The Immunological Effectiveness of Some Common Plants

Anwar I. S. Al-Assaf *

Zahraa H. M. Kadri*

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Abstract:

Three plant species were picked randomly and their alcoholic extracts have been screened to know their effects on the phagocytic capability and intracellular killing of yeast by human peripheral macrophages. Macrophage cultures were incubated with different concentration of each plant extract: for 15 min., 30 min and 45 min. The phagocytes activity in *Iresine herbstii* extract was significantly ($p \le 0.05$) increased with increasing dose and time of incubation. In *Mentha piperita* extract, increasing in dose of 20% and 25% of plant extract, perhaps because the antimicrobial and antiviral activities of this plant, as well as strong antioxidant and antitumor actions. While in *Elettaria cardamomum*, a significant elevation has been observed in phagocytic efficiency when the dose of extract increase to 15%, then decreased in the subsequent doses (20% and 25%), in three periods of time. These findings may suggest that cardamom exert immunomodulatory roles.

Key words: Phagocytosis, Iresine herbstii, Mentha piperita, Elettaria cardamomum

Introduction:

Phagocytosis is an innate immunity refers to antigen-nonspecific defense mechanisms that a host uses immediately or within several hours after exposure to an antigen [1]. Phagocytosis is a specific form of endocytosis involving the vesicular internalization of solids such as bacteria, and is, therefore, distinct from other forms of endocytosis such as the vesicular internalization of various liquids.

Phagocytosis is involved in the acquisition of nutrients for some cells, and, in the immune system, it is a major mechanism used to remove pathogens and cell debris. Bacteria, dead tissue cells, and small mineral particles are all examples of objects that may be phagocytosed [2]. Infectious diseases account for high proportion of health problems in the developing countries. Microorganism has developed resistance to many antibiotics and this

has created immune clinical problems in the treatment of infectious diseases and because of inadequate availability and high cost of new generation antibiotics, scientists are forced to search for new antimicrobial substances including from various sources medicinal plants [3]. Many of the plants used today were known to the people of ancient culture throughout the world for preservative their and medicinal property [4]. However several plants are used in the form of crude extracts, infusions or plaster to treat common infections without scientific evidence of efficacy [5]. Plants have been used to treat various ailments since the advent of human history, because the herbals have been usually considered to be safe and nontoxic compared to synthetic compounds. So, there are abundant studies about plant pharmacological properties [6,7, 8].

*Department of Biology, College of Education, Ibn Al-Haitham, for Pure Sciences, University of Baghdad.

There has been an increase in people trying to find naturaimmune system boosters. Herbal and homeopathic remedies have been used in traditional medicine for thousands of years to strengthen immune system functioning, acting as immune system tonics to encourage normal and efficient defense against pathogens and routine recovery [9].

Antimicrobial compounds of plant origin may occur in stems, roots, leaves, bark, flowers and fruits of plants [10]. For example: Iresine herbstii belongs to the family Amaranthaceae. It is commonly referred to as blood leaf, [11]. Iresine herbstii leaves are used as wound healing, anticancer agent [12], post-labor tonic [13], and externally against skin depurative such as eczemas, sores and pimples [11] as well as antimicrobial agents. It contains several bioactive substances and showed different biological activities and is used to treat various diseases. Leaves of I. diffusa are used to treat malaria [14]. Red-coloured plants in the family Amaranthaceae are recognized as a rich source of diverse and unique betacyanins such as acetylated and nonacetylated betacyanins. Acylated betacyanins are available with the highest proportion in I. herbstii and Gomphrena globosa [15] and [16]. We have selected Iresine herbstii, based on, these plants are more accessible and affordable [17] and can contribute to new bioactive compounds that are safe and effective. The sesquiterpene Iresine is found in high concentration in the plant "herb of the Maya" and the properties include anti-cancer, antiinflammatory, anti-allergic and Most terpenes are antiseptic. also substances with positive and stimulatory effect on the body in general. These substances are also able to shorten the menstrual period. Isoflavone tlatlancuayin has antioxidant, which captures free oxygen radicals. and contributes to cell powerful renewal. and also а antimicrobial agent [18]. The plant is used in astringent, diuretic. spasmolytic, whooping cough and roots in hemicranias [16]. Leaves and flowers are used in decoction, fever, relaxant and kidney problems [19] and also as an antipyretic [11]. Schmidt et al. [20], reported that this plant possessed antiinflammatory, cytotoxic and apoptotic activities and also has very low antioxidant activity [21].

