Five lichen species new to Poland

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Abstract: Athallia cerinelloides, Caloplaca ulcerosa, Flavoplaca arcis, Lecanora sinuosa and Sclerophora amabilis are reported for the first time from Poland. Descriptions, notes on similar species, habitat preferences and general distribution are provided for each species.

Keywords: lichenized fungi, distribution, Poland

INTRODUCTION

Lichens can be considered one of the best studied groups of fungi in Poland. Lichenological research on Polish territories began over 200 years ago and is being carried out with a constantly growing intensity to this day. The last edition of the list of Polish lichens (Faltynowicz & Kossowska, 2016) includes 1624 taxa; nevertheless, species new for Poland are still recorded (e.g., Kubiak & Wilk, 2016; Kukwa et al., 2017; Matura et al., 2017; Szczepańska et al., 2019; Szczepańska, 2020). The list of species is also enriched by taxa described as new to science (Guzow-Krzemińska et al., 2017, 2018, 2019; Czarnota & Guzow-Krzemińska, 2018; Ertz et al., 2018; Ossowska et al., 2021). In this article, we present five additional species that are reported from Poland for the first time.

MATERIAL AND METHODS

Specimens studied were collected by authors during various lichenological investigations and are deposited in following herbaria: OLTC, UGDA and hb. Kossowska. The taxa were identified by classical techniques, using dissecting and light microscopes and standard chemical reagents: 10% potassium dioxide (K), sodium hypochlorite (C), and p-phenylenediamine in ethanol (Pd). Descriptions of the species are based on own observations and measurements. Nomenclature of lichens follows van Herk & Aptroot (1999), Tibell (1999), Arup (2006), Vondrák et al. (2013) and Nimis (2016). Localities of each species are placed in ATPOL grid square system (Zając, 1978) modified by Cieśliński & Fałtynowicz (1993).

THE SPECIES

ATHALLIA CERINELLOIDES (Erichsen) Arup, Frödén & Søchting (syn. *Caloplaca cerinelloides* Erichsen) Thallus crustose, grey, thin, in parts immersed in the bark, smooth. Apothecia up to 0.4 mm diam., circular, grouped. Proper margin persistent, yellow; thalline margin inconspicuous, grey-yellow to yellow. Disc flat, yellow to pale orange-yellow. Hypothecium up to 43 µm thick. Asci 8-spored. Ascospores polarilocular, (8–)10–14 × (5–)6–7 µm, ellipsoid, septum 4–5(–6) µm wide. Thallus K negative, apothecial margin and discs K+ purple (see also Arup, 2009; Fletcher & Laundon, 2009).

The recently recognised genus Athallia Arup, Frödén & Søchting corresponds to the Caloplaca holocarpa group and contains mainly species with crustose thalli (Arup et al., 2013). Athallia cerinelloides is morphologically very similar

to A. cerinella (Nyl.) Arup, Frödén & Søchting, which differs in mostly 12-16-spored asci (Arup, 2009; Fletcher & Laundon, 2009). The species may also be confused with A. pyracea (Ach.) Arup, Frödén & Søchting; however, thalli of the latter species usually consist of slightly convex, greyish to pale orange areoles and produce larger (0.5–1 mm diam.), yellow-orange to orange apothecia, surrounded by thin, grey or yellowish grey thalline margin (Arup, 2009). Rare epiphytic or epixylic form of A. holocarpa (Hoffm.) Arup, Frödén & Søchting can also be similar to A. cerinelloides, but that species develops crowdedangular, larger apothecia (0.3-0.6(-1.0) mm) with usually darker discs (yellow-orange to orange, rarely yellow) and higher hypothecium (50-80 µm) (Arup, 2009; Fletcher & Laundon, 2009).

The species is a typical epiphyte which grows on various species of bushes and trees, especially Populus tremula, but also e.g. Betula, Fraxinus, Juglans, Juniperus, Picea, Ribes and Sambucus. It has also rarely been found on wood and rocks (Arup, 2009; Fletcher & Laundon, 2009). According to Arup (2009), it is likely that A. cerinelloides requires dust enrichment when growing on acidic substrata. In Poland it was found on bark of roadside trees and within a grey heron colony ("Czapliniec w Wierzysku" nature reserve). Accompanying species included Candelaria pacifica M. Westb. & Arup, Lecania cyrtella (Ach.) Th. Fr., L. naegelii (Hepp) Diederich & van den Boom, Myriolecis albescens (Hoffm.) Śliwa, Zhao Xin & Lumbsch, M. hagenii (Ach.) Śliwa, Zhao Xin & Lumbsch, M. sambuci (Pers.) Clem., Physcia tenella (Scop.) DC., Phaeophyscia nigricans (Flörke) Moberg, P. orbicularis (Neck.) Moberg and Xanthoria parietina (L.) Th. Fr. Athallia cerinelloides is a widespread, although locally uncommon species. It has been reported from many regions, namely Eastern, Central and Western Europe, Scandinavia, British Isles, Baltic Sea Region, Iberian Peninsula, Mediterranean and Black Sea Basin (e.g. Randlane & Saag 1999; Llimona & Hladun, 2001; Aptroot et al., 2004; Liška et al., 2008; Søchting & Alstrup, 2008; Abbott, 2009; Arup, 2009; Fletcher & Laundon, 2009; Urbanavichus, 2010; Vondrák & Liška, 2013). Outside Europe, it is also known from Northern Africa (Thor & Nascimbene, 2010), Asia (Fletcher & Laundon, 2009), New Zealand (Galloway, 2007) and North America (Esslinger, 2019).

