



## Effect of vegetable seed oils on methane emission and fermentation of feed *in vitro*\*

CHETNA MAHAJAN<sup>1</sup>, KAJAL MANGAL<sup>2</sup> and M WADHWA<sup>3</sup>

Guru Angad Dev Veterinary and Animal Sciences University, Ludhiana, Punjab 141 004 India

Received: 11 July 2015; Accepted: 12 November 2015

**Key words:** Alternate hydrogen sink, Methane mitigation *in-vitro*, Vegetable seed oils

In ruminants, the vast majority of enteric methane ( $\text{CH}_4$ ) production occurs in the reticulo-rumen. Reducing  $\text{CH}_4$  production not only improves feed energy utilization, but also enhances the system efficiency. Mitigation of  $\text{CH}_4$  emissions is receiving great attention, especially through dietary manipulations. The amount of methane produced in the rumen is affected by feed intake, other dietary factors like dietary fiber, starch, soluble sugars, dietary lipids, level of feeding, roughage to concentrate ratio, type of forage, stage of maturity of forage, rate of passage of digesta, efficiency of feed conversion, processing and supplementation (Blaxter and Clapperton 1965, Benchaar *et al.* 2001, Mills *et al.* 2001, Bakshi and Wadhwa 2009), plant secondary metabolites (Bakshi and Wadhwa 2010, 2012), feed additives (Wadhwa and Bakshi 2009), phylogenetics (Bakshi and Wadhwa 2011) and ambient temperature (McAllister *et al.* 1996). Adding fat to the diet reduces  $\text{CH}_4$  emission by decreasing organic matter fermentation in the rumen, reducing the activity of methanogens and protozoal numbers, and for lipids rich in unsaturated fatty acids, through hydrogenation of fatty acids (Maia *et al.* 2007, Beauchemin *et al.* 2008) consuming hydrogen in the process, which makes this process a potential alternate hydrogen sink. Fats and oils change the fermentation process in the rumen, producing more propionic acid and less methane. Since the effect of vegetable oils depends on diet, nature and level of supplementation, the present study was therefore planned to evaluate the effect of supplementation of vegetable oils at different levels on methane production, fermentation pattern and gas production *in vitro*.

The rumen contents were collected from 3 rumen fistulated male buffaloes in a thermos flask flushed with  $\text{CO}_2$  and maintained at 39°C. The rumen contents were blended for 2–3 min in a blender and strained through 4-

layered muslin cloth and processed for *in vitro* gas production measurements as per standard procedure (Menke *et al.* 1979, Menke and Steingass 1988). About 375 mg of the ground sample of complete feed (oat fodder and concentrate mixture in 60: 40 ratio on DM basis) was incubated at 39°C for 24 h in triplicate in 100 ml calibrated glass syringes with buffered rumen fluid for assessing the net gas production, digestibility of nutrients, volatile fatty acids (VFAs) production and ME availability. The set was repeated twice.

Methane was estimated by using the equation based on VFA proportions (Wolin 1960). VFA were estimated by using gas chromatograph equipped with a glass column (6 ft length and 1/8 inch diameter) packed with chromosorb 101 (Cottoyn and Boucque 1968). Samples were analysed for proximate (AOAC 2000) and cell wall components (Robertson and Van Soest 1981). Data were analysed by using 3×4 factorial design (Snedecor and Cochran 1994) by using SPSS (2007) Version 16 and the differences in means by using Tukey's b test.

The complete feed contained 13.5% CP and 2.8% EE. The carrot seed oil, rape seed oil and canola seed oil contained 61%, 12%, 13%; 60%, 28%, 12%; 61%, 27.8% and 10.6% mono unsaturated fatty acids, poly unsaturated fatty acids and saturated fatty acids, respectively. Supplementing the complete feed with different vegetable seed oils, irrespective of the level indicated that the NDF digestibility was highest ( $P<0.05$ ) in rape seed oil as compared to carrot seed oil supplemented group. However, no difference was observed between rape seed and canola seed oil (Table 1). The other parameters like net gas production (NGP), true OM digestibility, partitioning factor and ME availability were comparable in all the vegetable oil supplemented groups. The fermentative  $\text{CO}_2$  and methane production from complete feed was observed to be lowest ( $P<0.05$ ) with rape seed oil supplementation.

