REVIEW



Bevacizumab-Induced Hypertension in Glioblastoma Patients and Its Potential as a Modulator of Treatment Response

Kaitlin G. Scheer[®], Lisa M. Ebert, Michael S. Samuel[®], Claudine S. Bonder[®], Guillermo A. Gomez[®]

ABSTRACT: Glioblastoma invasion is the primary mechanism responsible for its dismal prognosis and is the direct result of interactions between glioblastoma cells and the tumor vasculature. The dysregulated microvasculature in glioblastoma tumors and vessels co-opted from surrounding brain tissue support rapid tumor growth and are utilized as pathways for invasive cancer cells. Attempts to target the glioblastoma vasculature with antiangiogenic agents (eg, bevacizumab) have nonetheless shown limited and inconsistent efficacy, and the underlying causes of such heterogeneous responses remain unknown. Several studies have identified that patients with glioblastoma who develop hypertension following treatment with bevacizumab show significant improvement in overall survival compared with normotensive nonresponders. Here we review these findings and discuss the potential of hypertension as a biomarker for glioblastoma treatment response in individual patients and the role of hypertension as a modulator of interactions between tumor cells and cells in the perivascular niche. We suggest that a better understanding of the actions of bevacizumab and hypertension at the cellular level will contribute to developing more effective personalized therapies that address glioblastoma tumor cell invasion.

Key Words: bevacizumab biomarkers brain glioblastoma hypertension invasion

G lioblastoma is the most common and aggressive form of brain cancer, with <5% of patients achieving 5-year survival.¹² Despite concerted research efforts over the past 3 decades, overall survival (OS) for glioblastoma has increased by only 3 months, extending median survival from 12 to 15 months following introduction of the Stupp protocol in 2005.³⁴ This protocol remains the standard of care for patients with glioblastoma and combines maximal surgical resection of the primary tumor and postoperative radiotherapy with concurrent Temozolomide administration.⁴ Unfortunately, patients treated with the Stupp protocol invariably experience therapy resistant tumor recurrence.⁵

Interactions between cancer cells and the microenvironment, particularly those cells comprising the vessels within the tumor, contribute to therapy resistance.⁶

Glioblastoma vascularization is a prognostic marker predictive of patient survival, with increased tumor microvessel density correlating with poorer OS.7-9 Angiogenesis in glioblastoma is predominantly attributed to the upregulation of VEGFA (vascular endothelial growth factor A), which stimulates endothelial cell (EC) survival, proliferation, and migration as well as EC progenitor differentiation.¹⁰⁻¹² Of note, the existing brain vasculature is insufficient to sustain rapidly growing glioblastoma, which inevitably results in the generation of hypoxic gradients within the tumour.¹³ Hypoxia-induced VEGFA expression subsequently stimulates the formation of new blood vessels13,14 (Figure 1A), which are tortuous, leaky, and poorly structured in comparison to vessels in the healthy brain.¹⁵ Thus, although glioblastoma tumors are highly

For Sources of Funding and Disclosures, see page XXX.

Correspondence to: Kaitlin G. Scheer, Centre for Cancer Biology, SA Pathology and the University of South Australia, South Australia, Email kaitlin.scheer@mymail.unisa. edu.au or Guillermo A. Gomez, Centre for Cancer Biology, SA Pathology and the University of South Australia, South Australia, Email guillermo.gomez@unisa.edu.au *M. Samuel and C. Bonder contributed equally.

Supplemental Material is available at https://www.ahajournals.org/doi/suppl/10.1161/HYPERTENSIONAHA.123.21119.

^{© 2023} The Authors. *Hypertension* is published on behalf of the American Heart Association, Inc., by Wolters Kluwer Health, Inc. This is an open access article under the terms of the Creative Commons Attribution Non-Commercial-NoDerivs License, which permits use, distribution, and reproduction in any medium, provided that the original work is properly cited, the use is noncommercial, and no modifications or adaptations are made.

Hypertension is available at www.ahajournals.org/journal/hyp

Nonstandard Abbreviations and Acronyms

EC	endothelial cell
eNOS	endothelial NO synthase
NO	nitric oxide
NOX1	NADPH oxidase 1
OS	overall survival
ROS	reactive oxygen species
VEGFA	vascular endothelial growth factor A

vascularized, the poorly formed neovessels that originate within the tumors do not alleviate hypoxia. As a result, cancer cells in hypoxic regions of the tumor often switch from a proliferative to a more migratory and invasive mesenchymal phenotype.^{16,17} These cells then not only escape and migrate away from hypoxic regions by invading the surrounding healthy tissue but also undergo transcriptional changes that additionally increase their resistance to therapy.¹⁸

Blockade of specific vascular development pathways with bevacizumab, a VEGFA inhibitor, was anticipated to be an effective treatment for glioblastoma.¹⁹ However, while bevacizumab showed promise in early clinical trials, it has not consistently yielded an OS benefit for patients.²⁰ Indeed, treatment with bevacizumab is associated with increased tumor invasion.^{21–23} Intriguingly, a subset (\approx 40%) of patients who develop hypertension following bevacizumab treatment exhibit significant survival benefits compared with normotensive patients.^{24–27}

Despite the limited success and its association with an invasive phenotype, bevacizumab is being broadly adopted to treat recurrent glioblastoma. This reliance on bevacizumab emphasizes the importance of unravelling the mechanisms contributing to disparate patient responses to this agent. In particular, understanding the role of tumorvessel interactions and the development of hypertension is likely to be critical for identifying glioblastoma patients who benefit the most from receiving bevacizumab.