Specimens examined. Poland. Bory Tucholskie, "Czapliniec w Wierzysku" nature reserve, 54.093806°N, 17.967444°E, elev. 190 m, ATPOL grid square Bc-16, on bark of fallen Fagus sylvatica, 14 Apr. 2012, A. Kowalewska 2012/27 (UGDA L-21274); Pojezierze Kaszubskie, Gdańsk Łostowice, 54.313972°N, 18.575056°E, elev. 90 m, ATPOL grid square Ad-90, orchard, on Malus sp., 16 Apr. 2016, A. Kowalewska s.n. (UGDA L-39662); Pobrzeże Kaszubskie, Gdańsk, Kartuska street, 54.342778°N, 18.547500°E, ATPOL grid square Ad-80, on Populus sp., 15 Oct. 1997, B. Ciechanowicz s.n. (UGDA L-49761); Pojezierze Kaszubskie, Gdynia Orłowo, 54.482222°N, 18.565278°E, elev. 3 m, ATPOL grid square Ad-70, open place, by the beach, on Acer platanoides, 29 May 2010, M. Kukwa 8020 (UGDA L-15969); Pojezierze Kaszubskie, Kaszubski Landscape Park, "Żurawie Błota" nature reserve, 54.410861°N, 18.006278°E, elev. 207 m, ATPOL grid square Ac-86, on Populus sp., 12 May 2012, A. Kowalewska s.n. (UGDA L-38484); Pradolina Łeby i Redy, Reda, by railway station, 54.594680°N, 18.354140°E, elev. 20 m, ATPOL grid square Ac-68, trees by road, on Acer pseudoplatanus, 18 Feb. 2020, M. Kukwa 20872 (UGDA L-29509).

CALOPLACA ULCEROSA Coppins & P. James

Thallus thin, flat, smooth to uneven, sometimes with shallow pustules, grey-white. Soralia scattered, discrete, flat to concave and ulcerose, with greenish soredia; exposed soredia never pigmented grey-green. Apothecia with thin, greywhite thalline margin, orange proper margin and orange discs. Ascospores eight in ascus, polarilocular, $9-12 \times 5-6.5 \mu m$, septum 4-5µm. Thallus and soralia K negative, disc and proper margin K+ purple (see also Coppins & James 1979; Fletcher & Laundon, 2009). According to Kondratyuk et al. (2018) the species belongs to the genus Coppinsiella S. Y. Kondr. & L. Lőkös (as C. ulcerosa (Coppins & P. James) S. Y. Kondr. & L. Lőkös). This genus has been recently described and comprises four species (one still undescribed), which form a well-supported sister clade to Athallia (Kondratyuk et al., 2018). Whether the genus should be considered as separate entity or be synonymized with Athallia (as suggested by Wijayawardene et al., 2022), requires further research. Caloplaca ulcerosa can be easily confused with C. obscurella (J. Lahm ex Körb.) Th. Fr., which develops similar soralia,

but differs mainly in the presence of brown apothecia (Fletcher & Laundon, 2009). Both species often occur in sterile state and can then be distinguished by the colour of external soredia, which are blue-grey (exposed hyphae with grey, K negative or rarely K+ violet, N± reddish pigment) in C. obscurella and unpigmented in C. ulcerosa (Fletcher & Laundon, 2009). Caloplaca substerilis Vondrák, Palice & van den Boom is another species with similar morphology but can be separated by its endophloeodal or minutely squamulose thallus and soralia formed in bark crevices or on margins of squamules (Vondrák et al., 2013). Caloplaca aff. ulcerosa (as C. "ulcerosa" in Vondrák et al., 2013) known from Austria and North America is morphologically more similar to C. substerilis than to C. ulcerosa (Vondrák et al., 2013), although North American records were previously subsumed under the latter name (Wetmore, 2005; Vondrák et al., 2009a, 2013). This material may represent another, as yet undescribed species (Vondrák et al., 2013).

