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Progress in biological and medical research in the deep underground: an update

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As the growing population of individuals residing or working in deep underground spaces for prolonged periods, it has become imperative to understand the influence of factors in the deep underground environment (DUGE) on living systems. Heping Xie has conceptualized the concept of deep underground medicine to identify factors in the DUGE that can have either detrimental or beneficial effects on human health. Over the past few years, an increasing number of studies have explored the molecular mechanisms that underlie the biological impacts of factors in the DUGE on model organisms and humans. Here, we present a summary of the present landscape of biological and medical research conducted in deep underground laboratories and propose promising avenues for future investigations in this field. Most research demonstrates that low background radiation can trigger a stress response and affect the growth, organelles, oxidative stress, defense capacity, and metabolism of cells. Studies show that residing and/or working in the DUGE has detrimental effects on human health. Employees working in deep mines suffer from intense discomfort caused by high temperature and humidity, which increase with depth, and experience fatigue and sleep disturbance. The negative impacts of the DUGE on human health may be induced by changes in the metabolism of specific amino acids; however, the cellular pathways remain to be elucidated. Biological and medical research must continue in deep underground laboratories and mines to guarantee the safe probing of uncharted depths as humans utilize the deep underground space.

KEYWORDS

deep underground laboratories, deep underground medicine, background radiation, cell growth, phenotypic changes

1. Introduction

As resources in the earth's superficial layers become scarce, many countries have initiated efforts to explore and exploit the deep-underground space (1, 2). Deep coal mining has occurred in Poland, Germany, the United Kingdom, Japan, and France since the 1980s, and currently, 55 coal mines have reached depths of over 1,000 m in China (3, 4).

Gaining insights into the effects of deep underground environment (DUGE) factors on living systems is essential for understanding the nature of the deep underground space and

implementing measures to protect the health of the humans that use the deep underground space (5). In 2018, Heping Xie advocated the need for deep underground medicine (DUGM), a multidisciplinary approach to exploring the physiological, pathological and psychological impacts of factors in the DUGE on humans (5). DUGM will identify detrimental factors in the DUGE and facilitate the development of efficient and secure methods to harness factors that are beneficial for life.

Research on factors in the DUGE is conducted in deep underground laboratories (DULs). Cosmic ray is reduced in DULs created under a rock overburden exceeding 1,000 m of water equivalent, creating a unique environment that allows investigations into rare occurrences in the fields of astroparticle physics and neutrino physics (6, 7). More recently, scientists have developed an interest in the biological impacts of low background radiation (LBR) on living organisms nurtured in DULs (6, 8). Currently, 12 DULs worldwide are engaged in biological research to elucidate the influence of factors in the DUGE on living systems (Figure 1).

Multiple experiments in DULs have revealed the notable detrimental impact of LBR in single-celled organisms (9). Since Prof. Xie proposed the concept of DUGM, an increasing number of DULs have extended their studies to multicellular organisms and started to explore the intricate molecular mechanisms that underlie the biological impacts of LBR (10, 11). Other studies are focused on the impact of factors in the DUGE on human physiological and psychological health. Here, we present a summary of the current landscape of biological and medical research in DULs and propose promising avenues for future investigations in this field.

2. Characteristics of the DUGE

To effectively comprehend the biological impacts of DUGE on living organisms, it is necessary to describe the various environmental

factors present, including radiation, temperature, humidity, and air pressure. The total background radiation level in DULs is 5–10 times lower than above-ground laboratories (Table 1). Natural background radiation comprises γ radiation, neutrons, muons, and radon gas. In DULs, γ radiation originates from nuclear decays of ^{222}Rn and ^{220}Rn and their daughter products in the rocks and atmosphere (21); neutron flux is usually low, resulting from the decay of uranium and thorium in rocks and building materials (22); muons are produced by the interaction of cosmic radiation with the Earth's atmosphere, with the rocky cover of the DULs providing effective shielding (23).

One further contribution to the natural background radiation comes from radon decay products. Radon is a naturally occurring radioactive gas generated by the decay of uranium in rocks and soil, and its decay releases α particles into the air (24, 25). And the hermetic nature of underground mines allows for the significant accumulation of radon, which not only increases the natural background radiation but also has detrimental effects on the health of workers involved in deep underground operations (26, 27). The World Health Organization has reported that radon gas was the second most significant cause of lung cancer after smoking (28). Besides, Rage and Richardson found that the incidence rate of lung cancer among deep underground uranium miners showed a positive correlation with cumulative radon exposure (29, 30). Therefore, it is necessary to focus on the levels and fluctuations of radon gas in the DUGE.