HYPERTENSION AS A BIOMARKER OF RESPONSE TO BEVACIZUMAB TREATMENT IN GLIOBLASTOMA

Antiangiogenic agents were tested with the goals of normalizing aberrant vasculature (and thus reducing edema), disrupting the glioblastoma perivascular niche, and improving access of other chemotherapeutics to the tumor.²⁸ A large majority of antiangiogenic compounds tested for efficacy against glioblastoma target VEGFA signaling. This is through inhibition of VEGFA ligand, its principal receptor VEGFR2/KDR, or downstream signaling molecules.²⁰ For a detailed review of antiangiogenic drugs trialed for glioblastoma, see Anthony et al.²⁰ The most extensively studied drug, bevacizumab, is a recombinant humanized monoclonal antibody that binds VEGFA, preventing receptor interaction and blocking VEGFA-mediated proangiogenic signaling.²⁹ In 2009, the United States Food and Drug Administration approved bevacizumab treatment for recurrent glioblastoma based on the promising results in 2 phase II clinical trials.³⁰ Elsewhere, applications for bevacizumab treatment of glioblastoma met with resistance due to insufficient evidence of efficacy.^{31,32} Approval was eventually granted in some countries^{33,34} and it is used off-label in others,³⁵ despite limited evidence of consistent or substantial efficacy.^{32,34} The improvement in progression-free survival commonly reported over the preceding 10 years of clinical trials is now referred to as a pseudoresponse.³⁶ This is because bevacizumab stabilizes the vasculature by inhibiting VEGFA-induced vessel permeabilization,³⁷ and thereby restores blood-brain barrier integrity, leading to skewed magnetic resonance imaging readouts due to marked decreases of hyperperfusion, edema, and contrast enhancement.36

The most reported adverse event following administration of bevacizumab is hypertension (grade 2-3), which affects ≈40% of treated patients^{24-27,38,39} (Figure 1B). Interestingly, with the exception of 1 small prospective study,38 patients who developed hypertension following bevacizumab treatment demonstrated prolonged progression-free survival and OS, with statistically significant increases in OS ranging from 5 to 9 months compared with normotensive patients.24-27 Furthermore, bevacizumab-induced hypertension was shown by multivariate analysis to independently predict improved OS.^{24-27,39} It is, therefore, possible that the inconsistencies of reported bevacizumab efficacy across different clinical trials may be a result of the large proportion of nonresponders statistically masking the benefit experienced by the smaller cohort of responders. Given these findings, it is now urgent to identify which patients will derive the most benefit from bevacizumab.

BEVACIZUMAB AND HYPERTENSION CAUSE MICROVASCULAR DEFECTS

It is thought that the vascular changes induced by bevacizumab confer the survival benefit in responsive patients, with these changes causing hypertension in many patients.⁴⁰ As bevacizumab causes a reduction in both vascular density (rarefaction) and production of the vasodilator nitric oxide (NO) production,⁴¹ these are suspected to be causal factors in hypertension pathogenesis following treatment.^{42,43} Microvascular rarefaction is also common early in the development of hypertension⁴⁴; however, it is unclear to what extent this contributes to the onset of increased blood pressure. Similarly, EC dysfunction and vascular constriction due to reduced availability of NO, an essential vasodilator, can be both cause

REVIEW



Figure 1. Glioblastoma microvasculature and the correlation between the development of hypertension and survival for glioblastoma patients treated with bevacizumab.

A, Glioblastomas commonly feature regions of hypoxia and necrosis. Tumor cells in the hypoxic areas express VEGFA (vascular endothelial growth factor A), inducing the formation of numerous new poorly formed and leaky blood vessels. The blood-brain barrier is often disrupted in the central tumor, causing edema and hemorrhage, while on the tumor periphery, this barrier variably remains intact. **B**, Roughly 4 of every 10 patients treated with bevacizumab will develop hypertension following treatment. Patients who develop hypertension have improved overall survival. This figure was created using BioRender.com.

and effect of hypertension.^{45–47} Under normal conditions, VEGFA stimulates activation and upregulation of eNOS (endothelial NO synthase), subsequently inducing or increasing NO release^{42,48} (Figure 2). Conversely, upregulation of angiotensin-II in hypertension causes increased production of reactive oxygen species (ROS) such as superoxide via NOX1 (NADPH oxidase 1), which then combines with NO to form peroxynitrite (ONOO⁻).^{45,47,49} Besides this reducing available NO, peroxynitrite destabilizes eNOS, inducing further superoxide production rather than NO in a feed-forward loop, which drives a reduction in the bioavailability of vasodilators alongside an increase

in constricting factors, a state known as endothelial dysfunction.^{45,47,49} Therefore, while bevacizumab reduces NO release through VEGFA inhibition, hypertension likely perturbs several alternate NO regulating pathways alongside increased ROS production,⁴⁵ thus likely increasing EC dysfunction and impairing the tumor vasculature.

It is, therefore, possible that hypertension contributes further to microvascular rarefaction independently of bevacizumab. Furthermore, development of hypertension may indicate sensitivity to endothelial dysfunction, whereas in other patients inhibition of VEGFA is insufficient to cause dysfunction-induced hypertension. REVIEW



Figure 2. Modulators of vascular tone and endothelial dysfunction in the perivascular niche.

Vascular tone is maintained by a careful balance of vasodilators and vasoconstrictors. VEGFA (vascular endothelial growth factor A) induces upregulation and activation of eNOS (endothelial nitric oxide synthase), which generates the key vasodilator NO. Inhibition of VEGFA signaling by bevacizumab reduces NO production. Under conditions of hypertension, angiotensin-II (Ang-II) signaling induces production of superoxide by NOX1 (NADPH oxidase 1), which combines with available NO to form peroxynitrite. This reactive oxygen species (ROS) then destabilizes eNOS by reducing the cofactor tetrahydrobiopterin (BH₄), causing eNOS to switch to production of superoxide, driving endothelial dysfunction through the loss of NO. This figure was created using BioRender.com.

Indeed, certain eNOS polymorphisms have been linked with significantly elongated progression-free survival for patients with metastatic colorectal cancer treated with bevacizumab, with a trend between polymorphisms associated with improved outcome and patients who experienced higher grades of hypertension.⁵⁰ However, little is known about whether hypertension itself could influence glioblastoma progression. Two studies showed no difference in survival between normotensive and hypertensive patients,^{51,52} which contrasts with a recent unbiased mortality study that showed a decreased risk of any cancer death in hypertensive participants.⁵³ Letourneur and colleagues⁵⁴ investigated the effects of chronic hypertension on glioma growth in a spontaneously hypertensive rat model. Intriguingly, tumors progressed slower and were significantly smaller at the experiment end point in spontaneously hypertensive rat compared with control Wistar-Kyoto rats.54 This was similar to the reduction in tumor size observed in rodents treated with antiangiogenic agents.55,56 The authors postulated that slower glioma growth in spontaneously hypertensive rat may be due to the known effects of hypertension on the vascular system, such as inhibition of angiogenesis.54,57 Microfluidic in vitro models of angiogenesis have demonstrated that neovessel growth is stimulated by increased interstitial flow on the basal side of EC monolayers.58-60 In contrast, flow in the direction of sprouting (apical/luminal side) was conversely shown to inhibit angiogenesis and even cause vessel regression.⁵⁹ More recently, interstitial flow was confirmed to induce angiogenesis by

mechanotransduction independent of VEGFA, although physiologically normal network formation required a balance between the 2.⁵⁸ Taken together, these results suggest that increased hydrostatic pressure against the lumen of blood vessels can modulate angiogenesis independently of VEGFA blockade by bevacizumab.

