



Effects of perinatal exposure to bisphenol A or S in EAE model of multiple sclerosis

Brigitta Bonaldo^{1,2} · Antonino Casile^{1,2,3} · Francesca Montarolo^{1,4,5} · Martina Bettarelli¹ · Francesca Napoli⁶ · Stefano Gotti^{1,2} · GianCarlo Panzica^{1,2} · Marilena Marraudino^{1,2}

Received: 23 June 2022 / Accepted: 23 January 2023
© The Author(s) 2023

Abstract

Epidemiological studies support the idea that multiple sclerosis (MS) is a multifactorial disease, overlapping genetic, epigenetic, and environmental factors. A better definition of environmental risks is critical to understand both etiology and the sex-related differences of MS. Exposure to endocrine-disrupting compounds (EDCs) fully represents one of these risks. EDCs are natural or synthetic exogenous substances (or mixtures) that alter the functions of the endocrine system. Among synthetic EDCs, exposure to bisphenol A (BPA) has been implicated in the etiology of MS, but to date, controversial data has emerged. Furthermore, nothing is known about bisphenol S (BPS), one of the most widely used substitutes for BPA. As exposure to bisphenols will not disappear soon, it is necessary to clarify their role also in this pathological condition defining their role in disease onset and course in both sexes. In this study, we examined, in both sexes, the effects of perinatal exposure to BPA and BPS in one of the most widely used mouse models of MS, experimental autoimmune encephalomyelitis (EAE). Exposure to bisphenols seemed to be particularly deleterious in males. In fact, both BPA- and BPS-treated males showed anticipation of the disease onset and an increased motoneuron loss in the spinal cord. Overall, BPA-treated males also displayed an exacerbation of EAE course and an increase in inflammation markers in the spinal cord. Analyzing the consequences of bisphenol exposure on EAE will help to better understand the role of both xenoestrogens and endogenous estrogens on the sexually dimorphic characteristics of MS.

Keywords Endocrine-disrupting chemicals · Environmental risk factor · BPA · BPS · Experimental autoimmune encephalomyelitis

We want to dedicate this paper in the memory of our mentor and friend GianCarlo Panzica who passed away on July 21, 2022: we miss you, and we will miss you a lot.

✉ Brigitta Bonaldo
brigitta.bonaldo@unito.it

¹ Neuroscience Institute Cavalieri Ottolenghi (NICO), Regione Gonzole, 10-10043 Orbassano, Turin, Italy

² Department of Neuroscience “Rita Levi-Montalcini”, University of Turin, Via Cherasco 15, Turin 10126, Italy

³ School of Pharmacy, Pharmacology Unit, University of Camerino, Via Madonna delle Carceri, 9, Camerino 62032, Italy

⁴ Neurobiology Unit, Neurology, CReSM (Regional Referring Center of Multiple Sclerosis), San Luigi Gonzaga University Hospital, Orbassano, Italy

⁵ Department of Molecular Biotechnology and Health Sciences, University of Turin, Turin, Italy

⁶ Department of Oncology, University of Turin, San Luigi Hospital, Orbassano, Turin, Italy

Introduction

Multiple sclerosis (MS) is a chronic autoimmune disease of the central nervous system (CNS), characterized by perivascular infiltration of inflammatory cells, demyelination, axonal loss, and gliosis (McGinley et al. 2021; Thompson et al. 2018). MS has a different prevalence in the sexes and the female-to-male *ratio* varies between 1.5:1 and 2.5:1 or more (i.e., 2.3–3.5:1), as reported in several studies (Harbo et al. 2013; Ortona et al. 2016; Voskuhl 2020). This data, indicating an increase in MS among women but not men, have brought about in-depth studies of sex differences in the nervous system or immune system, possibly due to genetic differences, gonadal hormones, environmental factors, and/or lifestyle (Alfredsson and Olsson 2019; Ascherio and Munger 2016; Hedström et al. 2021; Olsson et al. 2017).

Overall, women exhibit more robust responses of the immune system than men, and this is thought to influence

the different susceptibility to develop autoimmune diseases (Di Florio et al. 2020; Rubtsova et al. 2015). This quick increase could reveal unrecognized environmental or nutritional changes (Alfredsson and Olsson 2019; Ascherio and Munger 2016). Sex's effect on the clinical features of MS is unclear. Still, there are indications that women usually have earlier onset of disease, a lower prevalence of primary progressive disease course, and generally show less progression to disability than men (Bergamaschi 2007; Harbo et al. 2013; Olsson et al. 2017; Ramien et al. 2016; Voskuhl 2020).

Among the main factors affecting these sex differences, gonadal hormones and different responses to environmental factors appear to be particularly significant (Rubtsova et al. 2015). The role of sex hormones in MS appears to be limited to women; however, the situation is much more tangled (Ortona et al. 2016; Ramien et al. 2016; Shepherd et al. 2020). Because of the presence of hormone receptors on immune cells, sex hormones can affect the activities of the immune system and possibly influence different aspects of autoimmune diseases (i.e., risk, activity, and progression) (Moulton 2018). In general, estrogen and prolactin act as enhancers of humoral immunity, whereas testosterone and progesterone act as immunosuppressants (Pierdominici et al. 2010; Shepherd et al. 2020). The effects mediated by the estrogens depend on the dose: lower levels stimulate specific immune activities, while higher levels, for example, those observed in pregnancy, inhibit them (Merz et al. 2022; Whitacre et al. 1999). Hence, the different effects mediated by the sex hormones depend not only on the concentration but also on the cell type and receptor subtype expressed in a given cell type (Ortona et al. 2016).

In view of the wide range of effects that sex hormones play within the CNS (Spence and Voskuhl 2012), it is possible to hypothesize these hormones have a role in MS, acting not only on immune system cell populations but also on the CNS ones (Spence and Voskuhl 2012; Ysrraelit and Correale 2019): some endogenous and exogenous estrogens are useful in MS patients both during pregnancy (Gilli et al. 2010; Merz et al. 2022) and when using oral contraceptives (Chen et al. 2020; Sena et al. 2012). However, their action appears to be effective only in the early stages of MS (Spence and Voskuhl 2012). Estrogen can also act on astrocytes which modulate neuronal death and inflammation through several pathways. The action of estrogens is mediated through the estrogen receptor α (ER α), which reduces inflammation, demyelination, and axonal loss (Spence et al. 2011; Tiwari-Woodruff et al. 2007; Yilmaz et al. 2019), while the estrogen receptor β (ER β) has a more controversial role. It is not involved in endogenous estrogen protection but can respond to exogenous ligands, protecting against demyelination and axonal loss and stimulating endogenous myelination (Crawford et al. 2010; Spence and Voskuhl 2012). In women, studies are underway for estriol treatment with anti-inflammatory, neuroprotective, and immunomodulatory effects (Voskuhl et al. 2016). The

role of ER β in MS is less known, but it is an attractive therapeutic candidate in association with some anti-inflammatory drugs. Indeed, estriol binds to ER α and ER β weaker than estradiol, but it shows a higher affinity to ER β compared to ER α , the main cause of estrogenic effects on breast cancer and cardiovascular diseases (Voskuhl et al. 2016). Interestingly, a more recent study performed in the experimental autoimmune encephalomyelitis (EAE) model of MS demonstrates that, in this model, along with inflammation and demyelination in the spinal cord, it presents inflammation of the hypothalamic tissue in both females and males. This inflammation results in the downregulation of different genes in males and females, leading to sex-specific changes downstream in the hypothalamic-pituitary axis (HPA) (Milosevic et al. 2020), supporting the idea that EAE partially also models sex-specific characteristics of the disease (Ryan and Mills 2021).

Along with endogenous estrogens, the organism can be exposed to natural (phytoestrogens and mycoestrogens) or synthetic (xenoestrogens) compounds with estrogenic activity, i.e., endocrine-disrupting compounds (EDCs). There are thousands of chemicals, such as plastics, dust, pesticides, herbicides, and medical and/or dietary components, that have been classified as EDCs (Gore et al. 2015; Ho et al. 2022). Exposure to EDCs is more perilous if it occurs during peculiar "critical periods" of life (e.g., intrauterine, perinatal, juvenile, or puberty periods) when organisms are more sensitive to hormonal action. Nevertheless, exposure to EDCs in adulthood also can alter physiology (Frye et al. 2012; Kahn et al. 2020).

Synergic/additive effects might be displayed by environmental and endogenous estrogens, potentially also affecting the immune response. Moreover, there is numerous *in vitro* and *in vivo* evidence that these compounds may exert immunotoxic effects (Chighizola and Meroni 2012; Ortona et al. 2016). Many studies also highlighted the issues due to EDC exposure on steroid hormone receptors in the developing CNS, showing substantial effects on mRNA levels, protein expression, and neuroanatomical and functional consequences of altered receptor action (Gore et al. 2015).

There is evident crosstalk between the CNS and the immune system. In fact, on the one hand, lymphoid organs present receptors for neuropeptides, neurotransmitters, and hormones, and on the other one, immune system activation causes changes in hypothalamic, autonomic, and endocrine functions (Bahadar et al. 2015; Del Rey and Besedovsky 2017). Moreover, immune functions are also modulated by the interplay between the autonomic and the neuroendocrine systems via the pituitary-adrenal axis, which represents a crucial link between CNS-immune interaction and autoimmune diseases (Bahadar et al. 2015). Therefore, exposure to EDCs could increase the risks or intensify the aggressiveness of autoimmune diseases affecting the CNS, above all MS (Ascherio et al. 2012).