In addition to radiation, the temperature, humidity, and air pressure in the deep underground differ from above ground. Watson et al. (31) measured temperature change with increasing depth in mines and showed that temperatures increase by 3–5°C for every 100 m of depth in the deep underground. Relative humidity and air pressure also rise with increasing depth (32). Our previous studies found that the humidity exceeds 90% at a depth of 600 m in the Erdaogou mine of Jiapigou Minerals Limited Corporation of China National Gold Group Corporation (CJEM) and the China Pingmei Shenma Group (CPSG) mine, which exceeds the acceptable maximum

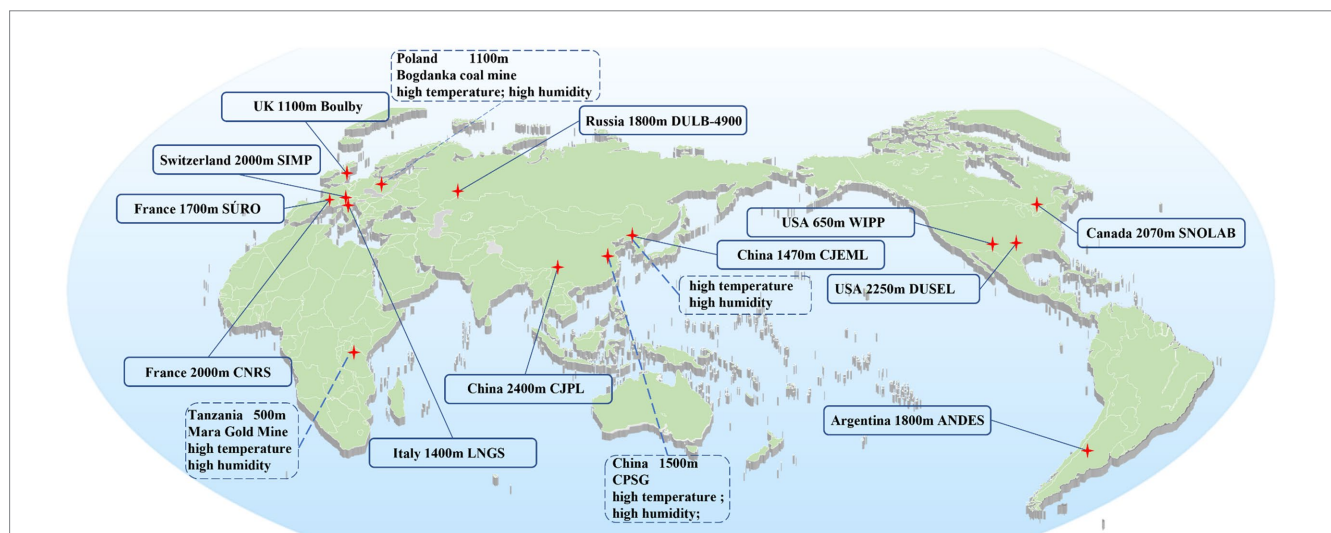


FIGURE 1

The location of DUGLs conducting biological research. SIMP, Simplon pass; WIPP, waste isolation pilot plant; CJEM, Laboratory of Jiapigou Minerals Limited Corporation of China National Gold Group Corporation; CJPL, Chinese Jinping Deep Underground Laboratory; DUSEL, Dusseldorf Underground Laboratory, United States; LNGS, the Gran Sasso National Laboratory; DULB-4900, the Baksan Neutrino Observatory; CPSG, China Pingmei Shenma Group; CNRS, Center National de la Recherche Scientifique; ANDES, Argentina Agua Negra Extremely Deep Underground Laboratory; and SNOLAB, Sudbury Neutrino Observatory Laboratory.

TABLE 1 Components of background radiation in deep underground laboratories.

