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**A New Species of Sucking Louse (Phthiraptera: Anoplura: Polyplacidae) from the Gray  
Mouse Lemur, *Microcebus murinus*, in Madagascar**

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Hasiniaina<sup>5,6</sup> and Sarah Zohdy<sup>7</sup>

23 **Abstract**

24 *Lemurpediculus madagascariensis* sp. nov. (Phthiraptera: Anoplura: Polyplacidae) is described  
25 from the Gray Mouse lemur, *Microcebus murinus* (J. F. Miller), from Ankarafantsika National  
26 Park, Madagascar. Lemurs were trapped using Sherman Live Traps and visually inspected for  
27 lice, which were preserved in 90% ethanol. Adults of both sexes and the third instar nymph of  
28 the new species are illustrated and distinguished from the four previously known species of  
29 *Lemurpediculus*: *L. verruculosus* (Ward), *L. petterorum* Paulian, *L. claytoni* Durden, Blanco and  
30 Seabolt, and *L. robbinsi* Durden, Blanco and Seabolt. It is not known if the new species of louse  
31 is a vector of any pathogens or parasites.

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33 **Key Words:** Phthiraptera, Anoplura, new species, mouse lemur, Madagascar

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45 The mouse and dwarf lemurs of Madagascar (family Cheirogaleidae) are among the smallest  
46 primates in the world (Zimmermann and Radespiel 2014, Lehman et al. 2016, Zohdy and Durden  
47 2016). Ectoparasites of cheirogaleid lemurs are inadequately known (Blanco et al. 2013, Zohdy  
48 and Durden 2016, Durden et al. 2017) and sucking lice (Phthiraptera: Anoplura) have been  
49 described from only three of the more than 30 species of cheirogaleids currently recognized  
50 (Hotaling et al. 2016). The three previously described species are *Lemurpediculus verruculosus*  
51 (Ward), an ectoparasite of the eastern mouse lemur, *Microcebus rufus* É. Geoffroy,  
52 *Lemurpediculus claytoni* Durden, Blanco and Seabolt, an ectoparasite of Sibree's dwarf lemur,  
53 *Cheirogaleus sibreei* Forsyth Major, and *Lemurpediculus robbinsi*, which parasitizes Crossley's  
54 dwarf lemur, *Cheirogaleus crossleyi* A. Grandidier (Durden et al. 2017). Another congeneric  
55 species, *Lemurpediculus petterorum* Paulian, parasitizes a different species of lemur which was  
56 stated to probably be *Lepilemur mustelinus* I. Geoffroy by Paulian (1958). Sucking lice are often  
57 host-specific (Durden and Musser 1994), and because few lemur species have been sampled for  
58 ectoparasites, there are undoubtedly additional undescribed species of *Lemurpediculus* associated  
59 with other species of cheirogaleids. Durden et al. (2017) amended the description of the genus  
60 *Lemurpediculus* to accommodate new developments in the systematics of Anoplura since the  
61 genus was erected by Paulian (1958). In this paper, we describe a new species of *Lemurpediculus*  
62 from the gray mouse lemur from Ankarafantsika National Park in northwestern Madagascar.

63

## 64 **Materials and Methods**

65 As part of a study on lemur health and communication, mouse lemurs were trapped in  
66 Ankarafantsika National Park, Madagascar in Jardin Botanique A using Sherman Live Traps (H.

67 B. Sherman Traps, Inc., Tallahassee, FL) baited with banana between May and November in  
68 2010 and 2011 (dry season). The study was conducted in the dry season because that is when the  
69 lemurs most readily enter traps for the banana. Lemurs were removed from the traps, manually  
70 restrained, and inspected for parasites by parting the fur down to the skin. Ectoparasites were  
71 collected primarily from the face, ears, legs, back and tail and stored in 90% ethanol in  
72 individually labeled vials. All lemurs were released at their capture site following collection of  
73 data and ectoparasites. This study was approved by Madagascar National Parks  
74 (N101/11/MEF/SG/DGF/DCB.SAP/SCB, N102/11/MEF/SG/DGF/DCB.SA/SCB) and the  
75 Arizona State University Institutional Animal Care and Use Committee (Protocol: 10-1077R).

