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No association between use of tenofovir disoproxil fumarate, etravirine, or integrase-strand transfer inhibitors and acquisition or severe outcomes of SARS-CoV-2 infection in people with HIV in the Netherlands

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In two Dutch observational cohorts of people with HIV, the use of TDF, ETR, or INSTIs was not independently associated with either the risk of incident SARS-CoV-2 infection or severe COVID-19 outcomes, as was suggested by previous observational and molecular docking studies. Our findings do not support a strategy of modifying antiretroviral therapy to include these agents to protect against SARS-CoV-2 infection and severe COVID-19 outcomes.

Since the start of the COVID-19 pandemic, several studies have tried to determine factors associated with acquisition of and clinical outcome of SARS-CoV-2 infection in people with HIV (PWH). Recent observational studies have suggested a protective effect of tenofovir disoproxil fumarate (TDF) against acquisition of SARS-CoV-2 [1,2] and severe COVID-19 outcomes [1,3,4], whereas other studies found no benefit of TDF or tenofovir alafenamide (TAF) in PWH [5,6] or adults without HIV [7,8].

Etravirine (ETR) and the integrase-strand transfer inhibitors (INSTIs) – specifically raltegravir (RAL) and dolutegravir (DTG) – were proposed as potential inhibitors of two major SARS-CoV-2 proteins in a molecular docking [9] and molecular dynamics simulation study [10]. One recent study showed that in-vitro docking by SARS-CoV-2 to the ACE2 receptor is inhibited by DTG and ETR [11]. Thus far, no studies in PWH have reported epidemiological evidence for a protective effect of the use of INSTIs or ETR against acquiring SARS-CoV-2 infection and severe COVID-19 outcomes.

We investigated the association between the above-mentioned antiretrovirals and incident SARS-CoV-2 infection and COVID-19-associated hospitalization and/or death in two Dutch observational cohorts of PWH.

First, we used data from the COVID-19 substudy of the AGE_hIV cohort collected from September 2020 until

April 2021 [12]. PWH and participants without HIV were assessed every 6 months for incident SARS-CoV-2 infection. Incident SARS-CoV-2 infection was defined as positive combined IgA/IgM/IgG SARS-CoV-2 nucleocapsid (N) antibody assay or a self-reported positive PCR test in participants without detectable N-antibodies. We previously reported that younger age and sub-Saharan African origin, but not HIV-status, were independently associated with higher risk of incident SARS-CoV-2 infection. However, we did not investigate the association with specific antiretrovirals in PWH.

Second, we used data from the Dutch national observational HIV cohort (ATHENA), containing data of more than 95% of PWH in care in one of the 24 HIV-treatment centers in the Netherlands [13]. Within this cohort, we recently reported that the risk of severe COVID-19 outcomes was increased in individuals with uncontrolled HIV replication, low CD4⁺ cell count and prior AIDS, independently of general risk factors such as age, comorbidity burden, and non-Western origin (F.W.N.M. Wit, P. Reiss, B. Rijnders, M. van der Valk, in preparation), but potential associations with specific antiretrovirals were not extensively analyzed.

Extending our earlier analyses, we now assessed whether use of TDF, ETR, and INSTIs were associated with the risk of incident SARS-CoV-2 infection in the AGE_hIV COVID-19 substudy and the risk of COVID-19-associated hospitalization and/or death in the ATHENA cohort. Associations between the aforementioned antiretrovirals and outcomes were assessed using multivariable logistic regression, both unadjusted and adjusted for age, sex at birth, ethnic origin, total comorbidity count (cardiovascular disease; non-AIDS-defining cancer; chronic kidney disease; diabetes mellitus; hypertension; and/or obesity (BMI ≥ 30 kg/m²), prior AIDS, current CD4⁺ cell count (categorized <200 , 200–499, ≥ 500 cells/ μ l), and current HIV-1 viral load (categorized ≤ 200 or >200 copies/ml).

In the current analysis, 239 PWH using antiretroviral therapy (ART) from the AGE_hIV COVID-19 substudy were included. Median age was 62.0 years [interquartile range (IQR) 57.7–67.2], 92.1% were men, and participants were living with HIV for a median of 21.6 years (IQR 15.2–27.2). All participants, except one (99.6%), were virally suppressed (<200 copies/ml) with a current CD4⁺ cell count of 670 cells/ μ l (IQR 530–834). By April 2021, 29 of 239 PWH had an incident SARS-CoV-2 infection. Those with an incident infection were significantly younger and more often of sub-Saharan

Table 1. Association between antiretroviral agents and incident SARS-CoV-2 infection in 239 AGE_{IV} COVID-19 substudy participants with known antiretroviral therapy regimen and association between antiretrovirals and severe COVID-19 (hospitalization and/or death) in 2189 ATHENA cohort participants.