Bisphenol A (BPA) is a compound mainly used for the production of clear and tough plastics, utilized for the manufacturing of many common consumer goods (Abraham and Chakraborty 2020). It represents one of the most known and studied EDCs (Abraham and Chakraborty 2020). Exposure to BPA has been described to alter the function of some systems, including the immune system (Kimber 2017; McDonough et al. 2021; Rochester 2013). In fact, in mice, BPA exposure resulted in augmented production of cytokine and antibodies and decreased numbers of regulatory T cells, even if many of the reports focused on adult, as opposed to gestational, exposure to this compound (Kimber 2017; Rochester 2013). The risk to public health due to BPA exposure was recognized by EFSA in 2015 (EFSA, 2015): the tolerable daily intake (TDI) for BPA was reduced from 50 to 4 µg/kg body weight/day, but the BPA substitutes, such as bisphenol S (BPS), have no specific limitations, even if they seem to have comparable, or even more alarming, endocrine disrupting properties as the BPA (den Braver-Sewradj et al. 2020; Eladak et al. 2015; Gramec Skledar and Peterlin Masic 2016; McDonough et al. 2021; Rochester and Bolden 2015).

Data on the effects of BPA exposure on different MS animal models appear to be particularly controversial, either excluding (Krementsov et al. 2013) or supporting (Brinkmeyer-Langford et al. 2014; Rogers et al. 2017) its potential effects on peculiar aspects of the disease.

Considering the increasing exposure to EDCs, and in particular to BPs, and that the environmental components which have been implicated in the etiology of MS, it is important to properly examine their role in the onset and course of the disease. Thus, taking the advantage of the EAE mouse model of MS, this study aimed to better understand the consequences of perinatal exposure to BPA and to evaluate and compare the one of BPS, in mice of both sexes. We assessed daily the severity of the disease both by carrying out a clinical evaluation and testing the motor symptoms, evaluating the performance with the rotarod. Finally, we evaluated the degree of inflammation and the motoneuron loss thanks to histological investigations.

Materials and methods

Animals

Adult C57BL/6 J mice from our colony at the Neuroscience Institute Cavalieri Ottolenghi (originally purchased from Envigo, S. Pietro al Natisone, Udine, Italy) were housed in standard conditions in 45 × 25 × 15 cm polypropylene mouse cages at 22 ± 2 °C, under 12:12 light–dark cycle (lights on at 08:00 AM). Food (standard mouse chow 4RF21, Mucedola srl, Settimo Milanese, Italy) and water were provided

ad libitum. One male and two female mice (3-month-old) were housed together to achieve a successful mating, assessed by the evaluation of the presence of the vaginal plug (assumed as gestational day 0, GD0) (Hasegawa et al. 2017).

The experimental design conforms to the ARRIVE guidelines originally published by Kilkenny et al. in 2010 (2010).

Treatments

BPA (Sigma-Aldrich, 239,658, CAS 80–05-7) or BPS (Sigma-Aldrich, 103,039, CAS 80–09-1) were prepared for oral administration by dissolving them in corn oil (Sigma-Aldrich, C8267). Twelve pregnant dams were assigned randomly to three experimental groups: oil-treated dams (receiving only vehicle, corn oil; $n=4$), BPA-treated dams (receiving 4 µg/kg BW/day of BPA, corresponding to the European TDI; $n=4$), and BPS-treated dams (receiving 4 µg/kg BW/day of BPS; $n=4$). The dose was calculated daily according to the dams' body weight, recorded with an electronic precision balance (Mod. Kern-440-47N, resolution 0.1 g).

We decided to test the same dose for both BPA and BPS to allow a precise comparison of the effects of the two bisphenols. Moreover, at present, although BPS is one of the most used BPA substitutes and it has already been detected in environmental and human samples (Catenza et al. 2021), at present, no user guidelines are available. Dams were treated starting at GD0, throughout pregnancy and lactation, until weaning of the offspring at postnatal day 28 (PND28). To resemble human exposure conditions, the daily treatment or the vehicle was given orally to the dams by means of a pipette to minimize the dams' stress (Bo et al. 2016; Palanza et al. 2002). This type of administration allowed us to perform a perinatal treatment (covering both prenatal and postnatal critical windows of development) (Neier et al. 2019) on the offspring. In fact, it is known that both BPA and BPS can pass first through the placenta and then into the milk during lactation (Cimmino et al. 2020; Mao et al. 2020).

Litters were reduced to 8 pups at birth to obtaining an equal number of pups of both sexes, sexed via the measurement of the anogenital distance (AGD) (Manno 3rd, 2008). The pups were weaned at PND28 and housed in monosexual groups of 4 mice. They were monitored weekly until adulthood (PND56), when the experimental procedures were performed.

EAE induction and clinical evaluation

Chronic EAE has been induced in 8 weeks-old mice of both sexes ($n=9$ /group) (Constantinescu et al. 2011). Briefly, mice have been immunized by subcutaneous immunization under the rostral part of the flanks and at the base of the tail with 300 µl of 200 µg/mouse of myelin oligodendrocyte glycoprotein (MOG_{35–55}; Espikem, Florence, Italy) in

incomplete Freund's adjuvant containing 8 mg/ml of *Mycobacterium tuberculosis* (strain H37Ra; Difco Laboratories Inc., St Henry, Detroit, Michigan, USA) and two intravenous injections of 500 ng of Pertussis toxin (Duotech, Milan, Italy) the day of immunization and 48 h after (i.e., 2 days post-immunization, dpi) (Montarolo et al. 2014, 2015).

Body weight (BW) and clinical score (CS; 0, healthy; 1, limp tail; 2, ataxia and/or paresis of hind limbs; 3, paralysis of hind limbs and/or paresis of forelimbs; 4, tetra paralysis; 5, dying or death) have been recorded daily by a blind investigator. This analysis allows to evaluate the clinical differences in the onset and progression of the disease (Constantinescu et al. 2011; Montarolo et al. 2014, 2015).

Furthermore, since the rotarod test could be used as a more quantitative and precise clinical assessment of the disease course than the clinical score alone (van den Berg et al. 2016), mice underwent a rotarod performance test daily (Mouse RotaRod, Ugo Basile 47600, Milan, Italy), starting from 6 dpi until the time of the sacrifice (28 dpi). The 1–5 dpi period has been used to train the animals in the use of the device and to obtain reference values (baseline). The test consisted of a single 300 s session during which the rod speed was increased linearly from 4 to 40 rpm (van den Berg et al. 2016). When the mouse was not capable of maintaining its balance and fell off the device, it fell and triggered a sensor, and the time (s) was recorded (latency).

Within the four weeks (0–28 dpi) of EAE follow-up, we also monitored the food intake (FI, g/animal/day; once a week), and, in females, we checked the estrous cycle for at least 2 cycles, evaluating the vaginal cytology smears (McLean et al. 2012). In particular, we focused on the evaluation of the estrous cycle between the first and the second week post-immunization (considering, in particular, the acute phase of the EAE) (Constantinescu et al. 2011) because it has been demonstrated that the immunization procedure causes the greatest alterations of the estrous cycle within this period (Rahn et al. 2014).

Fixation and tissue sampling

At 28 dpi, mice were sacrificed by deep irreversible anesthesia (intraperitoneal injection of Zoletil 80 mg/kg/ Rompum 10 mg/kg) and transcardially perfused with 4% paraformaldehyde (PFA) solution. Spinal cords were removed and stored in a 4% PFA solution for 24 h, followed by several washes in 0.01 M saline phosphate buffer (PBS). Finally, they were embedded in paraffin. Paraffin-embedded spinal cords were cut in the transversal plane at 10 μ m thickness with a microtome and collected on gelatin-coated slides. The plane of sectioning was oriented to match the drawings corresponding to the transversal sections of the mouse spinal cord atlas (Watson et al. 2009).

Histological evaluations

Ten- μ m-thick paraffin-embedded sections on gelatin-coated slides, representative of the entire spinal cord, were stained with Hematoxylin–Eosin (Montarolo et al. 2014, 2015) or Cresyl Violet (Nissl Staining) (Morales et al. 2006), to detect the presence of the perivascular inflammatory infiltrates (PvIIs; $n=9$ animals/group) and the motoneurons (MNs; $n=5$ animals/group), respectively. The presence of PvIIs and MNs loss is assessed as signs of the disease (Bolton and Smith 2015; Constantinescu et al. 2011; Frezel et al. 2016; Gushchina et al. 2018).

Briefly, the staining was performed as follows: after deparaffinization, sections were stained with the Hematoxylin and Eosin procedure by using Sigma-Aldrich (St. Louis, Missouri, USA) reagents, or they were Nissl-stained with 0.1% Cresyl Violet (Sigma-Aldrich, St. Louis, Missouri, USA). Dehydrated sections were covered with New-Entellan mounting medium (Merck, Milano, Italy).

Quantitative analysis

Neuropathological findings were quantified in 10 complete cross-sections of the spinal cord per mouse representative of whole spinal cord levels. The sections were acquired and analyzed with the NeuroLucida software connected to an E-800 Nikon microscope with a 20 \times objective (Glaser and Glaser 1990). The number of PvIIs or MNs was calculated and expressed as the number of PvIIs or MNs per mm².

The representative images in Figs. 3 and 4 were acquired with a NIKON DS-U1 digital camera (Software of acquisition: NIS-Element AR 2.10) connected to a NIKON Eclipse 90i microscope (Nikon Italia S.p.S., Firenze, Italy). Images were digitized by using a 10 \times or 40 \times objective for the acquisition.

Statistical analysis

BW, FI, CS, and latency at the rotarod performance test were analyzed by three-way (sex, treatment, and time as independent variables) analysis of variance (ANOVA). All other quantitative data were analyzed by two-way (sex and treatment as independent variables) ANOVA with SPSS 27 statistic software (SPSS Inc., Chicago, USA). If the ANOVA was significant, the post hoc analysis was performed using Tukey's HSD test. Comparison between the estrous cycle evaluations was performed using the Student's *t* test. Differences were considered statistically significant for values of $p \leq 0.05$. Data are shown as mean \pm SEM (mean standard error).