INTRATUMOR HETEROGENEITY DRIVEN BY CHANGES IN TUMOR CELL TRANSCRIPTIONAL STATES IS RELIANT ON A DELICATE BALANCE OF NO AND ROS

While control of rampant blood vessel growth reduces the availability of nutrients to the rapidly growing tumor, interactions between tumor and vascular cells within perivascular niches will remain, and these are of critical importance in other aspects of glioblastoma biology. Glioblastoma cells exhibiting stem-like properties have commonly been shown to reside in close contact with brain vasculature within a perivascular niche.^{20,61,62} Much like neural stem cell niches,63 these are spatially distinct microcompartments, where heterotypic interactions of stem-like glioblastoma cells with the surrounding microenvironment and nontumor cells can induce stem cell maintenance or differentiation and clonal selection.^{18,64} ECs provide cues to maintain stemness and self-renewal of tumor cells in the perivascular space, in addition to facilitating protection from radiation.62,65-68 Indeed, a high degree of cancer cell plasticity has been reported

to be a feature of glioblastoma, with cancer cells shifting between different transcriptional states depending on the local tumor microenvironment.^{18,64} Reference to glioblastoma cells in this review should thus be considered to encompass various subsets of tumor cell including those typically referred to as glioblastoma stem cells.

The vascular niches occupied by glioblastoma cells are suggested to be predominantly arteriolar,69 in a relatively hypoxic microenvironment with low levels of ROS, conditions known to maintain cancer cell stemness and self-renewal.⁷⁰ These conditions have been likened to those in adult stem cell niches^{69,71} and a delicate balance of hypoxia/ROS is indeed critical for stem cell maintenance. Quiescent or self-renewing stem cells display low levels of ROS, intermediate levels of ROS induce stem cell differentiation, and high ROS levels cause senescence and cell death.⁷² In the case of glioblastoma cancer cells, induction of intracellular ROS reduces tumorigenicity, decreases capacity for self-renewal, and induces differentiation.73 NO is also an important regulator shown to variably maintain cancer cell stemness, enhance invasiveness, or induce apoptosis under different conditions.^{66,74,75} In patient biopsies, eNOS is upregulated in ECs adjacent to perivascular glioblastoma cells expressing nestin, a stem cell marker.⁶⁶ PDGF-induced gliomas in mice with reduced NO due to eNOS knockdown had delayed tumor growth and improved survival.⁶⁶ Since this early study, a large volume of research has consolidated REVIEW

NO synthases in glioblastoma as potential therapeutic targets.⁷⁵ As previously mentioned, hypertension causes downregulation of NO with concomitant increases in ROS production, eNOS destabilization, and EC dysfunction (Figure 3).^{45–47,49} Whether hypertension may disrupt the perivascular glioblastoma cell niche by perturbing the balance of ROS and NO remains to be elucidated.

CONCLUSIONS

The prevailing wisdom is that hypertension may be a useful marker of patient responsiveness to bevacizumab to determine whether therapy is worth continuing. Here, we have reviewed the literature on the relationship between glioblastoma cancer-related outcomes and the development of hypertension after VEGFA inhibition using bevacizumab. So far, multiple reports suggest a positive correlation between the development of hypertension and a favorable tumor response to bevacizumab treatment. However, the causality of this relationship needs to be confirmed as, to date, no study has demonstrated whether effects mediated by hypertension increase the efficacy of these drugs or ameliorate adverse effects. This is an important distinction, because if hypertension improves treatment outcome, this may alter how hypertension is managed in patients treated with bevacizumab, particularly given the evidence for increased microvascular density and heightened risk of



Figure 3. VEGFA (vascular endothelial growth factor A) inhibition by bevacizumab normalizes vascular tone by reducing endothelial nitric oxide synthase (eNOS) production of NO.

Certain patients experience such a change in peripheral resistance due to capillary rarefaction and increase in vascular tone that they develop hypertension following treatment. These patients may experience further endothelial dysfunction due to the compounding effects of hypertension on NO availability. This figure was created using BioRender.com.

(any) cancer-related death following effective control of hypertension with antihypertensives.^{76,77} Furthermore, discovery of how hypertension affects survival may identify patients who will most benefit from bevacizumab treatment, or pathways that could be targeted in combination with bevacizumab.

No significant investigative effort has yet been made into the role of hypertension in glioblastoma, and extrapolation from currently available data is difficult due to continuing controversy in the literature. However, evidence gleaned from studies of hypertension demonstrate its potential impact on glioblastoma invasion-as discussed in the Supplemental Text^{21-23,55,56,78-106}-and stem cell maintenance, which could enhance patient response to bevacizumab. In this regard, the recent development of spatial omics approaches (spatial transcriptomics¹⁰⁷ and proteomics)¹⁰⁸ applied to patient tumor tissue (resected or postmortem) could help to elucidate the relationships between the onset of hypertension and response, including the potential influence of the immune system in hypertension development. Furthermore, it may be possible to identify new biomarkers that can in the future be used for the stratification of patients likely to respond to bevacizumab treatment, at the time of diagnosis.

ARTICLE INFORMATION

Affiliations

Centre for Cancer Biology, SA Pathology and the University of South Australia (K.G.S., L.M.E., M.S.S., C.S.B., G.A.G.). Adelaide Medical School, University of Adelaide, South Australia (L.M.E., C.S.B.). Royal Adelaide Hospital, Adelaide, South Australia (L.M.E.).

Acknowledgments

The authors apologize for not citing the work of many authors who contributed to the subject due to space limitations.

Sources of Funding

This work was supported by a McCleary Murchland Fellowship (to G. Gomez); grants from the NHMRC (Ideas grant 2021/GNT2013180) to G. Gomez and L. Ebert; the Cure Brain Cancer Foundation (to G. Gomez); the Neurosurgical Research Foundation (to L. Ebert and G. Gomez); the Cancer Council SA Beat Cancer Project (to G. Gomez); the Charlie Teo Foundation (Rebel Grant to G. Gomez), and a NRF Chris Adams award and a UniSA Research Training Program Scholarship to K. Scheer.

Disclosures

None.

