



**UNIVERSITY OF GENOA**



**Department of Neuroscience, Rehabilitation, Ophthalmology, Genetics,  
Maternal and Child Health (DINO GMI)**

**PHD COURSE IN NEUROSCIENCE**

**Curriculum in Clinical and Experimental Neuroscience (XXXV course)**

**Coordinator: Prof. Lino Nobili**

**Integrating cerebrospinal fluid  
and [ $^{18}\text{F}$ ]-fluorodeoxyglucose positron emission  
tomography to diagnose Alzheimer's disease and  
research its pathophysiological substrates**

**Supervisor:**

**Prof. Matteo Pardini**

**Candidate:**

**Dr. Federico Massa**



## Summary

1.	Introduction .....	4
1.1	Alzheimer's disease.....	4
1.2	Pathophysiology of AD .....	4
1.3	Clinical course .....	6
1.4	Biomarkers of AD pathology .....	7
1.4.1	Morphologic imaging.....	7
1.4.2	Cerebrospinal fluid biomarkers .....	8
1.4.3	Positron Emission Tomography .....	9
1.4.3.1.1	Amyloid-PET .....	9
1.4.3.1.2	[18F]-FDG-PET .....	10
1.5	A novel biological definition of AD: the AT(N) classification.....	12
1.6	Expanding the AT(N) system: focus on synaptopathy in AD.....	13
1.7	Issues with the use of diagnostic biomarkers in Alzheimer's disease .....	16
2.	Thesis aims and outline .....	19
3.	Reciprocal incremental value of 18F-FDG-PET and CSF biomarkers in MCI patients suspected for Alzheimer's disease and inconclusive first biomarker .....	22
4.	Exploring the brain metabolic correlates of process-specific CSF biomarkers in patients with MCI due to Alzheimer's disease: preliminary data .....	44
5.	Discussion and concluding remarks .....	62
6.	Appendix .....	63
6.1.	Publications during PhD.....	63
6.2.	References (in alphabetical order).....	69
7.	Acknowledgments .....	84

# 1.Introduction

## 1.1 Alzheimer's disease

Alzheimer's disease (AD) is a chronic neurodegenerative disease that is the leading cause of dementia worldwide (Levey 2021). It has a prevalence ranging from 10% to 30% of the population over the age of 65, with a 14-fold increase in over-85-year-old subjects. Sporadic presentation of AD is the most common (>95% of the time) and characterized by a later onset (80-90 years), whereas a little percentage (1%) is caused by gene mutations that affect  $\beta$ -amyloid transformation, resulting in an earlier onset of disease (median age 45 years) (Masters et al. 2015).

Therefore, AD is one of the main concerns for world health today in the context of the current "aging society". The inexorably expanding incidence and prevalence of this disorder, which has a negative impact on patients, their families, and society as a whole, results in an annual expenditure of around \$1 trillion on treatment. As the population over 65 grows, it is anticipated that these costs will continue to rise (Levey 2021).

## 1.2 Pathophysiology of AD

The fundamental anomaly in AD is the extracellular accumulation of  $\beta$ -amyloid protein aggregates, which come from the improper processing of amyloid precursor protein (APP), which is crucial for the development and repair of neurons. Normal breakdown of this protein takes place along what is known as the physiological 'nonamyloidogenic route', starting at the extracellular level with a first cut by the enzyme  $\alpha$ -secretase and ending with a second cut by  $\gamma$ -secretase at the intramembrane level, yielding soluble amyloid fragments. In the case of AD (amyloid cascade), the first step is carried out by a  $\beta$ -secretase in a different location of the APP, whereas a subsequent  $\gamma$ -secretase-mediated cut results in the production of non-solubilized  $\beta$ -amyloid fragments. These peptides are prone to aggregation as oligomers and neurofibrillary clusters before forming neuritic plaques in the brain parenchyma, triggering an inflammatory cascade driven by microglia and astrocytes. Astrocytic

activation caused by this prolonged process causes reactive gliosis, which worsens synaptic and neuronal damage by producing neuroinflammation and oxidative damage mediators. This state affects how proteins and neurotransmitters move from the soma to the synapse, which is crucial for neuronal function. The disturbance of neuronal cell homeostasis causes the Tau protein, an essential component of the axonal cytoskeleton, to be hyperphosphorylated, which leads to the formation of neurofibrillary tangles (NFTs) consisting of hyperphosphorylated Tau protein. These formations are harmful to cells and enhance synaptic dysfunction and neuronal death farther downstream.

It has been established that the most significant risk factors for sporadic forms of AD are familiarity with the condition, age greater than 65, lifestyle factors such as education and employment, and the presence of the APOE allele epsilon-4 ( $\epsilon 4$ ) located on chromosome 19. Noteworthy, the risk for people who carry  $\epsilon 4$  allele in either hetero- or homo-zigosis is increased by three to fifteen times, respectively (Masters et al. 2015). It is believed that a potent, dose-dependent influence on the impaired regulation of cerebral amyloid metabolism with aging mediates the elevated APO  $\epsilon 4$ -related risk for AD. Pathological and amyloid imaging investigations have demonstrated that APO  $\epsilon 4$  carriers had higher cerebral amyloid deposition in comparison to non-carriers in both experimental animals and humans. (Morris et al. 2010).

AD physiopathological changes start years before symptoms manifest, although they tend to plateau about the time when people begin to experience symptoms (Jack et al. 2013). According to neuropathological data, amyloid plaques and NFTs have distinct spatiotemporal trajectories of dissemination in the brain. While the latter spreads from transentorhinal and entorhinal regions to the neocortex at various stages, the former starts at the level of the neocortex and then spreads to the allocortex, basal ganglia, midbrain, and finally the pons and cerebellum (Thal et al. 2014). This process affects how the disease develops clinically, revealing a linear correlation between the severity of the illness and the quantity of NFTs and, to a lesser extent, amyloid neuritic plaques (Masters et al. 2015).

Recent research has combined the idea of synaptopathy with the "amyloid cascade" theory, focusing on synaptic damage that would occur before the death of the neuronal body itself. While the specific processes by which amyloid and tau

aggregates harm the synapse are yet poorly characterised, it has been observed that they behave as "neurotoxic species." These proceed to disrupt glutamatergic transmission pathways primarily at the postsynaptic level, specifically by interacting with and overstimulating the NMDA receptor, resulting in excitotoxic calcium influx, oxidative damage, altered axonal transport, disruption of synaptic networks, and ultimately cell death (Li et al. 2018). These mechanisms, which define AD as a proteinopathy and synaptopathy, lead to progressively severe parenchymal atrophy that is accompanied by cognitive impairment until overt dementia.

### **1.3 Clinical course**

An increasingly incapacitating loss of cognitive abilities, beginning with episodic, semantic, and visuospatial memory and progressing to semantic fluency, praxis, and executive skills, is a characteristic feature of AD. Behavioral and psychological symptoms of dementia (BPSD), which inevitably accompany the clinical progression, frequently also manifest from the outset, and include symptoms such as apathy and depression, anorexia, anxiety, and agitation.

Mild cognitive impairment (MCI) characterizes the early stages of AD and is a condition that resides in a "grey area" between a functional state that is preserved and decline into full-blown dementia. MCI is characterized by a broadly defined clinic, a subtle issue in one or more cognitive domains, objectified by neuropsychological testing, and differs from dementia in that everyday activities are unchanged (Petersen 2004). As reviewed by Roberts and Knopman (2013), approximately 15 to 20 percent of adults over 65 have MCI, and those with MCI and associated memory deficits are more likely to develop AD (or other dementias) than those without MCI. The cohorts analyzed show that transition to dementia occurs in more than 50% of individual with MCI, with an annual conversion rate varying from 4 to 31.1% (Bruscoli and Lovestone 2004).

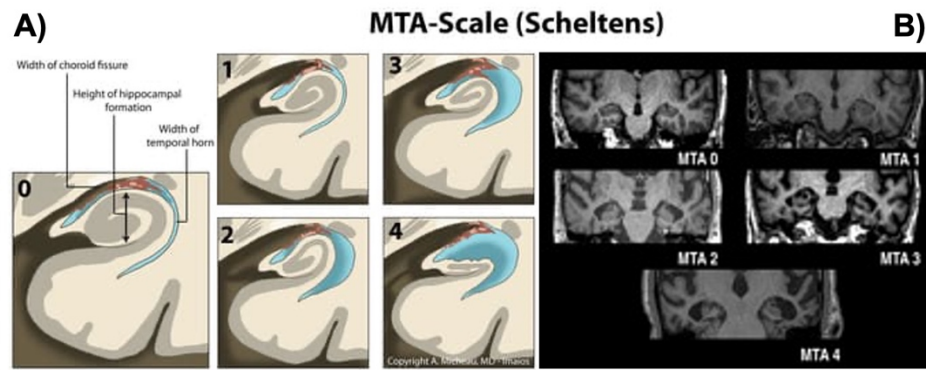
## **1.4 Biomarkers of AD pathology**

To examine the probability that the clinical state of MCI represents an early stage in the spectrum of AD (or other neurodegenerative disorders) that may later develop to dementia with a different likelihood, it is imperative to discover measurements that can be applied in the diagnostic process. This is consistent with how recently developed academic and clinical perspectives on neurodegenerative illnesses. In fact, the traditional diagnostic methods based only on clinical presentation, and eventually confirmed by post-mortem neuropathological findings, has been replaced by a biomarker-based approach involving different biological measures which can reflect pathological changes *in vivo*. By definition, biomarkers are measurable indicators of some biological state or condition allowing in-depth understanding of molecular aberrations and physiopathological processes concurring in neurodegenerative conditions, thus guiding clinicians in the differential diagnostic pathway and researchers in the development of new therapies.

Particularly significant for the AD condition are morphologic, molecular imaging and cerebrospinal fluid (CSF) biomarkers that enable capturing the three primary pathophysiological processes, namely amyloidosis, tauopathy, and neurodegeneration, which are outlined in the AT(N) framework (Jack et al. 2018) that will be thoroughly discussed in the paragraph 1.5.

### **1.4.1 Morphologic imaging**

Magnetic resonance imaging (MRI) can enable for estimation of the regional, structural integrity and volume loss and the detection of other brain disorders, such as space-occupying or vascular lesions, which may present with a cognitive impairment (Frisoni et al. 2010). Several studies have found that structural MRI estimations of tissue damage or loss in typically sensitive brain areas such as the hippocampus and entorhinal cortex are predictive of MCI progression to AD and can be quantified through visual metrics, such as the Medial Temporal Atrophy (MTA) scale (Scheltens et al. 1992) (Figure 1).



**Figure 1.** Graphical representation (A) and coronal T1-MRI sequence (B) for the assessment of mesial temporal lobe atrophy (Scheltens et al. 1992). The score value is determined by the size of the choroid fissure, the enlargement of the temporal horn, and the height of the hippocampal formation, as follows 0: no CSF is visible around the hippocampus; 1: choroid fissure is slightly widened; 2: moderate widening of the choroid fissure, mild enlargement of the temporal horn and mild loss of hippocampal height; 3: marked widening of the choroid fissure, moderate enlargement of the temporal horn, and moderate loss of hippocampal height; 4: marked widening of the choroid fissure, marked enlargement of the temporal horn, and the hippocampus is markedly atrophied and internal structure is lost

#### 1.4.2 Cerebrospinal fluid biomarkers

Alzheimer's disease (AD) may be accurately diagnosed with the use of cerebrospinal fluid (CSF) biomarker analysis, which further enables to thoroughly examine the pathophysiological mechanisms underlying each stage of these diseases. The analysis of CSF following lumbar sampling (L3-L4/L4-L5) is able to quantify the concentration reflecting the rates of both production (expressed, released, or secreted by neurons or glia) and clearance (degraded, removed) of specific proteins at the moment of sample, providing the *biological fingerprints* of AD.

With regard to the diagnosis of AD, the core CSF biomarkers specifically represent amyloid pathogenesis, leading to tauopathy (i.e., formation of the neurofibrillary tangles, NFT) and neurodegeneration according to the "amyloid cascade" theoretical model. These proteins include:

- Amyloid-1-42, alone or normalized based on the concentration of the A $\beta$ 40 isoform (A $\beta$ 42/40 ratio), which reduce in both cases at the CSF level by increased parenchymal sequestration and decreased clearance as an index of amyloid deposition in the brain parenchyma;
- Tau protein phosphorylated at threonine-181 (p-Tau), which represents NFT progressive formation and rises in CSF levels in response;



- total Tau protein (t-Tau), which increases in CSF in parallel to neuronal injury.

In addition to the proteins described above, the CSF level of proteins expressing numerous pathophysiological processes seen in AD, such as neuroinflammation, axonal and barrier/pericyte damage, axonal damage, and synaptic degeneration, can be measured. The latter will be covered in greater depth later.

### **1.4.3 Positron Emission Tomography**

Positron Emission Tomography, or PET, is a technique that uses organic compounds labelled with positron-emitting radioisotopes that enable to examine the regional glucose metabolism ([<sup>18</sup>F]-FDG PET), or the presence of pathological aggregates of proteins of interest. As for the latter, amyloid-PET will be discussed in the following paragraph due to its widespread usage and the substantial evidence supporting its diagnostic performance will be covered in the next paragraphs, whereas Tau-PET, which allows for detection of brain aggregates of Tau protein, still requires full validation and is still mostly available at a few research institutions.

#### **1.4.3.1.1 Amyloid-PET**

The produced fluorinated tracers [<sup>18</sup>F]-florbetaben, [<sup>18</sup>F]-florbetapir, and [<sup>18</sup>F]-flutemetamol) are highly selective for binding amyloid, but due of their lipophilicity, they are also diffusely and powerfully taken up at the level of the white myelin material. They lack horizontal specificity, failing to distinguish between amyloid in its various conformations, such as neuritic plaques (typical of AD and neurodegenerative diseases with cerebral amyloidosis), diffuse extracellular plaques (more typical of senile age), and amyloid angiopathy, but they do not bind other pathological proteins, such as tau and alpha-synuclein. AD, including prodromal forms (MCI-AD), and a significant fraction of individuals with dementia with Lewy bodies (DLB), in whom amyloidosis is linked with alpha-synuclein deposits, are all known to disclose significantly increased retention of amyloid tracers in the brain. Overall, the results for the three fluorinated tracers and for both visual and quantitative analyses ranged from 89% to 97% for sensitivity to distinguish AD patients from healthy people, but the values for specificity varied more significantly, from 63% to 93% (Morris et al. 2016). Indeed, a variable proportion of the aged population has been shown to have cerebral

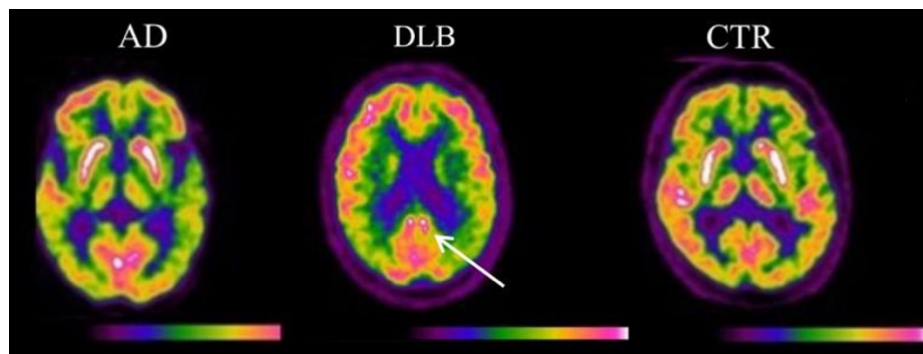
amyloidosis (10-15% at age 60, 30% at age 75, and higher after age 75), with a link to the Apo  $\epsilon 4+$  genotype (where 50% positive is attained at age 75 among cognitively healthy patients). Therefore, it is essential to understand the meaning of a positive amyloid PET scan, indicating the existence of cerebral amyloidosis that allow accomplishing the 'A' criteria of the AT(N) framework for the Alzheimer's continuum condition (Jack et al. 2018), but can be prone to false positives in the elderly.

#### 1.4.3.1.2 [ $^{18}\text{F}$ ]-FDG-PET

The ability to quantify the rate of glucose consumption in each region of the brain gives [ $^{18}\text{F}$ ]-FDG PET its distinctive quality, providing information on the distribution of synaptic dysfunction and *in vivo* neuronal death. Since [ $^{18}\text{F}$ ]-fluorodeoxyglucose (FDG) is a glucose analog, it is readily taken up by the cells with the highest metabolic activity, such as brain cells, where it is phosphorylated and rendered inert. The presence of fluorine-18 ([ $^{18}\text{F}$ ]), a radioisotope that may generate positrons that can be detected by the PET tomograph, results in a steric clutter that prohibits [ $^{18}\text{F}$ ]-FDG from undergoing glycolysis, unlike glucose. As a result, the molecule emits a signal that is "visible" to the tomograph for as long as it is radioactive and confined inside the cell. By analyzing the distribution of the tracer in various brain regions, it is possible to identify patterns that might aid in making the right diagnosis.

As a general interpretation, the region is less intact and synaptically functioning the less metabolically active (and so hypometabolic) it is, and vice versa. Because the metabolic consumption of glucose specifically takes place at the level of the neuron-astrocyte synaptic unit (neuron-glia metabolic coupling) (Magistretti 2000) hypometabolism in a given brain region is likely to be associated with lower synaptic density or neuronal suffering, or it could be an expression of deafferentation from another functionally-connected pathological region (diaschisis phenomenon), even before presenting neuronal damage. This latter point explains why [ $^{18}\text{F}$ ]-FDG-PET has higher sensitivity as a biomarker of neurodegeneration than traditional morphological imaging (MRI), particularly in the early stages of a neurodegenerative process, and a resulting significant negative predictive value (Nobili et al. 2018a). More particular patterns in the setting of dementia, and notably in AD, may be found exactly by studying the distribution of metabolism. In terms of AD, the temporo-parietal regions,

which comprise the angular gyrus, precuneus (PC), and posterior cingulate (PCC), are known to be the most often hypometabolic areas (Bohnen et al. 2012). Although asymmetrical, the basal hypometabolism detected is often bilateral. Conversely, it is typically challenging to detect frank hypometabolism at the mesial temporal level (entorhinal cortex, including the hippocampus, and amygdala), despite the fact that this area is primarily affected by the neurodegenerative process, as there is physiologically lower basal metabolism in these areas and the volume of these structures also varies across subjects. Therefore, it is thought that the most accurate and early indicator of AD is the hypometabolism of the posterior cingulate (PCC), which is frequently extended to the precuneus (PC) (Morbelli et al. 2015a). This pattern involving PC/PCC hypometabolism, which expresses a functional disconnect with the hippocampus, is helpful in the differential diagnosis of pathologies like dementia with Lewy bodies (DLB), which typically presents with occipital hypometabolism but with relative sparing of the posterior cingulate ("cingulate island sign") due to less hippocampal involvement (McKeith et al. 2017) (Figure 2).



**Figure 2.** [ $^{18}\text{F}$ ]-FDG-PET metabolic patterns of AD with the predominant involvement of the temporo-parietal cortex, posterior cingulate and precuneus of the classic form (AD); parietoccipital hypometabolism with relative sparing of the posterior cingulate, (cingulate island sign, CIS, white arrow) in dementia with Lewy bodies (DLB); normal metabolism of the healthy subject (CTR)

Automated semi-quantification methods can be used in conjunction with conventional visual analysis of FDG-PET images to increase diagnostic accuracy and reduce the weight of readers' expertise (Nobili et al. 2018b), especially in the early phases of cognitive loss, when there may only be minor changes in brain metabolism (Morbelli et al. 2015a; Massa et al. 2021). Among the many tools, there are specific software that differ primarily in the reference region used for normalization, analysis

methodologies (voxel-based, VBA, or volume-of-interest, VOI-based), the composition of the control group, and the selection of statistical significance threshold.

For instance, each PET voxel can be compared between an individual (single-subject analysis) or a group of subjects (group-analysis) and one or more control groups using a voxel-based analysis (VBA). This results in a parametric map that shows the voxels where the relative decrease in glucose metabolism is statistically significant (based on a pre-set threshold) (Perani et al. 2014). By defining VOIs (volumes of interest), this evaluation may also be applied to clusters made up of statistically significant groups of adjacent voxels. From these, mean metabolic density counts that are similar between groups can be calculated, adjusted to whole brain (WB) metabolism (VOI-based analysis). The semi-quantification technique permits correlation analyses between metabolism and other quantitative variables of interest, such as demographic data, neuropsychological scores, or biological indices.

### **1.5 A novel biological definition of AD: the AT(N) classification**

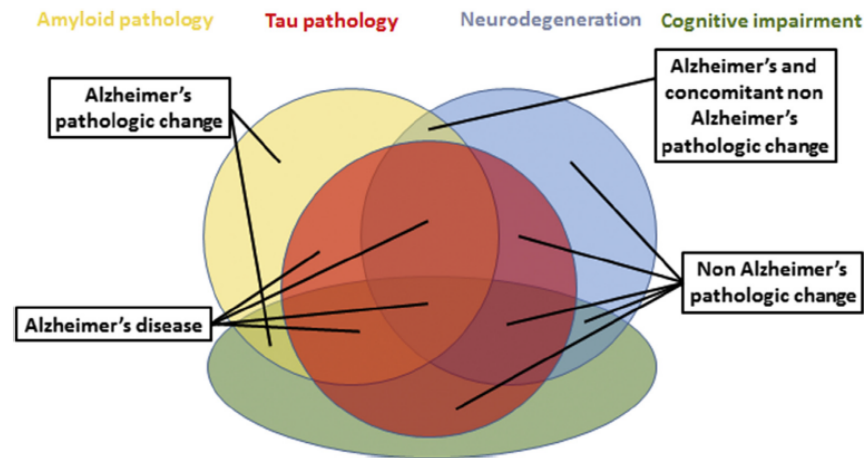
Through the AT(N) framework, the National Institute on Aging and Alzheimer's Association (NIA-AA) recommended updated research standards for AD, incorporating the idea of a "research framework" within the context of the biological development of the illness (AD-continuum) (Jack et al. 2018). This method shifts the emphasis from a solely clinical (signs/symptoms) and cognitive focus to the pathophysiology underpinnings of the illness. As a result, the new framework is based on the investigation of particular biomarkers that attest to the presence of toxic proteins and neurodegeneration for a definition of the disease that becomes biologically based. Therefore, the AT(N) approach enables classifying individuals based on their biomarker profiles outside of the clinic and using a common language with which researchers can comprehend and characterize a precise etiopathological sequence underlying the AD continuum.

Specifically, the use of biomarkers allows definition of:

- amyloidosis (A), namely extracellular amyloid deposits, that is indicated by either reduced A $\beta$ 42 levels, alone or normalized on the value of the A $\beta$ 40 isoform (A $\beta$ 42/40 ratio, or abnormal PET imaging with tracers for amyloid plaques;

- tauopathy (T), namely the presence of aggregates of hyperphosphorylated Tau protein (neurofibrillary tangles, NFT) connected to elevated CSF p-Tau or abnormal PET imaging with tracers for Tau protein;
- neurodegeneration or neuronal injury (N) that can be shown by volume loss at morphological imaging, decreased regional brain metabolism in [ $^{18}\text{F}$ ]-FDG-PET scan, or elevated t-Tau concentration in the CSF.

Precisely, the presence of amyloidosis determines if an individual is on the Alzheimer's continuum, whereas additional tauopathy whether an individual on the Alzheimer's continuum respects the pathophysiological requirements for AD., Neurodegeneration, on the other hand, is a non-specific process that may occur in a variety of diseases, and can be employed as a proxy for severity in AD staging (Figure 3).

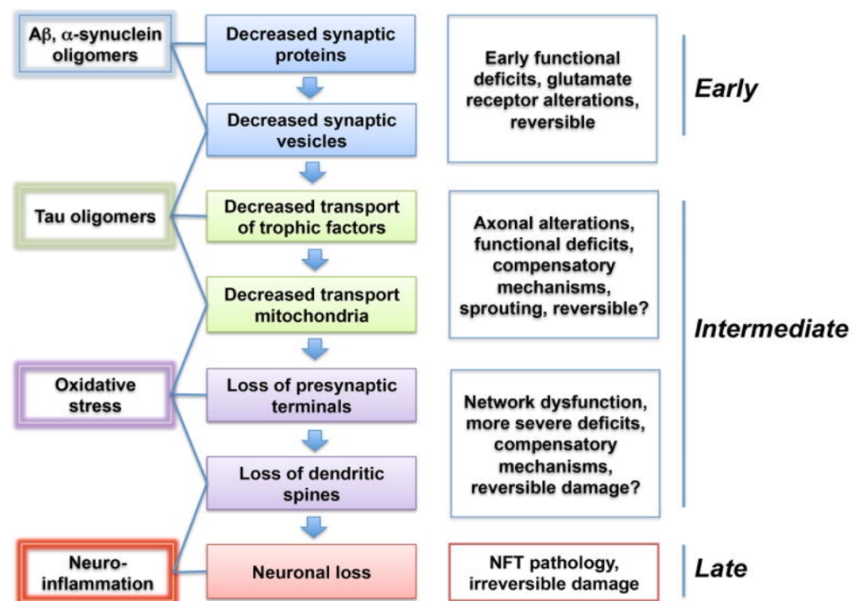


**Figure 3** depicts the interaction of AT(N) system categorization and cognitive state from Jack et al. (2018): Alzheimer's pathologic change (A+, in yellow); substantial biomarker alterations suggestive for Alzheimer's disease (A+T+, A+T+(N±)), in red; no evidence of amyloidosis, hence non-AD condition (A-T±(N±), in blue).