The NGP, irrespective of source of oil, was highest ( $P<0.05$ ) in the control group as compared to oil supplemented groups, but decreased linearly with the increase in level of oil supplementation. The digestibility of nutrients decreased with increase in levels of supplementation and this effect was more pronounced at

\*Part of Ph.D. research.

Present Address: <sup>1</sup>Ph.D. Scholar (dr.chetnamahajan@gmail.com), Department of Veterinary Physiology and Biochemistry, <sup>2</sup>Senior Research Fellow (kajalmangal@hotmail.com), <sup>3</sup>Senior Animal Nutritionist-cum-Head (mw\_7in@yahoo.co.in), Department of Animal Nutrition.

Table 1. Effect of vegetable oils on net gas production, digestibility of nutrients and ME availability

Parameter	Vegetable seed oil			PSE	Level of oil, %			PSE
	Carrot	Rape	Canola		0	1	2	
NGP, ml/g DM/24h	182.89	182.56	183.33	0.37	180.67 <sup>A</sup>	185.56 <sup>C</sup>	183.33 <sup>B</sup>	182.15 <sup>B</sup>
NDFD, %	37.18 <sup>a</sup>	38.86 <sup>b</sup>	38.66 <sup>ab</sup>	0.19	45.54 <sup>C</sup>	36.57 <sup>B</sup>	34.63 <sup>A</sup>	36.20 <sup>B</sup>
TOMD, %	63.24	64.24	64.11	0.21	68.08 <sup>C</sup>	62.95 <sup>B</sup>	61.76 <sup>A</sup>	62.66 <sup>B</sup>
PF	1.98	1.98	1.98	0.01	2.01 <sup>C</sup>	1.94 <sup>A</sup>	1.97 <sup>AB</sup>	1.98 <sup>B</sup>
NH <sub>3</sub> , mg/dl	0.029	0.030	0.030	-	0.030 <sup>B</sup>	0.030 <sup>B</sup>	0.029 <sup>A</sup>	0.030 <sup>B</sup>
FCO <sub>2</sub> , mM	3.07 <sup>a</sup>	3.06 <sup>a</sup>	3.12 <sup>b</sup>	0.004	3.07 <sup>B</sup>	3.16 <sup>D</sup>	3.10 <sup>C</sup>	3.04 <sup>A</sup>
FCH <sub>4</sub> , mM	2.06 <sup>b</sup>	2.03 <sup>a</sup>	2.06 <sup>b</sup>	0.01	1.981 <sup>A</sup>	2.111 <sup>D</sup>	2.083 <sup>C</sup>	2.022 <sup>B</sup>
ME, MJ/kg DM	8.30	8.29	8.31	0.01	8.25 <sup>A</sup>	8.36 <sup>C</sup>	8.31 <sup>B</sup>	8.28 <sup>B</sup>

NGP, net gas production; NDFD, neutral detergent fiber digestibility; TOMD, true OM digestibility; PF, partitioning factor; FCO<sub>2</sub>, fermentative CO<sub>2</sub>; FCH<sub>4</sub>, fermentative methane; ME, metabolizable energy. Figures with different superscripts <sup>a,b,c</sup> for different oils and different superscripts <sup>A,B,C,D</sup> for different levels of oil in a row differ significantly, P<0.05.

Table 2. Effect of vegetable oils on *in-vitro* volatile fatty acid production (mM/dl) after 24 h incubation