Laboratory name	Depth	γ flux	Muon flux ($\text{cm}^{-2} \text{s}^{-1}$)	Neutron flux ($\text{cm}^{-2} \text{s}^{-1}$)	Radon (Bq m^{-3})	Reference
WIPP	650 m	$8.27 * 10^{-9} \text{Gy/h}$	$4.77 * 10^{-7}$	$6 * 10^{-8}$	7	(12)
Boulby	1,100 m	$0.128 (\text{cm}^{-2} \text{s}^{-1})$	$4.5 * 10^{-8}$	$1.7 * 10^{-6}$ (E. 0.5 MeV)	2.4	(13, 14)
SNOLAB	2,000 m	$8.2 * 10^{-4} (\text{m}^{-2} \text{min}^{-1})$	$3 * 10^{-10}$	$9.3 * 10^{-6}$	120	(15)
LNGS	1,400 m	20 nSv / h	$3 * 10^{-8}$	$3.78 * 10^{-6}$	50–120	(16)
CNRS	1,700 m	0.301 or 0.622 ($\text{cm}^{-2} \text{s}^{-1}$)	$4.7 * 10^{-9}$	$5.6 * 10^{-6}$	15	(17)
DULB-4900	1,800 m	0.02 Gy/h	$3.0 * 10^{-9}$	$3.8 * 10^{-7}$	0.85	(18)
CJEM	1,470 m	0.04 $\mu\text{Sv/h}$	NA	NA	148	(19)
CJPL	2,400	NA	$2 * 10^{-10}$	$2.69 * 10^{-5}$	34–133	(20)

relative humidity for humans (70%) (33, 34). Carbon dioxide and oxygen concentrations in the CJEM and CPSG are similar to above ground, due to appropriate ventilation (33, 34).

3. Biological research in the DUGE

Natural background radiation has influenced the evolution of life on Earth for almost 4 billion years (7, 35). The “linear no-threshold” (LNT) model presumes that every exposure to radiation comes with an increased biological risk, and posits that there is no safe level below which harmful effects are not experienced (36, 37). Nevertheless, an expanding body of studies have contradicted the existing model and suggested that the dose–response relationship of cells and tissues under low radiation may not follow a linearity. Examples of phenomena that may deviate from the dose–response relationship include adaptive response and the bystander effects in cell populations (38). The former refers to the phenomenon where cells acquire resistance to the same or even higher doses of radiation after prior exposure to low doses of radiation (39, 40). The bystander effect suggests that cells exposed to low doses of radiation can affect non-irradiated cells through intercellular communication, which results in genomic instability and cellular damage (41). Current understanding of the biological effects of low radiation and LBR is limited. DUL provides a stable long-term chronic LBR environment, which contributes to refining the dose–response relationship of the LNT model and elucidating its underlying mechanisms (13, 42). At present, some DULs are conducting research on the biological effects of LBR in various organisms, including (i) unicellular organisms such as V79 cells, *Shewanella oneidensis*, and *Deinococcus radiodurans* and (ii) multicellular organisms such as *Drosophila melanogaster*, *Caenorhabditis elegans*, and Lake Whitefish. Research is focused on impacts of LBR on the growth, cellular functions, and phenotypes of the model organisms (Table 2).

3.1. LBR affects the growth of organisms

In 1964, Eugster indicated that when the cyanobacterium *Mastigocladus laminosus* was cultivated in the Simplon tunnel, it experienced mortality within a matter of week (8) (Figure 2). Planel et al. (43), Smith et al. (9), Kawanishi et al. (56), and Castillo et al. (44,

45) showed significantly reduced growth rates of single-celled organisms cultured in DULs for 3–10 days. In comparison to parallel populations that were cultured above the ground; however, the growth rates of the single-celled organisms cultured in the DULs could be rescued by introduction of an exogenous radiation source that simulated natural background levels (9, 56). Liu confirmed this phenomenon when studying growth rates of mammalian cells for 4–7 days in the CJEM (46, 47). Castillo et al. conducted a research that revealed no impact on the growth rates of *Shewanella oneidensis* when cultured in the Waste Isolation Pilot Plant (WIPP) for 4 days in New Mexico (45, 51). Satta et al. (48) found that V79 cells displayed normal growth rates when cultured in a DUL for 9 months, and Wadsworth et al. (13) elucidated that growth rates of *Bacillus subtilis* and *Escherichia coli* were not affected after 7 days of incubation in a DUL. Castillo et al. (49) discovered that V79 cells cultured for 23 days under LBR in the WIPP had a higher and more variable number of cells and more heterogeneous cell populations compared to cells grown above ground.

