

Article

Genomic Landscape, Clinical Features and Outcomes of Non-Small Cell Lung Cancer Patients Harboring *BRAF* Alterations of Distinct Functional Classes

Alessandro Di Federico ^{1,2,*} , Andrea De Giglio ^{1,2} , Francesco Gelsomino ^{1,2} , Dario De Biase ^{2,3} ,
Francesca Giunchi ⁴, Arianna Palladini ⁵ , Francesca Sperandi ^{1,2}, Barbara Melotti ^{1,2}  and Andrea Ardizzoni ^{1,2}

¹ Division of Medical Oncology, IRCCS Azienda Ospedaliero-Universitaria di Bologna, 40138 Bologna, Italy; dr.degiglio@gmail.com (A.D.G.); francesco_gelsomino@aosp.bo.it (F.G.); francesca.sperandi@aosp.bo.it (F.S.); barbara.melotti@aosp.bo.it (B.M.); andrea.ardizzoni2@unibo.it (A.A.)

² Department of Experimental, Diagnostic and Specialty Medicine (DIMES), University of Bologna, 40126 Bologna, Italy; dario.debiase@unibo.it

³ Department of Pharmacy and Biotechnology (FaBiT), University of Bologna, 40138 Bologna, Italy

⁴ Pathology Department, IRCCS Azienda Ospedaliero-Universitaria di Bologna, 40138 Bologna, Italy; francesca.giunchi@aosp.bo.it

⁵ Department of Molecular Oncology, University of Pavia, 27100 Pavia, Italy; arianna.palladini@unipv.it

* Correspondence: alessandrofederico1@gmail.com; Tel.: +39-3933051534

Simple Summary: Non-small cell lung cancer (NSCLC) patients harboring *BRAF* non-V600 alterations constitute a heterogeneous and poorly studied population orphan of targeted therapies. We conducted a systematic review to detect all *BRAF* alterations of defined functional class across different cancer types. Then, we searched for NSCLC patients harboring these alterations in the cancer bioportal and in POPLAR and OAK trials using patient-level data, to investigate clinical and genomic differences associated with each *BRAF* functional class and the prognostic impact of *BRAF* non-V600 mutations. We found that NSCLC patients harboring distinct classes of *BRAF* alterations have different clinical characteristics, clinical features and genomic landscape. Moreover, *BRAF* non-V600 alterations were associated with a poor prognostic impact, apparently regardless of the treatment received. These peculiar features may suggest the use of tailored treatments according to each class of *BRAF* alteration.

Abstract: Background: In non-small cell lung cancer (NSCLC), *BRAF* class 1 alterations are effectively targeted by *BRAF* inhibitors. Conversely, targeted therapies have very low or absent activity in patients carrying class 2 and 3 alterations. The spectrum of *BRAF* alterations in NSCLC patients, and their accompanying clinical features, genomic landscape and treatment outcomes have been poorly reported. Patients and methods: We identified *BRAF* alterations of defined functional class across different tumors through a systematic review. Then, we selected NSCLC patients carrying *BRAF* alterations, according to the systematic review, in the cBioPortal (cBioPortal cohort) to collect and analyze clinical, biomolecular and survival data. Finally, we identified NSCLC patients carrying *BRAF* non-V600 mutations enrolled in POPLAR and OAK trials (POPLAR/OAK cohort), extracting clinical and survival data for survival analyses. Results: 100 different *BRAF* non-V600 alterations were identified through the systematic review. In the cBioPortal cohort ($n = 139$), patients harboring class 2 and 3 alterations were more frequently smokers and had higher tumor mutational burden compared to those carrying class 1 alterations. The spectrum of most frequently co-altered genes was significantly different between *BRAF* alterations classes, including SETD2, STK11, POM121L12, MUC16, KEAP1, TERT, TP53 and other genes. In the POPLAR/OAK cohort, patients carrying non-V600 *BRAF* alterations were characterized by poor prognosis compared to *BRAF* wild-type patients. Conclusions: Different classes of *BRAF* alterations confer distinctive clinical features, biomolecular signature and disease behavior to NSCLC patients. Non-V600 alterations are characterized by poor prognosis, but key gene co-alterations involved in cancer cell survival and immune pathways may suggest their potential sensitivity to tailored treatments.



Citation: Di Federico, A.; De Giglio, A.; Gelsomino, F.; De Biase, D.; Giunchi, F.; Palladini, A.; Sperandi, F.; Melotti, B.; Ardizzoni, A. Genomic Landscape, Clinical Features and Outcomes of Non-Small Cell Lung Cancer Patients Harboring *BRAF* Alterations of Distinct Functional Classes. *Cancers* **2022**, *14*, 3472. <https://doi.org/10.3390/cancers14143472>

Academic Editor: Tatsuya Nagano

Received: 22 June 2022

Accepted: 15 July 2022

Published: 17 July 2022

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

Keywords: non-small cell lung cancer; BRAF; immunotherapy; survival; genomic; non-V600

1. Introduction

Non-small cell lung cancer (NSCLC) represents the primary cause of cancer-related deaths worldwide [1]. Recent treatment advances allowed significant extension of the life expectancy of patients diagnosed with locally advanced or metastatic disease. The advent of immunotherapy in non-oncogene addicted NSCLC approximately doubled the median survival, while targeted therapies revolutionized the therapeutic approach for patients carrying actionable oncogenic drivers [2–4]. *EGFR* mutations and *ALK* and *ROS1* rearrangements represent the first efficaciously druggable gene alterations in NSCLC [4]. However, more recently, new agents have been found to effectively target other specific molecular alterations such as *BRAF*, *KRAS*, *MET*, *HER2*, *RET* and *NTRK* [5]. Somatic *BRAF* alterations occur in approximately 2–4% of patients with NSCLC, and the V600E mutation has been reported to represent almost half of them [6]. Female sex and smoking history have been described most frequently in NSCLC patients harboring *BRAF* mutations [7,8]. *BRAF* alterations have been classified into three functional classes: class 1 alterations, represented by p.V600X mutations, are characterized by strong activity of BRAF kinase domain and constitutive activation of the MAPK pathway; class 2 alterations, with intermediate to high activity of BRAF kinase domain, activating RAS-independent signaling as dimers; class 3 alterations, characterized by low or complete lack of BRAF kinase domain activity and RAS dependence [9]. Agents targeting BRAF and MEK demonstrated their efficacy in NSCLC patients harboring class 1 *BRAF* mutations, and their use has been recently approved by most regulatory agencies [10,11]. On the contrary, patients whose tumors harbor *BRAF* alterations of class 2 or 3 are currently treated as non-oncogene addicted, since BRAF/MEK inhibitors demonstrated absent or very low activity [12]. However, prevalence, clinical features and treatment outcomes of class 2 and 3 *BRAF* alterations in patients affected by lung cancer are still poorly studied. Previous data suggest that tumors harboring *BRAF* alterations of different classes have distinct clinical characteristics, natural history of disease, and may show different responses to various available treatments. We hypothesized that a distinct molecular landscape might explain those differences, suggesting particular disease features and treatment outcomes. Herein, we first report a systematic review of the literature aiming to identify all *BRAF* gene alterations belonging to a defined functional class across different cancer types. Second, based on the results of the systematic review, we searched for all NSCLC patients harboring *BRAF* alterations of defined functional class in the cBioPortal, with the aim to analyze and compare genomic and clinical features of patients harboring distinct classes of *BRAF* alterations (cBioPortal cohort). Finally, we explored the prognostic impact of BRAF non-V600 mutations on the outcomes of patients enrolled into two randomized controlled trials, the phase II POPLAR trial and the phase III OAK trial (POPLAR/OAK cohort), which demonstrated the superiority of atezolizumab 1200 mg over standard chemotherapy with docetaxel 75 mg/m² in previously treated, squamous or non-squamous, advanced NSCLC patients [13–15]. POPLAR and OAK trials were selected for the availability of patient-level and mutation data [15].

2. Materials and Methods

2.1. Research Strategies

Papers published before 10 June 2021 reporting non-V600 *BRAF* alterations and their corresponding functional class (2 or 3) across all cancer types were searched through the online databases MEDLINE (PubMed) and Cochrane Database of Systematic Reviews and Central Register of Controlled Trials (Wiley, Hoboken, NJ, USA). Records from the Clinical Interpretation of Variants in Cancer (CIViC) were also searched.

Key words used for the research were: “BRAF”; “class”; “type”; “2”; “II”; “3”; “III”; “non-V600”.

Only articles published in peer-reviewed journals and written in the English language were considered.

Studies were retrieved and reviewed by two different authors.

Records underwent a first screening for title and/or abstract. Relevant articles were subsequently screened for full text and analyzed to identify those reporting *BRAF* non-V600 alterations with their respective functional class. Articles reporting non-V600 *BRAF* mutations that were already listed through previously screened papers or CIViC database were excluded. Articles not reporting the corresponding functional class of the described non-V600 *BRAF* mutation(s) were also excluded. The bibliography of each relevant article was finally searched.

The Preferred Reporting Items for Systematic Review and Meta-Analyses (PRISMA) guidelines were adopted to conduct this work (Supplementary Figure S1).

2.2. Study Population

For the cBioPortal cohort, all datasets including NSCLC patients harboring a *BRAF* gene alteration, including mutations, structural variants and copy number alterations (CNA), were searched in the cBioPortal [16,17]. NSCLC patients were subsequently classified based on the functional class of *BRAF* alteration, according to the results of the systematic review, into class 1, class 2, class 3 or unknown functional class. Data about clinical characteristics, genomic landscape, and overall survival (OS) for each patient derived from available online datasets were retrieved from the cBioPortal. For the POPLAR/OAK cohort, patient-level data of participants harboring *BRAF* mutations (all non-V600) were extracted from the available online dataset [15]. Data about clinical characteristics and OS were collected.