Supplemental Material

Supplemental text Figure S1 References 78–106

REFERENCES

- Australian Institute of Health and Welfare. Brain and other central nervous system cancers. 2017.
- Cantrell JN, Waddle MR, Rotman M, Peterson JL, Ruiz-Garcia H, Heckman MG, Quinones-Hinojosa A, Rosenfeld SS, Brown PD, Trifiletti DM. Progress toward long-term survivors of glioblastoma. *Mayo Clin Proc.* 2019;94:1278–1286. doi: 10.1016/j.mayocp.2018.11.031
- Ostrom QT, Bauchet L, Davis FG, Deltour I, Fisher JL, Langer CE, Pekmezci M, Schwartzbaum JA, Turner MC, Walsh KM, et al. The epidemiology of glioma

in adults: a "state of the science" review. *Neuro-Oncology.* 2014;16:896–913. doi: 10.1093/neuonc/nou087

- Stupp R, Mason WP, van den Bent MJ, Weller M, Fisher B, Taphoorn MJB, Belanger K, Brandes AA, Marosi C, Bogdahn U, et al. Radiotherapy plus concomitant and adjuvant temozolomide for glioblastoma. *N Engl J Med.* 2005;352:987–996. doi: 10.1056/NEJMoa043330
- Stupp R, Hegi ME, Mason WP, van den Bent MJ, Taphoorn MJB, Janzer RC, Ludwin SK, Allgeier A, Fisher B, Belanger K, et al; European Organisation for Research and Treatment of Cancer Brain Tumour and Radiation Oncology Groups. Effects of radiotherapy with concomitant and adjuvant temozolomide versus radiotherapy alone on survival in glioblastoma in a randomised phase iii study: 5-year analysis of the eortc-ncic trial. *Lancet Oncol* 2009;10:459–466. doi: 10.1016/S1470-2045(09)70025-7
- Pietras A, Katz AM, Ekström EJ, Wee B, Halliday JJ, Pitter KL, Werbeck JL, Amankulor NM, Huse JT, Holland EC. Osteopontin-cd44 signaling in the glioma perivascular niche enhances cancer stem cell phenotypes and promotes aggressive tumor growth. *Cell Stem Cell*. 2014;14:357–369. doi: 10.1016/j.stem.2014.01.005
- Leon SP, Folkerth RD, Black PM. Microvessel density is a prognostic indicator for patients with astroglial brain tumors. *Cancer.* 1996;77:362–372. doi: 10.1002/(SICI)1097-0142(19960115)77:2<362::AID-CNCR20>3.0.CO;2-Z
- Puig J, Blasco G, Daunis-i-Estadella J, Alberich-Bayarri A, Essig M, Jain R, Remollo S, Hernández D, Puigdemont M, Sánchez-González J, et al. High-resolution blood-pool-contrast-enhanced mr angiography in glioblastoma: tumorassociated neovascularization as a biomarker for patient survival. A preliminary study. *Neuroradiology.* 2016;58:17–26. doi: 10.1007/s00234-015-1599-0
- Fan C, Zhang J, Liu Z, He M, Kang T, Du T, Song Y, Fan Y, Xu J. Prognostic role of microvessel density in patients with glioma. *Medicine (Baltim)*. 2019;98:e14695–e14695. doi: 10.1097/MD.000000000014695
- Dieterich LC, Mellberg S, Langenkamp E, Zhang L, Zieba A, Salomäki H, Teichert M, Huang H, Edqvist P-H, Kraus T, et al. Transcriptional profiling of human glioblastoma vessels indicates a key role and tgfβ2 in vascular abnormalization. *J Pathol.* 2012;228:378–390. doi: 10.1002/path.4072
- Huang H, Held-Feindt J, Buhl R, Mehdorn HM, Mentlein R. Expression of vegf and its receptors in different brain tumors. *Neurol Res.* 2005;27:371– 377. doi: 10.1179/016164105X39833
- Plate KH, Breier G, Weich HA, Risau W. Vascular endothelial growth factor is a potential tumour angiogenesis factor in human gliomas in vivo. *Nature*. 1992;359:845–848. doi: 10.1038/359845a0
- Kaur B, Khwaja FW, Severson EA, Matheny SL, Brat DJ, Van Meir EG. Hypoxia and the hypoxia-inducible-factor pathway in glioma growth and angiogenesis. *Neuro-Oncology*. 2005;7:134–153. doi: 10.1215/S1152851704001115
- Onishi M, Ichikawa T, Kurozumi K, Date I. Angiogenesis and invasion in glioma. Brain Tumor Pathol. 2011;28:13–24. doi: 10.1007/s10014-010-0007-z
- Kane JR. The role of brain vasculature in glioblastoma. *Mol Neurobiol.* 2019;56:6645-6653. doi: 10.1007/s12035-019-1561-y
- Monteiro AR, Hill R, Pilkington GJ, Madureira PA. The role of hypoxia in glioblastoma invasion. *Cells.* 2017;6:45. doi: 10.3390/cells6040045
- Joseph JV, Conroy S, Pavlov K, Sontakke P, Tomar T, Eggens-Meijer E, Balasubramaniyan V, Wagemakers M, den Dunnen WFA, Kruyt FAE. Hypoxia enhances migration and invasion in glioblastoma by promoting a mesenchymal shift mediated by the hif1α-zeb1 axis. *Cancer Lett.* 2015;359:107– 116. doi: 10.1016/j.canlet.2015.01.010
- Varn FS, Johnson KC, Martinek J, Huse JT, Nasrallah MP, Wesseling P, Cooper LAD, Malta TM, Wade TE, Sabedot TS, et al; GLASS Consortium. Glioma progression is shaped by genetic evolution and microenvironment interactions. *Cell*. 2022;185:2184–2199.e16. doi: 10.1016/j.cell.2022.04.038
- Jain RK, di Tomaso E, Duda DG, Loeffler JS, Sorensen AG, Batchelor TT. Angiogenesis in brain tumours. *Nat Rev Neurosci*. 2007;8:610–622. doi: 10.1038/nrn2175
- Anthony C, Mladkova-Suchy N, Adamson DC. The evolving role of antiangiogenic therapies in glioblastoma multiforme: current clinical significance and future potential. *Expert Opin Investig Drugs.* 2019;28:787–797. doi: 10.1080/13543784.2019.1650019
- Narayana A, Kelly P, Golfinos J, Parker E, Johnson G, Knopp E, Zagzag D, Fischer I, Raza S, Medabalmi P, et al. Antiangiogenic therapy using bevacizumab in recurrent high-grade glioma: impact on local control and patient survival. *J Neurosurg.* 2009;110:173–180. doi: 10.3171/2008.4.17492
- Norden AD, Young GS, Setayesh K, Muzikansky A, Klufas R, Ross GL, Ciampa AS, Ebbeling LG, Levy B, Drappatz J, et al. Bevacizumab for recurrent malignant gliomas: efficacy, toxicity, and patterns of recurrence. *Neurol*ogy. 2008;70:779–787. doi: 10.1212/01.wnl.0000304121.57857.38
- Ribatti D, Annese T, Ruggieri S, Tamma R, Crivellato E. Limitations of antiangiogenic treatment of tumors. *Transl Oncol.* 2019;12:981–986. doi: 10.1016/j.tranon.2019.04.022