Another plant: Mentha piperita has antimicrobial and antiviral activities. antioxidant and strong antitumor actions. and some antiallergenic potential [22]. Mentha piperita is one of the world's oldest medicinal herbs and used in both Eastern and Western traditions. This plant is a perennial plant in *Lamiaceae* family and contains about 1.2-1.5% essential oils. The chemical composition of the essential oil from peppermint (Mentha piperita) was analyzed by GC/FID and GC-MS. The main constituents were menthol (40.7%)and menthone (23.4%). Further components were (+/-)-menthyl acetate, 1,8-cineole, limonene, betapinene and beta-caryophyllene [20,23, 24]. Mentha (also known as Mint, from Greek míntha. Linear B mi-ta) is a genus of flowering plants in the family Lamiaceae (mint family). The species are not clearly distinct and an estimate of the number of species varies from 13 to 18. Most Mentha grow best in wet environments and moist soils [25]. Animal studies show Mentha piperita a relaxation effect have to on gastrointestinal tissue, analgesic and anesthetic effects in the central and peripheral nervous system, immune system influencing actions and anti cancer potential. Human studies on the gastrointestinal (GI), respiratory tract and analgesic effects of Mentha piperita oil and its constituents have been reported [26].

Cardamom (*Elettaria cardamomum*) of the Zingiberacea family is one of the world's very ancient and expensive spices mainly grown in Sir Lanka and India. South Cardamom extract contains a number of volatile oils that provide numerous health benefits such as aiding digestion and improving metabolism. Another of the essential cardamom benefits is that is very helpful in removing toxins from the body. Regular use of cardamom gradually removes the accumulated toxins and improves the blood circulation. In this aspect, cardamom benefits are similar to cinnamon benefits. Although there are no known side effects of cardamom, it should be avoided during pregnancy [27]. The seeds of their ripe fruits are used medicinally, as a spice, and also as a flavoring agent in curries, coffee and cakes, particularly in the Arab countries. Some is used in the manufacture of liqueurs and a relatively small quantity in pharmacy, chiefly in the form of compound tincture of cardamom [28]. Cardamom seed yields 4% of volatile oils containing a high proportion of Terpinyl acetate and cincole and small quantities of other monoterpenes, including alcohols and esters [29]. Govindarajan, et al. [30] reported the presence of over 150 compounds in cardamom aroma. Many of these compounds are commonly found in cardamom oil [29, 30]. It is also thought to be supportive of the nervous system and could be useful in massage blends addressing sciatica. Finally. cardamom contains many minerals vitamins and including: niacin. vitamin riboflavin, С, magnesium and potassium that help ensure optimum health [29]. This study was to determine the possible effects of these three plants: Iresine herbstii, Mentha piperita and Elettaria cardamomum on phagocytic activity in human phagocytes.

Material and Methods:

I. Collection of blood:

Venous blood (5 ml) was collected in heparin tubes, By means of density gradient centrifugation modified by [31]; the lymphocytes were isolated from whole blood.

Π. Preparation of heat killed yeast:

Ten grams of yeast (Saccharomyces *cervisiae*) were suspended in a warm (37°C) sterilized physiological saline (one hindered-fifty milliliters), the cell suspension then heated in boiling water bath for sixty minutes, after heating cell suspension was cooled to (37° C) then filtered by tri stratified sterilized gaze. Yeast cells were adjusted to a concentration of $1X \quad 10^7$ cells /ml. distributed into small tubes and freeze at (-20° C) [32].