2.3. Statistical Analysis

Continuous and categorical variables were described as median values and proportions. T-test (or ANOVA or Pearson correlation test or Kruskal–Wallis test if needed) and chi2-test (or Fisher’s exact test, if needed) were performed to compare means and proportions. Shapiro test was performed to verify the normality of data distribution for each variable of interest. A p -value ≤ 0.05 was considered statistically significant. The Kaplan–Meier method was used to estimate median survival times. The log-rank test was used to compare survival outcomes. For the POPLAR/OAK cohort, OS was defined as the time from treatment initiation (docetaxel or atezolizumab) to death from any cause [15]. For the cBioPortal cohort, the definition of OS may vary depending on the study analyzed. Top 50 concurrently altered genes in each cohort of NSCLC (*BRAF* alterations of a known functional class; class 1; class 2; class 3) were retrieved from cBioPortal gene expression data. Mutual exclusiveness of top 50 concurrent gene alterations in each cohort was identified with the Fisher’s exact test, confirmed through the Benjamini–Hochberg false discovery rate (FDR) correction procedure expressed as q -values. Differential expression of top 50 co-altered genes in each class was measured among three *BRAF* functional classes. Genes differently expressed were identified as those meeting the expression fold-change threshold of absolute value greater than 2 and p value ≤ 0.05 . p -values were adjusted for multiple hypothesis testing via the FDR method. Statistical analyses were performed using RStudio Version 1.3.1093 and the cBioPortal online platform. The following R packages were used: ggplot2; ggrepel; ggstatsplot; DescTools; finalfit; dplyr; knitr; survival; EnancedVolcano; ggsvplot; survival.

3. Results

3.1. Systematic Review

A systematic review of the literature was performed to identify all *BRAF* gene alterations in cancer for which the corresponding functional class was reported.

The initial database search yielded a total of 5977 records. Through reviewing titles and abstracts of each article, 5907 records were excluded as they did not report non-V600 *BRAF*

alterations. Full texts of 70 remaining articles were accurately reviewed and analyzed. In total, 53 articles were excluded as they did not report alterations' functional class or reported already collected variants, in order to avoid duplicates. A total of 17 articles were finally included in the bibliography [9,18–33]. In total, 27 different non-V600 *BRAF* alterations were listed through the CIViC database [34]. Further, 73 different non-V600 *BRAF* alterations were collected by the 17 selected articles, accounting for a total of 100 different non-V600 *BRAF* alterations listed with the corresponding functional class (Supplementary Table S1).

3.2. Study Population

3.2.1. Clinical and Molecular Features and Survival Outcomes

For the cBioPortal cohort, 25 studies including patients with NSCLC were identified in the cBioPortal (Figure 1A) [35–50]. In total, 4065 patients with a total of 4658 samples were included. Of them, 3132 patients (3553 samples) had adenocarcinoma histology (LUAD), 590 patients (725 samples) had squamous cell histology (LUSC), while the remaining patients had other or not specified histological subtype.

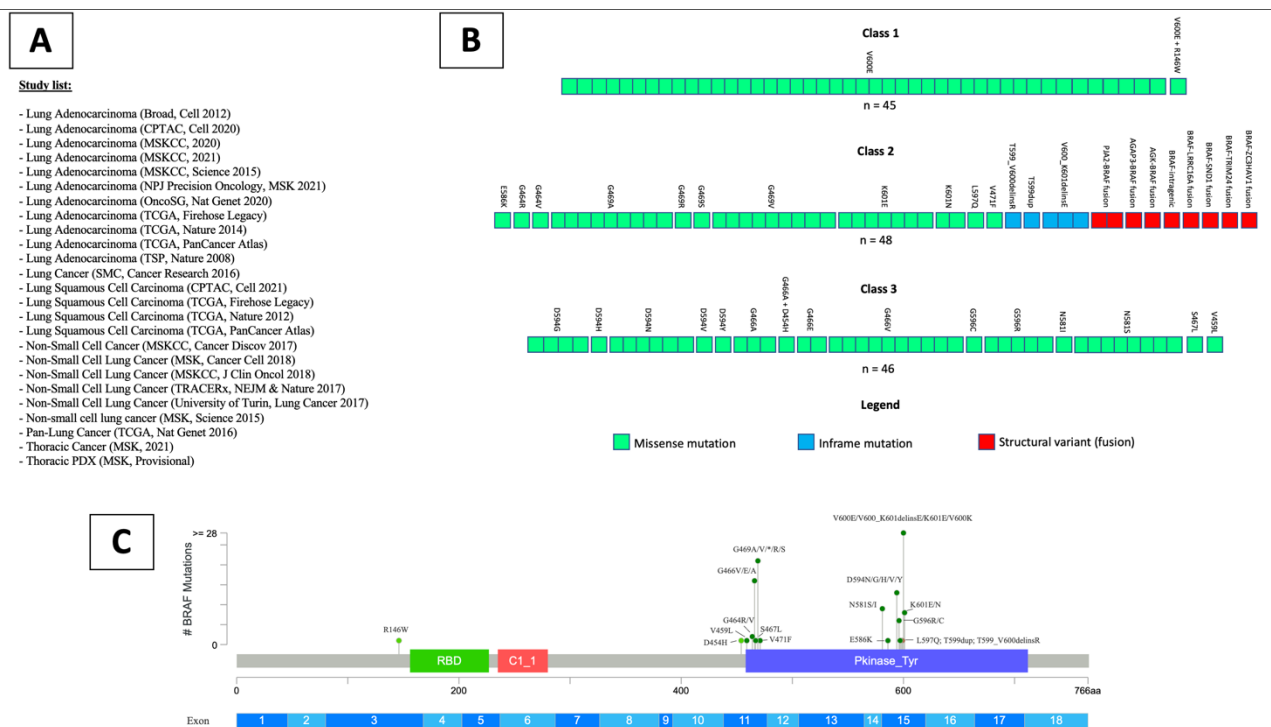


Figure 1. List of studies analyzed through the cBioPortal online platform containing available clinical and genomic data (A). Representation of patients (squares) analyzed for each class of BRAF alterations, and corresponding BRAF alteration detected (B). Lollipop plot showing the position of detected BRAF class 1, 2 and 3 mutations in the BRAF gene sequence (C).

BRAF gene alterations were identified in 236/3983 (5.92%) profiled patients with NSCLC. Mutations were found in 198 of 226 (87.6%) patients profiled for mutations, amplifications and deep deletions were found in 31 (14.0%) and 2 (0.9%) of 221 patients profiled for copy number alterations (CNA), and gene fusions were found in 9 of 226 (4.0%) patients profiled for structural variants. In total, 10 NSCLC patients were not profiled for mutations or fusions, and 15 were not profiled for amplifications or deletions. A concurrent *BRAF* mutation and amplification was found in 4 of 221 (1.8%) profiled NSCLC patients. In total, 4 of 226 (1.8%) profiled NSCLC patients had two concurrent different *BRAF* mutations. No patients with *BRAF* fusion had concurrent *BRAF* amplification or mutation. The prevalence of *BRAF* alterations was 6.44% (197/3057) in LUAD and 3.6% (21/583)

in LUSC. The remaining patients had other or not specified NSCLC histology subtype (Supplementary Figure S2).

According to the results of the systematic review, 45 (1.13%) NSCLC patients (all LUAD) had a *BRAF* class 1 alteration, 48 (1.21%) patients (of whom 45 LUAD) had a class 2 alteration, and 46 (1.15%) patients (of whom 43 LUAD) had a class 3 mutation (Figure 1B,C). The remaining 97 (2.44%) patients had a *BRAF* alteration of unknown functional class, including mutations, splice site variants and CNA (Supplementary Figure S2). Out of the 236 patients harboring *BRAF* alterations, 205 (86.9%) had LUAD histology, 118 (50%) were female and the mean age at diagnosis was 66 (95% CI, 65–68) (Table 1).

Table 1. Main clinical characteristics of patients harboring *BRAF* alterations detected through the analysis of the cBioPortal.

Clinical Characteristics	Total (<i>n</i> = 236)	Class 1 (<i>n</i> = 45)	Class 2 (<i>n</i> = 48)	Class 3 (<i>n</i> = 46)	Undefined Class (<i>n</i> = 97)	<i>p</i> Value
Age, Mean (SD)	66.0 (9.4)	66.6 (10.6)	68.5 (9.6)	65.7 (8.8)	64.8 (8.9)	0.243
Sex, <i>n</i> (%)						
Female	118 (51.5)	30 (68.2)	22 (47.8)	27 (58.7)	39 (41.9)	0.023
Male	111 (48.5)	14 (31.8)	24 (52.2)	19 (41.3)	54 (58.1)	
Histology, <i>n</i> (%)						
Adenocarcinoma	205 (86.9)	45 (100.0)	43 (89.6)	41 (89.1)	76 (78.4)	0.005
Squamous	23 (9.7)	0 (0)	2 (4.2)	3 (6.5)	18 (18.6)	
Non-Small Cell Lung Cancer NOS	8 (3.4)	0 (0)	3 (6.2)	2 (4.3)	3 (3.1)	
Geographical origin, <i>n</i> (%)						
Caucasian	65 (73.9)	5 (45.5)	13 (65.0)	12 (100.0)	35 (77.8)	0.059
Asian	17 (19.3)	5 (45.5)	6 (30.0)	0 (0)	6 (13.3)	
African	6 (6.8)	1 (9.1)	1 (5.0)	0 (0)	4 (8.9)	
Smoking habit, <i>n</i> (%)						
Yes	146 (83.4)	21 (56.8)	31 (93.9)	36 (92.3)	58 (87.9)	<0.001
No	29 (16.6)	16 (43.2)	2 (6.1)	3 (7.7)	8 (12.1)	

NOS: not otherwise specified.

According to known smoking habit (175 patients), 146 (83.4%) patients were current or former smokers. Smoking habit was significantly more common among patients harboring class 2 and class 3 alterations than in those with class 1 alterations (*p* for class 1 vs. class 2 or 3 = 0.003), while no difference was documented between patients with class 2 and class 3 alterations (Table 1). Consistently, pack-year was significantly lower in patients with class 1 alterations as compared to those harboring class 3 ones (*p* = 0.035) (Figure 2A). In this cohort (cBioPortal), no statistically significant difference in terms of overall survival (OS) was documented between a total of 71 patients of any disease stage and available survival data harboring class 1 (median OS: 37 months), 2 (median OS: 39 months) and 3 (median OS: 53 months) *BRAF* alterations (*p* = 0.482; Figure 2E).

For the POPLAR/OAK cohort, 35 patients with previously treated metastatic NSCLC harboring *BRAF* mutations were identified in the POPLAR (*n* = 7) and OAK (*n* = 28) trials (Figure 2C). All of them had *BRAF* non-V600 mutations (12 had class 2 mutations, 10 had class 3 mutations and 13 had *BRAF* mutations of undefined functional class). Patients' characteristics were consistent with that of the cBioPortal cohort, as they had a mean age of 64 years and almost all of them were previous or current smokers (Table 2).