- Lombardi G, Zustovich F, Farina P, Fiduccia P, Della Puppa A, Polo V, Bertorelle R, Gardiman MP, Banzato A, Ciccarino P, et al. Hypertension as a biomarker in patients with recurrent glioblastoma treated with antiangiogenic drugs: a single-center experience and a critical review of the literature. *Anticancer Drugs.* 2013;24:90–97. doi: 10.1097/CAD.0b013e32835aa5fd
- Carvalho B, Lopes RG, Linhares P, Costa A, Caeiro C, Fernandes AC, Tavares N, Osório L, Vaz R. Hypertension and proteinuria as clinical biomarkers of response to bevacizumab in glioblastoma patients. *J Neurooncol.* 2020;147:109–116. doi: 10.1007/s11060-020-03404-z
- Zhong J, Ali AN, Voloschin AD, Liu Y, Curran WJ Jr, Crocker IR, Shu H-KG. Bevacizumab-induced hypertension is a predictive marker for improved outcomes in patients with recurrent glioblastoma treated with bevacizumab. *Cancer*. 2015;121:1456–1462. doi: 10.1002/cncr.29234
- Liau C-T, Chou W-C, Wei K-C, Chang C-N, Toh C-H, Jung S-M. Female sex, good performance status, and bevacizumab-induced hypertension associated with survival benefit in asian patients with recurrent glioblastoma treated with bevacizumab. *Asia-Pacific Journal of Clinical Oncology*. 2018;14:e8–e14. doi: 10.1111/ajco.12747
- Kerbel RS. Antiangiogenic therapy: a universal chemosensitization strategy for cancer? *Science*. 2006;312:1171–1175. doi: 10.1126/science.1125950
- Ferrara N, Hillan KJ, Novotny W. Bevacizumab (avastin), a humanized antivegf monoclonal antibody for cancer therapy. *Biochem Biophys Res Commun.* 2005;333:328–335. doi: 10.1016/j.bbrc.2005.05.132
- Cohen MH, Shen YL, Keegan P, Pazdur R. Fda drug approval summary: bevacizumab (avastin
 [®]) as treatment of recurrent glioblastoma multiforme. *The Oncologist* 2009;14:1131–1138. doi: 10.1634/theoncologist.2009-0121
- Panel SAME. Evaluation summary bevacizumab for treatment of recurrent glioblastoma multiforme. 2014.
- Wick W, Weller M, Bent M, Stupp R. Bevacizumab and recurrent malignant gliomas: a european perspective. J Clin Oncol. 2010;28:e188–e189. doi: 10.1200/JCO.2009.26.9027
- Funakoshi Y, Hata N, Kuga D, Hatae R, Sangatsuda Y, Fujioka Y, Takigawa K, Mizoguchi M. Update on chemotherapeutic approaches and management of bevacizumab usage for glioblastoma. *Pharmaceuticals*. 2020;13:470. doi: 10.3390/ph13120470
- Committee PBA. Public summary document may 2019 pbac meeting. 2019;22.
- 35. Henaine AM, Paubel N, Ducray F, Diebold G, Frappaz D, Guyotat J, Cartalat-Carel S, Aulagner G, Hartmann D, Honnorat J, et al. Current trends in the management of glioblastoma in a french university hospital and associated direct costs. J Clin Pharm Ther. 2016;41:47–53. doi: 10.1111/jcpt.12346
- Arevalo OD, Soto C, Rabiei P, Kamali A, Ballester LY, Esquenazi Y, Zhu J-J, Riascos RF. Assessment of glioblastoma response in the era of bevacizumab: longstanding and emergent challenges in the imaging evaluation of pseudoresponse. *Front Neurol*. 2019;10:460–460. doi: 10.3389/fneur.2019.00460
- Zhao C, Wang H, Xiong C, Liu Y. Hypoxic glioblastoma release exosomal vegf-a induce the permeability of blood-brain barrier. *Biochem Biophys Res Commun*. 2018;502:324–331. doi: 10.1016/j.bbrc.2018.05.140
- Wagner CC, Held U, Kofmehl R, Battegay E, Zimmerli L, Hofer S. Role of arterial hypertension as a predictive marker for bevacizumab efficacy in recurrent glioblastoma – a prospective analysis. *Acta Oncol.* 2014;53:572– 575. doi: 10.3109/0284186X.2013.852240
- Khan KI, Ramesh P, Kanagalingam S, Zargham UI Haq F, Victory Srinivasan N, Khan AI, Mashat GD, Hazique M, Khan S. Bevacizumab-induced hypertension as a potential physiological clinical biomarker for improved outcomes in patients with recurrent glioblastoma multiforme: a systematic review. *Cureus*. 2022;14:e29269. doi: 10.7759/cureus.29269
- Higa GM, Abraham J. Biological mechanisms of bevacizumab-associated adverse events. *Expert Rev Anticancer Ther.* 2009;9:999–1007. doi: 10.1586/era.09.68
- Dinc E, Yildirim O, Ayaz L, Ozcan T, Yilmaz SN. Effects of intravitreal injection of bevacizumab on nitric oxide levels. *Eye.* 2015;29:436–442. doi: 10.1038/eye.2014.297
- Li M, Kroetz DL. Bevacizumab-induced hypertension: clinical presentation and molecular understanding. *Pharmacology & Therapeutics*. 2018;182:152–160. doi: 10.1016/j.pharmthera.2017.08.012
- Mourad JJ, des Guetz G, Debbabi H, Levy BI. Blood pressure rise following angiogenesis inhibition by bevacizumab. A crucial role for microcirculation. *Ann Oncol.* 2008;19:927–934. doi: 10.1093/annonc/mdm550
- Lai AY, Joo IL, Trivedi AU, Dorr A, Hill ME, Stefanovic B, McLaurin J. Cerebrovascular damage after midlife transient hypertension in non-transgenic and alzheimer's disease rats. *Brain Res.* 2021;1758:147369. doi: 10.1016/j.brainres.2021.147369