The species grows on bark of various deciduous trees and shrubs (Limonium); it was also reported from shaded limestone rock in the Czech Republic (Vondrák et al., 2009a). In Europe, Middle East and North Africa it was found mainly in the areas up to 50 km from a sea coast; however, in North America it was reported from further inland (Vondrák et al., 2009a). In Poland the species was recorded few meters from the sea shore on bark of Populus × canadensis. It was accompanied by Phaeophyscia orbicularis, Physcia tenella, Polycauliona phlogina (Ach.) Arup, Frödén & Søchting and Xanthoria parietina. Caloplaca ulcerosa is a widely distributed species. It is known from Northern Africa (Vondrák et al., 2009a; Thor & Nascimbene, 2010), the Middle East (Seaward et al., 2008; Vondrák et al., 2009a) and many regions of Europe, including Iberian Peninsula, British Isles, Scandinavia, Baltic Sea Region, Mediterranean and Black Sea Basin, as well as Western, Central and Eastern Europe (e.g. Randlane & Saag 1999; Llimona & Hladun, 2001; Aptroot et al., 2004; Abbott, 2009; Fletcher & Laundon, 2009; Vondrák et al., 2009a; Urbanavichus, 2010; Westberg et al., 2021). The species is also known from the Southern Hemisphere, namely Australia (Wetmore, 2005) and French Southern and Antarctic Lands in the southern Indian Ocean (Aptroot et al., 2011); however, due to the core occurrence of the species in the Northern Hemisphere, these records may belong to other similar species. Records from North America (Wetmore, 2005, 2009; Esslinger, 2019) probably represent an undescribed species (see comments above and Vondrák et al., 2009a, 2013).

Specimens examined. Poland. Mierzeja Helska, Nadmorski Landscape Park, SE of Chałupy, forest section No. 9H, 54.752491°N, 18.533442°E, elev. 5 m, ATPOL grid square Ad–40, group of *Populus* × *canadensis* and *Sorbus aucuparia* by dunes, on *Populus* × *canadensis*, 15 Aug. 2019, M. Kukwa 20647 & 20649 (UGDA L-29149 & 29151).

FLAVOPLACA ARCIS (Poelt & Vězda) Arup, Frödén & Søchting (syn. *Caloplaca arcis* (Poelt & Vězda) Arup)

Thallus crustose-subsquamulose, yellow, consisting of undulate, sometimes rosette-like areoles and minutely but distinctly lobate at the margins. Areoles rather thick, 0.4-2 mm in diam., some covered with coarse, globose blastidia, which are concolorous with thallus. Blastidia on both surface and margin of areoles, especially abundant in the centre of thalli. Apothecia absent in examined specimens; if present, zeorine, adnate to sessile, yellow and 0.3-1.5 mm wide. Thallus K+ purple (see Arup 2006). Flavoplaca arcis is a member of F. citrina group, easily distinguished by blastidiate thalli and the presence of short lobes at the margins. According to Arup (2006), this species is often fertile; while it may be true for Nordic populations, only 37% of specimens collected in the Black Sea region bore apothecia (Vondrák et al., 2009b). Polish specimens were sterile, covered with abundant vegetative propagules. Other features of the collected specimens are consistent with the description of Arup (2006). This species was formerly described as a variety within Caloplaca citrina complex by Poelt and Vězda (Vězda, 1990), but later raised to the species level by Arup (2006). As a result of multi-gene phylogenetic analyses of Teloschistaceae, Caloplaca arcis (Poelt & Vězda) Arup and other related taxa were transferred into newly established genus Flavoplaca Arup, Søchting & Frödén (Arup et al., 2013). Flavoplaca arcis may be confused with other similar members of F. citrina group, especially F. dichroa (Arup) Arup, Frödén & Søchting and F. limonia (Nimis &

Poelt) Arup, Frödén & Søchting. The former was reported from Poland by Wilk (2011); the latter is known from the neighbouring countries, i.e. Germany (Wirth et al., 2013) and Czech Republic (Vondrák et al., 2009b), and its occurrence in Poland is very probable. Both these taxa produce true soredia, developing from either cracked or eroded blastidia, and lack short lobes at the thallus margins (Arup, 2006; Vondrák et al., 2009b). Flavoplaca flavocitrina (Nyl.) Arup, Frödén & Søchting is another similar species. It is common in Poland (Wilk, 2011) and is truly sorediate with distinct, often labriform soralia developing from smaller (0.1-1.0(-1.5) mm across), somewhat squamulose areoles (Arup, 2006; Vondrák et al., 2009b).