	Association between antiretrovirals and incident SARS-CoV-2 infection (AGE _{IV} COVID-19 substudy)				Association between antiretrovirals and severe COVID-19 (hospitalization and/or death) (ATHENA cohort)			
	Unadjusted		Adjusted ^a		Unadjusted		Adjusted ^a	
	n/N ^b	OR (95% CI)	P	OR (95% CI)	P	n/N ^b	OR (95% CI)	P
NRTI backbone								
- Lamivudine only	0/7	-	-	-	-	8/157	0.83 (0.38–1.81)	0.63
- Abacavir-based	2/28	0.65 (0.13–3.28)	0.61	0.56 (0.11–2.95)	0.50	35/339	1.77 (1.10–2.86)	0.018
- Tenofovir alafenamide-based	17/107	1.61 (0.65–3.94)	0.30	1.33 (0.53–3.38)	0.55	67/946	1.18 (0.78–1.77)	0.43
- Tenofovir disoproxil fumarate-based	8/76	REF	REF	REF	REF	39/640	REF	REF
- No NRTI	2/21	0.89 (0.18–4.57)	0.89	0.96 (0.18–5.17)	0.96	9/107	1.42 (0.67–3.01)	.37
INSTI								
- Bictegravir	6/30	2.00 (0.55–7.27)	0.29	2.15 (0.56–8.26)	0.26	21/282	0.93 (0.55–1.59)	0.80
- Dolutegravir	5/45	REF	REF	REF	REF	50/626	REF	REF
- Elvitegravir/cobicistat	6/41	1.37 (0.38–4.89)	.63	0.77 (0.19–3.14)	.72	15/349	0.52 (0.29–0.94)	.029
- Raltegravir	1/12	0.73 (0.08–6.89)	0.78	0.97 (0.10–9.65)	0.98	7/51	1.83 (0.79–4.28)	.16
- No INSTI	11/111	0.88 (0.29–2.69)	0.82	0.83 (0.26–2.64)	0.75	65/881	0.92 (0.63–1.35)	0.66
NNRTI								
- Doravirine	2/11	2.00 (0.24–16.61)	0.52	2.59 (0.27–25.02)	0.41	6/140	0.49 (0.18–1.33)	0.16
- Efavirenz	2/20	REF	REF	REF	REF	12/142	REF	REF
- Etravirine	0/1	-	-	-	-	1/11	1.08 (0.13–9.20)	0.94
- Nevirapine	4/49	0.80 (0.13–4.76)	0.81	0.74 (0.11–5.02)	0.76	18/227	0.93 (0.44–2.00)	0.86
- Rilpivirine	2/13	1.64 (0.20–13.34)	0.65	2.22 (0.24–20.23)	0.48	11/172	0.74 (0.32–1.73)	0.49
- No NNRTI	19/145	1.36 (0.29–6.32)	0.70	1.15 (0.22–6.03)	0.87	110/1497	0.86 (0.46–1.60)	0.64
PI								
- Atazanavir/booster	1/5	5.25 (0.39–71.42)	0.21	9.72 (0.61–155.06)	0.11	2/22	0.72 (0.16–3.14)	0.67
- Darunavir/booster	2/44	REF	REF	REF	REF	33/270	REF	REF
- Lopinavir/booster	0/2	-	-	-	-	0/5	-	-
- No PI	26/188	3.37 (0.77–14.77)	0.11	3.51 (0.78–15.92)	.10	123/1892	0.50 (0.33–0.75)	<0.001

Values represent odds ratios (OR) with 95% confidence interval using logistic regression.

^aAdjusted for age, sex at birth, ethnic origin, total comorbidities count, prior AIDS, current CD4⁺ cell count, and current HIV-1 viral load.

^bNumber of participants with severe COVID-19 (hospitalization and/or death) per total number of participants for each variable category.

CI, confidence interval; INSTI, integrase strand transfer inhibitors; NNRTI, nonnucleoside reverse transcriptase inhibitors; NRTI, nucleoside reverse transcriptase inhibitors; OR, odds ratio; P, P-value; PI, protease inhibitors; REF, reference group.

African origin compared with those without incident SARS-CoV-2 infection (data not shown). ART regimen did not differ significantly between both groups.