- Li Q, Youn J-Y, Cai H. Mechanisms and consequences of endothelial nitric oxide synthase dysfunction in hypertension. *J Hypertens.* 2015;33:1128– 1136. doi: 10.1097/HJH.00000000000587
- Node K, Kitakaze M, Yoshikawa H, Kosaka H, Hori M. Reduced plasma concentrations of nitrogen oxide in individuals with essential hypertension. *Hypertension*. 1997;30:405–408. doi: 10.1161/01.hyp.30.3.405
- Schulz E, Gori T, Münzel T. Oxidative stress and endothelial dysfunction in hypertension. *Hypertens Res.* 2011;34:665–673. doi: 10.1038/hr.2011.39
- Kroll J, Waltenberger J. Vegf-a induces expression of enos and inos in endothelial cells via vegf receptor-2 (kdr). *Biochem Biophys Res Commun.* 1998;252:743–746. doi: 10.1006/bbrc.1998.9719
- Konukoglu D, Uzun H. Endothelial dysfunction and hypertension. Adv Exp Med Biol. 2017;956:511–540. doi: 10.1007/5584_2016_90
- Ulivi P, Scarpi E, Passardi A, Marisi G, Calistri D, Zoli W, Del Re M, Frassineti GL, Tassinari D, Tamberi S, et al. Enos polymorphisms as predictors of efficacy of bevacizumab-based chemotherapy in metastatic colorectal cancer: data from a randomized clinical trial. *J Transl Med.* 2015;13:258. doi: 10.1186/s12967-015-0619-5
- Wick A, Schäfer N, Dörner N, Schemmer D, Platten M, Bendszus M, Wick W. Arterial hypertension and bevacizumab treatment in glioblastoma: no correlation with clinical outcome. *J Neurooncol.* 2010;97:157–158. doi: 10.1007/s11060-009-0003-5
- Iwamoto FM, Cooper AR, Reiner AS, Nayak L, Abrey LE. Glioblastoma in the elderly. Cancer. 2009;115:3758–3766. doi: 10.1002/cncr.24413
- Wang J-B, Huang Q-C, Hu S-C, Zheng P-W, Shen P, Li D, Lu H-C, Gao X, Lin H-B, Chen K. Baseline and longitudinal change in blood pressure and mortality in a chinese cohort. *J Epidemiol Community Health*. 2018;72:1083– 1090. doi: 10.1136/jech-2018-211050
- Letourneur A, Roussel S, Bernaudin M, Fillesoye F, Toutain J, MacKenzie ET, Petit E, Touzani O, Valable S. Chronic arterial hypertension impedes glioma growth: a multiparametric mri study in the rat. *Hypertens Res.* 2015;38:723– 732. doi: 10.1038/hr.2015.66
- 55. Rubenstein JL, Kim J, Ozawa T, Zhang M, Westphal M, Deen DF, Shuman MA. Anti-vegf antibody treatment of glioblastoma prolongs survival but results in increased vascular cooption. *Neoplasia*. 2000;2:306–314. doi: 10.1038/sj.neo.7900102
- 56. Kunkel P, Ulbricht U, Bohlen P, Brockmann MA, Fillbrandt R, Stavrou D, Westphal M, Lamszus K. Inhibition of glioma angiogenesis and growth in vivo by systemic treatment with a monoclonal antibody against vascular endothelial growth factor receptor-2. *Cancer Res.* 2001;61:6624–6628.
- Humar R, Zimmerli L, Battegay E. Angiogenesis and hypertension: an update. J Hum Hypertens. 2009;23:773–782. doi: 10.1038/jhh.2009.63
- Abe Y, Watanabe M, Chung S, Kamm RD, Tanishita K, Sudo R. Balance of interstitial flow magnitude and vascular endothelial growth factor concentration modulates three-dimensional microvascular network formation. *APL Bioengineering*. 2019;3:036102. doi: 10.1063/1.5094735
- Kim S, Chung M, Ahn J, Lee S, Jeon NL. Interstitial flow regulates the angiogenic response and phenotype of endothelial cells in a 3d culture model. *Lab Chip.* 2016;16:4189–4199. doi: 10.1039/c6lc00910g
- Vickerman V, Kamm RD. Mechanism of a flow-gated angiogenesis switch: early signaling events at cell-matrix and cell-cell junctions. *Integr Biol.* 2012;4:863–874. doi: 10.1039/c2ib00184e
- Sanai N, Alvarez-Buylla A, Berger MS. Neural stem cells and the origin of gliomas. N Engl J Med. 2005;353:811–822. doi: 10.1056/nejmra043666
- Calabrese C, Poppleton H, Kocak M, Hogg TL, Fuller C, Hamner B, Oh EY, Gaber MW, Finklestein D, Allen M, et al. A perivascular niche for brain tumor stem cells. *Cancer Cell*. 2007;11:69–82. doi: 10.1016/j.ccr.2006.11.020
- Shen O, Goderie SK, Jin L, Karanth N, Sun Y, Abramova N, Vincent P, Pumiglia K, Temple S. Endothelial cells stimulate self-renewal and expand neurogenesis of neural stem cells. *Science*. 2004;304:1338–1340. doi: 10.1126/science.1095505
- Neftel C, Laffy J, Filbin MG, Hara T, Shore ME, Rahme GJ, Richman AR, Silverbush D, Shaw ML, Hebert CM, et al. An integrative model of cellular states, plasticity, and genetics for glioblastoma. *Cell.* 2019;178:835–849. e21. doi: 10.1016/j.cell.2019.06.024
- Infanger DW, Cho Y, Lopez BS, Mohanan S, Liu SC, Gursel D, Boockvar JA, Fischbach C. Glioblastoma stem cells are regulated by interleukin-8 signaling in a tumoral perivascular niche. *Cancer Res.* 2013;73:7079–7089. doi: 10.1158/0008-5472.CAN-13-1355
- Charles N, Ozawa T, Squatrito M, Bleau A-M, Brennan CW, Hambardzumyan D, Holland EC. Perivascular nitric oxide activates notch signaling and promotes stem-like character in pdgf-induced glioma cells. *Cell Stem Cell*. 2010;6:141-152. doi: 10.1016/j.stem.2010.01.001
- 67. Li D, Tian Y, Hu Y, Qi Y, Tian N, Li S, Hu P, Wu F, Wei Q, Wei Z, et al. Gliomaassociated human endothelial cell-derived extracellular vesicles specifically

promote the tumourigenicity of glioma stem cells via cd9. *Oncogene.* 2019;38:6898-6912. doi: 10.1038/s41388-019-0903-6