Flavoplaca arcis grows on base-rich silicate and calcareous rocks and stones (Arup, 2006; Vondrák at al., 2009b; Wirth et al., 2013). The species often inhabits also man-made substrata (Arup, 2006; Malíček et al., 2014; Wirth et al., 2013). It grows in rather exposed places on both vertical and horizontal surfaces (Arup, 2006). In Poland Flavoplaca arcis has been found on walls of an old mountain fortress, built in 18th century in the Sudety Mts (SW Poland). It grew there on vertical surfaces of brick walls facing south and south-east. Accompanying lichen species included Candelariella aurella (Hoffm.) Zahlbr., Circinaria contorta (Hoffm.) A. Nordin, Savić & Tibell, Caloplaca teicholyta (Ach.) J. Steiner, Lathagrium fuscovirens (With.) Otálora, P.M. Jørg. & Wedin, Myriolecis albescens, M. dispersa (Pers.) Śliwa, Zhao Xin & Lumbsch, Phaeophyscia orbicularis, Protoparmeliopsis muralis (Schreb.) M. Choisy, and Xanthoparmelia verruculifera (Nyl.) O. Blanco et al. The species is widespread in Europe, although not frequently recorded. It may be rare, or included in the wide concept of similar F. citrina (Hoffm.) Arup, Frödén & Søchting. Known localities include Western and Central Europe, Scandinavia, the Mediterranean together with the Balkan Peninsula, and the Canary Islands (Arup, 2006; Vondrák at al., 2009b).

Specimens examined. Poland. Sudety Mts, Kotlina Kłodzka Basin, Kłodzko Fortress, 50.442250°N, 16.652250°E, elev. 345 m, ATPOL grid square Fb–26, on brick walls, 15 August 2019, M. Kossowska & W. Fałtynowicz s.n. (hb. Kossowska 1569, 1579). LECANORA SINUOSA Herk & Aptroot

Thallus thick, whitish-grey. Apothecia sessile. Thalline margin thick, sinuous, crenulate, 0.12-0.2 mm in section, with angular groups of large crystals (amphithecium pulicaris-type). Hymenium ca. 70 um high. Epihymenium brown, *pulicaris*-type, sparsely filled with tiny, pale brownish crystals between the paraphyses (well-visible in polarised light). Asci 8-spored. Ascospores hyaline, broadly ellipsoidal, 12-14 \times 6–7.5 µm. Thallus and thalline margin C and Pd negative, K+ yellow (see also van Herk & Aptroot, 1999). Lecanora sinuosa is a member of L. subfusca group. The species is morphologically similar to L. argentata (Ach.) Malme (syn. L. subrugosa Nyl.) and L. chlarotera Nyl. Both these species have *pulicaris*-type amphithecium, but differ in *glabrata*-type and *chlarotera*-type epihymenium, respectively (Brodo, 1984; Jüriado, 1998; van Herk & Aptroot, 1999). Lecanora pulicaris (Pers.) Ach. shares the same type of amphithecium and epihymenium with L. sinuosa, but in the former species the apothecia have thinner and smooth (rarely coarse) thalline margin, which very often reacts Pd+ red due to the presence of fumarprotocetraric acid (Brodo, 1984; Jüriado, 1998; van Herk & Aptroot, 1999; Malíček, 2014). Anatomically Lecanora sinuosa resembles L. hybocarpa (Tuck.) Brodo, but the latter species has thinner thallus and regular and relatively thinner thalline margin (van Herk & Aptroot, 1999). Lecanora sinuosa is a typical epiphytic species found on well-lit and exposed trunks of roadside trees (Acer, Quercus, Populus, Ulmus) (van Herk & Aptroot, 1999; Wirth et al., 2013). In Poland it was found on bark of Populus balsamifera growing between the road and Pucka Bay in a well-lit situation. Van Herk and Aptroot (1999) reported that the lichen associations with Lecanora sinusoa are often extremely rich in species. It is also the case of Polish locality, where it was accompanied by several lichens: Alyxoria varia (Pers.) Ertz & Tehler, Athallia cerinella, Polycauliona phlogina, Calogaya saxicola (Hoffm.) Vondrák s.l., Lecania cyrtella, Lecania erysibe (Ach.) Mudd, Lecanora carpinea (L.) Vain., Myriolecis dispersa, M. hagenii, M. persimilis (Th. Fr.) Śliwa, Zhao Xin & Lumbsch, Lecidella elaeochroma (Ach.) M. Choisy, Phaeophyscia orbicularis, Physcia adscendens H. Olivier, Rinodina pyrina (Ach.) Arnold and Xanthoria parietina. So far, the species has been reported from scattered localities in

Western Europe, namely in Belgium, Germany and the Netherlands (van Herk & Aptroot, 1999; de Bruyn et al., 2000; Aptroot et al., 2004; Van den Broeck et al., 2006).

Specimen examined. Poland. Mierzeja Helska, Nadmorski Landscape Park, SE part of Kuźnica, 54.729167°N, 18.594722°E, elev. c. 2 m, ATPOL grid square Ad–40, roadside trees, on *Populus balsamifera*, 14 July 2019, M. Kukwa 20246a (UGDA L-29027).