For the ATHENA cohort, 2189 PWH using ART while being diagnosed with COVID-19 were included. Median age was 50.1 years (IQR 40.5–57.7), 80.3% were men, and participants were living with HIV for a median of 12.5 years (IQR 7.2–18.3). About 98.3% had an HIV-1 viral load less than 200 copies/ml and current CD4⁺ cell count of 710 cells/ μ l (IQR 530–900). One hundred fifty-eight participants had severe COVID-19: hospitalization ($n=149$) and/or death ($n=29$; 9/29 deaths occurred in individuals who were not hospitalized). Compared with those with mild COVID-19, participants with severe COVID-19 were significantly older, more often of non-white origin, had more comorbidities, and were living with HIV for more years. Moreover, those with severe COVID-19 had lower median nadir CD4⁺ cell count [153 (IQR 60–262) vs. 260 (130–390) cells/ μ l, $P<0.001$] and lower median current CD4⁺ cell count [593 (IQR 403–830) vs. 720 (IQR 540–910) cells/ μ l, $P<0.001$]. Those with severe COVID-19 outcomes were more often on a protease inhibitor-containing ART-regimen (22.2 vs. 12.9%), but otherwise ART-regimen did not differ significantly between the groups.

Use of TDF, ETR, or INSTIs was not significantly associated with risk of incident SARS-CoV-2 infection, or severe COVID-19 (Table 1) in both unadjusted and adjusted analyses. Moreover, no associations between use of other antiretrovirals and incident SARS-CoV-2 infection or severe COVID-19 were observed.

Our analyses in two cohorts of PWH support previous findings that TDF does not protect against SARS-CoV-2 infection or severe COVID-19 outcomes [5,6]. Similar to our analyses, these studies adjusted their outcome for baseline participant characteristics, such as country of origin, socioeconomic status, current CD4⁺ cell count and CD4/CD8 ratio, years on ART, presence of diabetes, chronic kidney disease, and metabolic disease [5]. Importantly, studies suggesting a protective effect of TDF in PWH did not adjust for these participant characteristics [2–4]. It is likely that the presence of risk factors such as higher age and comorbidities influence the choice of ART regimen, which may confound the association between ART use and COVID-19 outcomes, thereby explaining the difference in study findings.

With regard to INSTIs, two clinical studies likewise found no protective effect of INSTIs against acquisition of SARS-CoV-2 infection in PWH [2,6].

In conclusion, in two Dutch observational cohorts of PWH, the use of TDF, ETR, or INSTIs was not independently associated with a reduced risk of incident

SARS-CoV-2 infection or severe COVID-19 outcomes. Our findings do not support a strategy of modifying ART to include these antiretrovirals to protect against SARS-CoV-2 infection and severe COVID-19.

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Informed consent was obtained from all participants. The AGE_hIV cohort study was approved by the ethics committee of the Amsterdam UMC, location AMC, and is registered at www.clinicaltrials.gov (NCT01466582). The ATHENA cohort was approved by the institutional review boards of all participating HIV treatment centers.