III. Plant samples preparation:

Healthy, disease free (fresh) leafs and stems of Iresine herbstii. Mintha seeds piperta and of Elettaria cardamomum, were collected, washed properly in the tap water followed by detergent water and finally rinsed with distilled water until no foreign material (damaged leaves remained were removed). The fresh plant materials were left to dry in a closed room (25-28°C) for approximately five days. The dried plant parts were pulverized to obtain a powder by using sterile blender. The electrical powdered samples were stored in air tight container, protected from sunlight for further use [33].

IV. Alcoholic extracts:

Twenty grams of powdered materials of each of the three plants were continuously extracted with eighty ml of solvent like ethyl alcohol (99%) [34]. For successive solvent extraction on polarity using based soxhlet extraction apparatus at the boiling point of the respective solvents for 3-4 h or until the color of the extracted solvent become clear. Extracts were concentrated under reduced pressure using rotary evaporator and they were poured into a pre-weighed vial, further dried in a desiccating chamber until a constant dry weight was obtained. The extract vials were stored at 4°C for further studies [33].

VI.

0.25 ml Cells + 0.05 ml killed yeast suspension + 0.1 ml normal salin + 0.1ml different concentration of plant extracts (5%, 10%, 15%, 20% and 25%) which mixed before as in guidelines for laboratories and field testing from WHO [35].

All were incubated for different periods (15min., 30min. and 45min).

The phagocytic activity was determined, as in [36]

by counting the number of leukocytes (neutrophils and macrophages) that phagocytes yeast cells as in this equation:

Phagocytosis Index (%) = Number of phagocytotic Cells/ Total Count X 100

V. Statistical Analysis

The results were analyzed using the computer program SPSS (Statistical package for Social Sciences) version 13. Their data were presented in terms of means \pm standard errors (S.E), and differences between means were assessed by ANOVA and LSD tests.

Results and Discussion:

 Table (1): Phagocytic activity levels with different concentrations of Iresine herbstii extracts and control in three periods of time.

Concentration (%)	Number	Time per minutes Phagocytic Index (Mean ± S.E)		
		15min.	30 min.	45 min.
Control	4	$38.75 \pm 0.75^{\mathbf{D}}$	$45.75 \pm 0.85^{\circ}$	45.50 ± 1.32^{E}
5	4	$57.75 \pm 1.03^{\circ}$	$63.50 \pm 1.32^{\mathbf{B}}$	$63.00 \pm 1.22^{\mathbf{D}}$
10	4	$59.75 \pm 0.63^{\circ}$	66.25 ± 0.48^{B}	$67.00 \pm 0.91^{\circ}$
15	4			70.25 ± 1.43^{B}
20	4		70.00 ± 0.91^{A}	
25	4	71.25 ± 1.03^{A}	$72.50 \pm 0.86^{\mathrm{A}}$	76.25 ± 0.75^{A}

*Different letters: Significant difference ($P \le 0.05$) between mean values within the columns.

Phagocytes activity of monocytes using iresine herbstii extract was significantly $(p \le 0.05)$ increased with increasing dose and time of incubation, (Table -1). 15minits After of incubation. phagocytic percentages were increased gradually (57.75%, 59.75%, 63.00%, 69.25% and 71.25%) using plant extract concentrations (5, 10, 15, 20, 25%, respectively), as compared to control group (38.75 %). While, when incubated it for 30 minutes, phagocytic percentages also was elevated (63.50%, 66.25%, 66.50%, 70.00% and 72.50%) in extract concentrations (5, 10, 15, 20, 25%, respectively), as compared to

control group (45.75 %). When the incubation period extended to 45 minits, phagocytosis results were showed increased levels (63.00%, 67.00%,70.25%,73.00% and 76.25%) in extract concentrations (5, 10, 15, 20, 25%, respectively), as compared to control group (45.50 %). Such findings may highlight the importance of Iresine herbstii extract in defense against pathogens which in agreement with [11] and [20], whom reported that this plant possessed anti-inflammatory, cytotoxic and apoptotic activities and also has very low antioxidant activity [21].

