

1           **Ion-Imprinted Nanofilms Based on Tannic Acid and Silver**  
2                           **Nanoparticles for Sensing of Al(III)**

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12          **Keywords:** supramolecular film, electrochemical self-assembly, aluminum sensing, ion-  
13          printing, tailored sensor, human serum, electro-cross-linking

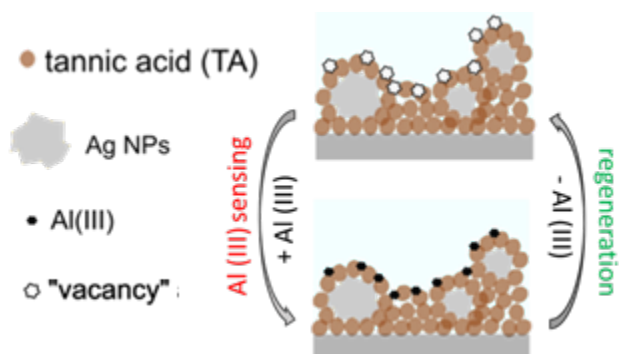
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15

16 **Abstract:** Electrochemically triggered self-assembly can be effectively utilized to produce  
17 electroactive materials of tailored properties for various applications, such as sensor  
18 development. Here, we present a thin sensor film based on tannic acid (TA) and silver  
19 nanoparticles (AgNPs), ionically imprinted via electrodeposition and tailor-designed for  
20 electrochemical tracing of aluminum ions, Al(III). In the first stage, the conditions for the  
21 Al(III)-printing of TA films onto indium-tin-oxide (ITO) electrode via electrodeposition are  
22 established and optimized. To form an AgNPs-containing film, AgNPs are pre-synthesized via  
23 a direct reduction of Ag(I) by TA resulting in TA-stabilized AgNPs (TA@AgNPs) of 1-4 nm  
24 in size, as observed by dynamic light scattering. Next, Al(III) ions are added to complex the TA  
25 molecules adsorbed on the surface of AgNPs. The resulting Al(III)/TA@AgNPs mixture is then  
26 electrodeposited onto ITO surface by applying an anodic potential to form a film. As a result,  
27 a mesh-structured layer composed of AgNPs with TA on their surface and electrochemically  
28 cross-linked via TA-TA covalent bonds at the Al(III)-free coordination sites is formed. The  
29 introduction of Al(III) ions bonded via coordination bonds with TA and their consecutive  
30 removal using sodium fluoride formed vacancies ready to bind Al(III) ions from the analyzed  
31 solution allowing their electrochemical sensing, as monitored by cyclic voltammetry, quartz  
32 crystal microbalance (QCM) and X-ray photoelectron spectroscopy. The film was employed  
33 for sensing of neurotoxic Al(III) in human serum. A linear correlation between the current value  
34 at 0.9 V and the concentration of Al(III) was obtained in the range between 0.10 and 0.298  $\mu\text{M}$ .  
35

36 **Graphical abstract**

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## 40 **1. Introduction**

41 Self-assembly technique is playing an important role in preparing well-defined multilevel  
42 nanostructures and the functionalized nanomaterials of designed and controlled properties.<sup>1</sup>

43 Molecular self-assembly usually takes advantage of supramolecular interactions (ionic,  
44 hydrophobic, van der Waals, hydrogen and coordination bonds), but can also make use of  
45 kinetically labile covalent bonds<sup>2</sup> or redox processes triggered by an electrical stimulus<sup>3</sup>.  
46 Electrochemically triggered self-construction of films is a relatively simple, promising and  
47 effective approach (i) providing covalent<sup>4-6</sup> and/or (ii) non-covalent<sup>4, 7, 8</sup> immobilization of  
48 compounds with excellent functionalities<sup>9</sup> or (bio)-activities<sup>4, 10</sup> and (iii) allowing selective  
49 functionalization of microelectrode arrays.<sup>4, 9, 10</sup>

50 Polyphenols, such as tannic acid, TA, are able to coordinate and to self-assemble with different  
51 metal ions into combined cross-linked networks of polyphenols and metal ions, the  
52 functionality of which can be versatile based on the incorporated polyphenols and metal ions<sup>11-</sup>  
53 <sup>13</sup>. Tannic acid, the polyphenol used in this work, is present in abundance in nature and easily  
54 accessible. It can also act as a reducing or stabilizing agent to form metal nanoparticles<sup>14</sup> by  
55 reacting covalently in its oxidative state forming intermolecular bonds leading to the formation  
56 of a film. The ability of TA to form covalent bonds between its own molecules under an  
57 electrical stimulus, as well as its ability to complex metals like iron, vanadium or aluminum to  
58 form electroactive nanocoatings<sup>15</sup> can be very well utilized for the fabrication of an  
59 electrochemical sensor, using, for example, a molecular imprinting<sup>16</sup> or an ion-printing  
60 approach<sup>17, 18</sup> and introduction of metal nanoparticles.<sup>19</sup>

61 Herein, an electrochemically triggered self-assembly approach was utilized to produce metal-  
62 polyphenol thin films tailor-designed for voltammetric sensing of aluminum, Al(III), via an ion-  
63 printing approach. Aluminum is widely used in almost every area of our everyday life, i.e. from  
64 food processing, water purification, cosmetics, medicines, vaccinations to clothing and building

65 construction, due to its incredibly useful physicochemical properties. On the other hand, Al(III),  
66 even at a trace level in biological fluids, is a dangerous neurotoxin with poisoning effects. It  
67 binds to the phosphate groups of DNA and RNA, influencing their topology, affects gene  
68 transcription and accumulates in the brain matter.<sup>20-22</sup> Currently, the determination of Al(III)  
69 levels in biological fluids of patients is conventionally performed using laboratory-based  
70 spectroscopic methods,<sup>23</sup> offering reasonable sensitivity and reliability, but requiring expensive  
71 instrumentation, expertise in handling and presenting the limitation of an ex-situ analysis.<sup>24</sup>  
72 With its high sensitivity and rapid analysis time, electrochemical determination of Al(III) level  
73 could be ideal for routine monitoring. A range of electrochemical sensors has been reported in  
74 the literature, based on enzyme inhibition,<sup>25, 26</sup> different complexing agents of metal ions<sup>27, 28</sup>  
75 such as (poly)phenols<sup>23, 29-33</sup> or zeolites.<sup>34, 35</sup> These sensors, however, suffer from different  
76 drawbacks: low sensitivity or precision, use of mercury electrodes, sensitivity to interfering  
77 species and none of them can be selectively deposited on microarrays electrode.<sup>23, 25, 26, 28, 31-33,</sup>  
78 <sup>36-38</sup> Most of them were designed for (i) Al(III) sensing in acidic conditions<sup>23, 27, 28, 32-35</sup> and/or  
79 (ii) in physiological conditions but in the absence of serum proteins.<sup>25, 26, 29</sup>  
80 Here, we report for the first time an application of electrochemically assisted self-assembly to  
81 form a complex supramolecular hybrid thin film composed of a biodegradable and non-toxic  
82 molecules, metal ions and metal nanoparticles. The film was obtained via a one pot process  
83 while taking advantage of multiple specific supramolecular and chemical interactions of each  
84 component of the system such to ensure the tailored properties of the sensor towards  
85 electrochemical sensing of Al(III) in biological fluids. TA has been selected as the suitable  
86 monomer candidate for the polymeric matrix of the sensor due to its ability to form relatively  
87 stable complexes with Al(III)<sup>15</sup> as well as to cross-link under an oxidant stimulus to form a  
88 film.<sup>3</sup> Due to the biological fluids being the targeted medium for the sensing process, the film  
89 was deposited onto indium tin oxide (ITO) to ensure the stability of the electrode upon an