76 Lice were cleared in 10% potassium hydroxide for ~24 h, rinsed in distilled water,  
77 transferred to 70% ethanol and then slide mounted in PVA medium (Bioquip Products, Rancho  
78 Dominguez, CA). Slide-mounted lice, including specimens of previously described congeneric  
79 species (in the Anoplura collections of LAD), were examined at high magnification under phase-  
80 contrast using an Olympus BH-2 microscope (Olympus Corporation of the Americas, Center  
81 Valley, PA) connected to an Ikegami MTV-3 video camera attachment and monitor (Ikegami  
82 Electronics, Neuss, Germany). Drawings of diagnostic morphological features were made from  
83 specimens examined at 100x – 400x. Specimen measurements were made using a calibrated  
84 graticule fitted into a microscope eyepiece.

85 Descriptive format for the new species follows Durden et al. (2010) and names and  
86 abbreviations of setae and morphological structures follow Kim and Ludwig (1978). Names of  
87 setae and certain structures are spelled out in full at first mention (with the abbreviation listed  
88 parenthetically) and then abbreviated when subsequently mentioned. The holotype male, allotype  
89 female and paratype third instar nymph of the new species are deposited in the U.S. National

90 Museum of Natural History (NMNH) (Smithsonian Institution), Department of Entomology,  
91 Washington DC.

92 Lemur taxonomy and common names used in this paper follow Groves (2005) and  
93 Hotaling et al. (2016).

94

## 95 **Results**

96 We trapped a total of 107 *M. murinus*. The entire ectoparasite faunas of these mouse lemurs will  
97 be reported in a separate paper.

98

99 *Lemurpediculus madagascariensis* **sp. nov.** (Figs. 1-3)

### 100 **Male (Fig. 1A,B,C)**

101 Total body length: 1.07–1.13 mm; mean, 1.10 mm (n=5). Head, thorax and abdomen lightly  
102 sclerotized.

103 Head: More heavily sclerotized along anterior dorsal margin and antero-laterally adjacent  
104 to first antennal segment; longer than broad with squarish anterior margin. One long Dorsal  
105 Principal Head Seta (DPHS), one small Dorsal Accessory Head Seta (DAcHS) anteromedial to  
106 DPHS, one Dorsal Posterior Central Head Seta (DPoCHS), 2-3 Dorsal Preantennal Head Setae  
107 (DPaHS), two Sutural Head Setae (SHS), three Dorsal Marginal Head Setae (DMHS), 3-4 Apical  
108 Head Setae (ApHS) and one Ventral Preantennal Head Seta (VPaHS) on each side. Antennae  
109 five-segmented with basal segment slightly wider than long and distinctly broader than second  
110 segment; fourth segment slightly extended posterolaterally.

111 Thorax: Much longer than wide, slightly wider than head. Thoracic sternal plate (Fig. 1B)  
112 lightly sclerotized, with narrow anterior extension and broadly curved lateral margins; tiny

113 sclerite bearing two long setae immediately posterior to thoracic sternal plate. Dorsal Principal  
114 Thoracic Seta (DPTS) mean length 0.13 mm (range, 0.12-0.14 mm, n=8), with adjacent small  
115 Dorsal Mesothoracic Seta (DMsS) on each side; mesothoracic spiracle mean maximum diameter  
116 0.025 mm (range, 0.023-0.027 mm, n=8). Legs with subtriangular coxae; forelegs each  
117 terminating in small tibio-tarsal claw; mid and hindlegs subequal in size, each terminating in  
118 large, robust tibio-tarsal claw.

119         Abdomen: Wider than thorax with six annulated spiracles on each side. Paratergal plates,  
120 tergites and sternites absent. One row of two long Dorsal Central Abdominal Setae (DCAS)  
121 anteriorly, followed by five rows of 4-6 long DCAS and then two rows of two shorter DCAS.  
122 Six Dorsal Lateral Abdominal Setae (DLAS) on each side, each adjacent to corresponding  
123 spiracle; DLAS 1-5 each with adjacent small seta; DLAS 1 and adjacent small seta both inserted  
124 on small ridge. DLAS 6 borne on small sclerite and distinctly longer than other DLAS and  
125 extending away from abdomen; five rows of four long Ventral Central Abdominal Setae  
126 (VCAS); VCAS in most posterior row slightly shorter than other VCAS. One posterior Ventral  
127 Lateral Abdominal Seta (VLAS) on each side adjacent to corresponding DLAS and most  
128 posterior spiracle. ~10 tiny to small dorsal setae near posterior apex of abdomen.