In recent years, research in the deep underground has extended to multicellular organisms. Morciano et al. (52) cultured the multicellular organism *Drosophila melanogaster* for 9 months in a DUL and found an increased lifespan but decreased fertility of adult males and females. *Drosophila melanogaster* cultured for five generations showed a stress response and chromosomal damage induced by exposure to LBR. This phenomenon had a cross-generational effect (53). Van Voorhies et al. (11) found that egg-laying rates and larval growth were increased in *Caenorhabditis elegans* exposed to LBR for 8 months compared to those observed under normal background radiation levels. Pirkkanen et al. (10) showed no significant differences in hatching time or survival rates in lake whitefish reared in SNOLAB (2 km underground) for 4 months and on the surface; however, embryos raised underground were larger. The authors speculated that higher radon levels underground ($100\text{--}150 \text{Bq/m}^3$) may potentially account for the notable increase in body size observed in embryos reared within the SNOLAB facility (57).

Although the different organisms maintained in LBR might exhibit different changes in growth, the single-celled organisms presented with growth inhibition in short time might be confirmed. Whereas, a few researches about multicellular cultures and longtime observation for single-celled organisms under LBR environment limited to get a solid conclusion, emphasizing the need for more precise long-term investigations (58).

TABLE 2 The different biological effects of low background radiation on different organisms.

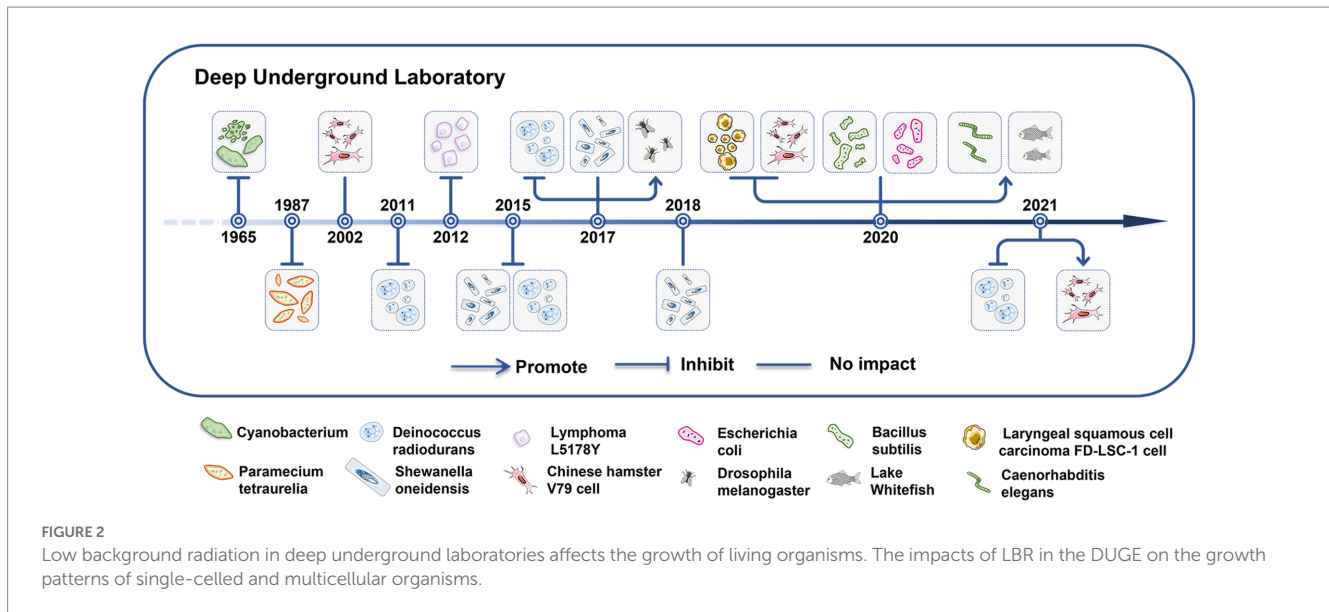
Reference	Laboratory	Depth (m)	Time	Value of radiation dose (LBR vs. CBR; nGy/h)	Cultures	Impact
(8)	SIMP	2,000	A few weeks	Not stated	<i>Mastigogladus I.</i>	Died
(43)	CNRS	200 + 5 cm lead	10 days	11.4 vs. 188.4	<i>Paramecium tetraurelia</i>	Growth↓
(9)	WIPP	650	75 h	2 vs. 31	<i>Deinococcus radiodurans</i>	Growth↓, HSP70↑
(44)	WIPP	650	72 h	2 vs. 31	<i>Shewanella oneidensis</i> , <i>Deinococcus radiodurans</i>	Growth↓, KatB↑, SOA0154↑, RecA↑, Dps ↓, GAPDH↓, and DnaK↑
(45)	WIPP	650	77 h	2 vs. 31	<i>Deinococcus radiodurans</i>	Growth↓, DnaK↑
(12)	WIPP	605	34 h	0.91 vs. 72.1	<i>Deinococcus radiodurans</i>	Growth↓, protein synthesis↓
(13)	Boulby	1,100	7 days	0.5 vs. 112.5	<i>Bacillus subtilis</i> , <i>Escherichia coli</i>	Growth (–)
(46)	CJEM	1,470	7 days	40 vs. 150	V79	Growth↓, mitochondrial volume↑, hypertrophic ER, ribosomal and spliceosomal protein pathway enrichment, oxidoreductase↓, UQCRH↑, ATP6V1G1↑, and DNA metabolic changes
(47)	CJEM	1,470	4 days	40 vs. 150	FD-LSC-1	Growth↓, ribosomal and spliceosomal protein pathway enrichment, mitochondrial volume↑, hypertrophic ER, UQCRH↑, ATP6V1G1↑, and DNA metabolic changes
(48)	LNGS	1,400	9 months	22 vs. 122	V79	Growth (–), GST (–), SOD↓, and HPRT mutation↑
(49)	WIPP	650	23 days	0.91 (LB) vs. 35 (CB) vs. 72(UB)	V79	Viability↓, transcription-related genes↓
(50)	LNGS	1,400	6 months	3.8 vs. 331.7	TK6	Growth (–), SOD↓, GSH-PX↓, and CAT↓,
(51)	WIPP	640	4 days	0.16 vs. 71	<i>Shewanella oneidensis</i>	Growth (–), protein synthesis↓
(52)	LNGS	1,400	9 months	22 vs. 122	<i>Drosophila melanogaster</i>	Lifespan↑; fertility↓
(11)	WIPP	650	8 months	15.6 vs. 67.4	<i>Caenorhabditis elegans</i> Nematode	Growth↑, egg-laying↑
(10)	SNOLAB	2,000	4 months	11.55 vs. 68.04	Lake Whitefish	Growth↑
(19)	CJEM	1,470	2 days	40 vs. 150	V79	Growth↓, RNA metabolic changes
(53)	LNGS	1,400	Five generations	22 vs. 122	<i>Drosophila melanogaster</i>	Chromosome breaks↑
(54)	LNGS	1,400	8 months	22 vs. 122	V79	SOD, CAT, GSH-PX, GSSG-RX, and GST change
(55)	LNGS	1,400	10 months	22.8 vs. 94.7	V79	GSH-Px↓, HPRT mutation↑