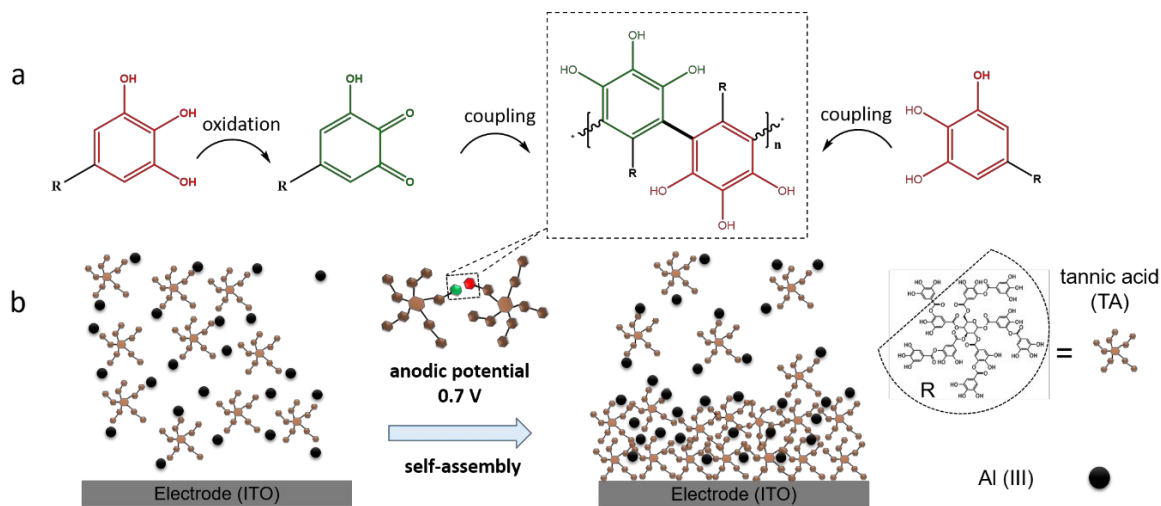
90 extended contact with a chloride solution as opposed to e.g., gold.<sup>39, 40</sup> Due to the use of a  
91 semiconductive substrate and low electrical conductivity of the cross-linked TA, AgNPs were  
92 chosen as the next component of the film due to their high electrical conductivity and ability to  
93 enhance the electron transfer between the organic coating and the electrode and thus, improving  
94 the overall sensitivity of the film.<sup>41-43</sup> Following, a mussel inspired electro-cross-linking of TA-  
95 capped silver nanoparticles (TA@AgNPs) complexed by Al(III) was utilized to develop  
96 ionically imprinted electroactive sensor coatings. A mixture of pre-synthesized TA@AgNPs  
97 complexed by Al(III) was used as a building solution in contact with ITO electrode. The  
98 application of an anodic potential induced the oxidation of TA gallol moieties into quinone  
99 moieties that led to TA cross-linking and the simultaneous covalent immobilization of  
100 TA@AgNP with the incorporation of Al(III) ions through the coordination bonds.

101 The obtained sensor, based on non-toxic and environmentally friendly and abundant species,  
102 was proven to be stable over a longer period of time under a physiological concentration of  
103 sodium chloride buffered at pH 7.4 and was applied for Al(III) sensing in human serum  
104 solution. The results provide new insight into complex supramolecular film formation and  
105 understanding of the voltammetric sensing of metal ions in blood serum samples as well as  
106 stability of AgNPs under physiological conditions. This new electrochemically assisted self-  
107 assembly approach could be easily generalized for detection of several metal ions, i.e. by  
108 choosing an appropriate type of molecule or macromolecule and metal nanoparticles or  
109 adjusting a fitting agent to remove the particular metal ion.<sup>44</sup>

110 **2. Results and discussion**

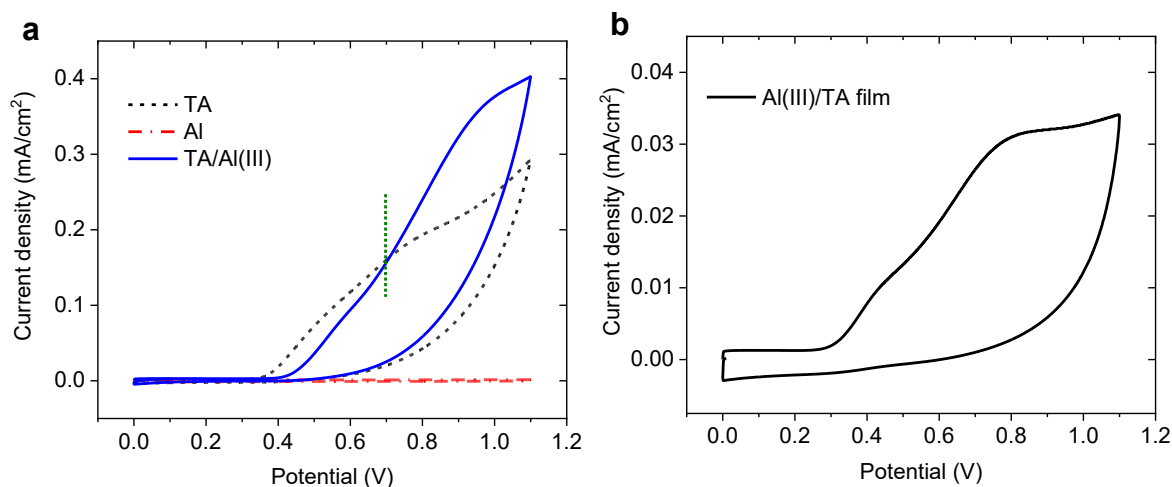
111 **2.1. Formation of Al(III)/TA films on ITO**

112 Initial experiments involved the study on the ability of TA to form films with Al(III) via  
113 electrochemically assisted self-assembly according to the scheme shown in **Figure 1**, as well  
114 as their electrochemical behavior to obtain a reference before involving AgNPs as the film  
115 building blocks. The experimental parameters of the electrodeposition were optimized  
116 previously where Fe(III)/TA films were deposited using the oxidation of Fe(II).<sup>3</sup> We noticed  
117 that the application of a potential higher than 0.5 V induces a cross-linking of TA. Following  
118 the procedure established, we adjusted the protocol accordingly using the same concentration  
119 of TA. As TA can coordinate up to three Al(III) ions<sup>45</sup>, the Al(III) to TA molar ratio of 2.5 was  
120 chosen for the film formed to both contain a sufficient number of Al(III) incorporated and also  
121 to assure a considerable number of TA free sites available for the TA-TA bonding during the  
122 electrochemically triggered self-assembly of the film.



124 **Figure 1** Formation of Al(III)-imprinted film via electrochemically triggered cross-linking of  
125 tannic acid molecules complexed by Al(III); a) mechanism of the TA-TA covalent bond  
126 formation upon an electrical stimulus and b) schematic presentation of the self-assembly of the  
127 Al(III)/TA film.

128 Due to the fact that the properties and physicochemical stability of the TA-based films vary  
129 depending on the pH, to ensure the stability of the films under the conditions intended for  
130 sensing later on, the self-assembly of the films was performed at the targeted physiological pH  
131 value of 7.4. In the first stage, the electrochemical response of TA and Al(III) in 0.15 M  $KPF_6$   
132 was studied by cyclic voltammetry (CV) to determine the applicable potential range for the self-  
133 assembly of the films onto ITO electrode surface. The CV registered for the of TA-Al(III)  
134 electrodeposition (ED) mixture (**Figure 2a**) exhibited an oxidation range between 0.4 and 1.1  
135 V with a peak centered at 0.9 V corresponding to the oxidation of TA-Al(III) complexes. In the  
136 case of CV where no Al(III) was added, the oxidation range of TA was slightly shifted towards  
137 the lower potential values in comparison to the Al(III)-TA curve prompting that the cross-  
138 linking of the not complexed TA molecules required a lower overpotential than in a complexed  
139 state.



140  
141 **Figure 2** a) Cyclic voltammograms registered for bare ITO electrode in TA and Al(III)  
142 solutions at concentrations corresponding to the ED and the Al(III)-TA ED mixture at 2.5  
143 Al(III)/TA molar ratio. b) Cyclic voltammogram of the as-deposited Al(III)-TA film. All curves  
144 registered at 50 mV/s in 0.15 M  $KPF_6$  adjusted to pH 7.4. The ED potential is marked with a  
145 dotted line in a).