Out of 35 patients analyzed for survival, 20 (57%) received atezolizumab and 15 (43%) received docetaxel. Patients harboring *BRAF* non-V600 mutations had significantly shorter OS compared to *BRAF* wild-type patients. Median OS was 8.4 (4.6–11.2) months in *BRAF* non-V600 mutated patients versus 11.5 (10.3–12.6) months in *BRAF* wild-type patients (HR: 1.70; 95% CI, 1.19–2.44; *p* = 0.0033) (Figure 2C). No significant OS differences were observed between *BRAF* non-V600 mutant patients treated with atezolizumab or docetaxel (HR: 0.84; 95% CI, 0.41–1.72; *p* = 0.63) (Figure 2D).

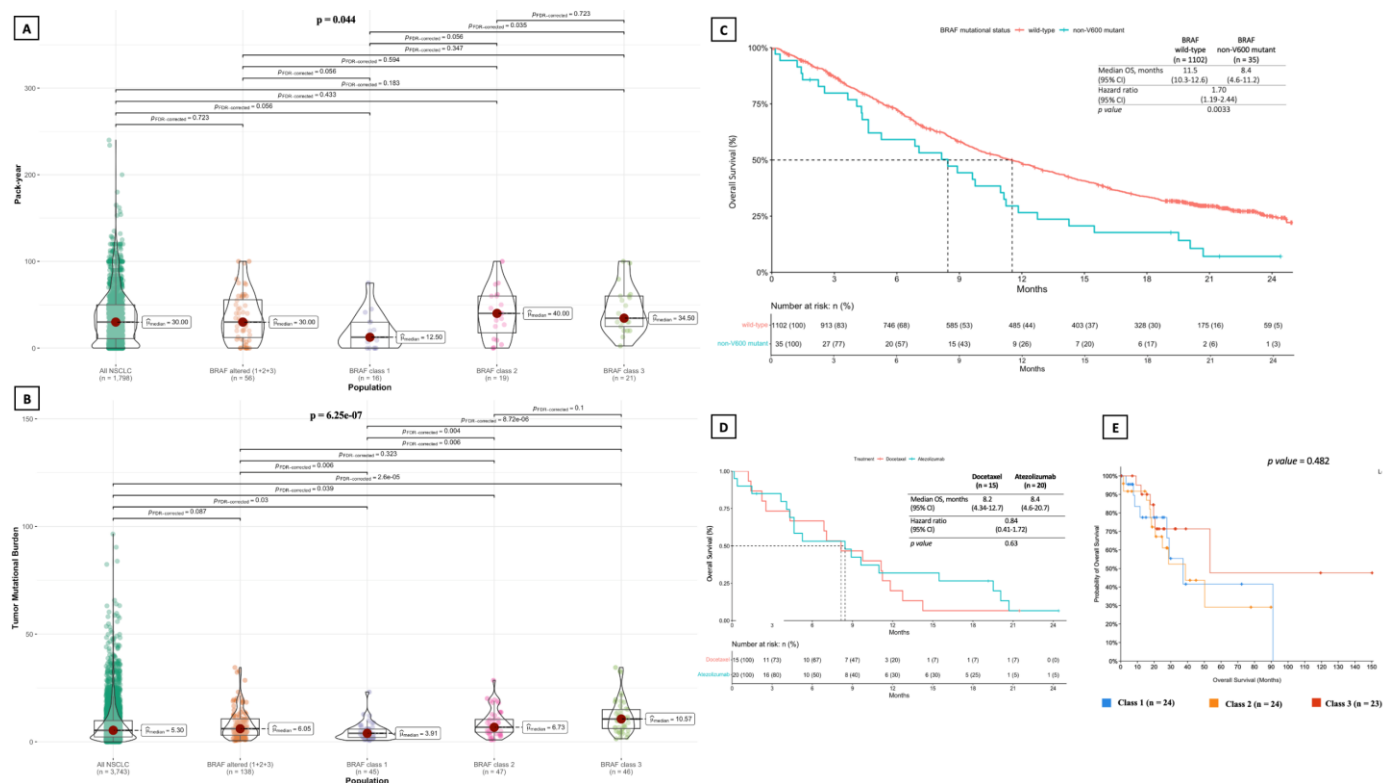


Figure 2. Comparison of smoking habit measured in pack-year among the three BRAF functional classes of alterations, showing a significantly greater pack-year value in patients harboring class 2 and class 3 BRAF alterations as compared to those with class 1 alterations (A). Consistently, tumor mutational burden (TMB) was significantly higher in patients harboring class 2 and class 3 BRAF alterations as compared to those with class 1 alterations (B). No statistically significant difference in terms of median TMB was found between patients harboring BRAF alterations of known functional class and all patients with NSCLC in the cBioPortal cohort. Kaplan–Meier of overall survival of patients with distinct BRAF alteration classes shows no statistically significant differences between classes in the POPLAR/OAK cohort (C), regardless of the treatment type (docetaxel or atezolizumab) (D). No statistically significant differences were found in terms of overall survival in patients harboring different classes of BRAF alterations in the cBioPortal cohort (E). TMB: tumor mutational burden; NSCLC: non-small cell lung cancer.

Table 2. Main clinical characteristics of patients harboring BRAF mutations in POPLAR/OAK cohort.

Clinical Characteristics	Total (n = 35)	Class 2 (n = 12)	Class 3 (n = 10)	Undefined Class (n = 13)	p Value
Age, Mean (SD)	64.1 (9.2)	65.1 (7.5)	65.4 (5.7)	62.1 (12.5)	0.415
Sex, n (%)					
Female	14 (40.0)	6 (50.0)	3 (30.0)	5 (38.5)	0.581
Male	21 (60.0)	6 (50.0)	7 (70.0)	8 (61.5)	
Histology, n (%)					
Adenocarcinoma	26 (74.3)	10 (83.3)	9 (90.0)	7 (53.8)	0.091
Squamous	9 (25.7)	2 (16.7)	1 (10.0)	6 (46.2)	
Geographical origin, n (%)					
Caucasian	27 (77.1)	9 (75.0)	7 (70.0)	11 (84.6)	0.669
Asian	6 (17.1)	2 (16.7)	2 (20.0)	2 (15.4)	
Other	2 (5.7)	1 (8.3)	1 (10.0)	0 (0)	
Smoking habit, n (%)					
Yes	33 (94.3)	12 (100)	9 (90.0)	12 (92.3)	0.431
No	2 (5.7)	0 (0)	1 (10.0)	1 (7.7)	

3.2.2. Concurrent Molecular Alterations

Among all NSCLC patients with *BRAF* alterations of any of the three functional classes ($n = 139$), *TP53* (75/139, 54%), *CSMD3* (25/53, 47%) and *TTN* (24/53, 45%) represented the most frequently co-altered genes (Figure 3).

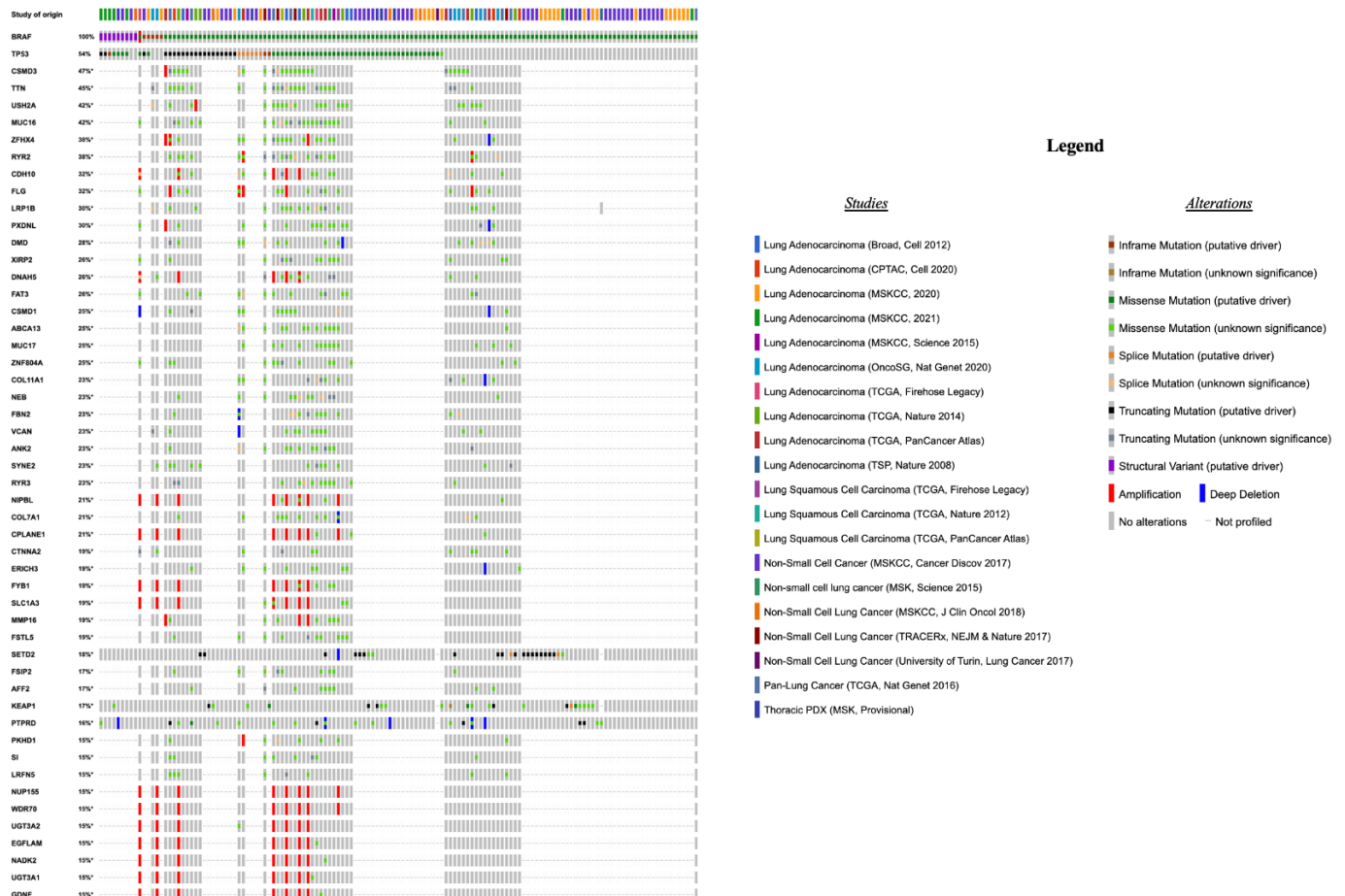


Figure 3. Top 50 most frequently altered genes in patients ($n = 139$) harboring *BRAF* alterations of any defined functional class, the type of alteration, and the study of origin of each patient.