SIMP, Simplon pass; CNRS, Center National de la Recherche Scientifique; LNGS, the Gran Sasso National Laboratory; WIPP, Waste Isolation Pilot Plant; SNOLAB, Sudbury Neutrino Observatory Laboratory; CJEM, Laboratory of Jiapigou Minerals Limited Corporation of China National Gold Group Corporation; LBR, low background radiation; CBR, control background radiation; UB, underground background contains K-40; ER, endoplasmic reticulum; UQCRH, ubiquinol cytochrome c reductase hinge; ATP6V1G1, ATPase H⁺ Transporting V1 Subunit G1; SOD, superoxide dismutase; CAT, catalase; GST, glutathione transferase; GSSG-PX, glutathione peroxidase; GSSG-RX, glutathione reductase; and HPRT, hypoxanthine guanine phosphoribosyl transferase.

3.2. LBR affects the function of organelles

Mitochondria are organelles surrounded by a double-membrane system that provide energy for the activities of eukaryotic cells (59).

Mitochondrial reactive oxygen species (ROS) generated during both physiological and pathological conditions have the potential to inflict cellular damage (60, 61). Liu et al. found that mitochondrial volume increased and mitochondria were largely devoid of cristae in V79 cells



and FD-LSC-1 cells cultured in a DUL compared to above ground (46, 47) (Figure 3).