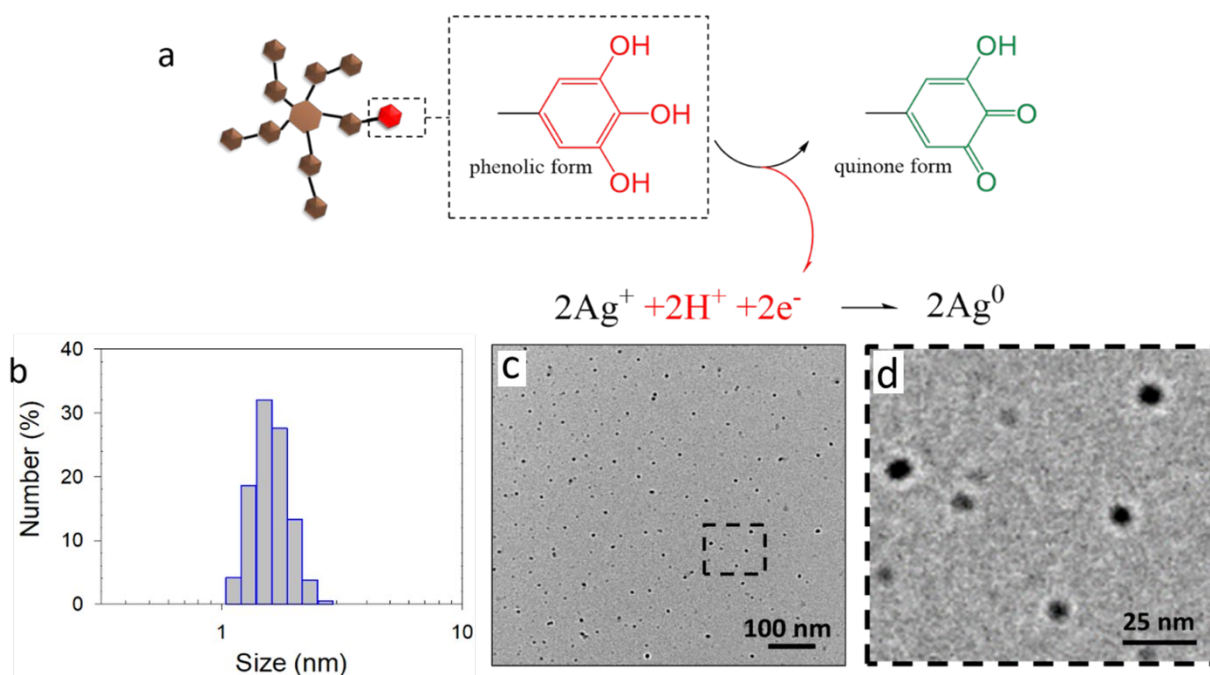


146 In both cases, the oxidation of TA molecules corresponds to the oxidative crosslinking of the  
147 TA-TA molecules. In the case of Al(III)/TA complex, a higher activation energy is required to  
148 decompose the complex first before the oxidation of TA. The current registered for the Al(III)  
149 solution is similar to 0.15 M KPF<sub>6</sub> medium. Al(III)/Al has a redox potential at -1.67 vs NHE (-  
150 1.87 vs Ag/AgCl) which is not in the potential range of the CV. The CV registered for the bare  
151 ITO electrode in a solution containing Al(III) exhibited no distinctive signals and the capacity  
152 current measured was of the same magnitude as for the KPF<sub>6</sub> solution (**Figure S1** in the SI),  
153 showing no influence of Al(III) on the current value when a non-coated ITO electrode was used.  
154 On the basis of the CV data, the potential value for the potentiostatic electrodeposition was set  
155 to 0.7 V, which was high enough to cause the crosslinking between the TA molecules in a  
156 complexed state (**Figure 2a**) and, on the other hand, too low to favor the oxidation and  
157 decomposition of TA-Al(III) complexes in the ED solution. The ED current and the quartz  
158 crystal microbalance (QCM) curve registered in-situ for the formation of the film are shown in  
159 **Figure S2** in the SI. The CV of the as-deposited film in the basic electrolyte (**Figure 2b**)  
160 exhibited the same distinctive features as in the case of CV for the TA-Al(III) mixture with the  
161 reduction wave centered at 0.8 V instead of 0.9V, revealing the presence of both TA and Al(III)  
162 in the films. This shift could be a result of the lower content of Al(III) and a different  
163 stoichiometry of the Al(III)/TA complexes in the film as compared to in the bulk. The current  
164 density registered for the film in the solution was lower in magnitude than the current density  
165 registered for Al(III) in the basic electrolyte by a factor of 10, prompting the fact that only a  
166 small portion of the TA-Al(III) complexes were deposited onto the surface of the QCM sensor  
167 from the solutions.

## 168 **2.2. Synthesis of Al(III)/(TA@AgNPs) films**

169 **2.2.1. Synthesis of TA@AgNPs.** To avoid the use of any additional species, colloidal AgNPs  
170 were synthesized in water using TA as both the reducing agent towards Ag(I) and capping agent

171 towards Ag(I) (**Figure 3a**).<sup>46-48</sup> Typically, the use of a single reagent for the synthesis of AgNPs  
 172 results in particles with a broad range of sizes and geometries with the size of AgNPs obtained  
 173 via reduction by TA ranging from 7 to 200nm and with the highest stability at 2:1 TA to Ag(I)  
 174 molar ratio.<sup>47</sup> In order to minimize the size and the polydispersity of AgNPs, the synthesis  
 175 procedure the overall reagent concentration was increased by 10-fold and performed at 20 ppm  
 176 in TA concentration with TA/Ag(I) molar ratio of 2:1. Due to the increase in the overall  
 177 concentration of both the reduced species, Ag(I), and the reducing and stabilizing agent, TA,  
 178 the decrease in the diameter of the nanoparticle was observed due to the steric limitation, but  
 179 for the cost of their stability in time.<sup>47</sup> The hydrodynamic size and stability of the TA@AgNPs  
 180 was assessed by DLS (**Figure 3b**).

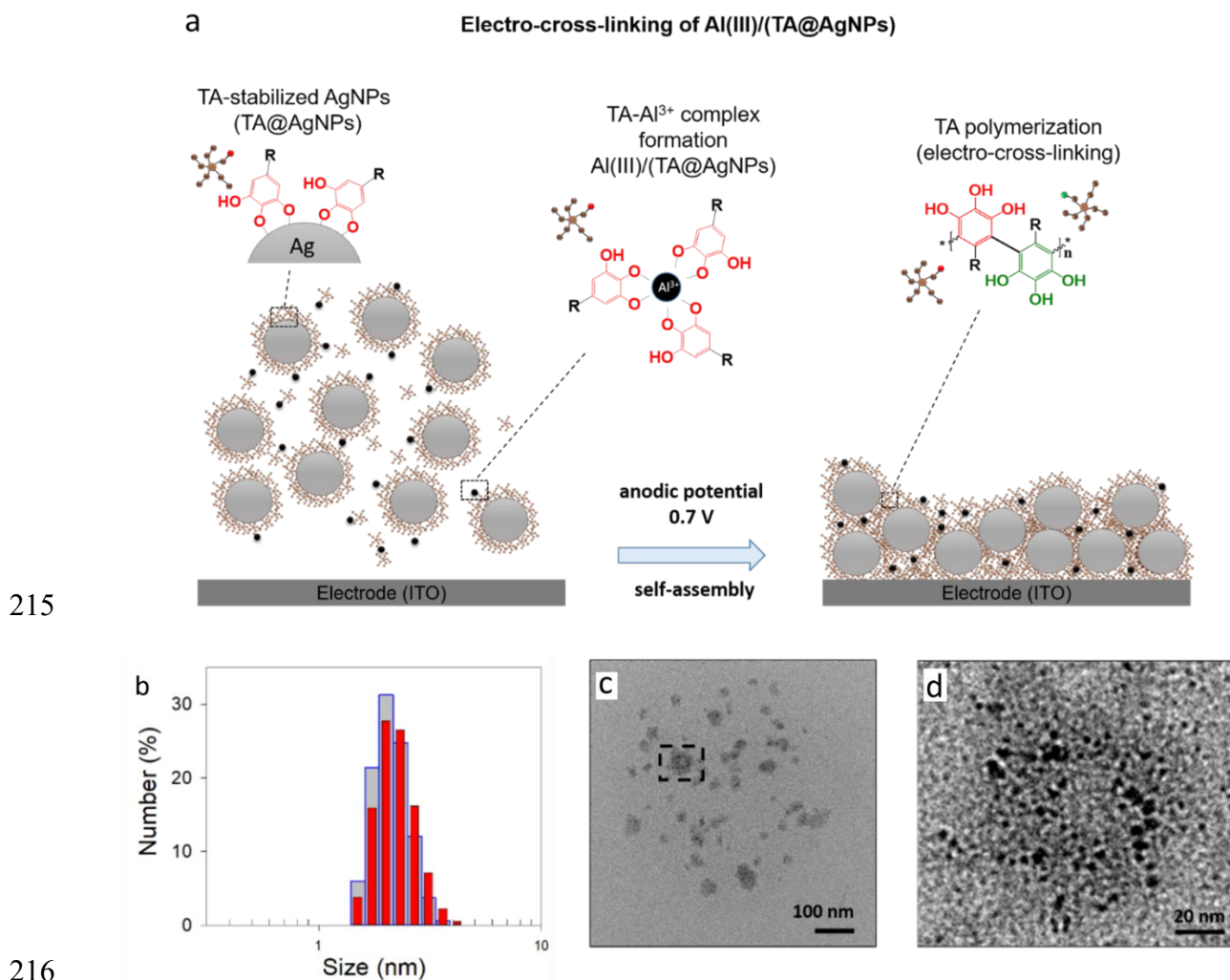


181  
 182 **Figure 3.** TA@AgNPs characterization (a) mechanism of TA@AgNPs formation via direct  
 183 reduction of Ag(I) ions by TA in water, (b) size distribution, determined by DLS, after synthesis  
 184 and (c) the corresponding TEM image in a dry state with (d) zoom-in of the dashed box. The  
 185 round-shaped objects marked in (d) correspond to the hydrodynamic size determined by DLS.