129         Genitalia (Figs 1A,C): Subgenital plate (Fig. 1A) well sclerotized, somewhat urn-shaped  
130 with bulging medio-lateral margins and small antero-lateral extensions. Basal apodeme longer  
131 than parameres and other genitalic components combined, slightly expanded posteriorly into two  
132 paddle-shaped plates on each side; C-shaped anterior endomere with posteriorly converging  
133 arms; anteriorly acuminate aedeagal sclerite located inside anterior endomere; posteriorly  
134 acuminate central endomere bordered laterally by one broad accessory sclerite on each side;

135 parameres broad anteriorly and tapering posteriorly to rounded apex; pseudopenis relatively  
136 small but extending posteriorly beyond apices of parameres and terminating in acute apex.

137

138 **Female (Fig. 2A,B,C)**

139 Body length: 1.32-1.50 mm; mean, 1.43 mm (n=6). Head, thorax and abdomen as in male unless  
140 indicated otherwise.

141       Head: Slightly wider than in male.

142       Thorax: Mesothoracic spiracle mean maximum diameter 0.0275 mm (range, 0.0250-  
143 0.0283, n=6).

144       Abdomen: Dorsally with eight rows of four long DCAS anteriorly followed by one row  
145 of six slightly shorter DCAS and one row six small Tergal Abdominal Setae (TeAS) inserted on  
146 broad, curved tergite immediately posterior to subgenital plate. One row of one DLAS on each  
147 side anteriorly followed by six rows of two DLAS on each side and then one very long DLAS  
148 borne on small sclerite posterior to last spiracle. One row of two long VCAS anteriorly followed  
149 by five rows of four VCAS. One very long VLAS posteriorly, associated with last DLAS and  
150 most posterior spiracle.

151       Genitalia (Fig. 2C): Subgenital plate broadly rounded anteriorly and extending  
152 posteriorly to broad apex, with small, distinct lateral lacuna on each side; each lacuna with four  
153 small setae inserted anteriorly; three small setae inserted on each side of subgenital plate near  
154 postero-lateral margins. Vulvar fimbriae distinct and extensive collectively forming a V shape;  
155 gonopods VIII and IX indistinct and with ~13 setae attached to each gonopod VIII and two  
156 slightly larger setae attached to each gonopod IX; gonopod setae collectively forming postero-

157 lateral fan-like patches. Curved subterminal transverse sclerite with small anterior apex situated  
158 between gonopods IX. Three small terminal setae ventrally on each side of genital opening.

159

### 160 **Third Instar Nymph (Fig. 3)**

161 Body length: 1.00-1.25 mm; mean, 1.11 mm (n=5).

162 Head: Shape as in male but with slightly more rounded anterior margin. One fairly long  
163 DPTS and one adjacent small DAcHS, one DPoCHS, two DMHS, two SHS, 3-4 ApHS and one  
164 VP aHS on each side. Antennae approximately as in male.

165 Thorax: Slightly wider than head, much longer than wide; one long DPTS (mean length,  
166 0.125 mm, range, 0.110-0.129 mm, n=4) adjacent to mesothoracic spiracle (mean maximum  
167 diameter, 0.025 mm, range, 0.020-0.028, n=4) on each side. Foreleg coxae subtriangular; mid  
168 and hind coxae more irregular; forelegs each terminating in small tibio-tarsal claw; mid and  
169 hindlegs subequal in size, each terminating in large, robust tibio-tarsal claw.

170 Abdomen: Wider than thorax with eight rows of two DCAS and nine rows of VCAS.  
171 Eight DLAS on each side; DLAS 2-7 each with accompanying spiracle; DLAS 2 with adjacent  
172 small accessory seta, both borne on small protuberance; two additional small setae adjacent to  
173 DLAS 2 on each side; one additional small accessory seta on each side adjacent to each of DLAS  
174 3-6; DLAS 7 and 8 both long, extending from postero-lateral abdomen and each associated with  
175 one VLAS on each side.