As a direct intracellular target of radiation, the endoplasmic reticulum (ER) is highly sensitive to changes in the internal environment (62). Mild ER stress initiates autophagy to restore ER homeostasis and protect cells from damage (63). Liu et al. showed that V79 and FD-LSC-1 cells, when cultivated in a DUL, had hypertrophic ER, obvious Golgi bodies, and downregulation of major ER stress proteins (WFS1, STT3B, CANX, ERP29, and HSPA5) compared to cells cultured above ground (46, 47). Transcriptomics demonstrated that differently expressed (DE) circRNAs in V79 cells cultured in the DUL or above ground were predominantly enriched in genes associated with protein processing in the endoplasmic reticulum (19).

Ribosomes are essential for protein translation and cell proliferation (64). Castillo et al. (51) found a downregulation of ribosomal proteins and tRNA genes in *S. oneidensis* deprived of background radiation compared to controls, suggesting a substantial decrease in protein translation. A similar observation was conducted by Liu et al. who indicated alterations in protein expression associated with the ribosome pathway in V79 and FD-LSC-1 cells cultivated in a DUL compared to above ground. KEGG enrichment analysis of DE proteins showed that ribosome biogenesis and ribosomal proteins (RPS6, RPS14, RPS16, RPL8, RPL23, RPL3, and RPS18) played a key role in regulating cell growth in cells cultured in the DUL (46, 47).

Spliceosomes are essential organelles that play a pivotal role in cell growth by regulating gene expression (65). Liu et al. found that DE proteins between V79 and FD-LSC-1 cells and DE lncRNAs between V79 cells cultured in a DUL or above ground were significantly enriched in the spliceosome pathway (19, 46, 47).

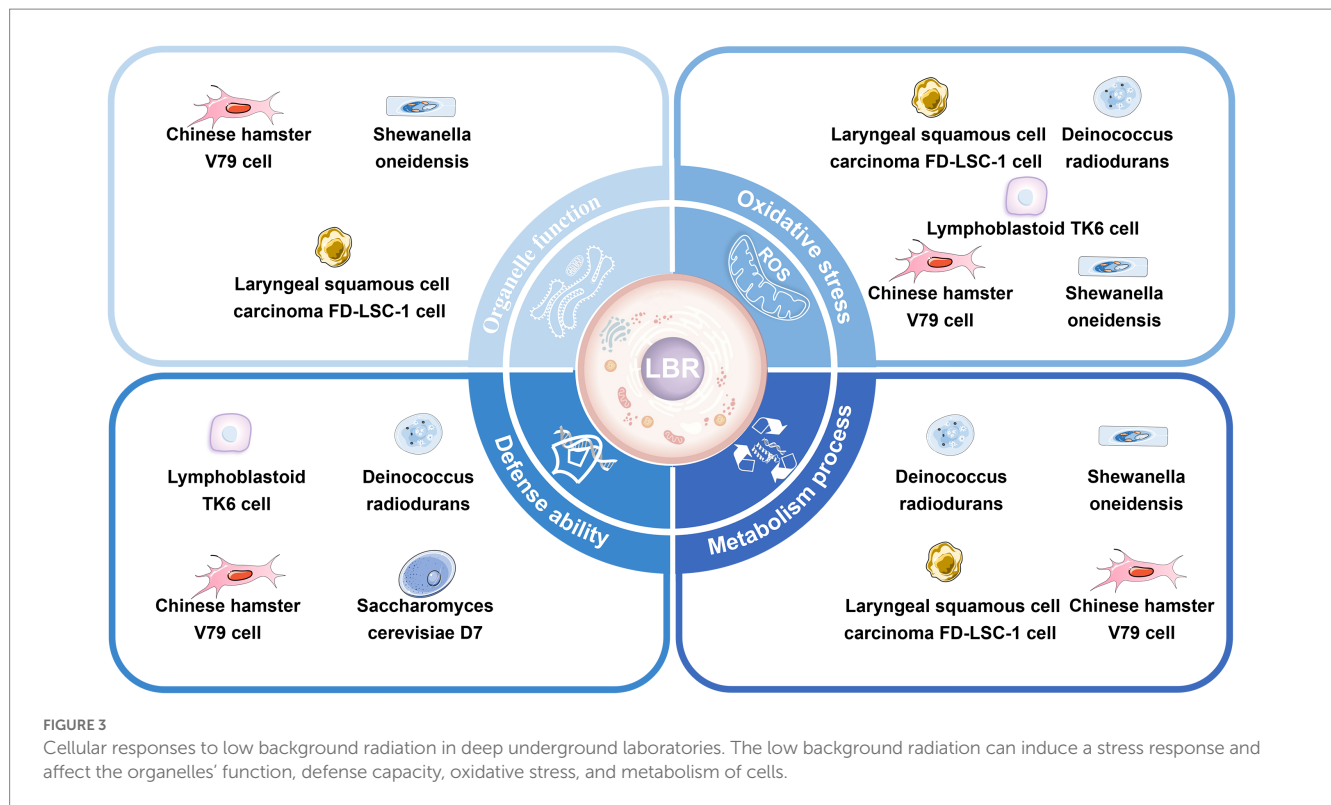
Together, these data imply that mitochondria, the ER, ribosomes, and the spliceosome are involved in the stress and adaptive responses in cells exposed to LBR. The mechanism underlying the role of these organelles in cellular adaptation in a DUGE require further research.