Results

Effects of BPs on body weight, food intake, and estrous cycle of EAE-affected mice

The analysis of BW showed no effects of the treatments ($F_{(2,48)}=0.472, p=0.627$; Fig. 1a). However, the sex differences in BW ($F_{(2,48)}=1.498, p=0.011$; Fig. 1a) were always maintained ($p<0.001$; Fig. 1a) despite the treatments and the disease progression. Finally, all the experimental groups

displayed a similar BW trend, showing a decrease (Fig. 1a) in the acute phase (within the second week post-immunization) of the disease due to the increased EAE severity.

The analysis of FI did not show any differences between the groups ($F_{(2,8)}=0.950, p=0.485$; Fig. 1b) but highlighted a significant decrease among all groups within the first and second post-immunization week ($p<0.001$), due to the increased EAE severity which caused difficulties in reaching the food placed in an upper container in the cage. To avoid further stress on the animals, the food was then placed on

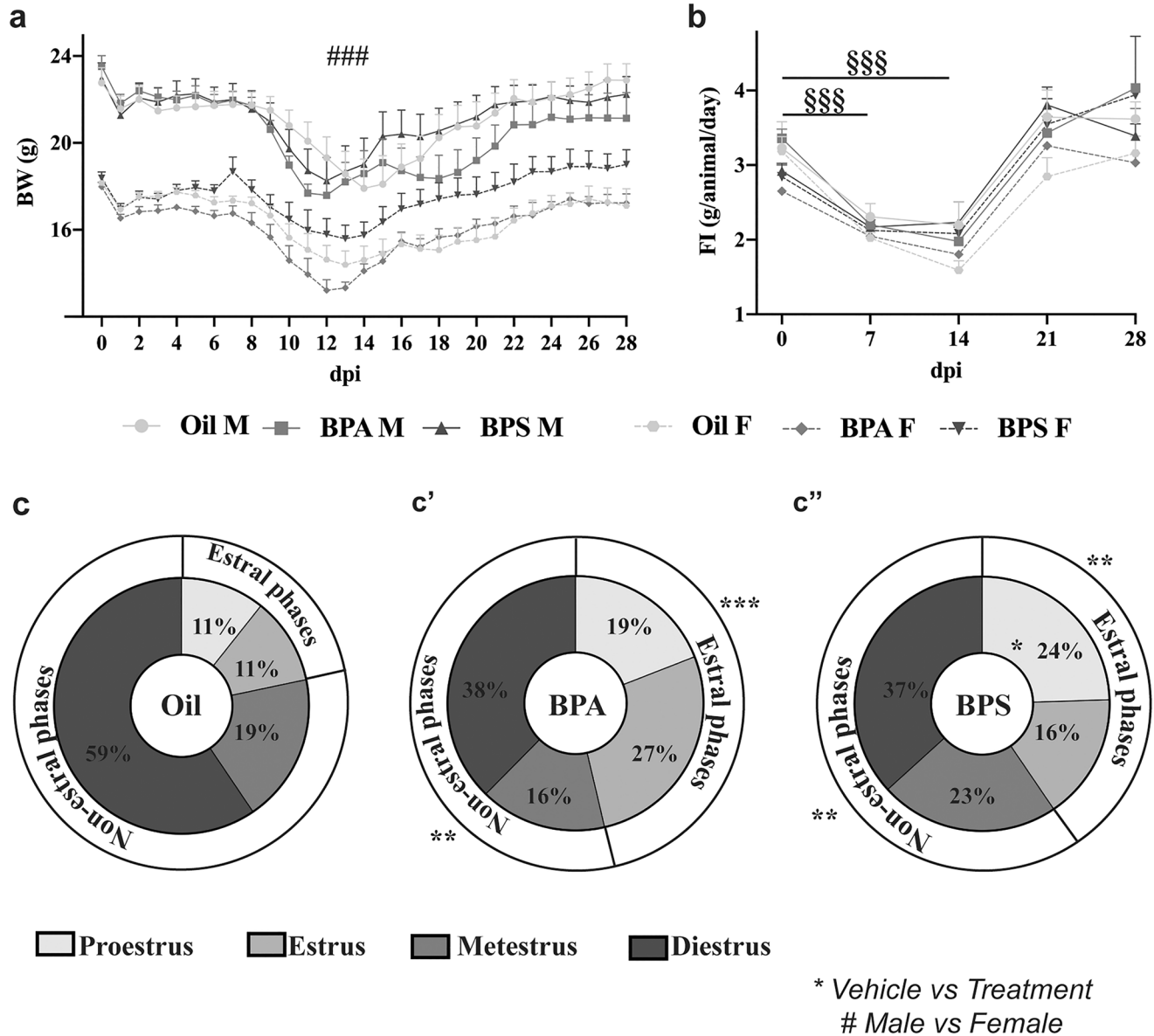


Fig. 1 Effects of BPA and BPS exposure on body weight, food intake, and estrous cycle of EAE-affected mice. Daily body weight (a) and weekly food intake (b) evaluation from the day of immunization (0 dpi) until the sacrifice (28 dpi) of the animals. Mean percentage of time spent in the different phases of the estrous cycle, assessed by vaginal

cytology smears, in the (c) oil-, (c') BPA-, or (c'') BPS-treated EAE-affected females. Data are expressed as mean ± SEM. Statistical analysis revealed a significant effect for $p \leq 0.05$ (*=vehicle vs. treatment; #=male vs. female; §=comparison between different timepoints). BW, body weight; FI, food intake; dpi, day post-immunization

the cage ground, so the FI returned to starter levels, and the BW partially recovered (Fig. 1b).

The analysis of the estrous cycle in EAE-affected females revealed that both BPA and BPS treatments caused an alteration in the time spent in the different phases of the estrous cycle (Fig. 1c, c', c''). Both BPS-treated females spent more time in estral phases (proestrus and estrus; oil vs. BPA, $p < 0.001$, Fig. 1c'; oil vs. BPS, $p = 0.004$, Fig. 1c'') and less in non-estral ones (metestrus and diestrus; oil vs. BPA, $p = 0.008$, Fig. 1c'; oil vs. BPS, $p = 0.004$, Fig. 1c'') compared to oil-treated ones (Fig. 1c). In particular, BPS-treated females spent more time in proestrus compared to oil-treated females ($p = 0.031$; Fig. 1c'').

Effects of BPs on EAE onset and course

The clinical evaluation of the EAE course was assessed daily, assigning both the CS and evaluating the rotarod performance (as latency of fall) in all experimental groups (Fig. 2).

First, the analysis of the daily CS showed some significant differences in the disease course ($F_{(2,48)} = 6.481$, $p = 0.003$; Fig. 2a). That is due to an increased CS among the BPA-treated males, which also displayed higher maximum reached CS (Fig. 2b) and a significantly higher cumulative CS ($p = 0.004$, Fig. 2c) compared to oil-treated males. Interestingly, BPS-treated males displayed a significant increase in CS, compared to oil-treated ones, only at 7 dpi ($p = 0.044$) and 8 dpi ($p = 0.004$). Furthermore, both BPA- ($p = 0.027$) and BPS- ($p < 0.001$) treated males showed anticipation in disease onset (Fig. 2d) compared to the oil-treated ones, which disrupted the sexual dimorphism existing among the oil-treated mice where the females displayed an anticipated onset compared to males ($p = 0.011$, Fig. 2d).

The analysis of the daily rotarod performance showed some significant differences among groups ($F_{(2,48)} = 4.069$, $p = 0.023$; Fig. 2e) due to the fact that BPS-treated males displayed lower latency at 7 dpi ($p = 0.020$) compared to oil-treated ones. Among oil-treated groups, females showed lower latency at 27 dpi ($p = 0.020$) and 28 dpi ($p = 0.043$) compared to males.

Effects of BPs on histological parameters in the spinal cord

The presence of PvIIs, observed in Hematoxylin–Eosin-stained sections of the spinal cord (*representative sections in Fig. 3a–b''*), and the MNs loss, measured in Cresyl Violet stained sections (*representative sections in Fig. 3c–d''*), are assessed as signs of disease severity (Fig. 4).

The quantification of PvIIs (Fig. 4a, a') highlighted some significant differences among the groups ($F_{(5,54)} = 12.656$, $p < 0.001$; Fig. 4c). First, oil-treated females displayed

Fig. 2 Effects of BPA and BPS exposure on EAE clinical evaluations. **a** Daily clinical score evaluation from the day of immunization (0 dpi) until the sacrifice (28 dpi) of the animals. Mean **b** maximum clinical score and **c** cumulative clinical score reached by the oil- (*left*), BPA- (*center*), or BPS- (*right*) treated EAE-affected male (*left side of the graph*) and female (*right side of the graph*) mice. **d** Mean dpi of disease onset in the oil- (*left*), BPA- (*center*), or BPS- (*right*) treated EAE-affected male (*left side of the graph*) and female (*right side of the graph*) mice. **e** Daily evaluation of rotarod performance (expressed as latency of fall) from 6 dpi (the 0 represents the baseline values obtained within the first 5 days of the test) until the sacrifice (28 dpi) of the animals. Data are expressed as mean \pm SEM. Statistical analysis revealed a significant effect for $p \leq 0.05$ (* = vehicle vs. treatment; § = BPA vs. BPS; # = male vs. female). CS, clinical score; dpi, day post-immunization