186 Directly after the synthesis, the size of the NPs ranged from 1 to 4 nm with a maximum at 2 nm  
187 and promptly increasing in time due to the proceeding aggregation of the nanoparticles. At the  
188 ED time (3 min after their synthesis), the lower-sized species have not been observed and the  
189 maximum in the histogram was shifted to 3 nm, with the sizes ranging between 2 and 4 nm  
190 (**Figure S3** in the SI). The TEM images revealed regular and uniformly distributed metal  
191 species of up to 10 nm in diameter (**Figure 3c** and **d**). The size of TA@AgNPs in TEM images  
192 is  $6 \pm 2$  nm by Image J treatment. The size obtained by DLS is between 2 and 4 nm. The sizes  
193 are similar which could be explained by the small thickness of TA adsorbed on the surface of  
194 the AgNPs.

195 **2.2.2. Formation of Al(III)/(TA@AgNPs) complexes.** **Figure 4a** shows the procedure  
196 adapted for the electrochemically triggered self-assembly of the Al(III)-imprinted film based  
197 on AgNPs and TA. TA@AgNPs obtained via the procedure described in the previous section  
198 were subjected to complexation by Al(III) at Al(III)/TA molar ratio of 2.5 in KPF<sub>6</sub> to form  
199 Al(III)/TA@AgNPs. As a result, a mixture of differently sized TA@AgNPs species complexed  
200 by Al(III) via TA-Al(III)<sup>49, 50</sup> coordination bonds at various ratios has been obtained. By  
201 comparing the strength and the character of TA-AgNPs interaction, as well as the TA-Al(III)  
202 complex strength, the addition of Al(III) would cause rather the aggregation of the TA@AgNPs  
203 species rather than destabilization of the whole mixture and aggregation of AgNPs into bulk  
204 silver<sup>45</sup>, similarly as observed for TA-stabilized gold nanoparticles.<sup>23</sup> The hydrodynamic  
205 diameters of Al(III)/TA@AgNPs measured by DLS ranged between 2 and 5 nm, with respect  
206 to their number with the maximum centered at 3 nm (**Figure 4b**). As compared to the size  
207 distribution versus intensity of the absorbed light, a negligible number of higher-sized particles  
208 were observed as well (**Figure S4** in the SI). While the size distribution shifted slightly towards  
209 higher diameters comparing to TA@AgNPs (**Figure 3a**), the overall range has not changed at  
210 the ED time, revealing an increased stability in time. In the corresponding TEM images, patch-

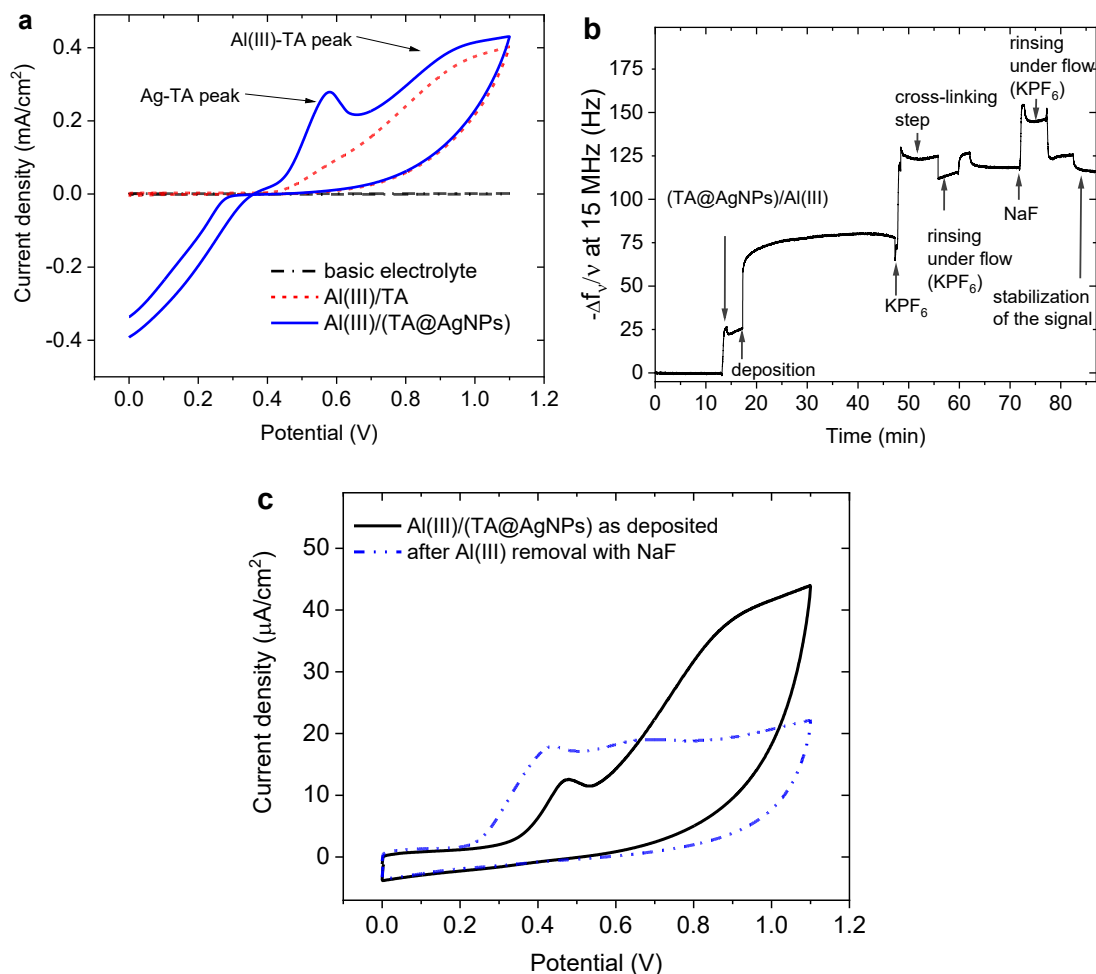
211 like aggregates upon addition of Al(III) (**Figure 4c and d**) were observed. This observation is  
 212 in agreement with recent work, reporting the agglomeration of AgNPs in the presence of Al(III)  
 213 ions followed by an aggregation-induced fluorescence emission.<sup>51</sup>  
 214



216  
 217 **Figure 4** (a) Schematic presentation of the electro-cross-linking of Al(III)/TA@AgNPs  
 218 suspension, obtained by application of an anodic potential value to form the film on ITO. The  
 219 ion-printing procedure achieved via incorporation of Al(III) through coordination bonds with  
 220 TA and inducing the oxidation of gallol moieties of TA into quinone followed by TA cross-  
 221 linking. (b) Size distribution of Al(III)/TA@AgNPs obtained by DLS for Al(III)/TA molar ratio  
 222 of 2.5, determined by DLS in 0.15 M KPF<sub>6</sub>, 1 min (blue) and 3 min (red) after complexation

223 with Al(III) and (c) The corresponding TEM image after 1 min of complexation with (d) zoom-  
224 in of the dashed box.

225 **2.2.3 Electrodeposition of Al(III)/(TA@AgNPs) sensor films.** The CV registered for ITO  
226 electrode in the Al(III)/(TA@AgNPs) solution at the ED concentration level is shown in **Figure**  
227 **5a.**



228

229

230 **Figure 5** (a) Cyclic voltammograms registered for bare ITO electrode in 0.15 M KPF<sub>6</sub> at pH  
231 7.4 (black dash-dotted line) and solutions containing Al(III)/TA@AgNPs (solid blue line) and  
232 Al(III)/TA (red dash line). (b) Normalized frequency shift, measured by QCM, as a function of  
233 time during the electrodeposition of Al(III)/TA@AgNPs (2.5 Al(III) to TA molar ratio) in 0.15  
234 M KPF<sub>6</sub> at pH 7.4, obtained by application of 0.7 V for 30 min, followed by a cross-linking  
235 step via application of CV (0 to 1.1 V at 50 mV/s scan rate) and rinsing steps using 0.15 M

236 KPF<sub>6</sub> at pH 7.4. (c) Cyclic voltammogram, performed at 50 mV/s in 0.15 M KPF<sub>6</sub> at pH 7.4,  
237 of Al(III)/TA@AgNPs coating before (blue dashed line) and after (solid black line) contact  
238 with 0.48 M NaF solution for 5 min. (color online)

