

## Deformation controlled load application in heart valve tissueengineering

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# **Deformation controlled load** application in heart valve tissue-engineering

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### Introduction

Mechanical behavior of tissue-engineered heart valves still needs improvement when native aortic valves are considered as a benchmark [1]. Although it is known that cyclic straining enhances tissue formation, optimal loading protocols have not been defined yet.



Figure 1: (a) Picture of 4 bioreactor systems. (b) Schematic drawing of a single bioreactor system, consisting of a pneumatic-air valve (A), a pulsatile pump (B), a bioreactor (C), including two pressure sensors (P) and a flow sensor (F), and a medium container (D).

To study the effect of mechanical conditioning on tissue development, it is desired to monitor and control induced deformations during load application.

#### **Objective**

To develop a bioreactor system for heart valve culture in which leaflet deformation is assessed and controlled in real time and non-invasively.

#### Materials & Methods

A combined experimental-numerical approach [2] was developed to assess volumetric and local leaflet deformation of the cultured heart valve in diastolic configuration (Fig. 1).



flow volumetric deformation

Figure 2: Flow-based deformation measurement principle. (a) A pressure difference over the heart valve causes the leaflets to bulge down; fluid exits the bioreactor (in red). (b) After deformation, the leaflets return to their undeformed state; fluid reenters the bioreactor (in blue). By time-integration of the flow signals, volumetric deformation is distinguished from leakage.

Volumetric deformation of the heart valve was measured using a non-contact measurement method based on flow monitoring (Fig. 2).

A numerical model was employed to relate volumetric deformation to local tissue strains in the valve leaflets. This approach was further developed and a PID controller to regulate deformation was incorporated into the bioreactor system. Ē 0.2



Figure 3: The protocols applied to the tissue-engineered heart valves.

To evaluate the functionality of the bioreactor system, 8 heart valves was tissue-engineered in 2 experiments. In each experiment 4 valves (A, B, C and D) were cultured by applying 2 different conditioning protocols (Fig. 3).

#### Results & Discussion

Good correspondence between the measured and the prescribed deformation values was found in both experiments (Fig. 4a ,b). Mean relative error values of all valves did not exceed 5%. However, controlled load application was not possible for heart valves having leak flows larger than ~10 ml/min (Fig. 4c,d).



Figure 4: Measured volumetric deformation values (a, b) and leak flows (c, d) of tissue-engineered heart valves, given as a function of the culture time for (a, c) experiment I and (b, d) experiment II.

#### Conclusion

This bioreactor system has promising possibilities to systematically elucidate the effects of temporal loading patterns on tissue properties, and to develop an optimal conditioning protocol for tissue-engineering of aortic heart valves.

#### References

- 1.Mol A et al., Circulation 2006; 114(19):152-158.
- 2. Kortsmit et al., Submitted for publication.