



Evaluation of Structural Flexibility and Cross-Reactivity of the T-Cell Receptor

Margaret He^{1,2}, Xianli Jiang, PhD², Kun Hee Kim^{2,3}, Ken Chen, PhD^{1,2,3}

¹ King Foundation High School Summer Program, School of Health Professions; ² Department of Bioinformatics and Computational Biology, UT MD Anderson Cancer Center; ³ UTMDACC UTHealth Graduate School of Biomedical Sciences, Houston, TX

Background

- T-cells have become of interest for cancer therapies due to their ability to recognize and engage with tumor antigens.^[1] T-cells receptors (TCR) interact with peptide-major histocompatibility complexes (pMHC) to trigger a response.^[2]
- During an immune response, T-cells proliferate until the pathogen is cleared, and a portion of them will develop an antigen-specific memory, allowing a stronger and faster immune response in an event of reactivation.^[3]
- There are two parts to each TCR chain (α and β): the variable and the constant region. Within each variable region, there are three complementarity-determining regions (CDR1, CDR2, and CDR3).^[2]
- While CDR1 and CDR2 more often interact with MHC helices, CDR3, the most structurally diverse of all loops, engages with peptides to a great extent.^[2] To advance therapeutic interventions, a greater understanding needs to be reached through the evaluation of structural flexibility and cross-reactivity of TCR.