## 1.6 Expanding the AT(N) system: focus on synaptopathy in AD

The AT(N) biomarker matrix theoretically might serve as the foundation for the creation of a larger system, ATX(N), where X denotes new potential biomarkers for other processes that contribute to the pathophysiology of AD, such as changes in the blood-brain barrier, synaptic dysfunction, and neuroimmune dysregulation. For example, vascular, inflammatory or synaptic biomarkers could be integrated to create ATV(N), ATI(N) or ATS(N) systems, respectively (Hampel et al. 2021).

Although the precise mechanisms are still uncertain, evidence suggests that a decrease in synaptic activity and density is one of the first events that would occur before neuronal death in a variety of CNS disorders, including Alzheimer's, which can therefore be classified as "synaptopathies" (Masliah et al. 2001; Taoufik et al. 2018). In AD, synaptic degradation outweighs and precedes neuronal loss at the cortical level and has stronger impact on cognitive symptoms than amyloid plaques or neurofibrillary tangles (Blennow et al. 1996). The amyloid cascade and subsequent buildup of oligomers that obstruct vesicular transport at the synapse level would be the mechanisms that lead to synaptic loss in AD. This multi-step process starts with a reversible initial phase that involves excitatory synapses and is characterized by dysregulation of NMDA and AMPA glutamatergic receptors and those membrane proteins ensuring proper positioning of these channels at the synaptic membrane. It may be assumed that the synapse and its network would already be dysfunctional in the early degenerative phases of disease, but as the synapse gets increasingly damaged, the pre-synaptic terminal and dendritic spines (post-synaptic side) really begin to deteriorate. According to some research, enhanced synaptic sprouting may be an initial protective mechanism, which is then followed by axonal degeneration and ultimately neuronal death in the pathology's later stages (Berchtold et al. 2013; Overk and Masliah 2014) (Figure 4).



**Figure 4.** Diagram of the evolution of the synaptic damage process in AD (Overk and Masliah 2014)

It has been postulated that altering synaptic plasticity is a critical component in the pathophysiology of AD. A significant component would be the neuron's aberrant self-organization, which arises from the dynamic process of compensation and then functional decompensation of the synapse (Arendt 2009). Furthermore, as a site of signal transmission between neurons, synapses may contribute to the spread of disease through a "prion-like" process (Guo and Lee 2014), permitting transmission of tau- and amyloid-oligomer aggregates from one neuron to the next.

In an effort to characterize synaptic dysfunction *in vivo*, several pre- and post-synaptic proteins were described in the last years, but evidence of their involvement in the pathophysiology of AD is sparse (Camporesi et al. 2020). Among them, neurogranin (Ng), a dendritic protein revealing post-synaptic integrity, and two pre-synaptic proteins of the family of synucleins (beta-synuclein,  $\beta$ -syn, and  $\alpha$ -synuclein,  $\alpha$ -syn) increase in the CSF of AD patients along with cognitive impairment and disease progression (Wellington et al. 2016; Oeckl et al. 2016, 2020; Monge-Argilés et al. 2020; Halbgebauer et al. 2021). As the focus of the study project, these proteins will be explained in more depth.

Neurogranin (Ng) is a neuron-specific postsynaptic protein involved in memory consolidation processes and long-term potentiation (LTP) circuits. It is highly expressed by excitatory neurons in the cerebral cortex, hippocampus, and amygdala at the cell body and dendritic spines levels that are involved in calmodulin regulation and commonly affected in AD. Hence, Ng it is seen as a potential marker of synapsis loss in the AD context (Kester et al. 2015; Blennow and Zetterberg 2018). Ng levels in the CSF have been shown to be much higher in AD patients than in those with other types of dementia as early as the MCI stage and to have a strong correlation with cognitive decline and the rate of progression to dementia. Furthermore, in individuals with MCI, the CSF concentration of Ng was found to be directly related to that of t-Tau and p-Tau and indirectly to that of A42, indicating how neurogranin is a sensitive marker for AD alterations (Kvartsberg et al. 2015; Wellington et al. 2016; Tarawneh et al. 2016; Wang 2019).

$\alpha$ -synuclein ( $\alpha$ -syn) is a protein predominantly expressed at the level of the pre-synaptic termination and marginally in the nuclear membrane of neurons in the cerebral cortex, cerebellum, striatum, thalamus, hippocampus, and olfactory bulb.

Alterations  $\alpha$ -syn often take place in the pathogenesis of AD despite typically being connected to synucleinopathies, i.e., Parkinson's disease, dementia with Lewy bodies and multisystem atrophy. It has been shown through research on murine and human cell cultures that  $\alpha$ -syn is involved in the phosphorylation processes of tau protein (mediated by the GSK3 system), which would explain the strong correlation between  $\alpha$ -syn and tau levels in CSF found in several neurodegenerative diseases, including AD. Furthermore, it appears that the presence of amyloid beta enhances GSK3 activity, increasing tau phosphorylation and  $\alpha$ -syn synthesis.  $\alpha$ -syn, tau, and amyloid complexes would encourage mutual aggregation, which might enhance cellular malfunction, and ultimately death (Twohig and Nielsen 2019). Additionally, several studies have shown a connection between cognitive impairment and high CSF levels of  $\alpha$ -syn, which are also connected to amyloid deposition in those patients at risk for sporadic AD and familial AD (Berge et al. 2016; Twohig and Nielsen 2019).

Another presynaptic protein that is a member of the synuclein family and that is mostly expressed in the striatum, thalamus, neocortex, cerebellum, and hippocampus is known as  $\beta$ -synuclein ( $\beta$ -syn). It is mostly concentrated in the pre-synaptic terminal, where it plays a part in a number of membrane activities (Barba et al. 2022), and during synaptic degeneration it is released into the synaptic space, the extracellular area and subsequently the CSF. In fact, CSF levels of  $\beta$ -syn were shown to be greater in the patients with AD at either MCI or dementia stage compared to other neurodegenerative conditions, indicating that this protein represents a promising biomarker of synaptic damage in AD (Oeckl et al. 2016; Halbgebauer et al. 2021). More recently, we showed that CSF  $\beta$ -syn increases in the whole AD continuum since the preclinical stage and may be a helpful early diagnostic/prognostic marker (Barba et al. 2023).

### **1.7 Issues with the use of diagnostic biomarkers in Alzheimer's disease**

The aforementioned biomarkers of AD are spread among centers dedicated to cognitive disorders and dementias, and several data are available about their specific features and accuracy (see for a review Fiandaca et al. 2014). Although, pushed by clinical needs, scientific and technical advances and commercial opportunities, the widespread use of diagnostic biomarkers has actually overtaken both the standardization and harmonization of the various methods and the definition and



validation of targeted diagnostic pathways by directly comparing performances among the various biomarkers (Frisoni et al. 2017).

Several initiatives promoted by the pertinent international scientific societies tried to address issues related to biomarker standardization and clinical application, mainly focusing on one biomarker or class of biomarkers, often without a common synergic framework which could have ensured a global consistency. As for an example, the CSF biomarkers for AD are at an advanced stage of development and harmonization; though, the manual enzyme immunoassays that have been used in most laboratories worldwide are sufficiently reliable and accurate only when used by an experienced staff with quality control procedures. A potentially important advancement is the development of ultra-sensitive, fully automated assays (Bittner et al. 2016; Mattsson-Carlgren et al. 2022) able to reduce the analytic variability. Moreover, efforts are focused on standardization of the operative protocols for CSF handling to reduce pre-analytical variability due to contamination, collection, and storage of samples with the aim to facilitate comparison of CSF biomarkers across studies and laboratories (Hansson et al., 2018). Consequently, a highest performance and reliability of CSF biomarkers analysis, by eliminating most confounding variables, may allow more accurate comparisons among different biomarkers in the diagnostic and prognostic evaluation of patients.

Furthermore, research on the most cost-effective and reliable flowchart for directing biomarker use is currently lacking. The studies comparing biomarker performance do not conclusively show that one is superior than the others (Herukka et al. 2017; Simonsen et al. 2017). As an example, a trend in favor of a higher accuracy of FDG-PET than CSF biomarkers reported by some papers (Shaffer et al. 2013; Prestia et al. 2015; Caminiti et al. 2018) has not been confirmed by a meta-analysis with pathological confirmation, reporting instead a similar accuracy (Cure et al. 2014). Several studies have indicated that combining several indicators enhances diagnostic value to using just one (Bouwman et al. 2007; Vemuri et al. 2009; Choo et al. 2013; Prestia et al. 2013). However, conducting all (or most) biomarkers in an MCI patient is not cost-effective because the majority of patients may be identified appropriately with a single biomarker and only those with ambiguous findings may have access to a

second examination. In this case, a progressive strategy might lead to a cost-effective and precise diagnosis.

There is considerable discussion over which biomarker should emerge first and for which clinical presentation. Both CSF biomarkers and FDG-PET have good diagnostic accuracy and are suggested as first-line biomarkers in MCI patients by scientific organizations (Herukka et al. 2017; Arbizu et al. 2018; Nobili et al. 2018a). When CSF biomarkers are available, they are increasingly recommended if the primary clinical suspicion is AD, but FDG-PET is recommended if alternative diagnoses are at least as likely as AD (Boccardi et al. 2019; Chételat et al. 2019).

Awaiting forthcoming internationally agreed-upon and evidence-based proposals (Festari et al. 2022), even in the clinical routine of centers with high-level of expertise in the management of neurodegenerative diseases, the choice of diagnostic biomarkers often relies on the confidence of the clinician, local facilities, waiting lists, cost, expertise and performance of local laboratories, rather than on the specific characteristics of the biomarker itself and on evidence-based considerations (Frisoni et al. 2017). Thus, to define an efficient combination of biomarkers in AD there is an urgent need to accumulate evidence on their analytical and diagnostic performance, when used alone, simultaneously or in a step-wise fashion. This is intended to assess the discriminant accuracy and the added role of one over another in the diagnostic flowchart, and complements the effort in validating and sharing accurate standardized operative procedures.

## 2. Thesis aims and outline

We have emphasized how various biomarkers are now essential to the diagnostic and research route in the context of research increasingly focusing on the series of pathophysiological events that characterize the course of AD.

In this research project, I have focused on two different types of biomarkers, namely CSF and [ $^{18}\text{F}$ ]-FDG-PET, that are each unique to different pathophysiological processes but that can be integrated to yield complementary data.

Specifically, my project intended to add to the body of knowledge regarding the diagnostic accuracy of biomarkers in AD and to expand the current understanding of some particular pathophysiological aberrations, including synaptic dysfunction.

The first study below (**Chapter 3**) is part of the efforts directed toward defining a diagnostic tree that considers the main characteristics of each biomarker, either alone or in sequential combination, and a cost-effective approach to identifying patients in whom the addition of a second biomarker might be truly useful. This addresses the need to find the best method to achieve the highest diagnostic accuracy using the fewest biomarkers for clinicians as well as for national health care systems, as the number of people with MCI is anticipated to gradually increase, along with the costs provided by diagnostic procedures, care, and hopefully forthcoming disease-modifying drugs. Precisely, we assessed the reciprocal incremental diagnostic value of CSF biomarkers *versus* [ $^{18}\text{F}$ ]-FDG-PET and *vice versa*, utilized one after the other, in individuals with MCI whose first biomarker result had been inconclusive in making the diagnosis. In order to achieve this, we looked on retrospective data collected for clinical purposes from two centers that specialize in the diagnosis and treatment of neurodegenerative disorders. According to clinical practice, either CSF biomarker assessment was used first, followed by [ $^{18}\text{F}$ ]-FDG-PET if this was insufficient for diagnosis, or *vice versa*. We first reported whether the second biomarker utilized in these two cohorts resulted in a diagnosis, and then we evaluated how many of these diagnoses were confirmed at longitudinal follow-up, which was regarded as the gold standard for confirming diagnoses. Finally, we could calculate an incremental relative diagnostic value of 30.6 percent for CSF compared with an inconclusive [ $^{18}\text{F}$ ]-FDG-

PET and vice versa of 38.5 percent. Therefore, we discovered a slight advantage for FDG-PET over CSF in making the most accurate diagnosis; consequently, in practical terms, we might suggest FDG-PET when the diagnostic scenario has not been narrowed by first-line assessments and still includes other diseases besides AD. On the other hand, adding CSF might be useful when we need a specific confirm of a valid suspicion of AD, which FDG-PET initially was unable to resolve.

In the second study of my research project (**Chapter 4**), we have leveraged the intrinsic abilities of [ $^{18}\text{F}$ ]-FDG-PET as a multifunctional measure of neuron-astrocyte metabolic unit activity, neuropil loss, and synaptic density, which is sensitive to the phenomenon of deafferentation (diaschisis) and axonal degeneration with neural network degradation (Herholz 2003; Nakashima et al. 2007; Zimmer et al. 2017; Provost et al. 2021). Through the course of AD, these mechanisms shift in relative importance, which affects the topography and degree of hypometabolism in individuals and stages differently (Brown et al., 2014; Mosconi, 2005; Silverman et al., 2001). According to this concept, specific topographic correlations of metabolic values with CSF proteins that represent distinctive underlying disease processes can be hypothesized. For this reason, we concentrated on a group of 26 patients with MCI-AD to investigate the brain metabolic correlates of certain newly discovered CSF synaptic biomarkers (Ng,  $\alpha$ -syn,  $\beta$ -syn) through the semiquantitative analysis of [ $^{18}\text{F}$ ]-FDG-PET scans. We additionally extended the analysis to the neurofilament light chain (NfL), whose level in CSF rises in AD as early as the MCI stage in correlation with cognition, amyloid load, and cortical atrophy (Zetterberg et al. 2016; Dhiman et al. 2020) as an expression of axonal damage (Gaetani et al. 2019). Our objectives were dual. Investigate the topographic relationship between these indicators and the metabolism of glucose in the brain, on the one hand. To determine if and to what extent these correlated locations relate to still largely unaltered regions or overlap with hypometabolic zones as determined from comparing MCI-AD patients with a group of healthy subjects. Overall, the alterations of different brain areas related to these proteins suggest they express distinct pathophysiological processes occurring in prodromal AD. Briefly, we observed negative correlations between the Ng and  $\alpha$ -syn concentrations in the CSF and the metabolism of the left precuneus/posterior cingulate

cortex (PC/PCC), which largely comprised regions of low metabolism. The metabolic correlates of  $\beta$ -syn and NfL regarded either the left or right lateral temporal areas, respectively, with some overlap with hypometabolism only for the  $\beta$ -syn-related volume. This leads us to hypothesize that the levels of i) CSF Ng and  $\alpha$ -syn express a hippocampal damage that has already developed and is causing PC/PCC deafferentation and hypometabolism; ii) CSF  $\beta$ -syn may represent the development of synaptopathy in the temporal lobe; and iii) CSF NfL reflect the axonal injury in right temporal regions where neuronal loss is not yet manifest.

### **3. Reciprocal incremental value of [<sup>18</sup>F]-FDG-PET and CSF biomarkers in MCI patients suspected for Alzheimer's disease and inconclusive first biomarker**

Massa F<sup>1</sup>, Farotti L<sup>2</sup>, Eusebi P<sup>3,4</sup>, Capello E<sup>5</sup>, Dottorini ME<sup>6</sup>, Tranfaglia C<sup>6</sup>, Bauckneht M<sup>7</sup>, Morbelli S<sup>7,8</sup>, Nobili F<sup>1,5</sup>, Parnetti L<sup>2</sup>

<sup>1</sup> Department of Neuroscience, Rehabilitation, Ophthalmology, Genetics, Maternal and Child Health (DINO GMI), University of Genoa, Genoa, Italy

<sup>2</sup> Center for Memory Disorders and Laboratory of Clinical Neurochemistry, Neurology Clinic, University of Perugia, Perugia, Italy

<sup>3</sup> Section of Neurology, Department of Medicine, University of Perugia, Perugia, Italy;

<sup>4</sup> Health Planning Service, Department of Epidemiology, Regional Health Authority of Umbria, Perugia, Italy

<sup>5</sup> Neurology Clinic, IRCCS Ospedale Policlinico San Martino, Genoa, Italy

<sup>6</sup> Nuclear Medicine Unit, "S. Maria della Misericordia" Hospital, Perugia, Italy

<sup>7</sup> Nuclear Medicine Unit, IRCCS Ospedale Policlinico San Martino, Genoa, Italy

<sup>8</sup> Department of Health Sciences (DISSAL), University of Genoa, Italy

*Journal of Alzheimer's disease: JAD*, 72(4),1193–1207.

doi: 10.3233/JAD-190539

## Abstract

**Background.** In Alzheimer's disease (AD) diagnosis, both cerebrospinal fluid (CSF) biomarkers and FDG-PET give sometimes inconclusive results. To evaluate the incremental diagnostic value of FDG-PET over CSF biomarkers, and *vice versa*, in patients with Mild Cognitive Impairment (MCI) and suspected AD, in which the first biomarker resulted inconclusive.

**Methods:** A consecutive series of MCI patients was retrospectively selected from two Memory Clinics where, as per clinical routine, either the first biomarker choice is FDG-PET and CSF biomarkers are only used in patients with uninformative FDG-PET, or viceversa. We defined criteria of uncertainty in interpretation of FDG-PET and CSF biomarkers, according to current evidence. The final diagnosis was established according to clinical-neuropsychological follow-up of at least one year (mean  $4.4 \pm 2.2$ ).

**Results.** When CSF was used as second biomarker after FDG-PET, 14 out of 36 (39%) received informative results. Among these 14 patients, 11 (79%) were correctly classified with respect to final diagnosis, thus with a relative incremental value of CSF over FDG-PET of 30.6%. When FDG-PET was used as second biomarker, 26 out of 39 (67%) received informative results. Among these 26 patients, 15 (58%) were correctly classified by FDG-PET with respect to final diagnosis, thus with a relative incremental value over CSF of 38.5%.

**Conclusions.** Our real-world data confirm the added values of FDG-PET (or CSF) in a diagnostic pathway where CSF (or FDG-PET) was used as first biomarkers in suspected AD. These findings should be replicated in larger studies with prospective enrolment according to a Phase III design.

## 1. Introduction

Mild Cognitive Impairment (MCI) is a challenging condition due to its heterogeneous clinical presentation and pathophysiology (Stephan et al. 2012; Petersen et al. 2014). The risk of progression to dementia mainly depends on the underlying etiology (Albert et al. 2011a) and current studies are aimed at defining those biomarkers able to elucidate the specific pathophysiology, guiding clinicians to the best management in terms of diagnosis, prognosis, care, and early therapies (Berk et al. 2014). Such biomarkers are spread among centers dedicated to neurodegenerative diseases and several data are available about their specific features and accuracy (see for a review Fiandaca et al. 2014) [5], although their clinical validation is still incomplete (Frisoni et al. 2017).

Moreover, evidence about the best cost-effective and accurate flowchart guiding the use of biomarkers is still incomplete. The studies comparing biomarker performances do not definitely demonstrate the superiority of one over the others (Herukka et al. 2017; Simonsen et al. 2017). As an example, a trend in favor of a higher accuracy of FDG-PET than CSF biomarkers reported by some papers (Shaffer et al. 2013; Prestia et al. 2015; Caminiti et al. 2018) has not been confirmed by a meta-analysis with pathological confirmation, reporting instead a similar accuracy (Cure et al. 2014).

Although MCI can evolve to different types of dementia, most studies focused on Alzheimer's disease (AD) (Albert et al. 2011a) and how biomarkers of amyloidosis, tauopathy, and neurodegeneration are useful to track the disease at the MCI stage (Jack et al. 2016a). Several studies have shown that the combinatorial use of these biomarkers adds diagnostic value to the use of a single one (Bouwman et al. 2007; Vemuri et al. 2009; Choo et al. 2013; Prestia et al. 2013). However, performing all (or most) biomarkers in a MCI patient is not cost-effective since the majority of patients can be diagnosed accurately with the use of a single biomarker and only those with inconclusive results may have access to a second assessment.

In this scenario a stepwise approach could reach a cost-effective and specific diagnosis. There is active debate on which biomarker should come first and precisely for which clinical presentation. CSF biomarkers are increasingly proposed, wherever



available, if the main clinical suspicion is AD whereas FDG-PET is mainly suggested if other diagnoses are at least as likely as AD (Boccardi et al. 2019; Chételat et al. 2019). Both CSF biomarkers and FDG-PET have high diagnostic accuracy and are indeed recommended by scientific societies (Herukka et al. 2017; Arbizu et al. 2018; Nobili et al. 2018a) as a first-line biomarker in MCI patients while amyloid PET and dopaminergic imaging would have a more limited use in specific conditions (Johnson et al. 2013; Boccardi et al. 2019).

Waiting for shared and evidence-based recommendations on their alternative use as a first-line biomarker, in the real-life the first choice often mainly relies on the confidence of the clinician, local facilities, waiting lists, expertise and performance of local laboratories. In the case the first choice gave inconclusive results thus the other may be used to achieve a correct diagnosis but their reciprocal relative incremental diagnostic value when the first one is inconclusive is poorly known and partly conflicting.

Indeed, the higher predictive value of FDG-PET toward conversion to AD dementia in MCI patients reported in the ADNI population (Shaffer et al. 2013) has not been confirmed by Choo et al. (Choo et al. 2013) who reported a similar power for both FDG-PET and total Tau (t-Tau) whereas in another study using a stepwise approach the hyperphosphorylated Tau (p-Tau) achieved the best result (Lange et al. 2017).

Rather than evaluating accuracy of individual biomarkers that has already been the object of previous investigations (Fiandaca et al. 2014) in this study we evaluated the relative and reciprocal incremental diagnostic value in the attempt to establish the etiological diagnosis of CSF biomarkers over FDG-PET, and *viceversa*, only in those patients with inconclusive results after examination of the first biomarker. To this purpose we retrospectively selected data acquired for clinical purpose in naturalistic populations in two memory clinics used to apply as per clinical routine either the CSF biomarkers as a first approach and then FDG-PET when the former gives inconclusive results, or *viceversa*. In these two cohorts we first reported when the second biomarker came to a likely diagnosis and then computed how many of such diagnoses were confirmed at clinical follow-up, used as the gold standard.

## 2. Material and methods

### 2.1. Patients

Patients were retrospectively and consecutively enrolled from two memory clinics. As per clinical routine, in a patient with MCI and suspected AD the first choice to try to establish an etiological diagnosis is FDG-PET in Genoa and CSF biomarkers are only used in patients with an uninformative FDG-PET; the opposite happens in Perugia. The retrospective patient selection criteria were that i) the first biomarker was assessed soon after (i.e., within 2 months) the initial clinical evaluation because of an uncertain diagnosis following standard clinical-neuropsychological assessment and MRI; ii) AD was one of the possible diagnosis; iii) the second biomarker was performed because the first one gave uncertain results (*uninformative*) and iv) patients were clinically followed-up (FU) for at least one year. It should be underscored that we did not select those patients whose differential diagnosis did not include AD.

All patients fitted the diagnostic criteria for MCI (Petersen et al. 2009), including amnesic and non-amnesic MCI, either single- or multi-domain. Activities of daily living (ADL) and instrumental ADL (IADL) were assessed through clinical interview and formal questionnaires, and clinical dementia rating (CDR) scale had to be 0.5. Patients underwent standard clinical and neuropsychological assessment exploring verbal and spatial memory, executive functions, attention, language, and visuo-construction, according to local clinical routine. The neuropsychological batteries included standard tests with available normative values in local language and only slightly differed between centers. Brain MRI (or CT when MRI was unfeasible) was available in all patients and allowed to exclude other causes of cognitive impairment, including vascular cognitive impairment (Gorelick et al. 2011). All other causes of secondary cognitive impairment were excluded based on clinical examination and blood tests.

From January 2012 to May 2018, we identified seventy-five patients (36 males, 39 females; mean age  $70.2 \pm 7.3$ ; mean MMSE score  $26.6 \pm 2.5$ ), including thirty-six patients enrolled in Genoa and thirty-nine in Perugia. We named the two cohorts according to the second biomarker used (i.e., CSF2 when FDG-PET came first and

was uninformative, and FDGPET2 when CSF biomarkers came first and were uninformative).