SCLEROPHORA AMABILIS (Tibell) Tibell

Thallus crustose, immersed, photobiont Trentepohlia. Ascomata stalked, rather tall (0.6-1.7 mm), stalks brown to yellowish, often with yellow or white pruina in uppermost part. Capitulum spherical, 0.4-0.5 mm diam., with a small collar-like basal extension of the excipulum, usually yellowish or whitish pruinose. Pruina on ascomata initially light yellow, during maturation turn violet-red. Ascospores globose, 4-6.5 µm, with minute verrucose ornamentation of the wall (see also Tibell, 1999). Sclerophora amabilis is similar to S. pallida (Pers.) Y. J. Yao & Spooner, S. peronella (Ach.) Tibell and S. farinacea (Chevall.) Chevall., but differs from them, among other things, in the size of ascospores, which are 7-8 µm in S. pallida and S. farinacea, and 3.0-3.5 µm in S. peronella (for other differences see Tibell, 1999). Tibell (1999) points out that European material of S. amabilis differs slightly from specimens from New Zealand, from where the species was originally described, and the European material may represent a distinct species (see Schultz & Steindl, 2018). The species occurs on the bark of old deciduous trees (Acer, Fagus, Fraxinus, Ulmus, Populus, and Tilia), and wood (Tibell, 1999; Diederich et al., 2012). In Poland Sclerophora amabilis was found on the trunk of an old roadside Acer negundo growing on the edge of the wide floodplain of the Vistula River. The following lichens have been noted as accompanying species: Alyxoria varia, Amandinea punctata (Hoffm.) Coppins & Scheid., Anisomeridium polypori (Ellis & Everh.) M.E. Barr, Bacidia rubella (Hoffm.) A. Massal., Chaenotheca trichialis (Ach.) Hellb., Lecania naegelii, Lepraria vouauxii (Hue) R.C. Harris, Phaeophyscia orbicularis, Physconia grisea (Lam.) Poelt, and P. *perisidiosa* (Erichsen) Moberg. Some European authors consider Sclerophora amabilis as a good indicator of natural habitats such as old-growth beech forests (Tibell, 1999; Malíček et al., 2014).

On the other hand, this species was reported from an avenue of roadside trees in a large city in Germany (Schultz & Steindl, 2018). In the newly discovered site of *S. amabilis* in Poland, several rare calicioid lichens (including *Chaenotheca brachypoda* (Ach.) Tibell, *C. stemonea* (Ach.) Müll. Arg., and *C. phaeocephala* (Turner) Th. Fr.) were noted (on willows growing nearby), which are mentioned by some authors as good indicators of ecological continuity (Cieśliński et al., 1996). It can be assumed that *S. amabilis* is a moderately hemerophilic species, perhaps spreading due to global warming.

Sclerophora amabilis was originally described from New Zealand (Tibell, 1982, 1984), but has also been discovered in North America (Goward et al., 1996) and several countries in Europe and East Asia (Tibell, 1999; Tibell & Thor, 2003). In the Baltic See Region, the species has been found in Denmark, Norway, Sweden (Tibell, 1999), Germany (Schultz & Steindl, 2018), Latvia (Plocina, 2007), Estonia (Oja et al., 2016), and European part of Russia (Muchnik & Konoreva, 2017). According to Tibell (1994), it is an antitropical species that is widely distributed in the cool temperate to temperate areas of both hemispheres. Its northernmost position (66°53'N) has recently been found in the Murmansk region (Urbanavichus & Urbanavichene, 2021).

Specimen examined. Poland. Kotlina Toruńska. Otłoczyn, Szlak Bursztynowy str., 52.920194°N, 18.719222°E, elev. 50 m, ATPOL grid square Cd-41, on *Acer negundo*, 22 May 2011, D. Kubiak s.n. (OLTC L-3018).

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REFERENCES

- Abbott, B. F. M. 2009. Checklist of the lichens and lichenicolous fungi of Greece. *Bibliotheca Liche*nologica 103: 1–368.
- Aptroot, A., van Herk, C. M., Sparrius, L. B. & Spier J. L. 2004. Checklist van de Nederlandse korstmossen en korstmosparasieten. *Buxbaumiella* 69: 17–55.
- Aptroot, A., Van de Vijver, B., Lebouvier, M. & Ertz, D. 2011. Lichens of Ile Amsterdam and

Ile Saint Paul (TAAF, southern Indian Ocean). *Nova Hedwigia* 92(3-4): 343-367. https://doi. org/10.1127/0029-5035/2011/0092-0343