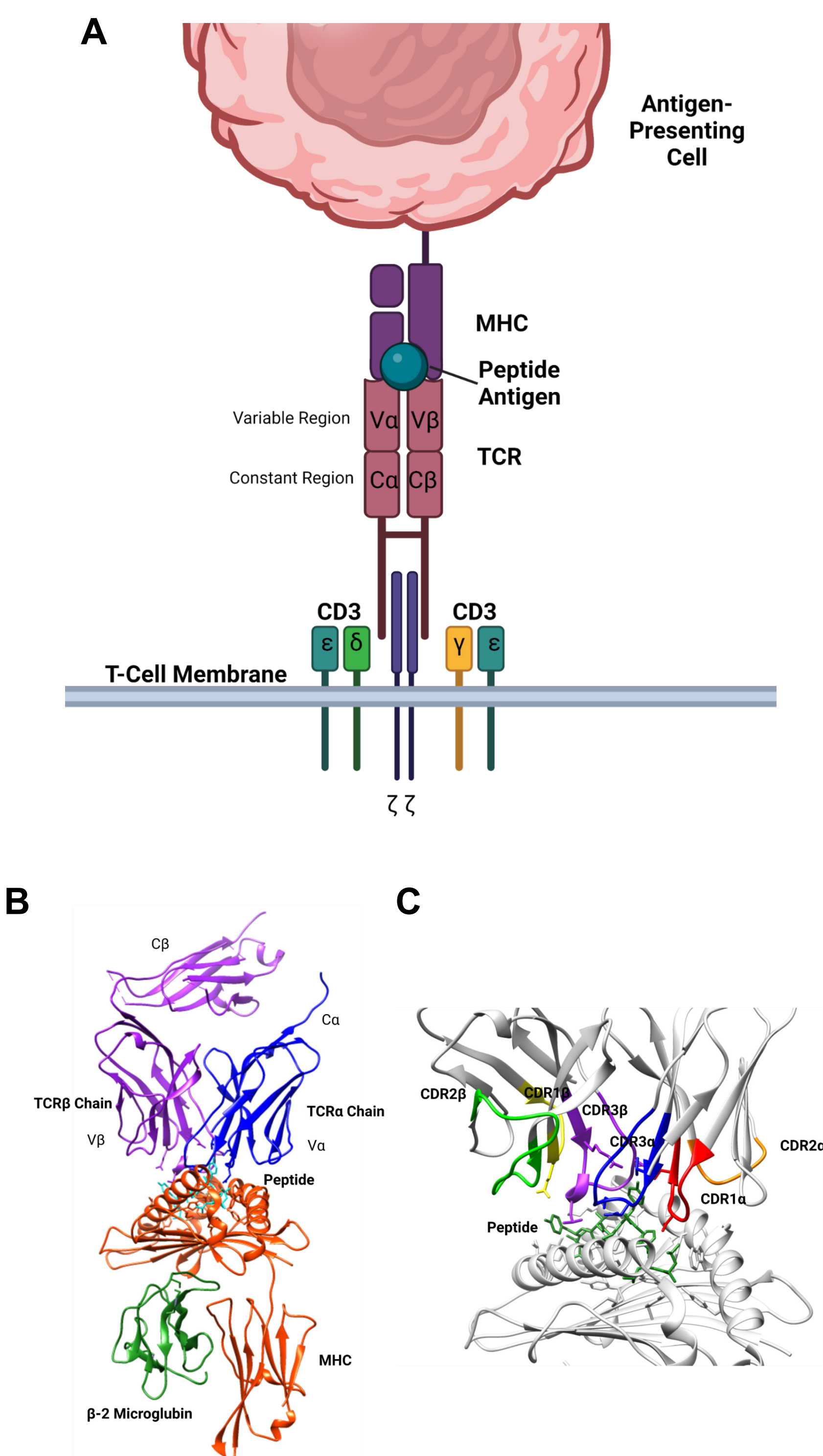


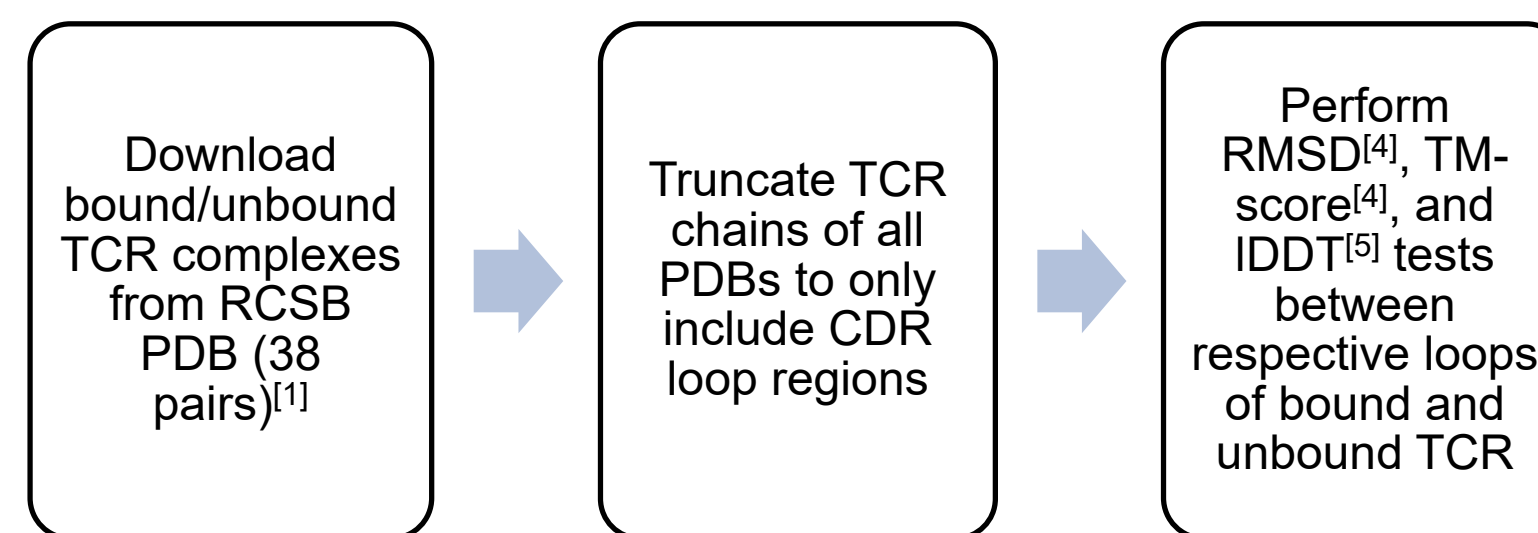
Figure 1: Structures of the TCR-pMHC complex.
(A) TCR chains engage with peptide presented by MHC.
(B) Structures as modeled by bound A6 TCR 1A07.
(C) CDR loops of bound A6 TCR 1A07.