239 In comparison to the CV curve registered without AgNPs (**Fig 2a, solid line**), an additional  
240 peak centered at ca. 0.55 V corresponding to TA-AgNPs interaction can be observed (**Figure**  
241 **2c** and **Figure S5** in the SI). Similarly, as in the case of CV registered for the Al(III)/TA  
242 mixture, a signal ascribed to the Al(III)/TA complex oxidation appeared in the solution  
243 containing all three species confirming their presence. The ED potential value was set at 0.7 V  
244 in order to ensure the efficient intermolecular cross-linking of TA and their bonding to AgNPs  
245 during the film buildup and to be below the oxidation of Al(III)/TA complexes (**Figure 5a,**  
246 **dashed line**). After injection of Al(III)/TA@AgNPs suspension into the QCM cell, an increase  
247 in the normalized frequency shift was observed due to non-specific adsorption (**Figure 5b**). In  
248 order to achieve cross-linking of TA and the ED of the film, the applied potential was set to 0.7  
249 V.<sup>52</sup> When the electric potential at 0.7 V was applied, the normalized frequency shift increased  
250 rapidly reaching a plateau at 80 Hz. Simultaneously, a steady decrease in the electrical current  
251 was observed, exhibiting the characteristics of a typical diffusion-controlled process, with the  
252 calculated overall charge density of ca. 0.6 mC/cm<sup>2</sup> (**Figure S6** in the SI). Coatings based on  
253 TA typically exhibit low electrical conductivity because of the lack of conjugated double bonds  
254 in the formed coatings.<sup>53</sup> The increase in thickness of the electrically insulating TA prevent  
255 further deposition as observed previously for catechol based molecules.<sup>6, 52, 53</sup> After 30 min of  
256 electrodeposition, the rinsing step using 0.15 M KPF<sub>6</sub> led to an increase in the frequency shift,  
257 which could be ascribed to the swelling of the coating. An additional cross-linking step was  
258 performed by the application of CV (20 cycles between 0 and 1.1 V at 50 mV/s scan rate) with  
259 no influence on the frequency shift but with a decrease in the peak current of TA oxidation

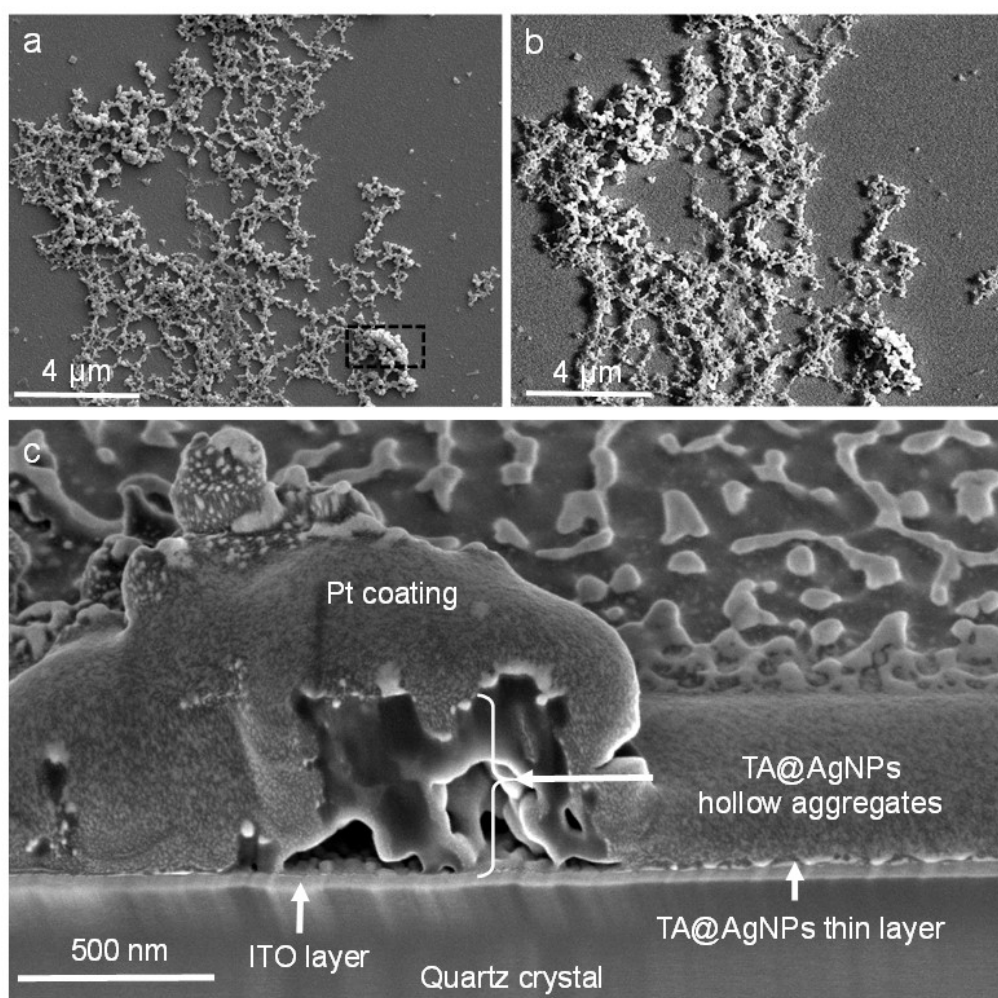
260 (Figure S7 in the SI). This means that sterically feasible cross-linking of TA occurred in the  
261 immobilized Al(III)/TA@AgNPs film.

262 The CV registered for Al(III)/TA@AgNPs film in 0.15 M KPF<sub>6</sub> (Figure 5c) exhibited a signal  
263 similar to TA-Al(III) complexes (Figure 2a), *i.e.* a large peak at 0.9 V, with an additional peak  
264 centered at ca. 0.6 V corresponding to TA@AgNPs (Figure 6). The deposition of a non-  
265 conductive layer of TA was expected to decrease the electroactive surface area of the electrode.  
266 In order to calculate the electroactive surface area and the coatings' capacity as a potential  
267 electrochemical sensor Randles-Sevcik equation<sup>54</sup> was used based on the position of the  
268 Fe<sup>2+</sup>/Fe<sup>3+</sup> redox pair in the ferro/ferricyanide solution (Figure S8 in the SI). The calculated  
269 electrochemical active surface was 0.66 cm<sup>2</sup> as compared to the geometric surface of 0.8 cm<sup>2</sup>,  
270 meaning a decrease of ca. 17.5 %. The incorporation of conductive AgNPs into the coating as  
271 well as metal ions allowed to lower this decrease.

272 **2.2.4. Chemical removal of Al(III) ions from Al(III)/TA@AgNPs films.** After the ED of  
273 Al(III)/TA@AgNPs coating, Al(III) ions were removed by soaking in 0.48 M NaF solution,  
274 prepared in 0.15 M KPF<sub>6</sub>. The principle of the removal was based on the formation of water-  
275 soluble Al-F complexes<sup>55-57</sup> with stability constants ( $\beta_n$ ) ranging from 7 to 19.8<sup>58, 59</sup> which are  
276 significantly higher than for the Al(III)-TA complexes ( $\beta_n = 5.25^{60}$ ). The injection of NaF  
277 solution resulted in an increase in the normalized frequency shift, probably due to the change  
278 of solvent bulk properties (viscosity and density) (Figure S9 in the SI). After the rinsing step  
279 using 0.15 M KPF<sub>6</sub> solution under a flow rate of 600  $\mu$ L/min, the stabilization of the signal was  
280 obtained at rest with no variation in the frequency value in comparison to before the contact  
281 with NaF. After the removal of Al(III) ions, the CV registered showed two typical irreversible  
282 oxidation peaks of TA bonded to a metal species at ca. 0.4 and 0.6 V corresponding to the  
283 AgNPs-TA bonds (Figure 5c). The signal ascribed previously to TA/Al(III) at ca. 0.9 V

284 disappeared which demonstrated that the removal of the Al(III) ions was successful and the  
285 film was robust upon removal of Al(III).

286 **2.2.5. Characterization of Al(III)/TA@AgNPs coatings.** The morphology of  
287 Al(III)/TA@AgNPs coatings was characterized by SEM (**Figure 6**). The surface of the coating  
288 exhibited irregularly distributed aggregates of comparable size. The phenomenon of strong  
289 backscattering of electrons (BSE) on heavy elements was used to follow the distribution of Ag  
290 over the surface area of Al(III)/TA@AgNPs coatings. Heavier elements appear brighter due to  
291 the higher intensity of the secondary electrons backscattered from the surface. The aggregates  
292 appeared significantly brighter than the rest of the surface when imaged in the LA-BSE mode  
293 which indicates the relatively high in Ag in the aggregates, as compared to the films deposited  
294 without the addition of AgNPs (**Figure S10 in the SI**).