Parameter	Vegetable seed oil			PSE	Level of oil, %			PSE
	Carrot	Rape	Canola		0	1	2	
TVFAs	5.78 <sup>a</sup>	5.92 <sup>b</sup>	6.0 <sup>c</sup>	0.01	5.87 <sup>B</sup>	6.04 <sup>D</sup>	5.94 <sup>C</sup>	5.76 <sup>A</sup>
Acetate (A)	4.06 <sup>a</sup>	4.10 <sup>b</sup>	4.12 <sup>b</sup>	0.01	3.98 <sup>A</sup>	4.21 <sup>B</sup>	4.18 <sup>B</sup>	4.00 <sup>A</sup>
Propionate (P)	0.95 <sup>a</sup>	1.07 <sup>b</sup>	1.09 <sup>b</sup>	0.007	1.11 <sup>C</sup>	1.04 <sup>B</sup>	1.02 <sup>B</sup>	0.97 <sup>A</sup>
Butyrate	0.54 <sup>b</sup>	0.50 <sup>a</sup>	0.54 <sup>b</sup>	0.002	0.53 <sup>B</sup>	0.53 <sup>B</sup>	0.50 <sup>A</sup>	0.53 <sup>B</sup>
Iso butyrate	0.036 <sup>a</sup>	0.067 <sup>b</sup>	0.069 <sup>b</sup>	0.063	0.067 <sup>D</sup>	0.060 <sup>C</sup>	0.049 <sup>A</sup>	0.054 <sup>B</sup>
Iso valerate	0.128 <sup>c</sup>	0.115 <sup>a</sup>	0.120 <sup>b</sup>	0.001	0.111 <sup>A</sup>	0.130 <sup>D</sup>	0.115 <sup>B</sup>	0.127 <sup>C</sup>
Valerate	0.064 <sup>a</sup>	0.076 <sup>c</sup>	0.071 <sup>b</sup>	0.07	0.062 <sup>A</sup>	0.072 <sup>B</sup>	0.075 <sup>B</sup>	0.072 <sup>B</sup>
A: P	4.31 <sup>b</sup>	3.84 <sup>a</sup>	3.79 <sup>a</sup>	0.03	3.59 <sup>A</sup>	4.07 <sup>B</sup>	4.09 <sup>B</sup>	4.16 <sup>B</sup>

TVFAs, total volatile fatty acids. Figures with different superscripts <sup>a,b,c</sup> for different oils and different superscripts <sup>A,B,C,D</sup> for different levels of oil in a row differ significantly, P<0.05.

2% level of supplementation. The partitioning factor decreased with increase in level of supplementation, in comparison to the unsupplemented feed. The fermentative methane production was higher (P<0.05) in oil supplemented than control group, but amongst the oil supplemented groups it was lowest (P<0.05) when oil was supplemented at 3% level. The availability of ME was improved (P<0.05) in all the oil supplemented groups as compared to control.

The VFAs, acetate, propionate and butyrate production, irrespective of level of vegetable oil was highest (P<0.05) in canola seed oil and lowest with carrot seed oil, except that of butyrate which was lowest in rape seed oil supplemented group, but A: P ratio followed the reverse trend (Table 2). TVFA and acetate levels were highest (P<0.05) at 1% level of supplementation, whereas propionate level was highest in control followed by 1% supplementation. The A: P ratio increased by 11.70% on an average compared to control. Machmüller *et al.* (2003) also reported that at supplementation of 50g/kg coconut oil, the molar proportions of propionate and isovalerate were reduced (P<0.01) and the molar proportion of butyrate (P<0.001) and A: P ratio were increased (P<0.05). On the contrary, Morales *et al.* (2012) obtained 28% decrease

methane in 24 h *in vitro* incubations of rumen digesta with added 0.2g ricinoleic acid/l. There was no effect on the TVFA after 24 h as a result of ricinoleic acid addition (castor oil), but the molar proportions of acetate and butyrate were decreased.

## SUMMARY

A study was conducted to assess the effect of carrot seed oil, canola seed oil and rape seed oil on rumen fermentation and methanogenesis *in-vitro*. The oils were supplemented to the complete feed (oat fodder and concentrate mixture in 60: 40 ratio) @ 1, 2 and 3% on DM basis, incubated for 24 h in 100 ml glass syringes. The digestibility of NDF varied significantly with highest in rapeseed and lowest in carrot seed, while OM digestibility and ME availability did not show any significant differences amongst the supplemented oils. The *in-vitro* methane production from complete feed supplemented with rape seed oil was observed to be the lowest. The TVFAs, acetate and propionate levels were highest in canola oil while A: P ratio was lowest.

TVFA and acetate levels were highest at 1% level of supplementation, whereas propionate level was highest in control followed by at 1% supplementation. The methane

production was significantly higher in oil supplemented groups as compared to control group, but amongst the oil supplemented groups it was significantly lowest when oil was supplemented at 3% level. Amongst the various oils evaluated for *in vitro* methane mitigation, the study conclusively revealed that the supplementation of diet with rape seed oil @ 2–3% on dry matter basis had an edge over other oils and levels.