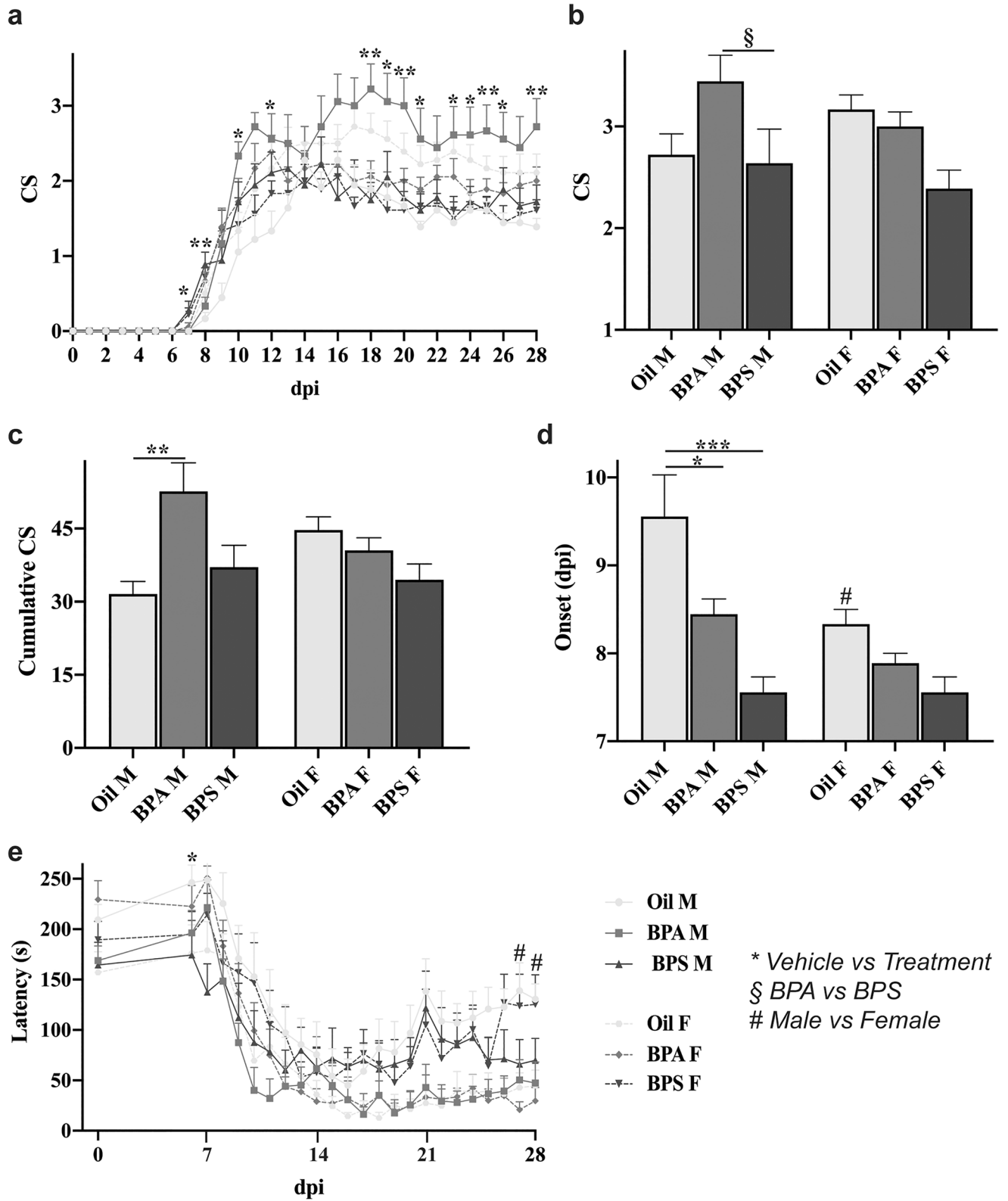
significantly higher values compared to oil-treated males ($p = 0.013$, Fig. 4c). This sexual dimorphism was disrupted in the treated groups. In fact, BPA-treated males showed a significant increase compared to oil-treated males ($p < 0.001$, Fig. 4c), while BPA-treated females showed a significant decrease compared to oil-treated females ($p = 0.006$, Fig. 4c), causing an opposite and extreme sexual difference ($p < 0.001$, Fig. 4c). On the other hand, BPS-treated males showed no difference compared to oil-treated ones ($p = 0.999$, Fig. 4c), while BPS-treated females showed a significant decrease compared to oil-treated females ($p = 0.033$, Fig. 4c), thus causing the disappearance of the sexual dimorphism in BPS-treated animals.

The analysis of MNs loss (Fig. 4b, b') showed some significant differences among the groups ($F_{(5,24)} = 3.189$, $p = 0.024$; Fig. 4d). In particular, both BPA- ($p = 0.044$) and BPS- ($p = 0.027$) treated males displayed a decreased number of MNs compared to the oil-treated ones (Fig. 4d), while we found no differences among the females (Fig. 4d).

Discussion

MS is a multifactorial disease that overlaps with genetic, epigenetic, and environmental factors (Ascherio et al. 2012; Ascherio and Munger 2016; Waubant et al. 2019). Thus, defining the environmental risks is a crucial turning point to better understand the great variability of the diseases in terms of etiology, progression, and sexual prevalence (Ascherio et al. 2012; Ascherio and Munger 2016; Hedström et al. 2021).

In our study, we highlighted, in the EAE model of MS, how exposure to either BPA or BPS during a critical period of development affected the disease onset and course differentially in the two sexes. BPs treatment seemed to be particularly serious in males. In fact, BPA-treated males displayed the greatest alterations, showing a more aggressive disease in terms of anticipation of disease onset, clinical score, inflammation, and motoneuron loss in the spinal cord. Furthermore, also BPS-treated males displayed anticipation



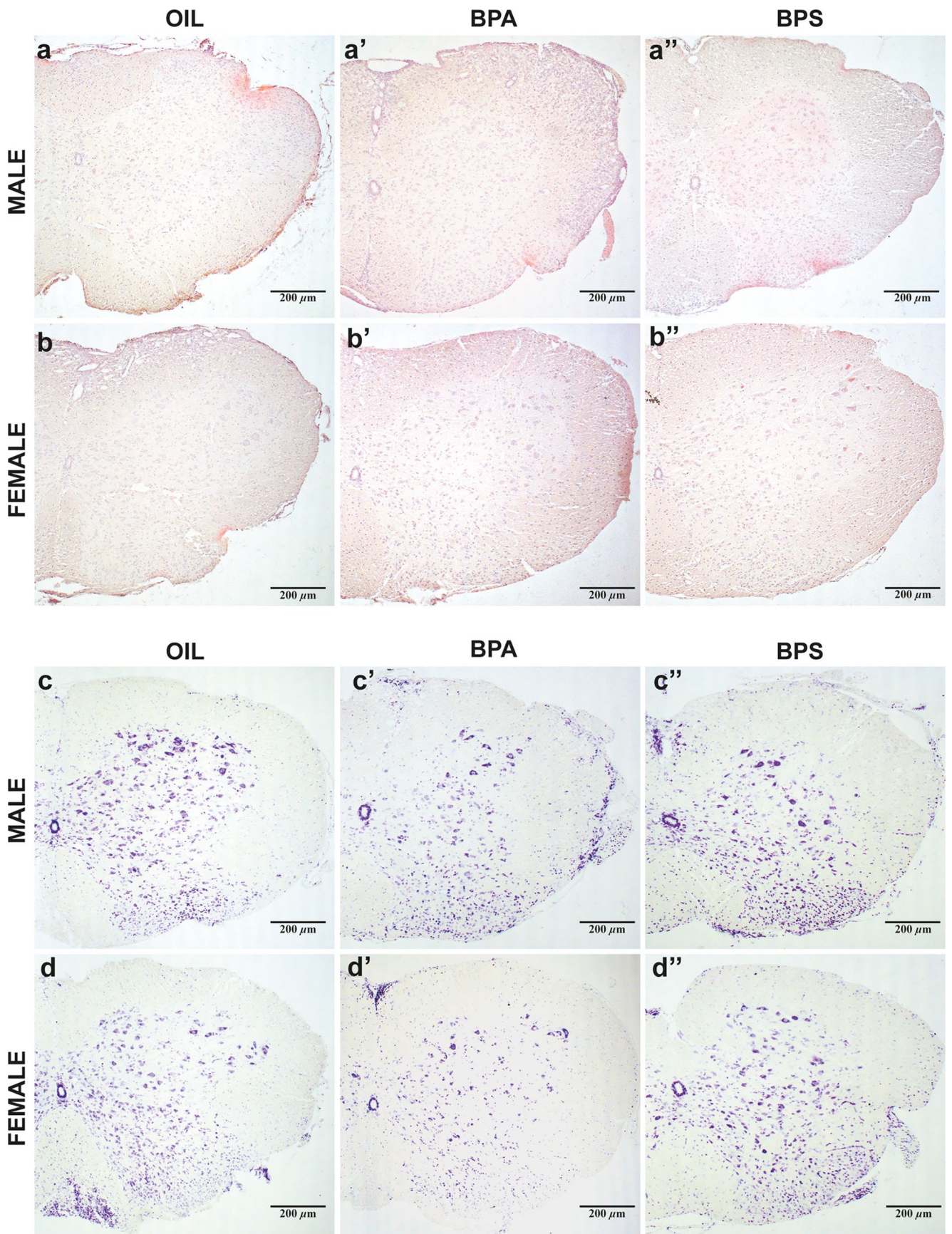


Fig. 3 Histological staining of EAE-affected mice's spinal cord. Representative images of transversal spinal cord sections stained with Hematoxylin–Eosin (*upper panel*) to assess the presence of PvIIs, of oil- (*left*), BPA- (*center*), or BPS- (*right*) treated male (**a, a', a''**) or female (**b, b', b''**) mice. Representative images of transversal spinal cord sections stained with Cresyl Violet (*lower panel*) to assess the number of MNs, of oil- (*left*), BPA- (*center*), or BPS- (*right*) treated male (**c, c', c''**) or female (**d, d', d''**) mice. Scale bar=200 μ m (10 \times magnification). PvIIs, perivascular inflammatory infiltrates; MNs, motoneurons

of the disease onset and a higher motoneuron loss in the spinal cord compared to oil-treated males. Among females, we did not notice any significant differences in the evaluated disease-related parameters, except for fewer PvIIs in the spinal cord, which did not come along with a recovered number of motoneurons.