**Table 1. Main clinical and demographic characteristics and CSF biomarkers values in CSF2 and FDGPET2 groups**

	CSF2 group (n = 36)	FDGPET2 group (n = 39)	p
Age (yrs)	70.9 ± 7.6	69.5 ± 7.0	n.s.
Sex (M/F)	16/20	20/19	n.s.
MMSE score	27.2 ± 2.4	26.1 ± 2.6	n.s.
CSF Aβ <sub>42</sub> (pg/mL)	682.2 ± 284.8	682.0 ± 333.6	n.s.
CSF t-Tau (pg/mL)	515.9 ± 368.9	413.9 ± 216.5	n.s.
CSF p-Tau (pg/mL)	66.1 ± 31.5	58.9 ± 20	n.s.
Δ-Time (yrs)	0.9 ± 0.5	0.3 ± 0.2	0.01
aMCI / naMCI	33 / 3	24 / 15	0.001
sdMCI / mdMCI	12 / 24	10 / 29	n.s.

Abbreviations: CSF, cerebrospinal fluid; CSF2, CSF as second biomarker; FDGPET2, [18F]-Fluorodeoxyglucose Positron Emission Tomography (FDG-PET) as second biomarker; MMSE, Mini Mental Score Examination; Aβ, amyloid-beta; t-Tau, total tau; p-Tau, hyperphosphorylated Tau; Δ-Time, mean time gap between the two biomarkers; aMCI, amnesic Mild Cognitive Impairment; naMCI, non-amnesic MCI; sdMCI, single domain MCI; mdMCI, multi-domain MCI

The final diagnosis was established according to the last available FU visit after at least one year (mean 4.4±2.2 years, range 1.02-10.61), according to current diagnostic criteria in those patients converting to dementia (Litvan et al. 1996; Gorno-Tempini et al. 2011; McKhann et al. 2011; Rascovsky et al. 2011; Postuma et al. 2015; Jack et al. 2016b; Respondek et al. 2017; McKeith et al. 2017). The managing clinician was a neurologist with expertise in dementia and was aware of biomarker results. Conversion to dementia was ascertained by clinical interview with patients and caregivers and by means of the ADL, IADL, and CDR scales.

## **2.2. CSF biomarkers**

According to the current standard operative procedures (Teunissen et al. 2009) CSF samples (6-8 mL) were collected by lumbar puncture in the L3-L4 or L4-L5 interspace and the procedure was always performed early in the morning. CSF was collected in sterile polypropylene tubes, centrifuged for 10 min at 4000g at 4 °C, and the aliquots stored in polypropylene tubes at –80 °C until analysis, to ensure long-term stability of proteins. CSF biomarkers included A $\beta$ <sub>42</sub>, t-Tau (total-Tau), and p-Tau (phosphorylated at threonine 181 or p-Tau181p) expressed in pg/mL, that were measured in both centers using commercially available enzyme-linked immunosorbent assays (ELISAs) kits (INNOTEST Fujirebio Europe, Gent, Belgium). Cut off-values have been computed according to ROC analysis and by using the highest Youden index in a large population of patients with longitudinal evaluations, diagnosed with AD and non-AD dementias, and neurological controls.

## **2.3. FDG-PET**

PET scan acquisitions were performed in both centers following standardized procedures, according to the European Association of Nuclear Medicine (EANM) guidelines (Varrone et al. 2009).-After at least 6-hour fasting, blood glucose level was measured and was lower than 7.8 mmol/l. 185 – 250MBq of [<sup>18</sup>F]-FDG were injected via a venous cannula after a 10-minute rest in a silent and obscured room, with closed eyes and unplugged ears. Subjects stayed in the room for 30 min, then they moved to the PET room where scanning started approximately 45 min. after the injection, lasting 15 min. Images were acquired by a Siemens Biograph 16 (Genoa) and a Discovery GE Healthcare ST (Perugia) PET/CT systems. Images were reconstructed with an ordered subset-expectation maximization algorithm following the standard protocols used for clinical purposes and embedded in the equipment workstations. Attenuation correction was based on CT scan.

Informed consent was obtained from all individual participants included in the study.

### 3. Data analysis and interpretation

#### 3.1. CSF biomarkers

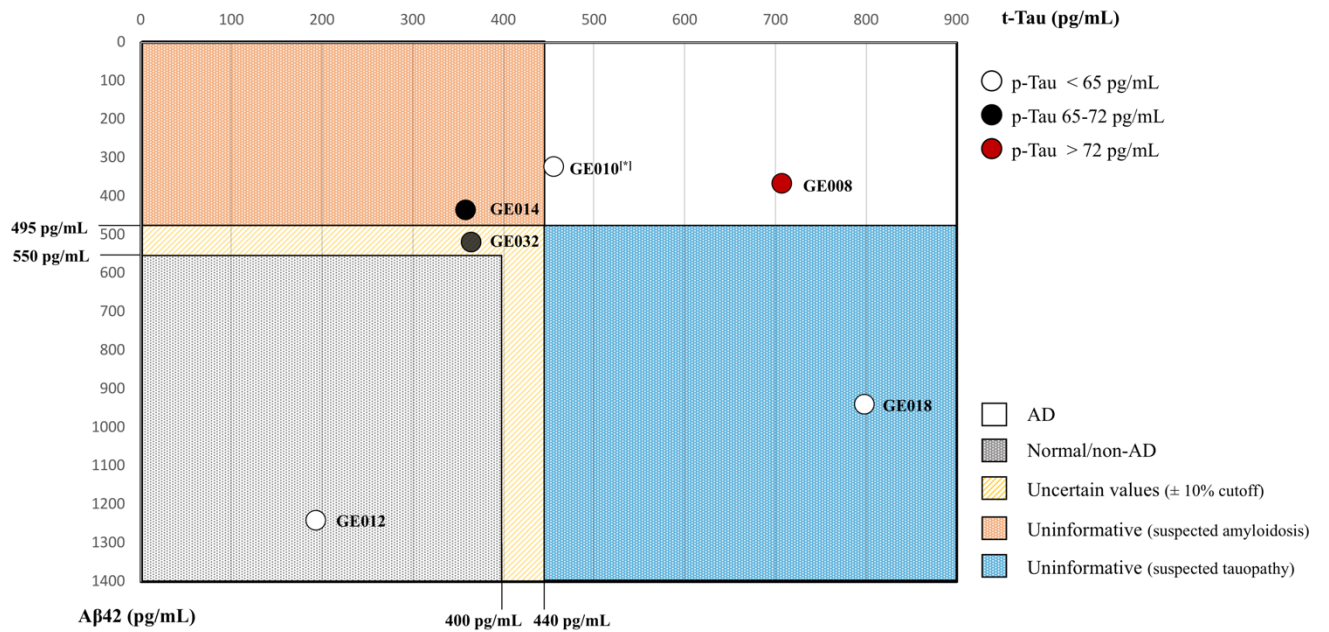
Due to small, negligible variation of the normal reference thresholds (within a 5% range) between the two Centers, we decided to choose the Perugia cutoffs. However, we wanted to further check whether the results of analyses would have substantially changed by applying the Genoa thresholds to CSF2 group (entirely made up by Genoa patients) and we obtained that a negligible percentage of patients changes category, which does not affect the results. Therefore, we choose to use the Perugia thresholds taking into consideration that this expert center regularly participates in the Alzheimer's Association external quality control program for cerebrospinal fluid biomarkers.

Cut-offs were as follows:  $A\beta_{42}$  550 pg/mL, t-Tau 400 pg/mL, p-Tau 65 pg/mL. Details of CSF biomarker results as well as borderline or uninformative results are reported in Table 2 (examples in Fig.1). In this last instance of uninformative results we computed the  $A\beta_{42}/p\text{Tau}$  ratio (cut-off 4.52, according to local normative data) to verify whether some more patient could be identified as affected by AD pathology, beyond the results of individual biomarkers (Welge et al. 2009).

**Table 2. Criteria for interpretation of CSF biomarkers**

CSF markers cutoffs	Uninformative CSF profile
<b><math>A\beta_{42} &gt; 550</math> pg/mL: normal (-)</b>	$A\beta_{42}$ (+ or +*) / p-Tau (-) / t-Tau (-)
$A\beta_{42}$ 495-550 pg/mL: uncertain (+*)	$A\beta_{42}$ (+ or +*) / p-Tau (+ or +*) / t-Tau (-)
$A\beta_{42} < 495$ pg/mL: abnormal (+)	$A\beta_{42}$ (+ or +*) / p-Tau (-) / t-Tau (+ or +*)
<b>t-Tau &lt; 400 pg/mL: normal (-)</b>	$A\beta_{42}$ (-) / p-Tau (+ or +*) / t-Tau (-)
t-Tau 400-440 pg/mL: uncertain (+*)	$A\beta_{42}$ (-) / p-Tau (-) / t-Tau (+ or +*)
t-Tau > 440 pg/mL: abnormal (+)	$A\beta_{42}$ (-) / p-Tau (+ or +*) / t-Tau (+ or +*)
<b>p-Tau &lt; 65 pg/mL: normal (-)</b>	$A\beta_{42}$ (+*) / p-Tau (+*) / t-Tau (+*)
p-Tau 65-72 pg/mL: uncertain (+*)	
p-Tau > 72 pg/mL: abnormal (+)	

\*A “grey zone” including +10% of t-Tau and p-Tau values and -10% of  $A\beta_{42}$  value was considered as uncertain (Molinuevo et al. 2014). Abbreviations: CSF, cerebrospinal fluid;  $A\beta$ , amyloid beta; t-Tau, total tau; p-Tau, hyperphosphorylated Tau



**Figure 1.** Examples of CSF biomarkers profiles in some patients of CSF2 group are shown with areas of different colours, according to the cut-offs. Each patient is expressed as a circle with different colors (white, black and red) which are determined by the value of p-Tau with respect to the cut-off.

Areas of interpretation of biomarkers (coloured squares) are: white (AD profile), grey (normal), yellow (uncertain area due to borderline values with respect to the cut-offs: +10% for p-Tau and t-Tau and -10% for Aβ<sub>42</sub>), orange (uninformative, with abnormal Aβ<sub>42</sub>, normal t-Tau and p-Tau) and light blue (uninformative, with abnormal t-Tau, normal Aβ<sub>42</sub> and p-Tau).

[\*] Notably, patient GE010 shows both abnormal Aβ<sub>42</sub> and t-Tau, though the normality of p-Tau leads to a conflicting result which is not classifiable as AD (uninformative analysis).

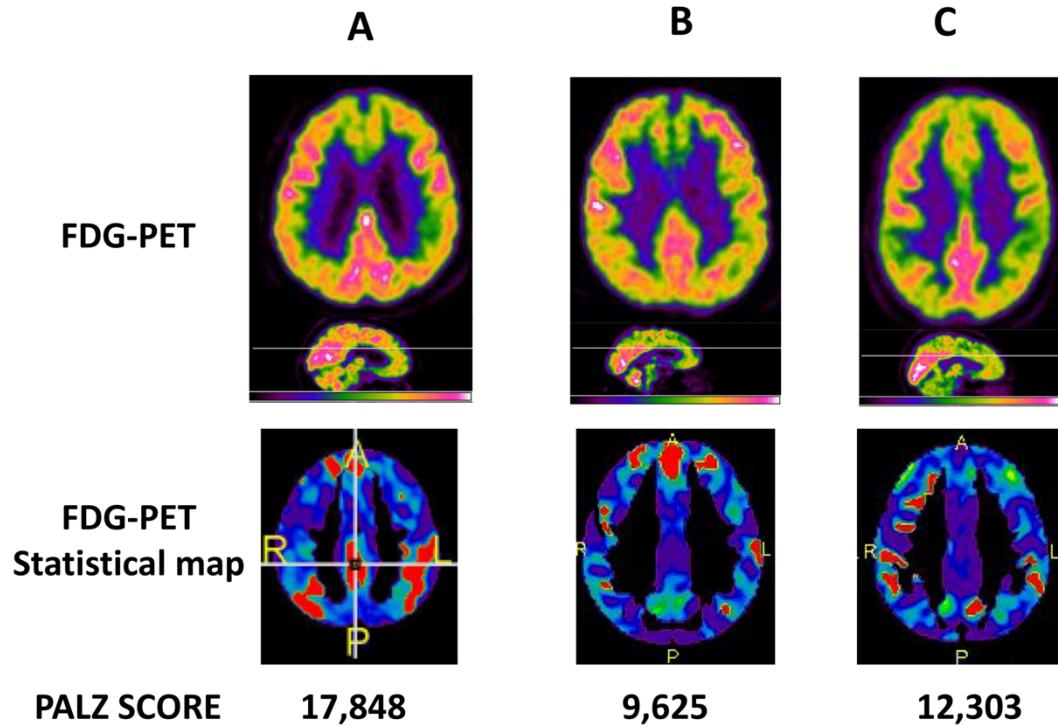
Abbreviations: CSF2, CSF as second biomarker; GE, Genoa cohort; CSF, cerebrospinal fluid; Aβ, amyloid-beta; t-Tau, total tau; p-Tau, hyperphosphorylated Tau; AD, Alzheimer's disease.

### 3.2. [<sup>18</sup>F]-FDG-PET

Scans were first visually and independently evaluated by two nuclear medicine experts (S.M. in Genoa, C.T. in Perugia), blinded to clinical history and CSF biomarker results. In fact, reading by experts is recommended by the EANM procedural guidelines (Varrone et al. 2009) and has been shown to be highly accurate (Morbelli et al. 2015a). Their judgment had to cover three options, according to brain metabolism distribution: i) normal (i.e., not compatible with either AD or other neurodegenerative diseases); ii) scan compatible with AD, which included either typical AD (clear hypometabolism in at least one brain region among precuneus/posterior cingulate and posterior temporo-parietal cortex) (Morbelli et al. 2015b) or atypical AD (left temporo-parietal cortex for logopenic variant AD (Gorno-Tempini et al. 2011); parieto-occipital cortex for posterior cortical atrophy (PCA) (Nestor et al. 2003); posterior temporo-parietal cortex plus frontal lobe involvement for frontal variant AD (Woodward et al. 2015)); iii) not compatible with AD but abnormal, when the pattern of hypometabolism involved other cortical areas and was inconsistent with those in ii). In this last instance the experts were asked to suggest an alternative diagnosis based on the metabolic pattern, also following the procedure adopted in previous papers (Grimmer et al. 2016) – i.e., behavioral variant frontotemporal dementia (bvFTD) (Rascovsky et al. 2011; Arbizu et al. 2018), semantic and non-fluent variants of frontotemporal lobe degeneration (Gorno-Tempini et al. 2011; Bouwman et al. 2018), dementia with Lewy bodies (DLB) (McKeith et al. 2017), corticobasal syndrome (CBS) and progressive supranuclear palsy (PSP) (Walker et al. 2018).

When the evaluation was discordant between the two experts, we computed the PALZ score (PMOD® Alzheimer's Discrimination Tool) obtained from semi-quantitative comparison of individual FDG-PET data with those from the normal reference population embedded in the software (cutoff > 11,090) (Herholz et al. 2002). If the PALZ score was above the cutoff (>11090) and the two experts agreed that the resulting statistical maps identified an AD hypometabolic pattern then the scan was judged to be positive for AD (example in Fig. 2-A). Otherwise FDG-PET was considered as uninformative if i) the PALZ score was below the cutoff (< 11090) (example in Fig. 2-B) or ii) the PALZ score was >11090 but the two experts did not

identify an AD typical hypometabolic pattern by evaluating the topography and extent of significant hypometabolic regions as showed by the statistical map (example in Fig. 2-C). To summarize, FDG-PET was ‘uninformative’ when it was abnormal but neither an AD-typical hypometabolic pattern was evidenced by the experts and supported by PALZ score, nor other specific hypometabolic pattern for a non-AD neurodegenerative disease was found.



**Figure 2.** Examples of reconstructed FDG-PET images showed in a trans-axial slice (according to the color bar below: white, red and yellow as ‘hot’ colors, green, blue and black as ‘cold’ colors) (top) with the corresponding trans-axial slice as shown by voxel-based analysis compared with healthy controls, corrected for age (bottom) where the significant hypometabolic voxels are reported in red.

In these three examples the two expert readers disagreed about the presence of an AD hypometabolic pattern and thus voxel-based statistical comparison was performed by means of the PALZ score computation embedded in the PMOD® software (see text for further details). A) in this patient the PALZ score was positive for AD and the two experts recognized an AD hypometabolic pattern in the statistical maps; B) in this case the PALZ score was negative for AD; C) in this last case the PALZ score was positive for AD but the two experts did not recognize an AD-typical hypometabolic pattern.



### 3.3. Diagnostic pathways

The possible pathways differed between CSF2 and FDGPET2 cohorts because pathway i) in FDGPET2 cohort ruled out all neurodegenerative conditions and not only AD as in cohort CSF2, and pathway iii) was limited to ‘uninformative’ in CSF2 cohort without the possibility to suggest an alternative diagnosis, while in FDGPET2 an alternative diagnosis could be suggested. The diagnoses suggested after obtaining the results of the second biomarker were compared with the final clinical diagnosis at FU taking into account all biomarker results that was regarded as the ‘gold standard’.

### 3.4. Statistics

Demographical features, MMSE score, clinical presentation (amnesic or non-amnesic MCI, single or multidomain) and CSF profile were compared between the two cohorts (CSF2 and FDGPET2) (t-test or chi-square test).

We then focused on the performance of the two biomarkers with a three-step computation. In either CSF2 or FDGPET2 cohort, respectively, we first computed the number (percentage) of patients in whom the second biomarker gave an informative result with respect to the CSF2 or FDGPET2 cohort. Second, we computed the number (percentage) of patients with an informative second biomarker whose diagnosis was confirmed at FU. Third, we expressed the number (percentage) of these last patients with respect to the CSF2 or FDGPET2 cohort, allowing us to compute the relative incremental value for each biomarker over the other one, as follows: relative incremental value (%) = (diagnosis suggested by the second biomarker and clinically confirmed at FU / all patients with *uninformative* first biomarker)\*100.

## 4. Results

The two cohorts showed similar age, sex distribution, MMSE score and CSF biomarkers values and only differed for the time lapse between the two biomarkers performance and for the distribution of amnestic and non-amnestic MCI (Table 1). During follow up 45/75 patients (60%) converted to and were diagnosed with a specific form of dementia, considering 18 in CSF2 group (16 AD, 2 bvFTD) and 27 in FDG2 group (17 AD, 9 bvFTD, 1 PSP).

### 4.1. CSF2 cohort (Table 3a)

CSF biomarkers gave an informative result in 14/36 patients (38.9%). In detail, CSF profile was normal (i.e., not compatible with AD, non-AD) in ten patients, compatible with AD in four patients, and was uninformative in the remaining twenty-two in whom also the A $\beta$ <sub>42</sub>/pTau ratio was not confirmatory of AD. During follow up evaluations among these 14 patients, 11 (78.6%) were confirmed at FU; indeed, all the four patients with a CSF profile compatible with AD were confirmed as affected with AD at FU, while three among the ten patients with a normal CSF were diagnosed as AD and seven were confirmed as non-AD (including three patients with masked, sub-threshold depression, two FTD phenocopies and two patients with obstructive sleep apnea syndrome).

Among the 22/36 patients with an uninformative CSF profile, nine patients had a final diagnosis of AD at FU and seven of non-AD pathologies (two with bvFTD, two with masked depression, three with amnestic MCI (aMCI) satisfying the criteria for suspected non-Alzheimer pathology (SNAP) according to CSF profile (Jack et al. 2016b), whereas six patients remained undiagnosed and phenotypically described according to their cognitive profile (Petersen et al. 2009), i.e., three with single-domain aMCI, two with multi-domain aMCI and one with non-amnestic MCI (naMCI).

Thus, 11/36 cases were clarified by CSF biomarkers and then clinically confirmed at FU (four with AD and seven non-AD final diagnoses) leading to a relative incremental diagnostic value of 30.6% of CSF biomarkers with respect to an uninformative FDG-PET to define an MCI as due to AD or non-AD.

**Table 3a. Summary of CSF2 group according to interpretation after second biomarker and the definite diagnoses at follow-up.**

CSF2					
Pt	AGE (y)	MMSE	MCI phenotype	CSF results	FINAL DIAGNOSIS
GE002	64.4	30	aMCI <sub>sd</sub>	NORMAL	AD
GE007	71.8	25	aMCI <sub>sd</sub>	NORMAL	AD
GE012	62.7	30	aMCI <sub>md</sub>	NORMAL	bvFTD phenocopy
GE016	65.6	29	aMCI <sub>md</sub>	NORMAL	DEP
GE020	73.8	26	naMCI <sub>md</sub>	NORMAL	OSAS
GE023	80.3	26	naMCI <sub>md</sub>	NORMAL	DEP
GE025	74.7	25	aMCI <sub>md</sub>	NORMAL	DEP
GE028	66.0	27	aMCI <sub>sd</sub>	NORMAL	AD
GE033	82.5	23	aMCI <sub>sd</sub>	NORMAL	bvFTD phenocopy
GE034	68.4	25	aMCI <sub>md</sub>	NORMAL	OSAS
GE008	73.1	26	aMCI <sub>md</sub>	AD	AD
GE024	72.7	29	aMCI <sub>md</sub>	AD	AD
GE030	66.9	20	aMCI <sub>md</sub>	AD	fvAD
GE035	77.7	25	aMCI <sub>md</sub>	AD	fvAD
GE001	65.3	29	aMCI <sub>sd</sub>	uninformative	aMCI <sub>sd</sub>
GE003	72.4	28	aMCI <sub>md</sub>	uninformative	aMCI <sub>md</sub> - SNAP
GE004	67.3	29	aMCI <sub>sd</sub>	uninformative	AD
GE005	73.0	29	aMCI <sub>sd</sub>	uninformative	DEP
GE006	72.6	28	aMCI <sub>md</sub>	uninformative	bvFTD
GE009	69.3	28	aMCI <sub>md</sub>	uninformative	AD
GE010	68.9	27	aMCI <sub>md</sub>	uninformative	fvAD
GE011	67.7	25	aMCI <sub>md</sub>	uninformative	AD
GE013	71.7	29	aMCI <sub>md</sub>	uninformative	aMCI <sub>md</sub> - SNAP
GE014	66.3	30	naMCI <sub>md</sub>	uninformative	naMCI <sub>md</sub>
GE015	82.1	29	aMCI <sub>md</sub>	uninformative	aMCI <sub>md</sub>
GE017	72.8	30	aMCI <sub>sd</sub>	uninformative	aMCI <sub>sd</sub>
GE018	66.4	25	aMCI <sub>sd</sub>	uninformative	aMCI <sub>sd</sub>
GE019	47.6	27	aMCI <sub>md</sub>	uninformative	DEP
GE021	76.3	29	aMCI <sub>md</sub>	uninformative	AD
GE022	76.1	27	aMCI <sub>md</sub>	uninformative	aMCI <sub>md</sub> - SNAP
GE026	81.1	29	aMCI <sub>md</sub>	uninformative	AD
GE027	78.0	22	aMCI <sub>md</sub>	uninformative	bvFTD
GE029	74.2	27	aMCI <sub>sd</sub>	uninformative	AD
GE031	71.7	29	aMCI <sub>sd</sub>	uninformative	AD
GE032	81.7	30	aMCI <sub>sd</sub>	uninformative	AD
GE036	50.4	26	aMCI <sub>md</sub>	uninformative	aMCI <sub>md</sub>

CSF2, CSF as second biomarker; Pt, patient; GE, Genoa cohort; MMSE, Mini Mental Score Examination score; CSF diagnosis, diagnosis suggested by CSF biomarker; FINAL DIAGNOSIS, diagnosis at follow-up; AD, Alzheimer's disease; fvAD, frontal variant AD; aMCI<sub>sd</sub>, amnesic Mild cognitive impairment, single-domain; aMCI<sub>md</sub>, amnesic Mild cognitive impairment, multi-domain; naMCI<sub>md</sub>, non-amnesic Mild cognitive impairment, multi-domain; bvFTD, behavioral variant-frontotemporal dementia; DEP, masked or sub-threshold depression; OSAS, obstructive sleep apnea syndrome; SNAP, suspected non-Alzheimer pathology.

#### 4.2. FDG-PET2 cohort (Table 3b)

FDG-PET gave informative results in 26/39 patients (66.7%). In detail, FDG-PET was normal in seven patients, showed a typical hypometabolic pattern for AD in twelve patients, and was not compatible with AD but showed a hypometabolic pattern suggesting another disease (i.e., FTD) in seven patients.

During follow up evaluations among these 26 patients, 15 (57.7%) were confirmed at FU – i.e. 10/12 patients with an AD typical hypometabolic pattern and 5/7 with a hypometabolic pattern suggesting FTD. On the other hand, two/12 patients with an AD hypometabolic pattern were diagnosed as bvFTD while in two patients the diagnosis suggested by FDG-PET (i.e., bvFTD and svPPA) was not confirmed at FU. Moreover, among the 7 patients with a normal scan, at FU three were diagnosed with AD, one with bvFTD and three remained undiagnosed.