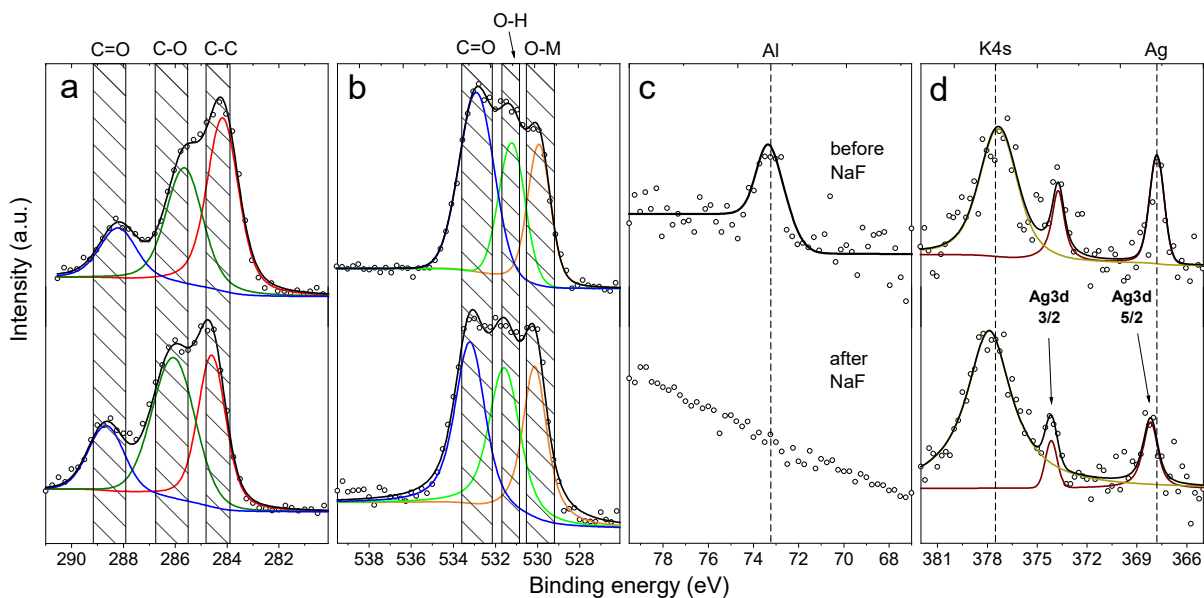
295



296 **Figure 6.** SEM images of Al(III)/TA@AgNPs coatings (a, b) top views obtained in SE (a)  
 297 and BSE (b) modes and (c) cross-section of the dashed rectangle area of image (a) observed  
 298 by FIB in SE mode after the deposition of the 200 nm Pt stripe.

299 The cross-sectional cut, performed with the focused ion beam (FIB) after the deposition of 200  
 300 nm platinum (Pt) layer onto the sample, revealed that the aggregates were partially hollow  
 301 structures of ca. 500 nm in height (**Figure 6c**). The granular surface under the hollow coatings  
 302 was attributed to the ITO layer grains of up to 50 nm in height. On the rest of the electrode  
 303 surface, a bright planar and thin coating was observed corresponding to a thin deposition of  
 304 Al(III)/TA@AgNPs.

305 XPS investigation allowed the determination of both atomic and chemical composition of the  
 306 coatings. In the XPS survey spectrum of Al(III)/TA@AgNPs coating carbon (C), oxygen (O),  
 307 silver (Ag) and aluminum were detected (**Figure S11** in the SI). The C1s peak at 285 eV (used  
 308 as an internal calibration peak) and O1s at peak 534 eV represented 50 and 48 at.%,  
 309 respectively, confirming the presence of TA, the only carbon-containing compound present in  
 310 the starting solution (**Figure 7**).



311

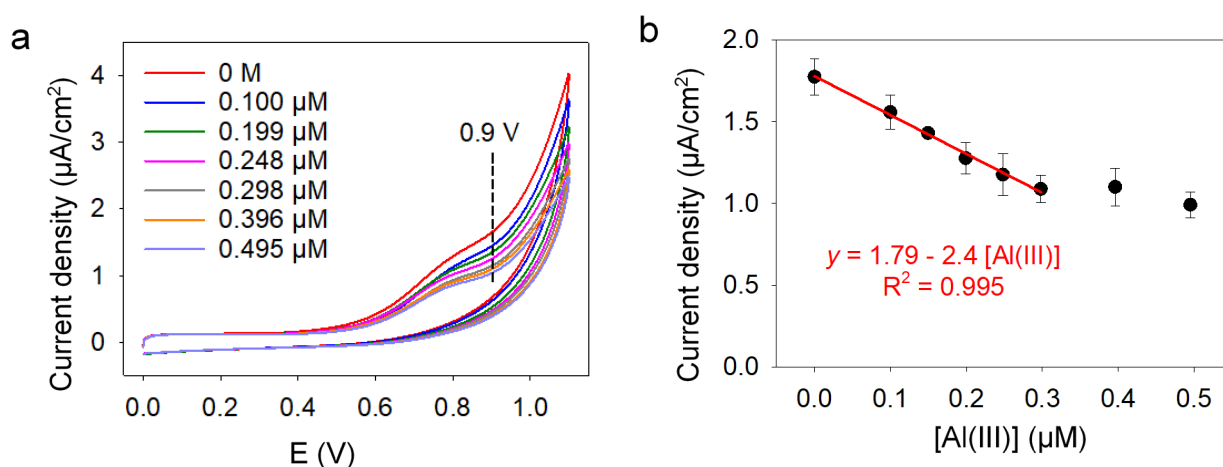
312 **Figure 7.** XPS (a) C1s, (b) O1s, (c) Al2p and (d) Ag3d core level spectra of Al(III)/TA@AgNPs  
313 coating obtained after 30 min deposition time before and after treatment with 0.48 M NaF  
314 solution.

315 The doublet in the Ag3d spectrum with the Ag3d<sub>5/2</sub> signal centered at ca. 368 eV corresponds  
316 to the metallic form of silver,<sup>61</sup> confirming the presence of AgNPs with an estimated atomic  
317 concentration of 0.9 at.%. In this region, a signal corresponding to K4s was deconvoluted as  
318 well, prompting that some content of the basic electrolyte persisted in the coatings after rinsing  
319 them with water. Al2p spectrum revealed a single signal at ca. 73 eV corresponding to metallic  
320 Al,<sup>61</sup> confirming the coordination character of the TA-Al bond with an atomic concentration of  
321 Al at 1.3 at.%.

322 After the chemical removal of Al(III), the Al2p signal in the XPS spectrum disappeared after  
323 the treatment with the fluoride solution, confirming the complete removal of Al(III) ions from  
324 the coating (**Figure 5c** and **Figure 7**). On the other hand, the intensity of the C1s component  
325 signal corresponding to carbon single-bonded to oxygen increased, confirming the release of  
326 Al(III) ions from a complex formed with the phenol groups. The atomic concentration of Ag  
327 remained at the same level before and after the removal of Al(III) ions.

328 **2.3. Al(III) sensing properties of TA@AgNPs coatings in physiological conditions.** After  
329 the complete removal of Al(III), the rebinding of Al(III) by TA@AgNPs coatings was first  
330 studied in HEPES-buffered 0.15 M NaCl solution at pH 7.4 (HEPES). **Figure S12** in the SI  
331 shows a typical cyclic voltammogram of TA@AgNPs coating registered in HEPES buffer. In  
332 contrast to the CV obtained in KPF<sub>6</sub> solution (**Figure 5c**), a sharp oxidation peak at ca. 0.12 V  
333 characteristic for AgNPs appeared because of the presence of NaCl.<sup>62, 63</sup> To move forward to a  
334 physiological medium, 10% of human serum (HS) was added to the HEPES buffer (labeled as  
335 HS-HEPES buffer).

336 While TA forms complexes with Al(III) that can be probed electrochemically,<sup>64</sup> it can also  
337 interact with proteins present in the human serum<sup>65</sup> and thus could interfere with the sensing of  
338 Al(III). To avoid this interference before each measurement, TA@AgNPs coatings in contact  
339 HS-HEPES buffer underwent the application of CV cycles until the stabilization of the CV  
340 signal (**Figure S10** in the SI). After this step, TA@AgNPs coating was put in contact for 5 min  
341 with Al(III) solutions, prepared at different concentrations in HS-HEPES buffer. To perform  
342 Al(III) sensing, a CV was performed between 0 and 1 V at 5 mV/s scan rate (**Figure 8a**).



343  
344 **Figure 8.** a) Cyclic voltammogram of TA@AgNPs coatings, registered at 5 mV/s in HS-  
345 HEPES buffer, in the presence of different concentrations of Al(III) ranging from 0 to 0.495  
346 µM. (b) Evolution of the current density, measured at 0.9 V in the CV, as a function of Al(III)  
347 concentration. The data represents the mean and standard deviation of three independent  
348 experiments. The point without an error bar presents an additional measurement of one of the  
349 independent experiment.

350 Only one oxidation signal can be distinguished spreading from ca. 0.5 to ca. 1 V, preceded by  
351 a current plateau and followed by a steep increase in the current magnitude corresponding to  
352 the continuous oxidation of the coating. After each sensing, TA@AgNPs coating was  
353 regenerated by NaF treatment and HS-HEPES soaking to be reused for the following Al(III)  
354 concentration. Following, after the rinsing step with HEPES buffer followed by removal of

355 Al(III) ions using NaF solution, another rinsing step with HEPES buffer and then with HS-  
356 HEPES. A decrease in the current density value of the oxidation wave was observed with the  
357 increase in Al(III) concentration. The difference in the current magnitude registered for a  
358 different level of Al(III) was the most visible at 0.9 V (**Figure 8a**, dashed line). The value of  
359 the current density measured at 0.9 V was then plotted as a function of Al(III) concentration to  
360 obtain a calibration curve. **Figure 8b** shows the evolution of the current density, mean and  
361 standard deviation of three independent experiments as a function of Al(III) concentration.