176 **HOLOTYPE** ♂ ex *Microcebus murinus* (J. F. Miller) (gray mouse lemur) (male, Animal  
177 25-09), MADAGASCAR: Boeny Region, Ankarafantsika National Park, Jardin Botanique A  
178 (46°48' E, 16°19' S), elevation 190 m, 17 October 2010, Coll: Sharon Kessler and Alida I. F.  
179 Hasiniaina. Deposited in NMNH (accession barcode, USNMENT00981907).



180           **ALLOTYPE** ♀ ex *M. murinus*, same data as Holotype except (male, Animal 82-10) and  
181 13 Nov. 2011. Deposited in NMNH (accession barcode, USNMENT00981908).

182           **PARATYPES** One nymph (third instar) same data as Holotype except (male, Animal 25-  
183 10), 15 October 2010 (accession barcode, USNMENT00981909); 2♂, 2♀, 2 nymphs (third  
184 instars) same data as Holotype except different individual lemurs and various dates in 2010 and  
185 2011; deposited in Georgia Southern University Insect Collection (1♂, 1♀) (accession no. L-  
186 3813) and Anoplura Collection of L. A. Durden (1♂, 1♀).

187 **ETYMOLOGY:** This species is named for the faunistically unique island of Madagascar where  
188 both the louse and its host co-occur.

189           For comparative purposes, the female subgenital plates and associated structures, for the  
190 four previously described congeneric species, *L. petterorum*, *L. claytoni*, *L. verruculosus*, and *L.*  
191 *robbinsi*, are illustrated in Fig. 4.

192

## 193 **Discussion**

### 194 **Males**

195           Males of *Lemurpediculus* spp. can easily be separated by examination of the genitalia in  
196 cleared slide-mounted specimens. In *L. petterorum* males, the parameres are about equal in  
197 length to the basal apodeme (shown in Paulian 1958, Fig. 1B), whereas *L. claytoni*, *L. robbinsi*,  
198 and *L. verruculosus*, the parameres are much shorter than the basal apodeme. The shape of the  
199 parameres and the presence or absence of genitalic endomeres and accessory sclerites can be  
200 used to separate these four species. The parameres have slightly concave medio-lateral margins  
201 in *L. claytoni* (shown in Durden et al. 2017, Fig. 3B) and distinctly rounded convex medio-lateral  
202 margins in both *L. verruculosus* (shown in Durden et al. 2010, Fig. 3) and *L. robbinsi* (shown in

203 Durden et al. 2017, Fig. 2B). The medial margins of the parameres of *L. robbinsi* have a  
204 distinctly rounded bulge (shown in Durden et al. 2017, Fig. 2B) which is absent in *L.*  
205 *verruculosus* (shown in Durden et al. 2010, Fig. 3). Further, the pseudopenis extends well  
206 beyond the posterior apices of the parameres in *L. robbinsi* (shown in Durden et al. 2017, Fig.  
207 2B) but just barely beyond the apices in *L. verruculosus* (shown in Durden et al. 2010, Fig. 3).  
208 The male genitalia of *L. madagascariensis* sp. nov. (Fig. 1C) have more acute anterior paramere  
209 margins than those of the other species in the genus and have additional adjacent small plates  
210 that are not present in the other species – a central endomere and a pair of lateral sclerites.  
211 Externally, the thoracic sternal plate of *L. petterorum* (shown in Paulian 1958, Fig. 2B) lacks an  
212 anterior projection which is clearly present in the other four species.

### 213 **Females**

214 Females of all five known species of *Lemurpediculus* can easily be separated based on  
215 the shape of the subgenital plate which can be observed in either cleared or uncleared specimens.  
216 In both *L. petterorum* and *L. madagascariensis* sp. nov., the anterior and posterior portions of the  
217 subgenital plate are joined centrally and laterally and have two lateral lacunae in the anterior  
218 portion (Figs. 2C, 4A), whereas the anterior and posterior sections of the subgenital plate are not  
219 joined laterally in the other three species Fig. 4B-D). The two lacunae in the female subgenital  
220 plate of *L. petterorum* are much larger than those in *L. madagascariensis*, collectively making up  
221 almost half of the plate size in the former species (Fig. 4A), but less than 10% in the latter  
222 species (Fig. 2C). Also, the thoracic sternal plate in the female of *L. petterorum* lacks the anterior  
223 extension (shown in Paulian 1958, Fig. 2B) that is present in females of *L. madagascariensis* sp.  
224 nov. (Fig. 2B in this paper). In females of *L. verruculosus*, the anterior portion of the subgenital  
225 plate is 3-4 times larger than the posterior portion (Fig. 4C). The anterior and posterior sections

226 of the subgenital plate are subequal in size in females of *L. claytoni* (Fig. 4B) whereas the  
227 anterior portion is slightly larger than the posterior portion in *L. robbinsi* (4D). Also, the shape of  
228 the female subgenital plate is very different between these species (Figs. 2C, 4A-D).