SETD2 (20/45, 44%), *CSMD3* (6/15, 40%) and *TP53* (17/45, 38%) gene co-alterations were the most common in NSCLC patients with *BRAF* class 1 alterations ($n = 45$) (Figure 4A), while *TP53* (28/48, 58%), *TTN* (9/18, 50%) and *CSMD3* (9/18, 50%) were the most commonly co-altered genes in patients with *BRAF* class 2 alterations ($n = 48$) (Figure 4B). Finally, among NSCLC patients with *BRAF* class 3 alterations ($n = 46$), the most frequent co-altered genes were *MUC16* (14/20, 70%), *TP53* (30/46, 65%), *ZFH4* (13/20, 65%) and *TTN* (12/20, 60%) (Figure 4C).

Concurrent gene alterations showed significant heterogeneity among the three *BRAF* functional classes (Figure 5C–E). In fact, 47 of the most commonly co-altered genes (top 50) and key genes in each class showed statistically significant different co-alteration frequency between the three functional classes of *BRAF* alterations, including *SETD2* ($p < 0.0001$), *STK11* ($p = 0.0002$), *POM121L12* ($p = 0.001$), *MUC16* ($p = 0.002$), *OVCH1* ($p = 0.003$), *ZFH4* ($p = 0.004$), *ITGA4* ($p = 0.004$), *KEAP1* ($p = 0.005$), *TERT* ($p = 0.002$), *RAS* ($p = 0.006$), *TP53* ($p = 0.024$), *FGFR1/2/3/4* ($p = 0.042$), *ALK* ($p = 0.047$) and DNA damage response and repair (DDR) genes ($p = 0.049$) (Supplementary Table S2). A statistically significant mutual exclusivity in patients with *BRAF* alteration of any class was documented between *STK11* and either *TP53* ($p = 0.011$; $q = 0.044$) or *TTN* ($p = 0.004$; $q = 0.023$) co-alterations, and between *PIK3CA* and *XIRP2* co-alterations ($p = 0.009$; $q = 0.039$).

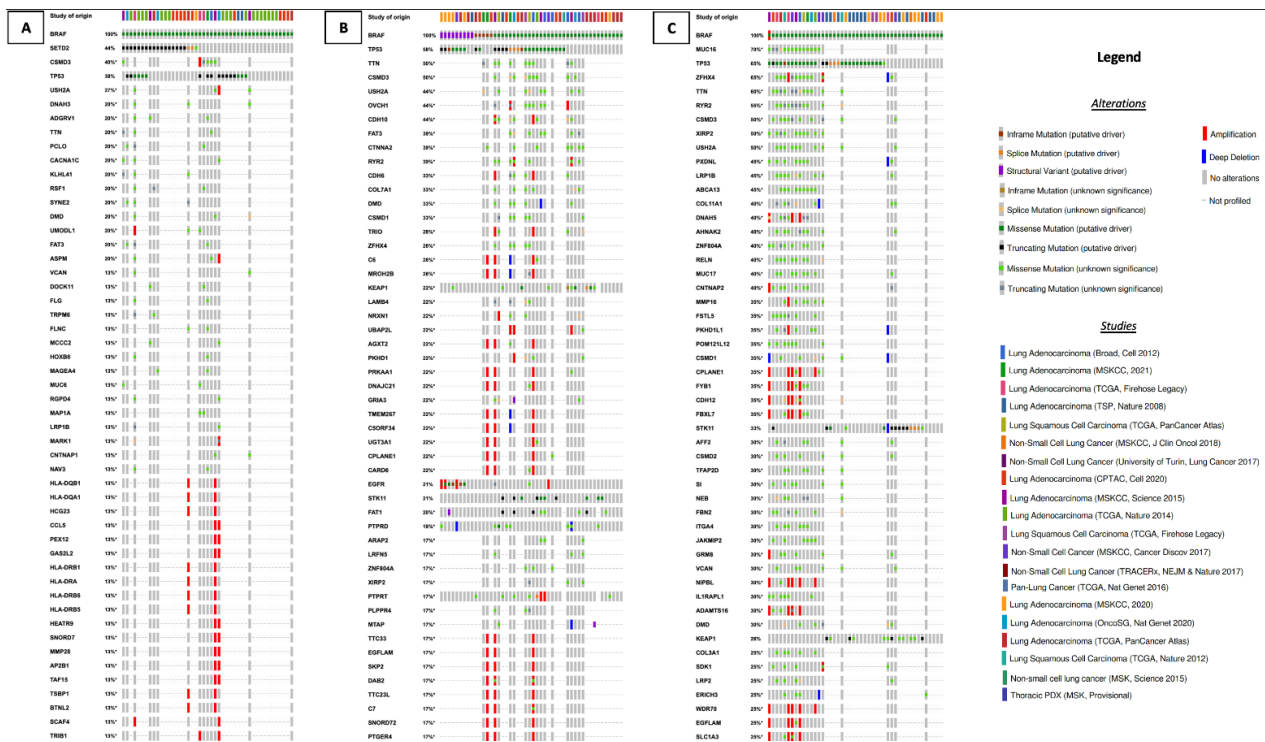


Figure 4. Top 50 most frequently altered genes in patients harboring BRAF alterations of class 1 (A), 2 (B), and 3 (C), the type of alteration, and the study of origin of each patient.

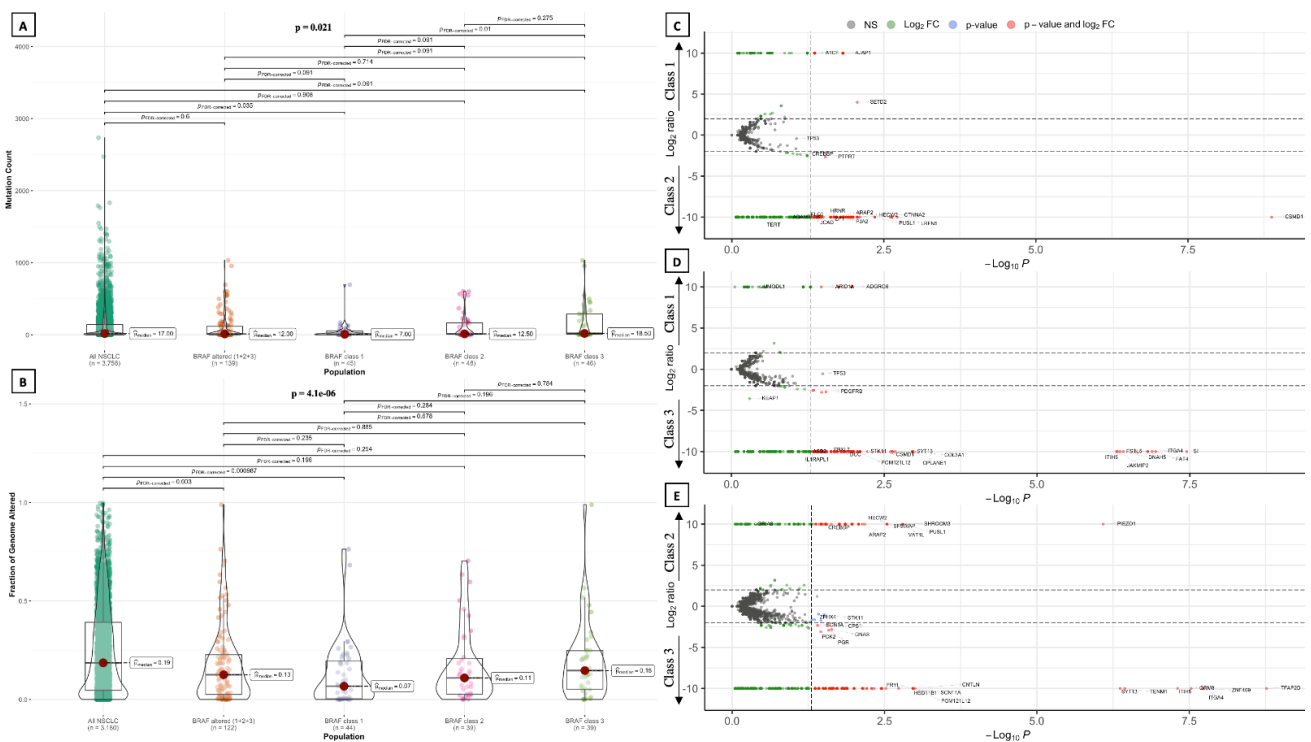


Figure 5. Violin plots show a comparison in terms of mutation count (A) and fraction of genome altered (B) among the three BRAF functional classes, showing a significantly higher mutation count in patients harboring class 3 alterations compared to those with class 1 alterations. Volcano plots comparing gene co-alteration frequency in patients harboring BRAF alterations of class 1 vs. class 2 (C), class 1 vs. class 3 (D), and class 2 vs. class 3 (E). NS: not significant.

3.2.3. Tumor Mutational Burden, Mutation Count and Fraction of Genome Altered

Median tumor mutational burden (TMB) was 7.83 mut/Mb (95% CI, 6.85–8.97) in patients with any *BRAF* alteration ($n = 236$). In tumors with *BRAF* alterations of a known functional class ($n = 139$), median TMB did not significantly differ from that of all NSCLC patients (6.05 mut/Mb vs. 5.30 mut/Mb, $p = 0.209$) (Figure 2B). Instead, a statistically significant difference was found between NSCLC harboring distinct classes of *BRAF* alterations ($p < 0.001$). In particular, median TMB was significantly lower in tumors with *BRAF* class 1 alterations (median TMB = 3.91 mut/Mb) than in those harboring class 2 (median TMB = 6.73 mut/Mb, $p = 0.004$) and class 3 (median TMB = 10.57 mut/Mb, $p < 0.001$) alterations, as well as compared to that of NSCLC with *BRAF* alterations of any known functional class ($p = 0.006$) and unselected NSCLC ($p = 0.03$) (Figure 2B). Class 3 alterations were associated with the highest median TMB, even compared to *BRAF* alterations of any known functional class ($p = 0.006$) and unselected NSCLC ($p < 0.001$) (Figure 2B). Total mutation count (MC), defined as the total number of mutations found in each patient's sample, was not significantly different between NSCLC patients with *BRAF* alterations of a known functional class and unselected NSCLC patients (Figure 5A). However, MC was significantly different among patients carrying distinct *BRAF* alterations classes, and was significantly higher in patients harboring class 3 alterations compared to those with class 1 alterations ($p = 0.01$) (Figure 5A). Likewise, the fraction of genome altered (FGA) was significantly lower in patients with *BRAF* alterations of a known functional class compared to the total of NSCLC patients, but did not show significant differences among the three *BRAF* classes (Figure 5B).