362 The curve showed a linear dependency ( $R^2 = 0.995$ ) in the concentration range of 0.10 to 0.298  
363  $\mu\text{M}$  in Al(III), which deviates from linearity at a higher concentration. The observed sensitivity,  
364 calculated from the slope of the calibration curve, was  $-2.4 \mu\text{A}/\text{mM cm}^2$  with a detection  
365 capacity of  $0.13 \mu\text{M}$  (limit of detection, at a signal-to-noise ratio of 3).<sup>66</sup> According to the  
366 literature, the toxic level of Al(III) in plasma (*i.e.* serum) is above  $2.3 \mu\text{M}$  for children and  $7.4$   
367  $\mu\text{M}$  for adults.<sup>67</sup> As the calibration curve was obtained in diluted human serum at 10%,  
368 TA@AgNPs coatings could be used on 10-fold diluted plasma samples of children to  
369 discriminate between Al(III) normal and toxic level, with a detection limit of  $1.3 \mu\text{M}$  and a  
370 linear range up to  $2.98 \mu\text{M}$  in the plasma.

371 **CONCLUSIONS:** A novel ionically-imprinted thin sensor film based on tannic acid and silver  
372 nanoparticles, tailor-designed for voltammetric tracing of aluminum ions in biological fluids  
373 was obtained via electrochemically assisted self-assembly onto ITO electrode. The modified  
374 colloidal synthesis of AgNPs via direct reduction by TA resulted in the fabrication of TA-  
375 stabilized nanoparticles of a significantly decreased size of a few nanometers, as compared to  
376 the literature. The XPS analysis revealed that the fabricated Al(III)/(TA@AgNPs) film  
377 contained ca. 2 at.% of Al, which could be completely removed from the layer by soaking in a  
378 fluoride solution, basing on the strong complexation of Al(III) by fluoride ions. The determined  
379 Ag content was ca. 1 at.% which persisted after the removal of Al(III) ions and the

380 corresponding BE value was assigned to Ag in the metallic form, proving the successful  
381 incorporation of AgNPs in situ during the self-assembly of the film. By chemical removal of  
382 Al(III) using NaF solution, ion-imprinted TA@AgNPs coating was obtained. A correlation  
383 between the current density value measured at 0.9 V and the concentration of Al(III) was  
384 established between 0.10 and 0.298  $\mu\text{M}$  (2.7-8.05  $\mu\text{g/L}$ ) with a LOD value of 0.13  $\mu\text{M}$ . These  
385 results show the high applicability of the electrochemically assisted self-assembly to produce  
386 complex supramolecular thin sensor films tailor-designed for the targeted metal ion sensing.  
387 This could contribute to the development of electrochemical sensors for the monitoring of  
388 Al(III) in 10-fold diluted plasma in particular for long-term dialysis patients.

389

### 390 **3. Material and methods**

391 **3.1 Materials.** Tannic acid (TA, A.C.S. reagent grade), potassium hexafluorophosphate ( $\text{KPF}_6$ ,  
392  $\geq 99\%$ ) and human serum (from human male AB plasma, USA origin, sterile-filtered) were  
393 purchased from Sigma-Aldrich. Aluminum nitrite ( $\text{Al}(\text{NO}_3)_3 \cdot 9\text{H}_2\text{O}$ , 98%), silver nitrite  
394 ( $\text{AgNO}_3$ , 99+%) and sodium fluoride (NaF, 99%) were purchased from Alfa Aesar. Potassium  
395 hexacyanoferrate (III) ( $\text{K}_4\text{Fe}(\text{CN})_6$ ) was acquired from Merck. 4-(2-hydroxyethyl)-1-  
396 piperazineethanesulfonic acid (HEPES, for molecular biology,  $\geq 99\%$ ), sodium chloride (NaCl,  
397 anal. reagent grade  $\geq 99.5\%$ ) and potassium chloride (KCl) grade were purchased from Fischer  
398 Scientific. All chemicals were used as received. All solutions were prepared using doubly  
399 distilled MilliQ water. All solutions used for electrodeposition were prepared in 0.15 M  $\text{KPF}_6$   
400 aqueous solutions adjusted at pH 7.4 with aqueous KOH solutions unless otherwise stated.

### 401 **3.2 Electrochemical Quartz Crystal Microbalance (QCM) with Dissipation Monitoring.**

402 QCM experiments were performed using a Q-Sense E1 apparatus from Q-Sense AB  
403 (Gothenburg, Sweden) by monitoring the changes in the resonance frequency  $f_v$  and the  
404 dissipation factor  $D_v$  of an oscillating quartz crystal upon adsorption of a viscoelastic layer ( $v$

405 represents the overtone number, equal to 1, 3, 5, 7). The quartz crystal was excited at its  
406 fundamental frequency (5 MHz), and the measurements were performed at the first, third, fifth,  
407 and seventh overtones, corresponding to 5, 15, 25, and 35 MHz, respectively. In each case, only  
408 the third overtone at 15 MHz is presented. QCM measurement is sensitive to the amount of  
409 water associated with the adsorbed molecules and senses the viscoelastic changes in the  
410 electrode/electrolyte interface. The indium tin oxide coated QCM (ITO-QCM, MicroVacuum  
411 Ltd., Budapest, Hungary) sensor acted as the working electrode. A platinum electrode (counter  
412 electrode) on the top of the electrochemical QCM cell and a no-leak Ag/AgCl reference  
413 electrode (Dri-Ref<sup>TM</sup>, World Precision Instruments, 3M KCl) fixed in the outlet flow channel  
414 was used as counter and reference electrodes, respectively. All potential values are given with  
415 respect to the Ag/AgCl reference electrode. Electrochemical measurements were performed  
416 using CHI660E apparatus from CH Instrument (Austin, Texas) coupled on the QCM-D  
417 apparatus. Before the buildup of the coating, the ITO-QCM crystal was cleaned by UV-ozone  
418 treatment for 15 min. The Ag/AgCl reference electrode was frequently tested by cyclic  
419 voltammetry (CV) in K<sub>4</sub>Fe(CN)<sub>6</sub> aqueous solution to ensure the potential stability and  
420 correctness. K<sub>4</sub>Fe(CN)<sub>6</sub> prepared in 0.15 M KPF<sub>6</sub> solution (pH 3.7) was routinely injected into  
421 the electrochemical QCM cell with the ITO crystal as the working electrode to monitor its cyclic  
422 voltammogram taken as reference. The presence of the two peaks and the potential values of  
423 redox reactions of the Fe(II)/Fe(III) redox pair were verified. A surface area of 0.8 cm<sup>2</sup>,  
424 corresponding to the exposed area of the ITO QCM sensor, was used for all current density  
425 calculations.

426 **3.3 Silver nanoparticles synthesis.** TA@AgNPs were obtained via direct reduction of AgNO<sub>3</sub>  
427 (1 mg/mL) dissolved in MilliQ water at 0.5 mg/mL by TA (10 mg/mL) corresponding to  
428 Ag(I)/TA molar ratio of 0.5. Following, the mixture was sonicated for 5 min at room

429 temperature to allow enough time for the Ag(I) reduction reaction and TA adsorption on the  
430 surface of the newly formed AgNPs while slowing down their aggregation rate.

431 **3.4 Buildup Procedure of Al(III)/TA coating.** The complexes of TA with Al(III) were  
432 obtained by mixing the TA solution with the same volume of 29  $\mu\text{M}$  Al(III) solution, both  
433 prepared in 0.15 M  $\text{KPF}_6$ , resulting in a mixture at Al(III)/TA molar ratio of 2.5. To ensure  
434 enough ionic conductivity of the building solutions, all the experiments of electrodeposition  
435 were performed in the presence of 0.15 M  $\text{KPF}_6$ . After the stabilization of the QCM signal in  
436 contact with 0.15 M  $\text{KPF}_6$  solution, Al(III)/TA@AgNPs suspension was injected into the  
437 electrochemical cell (600  $\mu\text{L}$ ) at a flow rate of 400  $\mu\text{L}/\text{min}$  with a peristaltic pump. After the  
438 stabilization of the signal, a constant potential of 0.7 V was applied for 30 min to trigger the  
439 crosslinking between TA molecules and start the electrodeposition of the coating. After the  
440 deposition, an aqueous solution of 0.15 M  $\text{KPF}_6$  (600  $\mu\text{L}$ ) was injected to rinse the coating.  
441 Then, the application of CV (20 cycles, 0-1.1 V at 50 mV/s in scan rate) was performed to  
442 obtain an additional cross-linking between the TA molecules of the coating to improve its  
443 mechanical stability.