- Arup, U. 2006. A new taxonomy of the Caloplaca citrina group in the Nordic countries, except Iceland. Lichenologist 38(1): 1–20. https://doi.org/10.1017/ S0024282905005402
- Arup, U. 2009. The Caloplaca holocarpa group in the Nordic countries, except Iceland. Lichenologist 41(2): 111–130. https://doi.org/10.1017/ S0024282909008135
- Arup, U., Søchting, U. & Frödén, P. 2013. A new taxonomy of the family Teloschistaceae. Nordic Jorunal of Botany 31(1): 16–83. https://doi. org/10.1111/j.1756-1051.2013.00062.x
- Brodo, I. M. 1984. The North American species of the Lecanora subfusca group. Beiheft zur Nova Hedwigia 79: 63–185.
- Cieśliński, S., Czyżewska, K., Faliński, J. B., Klama, H., Mułenko, W. & Żarnowiec, J. 1996. Relicts of the primeval (virgin) forest. Relict phenomena. In: Faliński, J. B. & Mułenko W. (eds). Cryptogamous plants in the forest communities of Białowieża National Park (Project CRYPTO 3). *Phytocoenosis* 8 (N.S.), Archivum Geobot. 6: 197–216.
- Cieśliński, S. & Fałtynowicz, W. 1993. Note from editors. In: Cieśliński, S. & Fałtynowicz, W. (eds). Atlas of the geographical distribution of lichens in Poland. 1. W. Szafer Institute of Botany, Polish Academy of Sciences, Kraków. 7–8 pp.
- Coppins, B. J. & James, P. W. 1979. New or interesting British lichens IV. *Lichenologist* 11(2): 139–179. https://doi.org/10.1017/S0024282979000190
- Czarnota, P. & Guzow-Krzemińska, B. 2018. Bacidina mendax sp. nov., a new widespread species in Central Europe, together with a new combination within the genus Bacidina. Lichenologist 50(1): 43–57. https://doi.org/10.1017/ S0024282917000627
- de Bruyn, U., Aptroot, A. & van Herk, K. 2000. Lichenized and lichenicolous fungi new to the flora of North West Germany. *Herzogia* 14: 218–221. https://doi.org/10.1127/herzogia/14/2000/218
- Diederich, P., Ertz, D., Eichler, M., Cezanne, R., van den Boom, P., Fischer, E., Killmann, D., Van den Broeck, D. & Sérusiaux, E. 2012. New or interesting lichens and lichenicolous fungi from Belgium, Luxembourg and northern France. XIV. Bulletin de la Société des Naturalistes Luxembourgeois 113: 95–115.
- Ertz, D., Sanderson, N., Łubek, A. & Kukwa, M. 2018. Two new species of Arthoniaceae from old-growth European forests, Arthonia thoriana and Inoderma sorediatum, and a new genus for Schismatomma niveum. Lichenologist 50(2): 161–172. http:// dx.doi.org/10.1017/S0024282917000688
- Esslinger, T. L. 2019. A cumulative checklist for the lichen-forming, lichenicolous and allied fungi of the continental United States and Canada, version 23. *Opuscula Philolichenum* 18: 102–378.

- Fałtynowicz, W. & Kossowska, M. 2016. The lichens of Poland. A fourth checklist. Acta Botanica Silesiaca, Monographiae 8: 1–121.
- Fletcher, A. & Laundon, J. R. 2009. Caloplaca Th. Fr. (1860). In: Smith, C. W., Aptroot, A., Coppins, B. J., Fletcher, A., Gilbert, O. L., James, P. W. & Wolseley, P. A. (eds). The lichens of Great Britain and Ireland. British Lichen Society, London, pp. 245–273.
- Galloway, D. J. 2007. Flora of New Zealand lichens. Revised second edition including lichen-forming and lichenicolous fungi. Volume 1. Manaaki Whenua Press, Lincoln. 1006 pp.
- Goward, T., Breuss, O., Ryan, B., McCune, B., Sipman, H. & Scheidegger, C. 1996. Notes on the lichens and allied fungi of British Columbia. III. *Bryologist* 99(4): 439–449. https://doi. org/10.2307/3244108
- Guzow-Krzemińska, B., Łubek, A., Kubiak, D., Ossowska, E. & Kukwa, M. 2018. Phylogenetic approaches reveal a new sterile lichen in the genus *Loxospora* (Sarrameanales, Ascomycota) in Poland. *Phytotaxa* 348(3): 211–220. http:// dx.doi.org/10.11646/phytotaxa.348.3.4
- Guzow-Krzemińska, B., Malíček, J., Tønsberg, T., Oset, M., Łubek, A. & Kukwa, M. 2017. Lecanora stanislai, a new, sterile, usnic acid containing lichen species from Eurasia and North America. Phytotaxa 329(3): 201–211. http://dx.doi. org/10.11646/phytotaxa.329.3.1
- Guzow-Krzemińska, B., Sérusiaux, E., van den Boom, P. P. G., Brand, A. M., Launis, A., Łubek, A. & Kukwa, M. 2019. Understanding the evolution of phenotypical characters in the *Micarea prasina* group (Pilocarpaceae) and descriptions of six new species within the group. *MycoKeys* 57: 1–30. https://doi.org/10.3897/mycokeys.57.33267
- Jüriado, I. 1998. A revision of the Lecanora subfusca group in Estonia. Folia Cryptogamica Estonica 32: 15–20.
- Kondratyuk, S. Y., Kärnefelt, I., Lőkös, L., Hur, J.-S. & Thell, A. 2018. *Coppinsiella* and *Seawardiella* – two new genera of the Xanthorioideae (Teloschistaceae, lichen-forming ascomycota). *Acta Botanica Hungarica* 60(3–4): 369–386. https://doi. org/10.1556/034.60.2018.3-4.8
- Kubiak, D. & Wilk, K. 2016. Caloplaca monacensis (Teloschistaceae, lichenized Ascomycota), a species new to Poland. Polish Botanical Journal 61(2): 279– 281. https://doi.org/10.1515/pbj-2016-0032
- Kukwa, M., Czarnota, P. & Łubek, A. 2017. Three lichen species in *Buellia*, *Catillaria*, and *Cheiromycina*, new to Poland. *Mycotaxon* 132: 177–182. https://doi.org/10.5248/132.177
- Liška, J., Palice, Z. & Slavíková, Š. 2008. Checklist and Red List of lichens of the Czech Republic. *Preslia* 80: 151–182.
- Llimona, X. & Hladun, N. L. 2001. Checklist of the lichens and lichenicolous fungi of the Iberian Peninsula and Balearic Islands. *Bocconea* 14: 1–581.