Sex's effect on the clinical features of MS is unclear. However, there are indications that women usually have an earlier onset of disease, a slightly lower prevalence of primary progressive disease course, and a minor progression to disability than men (Bergamaschi 2007; Harbo et al. 2004; Ramien et al. 2016; Voskuhl 2020). So, even if MS is more prevalent in women compared to men, men generally develop a more aggressive and progressive form of the disease (Harbo et al. 2013; Ortona et al. 2016). As environmental exposures play a role in determining those differences (Alfredsson and Olsson 2019; Ascherio and Munger 2016), our results support the idea that exposure to BPs could lead to an exacerbation of the diseases in males.

Data on the effects of BPA exposure on different MS animal models appear to be controversial, while we have no information about the BPS. A 2013 study in EAE-affected female mice did not support the hypothesis that gestational BPA exposure contributes to the increasing female MS risk (Krementsov et al. 2013). On the contrary, another 2013 study investigated, both in male and female mice, the effects of perinatal BPA exposure on Theiler's-virus-induced demyelination (TVID), another murine model of MS, showing that perinatal BPA exposure is linked to a decreased level of viral antibodies, an anticipation of the onset of TVID symptoms, an increased inflammation in both the spinal cord and digestive tract, and an intensified changes in immune-related gene expression caused by viral infection (Brinkmeyer-Langford et al. 2014). The controversial results could be linked to the fact that MS is modeled using different animal models, which reflect only partially the characteristic of the disease (Lassmann and Bradl 2017; Procaccini et al. 2015) and to the different periods and ways of administration and dose selected for BPA treatment (Panzica and Melcangi 2016).

However, our results are in line with the work of Rogers et al. (2017), which demonstrated that gestational exposure to BPA lowered the threshold for EAE onset, especially in

male mice. It is interesting to notice that we observed the deleterious effects of the exposure at a lower dose (4 μ g/kg BW/day vs. 1 or 3 mg/kg BW/day). Moreover, we also highlighted the pathological signs of the disease at spinal cord levels. Finally, for the first time, we described the effect of BPS exposure in a murine model of MS. A more recent paper shows that subchronic exposure to BPA in mice led to the deregulation of inflammatory cytokines and oxidative stress, possibly linked to neurotoxicity, axonal damage, and myelin degeneration (Khan et al. 2019). This mechanism could underlie the motoneuron loss in an immune system-independent way, which seemed to be the case, especially in BPS-treated males, displaying a significant decrease in the number of motoneurons without any increase in PvIIs compared to oil-treated ones. Increasing *in vitro* evidence describes the neurotoxic potential of BPS (Meng et al. 2021; Pang et al. 2019), while the inflammatory potential of BPS appears to be less compared to the one of BPA (Kobayashi et al. 2010; Profita et al. 2021).

In females, we did not observe any statistically relevant effects of both BPs on disease onset and course, except for a reduction in PvIIs in the spinal cord. This could be due to the effects of BPs on the estrous cycle. In fact, both BPA and BPS led to an increase in time spent in estral phases in treated females compared to oil-treated ones. This phase is characterized by increasing levels of estrogens which are known to exert an anti-inflammatory, neuroprotective effect (Spence and Voskuhl 2012). In particular, BPS-treated females spend more time in proestrus, which has been described as protective at least against neurological symptoms in the EAE models (Rahn et al. 2014).

It is important to underline the fact that the observed alterations are present in adult animals following perinatal exposure. Perinatal BPA and BPS could cause an impairment either in immune system cell populations or in motoneurons which is maintained in adulthood or may result in an altered response to stimuli. Moreover, both BPs presumably would accumulate within some compartments of the organism and face a slow release (Charisiadis et al. 2018; Venisse et al. 2019). Finally, different effects observed in males and females could be due also to the fact that males appeared to be particularly vulnerable to developmental exposure to BPs (Kobayashi et al. 2010).

Studying the effects of exogenous compounds with estrogenic activity can contribute to a better understanding of the role of endogenous hormones and identify the mechanisms underlying sex differences in MS (Harbo et al. 2013; Ortona et al. 2016; Ramien et al. 2016; Voskuhl 2020). Furthermore, a better definition of environmental risks is necessary. Investigating the effects of BPs exposure can support better determining their deleterious properties, which may

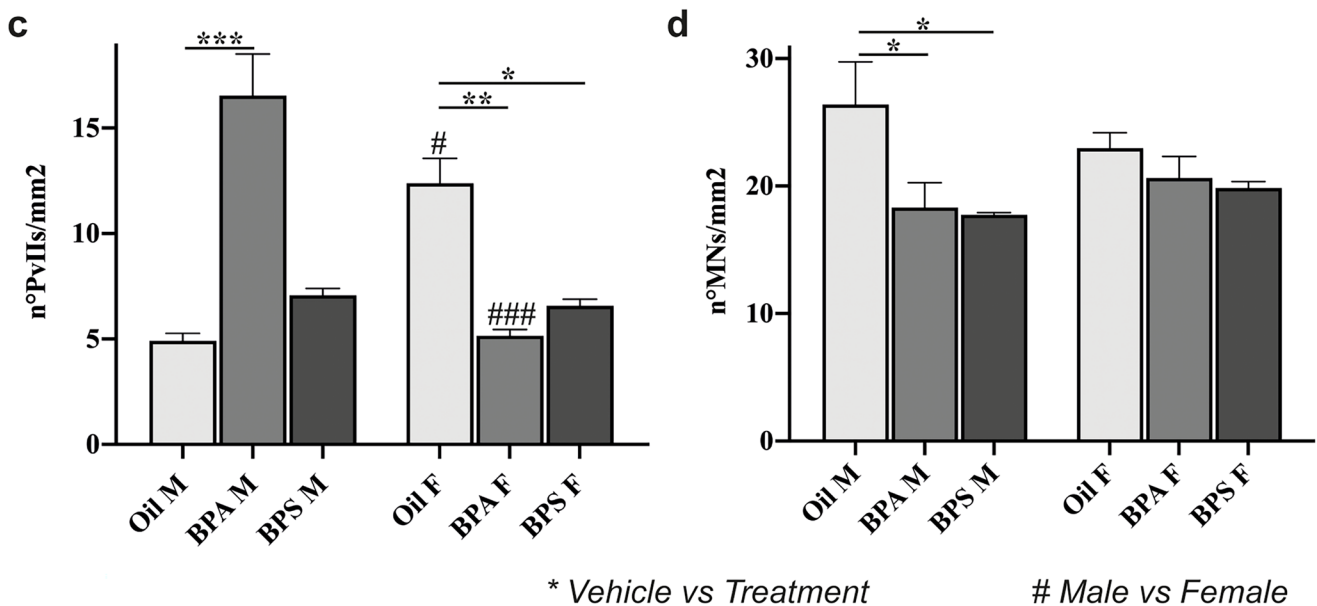
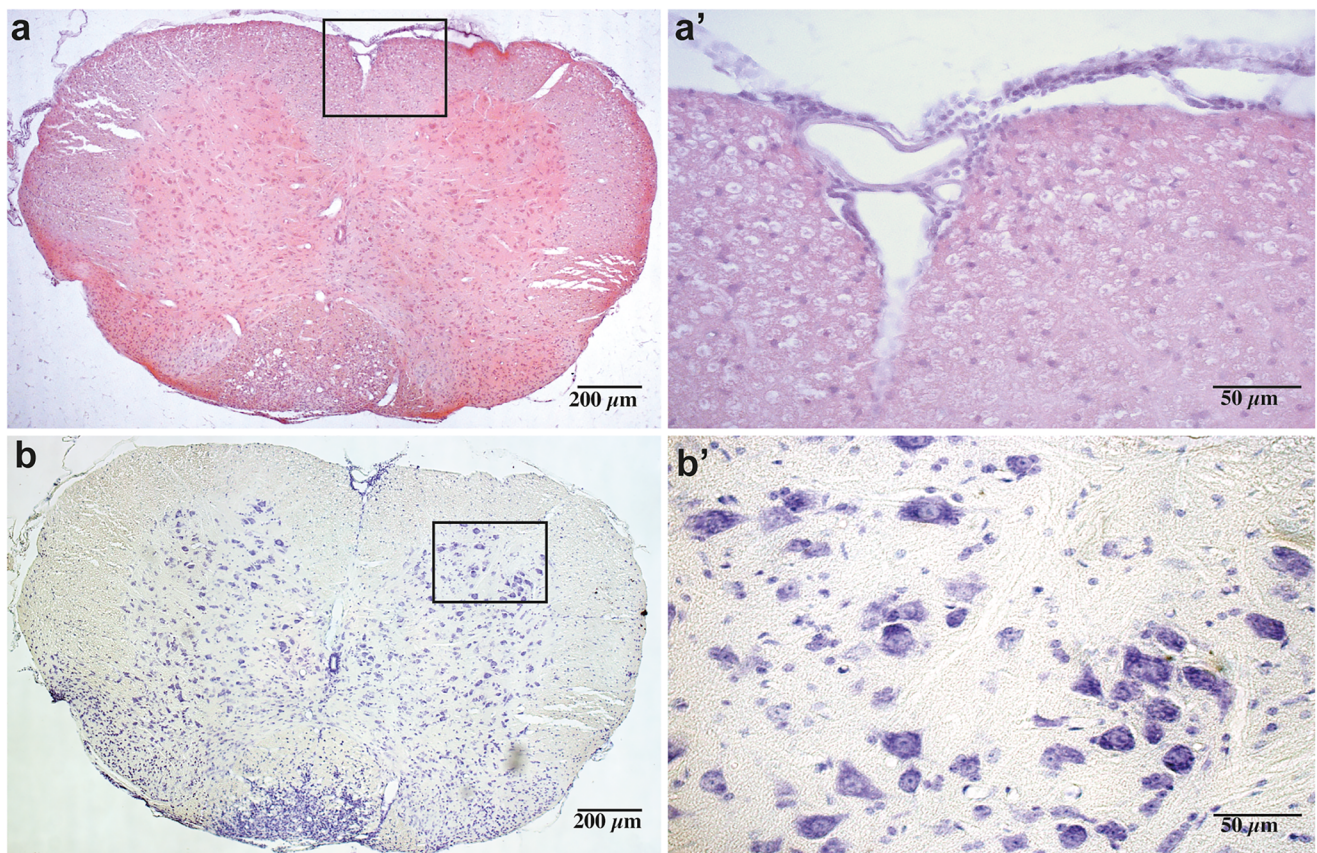