- Zhu TS, Costello MA, Talsma CE, Flack CG, Crowley JG, Hamm LL, He X, Hervey-Jumper SL, Heth JA, Muraszko KM, et al. Endothelial cells create a stem cell niche in glioblastoma by providing notch ligands that nurture selfrenewal of cancer stem-like cells. *Cancer Res.* 2011;71:6061–6072. doi: 10.1158/0008-5472.CAN-10-4269
- Hira VVV, Aderetti DA, van Noorden CJF. Glioma stem cell niches in human glioblastoma are periarteriolar. J Histochem Cytochem. 2018;66:349–358. doi: 10.1369/0022155417752676
- Mohyeldin A, Garzón-Muvdi T, Quiñones-Hinojosa A. Oxygen in stem cell biology: a critical component of the stem cell niche. *Cell Stem Cell*. 2010;7:150-161. doi: 10.1016/j.stem.2010.07.007
- Hira VVV, Wormer JR, Kakar H, Breznik B, van der Swaan B, Hulsbos R, Tigchelaar W, Tonar Z, Khurshed M, Molenaar RJ, et al. Periarteriolar glioblastoma stem cell niches express bone marrow hematopoietic stem cell niche proteins. *Journal of Histochemistry & Cytochemistry*. 2018;66:155– 173. doi: 10.1369/0022155417749174
- Bigarella CL, Liang R, Ghaffari S. Stem cells and the impact of ros signaling. Development 2014;141:4206–4218. doi: 10.1242/dev.107086
- Sato A, Okada M, Shibuya K, Watanabe E, Seino S, Narita Y, Shibui S, Kayama T, Kitanaka C. Pivotal role for ros activation of p38 mapk in the control of differentiation and tumor-initiating capacity of glioma-initiating cells. *Stem Cell Res.* 2014;12:119–131. doi: 10.1016/j.scr.2013.09.012
- Ridnour L, Thomas D, Donzelli S, Espey M, Roberts D, Wink D, Isenberg J. The biphasic nature of nitric oxide responses in tumor biology. *Antioxidants* & redox signaling. 2006;8:1329–1337. doi: 10.1089/ars.2006.8.1329
- Tran AN, Boyd NH, Walker K, Hjelmeland AB. Nos expression and no function in glioma and implications for patient therapies. *Antioxid Redox Signal.* 2017;26:986–999. doi: 10.1089/ars.2016.6820
- Debbabi H, Uzan L, Mourad JJ, Safar M, Levy BI, Tibiriçà E. Increased skin capillary density in treated essential hypertensive patients*. *Am J Hypertens*. 2006;19:477–483. doi: 10.1016/j.amjhyper.2005.10.021
- Swerdel JN, Janevic TM, Cabrera J, Cosgrove NM, Sedjro JE, Pressel SL, Davis BR, Kostis JB. Rapid decreases in blood pressure from antihypertensive treatment were associated with increased cancer mortality in the systolic hypertension in the elderly program. *Cancer Epidemiology Biomarkers & Prevention.* 2014;23:1589–1597. doi: 10.1158/1055-9965.EPI-14-0085
- Scherer HJ. Structural development in gliomas. *The American Journal of Cancer*. 1938;34:333–351.
- Tamura R, Miyoshi H, Sampetrean O, Shinozaki M, Morimoto Y, Iwasawa C, Fukaya R, Mine Y, Masuda H, Maruyama T, et al. Visualization of spatiotemporal dynamics of human glioma stem cell invasion. *Molecular Brain*. 2019;12:45. doi: 10.1186/s13041-019-0462-3
- Fujishima M, Ibayashi S, Fujii K, Mori S. Cerebral blood flow and brain function in hypertension. *Hypertens Res* 1995;18:111–117. doi: 10.1291/hypres.18.111
- Baker GJ, Yadav VN, Motsch S, Koschmann C, Calinescu A-A, Mineharu Y, Camelo-Piragua SI, Orringer D, Bannykh S, Nichols WS, et al. Mechanisms of glioma formation: iterative perivascular glioma growth and invasion leads to tumor progression, vegf-independent vascularization, and resistance to antiangiogenic therapy. *Neoplasia*. 2014;16:543–561. doi: 10.1016/j.neo.2014.06.003
- Paez-Ribes M, Allen E, Hudock J, Takeda T, Okuyama H, Vinals F, Inoue M, Bergers G, Hanahan D, Casanovas O. Antiangiogenic therapy elicits malignant progression of tumors to increased local invasion and distant metastasis. *Cancer Cell*. 2009;15:220–231. doi: 10.1016/j.ccr.2009.01.027
- Falchetti ML, D'Alessandris QG, Pacioni S, Buccarelli M, Morgante L, Giannetti S, Lulli V, Martini M, Larocca LM, Vakana E, et al. Glioblastoma endothelium drives bevacizumab-induced infiltrative growth via modulation of plxdc1. *Int J Cancer.* 2019;144:1331–1344. doi: 10.1002/ijc.31983
- Watkins S, Robel S, Kimbrough IF, Robert SM, Ellis-Davies G, Sontheimer H. Disruption of astrocyte–vascular coupling and the blood–brain barrier by invading glioma cells. *Nat Commun.* 2014;5:4196. doi: 10.1038/ncomms5196
- 85. Iliff JJ, Wang M, Liao Y, Plogg BA, Peng W, Gundersen GA, Benveniste H, Vates GE, Deane R, Goldman SA, et al. A paravascular pathway facilitates csf flow through the brain parenchyma and the clearance of interstitial solutes, including amyloid β. *Sci Transl Med.* 2012;4:147ra111–147ra111. doi: 10.1126/scitranslmed.3003748
- Rasmussen MK, Mestre H, Nedergaard M. Fluid transport in the brain. *Physiol Rev.* 2022;102:1025–1151. doi: 10.1152/physrev.00031.2020
- Nakada T, Kwee IL. Fluid dynamics inside the brain barrier: current concept of interstitial flow, glymphatic flow, and cerebrospinal fluid circulation in the brain. *The Neuroscientist*. 2019;25:155–166. doi: 10.1177/1073858418775027