Concentration (%)	Number	Time per minutes Phagocytic index (Mean ± S.E)		
		15 min.	30 min.	30 min.
Control	4	38.75 ± 0.75^{E}	$45.75 \pm 0.85^{\circ}$	$45.50 \pm 1.32^{\circ}$
5	4	$50.25 \pm 2.06^{\circ}$	61.75 ± 0.75^{B}	$56.75 \pm 2.14^{\mathbf{B}}$
10	4	$43.00 \pm 0.71^{\mathbf{D}}$	$63.50 \pm 1.19^{\mathbf{B}}$	67.00 ± 1.41^{A}
15	4	$47.00 \pm 1.76^{\circ}$	64.75 ± 1.38^{B}	$65.25\pm0.85^{\rm A}$
20	4	58.75 ± 1.11^{B}	64.23 ± 0.75^{B}	65.50 ± 0.65^{A}
25	4	$63.00 \pm 0.82^{\text{A}}$	70.50 ± 1.85^{A}	$69.00\pm0.71^{\mathbf{A}}$

 Table (2): Phagocytic activity levels in five concentrations of Mintha piperta

 extracts in three periods of time

Table-2, phagocytic demonstrates activity of monoctytes using Mentha *piperita* extract which was significantly $(p \le 0.05)$ increased with increasing time of incubation. In the first period (15 minutes), the phagocytosis using plant extract concentrations (5, 10, 15, 20, 25%) were (50.25%, 43.00%, 47.00%, 58.75% and 63.00% respectively) as compared to control group (38.75 %), whereas at the second period of incubation (30 minutes) phagocytosis percentages were (61.75%, 63.50%, 64.75%. 64.23% 70.50% and respectively) as compared to control group (45.75%), While after 45 minutes incubation with the same extract concentrations results were (56.75%, 67.00%, 65.25%, 65.50% and 69.00% respectively) when compared with control group(45.50%). Increasing in dose and time of incubation leads to elevate phagocytic capbility, especially in the dose of 20% and 25% of plant extract. Mentha is widely used as one of the important spices and traditional herbs in the world. Many reports have confirmed that the Mentha or its extracts has some pharmacological activities including, anti-tumor and anti-oxidation.

Studies have shown that Mentha *piperta* possess an anti-inflammatory effect against both acute and chronic models of inflammation [37]. In vitro data: Peppermint oil and menthol have moderate antibacterial effects against both Gram-positive and Gram- negative bacteria. Peppermint extracts are bacteriostatic against *Streptococcus* thermophilus and Lactobacillus bulgaricus. Menthol is bactericidal against Staphylococcus pyogenes, S. aureus. Streptococcus pyogenes, Serratia marcescens, Escherichia coli, and Mycobacterium avium [38]. It is well known that many diseases/disorders, that have immunomodulated components, can be modified administration by of biological compounds that activate key

pathways in the immune system. They

strengthen the defense and immune

mechanisms of the body and can be

used for stimulating the non-specific

immune responsiveness in both the

human and veterinary medical practice

[39].That is compatible with our

findings in increasing phagocytosis,

non-specific

which considered as

immune response.

^{*}Different letters: Significant difference ($P \le 0.05$) between mean values within the columns.

