller, Wirtswahlversuche mit Gynoparen von Myzus persicae (Sulz.) und von Aphis fabae Scop, Zeitschrift für Pflanzenkrankheiten und Pflanzenschutz 65 (1958) 405–411.
- [124] J.P. Sandstrom, J. Pettersson, Winter host plant specialization in a host-alternating aphid, Journal of Insect Behavior 13 (2000) 815–825.
- [125] J.A. Coyne, H.A. Orr, Speciation, Sinauer, Sunderland, USA, 2004.

- [126] J. Ferrari, H.C.J. Godfray, A.S. Faulconbridge, K. Prior, S. Via, Population differentiation and genetic variation in host choice among pea aphids from eight host plant genera, Evolution 60 (2006) 1574–1584.
- [127] S. Via, Reproductive isolation between sympatric races of pea aphids. I. Gene flow restriction and habitat choice, Evolution 53 (1999) 1446–1457
- [128] J. Sandström, Temporal changes in host adaptation in the pea aphid, *Acyrthosiphon pisum*, Ecological Entomology 21 (1996) 56–62.
- [129] F.P. Müller, Wirtspflanzen, Generationenfolge und reproduktive Isolation infraspezifischer Formen von Acyrthosiphon pisum, Entomologia Experimentalis et Applicata 28 (1980) 145–157.
- [130] M.R. Servedio, M.A.F. Noor, The role of reinforcement in speciation: theory and data, Annual Review of Ecology, Evolution and Systematics 34 (2003) 339–364.
- [131] J.A. Guldemond, A.F.G. Dixon, Specificity and daily cycle of release of sex-pheromones in aphids-a case of reinforcement, Biological Journal of the Linnean Society 52 (1994) 287–303.
- [132] J.A. Guldemond, W.T. Tigges, P.W.F. Devrijer, Circadian-rhythm of sexpheromone production and male activity of coexisting sibling species of *Cryptomyzus* aphids (homoptera, aphididae), European Journal of Entomology 91 (1994) 85–89.
- [133] G.K. Shaposhnikov, Origin and breakdown of reproductive isolation and the criterion of the species, Entomological Review 45 (1966) 1–
- [134] G.K. Shaposhnikov, Morphological divergence and convergence in an experiment with aphids (Homoptera, Aphidinea), Entomological Review 44 (1965) 1–12.
- [135] G.K. Shaposhnikov, Host specificity and adaptation to new hosts in aphids (Homoptera, Aphidoidea) in the process of natural selection (experimental research), Entomological Review 40 (1961) 412– 426.
- [136] S. Via, Sympatric speciation in animals: the ugly duckling grows up, Trends in Ecology and Evolution 16 (2001) 381–390.
- [137] J. Peccoud, J.-C. Simon, H.J. McLaughlin, N.A. Moran, Post-Pleistocene radiation of the pea aphid complex revealed by rapidly evolving endosymbionts, in: Proceedings of the National Academy of Sciences of the United States of America, 106, 2009, pp. 16315–16320.
- [138] D.J. Funk, K.E. Filchak, J.L. Feder, Herbivorous insects: model systems for the comparative study of speciation ecology, Genetica 116 (2002) 251–267.
- [139] D.J. Funk, P. Nosil, W.J. Etges, Ecological divergence exhibits consistently positive associations with reproductive isolation across disparate taxa, in: Proceedings of the National Academy of Sciences, USA, 103, (2006), pp. 3209–3213.
- [140] S. Via, J. West, The genetic mosaic suggests a new role for hitchhiking in ecological speciation, Molecular Ecology 17 (2008) 4334–4345.
- [141] B.B. Normark, Molecular systematics and evolution of the Aphid family Lachnidae, Molecular Phylogenetics and Evolution 14 (2000) 131–140.