Fig. 4 Analysis of perivascular inflammatory infiltrates and motoneuron loss in spinal cord sections of oil-, BPA- or BPS-treated EAE-affected mice. Representative images of (a, a') Hematoxylin-Eosin and (b, b') Nissl staining in a transversal section of spinal cord from an oil-treated EAE-affected male mice. Analysis of the (c) presence of PvIIs and (d) motoneuron loss in the spinal cords of oil- (left), BPA- (center), or BPS-

(right) treated EAE-affected male (left side of the graph) and female (right side of the graph) mice. Data are expressed as mean ± SEM. Statistical analysis revealed a significant effect for $p \leq 0.05$ (*=vehicle vs. treatment; #=male vs. female). Scale bar=200 μm (10×magnification) or 50 μm (40×magnification). PvIIs, perivascular inflammatory infiltrates; MNs, motoneurons

be particularly relevant in pathological conditions. Additionally, defining BPs as a real risk of developing or worsening MS can help devise new strategies to reduce exposure for sensitive people or patients (e.g., avoiding specific environments, do not use plastic food/water containers, etc.).

Acknowledgements Preliminary results were presented during the XXVIII national meeting of the Italian Group for the Study of Neuro-morphology (GISN), the Federation of European Neuroscience Societies (FENS) Virtual Forum 2020, and the 94th National Congress of the Italian Society for Experimental Biology (SIBS).

Author contribution BB designed and performed experiments, analyzed data, and wrote the draft. AC, FM, MB, and FN performed experiments. SG, GCP, and MM revised the draft and wrote the paper.

Funding Open access funding provided by Università degli Studi di Torino within the CRUI-CARE Agreement. This work has been supported by Ministero dell'Istruzione, dell'Università e della Ricerca-MIUR project Dipartimenti di Eccellenza 2018–2022 to Department of Neuroscience Rita Levi Montalcini; Cavaliere-Ottolenghi Foundation, Orbassano, Italy; University of Torino, RiLo to GCP. BB is fellow of INFRA-P, Piedmont Region (n.378–35). MM was a fellow of Prof. G.C. Bergui (2020) and is now a fellow of the Fondazione Umberto Veronesi (2021–2023).

Data availability All data generated or analyzed during this study are included in this article. Further enquiries can be directed to the corresponding author.

Declarations

Ethics approval Animal care and handling were according to the European Union Council Directive of 22nd September 2010 (2010/63/UE); all the procedures reported in the present study were approved by the Italian Ministry of Health (407/2018-PR) and by the Ethical Committee of the University of Torino (Project n° 360384).

Conflict of interest The authors declare no competing interests.

Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>.

References

- Abraham A, Chakraborty P (2020) A review on sources and health impacts of bisphenol A. *Rev Environ Health* 35(2):201–210. <https://doi.org/10.1515/revh-2019-0034>
- Alfredsson L, Olsson T (2019) Lifestyle and environmental factors in multiple sclerosis. *Cold Spring Harb Perspect Med* 9(4). <https://doi.org/10.1101/cshperspect.a028944>
- Ascherio A, Munger KL (2016) Epidemiology of multiple sclerosis: from risk factors to prevention—an update. *Semin Neurol* 36(2):103–114. <https://doi.org/10.1055/s-0036-1579693>
- Ascherio A, Munger KL, Lunemann JD (2012) The initiation and prevention of multiple sclerosis. *Nat Rev Neurol* 8(11):602–612. <https://doi.org/10.1038/nrneurol.2012.198>
- Bahadar H, Abdollahi M, Maqbool F, Baeri M, Niaz K (2015) Mechanistic overview of immune modulatory effects of environmental toxicants. *Inflamm Allergy Drug Targets* 13(6):382–386. <https://doi.org/10.2174/1871528114666150529103003>
- Bergamaschi R (2007) Prognostic factors in multiple sclerosis. *Int Rev Neurobiol* 79:423–447. [https://doi.org/10.1016/S0074-7742\(07\)79019-0](https://doi.org/10.1016/S0074-7742(07)79019-0)
- Bo E, Farinetti A, Marraudino M, Sterchele D, Eva C, Gotti S, Panzica G (2016) Adult exposure to tributyltin affects hypothalamic neuropeptide Y, Y1 receptor distribution, and circulating leptin in mice. *Andrology* 4(4):723–734. <https://doi.org/10.1111/andr.1222210.1111/andr.12222>. Epub 2016 Jun 16
- Bolton C, Smith P (2015) Defining and regulating acute inflammatory lesion formation during the pathogenesis of multiple sclerosis and experimental autoimmune encephalomyelitis. *CNS Neurol Disord: Drug Targets* 14(7):915–935. <https://doi.org/10.2174/1871527314666150716103629>
- Brinkmeyer-Langford C, Rodrigues A, Kochan KJ, Haney R, Rassu F, Steelman AJ, Young C, Riggs P, Storts R, Meagher MW, Welsh CJ (2014) Consequences of perinatal bisphenol A exposure in a mouse model of multiple sclerosis. *Autoimmunity* 47(1):57–66. <https://doi.org/10.3109/08916934.2013.832220>
- Catenza CJ, Farooq A, Shubear NS, Donkor KK (2021) A targeted review on fate, occurrence, risk and health implications of bisphenol analogues. *Chemosphere* 268:129273. <https://doi.org/10.1016/j.chemosphere.2020.129273>
- Charisiadis P, Andrianou XD, van der Meer TP, den Dunnen WFA, Swaab DF, Wolffenbuttel BHR, Makris KC, van Vliet-Ostapchouk JV (2018) Possible obesogenic effects of bisphenols accumulation in the human brain. *Sci Rep* 8(1):8186. <https://doi.org/10.1038/s41598-018-26498-y>
- Chen CS, Krishnakumar T, Rowles W, Anderson A, Zhao C, Do L, Bove R (2020) Comparison of MS inflammatory activity in women using continuous versus cyclic combined oral contraceptives. *Mult Scler Relat Disord* 41:101970. <https://doi.org/10.1016/j.msard.2020.101970>
- Chighizola C, Meroni PL (2012) The role of environmental estrogens and autoimmunity. *Autoimmun Rev* 11(6–7):A493–501. <https://doi.org/10.1016/j.autrev.2011.11.027>
- Cimmino I, Fiory F, Perruolo G, Miele C, Beguinot F, Formisano P, Oriente F (2020) Potential mechanisms of bisphenol A (BPA) contributing to human disease. *Int J Mol Sci* 21(16). <https://doi.org/10.3390/ijms21165761>
- Constantinescu CS, Farooqi N, O'Brien K, Gran B (2011) Experimental autoimmune encephalomyelitis (EAE) as a model for multiple sclerosis (MS). *Br J Pharmacol* 164(4):1079–1106. <https://doi.org/10.1111/j.1476-5381.2011.01302.x>
- Crawford AC, Riggins RB, Shajahan AN, Zwart A, Clarke R (2010) Co-inhibition of BCL-W and BCL2 restores antiestrogen sensitivity through BECN1 and promotes an autophagy-associated necrosis. *PLoS One* 5(1):e8604. <https://doi.org/10.1371/journal.pone.0008604>
- Del Rey A, Besedovsky HO (2017) Immune-neuro-endocrine reflexes, circuits, and networks: physiologic and evolutionary implications. *Front Horm Res* 48:1–18. <https://doi.org/10.1159/000452902>
- den Braver-Sewradj SP, van Spronsen R, Hessel EVS (2020) Substitution of bisphenol A: a review of the carcinogenicity, reproductive toxicity, and endocrine disruption potential of alternative substances. *Crit Rev Toxicol* 50(2):128–147. <https://doi.org/10.1080/10408444.2019.1701986>