Among the 13 patients with an uninformative FDG-PET, at FU three patients had a final diagnosis of AD, two patients received other diagnoses (one PSP, one bvFTD), two patients were labeled as SNAP while six remained patients were only labeled according to the MCI subtype.

Thus, in 15/39 patients a correct diagnosis was achieved by FDG-PET with a relative incremental value of 38.5% with respect to an uninformative CSF analysis to define MCI due to a specific neurodegenerative disease.

We lastly compared the main demographic and clinical characteristics between the 26/75 patients (34.7%) in whom the second biomarker allowed a corrected diagnosis as confirmed by the clinical FU (11 in CSF2 and 15 in FDG2), with the remaining 49 patients. We found that the MMSE score was significantly ( $p<0.05$ ) lower in these 26 patients (mean MMSE score  $25.8\pm2.6$ ) as compared to the remaining ones (mean MMSE score  $27.0\pm2.4$ ). No other significant difference was found as for age, sex, time gap between biomarkers, follow up time, MCI phenotype or CSF biomarker values (data not shown).

**Table 3b. Summary of FDGPET2 group according to diagnosis suggested by FDG-PET as second biomarker and the definite diagnoses at follow-up.**

FDGPET2					
Pt	AGE (y)	MMSE	MCI phenotype	FDG-PET diagnosis	FINAL DIAGNOSIS
PG001	62.5	28	aMCI <sub>sd</sub>	NORMAL	aMCI <sub>sd</sub>
PG007	75.1	21	aMCI <sub>md</sub>	NORMAL	bvFTD
PG013	72.8	28	naMCI <sub>sd</sub>	NORMAL	AD
PG028	67.3	29	naMCI <sub>md</sub>	NORMAL	AD
PG029	72.7	28	aMCI <sub>md</sub>	NORMAL	AD
PG031	64.7	26	aMCI <sub>md</sub>	NORMAL	aMCI <sub>md</sub>
PG036	52.6	30	aMCI <sub>md</sub>	NORMAL	aMCI <sub>md</sub>
PG002	65.7	26	aMCI <sub>sd</sub>	AD	bvFTD
PG005	63.7	22	aMCI <sub>md</sub>	AD	AD
PG006	61.0	23	aMCI <sub>md</sub>	AD	AD
PG010	69.6	28	aMCI <sub>sd</sub>	AD	AD
PG012	66.2	23	aMCI <sub>md</sub>	AD	bvFTD
PG025	71.5	27	aMCI <sub>sd</sub>	AD	AD
PG027	57.8	29	aMCI <sub>md</sub>	AD	AD
PG032	75.7	21	aMCI <sub>md</sub>	AD	AD
PG033	68.5	27	naMCI <sub>md</sub>	AD	AD
PG034	82.4	28	naMCI <sub>md</sub>	AD	AD
PG038	75.0	27	naMCI <sub>md</sub>	AD	AD
PG039	56.9	26	aMCI <sub>md</sub>	AD	AD
PG008	73.7	26	naMCI <sub>sd</sub>	NON-AD (bvFTD)	bvFTD
PG009	65.1	29	naMCI <sub>sd</sub>	NON-AD (bvFTD)	bvFTD
PG011	81.4	24	naMCI <sub>md</sub>	NON-AD (bvFTD)	bvFTD
PG017	72.7	25	aMCI <sub>md</sub>	NON-AD (bvFTD)	bvFTD
PG021	77.8	24	aMCI <sub>md</sub>	NON-AD (bvFTD)	bvFTD
PG023	72.2	26	aMCI <sub>md</sub>	NON-AD (svPPA)	AD
PG024	77.0	25	naMCI <sub>md</sub>	NON-AD (bvFTD)	PD-MCI
PG003	57.7	23	naMCI <sub>md</sub>	uninformative	naMCI <sub>md</sub>
PG004	71.8	29	naMCI <sub>md</sub>	uninformative	naMCI <sub>md</sub>
PG014	74.9	28	aMCI <sub>md</sub>	uninformative	aMCI <sub>md</sub> - SNAP
PG018	75.7	27	aMCI <sub>sd</sub>	uninformative	AD
PG019	72.4	28	aMCI <sub>md</sub>	uninformative	aMCI <sub>md</sub>
PG022	65.4	29	naMCI <sub>sd</sub>	uninformative	naMCI <sub>sd</sub>
PG026	57.6	26	aMCI <sub>md</sub>	uninformative	aMCI <sub>md</sub> - SNAP
PG030	74.1	20	aMCI <sub>md</sub>	uninformative	aMCI <sub>md</sub>

FDGPET2, [<sup>18</sup>F]-Fluoro-deoxyglucose Positron Emission Tomography (FDG-PET) as second biomarker; PG, Perugia cohort; FDG-PET diagnosis, diagnosis suggested by FDG-PET; svPPA, semantic variant-Primary Progressive Aphasia; PD, Parkinson's disease; PSP, progressive supranuclear palsy.

Other abbreviations as in Table 3a.

## 5. Discussion

We assessed the relative added diagnostic value of performing a second biomarker when the previous work-up, including clinical-neuropsychological evaluation, morphologic neuroimaging and a first-choice biomarker (CSF for Perugia, FDG-PET for Genoa) was inconclusive to define a MCI of a suspected AD origin.

Basing on the confirmed diagnoses at a mid-term follow-up (mean 4.4 years) used as the gold-standard, the relative incremental diagnostic values of CSF biomarkers and FDG-PET over one another was 30.6% and 38.5%, respectively (average 34.7% in the whole cohort), with a small advantage for FDG-PET.

Available literature data are somehow conflicting on the superiority of one biomarker over the other. In one study, both FDG-PET and total Tau (t-Tau) reached a similarly high predictive power toward conversion of MCI patients to AD dementia (Choo et al. 2013). In another study, hyperphosphorylated tau (p-tau) was highly sensitive to subsequent cognitive decline while FDG-PET was superior in predicting conversion to dementia in MCI patients (Fellgiebel et al. 2007). In the ADNI population, accuracy in predicting conversion to AD dementia was higher for FDG-PET (87.4%) than for CSF biomarkers (74.1%) and MRI (69.5%) with a combinatorial accuracy of 92.1% (Shaffer et al. 2013).

However, in those studies FDG-PET and CSF biomarkers, as well as other biomarkers, were performed and evaluated altogether rather than in a progressive fashion only when the first biomarker gave inconclusive results. Instead, our work focused on a ‘sequential’ approach in the decision-tree of MCI suspected for AD diagnosis, rather than a ‘in parallel’ scheme, in a similar way as in the study finding a 35% relative incremental diagnostic value of amyloid PET after an inconclusive CSF (Weston et al. 2016), practically an identical rate of increase as in the present study. Also, Lange et al. (Lange et al. 2017) meaningfully followed a stepwise order (namely ADAS-13 score, hippocampal volume (HV), CSF and FDG-PET) and showed that CSF p-Tau provided the best incremental risk stratification when added to ADAS-13 score with respect to HV and FDG-PET, while FDG-PET used in the second step outperformed HV in MCI subjects with relatively preserved cognition (ADAS-13

score <18). Moreover, CSF A $\beta$ <sub>42</sub> provided additional risk stratification in MCI subjects with increased CSF p-Tau.

Concerning the magnitude of relative incremental diagnostic value, whether the percentage of about one third of patients clarified by the second-line biomarker should be considered clinically relevant thus justifying the costs and the (mini)invasiveness of the procedures or not, this is matter of active investigation and is beyond the aims of the present study. Anyway, our results are in line with a study showing that adding CSF biomarkers to FDG-PET led to a small but measurable increase in diagnostic accuracy (Zhang et al. 2011). Also, an increase in diagnostic accuracy was shown using not precisely the same biomarkers as ours. A benefit of combining MRI and FDG-PET in predicting conversion to AD dementia with respect to the single biomarker was reported (Walhovd et al. 2010) and a further 6% AUC value increase was demonstrated from the best single (t-Tau, AUC 0.77) to the best three-predictors combination model (t-Tau/HCV/CDR-sum of boxes, AUC = 0.83) in predicting conversion to AD dementia (Frölich et al. 2017).

Taken together, these as well as our data suggest that using biomarkers with a progressive approach allows to increase the diagnostic accuracy although a non-trivial part of patients remains undiagnosed. This means that those patients leading to clinician uncertainty after basal clinical-neuropsychological assessment, MRI and a first biomarker are likely to be the most complex ones, including those with mixed pathologies, confounding factors, or atypical presentations, or a mix of these conditions (Jellinger and Attems 2010; Carotenuto et al. 2012; Dubois et al. 2014a). To reinforce this interpretation, in 20 patients of the whole our cohort (26.7 %) an etiologic diagnosis was not achieved even at follow-up, as MCI patients did not convert to dementia and, furthermore, no specific suggestive features about the underlying etiology could be unveiled, including 5 of them who satisfied the criteria for SNAP according to CSF profile (Jack et al. 2016b). The only factor that distinguished those patients taking advantage from a second biomarker assessment was a significantly lower MMSE score although the overlap with those remaining undiagnosed was considerable.

Another point of our results deserving discussion is that the majority (65%) of diagnoses suggested by an informative second-line biomarker were confirmed at

follow-up, in this case with an advantage of CSF biomarkers (78.6%) over FDG-PET (57.7%), although the clinician was not blind to the results of biomarker itself because this is a retrospective study with data collected from the real clinical practice. As we lacked pathological confirmation and used the clinical follow-up as the gold-standard, we cannot firmly establish that in the remaining 35% of patients the second biomarker ‘was wrong’ because we know the clinical diagnosis carries in turn a considerable risk of errors (Beach et al. 2012), especially if we consider that here the final clinical diagnosis tells against the suggestion given by the second biomarker. Moreover, even though mean clinical FU was  $4.4 \pm 2.2$  years, in 11 patients it was less than 2 years (of whom 6 patients < 18 months), so we might have missed some late converters who could have been properly diagnosed with a longer period of observation.

Moving to more technical considerations, as for CSF biomarker a grey zone of 10% above (t-Tau or p-Tau) or below ( $A\beta_{42}$ ) is admitted (Molinuevo et al. 2014; Lewczuk et al. 2015; Simonsen et al. 2017). Accordingly, we considered as uninformative CSF biomarkers when at least one of the three proteins was conflicting with the expected ‘AD signature’ or included in the grey zone (Molinuevo et al. 2014). Such strategy could be responsible for the high rate of uncertain interpretations (22/36), but was not mitigated by our attempt to overcome this limitation by using the  $A\beta_{42}/p\text{Tau}$  ratio. This is in line with the knowledge that almost half of the patients with AD may have conflicting CSF biomarkers results (Alexopoulos et al. 2016), only partly due to a false initial diagnosis or to adopted cutoffs and laboratory performance (Rosén et al. 2015; Herukka et al. 2017).

Indeed, these uncertain cases reflect at least in part the heterogeneous composition of MCI population as low CSF  $A\beta_{42}$  may be found in non-AD conditions such as Parkinson's disease and DLB (Kaerst et al. 2013), while Tau levels may be elevated in several other conditions expressing neurodegeneration (t-Tau), tauopathy (p-Tau), or both (van Harten et al. 2011).

Another part of variance may derive from pre-analytical and analytical issues since the sampling was realized in the course of years, and even if collection and analytical procedures were the exactly the same, some practical conditions may have changed, mainly depending on the technical skills which inevitably increase



paralleling experience. Lastly, some variance may derive from suboptimal inter- and intra-center reproducibility using commercial kits (Verwey et al. 2009).

Lastly, we now know that the  $A\beta_{42}/A\beta_{40}$  ratio is more accurate than the  $A\beta_{42}$  assay alone (Lewczuk et al. 2014; Dorey et al. 2015; Biscetti et al. 2019), but our casuistic was retrospectively collected when only the  $A\beta_{42}$  assay was available. It is likely that adding the ratio  $A\beta_{42}/A\beta_{40}$  could have solved some more cases, especially those with an uncertain  $A\beta_{42}$  value.

Obviously, when using such biomarkers we should have clearly in mind what information we expect from them. As said above, as for clinical routine the two centers have a different use of biomarkers, which mainly depends on a different diagnostic strategy to infer the pathological processes underlying MCI. Notwithstanding, as per recruitment criteria all patients should have had a MCI primarily suspected for AD. Such specific pre-requisite prevented any selection bias and allowed us to specifically focus on the reliability of CSF and [ $^{18}\text{F}$ ]-FDG PET to confirm or rule out AD.

Within the context of current diagnostic criteria (Albert et al. 2011a) our results confirm the high specificity of CSF markers in AD as they are able to simultaneously assess the main pathological features which confer the highest likelihood that AD is the cause of MCI (Dubois et al. 2014b; Jack et al. 2018). Moreover, our results enforced the role of [ $^{18}\text{F}$ ]-FDG PET as a reliable diagnostic tool not restricted only to AD (Nobili et al. 2018a). Thus, as for FDG-PET, the main advantage seems that an alternative diagnosis can be suggested when an AD-like hypometabolic pattern is not found, as in the case of FTD. This further stresses the role of FDG-PET in other etiologies than AD, in keeping with the recent EANM-EAN joint recommendations (Nobili et al. 2018a). On the other hand, FDG-PET is a non-pathology specific marker, thus in some instances the imaging endophenotype may mimic another disease by showing an unusual topographic pattern, and this is likely the explanation of some of the diagnostic mistakes we observed in the present series. Just as an example, the concomitant frontal and parietal hypometabolic pattern can be found in patients with fvAD but also in an occasional patient with FTD in which parietal hypometabolism may be due to the fronto-parietal disconnection through the fronto-parietal pathways (Johnson et al. 1999; Womack et al. 2011; Woodward et al. 2015).

Both for CSF analysis and FDG-PET we are aware of the paramount importance of laboratory and readers expertise in borderline or conflicting results which may predispose to misinterpretation (Herholz 2014; Fourier et al. 2015; Lewczuk et al. 2018). As a consequence, some of the mistakes in the present research may have been derived from analytical issues rather than from the intrinsic limitations of the diagnostic tools *per se*, but we described what happens in clinical practice in two expert centers, even with different field of expertise, thus very close to the real life in most centers worldwide. Also, the different time gap between biomarkers, which was significantly longer when CSF biomarkers were used after FDG-PET, is mainly be due to the reluctant approach (in the past years) of both physicians and patients/caregivers to the lumbar puncture in the Genoa center.

In conclusion, finding the best way to get to the highest diagnostic accuracy by using the least number of biomarkers is of paramount relevance not only for clinicians but also for national healthcare systems, as people with MCI are expected to progressively increase, along with costs given by diagnostic procedures, welfare and hopefully upcoming disease-modifying drugs. The main strength of our work is to have involved naturalistic cohorts, not in the set of clinical trials with selected patients, and thus it is more adherent to the real world; this let us to propose our evidences also in the clinical scenario in which MCI is a diagnostic challenge as a definite diagnosis could not be achieved even after a second-line biomarker evaluation in a substantial part of cases. Anyway, the retrospective nature of our study as well as to have focused on a selected cohort in which the first additional biomarker was unable to clarify the etiology are the main limits which require caution in generalizing the results.

Moreover, Amyloid PET has been routinely used in our centers only for 2-3 years and thus it is unavailable in the vast majority of our cases; consequently, we are far from having a sufficient number of patients to explore its role in this context. Amyloid PET has been reported to better predict AD-dementia conversion than CSF (Ben Bouallègue et al. 2017). Thus, it might be the case that some still uncertain diagnoses even after CSF analysis and [<sup>18</sup>F]-FDG PET or some cases which were not properly classified could have been clarified by amyloid PET, as well as by the ratio A $\beta$ 1-42/A $\beta$ 1-40, as discussed before.

The utility of our study is to have assessed the relative incremental value of two of the most used and widespread biomarkers in the diagnostic workup of MCI and to have highlighted the complexity of uncertain diagnoses. In particular we found a small advantage for FDG-PET with respect to CSF in achieving the most accurate diagnosis; thus, from a practical standpoint we might recommend FDG-PET when the diagnostic scenario has not been narrowed by first-line assessments, and still includes different diseases other than AD; on the other hand adding CSF might be helpful when we need a specific confirm of a justified suspicion of AD which FDG-PET at first was not able to solve.

According to this finding, one may want to access the second biomarker only if the cognitive deficit is more marked and, as a suggestion, if it is further confirmed in subsequent visits in the short term, but this hypothesis requires further studies. Moreover, as we found a prevalence of naMCI in the FDG2 group we hypothesize that a patient with naMCI could more likely have an inconclusive CSF compared to patients with aMCI, the typical presentation of AD and with a higher likelihood to exhibit a ‘typical’ CSF AD profile. Thus, naMCI patients who are still suspected of AD (as in our cohorts) are usually further assessed with a second biomarker (FDG-PET in our case) in the attempt to achieve an etiological diagnosis. On the other hand, aMCI might be underpinned by different conditions other than AD (such as SNAP, cerebrovascular disease, and even subthreshold depression); thus, an aMCI patient with an inconclusive FDG-PET might need CSF analysis as a second biomarker to suggest or rule out AD. Another unmet need is the potential role of either of the two biomarkers as gate keeper after neuropsychology and MRI, especially taking into account the age. Increasing evidence is supporting the use of amyloid biomarkers for subjects with MCI of suspected AD origin who are younger than 65, due to the high rate of cerebral amyloid pathology of unknown significance in cognitively unimpaired older people (Jansen et al. 2015). However, the specific role for FDG-PET as gate keeper for those who are older has never been investigated and goes beyond the aims of the present study.

Efforts should be directed towards the definition of a diagnostic-tree that considers either the main features of each biomarker, alone or in sequential combination, and a cost-effective approach to identify those patients in which adding

a second biomarker could be really useful. We believe the results of this retrospective and relatively small-number study would deserve larger and prospective studies to assess the reliability and clinical utility of sequential biomarker measurement, according to clinical presentation.

## **4. Exploring the brain metabolic correlates of process-specific CSF biomarkers in patients with MCI due to Alzheimer's disease: preliminary data**

<sup>1</sup>Federico Massa, <sup>2</sup>Steffen Halbgebauer, <sup>2,3</sup>Lorenzo Barba, <sup>2,4</sup>Patrick Oeckl, <sup>2</sup>Nerea Gómez de San José, <sup>5</sup>Matteo Bauckneht, <sup>6</sup>Francesco Lanfranchi, <sup>5</sup>Tiziana Vigo, <sup>1,5</sup>Dario Arnaldi, <sup>1,5</sup>Matteo Pardini, <sup>5,6</sup>Silvia Morbelli, <sup>7</sup>Andrea Chincarini, <sup>8</sup>Henryk Barthel, <sup>2,3</sup>Markus Otto and <sup>1,5</sup>Flavio Nobili

<sup>1</sup>Department of Neuroscience, Rehabilitation, Ophthalmology, Genetics, Maternal and Child Health (DINO GMI), University of Genoa, Genoa, Italy

<sup>2</sup>Department of Neurology, Ulm University Hospital, Ulm, Germany

<sup>3</sup>Department of Neurology, Martin-Luther-University of Halle-Wittenberg, Halle/Saale, Germany

<sup>4</sup>German Center for Neurodegenerative Diseases (DZNE e.v.), Ulm, Germany

<sup>5</sup>IRCCS Ospedale Policlinico San Martino, Genoa, Italy

<sup>6</sup>Department of Health Science (DISSAL), University of Genoa, Italy

<sup>7</sup>National Institute of Nuclear Physics (INFN), Genoa section, Genoa, Italy

<sup>8</sup>Department of Nuclear Medicine, University Hospital of Leipzig, Leipzig, Germany

*Neurobiology of aging*, 117, 212–221.

<https://doi.org/10.1016/j.neurobiolaging.2022.03.019>

## Abstract

**Introduction.** Cerebrospinal fluid (CSF) biomarkers can reflect several molecular aberrations and pathological changes in Alzheimer's disease (AD), e.g., neuronal death, synaptic and axonal injury. [<sup>18</sup>F]-fluorodeoxyglucose-PET(FDG-PET) brain metabolism might disclose a peculiar topography when correlated with the levels of some CSF proteins reflecting specific pathological processes.

**Methods.** We focused on some emergent CSF biomarkers, namely post-synaptic neurogranin (Ng), pre-synaptic  $\alpha$ - and  $\beta$ -synuclein ( $\alpha$ - and  $\beta$ -syn), and neurofilament light chain (NfL), as a marker of axonal damage, and explored the sites of correlation (volumes of interest, VOIs) of their levels with brain metabolism in a group of 26 patients with prodromal AD (16 females; age  $75.4 \pm 6.6$ ; MMSE score  $26.1 \pm 1.9$ ). We further assessed whether-and how extensively- these VOIs overlapped the hypometabolic areas resulting from comparing AD patients with 40 matched healthy controls (HC).

**Results.** Ng-VOI and  $\alpha$ -syn-VOI encompassed left precuneus/posterior cingulate cortex (PC/PCC) and partially overlapped hypometabolism at those sites.  $\beta$ -syn-VOI and NfL-VOI regarded either left or right lateral temporal areas, respectively, with partial overlap with hypometabolism for the  $\beta$ -syn-VOI, whereas the NfL-VOI did not include hypometabolic regions (Figure 1).

**Discussion.** We speculate that CSF levels of Ng and  $\alpha$ -syn express an already established hippocampal damage leading to PC/PCC deafferentation and hypometabolism.  $\beta$ -syn may represent the progression of synaptopathy in the temporal lobe, while NfL the axonal injury in less affected right temporal regions where neuronal loss is still subthreshold. These findings complement the information on the distribution of hypometabolism related to neuronal loss, which differs from the metabolic changes reflecting synaptic or axonal injury.

## 1. Introduction

In the past years, the research provided a deep insight into the pathogenesis of Alzheimer's disease (AD), and we can now assess many molecular aberrations and pathological changes through cerebrospinal fluid (CSF) analysis. Beyond the core CSF biomarkers of amyloidosis (amyloid  $\beta$  1-42 normalized for amyloid  $\beta$  1-40 values, A $\beta$ 42/A $\beta$ 40 ratio), tauopathy (phosphorylated Tau at threonine, position 181, p-Tau181), and neuronal death (total Tau, t-Tau), others can reveal concurrent pathological events of AD, e.g., axonal injury and synaptic degeneration. Both mechanisms have gained interest as early processes because of the consistent experimental and pathological evidence describing a particular dying-back pattern of neuronal degeneration in AD, where significant impairment in synaptic function and axonal connectivity precedes neuronal death (Kanaan et al. 2013). Neurofilament light chain (NfL) is the most reliable marker of axonal damage in inflammatory and degenerative CNS conditions (Gaetani et al. 2019). CSF NfL level rises in AD already in the mild cognitive impairment (MCI-AD) stage and correlates with cognition, amyloid load, and cortical atrophy (Zetterberg et al. 2016; Dhiman et al. 2020). Similarly, synaptic degeneration is an early pathogenic event in many neurodegenerative disorders and associates with cognitive symptoms in AD (Terry et al. 1991; Blennow et al. 1996; Selkoe 2002; Lleó et al. 2019). Several pre- and post-synaptic proteins were described in the last years, but evidence of their involvement in the pathophysiology of AD is sparse (Camporesi et al. 2020). Among them, neurogranin (Ng), a dendritic protein revealing post-synaptic integrity, and two pre-synaptic proteins of the family of synucleins (beta-synuclein,  $\beta$ -syn, and  $\alpha$ -synuclein,  $\alpha$ -syn) increase in the CSF of AD patients along with cognitive impairment and disease progression (Wellington et al. 2016; Oeckl et al. 2016, 2020; Monge-Argilés et al. 2020; Halbgebauer et al. 2021).

[18]F-fluorodeoxyglucose positron emission tomography (henceforth FDG-PET) is recommended for early and differential diagnosis in patients with suspected

AD (Garibotto et al. 2017; Nobili et al. 2018a) and is acknowledged as a biomarker of downstream neurodegeneration in the research framework for AD (Jack et al. 2018). FDG-PET is a multifaceted measure of neuronal-astrocyte activity, neuropil loss, and synaptic density (Herholz 2003; Zimmer et al. 2017), much more complex than the other neurodegeneration biomarkers and thus leading to suggest a ‘fourth,’ independent factor in the A/T/N system, the A/T/N/F system (Ou et al. 2019). FDG-PET is also sensitive to deafferentation, diaschisis phenomena, and axonal damage (Nakashima et al. 2007; Provost et al. 2021). These processes change their relative weight throughout the AD course and contribute to the differences in topography and intensity of hypometabolism between patients and across stages (Brown et al., 2014; Mosconi, 2005; Silverman et al., 2001). Within this framework, one might expect particular topographic correlations with those CSF proteins that reflect specific underlying pathological processes. Unveiling these complex relationships is even more relevant in the earliest phases of AD when understanding biomarkers changes could help to stage patients across the disease spectrum, improve phenotyping, and suggest the risk of progression to dementia.