4. Discussion

The significance and prevalence of the wide spectrum of *BRAF* gene alterations in cancer is largely unknown. Besides class 1 alterations, encompassing p.V600X mutations, little is known about the role of class 2 and 3 alterations, their prevalence in different tumor types, and their influence on clinical features and treatment outcomes. In NSCLC patients, *BRAF* non-V600 mutations are generally described as half of total *BRAF* mutations [6]. In the current study, following a comprehensive and cross-tumor systematic research of *BRAF* alterations of known functional class, we showed that each class of *BRAF* alterations approximately constitutes 1/3 of total *BRAF*-mutant NSCLC. We also widened the spectrum of *BRAF* alterations by functional classes previously reported in NSCLC patients. Nonetheless, the real prevalence of *BRAF* class 2 and 3 alterations is still to be considered underestimated, as we did not identify any reported corresponding functional class for many alterations found in literature. A deeper knowledge of the significance of these alterations and their clinical implication is thus of paramount importance, as it may lead to a more personalized approach for a considerable number of patients, including the identification of tailored treatments. Uncovering the molecular landscape accompanying *BRAF* alterations of distinct classes constitutes important aid in accomplishing this aim. We showed that class 1 alterations are associated with the lowest median TMB, significantly lower than class 3 ones and unselected NSCLC patients. Conversely, tumors harboring class 3 alterations have a median TMB greater than 10 mutation/Mb, significantly higher than the median of all NSCLC. These results are consistent with a heavier smoking habit in NSCLC patients with *BRAF* class 2 and 3 alterations compared to class 1. The presence of high TMB is a relevant biomarker of high tumor neoantigen load and, by consequence, a possible predictive factor of immunotherapy treatment outcome [51]. However, despite having demonstrated its ability to predict the outcomes of immune-checkpoint inhibitors (ICI) in many studies, the definitive predictive role of TMB in NSCLC is still debated, as its correlation with overall immunotherapy treatment outcome has been inconsistent in terms of survival benefit [51–54]. A recent retrospective study reported generally unsatisfactory outcomes with immunotherapy in NSCLC patients harboring *BRAF* alterations, although class 2 and 3 altered patients achieved numerically higher objective response rate (ORR) than those carrying class 1 mutations (26% vs. 9%; $p = 0.25$) [55]. Consistently

with our study, patients with class 2 and 3 mutations had significantly higher TMB than those harboring class 1 mutations [55]. However, the small sample size, the use of targeted therapy in patients with class 1 alterations and the heterogeneity of lines of ICI treatment constituted important limitations. Results from the IMMUNOTARGET registry, which included 43 NSCLC patients with *BRAF* alterations, showed higher activity of immunotherapy (ORR: 24.3%) compared to NSCLC patients carrying different oncogene alterations, such as *EGFR*, *MET*, *RET*, *ROS1*, *ALK* and *HER2* ones [56]. Median PFS in *BRAF*-mutant patients was also longer, especially in those harboring non-V600E mutations, compared to that of patients carrying several different driver gene mutations. However, conversely, median OS was remarkably shorter compared to that of patients carrying other driver alterations, such as *MET* or *RET* ones, supporting the negative prognostic value of *BRAF* mutations [56]. These results are consistent with that of a study from the Israeli Lung Cancer Group suggesting favorable outcomes with ICI in a smaller population of patients with *BRAF*-mutant NSCLC with either V600 or non-V600 alterations, as well as with an analysis of *BRAF*-mutant patients enrolled in the Italian Expanded Access Program of second-line nivolumab [57,58]. Less favorable survival outcomes in *BRAF*-altered patients of class 2 and 3 have also been reported with chemotherapy, mainly due to the presence of more aggressive clinical features compared to NSCLC patients with class 1 alterations, such as a higher frequency of extra-thoracic dissemination [27]. In fact, no survival difference was observed after the exclusion of patients with M1b disease and those treated with targeted therapy [27]. Our results support these findings, suggesting that *BRAF* non-V600 mutations confer a poor prognosis independently of the treatment received. However, a bigger sample size is necessary to determine whether immunotherapy performs better than chemotherapy in this population, and whether patients harboring different classes of *BRAF* alterations derive distinct benefit from specific treatment strategies. In accordance with what was observed with TMB, we showed that the MC and FGA progressively increased from class 1 to class 2 and 3 *BRAF*-altered patients. We also demonstrated that median FGA is significantly lower in patients with *BRAF* alteration of a known functional class compared to unselected NSCLC, but this difference is probably driven by the lower median FGA in patients with class 1 alterations. Similar to TMB, phenotypic implications of MC and FGA may impact patients' prognosis and immunotherapy efficacy [59–63]. Our work evidenced that distinct classes of *BRAF* alterations in NSCLC are associated with a broad and heterogeneous genomic landscape, and some gene alterations may help in explaining the peculiar behavior of each class. For example, *STK11* and *KEAP1* alterations, which we found with higher prevalence in tumors harboring class 2 and class 3 *BRAF* alterations than in those with class 1 alterations, where they were almost absent, have been associated with high TMB but immune “cold” tumor microenvironment and poor prognosis [64–66]. We showed that *TP53* alterations are also particularly enriched in class 2 and, particularly, class 3 *BRAF*-altered NSCLC patients compared to those with class 1 alterations, which may help in explaining the more aggressive behavior of these tumors and the poor outcomes reported in literature [27,67–69]. Likewise, we showed that *TERT* mutations, which are rare in lung cancer (approximate prevalence of 2%) and have been correlated with poor prognosis, are enriched in NSCLC patients harboring class 2 and 3 *BRAF* alterations and are absent in V600E mutants, which constituted the totality of class 1 patients [70]. This peculiar distribution among *BRAF* functional classes in NSCLC is in contrast with data from melanoma and thyroid carcinoma patients, where *TERT* mutations have been mainly described in *BRAF* V600E-mutant tumors [71]. We also found *MUC16* alterations in the majority of patients with class 3 *BRAF* alterations, but these occurred very less frequently in patients with class 2 and, especially, class 1 alterations. In melanoma, *MUC16* alterations have been frequently found to be associated with *BRAF* V600E mutations and higher TMB than wild-type patients. Interestingly, these alterations also occur in pancreatic cancer, where they have been associated with disease progression and metastasis through the activation of oncogenic pathways via the interaction between aberrant *MUC16* isoforms and epidermal growth factor (EGF) receptors [72]. On the contrary, consistently with the

current literature, *SETD2* co-alterations were present in many *BRAF* class 1 altered patients, but they were infrequent in non-V600 patients [29]. *SETD2* mutations have been associated with high TMB, microsatellite instability and favorable outcomes with ICI [73]. Further gene co-alterations found with high prevalence in one or more *BRAF* functional classes have been associated with higher TMB, such as *TTN*, *CSMD3*, *USH2A* and *RYR2* ones [74–77]. Together with a different distribution of DNA damage response gene alterations, which has been associated with enhanced ICI efficacy, these features suggest a potentially promising role of immunotherapy in selected patients [78]. The main limitation to our study is represented by the lack of data regarding tumor stage, metastatic sites and treatment outcomes in the cBioPortal cohort, as they were reported for too few patients to allow a proper analysis. Moreover, retrieving data from different studies included in the cBioPortal carries an intrinsic and not avoidable heterogeneity. However, meticulous data screening, cleaning and reporting reduced the risk of misinterpretations. In fact, one of the main strengths of this work is that it is represented by a rigorous methodology, which begins from the detection and collection of *BRAF* alterations of defined functional class through a comprehensive systematic review of the literature, leading to clinical and molecular data selection, retrieval and analysis from large and high-quality genomic studies and, finally, to the selection of a cohort of patients from two large, practice-changing, randomized clinical trials for survival analyses. Another strength is the production of original data from a large number of patients (271 patients harboring *BRAF* alterations taking into account both cohorts), considering the rarity of these alterations in NSCLC; moreover, many *BRAF* alterations of class 2 and 3 have not been previously described and analyzed in patients with NSCLC.

5. Conclusions

BRAF-altered NSCLCs encompass a broad and heterogeneous genomic spectrum of tumors, each with distinctive molecular signatures, clinical-biological behavior and potentially exploitable specific treatment strategies. NSCLC patients harboring non-V600 *BRAF* alterations constitute a considerable and underestimated population characterized by peculiar genomic landscape and poor prognosis compared to *BRAF* wild-type patients, warranting larger and deeper studies aiming to identify potential tailored therapies.

Supplementary Materials: The following supporting information can be downloaded at: <https://www.mdpi.com/article/10.3390/cancers14143472/s1>, Figure S1: Preferred Reported Items for Systematic Reviews and Meta-Analysis (PRISMA) flowchart of literature research adopted to conduct the systematic review; Figure S2: Consort Diagram summarizing the selection of NSCLC patients harboring *BRAF* alterations of defined functional class in the cBioPortal; Table S1: List of *BRAF* alterations (protein change) and corresponding functional class detected through the systematic review of the literature; Table S2: Comparative frequency of concurrent gene alterations in most commonly altered and key genes among NSCLC patients harboring *BRAF* class 1, class 2, and class 3 alterations; Table S3: Descriptions of detected *BRAF* structural variants in cBioPortal.