- Di Florio DN, Sin J, Coronado MJ, Atwal PS, Fairweather D (2020) Sex differences in inflammation, redox biology, mitochondria and autoimmunity. *Redox Biol* 31:101482. <https://doi.org/10.1016/j.redox.2020.101482>
- EFSA (European Food Safety Authority) (2015) Scientific opinion on the risks to public health related to the presence of bisphenol A (BPA) in foodstuffs. *EFSA J* 13(1):3978. <https://doi.org/10.2903/j.efsa.2015.3978>
- Eladak S, Grisin T, Moison D, Guerquin MJ, N'Tumba-Byn T, Pozzi-Gaudin S, Benachi A, Livera G, Rouiller-Fabre V, Habert R (2015) A new chapter in the bisphenol a story: bisphenol S and bisphenol F are not safe alternatives to this compound. *In Fertil Steril* 103(1):11–21. <https://doi.org/10.1016/j.fertnstert.2014.11.005>
- Frezel N, Sohet F, Daneman R, Basbaum AI, Braz JM (2016) Peripheral and central neuronal ATF3 precedes CD4+ T-cell infiltration in EAE. *Exp Neurol* 283(Pt A):224–234. <https://doi.org/10.1016/j.expneurol.2016.06.019>
- Frye CA, Bo E, Calamandrei G, Calza L, Dessi-Fulgheri F, Fernandez M, Fusani L, Kah O, Kajta M, Le Page Y, Patisaul HB, Venerosi A, Wojtowicz AK, Panzica GC (2012) Endocrine disruptors: a review of some sources, effects, and mechanisms of actions on behaviour and neuroendocrine systems. *J Neuroendocrinol* 24(1):144–159. <https://doi.org/10.1111/j.1365-2826.2011.02229.x>
- Gilli F, Lindberg RL, Valentino P, Marnetto F, Malucchi S, Sala A, Capobianco M, di Sapio A, Sperli F, Kappos L, Calogero RA, Bertolotto A (2010) Learning from nature: pregnancy changes the expression of inflammation-related genes in patients with multiple sclerosis. *PLoS One* 5(1):e8962. <https://doi.org/10.1371/journal.pone.0008962>
- Glaser JR, Glaser EM (1990) Neuron imaging with NeuroLucida—a PC-based system for image combining microscopy. *Comput Med Imaging Graph* 14(5):307–317. [https://doi.org/10.1016/0895-6111\(90\)90105-k](https://doi.org/10.1016/0895-6111(90)90105-k)
- Gore AC, Chappell VA, Fenton SE, Flaws JA, Nadal A, Prins GS, Toppari J, Zoeller RT (2015) Executive summary to EDC-2: the Endocrine Society's Second Scientific Statement on endocrine-disrupting chemicals. *Endocr Rev* 36(6):593–602. <https://doi.org/10.1210/er.2015-1093>
- Gramec Skledar D, Peterlin Masic L (2016) Bisphenol A and its analogs: do their metabolites have endocrine activity? *Environ Toxicol Pharmacol* 47:182–199. <https://doi.org/10.1016/j.etap.2016.09.014>
- Gushchina S, Pryce G, Yip PK, Wu D, Pallier P, Giovannoni G, Baker D, Bo X (2018) Increased expression of colony-stimulating factor-1 in mouse spinal cord with experimental autoimmune encephalomyelitis correlates with microglial activation and neuronal loss. *Glia* 66(10):2108–2125. <https://doi.org/10.1002/glia.23464>
- Harbo HF, Gold R, Tintore M (2013) Sex and gender issues in multiple sclerosis. *Ther Adv Neurol Disord* 6(4):237–248. <https://doi.org/10.1177/1756285613488434>
- Harbo HF, Lie BA, Sawcer S, Celius EG, Dai KZ, Oturai A, Hillert J, Lorentzen AR, Laaksonen M, Myhr KM, Ryder LP, Fredrikson S, Nyland H, Sørensen PS, Sandberg-Wollheim M, Andersen O, Svejgaard A, Edland A, Mellgren SI, ... Spurkland A (2004) Genes in the HLA class I region may contribute to the HLA class II-associated genetic susceptibility to multiple sclerosis. *Tissue Antigens* 63(3):237–247. <https://doi.org/10.1111/j.0001-2815.2004.00173.x>
- Hasegawa A, Mochida K, Ogonuki N, Hirose M, Tomishima T, Inoue K, Ogura A (2017) Efficient and scheduled production of pseudopregnant female mice for embryo transfer by estrous cycle synchronization. *J Reprod Dev* 63(6):539–545. <https://doi.org/10.1262/jrd.2017-068>
- Hedström AK, Hillert J, Olsson T, Alfredsson L (2021) Factors affecting the risk of relapsing-onset and progressive-onset multiple sclerosis. *J Neurol Neurosurg Psychiatry* 92(10):1096–1102. <https://doi.org/10.1136/jnnp-2020-325688>
- Ho V, Pelland-St-Pierre L, Gravel S, Bouchard MF, Verner MA, Labrèche F (2022) Endocrine disruptors: challenges and future directions in epidemiologic research. *Environ Res* 204(Pt A):111969. <https://doi.org/10.1016/j.envres.2021.111969>
- Kahn LG, Philippat C, Nakayama SF, Slama R, Trasande L (2020) Endocrine-disrupting chemicals: implications for human health. *Lancet Diabetes Endocrinol* 8(8):703–718. [https://doi.org/10.1016/S2213-8587\(20\)30129-7](https://doi.org/10.1016/S2213-8587(20)30129-7)
- Khan J, Salhotra S, Goswami P, Akhter J, Jahan S, Gupta S, Sharma S, Banerjee BD, Parvez S, Gupta S, Raisuddin S (2019) Bisphenol A triggers axonal injury and myelin degeneration with concomitant neurobehavioral toxicity in C57BL/6J male mice. *Toxicology* 428:152299. <https://doi.org/10.1016/j.tox.2019.152299>
- Kilkenny C, Browne WJ, Cuthill IC, Emerson M, Altman DG (2010) Improving bioscience research reporting: the ARRIVE guidelines for reporting animal research. *PLoS Biol* 8(6):e1000412. <https://doi.org/10.1371/journal.pbio.1000412>
- Kimber I (2017) Bisphenol A and immunotoxic potential: a commentary. *Regul Toxicol Pharmacol* 90:358–363. <https://doi.org/10.1016/j.yrtph.2017.08.022>
- Kobayashi K, Ohtani K, Kubota H, Miyagawa M (2010) Dietary exposure to low doses of bisphenol A: effects on reproduction and development in two generations of C57BL/6J mice. *Congenit Anom (kyoto)* 50(3):159–170. <https://doi.org/10.1111/j.1741-4520.2010.00279.x>
- Krementsov DN, Katchy A, Case LK, Carr FE, Davis B, Williams C, Teuscher C (2013) Studies in experimental autoimmune encephalomyelitis do not support developmental bisphenol a exposure as an environmental factor in increasing multiple sclerosis risk. *Toxicol Sci* 135(1):91–102. <https://doi.org/10.1093/toxsci/kft141>
- Lassmann H, Bradl M (2017) Multiple sclerosis: experimental models and reality. *Acta Neuropathol* 133(2):223–244. <https://doi.org/10.1007/s00401-016-1631-4>
- Manno FA 3rd (2008) Measurement of the digit lengths and the anogenital distance in mice. *Physiol Behav* 93(1–2):364–368. <https://doi.org/10.1016/j.physbeh.2007.09.011>
- Mao J, Jain A, Denslow ND, Nouri MZ, Chen S, Wang T, Zhu N, Koh J, Sarma SJ, Sumner BW, Lei Z, Sumner LW, Bivens NJ, Roberts RM, Tuteja G, Rosenfeld CS (2020) Bisphenol A and bisphenol S disruptions of the mouse placenta and potential effects on the placenta-brain axis. *Proc Natl Acad Sci USA* 117(9):4642–4652. <https://doi.org/10.1073/pnas.1919563117>
- McDonough CM, Xu HS, Guo TL (2021) Toxicity of bisphenol analogues on the reproductive, nervous, and immune systems, and their relationships to gut microbiome and metabolism: insights from a multi-species comparison. *Crit Rev Toxicol* 51(4):283–300. <https://doi.org/10.1080/10408444.2021.1908224>
- McGinley MP, Goldschmidt CH, Rae-Grant AD (2021) Diagnosis and treatment of multiple sclerosis: a review. *JAMA* 325(8):765–779. <https://doi.org/10.1001/jama.2020.26858>
- McLean AC, Valenzuela N, Fai S, Bennett SAL (2012) Performing vaginal lavage, crystal violet staining, and vaginal cytological evaluation for mouse estrous cycle staging identification. *J Vis Exp* 67:4389. <https://doi.org/10.3791/4389>
- Meng L, Liu J, Wang C, Ouyang Z, Kuang J, Pang Q, Fan R (2021) Sex-specific oxidative damage effects induced by BPA and its analogs on primary hippocampal neurons attenuated by EGCG. *Chemosphere* 264(Pt 1):128450. <https://doi.org/10.1016/j.chemosphere.2020.128450>
- Merz WM, Fischer-Betz R, Hellwig K, Lamprecht G, Gembruch U (2022) Pregnancy and autoimmune disease. *Deutsch Arztebl Int* 119(9):145–156. <https://doi.org/10.3238/arztebl.m2021.0353>
- Milosevic A, Janjic MM, Lavrnja I, Savic D, Bozic ID, Tesovic K, Jakovljevic M, Pekovic S, Stojilkovic SS, Bjelobaba I (2020) The sex-specific patterns of changes in