We focused on a cohort of patients with MCI-AD to explore the brain metabolic correlates of some emerging CSF biomarkers (Ng,  $\beta$ -syn,  $\alpha$ -syn, and NfL). Our goal was twofold. On the one hand, to investigate the topographic correlation of these biomarkers with brain glucose metabolism. On the other hand, to unravel whether -and to what extent- these correlating sites overlap the hypometabolic areas or regard still relatively preserved regions. This preliminary data may be helpful in terms of learning more about the value of altered CSF proteins in the prodromal phase of AD and the meaning of hypometabolism in specific areas.

## **2. Materials and methods**

### **2.1 Patients**

We retrospectively selected a cohort of 26 patients (16 females; age  $75.4 \pm 6.6$  (range 66.0-76.6); education  $9.6 \pm 4.0$  yrs. (range 5-17); MMSE score  $26.1 \pm 1.9$  (range 23-29) with typical, high-likelihood MCI-AD according to NIA-AA criteria (Albert et al. 2011b). They were extracted from the database of patients diagnosed with MCI-

AD at the memory clinic of the University of Genoa from June 2018 to January 2021. All patients had pathological values of A $\beta$ 42/A $\beta$ 40 ratio and p-Tau181, irrespective of the level of t-Tau, according to the AT(N) biomarker profile (Jack et al. 2018). Further details are in section 2.2.2.

These patients were selected because they also underwent FDG-PET during the diagnostic workup, within two months from CSF examination. At their first evaluation, they underwent neurological, general medical examinations, and an extended battery of neuropsychological tests, which defined their condition as amnesic MCI, either single- or multidomain (Petersen et al. 2009). We excluded patients with a non-amnesic presentation and performed blood tests and brain MRI to rule out non-degenerative causes of cognitive deficit. The presence of white matter hyperintensities was not an exclusion criterion if the Wahlund score was <2 in each site (Wahlund et al. 2001).

Eleven out of 26 patients converted to typical AD dementia after a mean of  $1.2 \pm 0.3$  years from baseline (range 0.65-1.8 years) (fast-converted), whereas the other 15 remained in the MCI stage after the last 2-year follow-up visit. We label these subjects as ‘non-converted’. Because follow-up time is still limited, we are aware that most of them will probably convert to AD dementia in later years, thus virtually including only ‘late’ converters.

Finally, we included a healthy control (HC) group made up of 40 age-, sex- and education-matched volunteers who performed FDG-PET during previous research (20 females; age  $75.6 \pm 5.0$  (range 54.9-85.1); education  $10.7 \pm 3.6$  yrs. (range 5-17); MMSE score  $28.9 \pm 1.0$  (range 27-29). We checked their health condition according to clinical history and examination, with MMSE score >27, CDR=0, and a normal FDG-PET scan as established by the independent visual reading of two expert nuclear medicine physicians

All subjects gave their written consent to use anonymized data for research purposes according to the university hospital’s rules for using retrospective data collected during the clinical routine. The study strictly followed the principles of the Helsinki declaration.



## **2.2 CSF biomarkers**

### **2.2.1. CSF collection and pre-processing**

According to the standard operating procedures, CSF samples (6–8 mL) were collected by lumbar puncture in the L3-L4 or L4-L5 interspace early in the morning (Teunissen et al. 2009). We collected CSF in sterile polypropylene tubes, centrifuged for 10 min at 4000 g at 4°C, and stored the aliquots in polypropylene tubes at –80°C until analysis to ensure long-term stability of proteins.

### **2.2.2. Measurement**

In all patients, we measured the core CSF AD biomarkers (A $\beta$ 42, A $\beta$ 40, p-Tau181, and t-Tau) through the Lumipulse G600 II® fully automated chemiluminescent enzyme immunoassay system (Fujirebio Europe, Gent, Belgium). The standard cut-off values reported in the datasheets of the biomarker assay cartridges by the manufacturers were validated in our center as well: 0.069 for A $\beta$ 42/A $\beta$ 40 ratio, 56.5 pg/mL for p-Tau181, and 404 pg/mL for t-Tau. Samples with pathological levels of both A $\beta$ 42/A $\beta$ 40 ratio and p-Tau181, with either positive or negative t-Tau levels (n=23 and n=3, respectively), were classified as AD (A+T+(N)) (Jack et al. 2018). However, those three patients had an impaired FDG-PET so that all patients were actually A+T+N+.

In the same samples, we performed the quantitative determination in CSF of i) Ng (trunc p75) and  $\alpha$ -syn using commercially available ELISA kits (Euroimmun, Lubeck, Germany) ii)  $\beta$ -syn by the recently established sandwich ELISA method, as described in detail in Halbgebauer et al. (2021) iii) NfL through Simple Plex™ Ella™ automated microfluidic platform (ProteinSimple). Concentration was in pg/ml for all these proteins. The average coefficient of variation (CV) for intra- and inter-test reproducibility was lower than 10% and 15%, respectively, for all the assays.

## **2.3 FDG-PET**

### **2.3.1 FDG-PET protocol and image pre-processing**

We acquired FDG-PET scans at baseline according to the European Association of Nuclear Medicine (EANM) guidelines (Varrone et al. 2009). The measured blood glucose level was less than 7.8 mmol/l after at least 6 hours of fasting. After a 10-minute rest in a quiet, dimly lit room with unplugged ears and open eyes, 185-250 MBq of [<sup>18</sup>F]-FDG was injected via a venous cannula. Approximately 45 minutes after the injection, the PET scan began and lasted 15 minutes (Siemens Biograph 16 PET/CT system). We reconstructed the acquired images with an ordered subset-expectation maximization algorithm following the standard protocols used for clinical purposes and embedded in the equipment workstations. Attenuation correction was based on CT scan.

We used MATLAB and Statistical Parametric Mapping software (SPM12, Wellcome Trust Center for Neuroimaging, London, UK) for image pre-processing. PET images were normalized into a specific FDG-PET template in the MNI stereotaxic space (Della Rosa et al. 2014) and resampled to 2x2x2 mm<sup>3</sup> voxels, then spatially smoothed using a 10-mm isotropic Gaussian filter.

A neurologist (FM) and a nuclear medicine physician (SM) with expertise in PET analysis and interpretation individually re-validated each image to check the quality and correctness of the pre-processing steps.

### **2.3.2 Voxel-based group analysis**

After pre-processing, smoothed images were subjected to whole-brain voxel-wise group analyses, including:

- 1) comparison between MCI-AD and HC groups (two-sample t-test; nuisance: age).
- 2) linear correlation analysis between relative brain metabolism and CSF levels of NfL, Ng,  $\beta$ -syn, and  $\alpha$ -syn, respectively, in MCI-AD patients (multiple regression analysis; nuisances: age and MMSE score).

We applied the standard 0.8 gray matter threshold masking and the default value of 50 for the grand mean scaling in all the analyses. The output was an SPM t-Map showing the statistically significant clusters using a height threshold of  $p < 0.001$ , uncorrected for multiple comparisons at the voxel level. We considered only clusters of at least 50 voxels if statistically significant with  $p < 0.05$ , family-wise corrected

(FWE) for multiple comparisons at the cluster level. We converted clusters coordinates using Ginger Ale (Eickhoff et al. 2009) and Talairach Client software to the Talairach's 3D coordinates system (Lancaster et al. 2000) to identify resultant cerebral areas according to the Brodmann classification.

### **2.3.3 Volumes of interest (VOI) analysis**

The volumes of interest (VOIs) were identified using the significant clusters deriving from i) comparison between MCI-AD and HC groups and ii) correlation of each CSF biomarker level with brain metabolism. The VOI count densities in each cluster were extracted using the Marsbar toolbox implemented in SPM, then scaled to the whole brain count density in each patient.

## **2.4. Statistics**

We compared demographic and clinical data between two subgroups of MCI-AD patients, namely fast- and non-converted. We used unpaired two-tailed T-test, or Wilcoxon rank-sum test when needed, to compare continuous data, and Fisher's exact test for categorical variables.

Furthermore, we explored differences between fast- and non-converted patients in terms of the whole-brain scaled average count densities of each of the significant VOIs (Wilcoxon rank-sum test with false discovery rate, FDR, method to account for multiple comparisons).

$p < 0.05$  was considered the first level of statistical significance for all the analyses.

We computed the overlap coefficient (in percentage) to estimate the spatial interactions among the different VOIs as: (the size of the intersection of two VOIs divided by the smaller of the two VOIs) \* 100.

### 3. Results

#### 3.1. Comparison between patients with MCI-AD and HC

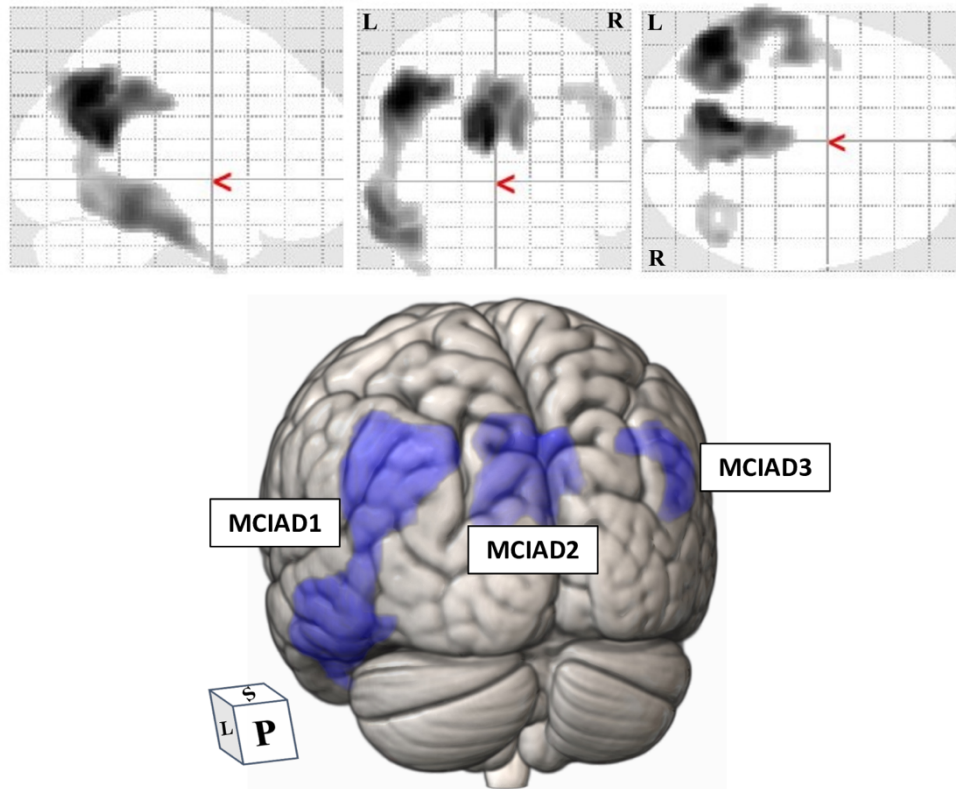
The main clinical features and biomarker values of MCI-AD patients are summarized in Table 1.

	<b>MCI-AD (n=26)</b>	<b>Late/non-converted (n=15)</b>	<b>Fast-converted (n=11)</b>	<b>p-value<sup>#</sup></b>
Age (y)	75.4 ± 6.6	74.4 ± 4.4	76.8 ± 8.7	0.42 <sup>*</sup>
Sex (M:F)	10:16	5:10	5:6	0.69 <sup>#</sup>
Education	9.6±4.0	10.7 ± 3.5	8.2±4.4	0.17 <sup>*</sup>
MMSE score	26.1±1.9	27.0±1.3	24.9 ±2.0	<b>0.007<sup>*</sup></b>
Right-handed	26/26	15/15	11/11	1
Aβ42 (pg/ml)	507.5±212.3	515.5±218.9	496.5±213.0	0.90 <sup>§</sup>
Aβ42/ Aβ40 ratio	0.043±0.01	0.041±0.010	0.046±0.009	0.28 <sup>§</sup>
p-Tau181 (pg/ml)	113.3±42.3	113.7±46.3	112.9±38.3	0.80 <sup>§</sup>
t-Tau (pg/ml)	718.5±253.2	731.4±275.3	700.3±231.5	0.80 <sup>§</sup>
Ng (pg/ml)	594.8±196.1.	572.6±191.3	625.0±207.8	0.36 <sup>§</sup>
α-syn (pg/ml)	2027.4±583.2	2015.0±626.1	2044.4±548.5	0.90 <sup>§</sup>
β-syn (pg/ml)	836.2±331.5	864.7±366.7	861.2±294.1	0.88 <sup>§</sup>
NfL (pg/ml)	1227.6±842.2	1044.6±631.1	1477.3±1047.3	0.33 <sup>§</sup>

\*p-Values for unpaired two-tailed T-test. <sup>#</sup>p-Values for Fischer's exact test; <sup>§</sup> p-values for Wilcoxon rank-sum test. Significant values are in **bold**.

Abbreviations: MCI-AD, Mild cognitive impairment due to Alzheimer's disease; F, female; M, male; MMSE, Mini Mental State Examination; Aβ42/Aβ40 ratio, amyloid Aβ1-42 normalized for amyloid Aβ1-40; pTau181, Tau phosphorylated at threonine 181; t-Tau, total Tau; Ng, neurogranin; α-syn, alpha-synuclein; β-syn, beta-synuclein; NfL, neurofilament light chain

We found three clusters (MCIAD1-VOI, MCIAD2-VOI, and MCIAD3-VOI) of significant relative hypometabolism in MCI-AD patients compared to healthy subjects. These clusters encompassed the bilateral precuneus/posterior cingulate (PC/PCC) and temporo-parietal junction, with maximal expression in the left hemisphere (Figure 1, Table 2).



**Figure 1.** In the upper part, T-maps as seen from the right, back and top of the three clusters (MCIAD1, MCIAD2, and MCIAD3) from the direct comparison between MCI-AD and HC groups (height threshold of  $p < 0.001$ , uncorrected for multiple comparisons at peak level;  $p < 0.05$  FWE for multiple comparisons at cluster level).

In the lower part, 3D rendering of the same clusters (shown in purple) in the MNI referential atlas (MRICroGL software, <https://www.nitrc.org/projects/mricrogl>).

Abbreviations: L, left; R, right; P, posterior; S, superior

**Table 2. Significant brain areas resulting from the comparisons of brain metabolism between MCI-AD and HC groups and from the correlation of CSF biomarkers with brain metabolism**

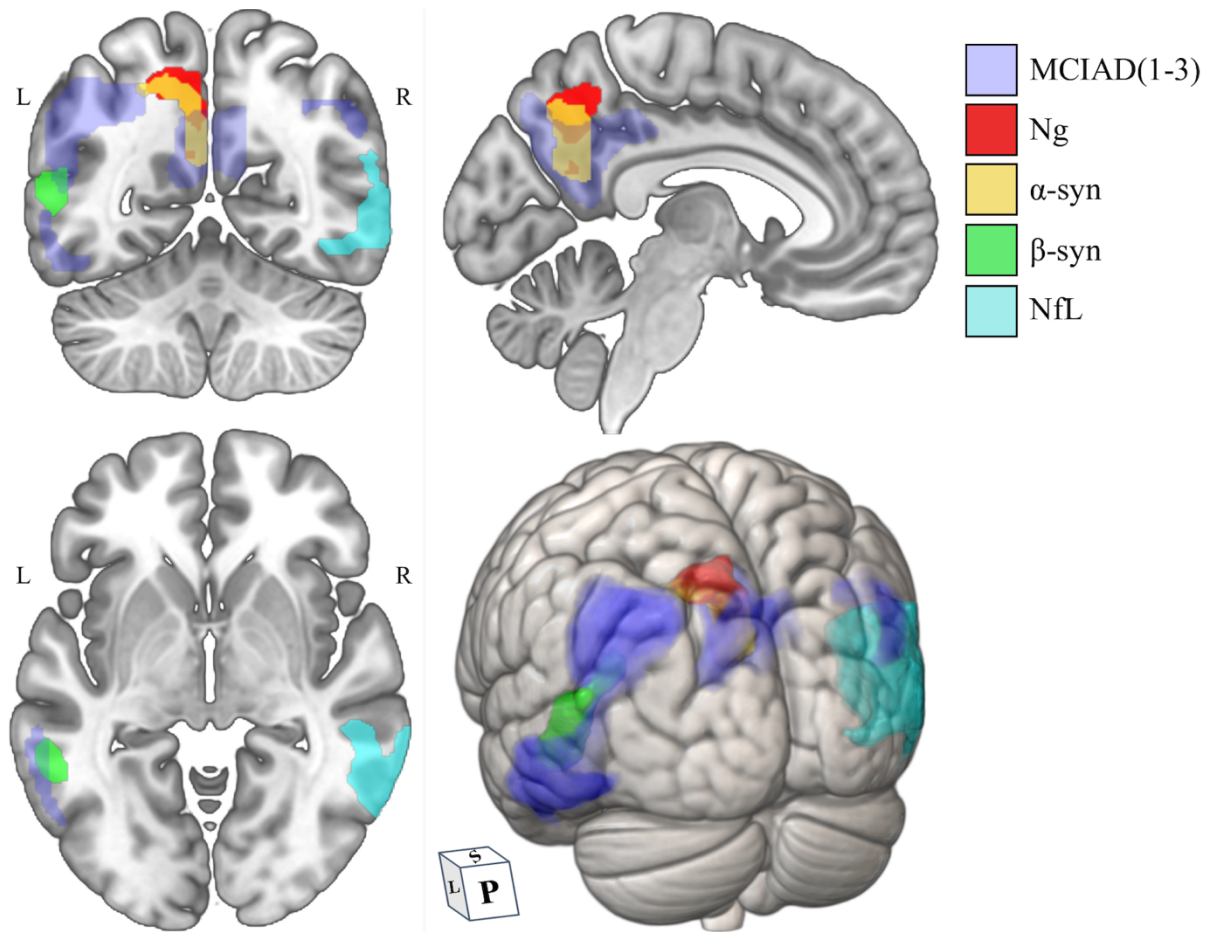
Hypometabolism in MCI-AD with respect to HC group (MCIAD-VOIs)						
Cluster extent	Cluster Significance	Cluster peaks coordinates			Cortical region (gyrus)	BA
		x	y	z		
4302 (MCIAD1-VOI)	p < .05 FWE corr	-46.04	-64.47	31.62	Left angular	39
		-33.09	-55.39	34.5	Left angular	39
		-56.43	-39.04	-16.59	Left inferior temporal	20
		-56.29	-19.71	-21.96	Left fusiform	20
		-56.36	-29.37	-19.28	Left inferior temporal	20
		-50.98	-58.05	-14.7	Left fusiform	37
		-51.2	-63.34	1.01	Left middle temporal	*
		-37.51	4.12	-35.6	Left middle temporal	21
		-41.24	3.97	-33.88	Left middle temporal	21
		-6.97	-55.82	18.69	Left posterior cingulate	23
2896 (MCIAD2-VOI)	p < .05 FWE corr	-8.94	-67.69	24.74	Left precuneus	31
		-12.54	-65.11	17.72	Left posterior cingulate	31
		-3.44	-42.51	36.23	Left cingulate	31
		5.99	-54.03	19.08	Right posterior cingulate	23
		7.72	-58.63	27.68	Right cingulate	31
		-1.49	-30.81	31.96	Left cingulate	31
		5.85	-66.08	26.94	Right precuneus	31
		46.55	-60.87	29.93	Right middle temporal	39
541 (MCIAD3-VOI)	p < .05 FWE corr	29.81	-66.9	34.48	Right precuneus	7
		46.68	-65.42	18.69	Right middle temporal	39
		39.06	-70.67	34.28	Right precuneus	39
		31.69	-57.59	35.39	Right angular	39
Correlation with neurogranin levels (Ng-VOI)						
575	p < .05 FWE corr	-16.57	-63.8	42.99	Left precuneus	7
		-7.22	-63.16	36.01	Left precuneus	7
		-5.44	-54.55	44.06	Left precuneus	7
		-8.82	-57.68	18.48	Left posterior cingulate	23
Correlation with $\alpha$ -synuclein levels ( $\alpha$ syn-VOI)						
530	p < .05 FWE corr	-7	-57.86	20.3	Left posterior cingulate	31
		-16.55	-65.49	41.03	Left precuneus	7
		-7.17	-60.94	32.61	Left precuneus	7
		-25.76	-59.68	39.63	Left superior parietal lobule	7
Correlation with $\beta$ -synuclein levels ( $\beta$ syn-VOI)						
573	p < .05 FWE corr	-54.92	-52.49	5.58	Left superior temporal	22
		-53.02	-50.29	2.22	Left middle temporal	22
		-44.02	-61.4	19.34	Left middle temporal	39
Correlation with neurofilament-light chain levels (NFL-VOI)						
583	p < .05 FWE corr	43.18	-42	10.04	Right superior temporal	41
		65.53	-41.07	-0.31	Right middle temporal	21
		57.98	-43.94	10.1	Right superior temporal	22
		47.08	-36.89	-3.83	Right middle temporal	37
		58.07	-61.52	-2.37	Right middle temporal	37
		63.76	-45.95	-8.01	Right middle temporal	21
		65.65	-40.19	-9.23	Right middle temporal	21
		60.23	-40.98	-20.21	Right inferior temporal	20
		58.3	-28.8	-10.08	Right middle temporal	21
		30.36	-62.89	-6.57	Right fusiform	19

Peak coordinates and cortical regions in each cluster are ordered downwards from the highest peak Z-score. In all the analyses (height threshold: uncorrected  $p < 0.001$  at peak level); clusters were considered only if they contained at least 50 voxels and were statistically significant with  $p < 0.05$ , family-wise corrected (FWE corr) for multiple comparisons at cluster level.

Abbreviations: BA, Brodmann area; HC, healthy controls; MCI-AD, Mild cognitive impairment due to Alzheimer's disease; VOI, volumes of interest

### 3.2. Brain metabolic correlates of CSF biomarkers in MCI-AD patients

We found significant negative correlations between regional metabolic levels and CSF values of i) Ng and  $\alpha$ -syn in similar regions of the left PC/PCC (hereafter referred to as Ng-VOI and  $\alpha$ -syn-VOI), ii)  $\beta$ -syn in the left superior and middle temporal gyri ( $\beta$ -syn-VOI) and iii) NfL in the superior, middle, and inferior temporal gyri as well as the occipitotemporal fusiform gyrus of the right hemisphere (NfL-VOI) (Figure 2). Clusters' extent, coordinates and Brodmann areas of correlation are summarized in Table 2.



**Figure 2.** The significant VOIs related to neurogranin (Ng, in red),  $\alpha$ -synuclein ( $\alpha$ -syn, in yellow),  $\beta$ -synuclein ( $\beta$ -syn, in green), and neurofilament light chain (NfL, in light blue) are overlaid to the MCIAD clusters (already described in Figure 1, in purple). The 2D representation (axial, coronal, and sagittal cuts) and 3D rendering refer to the MNI atlas ((MRICroGL software, <https://www.nitrc.org/projects/mricrogl>)).

**Table 3. Spatial interaction among significant VOIs expressed as overlap coefficient (%)**

	MCIAD1-VOI	MCIAD2-VOI	MCIAD3-VOI	NfL-VOI	Ng-VOI	$\alpha$ syn-VOI
MCIAD1-VOI						
MCIAD2-VOI	0%					
MCIAD3-VOI	0%	0%				
NfL-VOI	0%	0%	0%			
Ng-VOI	0.3%	17.4%	0%	0%		
$\alpha$ syn-VOI	2.5%	54.5%	0%	0%	59.6%	
$\beta$ syn-VOI	40.3%	0%	0%	0%	0%	0%

Abbreviations: MCI-AD, Mild cognitive impairment due to Alzheimer's disease; VOIs (volumes of interest): MCIAD1-2-3, clusters (1-2-3) showing significant hypometabolism with respect to the healthy controls group; NfL, cluster significantly correlated with neurofilament light chain; Ng, cluster significantly correlated with neurogranin;  $\alpha$ -syn, cluster significantly correlated with alpha-synuclein;  $\beta$ -syn, cluster significantly correlated with beta-synuclein; WB, whole brain

The overlap coefficients among VOIs are reported in Table 3. Briefly, the 2.5% of the  $\alpha$ syn-VOI was included in the MCIAD1-VOI and the 54.5% in the MCIAD2-VOI. The 0.3% of the Ng-VOI was contained in the MCIAD1-VOI and the 17.4% in the MCIAD-2VOI. The  $\alpha$ syn- and Ng-VOIs overlapped one another for the 59.6%. The 40.3% of the  $\beta$ syn-VOI was included in the MCIAD1-VOI, whereas the NfL-VOI had no overlap with other regions.

### 3.3. Converted *versus* non-converted subgroup comparisons

MMSE at baseline and the whole-brain scaled average count densities of MCIAD2-VOI and MCIAD3-VOI were significantly lower in fast-converted than in non-converted patients, whereas those of the NFL-VOI only showed a trend for significance when correcting for multiple comparisons (see Table 1 and 4 for details).