3.3. LBR causes phenotypic alterations in organisms

3.3.1. Oxidative stress

Most biological research on LBR conducted in DULs has focused on cellular oxidative stress. Antonelli et al. discovered changes in antioxidant enzyme activities [superoxide dismutase (SOD), catalase (CAT), glutathione peroxidase, reductase, and transferase (GSH-PX, GSSG-RX, and GST)] in V79 cells grown in the LNGS (54). Satta et al. (48) showed SOD activity was lower in V79 cells grown in LBR, as compared to those cultured in a standard environment for 9 months, while there was no difference in GST activity. Carbone et al. (50) found that SOD, GSH-PX, and CATase were reduced in human lymphoblastoid TK6 cells cultured in reduced compared to reference environmental radiation conditions. Smith et al. (9) reported an increase in the expression of the heat shock protein 70 (HSP70) in primary human lung fibroblasts and bronchial epithelial cells cultured in reduced radiation levels compared to background. Fratini et al. found a decrease in cellular GSH-Px activity in V79 cells cultured for 10 months in LBR, in comparison to cells cultured in a reference environment. This biological response did not revert in cells brought back to the reference-radiation environment for 6 months (55). Castillo et al. demonstrated that the oxidative stress-related genes *katB* and *SOA0154* and the DNA damage-related gene *recA* were upregulated in *S. oneidensis*, while *dps* (DNA protection against ROS) and *gapH* (ATP synthesis) genes were downregulated in *Deinococcus radiodurans* incubated in the WIPP for 76 h compared to background radiation control (44).

Liu et al. performed transcriptomic and proteomic analyses of V79 and FD-LSC-1 cells cultured in a DUL compared to above ground. Withdrawal of background radiation downregulated proteins enriched in oxidoreductase activity and the redox process (46) and upregulated ubiquinol cytochrome c reductase hinge (UQCRH) and ATPase H⁺ Transporting V1 Subunit G1 (ATP6V1G1) (46, 47). Transcriptomic analysis showed that the DE mRNAs between V79 cells grown in the DUL or above ground were mainly enriched in



redox activity and redox processes, and the DE mRNAs of FD-LSC-1 cells grown in a DUL or above ground were also enriched in SOD-related entities (19).

Together, these data imply that LBR in the DUGE could induce oxidative stress in cells. However, the core mechanism and target factors remain to be elucidated.

3.3.2. Defense against external factors

Satta et al. (66) found a higher frequency of radiomimetic induced recombination in *S cerevisiae* D7 exposed to recombinogenic doses of methyl methanesulfonate grown in LBR compared to a standard environment. A subsequent study showed that V79 cells grown in LBR for 9 months exhibited enhanced susceptibility to cycloheximide-induced apoptosis and had an increased frequency of mutations in the hypoxanthine guanine phosphoribosyl transferase (*hprt*) gene induced by γ -ray exposure compared to cells cultivated in normal background radiation (48). Similarly, Fratini et al. and Antonelli et al. observed an increased rate of spontaneous *hprt* mutations in V79 cells cultured in a LBR environment as opposed to a standard radiation (54, 55). Carbone et al. (50, 67) demonstrated that the degree of DNA damage and repair and management of ROS balance was different in TK6 cells exposed to 2Gy X-ray irradiation in a DUL as opposed to a standard environment for 6 months. Castillo et al. (44, 45) showed upregulation of *dnaK* in *D. radiodurans* cultured in LBR compared to background radiation.

Together, these findings imply that LBR may increase the frequency of gene recombination and susceptibility of cells to damaging factors.