Objective

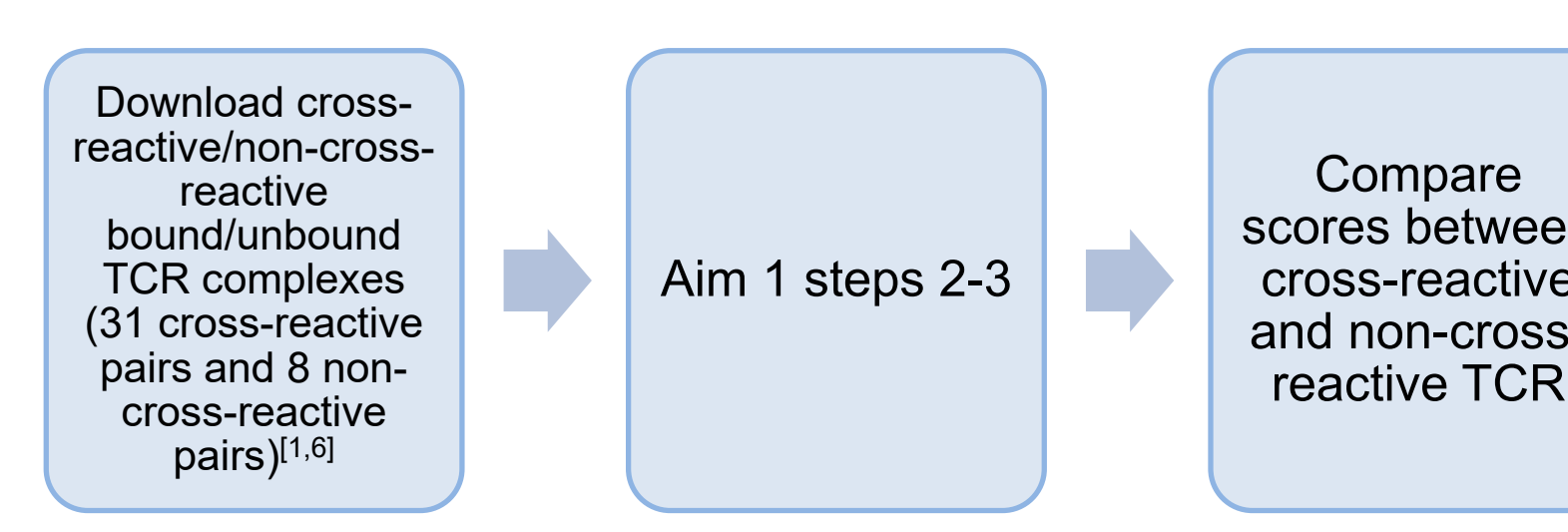
- Explore TCR structural flexibility upon binding to peptide presented by MHC.
- Evaluate role of TCR structural flexibility in cross-reactivity.

