Production of Slime Layer by *Staphylococcus epidermidis* Isolated From Corneal Infection

Munira CH. Ismail *

Fais I. Ali** Sin

Sinai W. Mohammed*

Received 31, May, 2009 Accepted 1, July, 2010

Abstract:

A total of 37 *Staphylococcus epidermidis* isolates, isolated from corneal scraping of patients with bacterial keratitis and 20 isolates from healthy eyes (as control) (all isolates, isolated from, Ibn Al- Haietham eye hospital / Baghdad), were tested for slime production, 52.63% of all isolates were positive-slime production (23 isolates from patients and 7 isolates from controls). It was found that positive-slime producing *S. epidermidis* were exhibited a high resistance to antibiotics as compared to negative-slime producing isolates.

Key words: Slime Layer, Staphylococcus epidermidis, Keratitis.

Introduction:

The coagulase-negative staphylococci (CONS) are widely distributed over the surface of human body, where they constitute the majority of the common nasal bacterial micro flora. Among the CONS, Staphylococcus epidermidis is the most frequently isolated species most common and the species responsible for infection [1]. One important property of S. epidermidis which is responsible for its persistence and / or opportunistic invasion in the tissues is its ability to produce slime [2, 3]. Slime not only helps the organism in adhesion to host cells, but also protects it from phagocytosis and from the action of antibiotics [4].

Despite being important ocular pathogens, S. epidermidis have so far received little attention in ophthalmology. The purpose of this determine study was to identify, antibiotic susceptibility and slime production of S. epidermidis isolated from patients with bacterial Keratitis.

Materials and Methods:

Bacteria: isolates of *S. epidermidis* (CONS) from 57 patients, who attended the Ibn Al-Haietham Eye Hospital, Baghdad, during October 2001 to October 2002.

Subjects: of the 57 patients, 37 had come for treatment and investigation of keratitis and 20 (from healthy eyes) served as controls.

Methods:

1. Corneal scrapings: were performed in each case under the slit– lamp biomicroscope. The scrapings were taken from the base and the margins of the ulcer and were then smeared on glass slides for Gram staining. The specimens also inoculated at 37 °C on to blood and chocolate agar.

2. Conjunctival swabs: were obtained from 20 control subjects. In order to obtain an ideal swab for culture.

3. Isolation and identification of bacteria: culture material from corneal scrapings and swab were routinely plated on the following media: Blood agar and Chocolate agar with 10% CO₂ (at 37°C for 24 hr.). In positive-culture cases: all bacteria (*Staphylococcus spp.*) were identified by API-system (API-staph) (Bio mereiux).

^{*} Tropical Biological Research Unit / Science College

^{**} Ibn Al- Haietham Eye Hospital / Ministry of Health

4. Slime-production test: isolates were tested for slime production with the use of a technique described by Christensen *et al.*, 1982 [5]. In brief, a loop of organisms from a pure growth on blood agar plate was inoculated onto 5ml of trypticase soy broth (oxoid), and incubated at 37°C for 48 hr. the contents of the tubes were aspirated and the tubes were stained with 1% safranine (BDH) for 7min. A visible safranine stained film lining the wall of the tubes indicated a positive test.

susceptibility: 5. Antibiotic The susceptibility of S. epidermidis isolates were performed by Kirby-Bauer disc diffusion assay [6]. We choose five effective antibiotics against most strains of corneal pathogens. The antibiotics and their concentrations/disc were: (μg) [Ciprofloxacin (5 µg), Gentamicin (10 μg), Cephalothin (30 μg), Rifampicin (5 µg) and Chloramphenicol (30 µg)] (oxoid).

6. Statistical analysis: Chi-Square test was used in the analysis of results [7].

Results and Discussion:

Slime test: A total of 57 CONS isolates were studied: 23 (62.16%) isolates from patients and 7(35%) from were positive-slime controls production. thus 30 (52.63%) of 57 isolates were positive-slime producers. Antibiotics susceptibility: The results of antibiotic susceptibility testing of S. epidermidis isolates were isolated from patients with bacterial keratitis are given in Table (1)& Figure (1). The results showed a high resistance to rifampicin and chloramphenicol [16 (69.5%) and 14 (60.8%) respectively]. While cephalothin, ciprofloxacin and gentamicin had a low resistance [7 (30.43%). (34.78%) 8 and 10 (43.47%)] respectively in positiveslime producing isolates. While negative-slime producing isolates exhibited a low resistant against antibiotics. there was significant difference $(X^2 = 25.8, P < 0.05)$ between them. Table (2) & Figure (2) shows antibiotic resistance of control isolates. Results showed a low resistant against all antibiotics which are used particularly in negative-slime production isolates there was significant difference ($X^2 = 42.7$, P < 0.05) between them.

Slime layer has been documented to be one of the virulence markers of *S. epidermidis* because of the close association of slime producing strains with infections related to indwelling medical devices including intraocular lenses [8]. Positive-slime producing isolates were isolated in high numbers from patients as compared to control [9].

We found a positive association between positive-slime production and resistance to antibiotics. This is supported by observation made presently in some reports [2, 9, 10], that slime not only helped the organism to colonize the host tissues, but it also protected it from the action of antibiotics.

Cephalothen and Ciprofloxacin a new broad spectrum antibiotics were found an effective agent in this study, they shows a low resistance for most of S. epidermidis isolates and bacteria had no chance to develop resistance to these antibiotics was expected to be slow because they requires chromosomal mutation, and resistance can not to be transferred by plasmid mediated mechanisms [11]. The results are similar to those reported by other authors [12], who found that slime layer and multi drug resistance were the important virulence factors of S. epidermidis in bacterial keratitis.