## 229 **Nymph**

230 The third instar nymph of only one other species of *Lemurpediculus* has been described.  
231 This nymphal stage of *L. verruculosus* was described and illustrated by Durden et al. (2010). The  
232 third instar nymph is easily separated between these two species because *L. verruculosus* lacks  
233 DLAS next to abdominal spiracles 2-5 (see Durden et al. 2010, Fig. 3) whereas *L.*  
234 *madagascariensis* sp. nov. has one long DLAS next to each of these spiracles on each side (Fig.  
235 2C).

236 With the description of the new species included in this paper, there are now five  
237 recognized species of *Lemurpediculus*. Four of these species, *L. verruculosus*, *L. robbinsi*, *L.*  
238 *claytoni*, and *L. madagasarensis* sp. nov., parasitize cheirogaleid lemurs while the host of the  
239 fifth species, *L. petterorum* Paulian, was stated by Paulian (1958) to probably be *Lepilemur*  
240 *mustelinus* (weasel sportive lemur) another nocturnal species which belongs to a different lemur  
241 family, the Lepilemuridae. All five of these species of lice appear to be host-specific but the  
242 host/s of *L. petterorum* requires verification. It would be premature to provide a dichotomous  
243 identification key for known *Lemurpediculus* species because we anticipate the collection and  
244 description of additional species in this genus in the future, especially considering the highly  
245 diverse radiation of mouse lemur species around Madagascar (Yoder et al. 2010, 2016).

246 With few exceptions, very little is known about the potential for sucking lice of wild  
247 mammals to transmit pathogens to their hosts (Durden 2001) and nothing is currently known  
248 about any potential vectorial role of lice that parasitize lemurs. However, some pathogens and

249 parasites of lemurs, including certain viruses, bacteria and protozoans could feasibly be  
250 transmitted by sucking lice. Future research should address the potential for blood-feeding  
251 ectoparasites, including sucking lice, to transmit pathogens to lemurs, particularly in light of the  
252 threatened or endangered status of many species of these primates.

253 Many authors have advocated conserving (or co-conserving) parasites of rare hosts (e.g.,  
254 Durden and Keirans 1996, Whiteman and Parker 2005, Dunn et al. 2009) and we likewise  
255 advocate co-conservation of mouse lemurs and their unique host-specific parasites including  
256 their sucking lice.

257

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273

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325

326 **Footnotes**

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337

338

339 **Figure Legends**

340 **Fig. 1.** *Lemurpediculus madagascariensis* sp. nov., adult male. A: Dorsoventral view. B:  
341 Thoracic sternal plate. C. Genitalia. All scale bars, 0.1 mm.

342 **Fig. 2.** *Lemurpediculus madagascariensis* sp. nov., adult female. A: Dorsoventral view. B:  
343 Thoracic sternal plate. C. Genitalia. All scale bars, 0.1 mm.

344 **Fig. 3.** *Lemurpediculus madagascariensis* sp. nov., third instar nymph: Dorsoventral view. Scale  
345 bar, 0.1 mm.