Methods

Aim 1: Structural Flexibility



Aim 2: Cross-Reactivity



Results

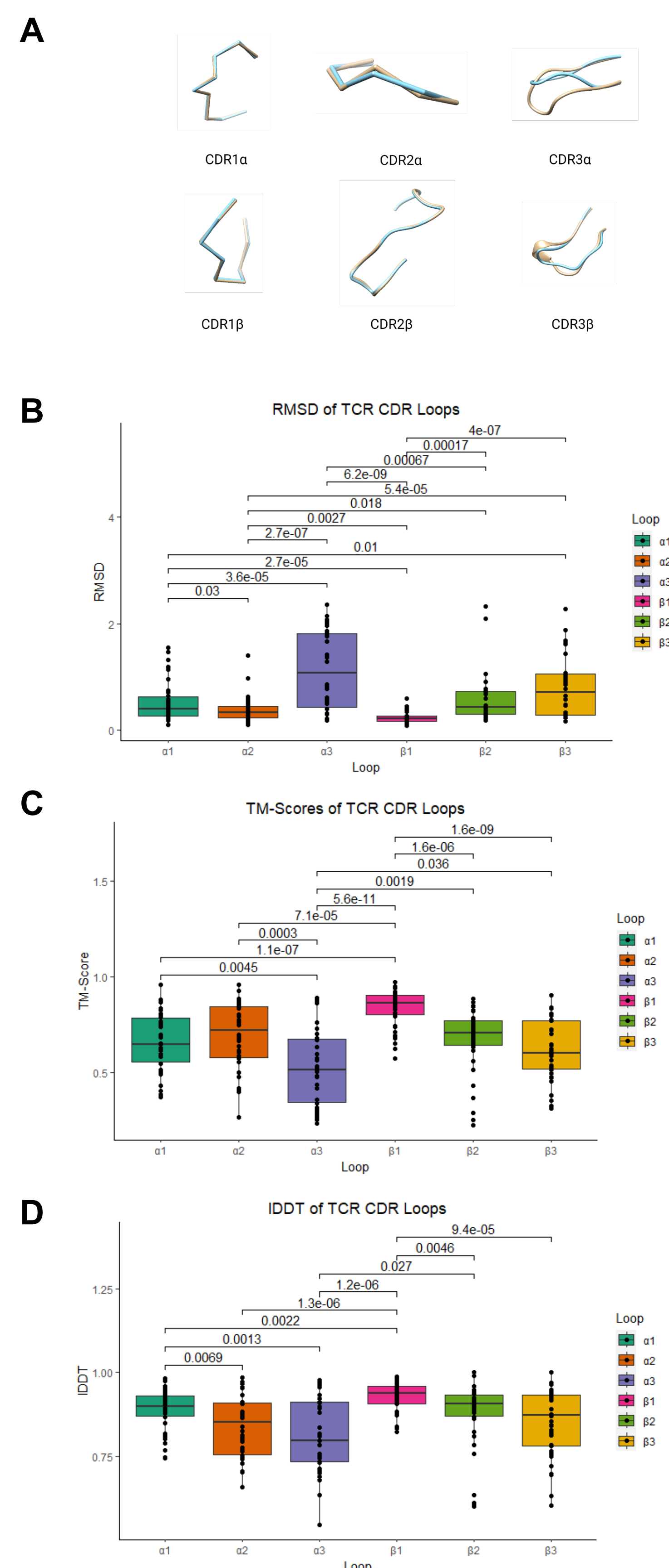


Figure 2: Results for Aim 1: Structural Flexibility.
(A) The backbone of CDR loops adopts different conformations between bound and unbound states (gold: 1A07 for A6 TCR bound to pMHC; blue: 3QH3 for unbound A6 TCR).
(B) Greatest RMSD range (Å) in CDR3 α/β loops.
(C) Lowest TM-score range in CDR3 α/β loops.
(D) Lowest IDDT range in CDR3 α/β loops.