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References

- Li G, Park LS, Lodi S, Logan RW, Cartwright EJ, Barakat LA, et al. **Tenofovir disoproxil fumarate and COVID-19 outcomes in men with HIV.** *AIDS* 2022; **36**:1689–1696.
- Berenguer J, Diez C, Martin-Vicente M, Mican R, Perez-Elias MJ, Garcia-Fraile LJ, et al. **Prevalence and factors associated with SARS-CoV-2 seropositivity in the Spanish HIV Research Network Cohort.** *Clin Microbiol Infect* 2021; **27**:1678–1684.
- Del Amo J, Polo R, Moreno S, Diaz A, Martinez E, Arribas JR, et al. **Incidence and severity of COVID-19 in HIV-positive persons receiving antiretroviral therapy: a cohort study.** *Ann Intern Med* 2020; **173**:536–541.
- Western Cape Department of Health in collaboration with the National Institute for Communicable Diseases SA. **Risk factors for coronavirus disease 2019 (COVID-19) death in a population cohort study from the Western Cape Province, South Africa.** *Clin Infect Dis* 2021; **73**:e2005–e2015.
- Nomah DK, Reyes-Uruena J, Diaz Y, Moreno S, Aceiton J, Bruguera A, et al. **Impact of tenofovir on SARS-CoV-2 infection and severe outcomes among people living with HIV: a propensity score-matched study.** *J Antimicrob Chemother* 2022; **77**:2265–2273.
- De Lazzari E, Blanco J, Rico N, Filella X, Egri N, Ruiz R, et al. **MO43 Prevalence, risk factors and the impact of antiretroviral treatment in SARS-CoV-2 infection in people with HIV: a cross-sectional study.** *J Int AIDS Soc* 2022; **25** (Suppl 6): e26009.
- Montejano R, de la Calle-Prieto F, Velasco M, Guijarro C, Queiruga-Parada J, Jimenez-Gonzalez M, et al. **Tenofovir disoproxil fumarate/emtricitabine and baricitinib for patients at high risk of severe COVID-19: the PANCOVID Randomized Clinical Trial.** *Clin Infect Dis* 2023; **76**:e116–e125.
- Ayerdi O, Puerta T, Clavo P, Vera M, Ballesteros J, Fuentes ME, et al. **Preventive efficacy of tenofovir/emtricitabine against severe acute respiratory syndrome coronavirus 2 among pre-exposure prophylaxis users.** *Open Forum Infect Dis* 2020; **7**: ofaa455.
- Indu P, Rameshkumar MR, Arunagirinathan N, Al-Dhabi NA, Valan Arasu M, Ignacimuthu S. **Raltegravir, Indinavir, Tipranavir, Dolutegravir, and Etravirine against main protease and RNA-dependent RNA polymerase of SARS-CoV-2: a molecular docking and drug repurposing approach.** *J Infect Public Health* 2020; **13**:1856–1861.
- Khan RJ, Jha RK, Amara GM, Jain M, Singh E, Pathak A, et al. **Targeting SARS-CoV-2: a systematic drug repurposing approach to identify promising inhibitors against 3C-like proteinase and 2'-O-ribose methyltransferase.** *J Biomol Struct Dyn* 2021; **39**:2679–2692.
- Lee RK, Li TN, Chang SY, Chao TL, Kuo CH, Pan MY, et al. **Identification of entry inhibitors against Delta and Omicron variants of SARS-CoV-2.** *Int J Mol Sci* 2022; **23**:4050.
- Verburgh ML, Boyd A, Wit FWNM, Schim van der Loeff MF, van der Valk M, Bakker M, et al. **Similar risk of severe acute respiratory syndrome Coronavirus 2 Infection and similar nucleocapsid antibody levels in people with well controlled human immunodeficiency virus (HIV) and a comparable cohort of people without HIV.** *J Infect Dis* 2022; **225**:1937–1947.
- Boender TS, Smit C, Sighem AV, Bezemer D, Ester CJ, Zaheri S, et al. **AIDS Therapy Evaluation in the Netherlands (ATHENA) national observational HIV cohort: cohort profile.** *BMJ Open* 2018; **8**:e022516.

A moving target: impacts of lowering viral load suppression cutpoints on progress towards HIV epidemic control goals

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Redefining viral load suppression (VLS) using lower cutpoints could impact progress towards the United Nations Programme on HIV/AIDS 95–95–95 targets. We assessed impacts of lowering the VLS cutpoint on achieving the ‘third 95’ in the Rakai Community Cohort Study. Population VLS would fall from 86% to 84% and 76%, respectively, after lowering VLS cutpoints from <1000 to <200 and <50 copies/ml. The fraction of viremic persons increased by 17% after lowering the VLS cutpoint from <1000 to <200 copies/ml.

The Joint United Nations Programme on HIV/AIDS (UNAIDS) 95–95–95 targets aim to achieve HIV elimination by 2030 and optimize health outcomes among persons with HIV [1]. To reach the ‘third 95’, 86% of people living with HIV [95% of whom are receiving antiretroviral therapy (ART)] must achieve viral load suppression (VLS). The World Health Organization (WHO) defines VLS as <1000 RNA copies/ml [2], a threshold associated with reduced HIV transmission risk [3,4]. This definition is also used by national surveys in Africa, including the population-based HIV impact assessments [5].

A VLS cutpoint of <1000 copies/ml, however, potentially underestimates the proportion of individuals on ART experiencing negative consequences of viremia. A study in Lesotho estimated that 94% of treatment-experienced persons with viral loads 80–999 copies/ml harbored drug-resistant mutations [6]. Likewise, persistent low-level viremia (50–999 copies/ml over >6 months) has been linked to residual inflammation and subsequent virologic failure [7–9]. These findings, coupled with rising levels of HIV-1 drug resistance across Africa, have prompted calls to redefine VLS using lower cutpoints [10,11].

In addition to identifying persistent low-level viremia of potential clinical significance, lowering VLS cutpoints will also result in the detection and escalated management of people experiencing clinically insignificant transient viremia, or viral ‘blips’. Emerging evidence also suggests viremic blips of low magnitude (<500 copies/ml) unassociated with subsequent virologic failure are common in persons with prolonged ART use [12].