**Table 4. Whole brain scaled average count densities of each of the significant VOI**

	MCI-AD (n=26)	Late/non-converted (n=15)	Fast-converted (n=11)	p-value <sup>#</sup>	q-value <sup>*</sup>
MCIAD1-VOI/WB	0.83±0.12	0.84±0.11	0.81±0.14	0.24	0.30
MCIAD2-VOI /WB	1.12±0.14	1.18±0.11	1.04±1.14	<b>0.01</b>	<b>0.047</b>
MCIAD3-VOI /WB	0.95±0.22	1.05±0.16	0.81±0.21	<b>0.003</b>	<b>0.019</b>
Ng-VOI /WB	0.95±0.16	0.97±0.14	0.92±0.19	0.44	0.46
$\alpha$ -syn-VOI /WB	0.12±0.06	1.07±0.14	1.02±0.16	0.24	0.30
$\beta$ -syn-VOI /WB	0.86±0.17	0.86±0.17	0.88±0.18	0.80	0.72
NfL-VOI WB	0.74±0.16	0.81±0.09	0.66±0.2	<b>0.03</b>	0.066

<sup>#</sup> p-values for Wilcoxon rank-sum test; <sup>\*</sup> FDR adjusted p-value. Significant values are in **bold**. Abbreviations: WB, whole brain, others as in Table 3.



#### 4. Discussion

This exploratory study revealed the brain metabolic correlates of some emerging CSF biomarkers in a group of MCI-AD patients through FDG-PET voxel-based analysis. The different brain areas related to these proteins suggest they express distinct pathophysiological processes occurring in prodromal AD.

First, the metabolism in regions included in the left PC/PCC negatively correlated with CSF Ng and  $\alpha$ -syn. Ng is a dendritic post-synaptic protein with a critical role in calmodulin-dependent memory consolidation and long-term potentiation (Hayashi 2009). Its changes in CSF closely reflect synaptic loss, cognitive decline, and neurodegeneration in AD (Portelius et al. 2015; Wellington et al. 2016; Tarawneh et al. 2016). CSF Ng values in our cohort were consistent with those previously reported with ELISA assays (Portelius et al. 2015; Sanfilippo et al. 2016; Wellington et al. 2016; Schipke et al. 2018; Willemse et al. 2018; Wang 2019), as a consequence of leakage or active secretion in the CSF after synaptic injury in AD-specific areas, such as that described in the hippocampus by pathological studies (Davidsson and Blennow 1998; Reddy et al. 2005). The CSF  $\alpha$ -syn level similarly relates to synaptic damage and cognitive impairment in AD (Twhig and Nielsen 2019). Our results closely overlap those by Chiasserini et al. (2017) and Majbour et al. (2017) using the same ELISA assay and fit with the hypothesis of a release of  $\alpha$ -syn from damaged neurons in prodromal AD (Twhig and Nielsen 2019). Notably, Ng is enriched primarily in the dendrites of entorhinal and hippocampal neurons (Represa et al. 1990; Guadaño-Ferraz et al. 2005) while  $\alpha$ -syn locates in the pre-synaptic terminals of the hippocampus, amygdala, and entorhinal cortex (Twhig and Nielsen 2019). The Ng- and  $\alpha$ syn-VOIs largely overlapped one another (59.6%), both similarly reflecting deafferentation of the PC/PCC regions from damaged medial temporal lobe (MTL) structures. In fact, altered metabolism of remote but connected posterior areas, such as PC/PCC, is typically described in AD as a consequence of deafferentation from injured hippocampus (Matsuda 2001). This is evidenced by the overlap of hypometabolic VOIs with the  $\alpha$ syn-VOI (2.5% with MCIAD1 and 54.5% with MCIAD2) and Ng-VOI (0.3% with MCIAD1 and 17.4% with MCIAD2). Our results complement the findings of Portelius et al. (2015), who described a relationship between CSF Ng levels and reduced brain glucose uptake. However, they used a global PET metabolic

score obtained by averaging values of several brain regions rather than voxel-wise correlation analysis, thus a detailed topographic comparison with our results is unfeasible.

$\beta$ -syn correlation sites were identified in the left superior and middle temporal areas. CSF  $\beta$ -syn values in our study closely overlapped with those described by Halbgebauer et al. (2021), who established the ELISA assay used in this study and confirmed the previous evidence on  $\beta$ -syn as a marker of pre-synaptic damage closely related to AD (Oeckl et al. 2016, 2020). Given the physiological expression of  $\beta$ -syn throughout cortical and subcortical brain areas (George 2001), the correlation with left lateral temporal cortex metabolism suggests an AD-specific process. This result is even more intriguing considering that a limited part of the  $\beta$ syn-VOI was encompassed in the hypometabolic MCIAD1-VOI. Hence, the larger part of  $\beta$ syn-VOI could express an early synaptic impairment but still not leading to overt hypometabolism in those regions. Notably, both the pre-synaptic proteins ( $\alpha$ -syn and  $\beta$ -syn) were more extensively related to hypometabolic regions than post-synaptic Ng. These findings confirm the critical involvement of the pre-synaptic compartment in the dynamic of [ $^{18}$ F]-FDG in the astrocyte-neuron functional unit (Lucas et al. 2018), but also that synaptic impairment precedes neuronal damage, as suggested by previous PET studies using synaptic-specific tracers (Chen et al. 2018; Bastin et al. 2020; Mecca et al. 2020).

The correlation with areas included in the left hemisphere may not be casual, as metabolic asymmetries are frequent in early AD. Several studies highlighted the greater susceptibility of the left hemisphere to metabolic dysfunction, vascular injury, and neurodegeneration in AD (Loewenstein et al. 1989; Thompson et al. 2003; Giannakopoulos et al. 2008) and, in particular, of the left temporal lobe to altered glucose utilization due to oxidative stress (Picco et al. 2014).

The correlation of synaptic proteins with dysmetabolism in temporal areas may be read in the light of the amyloid hypothesis postulating that amyloid beta triggers a cascade harming synaptic functioning and ultimately neurons through the formation of paired helical filaments of Tau aggregates. In particular, the involvement of the lateral temporal region could reflect the progression of Alzheimer's pathology from medial to lateral superficial temporal areas (i.e., from archil- to iso- and neocortex), as

derived from neuropathological (Braak and Braak 1991; Braak et al. 2006) and Tau-PET studies (Chiotis et al. 2018).

In line with this pattern of disease progression, CSF NfL correlated negatively with lateral temporal metabolism and, albeit marginally, with the fusiform gyrus in the right hemisphere. Noteworthy, the NfL-VOI was utterly independent of the hypometabolic clusters. We can only speculate about the meaning of such intriguing finding, which might reflect an early pathological process where axonal damage - paralleled by the CSF NfL level – even precedes synaptic loss and neurodegeneration - and thus regional hypometabolism - in the less affected hemisphere.

In the same vein, previous work by our group showed that the metabolic changes of right prefrontal areas correlated with specific prospective memory tasks in MCI-AD patients, and early network disruption was significant, primarily in the interhemispheric connection (Massa et al. 2020). To note, those prefrontal regions were not hypometabolic compared to controls, like the NfL-VOI in the present study. This reinforces that progressive network disruption is critical in AD (Delbeuck et al. 2003; Agosta et al. 2011; Pagani et al. 2017) and parallels subtle metabolic changes that, at least in the earliest stages, do not result in overt hypometabolism. In agreement with the dying back hypothesis (Kanaan et al. 2013), the significant impairment in axonal connectivity precedes neurodegeneration and consequent leakage of neuronal (e.g. synaptic) proteins in CSF, hence possibly explaining why synaptic markers correlated with more damaged, dysmetabolic brain areas.

Interestingly, the whole-brain scaled mean count density of NfL-VOI was lower in fast- than non-converted patients, although such finding only showed a trend to significance ( $q=0.066$ ) when correcting for multiple comparisons. Even if we need caution when interpreting such rather spurious result, it is concordant with a relative sparing of right hemisphere metabolism in our cohort to be associated with slower progression to dementia, as MCIAD3-VOI in the right hemisphere was significantly preserved in late converters ( $q=0.019$ ). This is consistent with previous evidence of relatively preserved metabolism in regions within the right hemisphere (temporal areas and fusiform gyrus), acting as sites of resilience and directly associating with conversion time to dementia (Morbelli et al. 2017; Bauckneht et al. 2018).

In summary, we may put forward some hypotheses drawn by the correlations where i) Ng and  $\alpha$ -syn reflected established hippocampal damage referring to largely deafferented and thus dysmetabolic posterior areas (PCC/PC); ii)  $\beta$ -syn points to the progressive synaptic and metabolic failure in left lateral temporal areas; iii) NfL relates to still relatively preserved temporal areas of the less affected (right) hemisphere, whose progressive deterioration parallels progression to dementia.

We acknowledge some limitations of this study. The relatively small sample size was due to the concomitant need to have two second-level biomarkers available (e.g., CSF and FDG-PET) and to the assessment of non-routine CSF proteins. However, the careful selection of patients with a high-likelihood AD diagnosis (Albert et al. 2011b) partially compensates for this limitation. Most patients were part of non-pharmacological clinical trials, so an atypical course or a more complex diagnosis were not the reasons they underwent both FDG-PET and CSF analysis. Furthermore, we adjusted all our analyses for age to limit its influence on CSF biomarker level, primarily NfL (Bridel et al. 2019). We included healthy subjects only to compare their brain metabolism with that of MCI-AD patients but not to assess the metabolic correlates of CSF biomarkers. The use of both biomarkers (i.e., CSF and FDG-PET) is rare in the healthy, even when included in research protocols. Without a control group for CSF biomarkers, we referenced published works that used the same assays and found similar levels in CSF. Moreover, the relatively small sample of patients led us to use a univariate analysis - where the different proteins were independently correlated with brain metabolism - rather than a multivariate one that might more appropriately estimate the relative contribution of each protein to the total variance. For the same reason, the correction for multiple comparisons limited the significance of the NfL-VOI comparison between converted patients and those not converted yet, thus suggesting that the prognostic value of NfL metabolic correlates should be cautiously interpreted and additional data are needed.

Given such limitations, our preliminary findings should be considered explorative for the MCI-AD condition with a typical amnesic presentation and need to be replicated in larger samples with a prospective design and more conservative statistical thresholds.

In conclusion, by providing evidence for the metabolic correlates of several process-specific CSF biomarkers, the present results shed light on their pathophysiological role in prodromal AD. These findings complement the valuable information on the distribution of hypometabolism related to neuronal loss, which differs from the metabolic changes reflecting synaptic loss or axonal damage. Further studies including also pre-symptomatic and demented subjects are worthwhile to assess the dynamic changes of CSF proteins and brain metabolism across different AD stages. This would give insight into the pathological processes occurring at different times in distinct brain regions and the resilience mechanisms that counteract them.

## 5. Discussion and concluding remarks

Revealing the complex interactions and assessing potential integration between biomarkers is essential, especially in the early stages of AD, when biomarker alterations may serve to stage patients throughout the disease spectrum, improve phenotyping, and indicate the likelihood of progression to dementia.

In this research, the integration of [ $^{18}\text{F}$ ]-FDG-PET and CSF biomarkers, two of the most used biomarkers in centers focused on neurocognitive disorders, enabled us to collect evidence on their analytical and diagnostic performance when used in a step-wise fashion. As part of the ongoing endeavor to create a common diagnostic chart for the precise and cost-effective use of biomarkers in neurocognitive diseases with neurodegenerative origin (Festari et al. 2022), these data gain further significance.

Additionally, by combining semiquantitative [ $^{18}\text{F}$ ]-FDG-PET and CSF data, we were able to identify precise topographic correlations between metabolic values and CSF proteins that indicated distinct underlying disease processes. These findings add to the knowledge regarding the distribution of hypometabolism linked to neuronal loss, which is distinct from metabolic changes reflecting synaptic or axonal injury, and provide an indirect insight of the pathological processes taking place at various times in different parts of the brain.

These results will be expanded into bigger cohorts in future research, which will also integrate additional newly discovered synaptopathy-expressing proteins for diagnostic and prognostic purposes.

## 6. Appendix

### 6.1. Publications during PhD

1. Benedetti L, Garnero M, Demichelis C, Grandis M, Briani C, Beltramini S, Bellucci M, Prada V, **Massa F**, Gastaldi M, Schenone A, Franciotta D. Outcomes after single-cycle rituximab monotherapy in patients with anti-MAG polyneuropathy: A bi-center experience with an average follow-up of 11 years. *J Neuroimmunol*. 2019 Oct 15;337:577081
2. Prada V, **Massa F**, Salerno A, Fregosi D, Beronio A, Serrati C, Mannironi A, Mancardi G, Schenone A, Benedetti L. Importance of intensive and prolonged rehabilitative treatment on the Guillain-Barré syndrome long-term outcome: a retrospective study. *Neurol Sci*. 2020 Feb;41(2):321-327
3. **Massa F**, Farotti L, Eusebi P, Capello E, Dottorini ME, Tranfaglia C, Bauckneht M, Morbelli S, Nobili F, Parnetti L. Reciprocal incremental value of 18F-FDG-PET and cerebrospinal fluid biomarkers in Mild Cognitive Impairment patients suspected for Alzheimer's disease and inconclusive first biomarker. *J Alzheimers Dis*. 2019;72(4):1193–1207
4. Gastaldi M, Mariotto S, Giannoccaro MP, Iorio R, Zoccarato M, Nosadini M, Benedetti L, Casagrande S, Di Filippo M, Valeriani M, Ricci S, Bova S, Arbasino C, Mauri M, Versino M, Vigeveno F, Papetti L, Romoli M, Lapucci C, **Massa F**, Sartori S, Zuliani L, Barilaro A, De Gaspari P, Spagni G, Evoli A, Liguori R, Ferrari S, Marchioni E, Giometto B, Massacesi L, Franciotta D. Subgroup comparison according to clinical phenotype and serostatus in autoimmune encephalitis: a multicenter retrospective study. *Eur J Neurol*. 2020; 27:633-643
5. Orso B, Mattei C, Arnaldi D, **Massa F**, Serafini G, Plantone D, Doglione E, Grafman J, Nobili F, Pardini M. Clinical and MRI predictors of conversion from Mild Behavioural Impairment to dementia. *Am J Geriatr Psych*. 2020;28:755–76
6. Arnaldi D, Donniaquio A, Mattioli P, **Massa F**, Grazzini M, Meli R, Filippi L, Grisanti S, Famà F, Terzaghi M, Girtler N, Brugnolo A, Doglione E, Pardini M, Villani F, Nobili F. Epilepsy in Neurodegenerative Dementias: A Clinical, Epidemiological, and EEG Study. *J Alzheimers Dis*. 2020;74(3):865-874

7. Arnaldi D, Chincari A, De Carli F, Famà F, Girtler N, Brugnolo A, Pardini M, **Massa F**, Meli R, Schenone C, Bauckneht M, Morbelli S, Nobili F. The fate of patients with REM sleep behavior disorder and Mild Cognitive Impairment. *Sleep Medicine*, 2021;79:205-210
8. **Massa F**, Zuppa A, Pesce G, Demichelis C, Bergamaschi M, Garnero M, Briani C, Ferrari S, Schenone A, Benedetti L. Bendamustine–rituximab (BR) combined therapy for treatment of immuno-mediated neuropathies associated with hematologic malignancy. *J Neurol Sci*. 2020 Jun 15;413:116777
9. Rosa G, Giannotti C, Martella L, **Massa F**, Serafini G, Pardini M, Nobili F, Monacelli F & Disease Management Team on Dementia of the IRCCS Ospedale Policlinico San Martino (Genoa, I). Brain Aging, Cardiovascular Diseases, Mixed Dementia, and Frailty in the Oldest Old: From Brain Phenotype to Clinical Expression. *J Alzheimers Dis*. 2020;75(4):1083-1103
10. **Massa F**, Meli R, Grazzini M, Famà F, De Carli F, Filippi L, Arnaldi A, Pardini M, Morbelli S, Nobili F. Utility of quantitative EEG in early Lewy body disease, *Park Rel Dis*. 2020;75:70-75
11. Orso B, Arnaldi D, Famà F, Girtler N, Brugnolo A, Doglione E, Filippi L, **Massa F**, Peira E, Bauckneht M, Morbelli S, Nobili F, Pardini M, Anatomical and neurochemical bases of theory of mind in de novo Parkinson's Disease, *Cortex*, 2020 Sep;130:401-412
12. **Massa F**, Grisanti S, Brugnolo A, Doglione E, Orso B, Morbelli S, Bauckneht M, Origone P, Filippi L, Arnaldi D, De Carli F, Pardini M, Pagani M, Nobili F, Girtler N. The role of anterior prefrontal cortex in prospective memory: an exploratory FDG-PET study in early Alzheimer's disease. *Neurobiol Aging*, 2020; 96: 117-127
13. Morbelli S, Arnaldi D, Cella E, Raffa S, Donegani MI, Capitanio S, **Massa F**, Miceli A, Filippi L, Chincari A, Nobili F. Striatal dopamine transporter SPECT quantification: head-to-head comparison between two three-dimensional automatic tools. *EJNMMI Res*. 2020 Nov 7;10(1):137
14. Arnaldi D, Famà F, Girtler N, Brugnolo A, Pardini M, Mattioli P, Meli R, **Massa F**, Orso B, Sormani MP, Donegani MI, Bauckneht M, Morbelli S, Nobili F. REMeDio: a proof-of-concept neuroprotection study for prodromal synucleinopathies. *Eur J Neurol*. 2021 Apr;28(4):1210-1217



15. Brugnolo A, Girtler N, Doglione E, Orso B, **Massa F**, Donegani MI, Bauckneht M, Morbelli S, Arnaldi D, Nobili F, Pardini M. Brain Resources: How Semantic Cueing Works in Mild Cognitive Impairment due to Alzheimer's Disease (MCI-AD). *Diagnostics*. 2021; 11(1):108
16. Mattioli P, Pardini M, Famà F, Girtler N, Brugnolo A, Orso B, Meli R, Filippi L, Grisanti S, **Massa F**, Bauckneht M, Miceli A, Terzaghi M, Morbelli S, Nobili F, Arnaldi D. Cuneus/precuneus as a central hub for brain functional connectivity of mild cognitive impairment in idiopathic REM sleep behavior patients. *Eur J Nucl Med Mol Imaging*. 2021 Aug;48(9):2834-2845
17. **Massa F**, Filippi L, Benedetti L, Morbelli S, Nobili F. FDG PET Unveils the Course of Paraneoplastic Cerebellar Degeneration: A Semiquantitative Analysis. *Clin Nucl Med*. 2021 Jun 1;46(6):e327-e328
18. Donegani MI, Miceli A, Pardini M, Bauckneht M, Chiola S, Pennone M, Marini C, **Massa F**, Raffa S, Ferrarazzo G, Arnaldi D, Sambuceti G, Nobili F, Morbelli S. Brain Metabolic Correlates of Persistent Olfactory Dysfunction after SARS-Cov2 Infection. *Biomedicines*. 2021 Mar 12;9(3):287
19. Orso B, Arnaldi D, Girtler N, Brugnolo A, Doglione E, Mattioli P, Biassoni E, Fancellu R, **Massa F**, Bauckneht M, Chiola S, Morbelli S, Nobili F, Pardini M. Dopaminergic and Serotonergic Degeneration and Cortical [<sup>18</sup>F]Fluorodeoxyglucose Positron Emission Tomography in De Novo Parkinson's Disease. *Mov Disord*. 2021 May 22; 36: 2293-2302
20. Grisanti SG, Franciotta D, Garnero M, Zuppa A, **Massa F**, Mobilia EM, Pesce G, Schenone A, Benedetti L. A case series of parainfectious Guillain-Barré syndrome linked to influenza A (H1N1) virus infection. *J Neuroimmunology*. 2021 Aug 15;357:577605
21. Demichelis C, Grisanti S, **Massa F** ✉, Serrati C, Beronio A, Mannironi A, Mobilia E, Briani C, Schenone A, Benedetti L. Abnormal sweating and "skin flushing" as possible predictive factor for treatment related fluctuations in Guillain-Barré syndrome: a case series and a review of the literature. *J Neurol Sci*. 2021 Sep 15;428:117589.
22. Gómez de San José N, **Massa F**, Halbgebauer S, Oeckl P, Steinacker P, Otto M. Neuronal pentraxins as biomarkers of synaptic activity: from physiological functions to pathological changes in neurodegeneration. *J. Neural Transm*. 2022 Feb;129(2):207-230.

23. Arnaldi D, Mattioli P, Famà F, Girtler N, Brugnolo A, Pardini M, Donniaquio A, **Massa F**, Orso B, Raffa S, Bauckneht M, Morbelli S, Nobili F. Stratification Tools for Disease-Modifying Trials in Prodromal Synucleinopathy. *Mov Disord* 2022 Jan;37(1):52-61
24. Orso B, Famà F, Giorgetti L, Mattioli P, Donniaquio A, Girtler N, Brugnolo A, **Massa F**, Peira E, Pardini M, Morbelli S, Nobili F, Arnaldi D. Polysomnographic correlates of sleep disturbances in de novo, drug naïve Parkinson's Disease. *Neurol Sci.* 2022 Apr;43(4):2531-2536
25. **Massa F**, Chincarini A, Bauckneht M, Raffa S, Peira E, Arnaldi A, Pardini M, Pagani M, Orso B, Donegani MI, Brugnolo A, Biassoni E, Mattioli P, Girtler N, Guerra UP, Morbelli S, Nobili F. Added value of semi-quantitative analysis of brain FDG-PET for the differentiation between MCI-Lewy bodies and MCI due to Alzheimer's disease. *Eur J Nucl Med Mol Imaging.* 2022; 49:1263–1274
26. Orso B, Lorenzini L, Arnaldi D, Girtler N, Brugnolo A, Doglione E, Mattioli P, Biassoni E, **Massa F**, Peira E, Bauckneht M, Donegani MI, Morbelli S, Nobili F, Pardini M. The Role of Hub and Spoke Regions in Theory of Mind in Early Alzheimer's Disease and Frontotemporal Dementia. *Biomedicines.* 2022; 10(3):544
27. Girtler N, Chincarini A, Brugnolo A, Doglione E, Orso B, Morbelli S, **Massa F**, Peira E, Biassoni E, Donniaquio A, Grisanti S, Pardini M, Arnaldi D, Nobili F. The Free and Cued Selective Reminding Test: Discriminative Values in a Naturalistic Cohort. *J Alzheimers Dis.* 2022;87(2):887-899
28. Grisanti SG, Garbarino S, Barisione E, Aloè T, Grosso M, Schenone C, Pardini M, Biassoni E, Zaottini F, Picasso R, Morbelli S, Campi C, Pesce G, **Massa F**, Girtler N, Battaglini D, Cabona C, Bassetti M, Uccelli A, Schenone A, Piana M, Benedetti L. Neurological long-COVID in the outpatient clinic: Two subtypes, two courses. *J Neurol Sci.* 2022 Aug 15;439:120315
29. **Massa F**, Halbgebauer S, Barba L, Oeckl P, Gómez de San José N, Bauckneht M, Lanfranchi F, Vigo T, Arnaldi D, Pardini M, Morbelli S, Chincarini A, Barthel H, Otto M, Nobili F. Exploring the brain metabolic correlates of process-specific CSF biomarkers in patients with MCI due to Alzheimer's disease: preliminary data. *Neurobiol Aging.* 2022 Sep;117:212-221