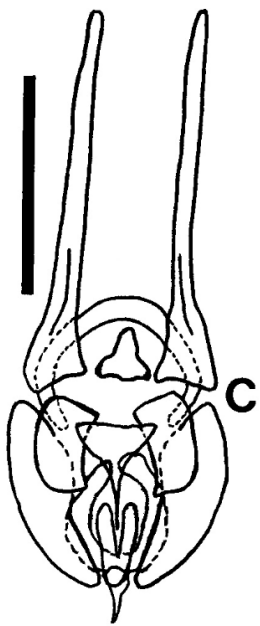
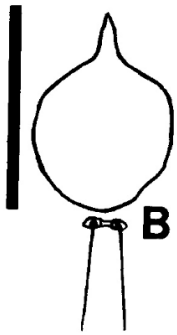
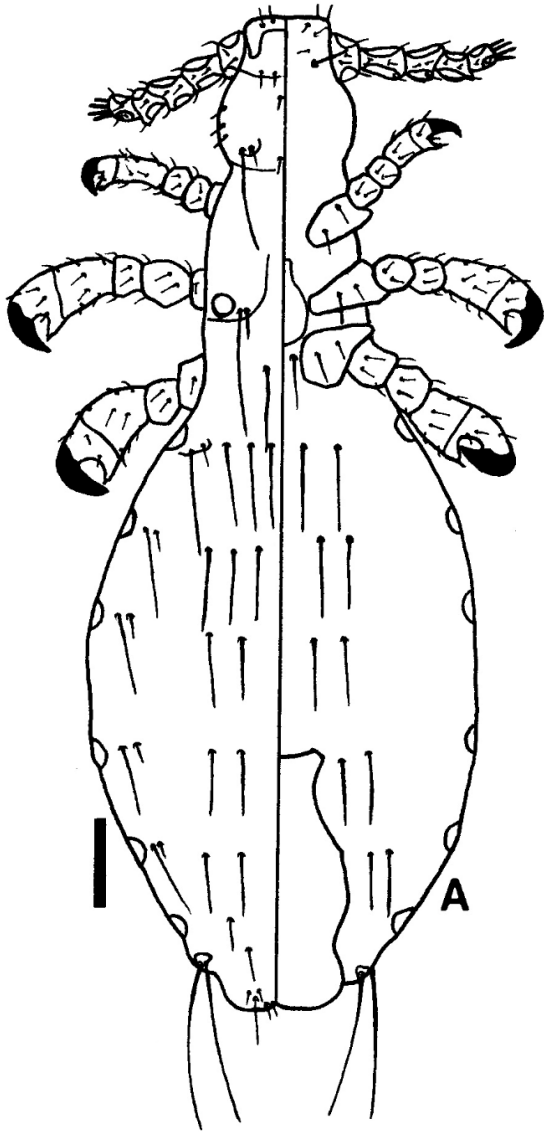
346 **Fig. 4.** Female subgenital plates, associated setae, and vulvar fimbriae of the four previously  
347 described species of *Lemurpediculus*. A: *L. petterorum* Paulian ex (probably) *Lepilemur*  
348 *mustelinus*, Ambatolampy District, Madagascar. B: *L. claytoni* Durden, Blanco and Seabolt ex  
349 *Cheirogaleus sibreei*, Tsinjoarivo, Amabatolampy District, Madagascar. C: *L. verruculosus*  
350 (Ward) ex *Microcebus rufus*, Ranomafana National Park, Madagascar. D: *L. robbinsi* Durden,  
351 Blanco and Seabolt ex *Cheirogaleus crossleyi*, Tsinjoarivo, Amabatolampy District,  
352 Madagascar. Scale bar = 0.05 mm.