- Malíček, J. 2014. A revision of the epiphytic species of the *Lecanora subfusca* group (Lecanoraceae, Ascomycota) in the Czech Republic. *Lichenologist* 46(4): 489–513. https://doi.org/10.1017/ S0024282914000139
- Malíček, J., Palice, Z. & Vondrák, J. 2014. New lichen records and rediscoveries from the Czech Republic and Slovakia. *Herzogia* 27(2): 257–284. https:// doi.org/10.13158/heia.27.2.2014.257
- Matura, N., Krzewicka, B. & Flakus, A. 2017. Seven species of freshwater lichen-forming fungi newly recorded from Poland. *Polish Botanical Journal* 62(2): 273–278.https://doi.org/10.1515/pbj-2017-0029
- Muchnik, E. & Konoreva, L. 2017. New and noteworthy records of lichens and allied fungi from central European Russia. *Herzogia* 30(2): 509–514. https://doi.org/10.13158/heia.30.2.2017.509
- Nimis, P. L. 2016. The lichens of Italy. A second annotated catalogue. Edizioni Università di Trieste, Trieste. 739 pp.
- Oja, E., Gerasimova, J., Suija, A., Lõhmus, P. & Randlane, T. 2016. New Estonian records and amendments: Lichenized fungi. *Folia Cryptogamica Estonica* 53: 123–126. https://doi.org/10.12697/ fce.2016.53.14
- Ossowska, E., Guzow-Krzemińska, B., Szymczyk, R. & Kukwa, M. 2021. A molecular re-evaluation of *Parmelia encryptata* with notes on its distribution. *Lichenologist* 53(4): 341–345. https://doi. org/10.1017/S0024282921000219
- Plocina, J. 2007. Monitoring of woodland meadow management using epiphytic lichens. In: 4th International Conference "Research and Conservation of Biological Diversity in Baltic Region" Daugavpils, 25-27 April, 2007 Book of Abstracts. Daugavpils University Academic Press "Saule", Daugavpils, p. 88.
- Randlane, T. & Saag, A. (eds) 1999. Second checklist of lichenized, lichenicolous and allied fungi of Estonia. Folia Cryptogamica Estonica 35: 1–132.
- Schultz, M. & Steindl, P. 2018. Erstnachweis von Sclerophora amabilis in Deutschland [First record of Sclerophora amabilis in Germany]. Herzogia 31(1): 317-321. https://doi. org/10.13158/099.031.0126
- Seaward, M. R. D., Sipman, H. J. M. & Sohrabi, M. 2008. A revised checklist of lichenized, lichenicolous and allied fungi for Iran. Sauteria 15: 459–520.
- Søchting, U. & Alstrup, V. 2008. Danish lichen checklist. Faculty of Science, University of Copenhagen, Copenhagen. 46 pp.
- Szczepańska, K. 2020. Carbonea assimilis and Rinodina aspersa, new to Poland. Mycotaxon 135(2): 355–363. https://doi.org/10.5248/135.355
- Szczepańska, K., Rodriguez-Flakus, P., Urbaniak, J. & Śliwa, L. 2019. Neotypification of *Protoparmeliopsis garovaglii* and molecular evidence of its occurrence in Poland and South America.