Concentration (%)	Number	Time per minutes Phagocytic Index (Mean ± S.E)		
		15 min.	30 min.	45 min.
control	4	$38.75 \pm 0.75^{\mathbf{D}}$	45.75 ± 0.85^{D}	$45.50 \pm 1.32^{\mathbf{D}}$
5	4	$42.75 \pm 2.29^{\text{CD}}$	$42.25 \pm 2.66^{\mathbf{D}}$	$43.50 \pm 1.94^{\mathbf{D}}$
10	4	$43.75 \pm 2.32^{\text{BCD}}$	$56.00 \pm 2.16^{\circ}$	66.7 ± 1.49 ^{B}
15	4	67.25 ± 1.11^{A}	72.75 ± 1.03^{A}	$89.25 \pm 1.11^{\mathbf{A}}$
20	4	$47.25 \pm 1.25^{\mathbf{B}}$	$63.00 \pm 1.08^{\mathbf{B}}$	$52.25 \pm 2.50^{\circ}$
25	4	$45.25 \pm 2.78^{\mathbf{B}}$	$42.75 \pm 1.31^{\mathbf{D}}$	$44.25 \pm 1.11^{\mathbf{D}}$

Table 3: Phagocytic activity levels in five concentrations of <i>Elettaria cardamomum</i>
extracts in three periods of time.

In the first period of incubation with concentration (15minits). extract phagocytic percentages were (42.75%, 43.75%, 67.25%, 47.25% and 45.25%) using the concentrations (5, 10, 15, 20, 25%, respectively), as compared to control group (38.75 %). But, after incubation for 30 minutes, percentages 72.75%, were (42.25%)56.00%. 42.75%) 63.00% and using concentrations (5, 10, 15, 20, 25%, respectively), as compared to control group (45.75 %). While phagocytic percentages were (43.50%, 66.7%, 89.25%, 52.25% and 44.25%) using concentrations (5, 10, 15, 20, 25%, respectively), as compared to control group (45.50%). Here we observed that there was a significant elevation in phagocytic efficiency when the dose of extract increased to 15%. then decreased in the subsequent doses (20% and 25%), in three periods of time.

The findings indicated that immune defense may be diminished in high concentration of cardamom and exert immunomodulatory roles, as suggested by Majdalawieh and Carr [40], whom also confirmed that nitric oxide macrophages production by is significantly augmented and reduced by black pepper and cardamom, respectively and they suggested that black pepper and cardamom exert immunomodulatory roles and antitumor activities, and hence they manifest themselves as natural agents that can promote the maintenance of a healthy immune system. Generally, our results agreed with Acharya, *et al.* [41] that Cardamom oil is also noted for its antiseptic properties, and may stimulate phagocytic action of the immune system.

References:

- 1. Peter, P., 2000. The Immune System. Garland Science, 2nd edition.
- Ishimoto, H., Yanagihara, K., Araki, N., *et al.* 2008. "Single-cell observation of phagocytosis by human blood dendritic cells". Jpn. J. Infect. Dis .61(4):294–7. PMID 18653972.
- Sashikumar, J.M., Remya, M. and 3. Janardhanan, K. 2003. Antimicrobial activity of ethanol medicinal plants of Nilgiri Bioshpere reserve and Western Ghats. J. Microbial. Asian Biotechnol. 5: 183-185.
- **4.** Zaika, L.I. 1975. Spices and Herbs, their antimicrobial activity and its determination. J. Food Safety. 9: 97–118.
- Ahmed, I., Mehmood, Z., Mohammad, F. 1998. Screening of some Indian medicinal plants for their antimicrobial properties. J. Ethnopharmacol. 62: 183–193.

^{*}Different letters: Significant difference ($P \le 0.05$) between mean values within the columns.