Author Contributions: Conceptualization, A.D.F.; methodology, A.D.F., A.D.G., F.G. (Francesco Gelsomino) and A.A.; validation, A.D.G., F.G. (Francesca Giunchi), D.D.B., A.P., F.G. (Francesco Gelsomino) and A.A.; formal analysis, A.D.F.; investigation, A.D.F. and A.D.G.; data curation, A.D.F.; writing—original draft preparation, A.D.F. and A.D.G.; writing—review and editing, F.S., B.M., A.A., D.D.B., F.G. (Francesca Giunchi) and F.G. (Francesco Gelsomino); supervision, F.G. (Francesco Gelsomino) and A.A.; project administration, A.A.; All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

Data Availability Statement: Patient-level data from the POPLAR and OAK trials are available as supplementary information of the Gandara DR et al. [15]. Data extracted from the 25 studies (Figure 1A) included in the cBioPortal are available at www.cbioportal.org, last accessed on 18 February 2022.

Conflicts of Interest: AA reports grants and personal fees from BMS, personal fees from MSD, personal fees from Eli-Lilly, personal fees from Boehringer, personal fees from Pfizer, grants from Celgene and grants and personal fees from Roche, outside the submitted work. Francesco Gelsomino received honoraria for advisory board participation: Eli-Lilly. The other authors have no disclosure or conflict of interest to declare.

References

1. Global Cancer Observatory. Available online: <https://gco.iarc.fr/> (accessed on 26 January 2022).
2. Reck, M.; Rodríguez-Abreu, D.; Robinson, A.G.; Hui, R.; Csósz, T.; Fülöp, A.; Gottfried, M.; Peled, N.; Tafreshi, A.; Cuffe, S.; et al. Updated Analysis of KEYNOTE-024: Pembrolizumab Versus Platinum-Based Chemotherapy for Advanced Non-Small-Cell Lung Cancer with PD-L1 Tumor Proportion Score of 50% or Greater. *J. Clin. Oncol.* **2019**, *37*, 537–546. [[CrossRef](#)] [[PubMed](#)]
3. Gadgeel, S.; Rodríguez-Abreu, D.; Speranza, G.; Esteban, E.; Felip, E.; Dómine, M.; Hui, R.; Hochmair, M.J.; Clingan, P.; Powell, S.F.; et al. Updated Analysis From KEYNOTE-189: Pembrolizumab or Placebo Plus Pemetrexed and Platinum for Previously Untreated Metastatic Nonsquamous Non-Small-Cell Lung Cancer. *J. Clin. Oncol.* **2020**, *38*, 1505–1517. [[CrossRef](#)] [[PubMed](#)]
4. Imyanitov, E.N.; Iyevleva, A.G.; Levchenko, E.V. Molecular testing and targeted therapy for non-small cell lung cancer: Current status and perspectives. *Crit. Rev. Oncol. Hematol.* **2021**, *157*, 103194. [[CrossRef](#)] [[PubMed](#)]
5. Lamberti, G.; Andrini, E.; Sisi, M.; Rizzo, A.; Parisi, C.; Di Federico, A.; Gelsomino, F.; Ardizzoni, A. Beyond EGFR, ALK and ROS1: Current evidence and future perspectives on newly targetable oncogenic drivers in lung adenocarcinoma. *Crit. Rev. Oncol. Hematol.* **2020**, *156*, 103119. [[CrossRef](#)]
6. Paik, P.K.; Arcila, M.E.; Fara, M.; Sima, C.S.; Miller, V.A.; Kris, M.G.; Ladanyi, M.; Riely, G.J. Clinical Characteristics of Patients with Lung Adenocarcinomas Harboring BRAF Mutations. *J. Clin. Oncol.* **2011**, *29*, 2046–2051. [[CrossRef](#)] [[PubMed](#)]
7. Leonetti, A.; Facchinetti, F.; Rossi, G.; Minari, R.; Conti, A.; Friboulet, L.; Tiseo, M.; Planchard, D. BRAF in non-small cell lung cancer (NSCLC): Pickaxing another brick in the wall. *Cancer Treat. Rev.* **2018**, *66*, 82–94. [[CrossRef](#)] [[PubMed](#)]
8. Litvak, A.M.; Paik, P.K.; Woo, K.M.; Sima, C.S.; Hellmann, M.D.; Arcila, M.E.; Ladanyi, M.; Rudin, C.M.; Kris, M.G.; Riely, G.J. Clinical Characteristics and Course of 63 Patients with BRAF Mutant Lung Cancers. *J. Thorac. Oncol.* **2014**, *9*, 1669–1674. [[CrossRef](#)]
9. Dankner, M.; Rose, A.A.N.; Rajkumar, S.; Siegel, P.M.; Watson, I.R. Classifying BRAF alterations in cancer: New rational therapeutic strategies for actionable mutations. *Oncogene* **2018**, *37*, 3183–3199. [[CrossRef](#)]
10. Planchard, D.; Besse, B.; Groen, H.J.M.; Hashemi, S.M.S.; Mazieres, J.; Kim, T.M.; Quoix, E.; Souquet, P.-J.; Barlesi, F.; Baik, C.; et al. Phase 2 Study of Dabrafenib Plus Trametinib in Patients with BRAF V600E-Mutant Metastatic NSCLC: Updated 5-Year Survival Rates and Genomic Analysis. *J. Thorac. Oncol.* **2022**, *17*, 103–115. [[CrossRef](#)]
11. Planchard, D.; Besse, B.; Groen, H.J.M.; Souquet, P.-J.; Quoix, E.; Baik, C.S.; Barlesi, F.; Kim, T.M.; Mazieres, J.; Novello, S.; et al. Dabrafenib plus trametinib in patients with previously treated BRAFV600E-mutant metastatic non-small cell lung cancer: An open-label, multicentre phase 2 trial. *Lancet Oncol.* **2016**, *17*, 984–993. [[CrossRef](#)]
12. Dankner, M.; Lajoie, M.; Moldoveanu, D.; Nguyen, T.-T.; Savage, P.; Rajkumar, S.; Huang, X.; Lvova, M.; Protopopov, A.; Vuzman, D.; et al. Dual MAPK Inhibition Is an Effective Therapeutic Strategy for a Subset of Class II BRAF Mutant Melanomas. *Clin. Cancer Res.* **2018**, *24*, 6483–6494. [[CrossRef](#)] [[PubMed](#)]
13. Fehrenbacher, L.; Spira, A.; Ballinger, M.; Kowanzet, M.; Vansteenkiste, J.; Mazieres, J.; Park, K.; Smith, D.; Artal-Cortes, A.; Lewanski, C.; et al. Atezolizumab versus docetaxel for patients with previously treated non-small-cell lung cancer (POPLAR): A multicentre, open-label, phase 2 randomised controlled trial. *Lancet* **2016**, *387*, 1837–1846. [[CrossRef](#)]
14. Rittmeyer, A.; Barlesi, F.; Waterkamp, D.; Park, K.; Ciardiello, F.; von Pawel, J.; Gadgeel, S.M.; Hida, T.; Kowalski, D.M.; Dols, M.C.; et al. Atezolizumab versus docetaxel in patients with previously treated non-small-cell lung cancer (OAK): A phase 3, open-label, multicentre randomised controlled trial. *Lancet* **2017**, *389*, 255–265. [[CrossRef](#)]
15. Gandara, D.R.; Paul, S.M.; Kowanzet, M.; Schleifman, E.; Zou, W.; Li, Y.; Rittmeyer, A.; Fehrenbacher, L.; Otto, G.; Malboeuf, C.; et al. Blood-based tumor mutational burden as a predictor of clinical benefit in non-small-cell lung cancer patients treated with atezolizumab. *Nat. Med.* **2018**, *24*, 1441–1448. [[CrossRef](#)] [[PubMed](#)]
16. Cerami, E.; Gao, J.; Dogrusoz, U.; Gross, B.E.; Sumer, S.O.; Aksoy, B.A.; Jacobsen, A.; Byrne, C.J.; Heuer, M.L.; Larsson, E.; et al. The cBio Cancer Genomics Portal: An Open Platform for Exploring Multidimensional Cancer Genomics Data. *Cancer Discov.* **2012**, *2*, 401–404. [[CrossRef](#)]
17. Gao, J.; Aksoy, B.A.; Dogrusoz, U.; Dresdner, G.; Gross, B.; Sumer, S.O.; Sun, Y.; Jacobsen, A.; Sinha, R.; Larsson, E.; et al. Integrative Analysis of Complex Cancer Genomics and Clinical Profiles Using the cBioPortal. *Sci. Signal.* **2013**, *6*, p11. [[CrossRef](#)]
18. Negrao, M.V.; Raymond, V.M.; Lanman, R.B.; Robichaux, J.P.; He, J.; Nilsson, M.B.; Ng, P.K.S.; Amador, B.E.; Roarty, E.B.; Nagy, R.J.; et al. Molecular Landscape of BRAF-Mutant NSCLC Reveals an Association between Clonality and Driver Mutations and Identifies Targetable Non-V600 Driver Mutations. *J. Thorac. Oncol.* **2020**, *15*, 1611–1623. [[CrossRef](#)]
19. Schirripa, M.; Biondi, P.; Lonardi, S.; Pella, N.; Pino, M.S.; Urbano, F.; Antoniotti, C.; Cremolini, C.; Corallo, S.; Pietrantonio, F.; et al. Class 1, 2, and 3 BRAF-Mutated Metastatic Colorectal Cancer: A Detailed Clinical, Pathologic, and Molecular Characterization. *Clin. Cancer Res.* **2019**, *25*, 3954–3961. [[CrossRef](#)]