30. Barba L, Halbgebauer S, Paolini Paoletti F, Bellomo G, Abu-Rumeileh S, Steinacker P, **Massa F**, Parnetti L, Otto M. Specific Cerebrospinal Fluid SerpinA1 Isoform Pattern in Alzheimer's Disease. *Int J Mol Sci.* 2022; 23(13):6922
31. Bellucci M, Germano F, Grisanti S, Castellano C, Tazza F, Mobilia EM, Visigalli D, Novi G, **Massa F**, Rossi S, Durando P, Cabona C, Schenone A, Franciotta D, Benedetti L. Case Report: Post-COVID-19 Vaccine Recurrence of Guillain-Barré Syndrome (GBS) Following an Antecedent Para-infectious COVID-19-related GBS. *Front Immunol.* 2022 Jul 18;13:894872
32. **Massa F**, Franciotta D, Grisanti S, Roccatagliata L, Morbelli S, Beltramini S, Uccelli A, Schenone A, Benedetti L. Intravenous immunoglobulin bridging to rituximab in NMDAR encephalitis patients non-responders to first-line treatments. *Neurol Sci.* 2022 **43**, 6441–6447
33. Barba L, Abu Rumeileh S, Bellomo G, Paolini Paoletti F, Halbgebauer S, Oeckl P, Steinacker P, **Massa F**, Gaetani L, Parnetti L, Otto M. Cerebrospinal fluid  $\beta$ -synuclein as a synaptic biomarker for preclinical Alzheimer's disease. *J Neurol Neurosurg Psychiatry.* 2023 Jan;94(1):83-86
34. Orso B, Arnaldi D, Peira E, Famá F, Giorgetti L, Girtler N, Brugnolo A, Mattioli P, Biassoni E, Donniaquio A, **Massa F**, Bauckneht M, Miceli A, Morbelli S, Nobili F, Pardini M. The Role of Monoaminergic Tones and Brain Metabolism in Cognition in de novo Parkinson's Disease. *J Parkins Dis.* 2022;12(6):1945-1955
35. Matricardi S, Casciato S, Bozzetti S, Mariotto S, Stabile A, Freri E, Deleo F, Sartori S, Nosadini M, Pappalardo I, Meletti S, Giovannini G, Zucchi E, Di Bonaventura C, Di Gennaro G, Ferrari S, Zuliani L, Zoccarato M, Vogrig A, Lattanzi S, Michelucci R, Gambardella A, Ferlazzo E, Fusco L, Granata T, Villani F; **Immune Epilepsies Study Group of the Italian League Against Epilepsy\***. Epileptic phenotypes in autoimmune encephalitis: from acute symptomatic seizures to autoimmune-associated epilepsy. *J Neurol Neurosurg Psychiatry.* 2022 Jul 25:jnnp-2022-329195. \* as member of the study group
36. Mattioli P, Pardini M, Girtler N, Brugnolo A, Orso B, Donniaquio A, Calizzano F, Mancini R, **Massa F**, Terzaghi M, Bauckneht M, Morbelli S, Sambuceti G, Nobili F, Arnaldi D. Cognitive and brain metabolism profiles of mild cognitive impairment in prodromal synucleinopathy. *J Alzheimers Dis.* 2022;90(1):433-444

37. Festari C, **Massa F**, Cotta Ramusino M, Gandolfo F, Nicolosi V, S Orini, Aarsland D, Agosta F, Babiloni C, Boada M, Borroni B, Cappa S, Dubois B, Frederiksen KS, Frölich L, Garibotto V, Georges J, Haliassos A, Hansson O, Jessen F, Kamondi A, Kessels RPC, Morbelli S, O'Brien JT, Otto M, Perret-Liaudet A, Pizzini FB, Ritchie CW, Scheltens P, van der Flier WM, Vandenbulcke M, Vanninen R, Verhey FRJ, Vernooij MW, Yousry T, Nobili F, Frisoni GB. European consensus for the diagnosis of MCI and mild dementia: preparatory phase. *Alzheimers dement.* 2022 Oct 9.doi:10.1002/alz.12798.
38. Ruscitti F, Origone P, Rosti G, Trevisan L, Marchese R, Brugnolo A, **Massa F**, Castellini P, Mandich P. A case of Huntington disease-like 2 in a patient of African ancestry: the everlasting support of clinical examination in the molecular era. *Clin case rep.* 2022; 10:e06308
39. Mattioli P, Orso B, Liguori C, Famà F, Giorgetti L, Donniaquio A, **Massa F**, Giberti A, Vállez García D, Meles SK, Leenders KL, Placidi F, Spanetta M, Chiaravallotti A, Camedda R, Schillaci O, Izzi F, Mercuri NB, Pardini M, Bauckneht M, Morbelli S, Nobili F, Arnaldi D. Derivation and Validation of a Phenoconversion-Related Pattern in Idiopathic Rapid Eye Movement Behavior Disorder. *Mov Disord.* 2022 Oct 3. doi: 10.1002/mds.29236 (IF=9.698)
40. Ferraro PM, Gualco L, Costagli M, Schiavi S, Ponzano M, Signori A, Massa F, Pardini M, Castellan L, Levrero F, Zacà D, Piredda GF, Hilbert T, Kober T, Roccatagliata L. Compressed sensing (CS) MP2RAGE versus standard MPRAGE: A comparison of derived brain volume measurements. *Phys Med.* 2022 Nov 8;103:166-174
41. Lanfranchi F, Arnaldi D, Miceli A, Mattioli P, D'Amico F, Raffa S, Donegani MI, Chiola S, Massa F, Pardini M, Di Raimondo T, Sambuceti G, Bauckneht M, Nobili F, Morbelli S. Different z-score cut-offs for Striatal Binding Ratio (SBR) of DaT SPECT are needed to support the diagnosis of Parkinson's Disease (PD) and Dementia with Lewy Bodies (DLB). *EJNMMI.* 2022 Dec 6. doi: 10.1007/s00259-022-06069-0.

## 6.2. References (in alphabetical order)

- Agosta F, Pievani M, Sala S, et al (2011) White Matter Damage in Alzheimer Disease and Its Relationship to Gray Matter Atrophy. *Radiology* 258:853–863. <https://doi.org/10.1148/radiol.10101284>
- Albert MS, DeKosky ST, Dickson D, et al (2011a) The diagnosis of mild cognitive impairment due to Alzheimer's disease: Recommendations from the National Institute on Aging-Alzheimer's Association workgroups on diagnostic guidelines for Alzheimer's disease. *Alzheimer's Dement* 7:270–279. <https://doi.org/10.1016/j.jalz.2011.03.008>
- Albert MS, DeKosky ST, Dickson D, et al (2011b) The diagnosis of mild cognitive impairment due to Alzheimer's disease: Recommendations from the National Institute on Aging-Alzheimer's Association workgroups on diagnostic guidelines for Alzheimer's disease. *Alzheimer's Dement* 7:270–279. <https://doi.org/10.1016/j.jalz.2011.03.008>
- Alexopoulos P, Werle L, Roesler J, et al (2016) Conflicting cerebrospinal fluid biomarkers and progression to dementia due to Alzheimer's disease. *Alzheimers Res Ther* 8:51. <https://doi.org/10.1186/s13195-016-0220-z>
- Arbizu J, Festari C, Altomare D, et al (2018) Clinical utility of FDG-PET for the clinical diagnosis in MCI. *Eur J Nucl Med Mol Imaging* 45:1497–1508. <https://doi.org/10.1007/s00259-018-4039-7>
- Arendt T (2009) Synaptic degeneration in Alzheimer's disease. *Acta Neuropathol* 118:167–179. <https://doi.org/10.1007/s00401-009-0536-x>
- Barba L, Abu Rumeileh S, Bellomo G, et al (2023) Cerebrospinal fluid  $\beta$ -synuclein as a synaptic biomarker for preclinical Alzheimer's disease. *J Neurol Neurosurg & Psychiatry* 94:83 LP – 86. <https://doi.org/10.1136/jnnp-2022-329124>
- Barba L, Paolini Paoletti F, Bellomo G, et al (2022) Alpha and Beta Synucleins: From Pathophysiology to Clinical Application as Biomarkers. *Mov Disord* 37:669–683. <https://doi.org/10.1002/mds.28941>
- Bastin C, Bahri MA, Meyer F, et al (2020) In vivo imaging of synaptic loss in Alzheimer's disease with [18F]UCB-H positron emission tomography. *Eur J Nucl Med Mol Imaging* 47:390–402. <https://doi.org/10.1007/s00259-019-04461-x>
- Bauckneht M, Chincarini A, Piva R, et al (2018) Metabolic correlates of reserve and resilience in MCI due to Alzheimer's Disease (AD) Rik Ossenkoppele. *Alzheimer's Res Ther* 10:. <https://doi.org/10.1186/s13195-018-0366-y>
- Beach TG, Monsell SE, Phillips LE, Kukull W (2012) Accuracy of the Clinical Diagnosis of

- Alzheimer Disease at National Institute on Aging Alzheimer Disease Centers, 2005–2010. *J Neuropathol Exp Neurol* 71:266–273. <https://doi.org/10.1097/NEN.0b013e31824b211b>
- Ben Bouallègue F, Mariano-Goulart D, Payoux P, Alzheimer's Disease Neuroimaging Initiative (ADNI) the ADNI (2017) Comparison of CSF markers and semi-quantitative amyloid PET in Alzheimer's disease diagnosis and in cognitive impairment prognosis using the ADNI-2 database. *Alzheimers Res Ther* 9:32. <https://doi.org/10.1186/s13195-017-0260-z>
- Berchtold NC, Coleman PD, Cribbs DH, et al (2013) Synaptic genes are extensively downregulated across multiple brain regions in normal human aging and Alzheimer's disease. *Neurobiol Aging* 34:1653–1661. <https://doi.org/10.1016/j.neurobiolaging.2012.11.024>
- Berge G, Sando SB, Albrektsen G, et al (2016) Alpha-synuclein measured in cerebrospinal fluid from patients with Alzheimer's disease, mild cognitive impairment, or healthy controls: a two year follow-up study. *BMC Neurol* 16:180. <https://doi.org/10.1186/s12883-016-0706-0>
- Berk C, Paul G, Sabbagh M (2014) Investigational drugs in Alzheimer's disease: current progress. *Expert Opin Investig Drugs* 23:837–846. <https://doi.org/10.1517/13543784.2014.905542>
- Biscetti L, Salvadori N, Farotti L, et al (2019) The added value of A $\beta$ 42/A $\beta$ 40 in the CSF signature for routine diagnostics of Alzheimer's disease. *Clin Chim Acta* 494:71–73. <https://doi.org/10.1016/j.cca.2019.03.001>
- Bittner T, Zetterberg H, Teunissen CE, et al (2016) Technical performance of a novel, fully automated electrochemiluminescence immunoassay for the quantitation of  $\beta$ -amyloid (1–42) in human cerebrospinal fluid. *Alzheimer's Dement* 12:517–526. <https://doi.org/https://doi.org/10.1016/j.jalz.2015.09.009>
- Blennow K, Bogdanovic N, Alafuzoff I, et al (1996) Synaptic pathology in Alzheimer's disease: Relation to severity of dementia, but not to senile plaques, neurofibrillary tangles, or the ApoE4 allele. *J Neural Transm* 103:603–618. <https://doi.org/10.1007/BF01273157>
- Blennow K, Zetterberg H (2018) The Past and the Future of Alzheimer's Disease Fluid Biomarkers. *J Alzheimer's Dis* 62:1125–1140. <https://doi.org/10.3233/JAD-170773>
- Boccardi M, Nicolosi V, Festari C, et al (2019) Italian consensus recommendations for the biomarker-based etiological diagnosis in MCI patients. *Eur J Neurol* under review
- Bohnen NI, Djang DSW, Herholz K, et al (2012) Effectiveness and safety of 18F-FDG PET

- in the evaluation of dementia: a review of the recent literature. *J Nucl Med* 53:59–71. <https://doi.org/10.2967/jnumed.111.096578>
- Bouwman F, Orini S, Gandolfo F, et al (2018) Diagnostic utility of FDG-PET in the differential diagnosis between different forms of primary progressive aphasia. *Eur J Nucl Med Mol Imaging* 45:1526–1533. <https://doi.org/10.1007/s00259-018-4034-z>
- Bouwman FH, Schoonenboom SNM, van der Flier WM, et al (2007) CSF biomarkers and medial temporal lobe atrophy predict dementia in mild cognitive impairment. *Neurobiol Aging* 28:1070–1074. <https://doi.org/10.1016/j.neurobiolaging.2006.05.006>
- Braak H, Alafuzoff I, Arzberger T, et al (2006) Staging of Alzheimer disease-associated neurofibrillary pathology using paraffin sections and immunocytochemistry. *Acta Neuropathol* 112:389–404. <https://doi.org/10.1007/s00401-006-0127-z>
- Braak H, Braak E (1991) Neuropathological staging of Alzheimer-related changes. *Acta Neuropathol* 82:239–259. <https://doi.org/10.1007/BF00308809>
- Bridel C, van Wieringen WN, Zetterberg H, et al (2019) Diagnostic Value of Cerebrospinal Fluid Neurofilament Light Protein in Neurology: A Systematic Review and Meta-analysis. *JAMA Neurol* 76:1035–1048. <https://doi.org/10.1001/jamaneurol.2019.1534>
- Brown RKJ, Bohnen NI, Wong KK, et al (2014) Brain PET in Suspected Dementia: Patterns of Altered FDG Metabolism. *RadioGraphics* 34:684–701. <https://doi.org/10.1148/rg.343135065>
- Bruscoli M, Lovestone S (2004) Is MCI really just early dementia? A systematic review of conversion studies. *Int Psychogeriatrics* 16:129–140. <https://doi.org/DOI:10.1017/S1041610204000092>
- Caminiti SP, Ballarini T, Sala A, et al (2018) FDG-PET and CSF biomarker accuracy in prediction of conversion to different dementias in a large multicentre MCI cohort. *NeuroImage Clin* 18:167–177. <https://doi.org/10.1016/j.nicl.2018.01.019>
- Camporesi E, Nilsson J, Brinkmalm A, et al (2020) Fluid Biomarkers for Synaptic Dysfunction and Loss. *Biomark Insights* 15:1177271920950319. <https://doi.org/10.1177/1177271920950319>
- Carotenuto A, Rea R, Colucci L, et al (2012) Late and early onset dementia: What is the role of vascular factors? A retrospective study. *J Neurol Sci* 322:170–175. <https://doi.org/10.1016/j.jns.2012.07.066>
- Chen MK, Mecca AP, Naganawa M, et al (2018) Assessing Synaptic Density in Alzheimer Disease with Synaptic Vesicle Glycoprotein 2A Positron Emission Tomographic Imaging. *JAMA Neurol* 75:1215–1224. <https://doi.org/10.1001/jamaneurol.2018.1836>
- Chételat G, Arbizu J, Barthel H, et al (2019) Molecular neuroimaging in dementia clinical

- diagnosis and research: where are we headed? *Lancet Neurol* under review
- Chiasserini D, Biscetti L, Eusebi P, et al (2017) Differential role of CSF fatty acid binding protein 3,  $\alpha$ -synuclein, and Alzheimer's disease core biomarkers in Lewy body disorders and Alzheimer's dementia. <https://doi.org/10.1186/s13195-017-0276-4>
- Chiotis K, Saint-Aubert L, Rodriguez-Vieitez E, et al (2018) Longitudinal changes of tau PET imaging in relation to hypometabolism in prodromal and Alzheimer's disease dementia. *Mol Psychiatry* 23:1666–1673. <https://doi.org/10.1038/mp.2017.108>
- Choo IH, Ni R, Schöll M, et al (2013) Combination of 18F-FDG PET and Cerebrospinal Fluid Biomarkers as a Better Predictor of the Progression to Alzheimer's Disease in Mild Cognitive Impairment Patients. *J Alzheimer's Dis* 33:929–939. <https://doi.org/10.3233/JAD-2012-121489>
- Cure S, Abrams K, Belger M, et al (2014) Systematic Literature Review and Meta-Analysis of Diagnostic Test Accuracy in Alzheimer's Disease and Other Dementia Using Autopsy as Standard of Truth. *J Alzheimer's Dis* 42:169–182. <https://doi.org/10.3233/JAD-131559>
- Davidsson P, Blennow K (1998) Neurochemical dissection of synaptic pathology in Alzheimer's disease. *Int psychogeriatrics* 10:11–23. <https://doi.org/10.1017/S1041610298005110>
- Delbeuck X, Van Der Linden M, Collette F (2003) Alzheimer's Disease as a Disconnection Syndrome? *Neuropsychol. Rev.* 13:79–92
- Della Rosa PA, Cerami C, Gallivanone F, et al (2014) A Standardized [18F]-FDG-PET Template for Spatial Normalization in Statistical Parametric Mapping of Dementia. *Neuroinformatics* 12:575–593. <https://doi.org/10.1007/s12021-014-9235-4>
- Dhiman K, Gupta VB, Villemagne VL, et al (2020) Cerebrospinal fluid neurofilament light concentration predicts brain atrophy and cognition in Alzheimer's disease. *Alzheimer's Dement Diagnosis, Assess Dis Monit* 12:e12005. <https://doi.org/https://doi.org/10.1002/dad2.12005>
- Dorey A, Perret-Liaudet A, Tholance Y, et al (2015) Cerebrospinal Fluid A $\beta$ 40 Improves the Interpretation of A $\beta$ 42 Concentration for Diagnosing Alzheimer's Disease. *Front Neurol* 6:. <https://doi.org/10.3389/fneur.2015.00247>
- Dubois B, Feldman HH, Jacova C, et al (2014a) Advancing research diagnostic criteria for Alzheimer's disease: the IWG-2 criteria. *Lancet Neurol* 13:614–629. [https://doi.org/10.1016/S1474-4422\(14\)70090-0](https://doi.org/10.1016/S1474-4422(14)70090-0)
- Dubois B, Feldman HH, Jacova C, et al (2014b) Advancing research diagnostic criteria for Alzheimer's disease: the IWG-2 criteria. *Lancet Neurol* 13:614–629.



[https://doi.org/10.1016/S1474-4422\(14\)70090-0](https://doi.org/10.1016/S1474-4422(14)70090-0)

- Eickhoff SB, Laird AR, Grefkes C, et al (2009) Coordinate-based activation likelihood estimation meta-analysis of neuroimaging data: a random-effects approach based on empirical estimates of spatial uncertainty. *Hum Brain Mapp* 30:2907–26. <https://doi.org/10.1002/hbm.20718>
- Fellgiebel A, Scheurich A, Bartenstein P, Müller MJ (2007) FDG-PET and CSF phospho-tau for prediction of cognitive decline in mild cognitive impairment. *Psychiatry Res Neuroimaging* 155:167–171. <https://doi.org/10.1016/j.psychresns.2006.12.002>
- Festari C, Massa F, Cotta Ramusino M, et al (2022) European consensus for the diagnosis of MCI and mild dementia: Preparatory phase. *Alzheimer's Dement* n/a: <https://doi.org/https://doi.org/10.1002/alz.12798>
- Fiandaca MS, Mapstone ME, Cheema AK, Federoff HJ (2014) The critical need for defining preclinical biomarkers in Alzheimer's disease. *Alzheimer's Dement* 10:S196–S212. <https://doi.org/10.1016/j.jalz.2014.04.015>
- Fourier A, Portelius E, Zetterberg H, et al (2015) Pre-analytical and analytical factors influencing Alzheimer's disease cerebrospinal fluid biomarker variability. *Clin Chim Acta* 449:9–15. <https://doi.org/10.1016/j.cca.2015.05.024>
- Frisoni GB, Boccardi M, Barkhof F, et al (2017) Strategic roadmap for an early diagnosis of Alzheimer's disease based on biomarkers. *Lancet Neurol* 16:661–676. [https://doi.org/10.1016/S1474-4422\(17\)30159-X](https://doi.org/10.1016/S1474-4422(17)30159-X)
- Frisoni GB, Fox NC, Jack CR, et al (2010) The clinical use of structural MRI in Alzheimer disease. *Nat Rev Neurol* 6:67–77. <https://doi.org/10.1038/nrneurol.2009.215>
- Frölich L, Peters O, Lewczuk P, et al (2017) Incremental value of biomarker combinations to predict progression of mild cognitive impairment to Alzheimer's dementia. *Alzheimers Res Ther* 9:84. <https://doi.org/10.1186/s13195-017-0301-7>
- Gaetani L, Blennow K, Calabresi P, et al (2019) Neurofilament light chain as a biomarker in neurological disorders. *J. Neurol. Neurosurg. Psychiatry* 90
- Garibotto V, Herholz K, Boccardi M, et al (2017) Clinical validity of brain fluorodeoxyglucose positron emission tomography as a biomarker for Alzheimer's disease in the context of a structured 5-phase development framework. *Neurobiol Aging* 52:183–195. <https://doi.org/10.1016/j.neurobiolaging.2016.03.033>
- George JM (2001) The synucleins. *Genome Biol* 3:reviews3002.1. <https://doi.org/10.1186/gb-2001-3-1-reviews3002>
- Giannakopoulos P, Kövari E, Herrmann FR, et al (2008) Interhemispheric distribution of Alzheimer disease and vascular pathology in brain aging.

- <https://doi.org/10.1161/STROKEAHA.108.530337>
- Gorelick PB, Scuteri A, Black SE, et al (2011) Vascular Contributions to Cognitive Impairment and Dementia. *Stroke* 42:2672–2713. <https://doi.org/10.1161/STR.0b013e3182299496>
- Gorno-Tempini ML, Hillis AE, Weintraub S, et al (2011) Classification of primary progressive aphasia and its variants. *Neurology* 76:1006–14. <https://doi.org/10.1212/WNL.0b013e31821103e6>
- Grimmer T, Wutz C, Alexopoulos P, et al (2016) Visual Versus Fully Automated Analyses of 18F-FDG and Amyloid PET for Prediction of Dementia Due to Alzheimer Disease in Mild Cognitive Impairment. *J Nucl Med* 57:204–207. <https://doi.org/10.2967/jnumed.115.163717>
- Guadaño-Ferraz A, Viñuela A, Oeding G, et al (2005) RC3/neurogranin is expressed in pyramidal neurons of motor and somatosensory cortex in normal and denervated monkeys. *J Comp Neurol* 493:554–570. <https://doi.org/10.1002/cne.20774>
- Guo JL, Lee VMY (2014) Cell-to-cell transmission of pathogenic proteins in neurodegenerative diseases. *Nat Med* 20:130–138. <https://doi.org/10.1038/nm.3457>
- Halbgebauer S, Oeckl P, Steinacker P, et al (2021) Beta-synuclein in cerebrospinal fluid as an early diagnostic marker of Alzheimer’s disease. *J Neurol Neurosurg & Psychiatry* 92:349 LP – 356. <https://doi.org/10.1136/jnnp-2020-324306>
- Hampel H, Cummings J, Blennow K, et al (2021) Developing the ATX(N) classification for use across the Alzheimer disease continuum. *Nat Rev Neurol* 17:580–589. <https://doi.org/10.1038/s41582-021-00520-w>
- Hayashi Y (2009) Long-term potentiation: two pathways meet at neurogranin. *EMBO J* 28:2859–2860. <https://doi.org/10.1038/EMBOJ.2009.273>
- Herholz K (2014) Guidance for reading FDG PET scans in dementia patients. *Q J Nucl Med Mol Imaging* 58:332–43
- Herholz K (2003) PET studies in dementia. *Ann Nucl Med* 17:79–89
- Herholz K, Salmon E, Perani D, et al (2002) Discrimination between Alzheimer dementia and controls by automated analysis of multicenter FDG PET. *Neuroimage* 17:302–316. <https://doi.org/10.1006/nimg.2002.1208>
- Herukka S-K, Simonsen AH, Andreasen N, et al (2017) Recommendations for cerebrospinal fluid Alzheimer’s disease biomarkers in the diagnostic evaluation of mild cognitive impairment. *Alzheimer’s Dement* 13:285–295. <https://doi.org/10.1016/j.jalz.2016.09.009>
- Jack CR, Bennett DA, Blennow K, et al (2018) NIA-AA Research Framework: Toward a

- biological definition of Alzheimer's disease. *Alzheimer's Dement* 14:535–562. <https://doi.org/10.1016/j.jalz.2018.02.018>
- Jack CR, Bennett DA, Blennow K, et al (2016a) A/T/N: An unbiased descriptive classification scheme for Alzheimer disease biomarkers. *Neurology* 87:539–547. <https://doi.org/10.1212/WNL.0000000000002923>
- Jack CR, Jr, Knopman DS, et al (2016b) Suspected non-Alzheimer disease pathophysiology—concept and controversy. *Nat Rev Neurol* 12:117. <https://doi.org/10.1038/NRNEUROL.2015.251>
- Jansen WJ, Ossenkoppele R, Knol DL, et al (2015) Prevalence of Cerebral Amyloid Pathology in Persons Without Dementia: A Meta-analysis. *JAMA* 313:1924. <https://doi.org/10.1001/JAMA.2015.4668>
- Jellinger KA, Attems J (2010) Prevalence of dementia disorders in the oldest-old: an autopsy study. *Acta Neuropathol* 119:421–433. <https://doi.org/10.1007/s00401-010-0654-5>
- Johnson JK, Head E, Kim R, et al (1999) Clinical and pathological evidence for a frontal variant of Alzheimer disease. *Arch Neurol* 56:1233–9
- Johnson KA, Minoshima S, Bohnen NI, et al (2013) Appropriate use criteria for amyloid PET: A report of the Amyloid Imaging Task Force, the Society of Nuclear Medicine and Molecular Imaging, and the Alzheimer's Association. *Alzheimer's Dement* 9:E1–E16. <https://doi.org/10.1016/j.jalz.2013.01.002>
- Kaerst L, Kuhlmann A, Wedekind D, et al (2013) Using Cerebrospinal Fluid Marker Profiles in Clinical Diagnosis of Dementia with Lewy Bodies, Parkinson's Disease, and Alzheimer's Disease. *J Alzheimer's Dis* 38:63–73. <https://doi.org/10.3233/JAD-130995>
- Kanaan NM, Pigino GF, Brady ST, et al (2013) Axonal degeneration in Alzheimer's disease: When signaling abnormalities meet the axonal transport system. *Exp. Neurol.* 246:44–53
- Kester MI, Teunissen CE, Crimmins DL, et al (2015) Neurogranin as a cerebrospinal fluid biomarker for synaptic loss in symptomatic Alzheimer disease. *JAMA Neurol* 72:1275–1280. <https://doi.org/10.1001/jamaneurol.2015.1867>
- Kvartsberg H, Duits FH, Ingelsson M, et al (2015) Cerebrospinal fluid levels of the synaptic protein neurogranin correlates with cognitive decline in prodromal Alzheimer's disease. *Alzheimer's Dement* 11:1180–1190. <https://doi.org/10.1016/j.jalz.2014.10.009>
- Lancaster JL, Woldorff MG, Parsons LM, et al (2000) Automated Talairach Atlas Labels For Functional Brain Mapping
- Lange C, Suppa P, Pietrzyk U, et al (2017) Prediction of Alzheimer's Dementia in Patients with Amnesic Mild Cognitive Impairment in Clinical Routine: Incremental Value of