- Yuan, D., Sunouchi, H., Sakurai, T., Saito, K., and Kano, Y. 2002. Pharmacological properties of traditional medicines (XXVII). Interaction between Ephedra Herb and Gypsum under hyperthermal conditions in rats. Biol. Pharm. Bull. 25(7): 872-874.
- 7. Gubaev, A.G., Ortenberg, E.A., Rusakova, O.A. and Chiriat'ev, E.A. 1996. The pharmacological properties of a direct-action anticoagulant from the herb Nonea poulla (L.) D. C. Eksp. Klin. Farmakol. 59(1): 40-42.
- Liu, Y., Ling, Y., Hu, W., Xie, L., 8. Yu, L., Qian, X., Zhang, B. and Liu, B. 2009. The Herb Medicine Formula 'Chong Lou Fu Fang' Increases the Cytotoxicity of Chemotherapeutic Agents and Down-regulates the Expression of Chemotherapeutic Agent **Resistance-related** Genes in Human Gastric Cancer Cells In Vitro. eCAM: nep175.
- **9.** Bergner P. L. 1993. Medical Herbalism; 5(4):3
- Borchardt, J.o.y. R., Wyse, D. L., Sheaffer, C., C, Kauppi, K. L., Fulcher, R., Gary, E., Nancy, J., Biesboer, D. D., Russell, F. B. 2008. Antimicrobial activity of native and naturalized plants of Minnesota and Wisconsin. J. Medic. Plants, Res. 2 (5): 098–110.
- **11.** De, Feo, V. 2003. Ethnomedical field study in northern Peruvian Andes with particular reference to divination practices. J. Ethnopharmacol.; 85:243–256.
- Sebold, D.F., Levantamento, etnobotânico, 2003. De plantas de uso medicinal no município de Campo Bom, Rio Grande do Sul, Brasil. Universidade Federal do Rio Grande do Sul; p. 107. Master thesis.
- **13.** Srithi, K., Balslev, Henrik, 2009. Medicinal plant knowledge and its

erosion Wangpakapattanawong Prasit, Srisanga Prachaya, Trisonthi Chusie. among the Mien (Yao)in northern Thailand. J Ethnopharmacol.;123:335–342.

- 14. Céline, V., Pabon, A., Deharo, E., Albán-C. J., Estevez, Y., Lores, F., Augusto, R. R., Gamboa, D., Sauvain, M. and Castillo, D. and Bourdy, G. 2009. Medicinal plants from the Yanesha (Peru): Evaluation of the leishmanicidal and antimalarial activity of selected extracts. J. Ethnopharmacol.; 123:413–422.
- 15. Cai, Y., Sun, M. and Corke, H. 2001. Identification and distribution of simple and acylated betacyanins in the Amaranthaceae. J. Agri. and Food Chem.; 49(4):1971–1978.
- **16.** Khare, C.P. 2007. Indian Medicinal Plants: An Illustrated Dictionary. Springer.
- **17.** Mander, M. 1998. Marketing of indigenous medicinal plants in South Africa A case study in Kwa-Zulu Natal. Rome: FAO.
- Herdocia, Alfonso and Guido Lea. 1983. Minister of Health of Nicaragua.
- Vicente, T., Omar, M., Paola, F. Vidari, V., Chabaco, A. and Tomás, Z. 2007. An ethnobotanical survey of medicinal plants used in Loja and Zamora-Chinchipe, Ecuador. J Ethnopharmacol.; 111:63–81.
- 20. Schmidt, C., Fronza, M., Goettert, M., Geller, F., Luik, S., Flores, EMM., Bittencourt, C.F., Zanettie, G.D., Heinzmann, B.M., Laufer, S. and Merfort, I. 2009. Biological studies on Brazilian plants used in wound healing. J Ethnopharmacol. ;122:523–532. [PubMed]
- **21.** Cai, Y., Sun, M. and Corke, H. 2003. Antioxidant Activity of Betalains from Plants of the

Amaranthaceae. J Agric. Food Chem.; 51:2288–2294.