MycoKeys 57: 31–46. https://doi.org/10.3897/ mycokeys.57.34501

- Thor, G. & Nascimbene, J. 2010. An annotated checklist and bibliography of lichens and lichenicolous fungi of Libya. *Cryptogamie Mycologie* 31(1): 67–95.
- Tibell, L. 1982. Caliciales Exsiccatae. Fasc. III (No. 51–75). Publications from the Herbarium, University of Uppsala, Sweden 10: 1–10. https://doi. org/10.1017/S0024282981000066
- Tibell, L. 1984. A reappraisal of the taxonomy of Caliciales. *Beihefte zur Nova Hedwigia* 79: 597–713.
- Tibell L. 1994. Distribution patterns and dispersal strategies of Caliciales. *Botanical Journal of the Linnean Society* 116: 159–202. https://doi. org/10.1006/bojl.1994.1059
- Tibell, L. 1999. Calicioid lichens and fungi. Nordic Lichen Flora 1: 20-94. https://doi. org/10.1111/j.1756-1051.2000.tb00759.x
- Tibell, L. & Thor, G. 2003. Calicioid lichens and fungi of Japan. *Journal of the Hattori Botanical Laboratory* 94: 205–259.
- Urbanavichus, G. 2010. A checklist of the lichen flora of Russia. Nauka, St. Petersburg. 194 pp.
- Urbanavichus, G. & Urbanavichene, I. 2021. New records of lichens and lichenicolous fungi from Murmansk Region, Russia. *Folia Cryptog. Estonica* 58: 35–40. https://doi.org/10.12697/fce.2021.58.04
- Van den Broeck, D., Aptroot, A. & Jordaens, D. 2006. Een lichenologisch verslag van het voorjaarsweekend 2006 naar Zeeuws-Vlanderen en aangrenzend België. *Buxbaumiella* 75: 16–25.
- van Herk, C. M. & Aptroot, A. 1999. Lecanora compallens and L. sinuosa, two new overlooked corticolous lichen species from Western Europe. Lichenologist 31(6): 543–553. https://doi.org/10.1006/ lich.1999.0216
- Vězda, A. 1990. Lichenes selecti exsiccati, Fasc. 99. Průhonice: Instituto Botanico Academiae Scientiarum Čechoslovacae.
- Vondrák, J. & Liška, J. 2013. Lichens and lichenicolous fungi from the Retezat Mts and overlooked records for the checklist of Romanian lichens. *Herzogia* 26(2): 293–305. https://doi.org/10.13158/ heia.26.2.2013.293
- Vondrák, J., Šoun, J., Arup, U., Aptroot, A. & Redchenko, O. 2009a. *Caloplaca ulcerosa*, a maritime species in Europe with a remarkable occurrence in the Czech Republic. *Bryonora* 1–7.
- Vondrák, J., Říha, P., Arup, U. & Søchting, U. 2009b. The taxonomy of the *Caloplaca citrina* group (*Theloschistaceae*) in the Black Sea region; with contributions to the cryptic species concept in lichenology. *Lichenologist* 41(6): 571–604. https:// doi.org/10.1017/S0024282909008317
- Vondrák, J., Frolov, I., Øíha, P., Hrouzek, P., Palice, Z., Nadyeina, O., Halıcı, G., Khodosovtsev, A. & Roux, C. 2013. New crustose *Teloschistaceae* in Central Europe. *Lichenologist* 45(6): 701–722. https://doi.org/10.1017/S0024282913000455

- Westberg, M., Moberg, R., Myrdal, M., Nordin, A. & Ekman, S. 2021. Santesson's checklist of Fennoscandian lichen-forming and lichenicolous fungi. Museum of Evolution, Uppsala University, Uppsala. 933 pp.
- Wetmore, C. M. 2005. The sorediate, corticolous species of *Caloplaca* in North and Central America. *Bryologist* 107(4): 505–520. https:// doi.org/10.1639/0007-2745(2004)107[505:TSC SOC]2.0.CO;2
- Wetmore, C. M. 2009. New species of Caloplaca (Teloschistaceae) from North America. Bryologist 112(2): 379–386. https://doi.org/10.1639/0007-2745-112.2.379
- Wilk, K. 2011. New or noteworthy records of Caloplaca (Theloschistaceae) from Poland. Mycotaxon 115: 83–98. https://doi.org/10.5248/115.83
- Wirth, V., Hauck, M. & Shultz, M. 2013. Die Flechten Deutschlands. Band 1. Eugen Ulmer, Stuttgart. 672 pp.
- Wijayawardene, N. N., Hyde, K. D., Dai, D. Q., Sánchez-García, M., Goto, B. T., Saxena, R. K., ... & Thines, M. 2022. Outline of fungi and fungus-like taxa-2021. *Mycosphere* 13(1), 53–453. https://10.5943/mycosphere/13/1/2
- Zając, A. 1978. Atlas of distribution of vascular plants in Poland (ATPOL). *Taxon* 27: 1–481. https://doi. org/10.2307/1219899