444 **3.5 Buildup Procedure of Al(III)/TA@AgNPs coating.** The complexation of TA@AgNPs  
445 with Al(III) was performed by mixing the synthesized TA@AgNPs suspension with the same  
446 volume of 29  $\mu\text{M}$  Al(III) solution in 0.3 M  $\text{KPF}_6$  to obtain a final electrodeposition (ED)  
447 mixture at Al(III)/TA molar ratio of 2.5 in 0.15 M  $\text{KPF}_6$ . Next, the same procedure as in part  
448 3.4 was followed for the injection and deposition of the coating. After the cross-linking step, a  
449 0.48 M NaF solution, prepared in 0.15 M  $\text{KPF}_6$  solution, was injected in order to remove Al(III)  
450 ions from the coating and thus, create “holes” ready to bind the Al(III). After the stabilization  
451 of the QCM signal, the coating was rinsed by 0.15 M  $\text{KPF}_6$  for 2 min under flow. To ensure the  
452 complete removal of the Al(III) ions, CV monitoring after each injection of NaF solution was  
453 performed. Prior to further microscopic and spectroscopic characterization, MilliQ water was

454 injected into the cell after the buildup to rinse the coating and avoid any additional signals were  
455 present in the spectra of the coatings.

456 **3.6 Electroactive surface area of the coatings.** In order to calculate the electroactive surface  
457 area and the coatings' capacity as a potential electrochemical sensor Randles-Sevcik equation<sup>54</sup>  
458 was utilized (**Equation 1**) basing on the position of the Fe<sup>2+</sup>/Fe<sup>3+</sup> redox pair in the ferro-  
459 /ferricyanide solution (**Figure S7** in the SI).

$$460 \quad I_p = 0.4463 \times \left(\frac{F^3}{RT}\right)^{1/2} \times n^{3/2} \times A_{eff} \times D^{1/2} \times C \times v^{1/2} \quad (1)$$

461 where  $I_p$  is the peak current,  $F$  is the Faraday constant (96485.3 C/mol),  $R$  is the universal gas  
462 constant (8.31 J/Kmol),  $T$  is the temperature (293.15 K),  $n$  the number of moles of electrons  
463 transferred in the reaction (for ferrocyanide/ferricyanide  $n = 1$ ),  $A_{eff}$  is the active surface of the  
464 electrode (cm<sup>2</sup>),  $D$  the diffusion coefficient of electroactive species ( $0.72 \times 10^{-5}$  and  
465  $0.67 \times 10^{-5}$  cm<sup>2</sup>/s<sup>1</sup> for ferricyanide and ferrocyanide, respectively)<sup>68</sup>,  $C$  is the concentration of  
466 electroactive species (1.18 mM) and  $v$  is the potential sweep rate (50 mV/s).

467 **3.7 Voltammetric sensing of Al(III) in HS using TA@AgNPs coating.** Human serum  
468 solution was prepared at 10% in volume in 10 mM HEPES 0.15 M NaCl at pH 7.4, referred to  
469 as HS-HEPES. The pH value of HS-HEPES was adjusted at pH 7.4 with aqueous NaOH  
470 solution. After the removal of Al(III) by NaF treatment and a rinsing step of TA@AgNPs  
471 coating with 0.15 M KPF<sub>6</sub>, 500  $\mu$ l of HS-HEPES solution was injected into the QCM cell before  
472 the application of CV (10 cycles at 50 mV/s sweep rate) to satiate the coatings with proteins  
473 from HS. In the following, Al(III) solution prepared in HS-HEPES was injected in the QCM  
474 cell and a CV scan at 5 mV/s in scan rate was recorded for Al(III) sensing. After each sensing,  
475 TA@AgNPs coating was regenerated by NaF treatment and HS-HEPES soaking to be reused  
476 for the following Al(III) concentration.



477 **3.8 Spectroscopic characterization of the coatings.** X-ray Photoelectron Spectroscopy (XPS)  
478 measurements were performed using a VG ESCALAB 210 system equipped with a  
479 monochromatized Al K $\alpha$  ( $h\nu=1486.6$  eV) radiation source with the pass energy of 20 eV used  
480 for all narrow scan measurements. The photoemission spectra were recorded at normal emission  
481 with an overall resolution of about 0.6 eV. The energy positions of the spectra were calibrated  
482 with reference to the 4f $_{7/2}$  level of a clean gold sample at 84.0 eV binding energy (BE) value.  
483 Fitting of the data was performed using Doniach-Sunjic functions,<sup>69</sup> after a Shirley background  
484 fitting<sup>70</sup> with the help of UNIFIT 2016 software.<sup>71</sup> A convolution of Lorentzian and Gaussian  
485 line shapes was used to fit the individual peaks. After this, the intensities were estimated by  
486 calculating the integral of each peak. The atomic concentrations were then derived using  
487 Scofield sensitivity factors<sup>72</sup>. No charging of the surface during the measurement was observed.

488 **3.9 Dynamic light scattering.** Size distribution histograms of AgNPs were recorded using  
489 dynamic light scattering (DLS) method on Zetasizer Nano ZS (Malvern Instruments Ltd., GB)  
490 in 12 mm disposable polystyrene cuvettes (DTS0012, Brand, Germany). The measurements  
491 were performed at an ambient temperature of 25°C after 30 s of equilibration time with 0.135  
492 and 3.990 as the refractive index and adsorption coefficient of AgNPs, respectively.<sup>73</sup> The  
493 viscosity for the dispersant (water) was set to 0.8872 cP and the refractive index to 1.330.

494 **3.10 Microscopic characterization.** Scanning electron microscopy (SEM) images were  
495 registered with a Hitachi S-4800 microscope in secondary electron (SE) and back-scattered  
496 secondary electron (LA-BSE) mode. To obtain the cross-section of the coating, NanoLab 600  
497 Helios (FEI company) device, which is a combined SEM and focused ion beam (FIB) facility  
498 using 30 keV Ga<sup>+</sup> ions for cutting, was used. A layer of 200 nm platinum (Pt) was used to  
499 protect the sample surface at the cross-section edge. For the transmission electron micrographs,  
500 5  $\mu$ l of the nanoparticles suspension (diluted by a factor of 3) was deposited onto a freshly glow  
501 discharged carbon-covered grid (400 mesh). The suspension was left for 2 minutes and then

502 dried using a filter paper. The grids were observed at 200 kV with a Technai G2 (FEI)  
503 microscope. Images were acquired with an Eagle 2k (FEI) ssCCD camera.

#### 504 ASSOCIATED CONTENT

505 **Supporting Information.** Normalized frequency shift and ED current registered during the  
506 deposition of Al(III)-TA films onto ITO electrode; Cyclic voltammograms of ITO electrode in  
507 KPF<sub>6</sub> and Al(III)/KPF<sub>6</sub> solutions; TA@AgNPs size distribution determined by DLS in water  
508 directly after the synthesis and at the ED time; Cyclic voltammogram of Al(III)/TA@AgNPs  
509 coating electrodeposited at the cross-linking step after electrodeposition; Cyclic  
510 voltammograms of K<sub>2</sub>Fe(CN)<sub>6</sub> before and after the electrodeposition of the Al(III)/TA@AgNPs  
511 film on ITO-QCM electrode; SEM top-view images of Al(III)/TA coatings deposited onto ITO  
512 surface obtained in SE and BSE modes; XPS survey spectra of Al(III)/(TA@AgNPs) film  
513 before and after NaF treatment.

514

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522 The manuscript was written through contributions of all authors.

#### 523 ACKNOWLEDGMENT

524 The authors acknowledge the platform of characterization of ICS for the use of the Dynamic  
525 Light Scattering. AKC acknowledges Initiative of Excellence (Idex) Post doctorants from  
526 Université de Strasbourg and Région Grand Est « Jeunes Chercheurs » for financial support.  
527 The authors acknowledge Swiss Federal Office for Education and Science (grant no.  
528 1315001762), Swiss National Science Foundation (SNF) and Swiss Nanoscience Institute  
529 (SNI) for their financial support. This work of the Interdisciplinary Institute HiFunMat, as part  
530 of the ITI 2021-2028 program of the University of Strasbourg, CNRS and Inserm, was  
531 supported by IdEx Unistra (ANR-10-IDEX-0002) and SFRI (STRAT'US project, ANR-20-  
532 SFRI-0012) under the framework of the French Investments for the Future Program.

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652 higher wavenumbers with increasing electrode potential, but the SERS intensity of this band  
653 is significantly diminished when leaching commences. In cyanide solutions, the presence of  
654 specifically adsorbed cyanide ions at the electrode surface at the lower potential limit was  
655 indicated by Stark-shifted Raman bands characteristic of carbon-nitrogen stretching. Bond  
656 formation between cyanide and gold, evidenced by vibrational modes due to gold-carbon  
657 stretching and gold-carbon-nitrogen bending, commenced near the potential at which  
658 voltammetric currents consistent with gold dissolution were observed. At high potentials,  
659 cyanide was displaced from the surface and a Raman band characteristic of gold-oxygen  
660 stretching appeared. Oxidation of cyanide to cyanate was also indicated in this region by a  
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