- **22.** Anthony, G., Paul, S., Kerry, B. and Matt, G. 2011. Principles and Practices of Naturopathic Botanical Medicine, Advanced Botanical Medicine. V3 CCNM Press, Toronto.
- **23.** Escop 2003. 'Thymi herba'' Monographs on the Medicinal uses of plant Drugs. Exeter,U.K: European scientific cooperative on phytotherapy.
- 24. Schmidt E., Bail S., Buchbauer G., Stoilova Ι., Atanasova Т.. Stoyanova A., Krastanov A., Jirovetz L. 2009. "Chemical composition, olfactory evaluation and antioxidant effects of essential oil from Mentha x piperita." Natural product communications. 4 (8):1107-1112.
- 25. Christopher, B. and Tevor, C. 2002. The American Horticultural Society: Encyclopedia of Plants & Flowers. New York, NY, USA: DK Publishing. p.605. ISBN 0-7894-8993-7.
- **26.** Bebell, S., M., Jonathan, T., Dwight L, M. 2008. Herb, Nutrient and Drug Interactions: Clinical Implications and Therapeutic Strategies.
- **27.** al Zuhair, H., el Sayeh, B., Ameen, H.A., et al. 1996. Pharmacological studies of cardamom oil in animals. Pharmacol Res; 34(1-2):79-82.
- **28.** J. S. Ajarem, J. S. and Ahmed, M. 1992. Effects of Perinatal Exposure of Cardamom (*Elettaria cardamomum*) on The Post- natal Development and Social Behavior of Mice Offspring. Journal of King Saud University, 4(2): 34-57.
- **29.** Abo Khalwa, and Kubo, J. 1987. Chemical Composition of the Essential Oil of Cardamom Seeds (*Elettaria cardamomum*), Proc. Saudi of Biology Science, 10: 297-305.

- Govindarajan, V.S., Narasimhan, S. Raghuveer K.G. and Lewis, S. 1982. Cardamom- Production, technology, chemistry and quality. CRC Critical Reviews in Food Science and Nutrition, 16: 229-305.
- **31.** A d'hiah, A. H. 1990. Immunogenetics Studies in Selected Human Diseases. Ph.D. Thesis. University of Newcastle upon Tyne, England.
- **32.** Metcalf, D., Begley, C. G., Jophnson, N. A., Vadas, M. A., Lopez, A. F., Williamson, D. J., Wong, G. G., Clark, S. C. And Wang, E. A. 1986. Biologic properties *in vitro* of a recombinant human granulocyte-macrophage colony-stimulating factor. Blood 67, 37-45.
- 33. Chaudhuri, D. and Sevanan, M. 2012. Investigation on Phytochamicals and antibacterial activity of the leaf and stem extracts of *Iresine herbstii*. Int. J. Pharm.Bio. Sci. Oct; 3(4): 697 705.
- **34.** Baht, A. and Bahc, B. S. 2005. A text book of organic chemistry. S. chand company LTD.
- **35.** WHO. 2005. Guidelines for laboratory and field testing of Mosquito larvicides. WHO/CDs/ WHOPES/ GCDPP/13.
- **36.** Cech, P.; and Lehrer, R.I. 1984. Heterogenicity of human neutrophil phagolysosome: Functional consequences for candidacial activity. Blood. 64: 147 – 15.
- **37.** Atta, A.H. and Alkofahi, A. S. 1998. Anti-nociceptive and antiinflammatory effects of some Jordanian medicinal plant extracts. Journal of ethnopharmacology, 60(2):117-24.
- **38.** Jeyakumar, E., Lawrence, R. and Pal, T. 2011. Comparative evaluation in the efficacy of peppermint (*Mentha piperita*) oil with standards antibiotics against

selected bacterial pathogens. Asian Pacific Journal of Tropical Biomedicine. S253-S257 .

- **39.** Awaad. M. H. H., Abdel-Alim, G. A., Sayed, K. S. S., Kawkab, A. A., Nada, A. A., Metwalli, A. S. Z. and Alkhalaf, A. N. 2010. Immunostimulant Effects of Essential Oils of Peppermint and Eucalyptus in Chickens, Pakistan Veterinary Journal, 30(2): 61-66.
- **40.** Majdalawieh, A. F., Carr, R.,I. 2010. In vitro investigation of the

potential immunomodulatory and anti-cancer activities of black pepper (*Piper igrum*) and cardamom (*Elettaria cardamomum*).