- Biomarkers of Neurodegeneration and Brain Amyloidosis Added Stepwise to Cognitive Status. *J Alzheimer's Dis* 61:373–388. <https://doi.org/10.3233/JAD-170705>
- Levey AI (2021) Progress with Treatments for Alzheimer's Disease. *N Engl J Med* 384:1762–1763. <https://doi.org/10.1056/NEJMe2103722>
- Lewczuk P, Kornhuber J, Toledo JB, et al (2015) Validation of the Erlangen Score Algorithm for the Prediction of the Development of Dementia due to Alzheimer's Disease in Pre-Dementia Subjects. *J Alzheimer's Dis* 48:433–441. <https://doi.org/10.3233/JAD-150342>
- Lewczuk P, Lelental N, Spitzer P, et al (2014) Amyloid- $\beta$  42/40 Cerebrospinal Fluid Concentration Ratio in the Diagnostics of Alzheimer's Disease: Validation of Two Novel Assays. *J Alzheimer's Dis* 43:183–191. <https://doi.org/10.3233/JAD-140771>
- Lewczuk P, Riederer P, O'Bryant SE, et al (2018) Cerebrospinal fluid and blood biomarkers for neurodegenerative dementias: An update of the Consensus of the Task Force on Biological Markers in Psychiatry of the World Federation of Societies of Biological Psychiatry. *World J Biol Psychiatry* 19:244–328. <https://doi.org/10.1080/15622975.2017.1375556>
- Li K, Wei Q, Liu F-F, et al (2018) Synaptic Dysfunction in Alzheimer's Disease: A $\beta$ , Tau, and Epigenetic Alterations. *Mol Neurobiol* 55:3021–3032. <https://doi.org/10.1007/s12035-017-0533-3>
- Litvan I, Agid Y, Calne D, et al (1996) Clinical research criteria for the diagnosis of progressive supranuclear palsy (Steele-Richardson-Olszewski syndrome): report of the NINDS-SPSP international workshop. *Neurology* 47:1–9
- Lleó A, Núñez-Llaves R, Alcolea D, et al (2019) Changes in Synaptic Proteins Precede Neurodegeneration Markers in Preclinical Alzheimer's Disease Cerebrospinal Fluid\*. *Mol Cell Proteomics* 18:546–560. <https://doi.org/https://doi.org/10.1074/mcp.RA118.001290>
- Loewenstein DA, Barker WW, Chang J-Y, et al (1989) Predominant Left Hemisphere Metabolic Dysfunction in Dementia. *Arch Neurol* 46:146–152. <https://doi.org/10.1001/archneur.1989.00520380046012>
- Lucas SJ, Michel CB, Marra V, et al (2018) Glucose and lactate as metabolic constraints on presynaptic transmission at an excitatory synapse. *J Physiol* 596:1699–1721. <https://doi.org/10.1113/JP275107>
- Magistretti PJ (2000) Cellular bases of functional brain imaging: insights from neuron-glia metabolic coupling. *Brain Res* 886:108–112
- Majbour NK, Chiasserini D, Vaikath NN, et al (2017) Increased levels of CSF total but not oligomeric or phosphorylated forms of alpha-synuclein in patients diagnosed with

- probable Alzheimer's disease. *Sci Rep* 7:40263. <https://doi.org/10.1038/srep40263>
- Masliah E, Mallory M, Alford M, et al (2001) Altered expression of synaptic proteins occurs early during progression of Alzheimer's disease. *Neurology* 56:127 LP – 129. <https://doi.org/10.1212/WNL.56.1.127>
- Massa F, Chincari A, Bauckneht M, et al (2021) Added value of semiquantitative analysis of brain FDG-PET for the differentiation between MCI-Lewy bodies and MCI due to Alzheimer's disease. *Eur J Nucl Med Mol Imaging*. <https://doi.org/10.1007/s00259-021-05568-w>
- Massa F, Grisanti S, Brugnolo A, et al (2020) The role of anterior prefrontal cortex in prospective memory: an exploratory FDG-PET study in early Alzheimer's disease. *Neurobiol Aging*. <https://doi.org/10.1016/j.neurobiolaging.2020.09.003>
- Masters CL, Bateman R, Blennow K, et al (2015) Alzheimer's disease. *Nat Rev Dis Prim* 1:15056. <https://doi.org/10.1038/nrdp.2015.56>
- Matsuda H (2001) Cerebral blood flow and metabolic abnormalities in Alzheimer's disease. *Rev Ann Nucl Med* 15:85–92
- Mattsson-Carlsson N, Grinberg LT, Boxer A, et al (2022) Cerebrospinal Fluid Biomarkers in Autopsy-Confirmed Alzheimer Disease and Frontotemporal Lobar Degeneration. *Neurology* 98:e1137 LP-e1150. <https://doi.org/10.1212/WNL.0000000000200040>
- McKeith IG, Boeve BF, Dickson DW, et al (2017) Diagnosis and management of dementia with Lewy bodies. *Neurology* 89:88–100. <https://doi.org/10.1212/WNL.0000000000004058>
- McKhann GM, Knopman DS, Chertkow H, et al (2011) The diagnosis of dementia due to Alzheimer's disease: Recommendations from the National Institute on Aging-Alzheimer's Association workgroups on diagnostic guidelines for Alzheimer's disease. *Alzheimer's Dement* 7:263–269. <https://doi.org/10.1016/j.jalz.2011.03.005>
- Mecca AP, Chen M-K, O'Dell RS, et al (2020) In vivo measurement of widespread synaptic loss in Alzheimer's disease with SV2A PET. *Alzheimer's Dement* 16:974–982. <https://doi.org/10.1002/alz.12097>
- Molinie JL, Blennow K, Dubois B, et al (2014) The clinical use of cerebrospinal fluid biomarker testing for Alzheimer's disease diagnosis: A consensus paper from the Alzheimer's Biomarkers Standardization Initiative. *Alzheimer's Dement* 10:808–817. <https://doi.org/10.1016/j.jalz.2014.03.003>
- Monge-Argilés JA, García-Ayllón M-S, García VM, et al (2020) Relation between alpha-synuclein and core CSF biomarkers in Alzheimer's disease. *Alzheimer's Dement* 16:e041147. <https://doi.org/https://doi.org/10.1002/alz.041147>

- Morbelli S, Bauckneht M, Arnaldi D, et al (2017) 18F-FDG PET diagnostic and prognostic patterns do not overlap in Alzheimer's disease (AD) patients at the mild cognitive impairment (MCI) stage. *Eur J Nucl Med Mol Imaging* 44:2073–2083. <https://doi.org/10.1007/s00259-017-3790-5>
- Morbelli S, Brugnolo A, Bossert I, et al (2015a) Visual Versus Semi-Quantitative Analysis of 18F-FDG-PET in Amnestic MCI: An European Alzheimer's Disease Consortium (EADC) Project. *J Alzheimer's Dis* 44:815–826. <https://doi.org/10.3233/JAD-142229>
- Morbelli S, Brugnolo A, Bossert I, et al (2015b) Visual Versus Semi-Quantitative Analysis of 18F-FDG-PET in Amnestic MCI: An European Alzheimer's Disease Consortium (EADC) Project. *J Alzheimer's Dis* 44:815–826. <https://doi.org/10.3233/JAD-142229>
- Morris E, Chalkidou A, Hammers A, et al (2016) Diagnostic accuracy of (18)F amyloid PET tracers for the diagnosis of Alzheimer's disease: a systematic review and meta-analysis. *Eur J Nucl Med Mol Imaging* 43:374–385. <https://doi.org/10.1007/s00259-015-3228-x>
- Morris JC, Roe CM, Xiong C, et al (2010) APOE predicts amyloid-beta but not tau Alzheimer pathology in cognitively normal aging. *Ann Neurol* 67:122–131. <https://doi.org/https://doi.org/10.1002/ana.21843>
- Mosconi L (2005) Brain glucose metabolism in the early and specific diagnosis of Alzheimer's disease. *Eur J Nucl Med Mol Imaging* 32:486–510. <https://doi.org/10.1007/s00259-005-1762-7>
- Nakashima T, Nakayama N, Miwa K, et al (2007) Focal Brain Glucose Hypometabolism in Patients with Neuropsychologic Deficits after Diffuse Axonal Injury. *Am J Neuroradiol* 28:236 LP – 242
- Nestor PJ, Caine D, Fryer TD, et al (2003) The topography of metabolic deficits in posterior cortical atrophy (the visual variant of Alzheimer's disease) with FDG-PET. *J Neurol Neurosurg Psychiatry* 74:1521–9
- Nobili F, Arbizu J, Bouwman F, et al (2018a) European Association of Nuclear Medicine and European Academy of Neurology recommendations for the use of brain <sup>18</sup>F-fluorodeoxyglucose positron emission tomography in neurodegenerative cognitive impairment and dementia: Delphi consensus. *Eur J Neurol* 25:1201–1217. <https://doi.org/10.1111/ene.13728>
- Nobili F, Festari C, Altomare D, et al (2018b) Automated assessment of FDG-PET for differential diagnosis in patients with neurodegenerative disorders. *Eur. J. Nucl. Med. Mol. Imaging* 45:1557–1566
- Oeckl P, Halbgebauer S, Anderl-Straub S, et al (2020) Targeted Mass Spectrometry Suggests Beta-Synuclein as Synaptic Blood Marker in Alzheimer's Disease. *J Proteome Res*

- 19:1310–1318. <https://doi.org/10.1021/acs.jproteome.9b00824>
- Oeckl P, Metzger F, Nagl M, et al (2016) Alpha-, Beta-, and Gamma-synuclein Quantification in Cerebrospinal Fluid by Multiple Reaction Monitoring Reveals Increased Concentrations in Alzheimer's and Creutzfeldt-Jakob Disease but No Alteration in Synucleinopathies. *Mol Cell Proteomics* 15:3126–3138. <https://doi.org/10.1074/mcp.M116.059915>
- Ou Y-N, Xu W, Li J-Q, et al (2019) FDG-PET as an independent biomarker for Alzheimer's biological diagnosis: a longitudinal study. *Alzheimers Res Ther* 11:57. <https://doi.org/10.1186/s13195-019-0512-1>
- Overk CR, Masliah E (2014) Pathogenesis of synaptic degeneration in Alzheimer's disease and Lewy body disease. *Biochem Pharmacol* 88:508–516. <https://doi.org/10.1016/j.bcp.2014.01.015>
- Pagani M, Giuliani A, Öberg J, et al (2017) Progressive Disintegration of Brain Networking from Normal Aging to Alzheimer Disease: Analysis of Independent Components of 18F-FDG PET Data. *J Nucl Med* 58:1132–1139. <https://doi.org/10.2967/jnumed.116.184309>
- Perani D, Della Rosa PA, Cerami C, et al (2014) Validation of an optimized SPM procedure for FDG-PET in dementia diagnosis in a clinical setting. *NeuroImage Clin* 6:445–454. <https://doi.org/10.1016/j.nicl.2014.10.009>
- Petersen RC (2004) Mild cognitive impairment as a diagnostic entity. *J Intern Med* 256:183–194. <https://doi.org/https://doi.org/10.1111/j.1365-2796.2004.01388.x>
- Petersen RC, Caracciolo B, Brayne C, et al (2014) Mild cognitive impairment: a concept in evolution. *J Intern Med* 275:214–228. <https://doi.org/10.1111/joim.12190>
- Petersen RC, Roberts RO, Knopman DS, et al (2009) Mild Cognitive Impairment. *Arch Neurol* 66:1447–55. <https://doi.org/10.1001/archneurol.2009.266>
- Picco A, Polidori MC, Ferrara M, et al (2014) Plasma antioxidants and brain glucose metabolism in elderly subjects with cognitive complaints. *Eur J Nucl Med Mol Imaging* 41:764–775. <https://doi.org/10.1007/s00259-013-2638-x>
- Portelius E, Zetterberg H, Skillbäck T, et al (2015) Cerebrospinal fluid neurogranin: relation to cognition and neurodegeneration in Alzheimer's disease. *Brain* 138:3373–85. <https://doi.org/10.1093/brain/awv267>
- Postuma RB, Berg D, Stern M, et al (2015) MDS clinical diagnostic criteria for Parkinson's disease. *Mov Disord* 30:1591–1601. <https://doi.org/10.1002/mds.26424>
- Prestia A, Caroli A, Herholz K, et al (2013) Diagnostic accuracy of markers for prodromal Alzheimer's disease in independent clinical series. *Alzheimer's Dement* 9:677–686. <https://doi.org/10.1016/j.jalz.2012.09.016>

- Prestia A, Caroli A, Wade SK, et al (2015) Prediction of AD dementia by biomarkers following the NIA-AA and IWG diagnostic criteria in MCI patients from three European memory clinics. *Alzheimer's Dement* 11:1191–1201. <https://doi.org/10.1016/j.jalz.2014.12.001>
- Provost K, La Joie R, Strom A, et al (2021) Crossed cerebellar diaschisis on 18F-FDG PET: Frequency across neurodegenerative syndromes and association with 11C-PIB and 18F-Flortaucipir. *J Cereb Blood Flow Metab* 0271678X211001216. <https://doi.org/10.1177/0271678X211001216>
- Rascovsky K, Hodges JR, Knopman D, et al (2011) Sensitivity of revised diagnostic criteria for the behavioural variant of frontotemporal dementia. *Brain* 134:2456–2477. <https://doi.org/10.1093/brain/awr179>
- Reddy PH, Mani G, Park BS, et al (2005) Differential loss of synaptic proteins in Alzheimer's disease: Implications for synaptic dysfunction. *J Alzheimer's Dis* 7:103–117. <https://doi.org/10.3233/JAD-2005-7203>
- Represa A, Deloulme J, Sensenbrenner M, et al (1990) Neurogranin: immunocytochemical localization of a brain-specific protein kinase C substrate. *J Neurosci* 10:3782–3792. <https://doi.org/10.1523/JNEUROSCI.10-12-03782.1990>
- Respondek G, Kurz C, Arzberger T, et al (2017) Which ante mortem clinical features predict progressive supranuclear palsy pathology? *Mov Disord* 32:995–1005. <https://doi.org/10.1002/mds.27034>
- Roberts R, Knopman DS (2013) Classification and Epidemiology of MCI. *Clin Geriatr Med* 29:753–772. <https://doi.org/10.1016/j.cger.2013.07.003>
- Rosén C, Farahmand B, Skillbäck T, et al (2015) Benchmarking biomarker-based criteria for Alzheimer's disease: Data from the Swedish Dementia Registry, SveDem. *Alzheimer's Dement* 11:1470–1479. <https://doi.org/10.1016/j.jalz.2015.04.007>
- Sanfilippo C, Forlenza O, Zetterberg H, Blennow K (2016) Increased neurogranin concentrations in cerebrospinal fluid of Alzheimer's disease and in mild cognitive impairment due to AD. *J Neural Transm* 123. <https://doi.org/10.1007/s00702-016-1597-3>
- Scheltens P, Kuiper M, Ch Wolters E, et al (1992) Atrophy of medial temporal lobes on MRI in “probable” Alzheimer's disease and normal ageing: diagnostic value and neuropsychological correlates. *J Neurol Neurosurg Psychiatry* 55:967–972. <https://doi.org/10.1136/jnnp.55.10.967>
- Schipke CG, De Vos A, Fuentes M, et al (2018) Neurogranin and BACE1 in CSF as Potential Biomarkers Differentiating Depression with Cognitive Deficits from Early Alzheimer's



- Disease: A Pilot Study. *Dement Geriatr Cogn Dis Extra* 8:277–289.  
<https://doi.org/10.1159/000489847>
- Selkoe DJ (2002) Alzheimer's disease is a synaptic failure. *Science* (80-. ). 298:789–791
- Shaffer JL, Petrella JR, Sheldon FC, et al (2013) Predicting Cognitive Decline in Subjects at Risk for Alzheimer Disease by Using Combined Cerebrospinal Fluid, MR Imaging, and PET Biomarkers. *Radiology* 266:583–591. <https://doi.org/10.1148/radiol.12120010>
- Silverman DHS, Chen W, Czernin J, et al (2001) Positron emission tomography in evaluation of dementia: Regional brain metabolism and long-term outcome. *J Am Med Assoc* 286:2120–2127. <https://doi.org/10.1001/jama.286.17.2120>
- Simonsen AH, Herukka S-K, Andreasen N, et al (2017) Recommendations for CSF AD biomarkers in the diagnostic evaluation of dementia. *Alzheimer's Dement* 13:274–284. <https://doi.org/10.1016/j.jalz.2016.09.008>
- Stephan BCM, Hunter S, Harris D, et al (2012) The neuropathological profile of mild cognitive impairment (MCI): a systematic review. *Mol Psychiatry* 17:1056–1076. <https://doi.org/10.1038/mp.2011.147>
- Taoufik E, Kouroupi G, Zygogianni O, Matsas R (2018) Synaptic dysfunction in neurodegenerative and neurodevelopmental diseases: an overview of induced pluripotent stem-cell-based disease models. *Open Biol* 8:180138. <https://doi.org/10.1098/rsob.180138>
- Tarawneh R, D'Angelo G, Crimmins D, et al (2016) Diagnostic and prognostic utility of the synaptic marker neurogranin in Alzheimer disease. *JAMA Neurol* 73:561–571. <https://doi.org/10.1001/jamaneurol.2016.0086>
- Terry RD, Masliah E, Salmon DP, et al (1991) Physical basis of cognitive alterations in alzheimer's disease: Synapse loss is the major correlate of cognitive impairment. *Ann Neurol* 30:572–580. <https://doi.org/10.1002/ana.410300410>
- Teunissen CE, Petzold A, Bennett JL, et al (2009) A consensus protocol for the standardization of cerebrospinal fluid collection and biobanking. *Neurology* 73:1914–1922. <https://doi.org/10.1212/WNL.0b013e3181c47cc2>
- Thal DR, Attems J, Ewers M (2014) Spreading of Amyloid, Tau, and Microvascular Pathology in Alzheimer's Disease: Findings from Neuropathological and Neuroimaging Studies. *J Alzheimer's Dis* 42:S421–S429. <https://doi.org/10.3233/JAD-141461>
- Thompson PM, Hayashi KM, Zubicaray G de, et al (2003) Dynamics of Gray Matter Loss in Alzheimer's Disease. *J Neurosci* 23:994–1005. <https://doi.org/10.1523/JNEUROSCI.23-03-00994.2003>
- Twohig D, Nielsen HM (2019)  $\alpha$ -synuclein in the pathophysiology of Alzheimer's disease.

- Mol Neurodegener 14:. <https://doi.org/10.1186/S13024-019-0320-X>
- van Harten AC, Kester MI, Visser P-J, et al (2011) Tau and p-tau as CSF biomarkers in dementia: a meta-analysis. Clin Chem Lab Med 49:353–66. <https://doi.org/10.1515/CCLM.2011.086>
- Varrone A, Asenbaum S, Vander Borgh T, et al (2009) EANM procedure guidelines for PET brain imaging using [18F]FDG, version 2. Eur J Nucl Med Mol Imaging 36:2103–2110. <https://doi.org/10.1007/s00259-009-1264-0>
- Vemuri P, Wiste HJ, Weigand SD, et al (2009) MRI and CSF biomarkers in normal, MCI, and AD subjects: Predicting future clinical change. Neurology 73:294–301. <https://doi.org/10.1212/WNL.0b013e3181af79fb>
- Verwey NA, van der Flier WM, Blennow K, et al (2009) A worldwide multicentre comparison of assays for cerebrospinal fluid biomarkers in Alzheimer’s disease. Ann Clin Biochem 46:235–240. <https://doi.org/10.1258/acb.2009.008232>
- Wahlund LO, Barkhof F, Fazekas F, et al (2001) A new rating scale for age-related white matter changes applicable to MRI and CT. Stroke 32:1318–1322. <https://doi.org/10.1161/01.STR.32.6.1318>
- Walhovd KB, Fjell AM, Brewer J, et al (2010) Combining MR Imaging, Positron-Emission Tomography, and CSF Biomarkers in the Diagnosis and Prognosis of Alzheimer Disease. Am J Neuroradiol 31:347–354. <https://doi.org/10.3174/ajnr.A1809>
- Walker Z, Gandolfo F, Orini S, et al (2018) Clinical utility of FDG PET in Parkinson’s disease and atypical parkinsonism associated with dementia. Eur J Nucl Med Mol Imaging 45:1534–1545. <https://doi.org/10.1007/s00259-018-4031-2>
- Wang L (2019) Association of cerebrospinal fluid Neurogranin with Alzheimer’s disease. Aging Clin Exp Res 31:185–191. <https://doi.org/10.1007/s40520-018-0948-3>
- Welge V, Fiege O, Lewczuk P, et al (2009) Combined CSF tau, p-tau181 and amyloid- $\beta$  38/40/42 for diagnosing Alzheimer’s disease. J Neural Transm 116:203–212. <https://doi.org/10.1007/s00702-008-0177-6>
- Wellington H, Paterson RW, Portelius E, et al (2016) Increased CSF neurogranin concentration is specific to Alzheimer disease. Neurology 86:829–835. <https://doi.org/10.1212/WNL.0000000000002423>
- Weston PSJ, Paterson RW, Dickson J, et al (2016) Diagnosing Dementia in the Clinical Setting: Can Amyloid PET Provide Additional Value Over Cerebrospinal Fluid? J Alzheimer’s Dis 54:1297–1302. <https://doi.org/10.3233/JAD-160302>
- Willemse EAJ, De Vos A, Herries EM, et al (2018) Neurogranin as Cerebrospinal Fluid Biomarker for Alzheimer Disease: An Assay Comparison Study. Clin Chem 64:927–

937. <https://doi.org/10.1373/clinchem.2017.283028>
- Womack KB, Diaz-Arrastia R, Aizenstein HJ, et al (2011) Temporoparietal Hypometabolism in Frontotemporal Lobar Degeneration and Associated Imaging Diagnostic Errors. *Arch Neurol* 68:329–37. <https://doi.org/10.1001/archneurol.2010.295>
- Woodward MC, Rowe CC, Jones G, et al (2015) Differentiating the Frontal Presentation of Alzheimer’s Disease with FDG-PET. *J Alzheimer’s Dis* 44:233–242. <https://doi.org/10.3233/JAD-141110>
- Zetterberg H, Skillbäck T, Mattsson N, et al (2016) Association of Cerebrospinal Fluid Neurofilament Light Concentration With Alzheimer Disease Progression. *JAMA Neurol* 73:60–67. <https://doi.org/10.1001/jamaneurol.2015.3037>
- Zhang D, Wang Y, Zhou L, et al (2011) Multimodal classification of Alzheimer’s disease and mild cognitive impairment. *Neuroimage* 55:856–867. <https://doi.org/10.1016/j.neuroimage.2011.01.008>
- Zimmer ER, Parent MJ, Souza DG, et al (2017) [18F]FDG PET signal is driven by astroglial glutamate transport. *Nat Neurosci* 20:393–395. <https://doi.org/10.1038/nn.4492>

## 7. Acknowledgments

I want to express my gratitude to Prof. Flavio Nobili, who served as my former supervisor. Thank you for having faith in me from the begin, instilling in me a work ethic and the capability to take responsibility in my role as a clinician, and for standing as my mentor while I proceeded along the road of research.

Thanks to my current supervisor, Prof. Matteo Pardini, with whom I've made the first steps toward completing my degree in Medicine and Surgery and who I can now cooperatively collaborate, as well as to the entire research team, everyone from the youngest to the oldest.

A particular thank goes to Prof. Silvia Morbelli, who has always supported me during my early research experiences and who holds a unique place in my heart as an 'elder' sister in neuroscience.

And in particular, I recognize Dr. Luana Benedetti, who has a position of honor. She first saw potential in me that I wasn't even aware of, and she inspired me to develop as a doctor and researcher by imparting her enthusiasm and conviction to achieve ever-new goals, day after day.

What I've accomplished in my career would not have been possible without the support provided by my family and other close friends.

Thanks to Ilaria and little Benny. Tutto qui accade per noi...e per chi verrà.