20. Lokhandwala, P.M.; Tseng, L.-H.; Rodriguez, E.; Zheng, G.; Pallavajjalla, A.; Gocke, C.D.; Eshleman, J.R.; Lin, M.-T. Clinical mutational profiling and categorization of BRAF mutations in melanomas using next generation sequencing. *BMC Cancer* **2019**, *19*, 665. [[CrossRef](#)]
21. Lin, Q.; Zhang, H.; Ding, H.; Qian, J.; Lizaso, A.; Lin, J.; Han-Zhang, H.; Xiang, J.; Li, Y.; Zhu, H. The association between BRAF mutation class and clinical features in BRAF-mutant Chinese non-small cell lung cancer patients. *J. Transl. Med.* **2019**, *17*, 298. [[CrossRef](#)]
22. Bracht, J.W.P.; Karachaliou, N.; Bivona, T.; Lanman, R.B.; Faull, I.; Nagy, R.J.; Drozdowskyj, A.; Berenguer, J.; Fernandez-Bruno, M.; Molina-Vila, M.A.; et al. BRAF Mutations Classes I, II, and III in NSCLC Patients Included in the SLLIP Trial: The Need for a New Pre-Clinical Treatment Rationale. *Cancers* **2019**, *11*, 1381. [[CrossRef](#)]
23. Owsley, J.; Stein, M.K.; Porter, J.; In, G.K.; Salem, M.; O'Day, S.; Elliott, A.; Poorman, K.; Gibney, G.; VanderWalde, A. Prevalence of class I–III BRAF mutations among 114,662 cancer patients in a large genomic database. *Exp. Biol. Med.* **2021**, *246*, 31–39. [[CrossRef](#)] [[PubMed](#)]
24. Zhao, Y.; Yu, H.; Ida, C.M.; Halling, K.C.; Kipp, B.R.; Geiersbach, K.; Rumilla, K.M.; Gupta, S.; Lin, M.-T.; Zheng, G. Assessment of RAS Dependency for BRAF Alterations Using Cancer Genomic Databases. *JAMA Netw. Open* **2021**, *4*, e2035479. [[CrossRef](#)] [[PubMed](#)]
25. Yao, Z.; Yaeger, R.; Rodrik-Outmezguine, V.S.; Tao, A.; Torres, N.M.; Chang, M.T.; Drosten, M.; Zhao, H.; Cecchi, F.; Hembrough, T.; et al. Tumours with class 3 BRAF mutants are sensitive to the inhibition of activated RAS. *Nature* **2017**, *548*, 234–238. [[CrossRef](#)] [[PubMed](#)]
26. Yaeger, R.; Kotani, D.; Mondaca, S.; Parikh, A.R.; Bando, H.; Van Seventer, E.E.; Taniguchi, H.; Zhao, H.; Thant, C.N.; de Stanchina, E.; et al. Response to Anti-EGFR Therapy in Patients with BRAF non-V600–Mutant Metastatic Colorectal Cancer. *Clin. Cancer Res.* **2019**, *25*, 7089–7097. [[CrossRef](#)]
27. Dagogo-Jack, I.; Martinez, P.; Yeap, B.Y.; Ambrogio, C.; Ferris, L.A.; Lydon, C.; Nguyen, T.; Jessop, N.A.; Iafrate, A.J.; Johnson, B.E.; et al. Impact of BRAF Mutation Class on Disease Characteristics and Clinical Outcomes in BRAF-mutant Lung Cancer. *Clin. Cancer Res.* **2019**, *25*, 158–165. [[CrossRef](#)]
28. Osumi, H.; Shinozaki, E.; Wakatsuki, T.; Suenaga, M.; Ichimura, T.; Ogura, M.; Takahari, D.; Ooki, A.; Suzuki, T.; Ota, Y.; et al. Non-V600E BRAF mutations and EGFR signaling pathway in colorectal cancer. *Int. J. Cancer* **2019**, *145*, 2488–2495. [[CrossRef](#)]
29. Sheikine, Y.; Pavlick, D.; Klemptner, S.J.; Trabucco, S.E.; Chung, J.H.; Rosenzweig, M.; Wang, K.; Velcheti, V.; Frampton, G.M.; Peled, N.; et al. BRAF in Lung Cancers: Analysis of Patient Cases Reveals Recurrent BRAF Mutations, Fusions, Kinase Duplications, and Concurrent Alterations. *JCO Precis. Oncol.* **2018**, *2*, PO.17.00172. [[CrossRef](#)]
30. Lobo-Martins, S.; Pais, H.L.; Soares-de-Almeida, L.; Costa, L.; Mansinho, A.; Teixeira de Sousa, R. BRAF L597K mutation: An opportunity to treat. *Dermatol. Online J.* **2021**, *27*. [[CrossRef](#)]
31. Wang, Y.; Ji, M.; Wang, W.; Miao, Z.; Hou, P.; Chen, X.; Xu, F.; Zhu, G.; Sun, X.; Li, Y.; et al. Association of the T1799A BRAF mutation with tumor extrathyroidal invasion, higher peripheral platelet counts, and over-expression of platelet-derived growth factor-B in papillary thyroid cancer. *Endocr. Relat. Cancer* **2008**, *15*, 183–190. [[CrossRef](#)]
32. Mayank, M.; Kaur, N.; Singh, N. Structural insights and influence of V599 mutations on the overall dynamics of BRAF protein against its kinase domains. *Integr. Biol.* **2018**, *10*, 646–657. [[CrossRef](#)] [[PubMed](#)]
33. Cañadas-Garre, M.; Fernandez-Escamilla, A.M.; Fernandez-Ballester, G.; Becerra-Massare, P.; García-Calvente, C.; Ramos, J.L.; Llamas-Elvira, J.M. Novel BRAF1599Ins Mutation Identified in a Follicular Variant of Papillary Thyroid Carcinoma: A Molecular Modeling Approach. *Endocr. Pract.* **2014**, *20*, e75–e79. [[CrossRef](#)] [[PubMed](#)]
34. Griffith, M.; Spies, N.C.; Krysiak, K.; McMichael, J.F.; Coffman, A.C.; Danos, A.M.; Ainscough, B.J.; Ramirez, C.A.; Rieke, D.T.; Kujan, L.; et al. CIViC is a community knowledgebase for expert crowdsourcing the clinical interpretation of variants in cancer. *Nat. Genet.* **2017**, *49*, 170–174. [[CrossRef](#)] [[PubMed](#)]
35. Imielinski, M.; Berger, A.H.; Hammerman, P.S.; Hernandez, B.; Pugh, T.J.; Hodis, E.; Cho, J.; Suh, J.; Capelletti, M.; Sivachenko, A.; et al. Mapping the Hallmarks of Lung Adenocarcinoma with Massively Parallel Sequencing. *Cell* **2012**, *150*, 1107–1120. [[CrossRef](#)] [[PubMed](#)]
36. Gillette, M.A.; Satpathy, S.; Cao, S.; Dhanasekaran, S.M.; Vasaikar, S.V.; Krug, K.; Petralia, F.; Li, Y.; Liang, W.-W.; Reva, B.; et al. Proteogenomic Characterization Reveals Therapeutic Vulnerabilities in Lung Adenocarcinoma. *Cell* **2020**, *182*, 200–225.e35. [[CrossRef](#)] [[PubMed](#)]
37. Rizvi, N.A.; Hellmann, M.D.; Snyder, A.; Kvistborg, P.; Makarov, V.; Havel, J.J.; Lee, W.; Yuan, J.; Wong, P.; Ho, T.S.; et al. Mutational landscape determines sensitivity to PD-1 blockade in non-small cell lung cancer. *Science* **2015**, *348*, 124–128. [[CrossRef](#)] [[PubMed](#)]
38. Chen, J.; Yang, H.; Teo, A.S.M.; Amer, L.B.; Sherbaf, F.G.; Tan, C.Q.; Alvarez, J.J.S.; Lu, B.; Lim, J.Q.; Takano, A.; et al. Genomic landscape of lung adenocarcinoma in East Asians. *Nat. Genet.* **2020**, *52*, 177–186. [[CrossRef](#)] [[PubMed](#)]
39. Cancer Genome Atlas Research Network. Comprehensive molecular profiling of lung adenocarcinoma. *Nature* **2014**, *511*, 543–550, Erratum in *Nature* **2014**, *514*, 262. [[CrossRef](#)] [[PubMed](#)]
40. Liu, J.; Lichtenberg, T.; Hoadley, K.A.; Poisson, L.M.; Lazar, A.J.; Cherniack, A.D.; Kovatich, A.J.; Benz, C.C.; Levine, D.A.; Lee, A.V.; et al. An Integrated TCGA Pan-Cancer Clinical Data Resource to Drive High-Quality Survival Outcome Analytics. *Cell* **2018**, *173*, 400–416.e11. [[CrossRef](#)] [[PubMed](#)]