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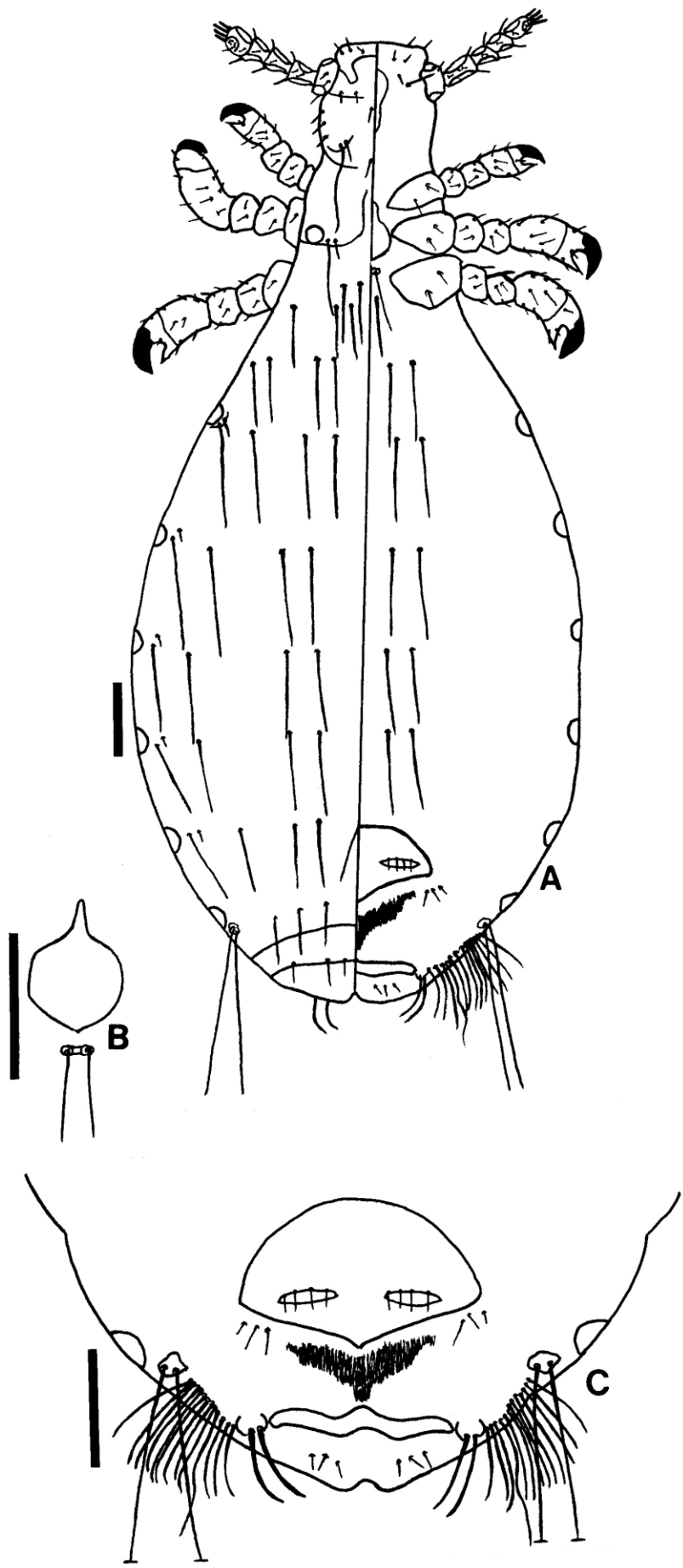
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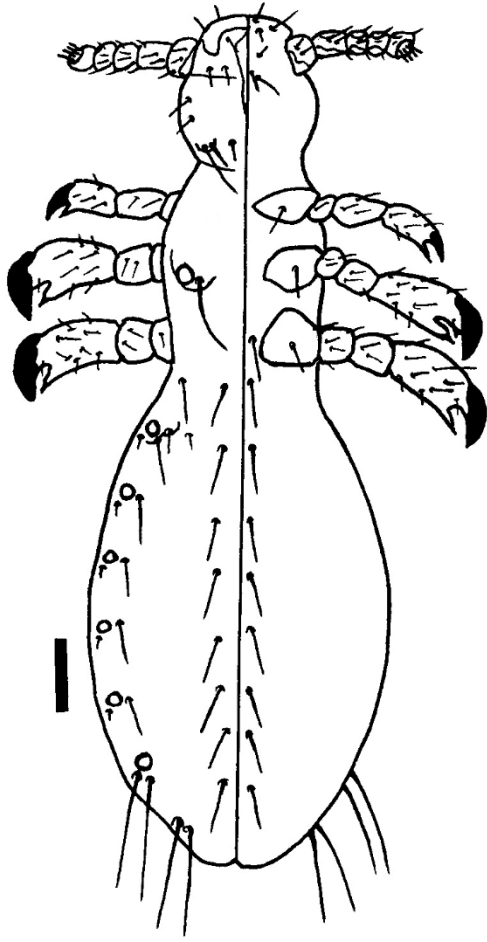


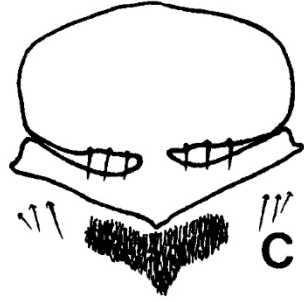
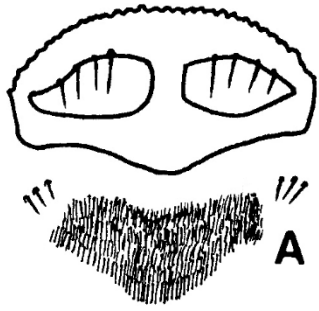












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