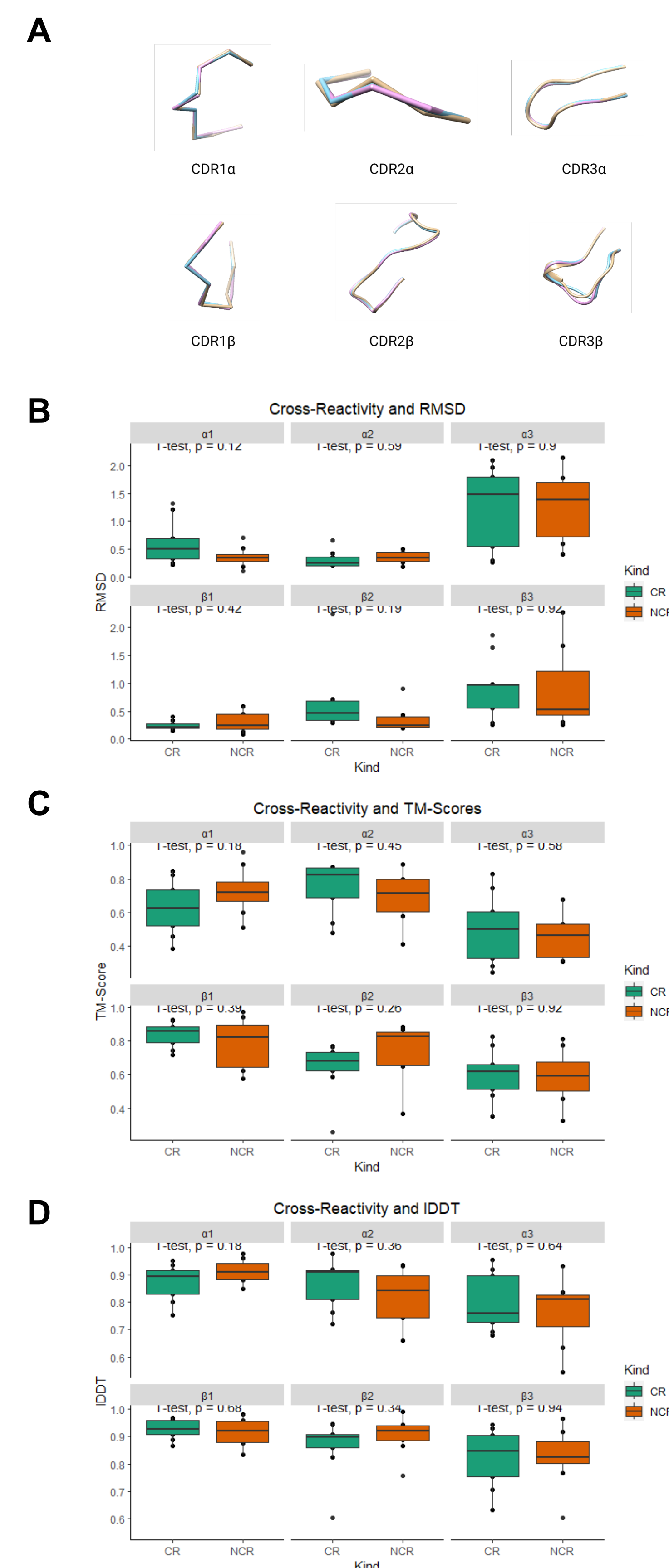


Figure 3: Results for Aim 2: Cross-Reactivity.
(A) The backbone of CDR loops adopts different conformations among three A6 TCR bound to different peptides (gold: 1A07 bound to LLFGYPVYV; blue: 3H9S bound to MLWGYLQYV; pink: 3PWP bound to LGYGFVNYI).
(B) Greatest RMSD range (Å) in cross-reactive CDR3 α/β loops.
(C) Lowest TM-score range in cross-reactive CDR3 α/β loops.
(D) Lowest IDDT range in cross-reactive CDR3 α/β loops.

Definitions

LYRA 1.0^[7]- used to determine CDR regions through input of TCR chain sequences

UCSF Chimera^[8]- used to visualize structural differences through alignment function

RMSD- root-mean-square deviation, measures the average distance between corresponding atoms
Higher value indicates greater structural difference

TM-score- template modeling score, measures the alignment and structural similarity between two protein structures
Lower value indicates greater structural difference

IDDT- Local Distance Difference Test, measures the distance differences between atom pairs in model and reference structures
Lower value indicates greater structural difference

Conclusions

- Aim 1:** CDR3 α/β loops display a greater degree of structural flexibility than other CDR loops.
- Aim 2:** CDR3 α/β loop-mediated structural flexibility is found in cross-reactivity.

Future Directions

- Increase the number of TCR complexes studied for more definitive conclusions.
- Implement more structural difference methods.^[9]
- Evaluate the role of MHC flexibility in cross-reactivity.

References

- Waldman, et al. A guide to cancer immunotherapy: from T cell basic science to clinical practice. *Nature Reviews Immunology*, 2020, 20:651-668. <https://doi.org/10.1038/s41577-020-0306-5>
- Wong, et al. Comparative Analysis of the CDR Loops of Antigen Receptors. *Front. Immunol.*, 2019, 10. <https://doi.org/10.3389/fimmu.2019.02454>
- Mangani, et al. Learning from the nexus of autoimmunity and cancer. *Immunity*, 2023, 56(2):256-271. <https://doi.org/10.1016/j.immuni.2023.01.022>
- Zhang & Skolnick. TM-align: A local superposition-free score for comparing protein structures and models using distance difference tests. *Nucleic Acids Research*, 2005, 33(7):2302-2309. <https://doi.org/10.1093/nar/gki524>
- Mariani, et al. IDDT: a local superposition-free score for comparing protein structures and models using distance difference tests. *Bioinformatics*, 2013, 29(21):2722-2728. <https://doi.org/10.1093/bioinformatics/btt473>
- Armstrong, et al. Conformational changes and flexibility in T-cell receptor recognition of peptide-MHC complexes. *Biochem J.*, 2008, 415(2):183-196. <https://doi.org/10.1042/BJ20080850>
- Klausen, et al. LYRA, a webserver for lymphocyte receptor structural modeling. *Nucleic Acids Research*, 2015, 43(W1):W349-W355. <https://doi.org/10.1093/nar/gkv535>
- Petterson, et al. UCSF Chimera—a visualization system for exploratory research and analysis. *Journal of Computational Chemistry*, 2004, 25(13):1605-1612. <https://doi.org/10.1002/jcc.20084>
- Olechovic, et al. Comparative analysis of methods for evaluation of protein models against native structures. *Bioinformatics*, 2019, 35(6):937-944. <https://doi.org/10.1093/bioinformatics/bty760>

Acknowledgments

Special thanks to the King Foundation and UTMDACC SHP for their support. Diagrams created with Biorender.com. Molecular graphics generated with UCSF Chimera.