3.3.3. Metabolic responses

Metabolic processes involving DNA, RNA, and proteins control cell growth and proliferation. Metabolic processes in organisms

exposed to LBR are altered. Castillo et al. found that protein synthesis was inhibited, reflected by a reduced number of ribosomes and downregulation of tRNA, and transcription of HSP genes was reduced, in *D. radiodurans* exposed to LBR environment as opposed to a reference environment (12, 51). In another study, Castillo et al. (49) used GO analysis to show that transcription-related genes were downregulated in V79 cells cultured in a DUL compared to above ground. Duan et al. (19) and Liu et al. (68) revealed an altered RNA profile compared to control, which may have negative impacts on metabolic processes, in V79 and FD-LSC-1 cells grown in LBR in the DUGE. Liu et al. performed proteomic analysis of V79 and FD-LSC-1 cells cultivated in a DUL compared to above ground and demonstrated modifications in DNA replication, transcription, translation, and protein modification, which may be related to a LBR stress response (46, 47). Liu et al. (68) also showed that the TGF- β and Hippo signaling pathway and the cell proliferation-related genes SMAD, SMAD7, CDH1, EGR1, and BMP2 played key roles in the transcriptional regulation of FD-LSC-1 cells grown in an LBR environment.

Together, these data imply that living systems can perceive LBR in the DUGE and generate stress responses that result in changes to metabolic processes involving protein synthesis.

4. Advancing deep underground medical research

With the prospect of a growing number of individuals inhabiting and working in the deep underground space, some research has been dedicated to exploring the effect of the DUGE on human pathology, psychology, and physiology.

In relation to human pathology, Strzemecka et al. surveyed 700 people working underground in the Bogdanka coal mine at a depth of 1,100 m. They revealed that 50% of the miners perceived the microclimate in the mine, including high humidity and high temperatures, threatened their health (32). Meshi et al. reported that 78.4% of underground miners of the Mara gold mine in Tanzania suffered from moderate heat-related illness, manifested by high body temperature and hot and dry skin. The Mara gold mine is located in the tropics at a depth of 500 m and has an ambient temperature of 28.4°C and relative humidity over 58.2% (69). Liu et al. described the subjective perceptions and mental state of employees working in the CJEM (1,470 m) and CPSG (1,500 m). Moisture and heat were the most commonly perceived adverse factors in the deep underground spaces (33, 34).

In relation to human psychology, it is known that employees working for long periods in shallow coal mines suffer depression and anxiety (70–72). Liu et al. investigated the mental state of 496 miners who worked for long periods of time at a depth below 600 m in the CJEM and CPSG. Workers at both mines reported being easily fatigued and having trouble sleeping and waking up early in the morning (33, 34). In relation to human physiology, urine metabolomics analysis indicates that the DUGE induces specific changes in the metabolism of amino acids (L-phenylalanine, L-tyrosine, and L-glutamine) in humans (73).

5. Conclusion and perspective research in the deep underground

The utilization of deep underground resources and the development of deep underground spaces have become a major strategy for many developed countries. However, research suggests that the unique DUGE could induce a stress response in living organisms and be harmful to humans. Currently, knowledge about the DUGE is limited, making it difficult to support further development. In future, it is necessary to explore the biological impacts of LBR in the DUGE on living systems. DULs provide a unique platform where the biological effects of various components of radiation (radon gas, γ , etc.) and dose responses can be explored. Some data imply that mitochondria, the ER, ribosomes, and the spliceosome are responsible for a stress response in cells cultured in a LBR environment. However, the interconnections and feedback control between organelles and underlying molecular mechanisms remain unclear. Interestingly, LBR in the DUGE has an inhibitory effect on the proliferation of cancer cells, especially radiation-sensitive cancer cells, providing a potential new perspective for cancer control.

Some studies have shown that the DUGE affects human pathology, psychology, and physiology. Therefore, a comprehensive qualitative

and quantitative assessment is imperative to accurately characterize the factors in the DUGE (rock, humidity, temperature, illumination etc.) and their potential advantageous or detrimental impacts on humans. Findings may inform strategies that facilitate human adaptation and habituation to the DUGE. The long-term consequences of exposure to factors in the DUGE in humans must be monitored, and cross-disciplinary research between DUGM and various clinical disciplines is required to support sustained development and utilization of the deep underground space.

Author contributions

JWe, JC, and CL collated and analyzed the literature. ZH, JZ, and MG are helpful for completing diagrams and writing papers. WL, JWu, and HX wrote sections of the manuscript. All authors contributed to the article and approved the submitted version.

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Conflict of interest

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