Studies on biofilms have shown that *S. epidermidis* is the most frequently isolated slime-producing CNS and is also the most common cause of nosocomial infections in patients with catheters, medical implants or other invasive devices [13].

Table	(1):	Antibiotic	resistance	of
keratit	is iso	lates (No. of	f isolates=37	7).

Antibiotics	Positive Slime production (NO. of isolates=23)	%	Negative Slime production (NO. of isolates=14)	%
Chloramphenicol	14	60.8	2	15
Ciprofloxacin	8	34.78	2	15
Cephalothen	7	30.43	5	35
Rifampicin	16	69.5	4	29
Gentamycin	10	43.47	4	29

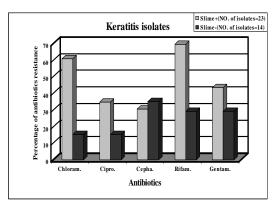


Fig. (1): Antibiotic resistance pattern of Keratitis isolates.

Table	(2):	Antibiotic	resistance of		
control isolates (No. of isolates=20).					

Antibiotics	Positive Slime production (NO. of isolates=7)	%	Negative Slime production (NO. of isolates=13)	%
Chloramphenicol	4	59	5	39
Ciprofloxacin	3	39	3	25
Cephalothen	3	44	5	39.5
Rifampicin	2	29.5	4	31
Gentamycin	2	29	3	23

In conclusion, our findings showed that the slime layer, was responsible for resistance to antibiotics

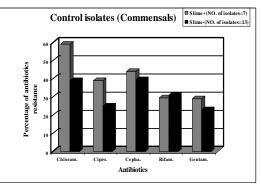


Fig. (2): Antibiotic resistance pattern of Control isolates.

References:

- 1. O'Gara, J.P., Humphreys, H. 2001. *Staphylococcus epidermidis* biofilms: importance and implications (Review article). J. Med. Microbiol. 50: 582-587.
- 2. Nayak, N., Satpathy, G. 2000. Slime production as a virulence factor in *Staphylococcus epidermidis* isolated from bacterial keratitis. Indian J. med. Res. 111: 6-10.
- **3.** Beachey, E.H. 1981. Bacterial adherence: adhesion-receptor interactions mediating the attachment of bacteria to mucosal surface. J. Infect. Dis. 143: 325-345.
- **4.** Quie, P.G., Belani, k. k. 1987. Coagulase-negative staphylococcal adherence. J. Infect. Dis. 156: 543-547.
- Christensen, G. D., Simpson, W. A., Bisno, A. L. and Beachey, E. H. 1982. Adherence of slimeproducing strains of *Staphylococcus epidermidis* to smooth surfaces. Infect. immune. 37: 318-326.
- 6. Baur, A. W., Sheris, J. G. and Truck, M. 1966. Antibiotic susceptibility testing by

standardized single disk method. Am. J. Clin. Path. 43: 493-496.

- Negi, K. S. 2008. Biostatistics with latest MCQs. A.I.T.B.S. publishers, 2nd. India pp146.
- 8. Raskin, E. M., Speaker, M. G., McCormick, S. A., Wong, D., Menikoff, S. A. and Pelton. H. k. 1993. Influence of hepatic materials on adherence of staphylococci to intraocular lenses. Arch. Ophthalmol, 111: 250-253.
- Peters, G., Locci, R. and Pulverer, G. 1980. Adherence and growth of coagulase-negative staphylococci on surface of intravenous catheters. J. Infect. Dis. 146: 479-482.
- **10.** Arslan, S., Özkardes, F. 2007. Slime production and antibiotic susceptibility in staphylococci

isolated from clinical samples. Mem. Inst Oswaldo Cruz, Rio de Janeiro. 102(1): 29-33

- **11.** Tuft, S. J., Metheson, M. 2000. In vitro antibiotic resistance in bacterial keratitis in London. Br. J. Opthalmol. 84: 687-691.
- 12. Nayak, N., Nag, T.C., Satpathy, G. & Ray, S.B. 2007. Ultrastructural analysis of slime positive & slime negative *Staphylococcus epidermidis* isolates in infectious keratitis. Indian J Med Res 125: 767-771.
- **13.** Oliveira, A., Cunha, M. L. R. S. 2008. Bacterial Biofilms with Emphasis on Coagulase-Negative Staphylococci. J. Venom. Anim. Toxins Incl. Trop. Dis., 14, 4, 588

أنتاج الطبقة اللزجة من بكتريا المكورات العنقوديه للجلد المعزولة من أصابة القرنية

منيرة اسماعيل جلوب * فائز اسماعيل على * * سيناء وليد محدسعيد *

* وحدة الابحاث البايولوجية للمناطق الحارة / كلية العلوم ** مستشفى ابن الهيثم للعيون / وزارة الصحة

الخلاصة:

من المجموع الكلي لـ 37 عزلة لبكتريا المكورات العنقوديه للجلد عزلت من قرنيات مرضى مصابين بالتهاب القرنية البكتيري و20عزلة عزلت من قرنيات عيون غير مصابة (السيطرة) (جميع العزلات, عزلت من مستشفى ابن الهيثم للعيون / بغداد), ظهر 52.63 % منها (23 عزلة من المصابين و7 من غير المصابين) منتجة للطبقة اللزجة وقد أظهرت النتائج إن جميع العزلات المنتجة لهذه الطبقة لها مقاومة عالية تجاه المصادت الحيوية مقارنة بالعزلات غير المنتجة.