- hypothalamic-pituitary-gonadal axis during experimental autoimmune encephalomyelitis. *Brain Behav Immun* 89:233–244. <https://doi.org/10.1016/j.bbi.2020.06.025>
- Montarolo F, Perga S, Martire S, Bertolotto A (2015) Nurr1 reduction influences the onset of chronic EAE in mice. *Inflamm Res* 64(11):841–844. <https://doi.org/10.1007/s00011-015-0871-4>
- Montarolo F, Raffaele C, Perga S, Martire S, Finardi A, Furlan R, Hintermann S, Bertolotto A (2014) Effects of isoxazolo-pyridinone 7e, a potent activator of the Nurr1 signaling pathway, on experimental autoimmune encephalomyelitis in mice. *PLoS One* 9(9):e108791. <https://doi.org/10.1371/journal.pone.0108791>
- Morales LB, Loo KK, Liu HB, Peterson C, Tiwari-Woodruff S, Voskuhl RR (2006) Treatment with an estrogen receptor alpha ligand is neuroprotective in experimental autoimmune encephalomyelitis. *J Neurosci* 26(25):6823–6833. <https://doi.org/10.1523/JNEUROSCI.0453-06.2006>
- Moulton VR (2018) Sex hormones in acquired immunity and autoimmune disease. *Front Immunol* 9:2279. <https://doi.org/10.3389/fimmu.2018.02279>
- Neier K, Marchlewicz EM, Bedrosian LD, Dolinoy DC, Harris C (2019) Characterization of the mouse white adipose tissue redox environment and associations with perinatal environmental exposures to bisphenol A and high-fat diets. *J Nutr Biochem* 66:86–97. <https://doi.org/10.1016/j.jnutbio.2019.01.005>
- Olsson T, Barcellos LF, Alfredsson L (2017) Interactions between genetic, lifestyle and environmental risk factors for multiple sclerosis. *Nat Rev Neurol* 13(1):25–36. <https://doi.org/10.1038/nrneuro.2016.187>
- Ortona E, Pierdominici M, Maselli A, Veroni C, Aloisi F, Shoenfeld Y (2016) Sex-based differences in autoimmune diseases. *Ann Ist Super Sanita* 52(2):205–212. https://doi.org/10.4415/ANN_16_02_12
- Palanza P, Howdeshell KL, Parmigiani S, vom Saal FS (2002) Exposure to a low dose of bisphenol A during fetal life or in adulthood alters maternal behavior in mice. *Environ Health Perspect* 110(SUPPL. 3):415–422. <https://doi.org/10.1289/ehp.02110s3415>
- Pang Q, Li Y, Meng L, Li G, Luo Z, Fan R (2019) Neurotoxicity of BPA, BPS, and BPB for the hippocampal cell line (HT-22): an implication for the replacement of BPA in plastics. *Chemosphere* 226:545–552. <https://doi.org/10.1016/j.chemosphere.2019.03.177>
- Panzica G, Melcangi RC (2016) Structural and molecular brain sexual differences: a tool to understand sex differences in health and disease. *Neurosci Biobehav Rev* 67:2–8. <https://doi.org/10.1016/j.neubiorev.2016.04.017>
- Pierdominici M, Maselli A, Colasanti T, Giammarioli AM, Delunardo F, Vacirca D, Sanchez M, Giovannetti A, Malorni W, Ortona E (2010) Estrogen receptor profiles in human peripheral blood lymphocytes. *Immunol Lett* 132(1–2):79–85. <https://doi.org/10.1016/j.imlet.2010.06.003>
- Procaccini C, De Rosa V, Pucino V, Formisano L, Matarese G (2015) Animal models of Multiple Sclerosis. *Eur J Pharmacol* 759:182–191. <https://doi.org/10.1016/j.ejphar.2015.03.042>
- Profita M, Fabbri E, Spisni E, Valbonesi P (2021) Comparing effects and action mechanisms of BPA and BPS on HTR-8/SVneo placental cells. *Biol Reprod* 105(5):1355–1364. <https://doi.org/10.1093/biolre/ibob139>
- Rahn EJ, Iannitti T, Donahue RR, Taylor BK (2014) Sex differences in a mouse model of multiple sclerosis: neuropathic pain behavior in females but not males and protection from neurological deficits during proestrus. *Biol Sex Differ* 5(1):4. <https://doi.org/10.1186/2042-6410-5-4>
- Ramien C, Taenzer A, Lupu A, Heckmann N, Engler JB, Patas K, Friese MA, Gold SM (2016) Sex effects on inflammatory and neurodegenerative processes in multiple sclerosis. *Neurosci Biobehav Rev* 67:137–146. <https://doi.org/10.1016/j.neubiorev.2015.12.015>
- Rochester JR (2013) Bisphenol A and human health: a review of the literature. *Reprod Toxicol* 42:132–155. <https://doi.org/10.1016/j.reprotox.2013.08.008>
- Rochester JR, Bolden AL (2015) Bisphenol S and F: a systematic review and comparison of the hormonal activity of bisphenol A substitutes. *Environ Health Perspect* 123(7):643–650. <https://doi.org/10.1289/ehp.1408989>
- Rogers JA, Mishra MK, Hahn J, Greene CJ, Yates RM, Metz LM, Yong VW (2017) Gestational bisphenol-A exposure lowers the threshold for autoimmunity in a model of multiple sclerosis. *Proc Natl Acad Sci USA* 114(19):4999–5004. <https://doi.org/10.1073/pnas.1620774114>
- Rubtsova K, Marrack P, Rubtsov AV (2015) Sexual dimorphism in autoimmunity. *J Clin Invest* 125(6):2187–2193. <https://doi.org/10.1172/JCI78082>
- Ryan L, Mills KHG (2021) Sex differences regulate immune responses in experimental autoimmune encephalomyelitis and multiple sclerosis. *Eur J Immunol*. <https://doi.org/10.1002/eji.202149589>
- Sena A, Couderc R, Vasconcelos JC, Ferret-Sena V, Pedrosa R (2012) Oral contraceptive use and clinical outcomes in patients with multiple sclerosis. *J Neurol Sci* 317(1–2):47–51. <https://doi.org/10.1016/j.jns.2012.02.033>
- Shepherd R, Cheung AS, Pang K, Saffery R, Novakovic B (2020) Sexual dimorphism in innate immunity: the role of sex hormones and epigenetics. *Front Immunol* 11:604000. <https://doi.org/10.3389/fimmu.2020.604000>
- Spence RD, Hamby ME, Umeda E, Itoh N, Du S, Wisdom AJ, Cao Y, Bondar G, Lam J, Ao Y, Sandoval F, Suriyan S, Sofroniew MV, Voskuhl RR (2011) Neuroprotection mediated through estrogen receptor-alpha in astrocytes. *Proc Natl Acad Sci USA* 108(21):8867–8872. <https://doi.org/10.1073/pnas.1103833108>
- Spence RD, Voskuhl RR (2012) Neuroprotective effects of estrogens and androgens in CNS inflammation and neurodegeneration. *Front Neuroendocrinol* 33(1):105–115. <https://doi.org/10.1016/j.yfrne.2011.12.001>
- Thompson AJ, Baranzini SE, Geurts J, Hemmer B, Ciccarelli O (2018) Multiple sclerosis. *Lancet* 391(10130):1622–1636. [https://doi.org/10.1016/S0140-6736\(18\)30481-1](https://doi.org/10.1016/S0140-6736(18)30481-1)
- Tiwari-Woodruff S, Morales LB, Lee R, Voskuhl RR (2007) Differential neuroprotective and antiinflammatory effects of estrogen receptor (ER)alpha and ERbeta ligand treatment. *Proc Natl Acad Sci USA* 104(37):14813–14818. <https://doi.org/10.1073/pnas.0703783104>
- van den Berg R, Laman JD, van Meurs M, Hintzen RQ, Hoogenraad CC (2016) Rotarod motor performance and advanced spinal cord lesion image analysis refine assessment of neurodegeneration in experimental autoimmune encephalomyelitis. *J Neurosci Methods* 262:66–76. <https://doi.org/10.1016/j.jneumeth.2016.01.013>
- Venisse N, Cambien G, Robin J, Rouillon S, Nadeau C, Charles T, Rabouan S, Migeot V, Dupuis A (2019) Development and validation of an LC-MS/MS method for the simultaneous determination of bisphenol A and its chlorinated derivatives in adipose tissue. *Talanta* 204:145–152. <https://doi.org/10.1016/j.talanta.2019.05.103>
- Voskuhl RR, Wang H, Wu TC, Sicotte NL, Nakamura K, Kurth F, Itoh N, Bardens J, Bernard JT, Corboy JR, Cross AH, Dhib-Jalbut S, Ford CC, Frohman EM, Giesser B, Jacobs D, Kasper LH, Lynch S, Parry G, ... Elashoff R (2016) Estriol combined with glatiramer acetate for women with relapsing-remitting

- multiple sclerosis: a randomised, placebo-controlled, phase 2 trial. *Lancet Neurol* 15(1):35–46. [https://doi.org/10.1016/S1474-4422\(15\)00322-1](https://doi.org/10.1016/S1474-4422(15)00322-1)
- Voskuhl RR (2020) The effect of sex on multiple sclerosis risk and disease progression. *Mult Scler* (houndmills, Basingstoke, England) 26(5):554–560. <https://doi.org/10.1177/1352458519892491>
- Watson C, Paxinos G, Kayalioglu G, Heise C (2009) Atlas of the mouse spinal cord. *Spinal Cord* 308–379. <https://doi.org/10.1016/B978-0-12-374247-6.50020-1>
- Waubant E, Lucas R, Mowry E, Graves J, Olsson T, Alfredsson L, Langer-Gould A (2019) Environmental and genetic risk factors for MS: an integrated review. *Ann Clin Transl Neurol* 6(9):1905–1922. <https://doi.org/10.1002/acn3.50862>
- Whitacre CC, Reingold SC, O’Looney PA (1999) A gender gap in autoimmunity. *Science* 283(5406):1277–1278. <https://doi.org/10.1126/science.283.5406.1277>
- Yilmaz C, Karali K, Fodelianaki G, Gravanis A, Chavakis T, Charalampopoulos I, Alexaki VI (2019) Neurosteroids as regulators of neuroinflammation. *Front Neuroendocrinol* 55:100788. <https://doi.org/10.1016/j.yfrne.2019.100788>
- Ysraelit MC, Correale J (2019) Impact of sex hormones on immune function and multiple sclerosis development. *Immunology* 156(1):9–22. <https://doi.org/10.1111/imm.13004>

Publisher's Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.