- Iliff J, Simon M. Crosstalk proposal: the glymphatic system supports convective exchange of cerebrospinal fluid and brain interstitial fluid that is mediated by perivascular aquaporin-4. *J Physiol.* 2019;597:4417–4419. doi: 10.1113/JP277635
- Smith AJ, Verkman AS. Crosstalk opposing view: going against the flow: interstitial solute transport in brain is diffusive and aquaporin-4 independent. *J Physiol.* 2019;597:4421–4424. doi: 10.1113/JP277636
- Mestre H, Tithof J, Du T, Song W, Peng W, Sweeney AM, Olveda G, Thomas JH, Nedergaard M, Kelley DH. Flow of cerebrospinal fluid is driven by arterial pulsations and is reduced in hypertension. *Nat Commun.* 2018;9:4878. doi: 10.1038/s41467-018-07318-3
- Sweeney AM, Plá V, Du T, Liu G, Sun O, Peng S, Plog BA, Kress BT, Wang X, Mestre H, et al. In vivo imaging of cerebrospinal fluid transport through the intact mouse skull using fluorescence macroscopy. *J Vis Exp.* 2019;10.3791/59774. doi: 10.3791/59774
- Abbott NJ, Pizzo ME, Preston JE, Janigro D, Thorne RG. The role of brain barriers in fluid movement in the cns: is there a "glymphatic" system? *Acta Neuropathol.* 2018;135:387–407. doi: 10.1007/s00401-018-1812-4
- Raghunandan A, Ladron-de-Guevara A, Tithof J, Mestre H, Du T, Nedergaard M, Thomas JH, Kelley DH. Bulk flow of cerebrospinal fluid observed in periarterial spaces is not an artifact of injection. *eLife*. 2021;10:e65958. doi: 10.7554/eLife.65958
- Verheggen ICM, Van Boxtel MPJ, Verhey FRJ, Jansen JFA, Backes WH. Interaction between blood-brain barrier and glymphatic system in solute clearance. *Neuroscience & Biobehavioral Reviews*. 2018;90:26–33. doi: 10.1016/j.neubiorev.2018.03.028
- Xu D, Zhou J, Mei H, Li H, Sun W, Xu H. Impediment of cerebrospinal fluid drainage through glymphatic system in glioma. *Front Oncol.* 2021;11:790821. doi: 10.3389/fonc.2021.790821
- Pacioni S, D'Alessandris OG, Buccarelli M, Boe A, Martini M, Larocca LM, Bolasco G, Ricci-Vitiani L, Falchetti ML, Pallini R. Brain invasion along perivascular spaces by glioma cells: relationship with blood-brain barrier. *Cancers (Basel)*. 2019;12:18. doi: 10.3390/cancers12010018
- Sarkaria JN, Hu LS, Parney IF, Pafundi DH, Brinkmann DH, Laack NN, Giannini C, Burns TC, Kizilbash SH, Laramy JK, et al. Is the blood-brain barrier really disrupted in all glioblastomas? A critical assessment of existing clinical data. *Neuro-Oncology*. 2017;20:184–191. doi: 10.1093/neuonc/nox175
- Munson JM, Bellamkonda RV, Swartz MA. Interstitial flow in a 3d microenvironmentincreases glioma invasion by a cxcr4-dependent mechanism. *Cancer Res.* 2013;73:1536–1546. doi: 10.1158/0008-5472.CAN-12-2838
- Geer CP, Grossman SA. Interstitial fluid flow along white matter tracts: a potentially important mechanism for the dissemination of primary brain tumors. J Neurooncol. 1997;32:193–201. doi: 10.1023/a:1005761031077
- 100. Cornelison RC, Brennan CE, Kingsmore KM, Munson JM. Convective forces increase cxcr4-dependent glioblastoma cell invasion in gl261 murine model. *Sci Rep.* 2018;8:17057. doi: 10.1038/s41598-018-35141-9
- 101. Kingsmore KM, Logsdon DK, Floyd DH, Peirce SM, Purow BW, Munson JM. Interstitial flow differentially increases patient-derived glioblastoma stem cell invasion via cxcr4, cxcl12, and cd44-mediated mechanisms. *Integr Biol.* 2016;8:1246–1260. doi: 10.1039/c6ib00167j
- 102. Shields JD, Fleury ME, Yong C, Tomei AA, Randolph GJ, Swartz MA. Autologous chemotaxis as a mechanism of tumor cell homing to lymphatics via interstitial flow and autocrine ccr7 signaling. *Cancer Cell*. 2007;11:526– 538. doi: 10.1016/j.ccr.2007.04.020
- 103. Hira VV, Verbovsek U, Breznik B, Srdic M, Novinec M, Kakar H, Wormer J, der Swaan BV, Lenarcic B, Juliano L, et al. Cathepsin k cleavage of sdf-1alpha inhibits its chemotactic activity towards glioblastoma stem-like cells. *Biochim Biophys Acta Mol Cell Res.* 2017;1864:594–603. doi: 10.1016/j.bbamcr.2016.12.021
- 104. Stevenson CB, Ehtesham M, McMillan KM, Valadez JG, Edgeworth ML, Price RR, Abel TW, Mapara KY, Thompson RC. Cxcr4 expression is elevated in glioblastoma multiforme and correlates with an increase in intensity and extent of peritumoral t2-weighted magnetic resonance imaging signal abnormalities. *Neurosurgery*. 2008;63:560–9; discussion 569. doi: 10.1227/01.NEU.0000324896.26088.EF
- 105. Pires PW, Dams Ramos CM, Matin N, Dorrance AM. The effects of hypertension on the cerebral circulation. Am J Physiol Heart Circ Physiol. 2013;304:H1598–H1614. doi: 10.1152/ajpheart.00490.2012
- 106. Kokovay E, Goderie S, Wang Y, Lotz S, Lin G, Sun Y, Roysam B, Shen Q, Temple S. Adult svz lineage cells home to and leave the vascular niche via differential responses to sdf1/cxcr4 signaling. *Cell Stem Cell*. 2010;7:163– 173. doi: 10.1016/j.stem.2010.05.019
- 107. Marx V. Method of the year: spatially resolved transcriptomics. Nat Methods. 2021;18:9–14. doi: 10.1038/s41592-020-01033-y
- 108. Doerr A. Mass spectrometry imaging takes off. *Nat Methods*. 2018;15:32– 32. doi: 10.1038/nmeth.4546