#### REFERENCES

- AOAC. 2000. *Official Methods of Analysis*. 7<sup>th</sup> edn. Association of Analytical Chemists, Gaithersburg, Maryland, USA.
- Bakshi M P S and Wadhwa M. 2009. Dietary manipulation for mitigation of enteric methane emission. Lead paper 5<sup>th</sup> Asian Buffalo Conference, Lahore. Proceedings in *Pakistan Journal of Zoology* **9**: 887–93. (Environ-SPJZ-2).
- Bakshi M P S and Wadhwa M. 2010. Significance of plant secondary metabolites in relation to methane produced by ruminants. *Proceedings in Animal Nutrition Association Conference*, held at NAASC complex, New Delhi. Vol I (Lead papers). pp 25–28.
- Bakshi M P S and Wadhwa M. 2011. Phylogenics: Role in enteric methane mitigation and performance of animals. *Nutritional interventions for clean and green livestock production*. CAS in Animal Nutrition, IVRI, Izatnagar. pp 180–84.
- Bakshi M P S and Wadhwa M. 2012. Herbal feed additives- Role in Animal Nutrition. *Animal Nutrition-Advances and Developments*. (Eds) Mehra U R, Singh P and Verma A K. pp 707–33. Satish Serial Publishing House, New Delhi.
- Beauchemin K A, Kreuzer M, O'Mara F and McAllister T A. 2008. Nutritional management for enteric methane abatement: a review. *Australian Journal of Experimental Agriculture* **48**: 21–27.
- Benchaar C, Pomar C, Chiquette J. 2001. Evaluation of diet strategies to reduce methane production in ruminants: a modelling approach. *Canadian Journal of Animal Science* **81**: 563–74.
- Blaxter K L and Clapperton J L. 1965. Prediction of the amount of methane produced by ruminants. *British Journal of Nutrition* **19**: 511–22.
- Cottyn B G and Boucque C V. 1968. Rapid methods for the gas chromatographic determination of volatile acids in rumen fluid. *Journal of Agricultural and Food Chemistry* **16**: 105–07.
- Machmüller A, Soliva C R and Kreuzer M. 2003a. Effect of coconut oil and defaunation treatment on methanogenesis in sheep. *Reproductive and Nutritional Development* **43**: 41–55.
- Maia M R G, Chaudhary L C, Figueres L and Wallace R J 2007 Metabolism of polyunsaturated fatty acids and their toxicity to the microflora of the rumen. *Antonie Van Leeuwenhoek* **91**: 303–14.
- McAlister T A, Okine E K, Mathison G W and Cheng K J. 1996. Dietary, Environment and microbiological aspects of methane production in ruminants. *Canadian Journal of Animal Science* **76**: 231–43.
- Menke K H and Steingass H. 1988. Estimation the energetic feed value obtained by chemical analysis and *in vitro* gas production using rumen fluid. *Animal Research and Development* **28**: 7–55.
- Menke K H, Raab L, Salweski A, Steingass H, Fritz D and Scheider W. 1979. The estimation of digestibility and metabolizable energy content of ruminant feedstuffs from the gas production when they are incubated with rumen liquor *in vitro*. *Journal of Agricultural Science Cambridge* **93**: 217–22.
- Mills J A N, Dijkstra J, Bannink A, Cammell, S B Kebreab, E and France J. 2001. A mechanistic model of whole-tract digestion and methanogenesis in the lactating dairy cow: Model development, evaluation, and application. *Journal of Animal Science* **79**: 1584–97.
- Morales E R, Espinosa M A M, McKain N and Wallace R J. 2012. Ricinoleic acid inhibits methanogenesis and fatty acid biodehydrogenation in ruminal digesta from sheep and in bacterial cultures. *Journal of Animal Science* **90**: 4943–50.
- Robertson J A and Van Soest P J. 1981. The Detergent system of analysis and its application to human food. *The Analysis of Dietary Fibre in Food*. (Eds) W P T James and O Theander. pp. 123–58. Marcel Dekker Inc., New York.
- Snedecor G W and W G Cochran. 1994. *Statistical Methods*. Oxford and IBH Publications, New Delhi.
- SPSS. 2007. *Statistical Packages for Social Sciences*. Version 16, SPSS Inc., Linois, USA.
- Wadhwa M and Bakshi M P S. 2010. *Animal Agriculture and Greenhouse Gas Emission: Mitigation Strategies*. Vol.I (Lead Papers). (Eds) Pattanaik A K, Verma A K, Kamra D N and Sharma K. p. 119–22.
- Wolin M J. 1960. A theoretical rumen fermentation balance. *Journal of Dairy Science* **43**: 1452–59.