41. Acharya, A., Das. I., Singh, S. and Saha, T. 2010. Chemopreventive properties of indole- 3-carbinol, diindolylmethane and other constituents of cardamom against carcinogenesis.Recent Pat Food Nutr. Agric. 2(2):166-77.

الفعالية المناعية لبعض النباتات الشائعة

ز هراء حسين محمد قدري *

أنوار ادريس سليمان العساف*

*كلية التربية - ابن الهيثم للعلوم الصرفة/ جامعة بغداد/ قسم علوم الحياة

الخلاصة:

تم تحضير المستخلص الكحولي (بتراكيز مختلفة) لبعض الأنواع من النباتات الشائعة مثل: دم العاشق Iresine ولنعناع Elettaria cardamomum اضافة الى نبات الهيل Elettaria cardamomum ، حيث تم حضن الخلايا في عملية البلعمة باستخدام خميرة الخبز المعروفة Saccharomyces cervisiae ، حيث تم حضن الخلايا العدلى (متعددة أشكال النوى) للانسان مع تلك المستخلصات لفترات متعددة (15 دقيقة، 30 دقيقة و45 دقيقة) وقد حدد التأثير المحتمل لتلك النباتات في فعالية الالتهام وعلى معدل ابتلاع الخميرة من قبل الخلايا البلعمية ، اذ لوحظ ود4 دقيقة و45 دقيقة) وقد ريادة المعروفة Saccharomyces cervisiae ، 30 دقيقة و45 دقيقة) وقد دد التأثير المحتمل لتلك النباتات في فعالية الالتهام وعلى معدل ابتلاع الخميرة من قبل الخلايا البلعمية ، اذ لوحظ ريادة معنوية (10.00) في مستوى البلعمة باستخدام مستخلص نبات (دم العاشق) بزيادة الوقت وزيادة التركيز، وقد يعزى السبب في ذلك لكون النبات يمتلك خصائص سمية ومضادة للالتهاب أما فيما يخص نبات (النعناع)، وقد ارتفعت مستويات البلعمة باستخدام مستخلص نبات (دم العاشق) بزيادة الوقت وزيادة التركيز، وقد يعزى السبب في ذلك لكون النبات يمتلك خصائص سمية ومضادة للالتهاب أما فيما يخص نبات (النعناع)، وقد ارتفعت مستويات البلعمة أيضا بزيادة الوقت والتركيز ولا سيما عند استخدام النبات بتركيز 20% و25%. وومحتمل ان يعود ذلك للدور الفعال لهذا النبات كمضاد للبكتريا والفايروسات ومضاد قوي للأكسدة والاورام. وومحتمل ان يعود ذلك للدور الفعال لهذا النبات كمضاد للبكتريا والفايروسات ومضاد قوي للأكسدة والاورام. وومحتمل ان يعود ذلك للدور الفعال لهذا النبات كمضاد للبكتريا والفايروسات ومضاد قوي للأكسدة والاورام. وومحتمل ان يعود ذلك للدور الفعال لهذا النبات كمضاد للبكتريا والفايروسات ومضاد قوي للأكسدة والاورام. وومحتمل ان يعود ذلك للدور الفعال لهذا النبات كمضاد للبكتريا والفايروسات ومضاد قوي للأكسدة والاورام. وومحتمل ان يعود ذلك للدور الفعال لهذا النبات كمضاد للبكتريا والفايروسات ومضاد قوي للأكسدة والاورام. وومحتمل ان يعود ذلك للدور الفعال لهذا النبات كمضاد للبكتريا والفايروسات ومضاد قوي للأكسدة والاورام. وومات في محتا وي لمعنوي في في عالية البلعمة باستخدام المستخلص باليرييز قارم ألورام. وومات في محل القوما في ممدي القدم معنوي في في علم معناي وومات ومحاص في مدى الخاص ف