41. Ding, L.; Getz, G.; Wheeler, D.A.; Mardis, E.R.; McLellan, M.D.; Cibulskis, K.; Sougnez, C.; Greulich, H.; Muzny, D.M.; Morgan, M.B.; et al. Somatic mutations affect key pathways in lung adenocarcinoma. *Nature* **2008**, *455*, 1069–1075. [[CrossRef](#)] [[PubMed](#)]
42. Um, S.-W.; Joung, J.-G.; Lee, H.; Kim, H.; Kim, K.-T.; Park, J.; Hayes, D.N.; Park, W.-Y. Molecular Evolution Patterns in Metastatic Lymph Nodes Reflect the Differential Treatment Response of Advanced Primary Lung Cancer. *Cancer Res.* **2016**, *76*, 6568–6576. [[CrossRef](#)]
43. Satpathy, S.; Krug, K.; Jean Beltran, P.M.; Savage, S.R.; Petralia, F.; Kumar-Sinha, C.; Dou, Y.; Reva, B.; Kane, M.H.; Avanesian, S.C.; et al. A proteogenomic portrait of lung squamous cell carcinoma. *Cell* **2021**, *184*, 4348–4371.e40. [[CrossRef](#)]
44. Cancer Genome Atlas Research Network. Comprehensive genomic characterization of squamous cell lung cancers. *Nature* **2012**, *489*, 519–525, Erratum in *Nature* **2012**, *491*, 288. [[CrossRef](#)] [[PubMed](#)]
45. Hoadley, K.A.; Yau, C.; Hinoue, T.; Wolf, D.M.; Lazar, A.J.; Drill, E.; Shen, R.; Taylor, A.M.; Cherniack, A.D.; Thorsson, V.; et al. Cell-of-Origin Patterns Dominate the Molecular Classification of 10,000 Tumors from 33 Types of Cancer. *Cell* **2018**, *173*, 291–304.e6. [[CrossRef](#)] [[PubMed](#)]
46. Jordan, E.J.; Kim, H.R.; Arcila, M.E.; Barron, D.; Chakravarty, D.; Gao, J.; Chang, M.T.; Ni, A.; Kundra, R.; Jonsson, P.; et al. Prospective Comprehensive Molecular Characterization of Lung Adenocarcinomas for Efficient Patient Matching to Approved and Emerging Therapies. *Cancer Discov.* **2017**, *7*, 596–609. [[CrossRef](#)]
47. Hellmann, M.D.; Nathanson, T.; Rizvi, H.; Creelan, B.C.; Sanchez-Vega, F.; Ahuja, A.; Ni, A.; Novik, J.B.; Mangarin, L.M.B.; Abu-Akeel, M.; et al. Genomic Features of Response to Combination Immunotherapy in Patients with Advanced Non-Small-Cell Lung Cancer. *Cancer Cell* **2018**, *33*, 843–852.e4. [[CrossRef](#)] [[PubMed](#)]
48. Rizvi, H.; Sanchez-Vega, F.; La, K.; Chatila, W.; Jonsson, P.; Halpenny, D.; Plodkowski, A.; Long, N.; Sauter, J.L.; Rekhman, N.; et al. Molecular Determinants of Response to Anti-Programmed Cell Death (PD)-1 and Anti-Programmed Death-Ligand 1 (PD-L1) Blockade in Patients with Non-Small-Cell Lung Cancer Profiled with Targeted Next-Generation Sequencing. *J. Clin. Oncol.* **2018**, *36*, 633–641. [[CrossRef](#)] [[PubMed](#)]
49. Jamal-Hanjani, M.; Wilson, G.A.; McGranahan, N.; Birkbak, N.J.; Watkins, T.B.K.; Veeriah, S.; Shafi, S.; Johnson, D.H.; Mitter, R.; Rosenthal, R.; et al. Tracking the Evolution of Non-Small-Cell Lung Cancer. *N. Engl. J. Med.* **2017**, *376*, 2109–2121. [[CrossRef](#)]
50. Vavalà, T.; Monica, V.; Lo Iacono, M.; Mele, T.; Busso, S.; Righi, L.; Papotti, M.; Scagliotti, G.V.; Novello, S. Precision medicine in age-specific non-small-cell-lung-cancer patients: Integrating biomolecular results into clinical practice—A new approach to improve personalized translational research. *Lung Cancer* **2017**, *107*, 84–90. [[CrossRef](#)]
51. Chan, T.A.; Yarchoan, M.; Jaffee, E.; Swanton, C.; Quezada, S.A.; Stenzinger, A.; Peters, S. Development of tumor mutation burden as an immunotherapy biomarker: Utility for the oncology clinic. *Ann. Oncol.* **2019**, *30*, 44–56. [[CrossRef](#)]
52. Hellmann, M.D.; Paz-Ares, L.; Bernabe Caro, R.; Zurawski, B.; Kim, S.-W.; Carcereny Costa, E.; Park, K.; Alexandru, A.; Lupinacci, L.; de la Mora Jimenez, E.; et al. Nivolumab plus Ipilimumab in Advanced Non-Small-Cell Lung Cancer. *N. Engl. J. Med.* **2019**, *381*, 2020–2031. [[CrossRef](#)] [[PubMed](#)]
53. Si, H.; Kuziora, M.; Quinn, K.J.; Helman, E.; Ye, J.; Liu, F.; Scheuring, U.; Peters, S.; Rizvi, N.A.; Brohawn, P.Z.; et al. A Blood-based Assay for Assessment of Tumor Mutational Burden in First-line Metastatic NSCLC Treatment: Results from the MYSTIC Study. *Clin. Cancer Res.* **2021**, *27*, 1631–1640. [[CrossRef](#)] [[PubMed](#)]
54. Dziadziusko, R.; Peters, S.; Gadgeel, S.; Mathisen, M.S.; Shagan, S.M.; Felip, E.; Morabito, A.; Cheema, P.; Cobo Dols, M.; Andric, Z.; et al. Atezolizumab (atezo) vs. platinum-based chemo in bloodbasedtumour mutational burden-positive (bTMB+) patients(pts) with first-line (1L) advanced/metastatic (m)NSCLC: Results of the Blood First Assay Screening Trial (BFAST)phase III cohort C. *Ann. Oncol.* **2021**, *32*, S949–S1039. [[CrossRef](#)]
55. Murciano-Goroff, Y.R.; Pak, T.; Mondaca, S.; Flynn, J.R.; Montecalvo, J.; Rekhman, N.; Halpenny, D.; Plodkowski, A.J.; Wu, S.L.; Kris, M.G.; et al. Immune biomarkers and response to checkpoint inhibition of BRAFV600 and BRAF non-V600 altered lung cancers. *Br. J. Cancer* **2022**, *126*, 889–898. [[CrossRef](#)] [[PubMed](#)]
56. Mazieres, J.; Drilon, A.; Lusque, A.; Mhanna, L.; Cortot, A.B.; Mezquita, L.; Thai, A.A.; Mascaux, C.; Couraud, S.; Veillon, R.; et al. Immune checkpoint inhibitors for patients with advanced lung cancer and oncogenic driver alterations: Results from the IMMUNOTARGET registry. *Ann. Oncol.* **2019**, *30*, 1321–1328. [[CrossRef](#)]
57. Dudnik, E.; Peled, N.; Nechushtan, H.; Wollner, M.; Onn, A.; Agbarya, A.; Moskovitz, M.; Keren, S.; Popovits-Hadari, N.; Urban, D.; et al. BRAF Mutant Lung Cancer: Programmed Death Ligand 1 Expression, Tumor Mutational Burden, Microsatellite Instability Status, and Response to Immune Check-Point Inhibitors. *J. Thorac. Oncol.* **2018**, *13*, 1128–1137. [[CrossRef](#)]
58. Rihawi, K.; Giannarelli, D.; Galetta, D.; Delmonte, A.; Giavarrà, M.; Turci, D.; Garassino, M.; Tiseo, M.; Barbieri, F.; Panni, S.; et al. BRAF Mutant NSCLC and Immune Checkpoint Inhibitors: Results From a Real-World Experience. *J. Thorac. Oncol.* **2019**, *14*, e57–e59. [[CrossRef](#)]
59. Liu, Z.; Zhang, Y.; Shi, C.; Zhou, X.; Xu, K.; Jiao, D.; Sun, Z.; Han, X. A novel immune classification reveals distinct immune escape mechanism and genomic alterations: Implications for immunotherapy in hepatocellular carcinoma. *J. Transl. Med.* **2021**, *19*, 5. [[CrossRef](#)]
60. Jones, G.D.; Brandt, W.S.; Shen, R.; Sanchez-Vega, F.; Tan, K.S.; Martin, A.; Zhou, J.; Berger, M.; Solit, D.B.; Schultz, N.; et al. A Genomic-Pathologic Annotated Risk Model to Predict Recurrence in Early-Stage Lung Adenocarcinoma. *JAMA Surg.* **2021**, *156*, e205601. [[CrossRef](#)]

61. Mehta, K.R.; Nakao, K.; Zuraek, M.B.; Ruan, D.T.; Bergsland, E.K.; Venook, A.P.; Moore, D.H.; Tokuyasu, T.A.; Jain, A.N.; Warren, R.S.; et al. Fractional Genomic Alteration Detected by Array-Based Comparative Genomic Hybridization Independently Predicts Survival after Hepatic Resection for Metastatic Colorectal Cancer. *Clin. Cancer Res.* **2005**, *11*, 1791–1797. [[CrossRef](#)]
62. Chakraborty, G.; Atiq, M.; Nandakumar, S.; Mazzu, Y.Z.; Armenia, J.; Yoshikawa, Y.; Khan, N.; Lee, G.-S.M.; Mucci, L.; Kantoff, P.W. Abstract 2534: A comparative analysis of fraction genome altered vs tumor mutational count in prostate cancer. *Cancer Res.* **2019**, *79*, 2534. [[CrossRef](#)]
63. Chakraborty, G.; Ghosh, A.; Nandakumar, S.; Armenia, J.; Mazzu, Y.Z.; Atiq, M.O.; Lee, G.-S.M.; Mucci, L.A.; Merghoub, T.; Wolchok, J.D.; et al. Fraction genome altered (FGA) to regulate both cell autonomous and non-cell autonomous functions in prostate cancer and its effect on prostate cancer aggressiveness. *J. Clin. Oncol.* **2020**, *38*, 347. [[CrossRef](#)]
64. Marinelli, D.; Mazzotta, M.; Scalera, S.; Terrenato, I.; Sperati, F.; D'Ambrosio, L.; Pallocca, M.; Corleone, G.; Krasniqi, E.; Pizzuti, L.; et al. KEAP1-driven co-mutations in lung adenocarcinoma unresponsive to immunotherapy despite high tumor mutational burden. *Ann. Oncol.* **2020**, *31*, 1746–1754. [[CrossRef](#)] [[PubMed](#)]
65. Cho, B.C.; Lopes, G.; Kowalski, D.M.; Kasahara, K.; Wu, Y.-L.; Castro, G.; Turna, H.Z.; Cristescu, R.; Aurora-Garg, D.; Loboda, A.; et al. Abstract CT084: Relationship between STK11 and KEAP1 mutational status and efficacy in KEYNOTE-042: Pembrolizumab monotherapy versus platinum-based chemotherapy as first-line therapy for PD-L1-positive advanced NSCLC. *Cancer Res.* **2020**, *80*, CT084. [[CrossRef](#)]
66. Di Federico, A.; De Giglio, A.; Parisi, C.; Gelsomino, F. STK11/LKB1 and KEAP1 mutations in non-small cell lung cancer: Prognostic rather than predictive? *Eur. J. Cancer* **2021**, *157*, 108–113. [[CrossRef](#)]
67. Gu, J.; Zhou, Y.; Huang, L.; Ou, W.; Wu, J.; Li, S.; Xu, J.; Feng, J.; Liu, B. TP53 mutation is associated with a poor clinical outcome for non-small cell lung cancer: Evidence from a meta-analysis. *Mol. Clin. Oncol.* **2016**, *5*, 705–713. [[CrossRef](#)]
68. Tsao, M.-S.; Aviel-Ronen, S.; Ding, K.; Lau, D.; Liu, N.; Sakurada, A.; Whitehead, M.; Zhu, C.-Q.; Livingston, R.; Johnson, D.H.; et al. Prognostic and Predictive Importance of p53 and RAS for Adjuvant Chemotherapy in Non-Small-Cell Lung Cancer. *J. Clin. Oncol.* **2007**, *25*, 5240–5247. [[CrossRef](#)]
69. Custodio, A.B.; González-Larriba, J.L.; Bobokova, J.; Calles, A.; Álvarez, R.; Cuadrado, E.; Manzano, A.; Díaz-Rubio, E. Prognostic and Predictive Markers of Benefit from Adjuvant Chemotherapy in Early-Stage Non-small Cell Lung Cancer. *J. Thorac. Oncol.* **2009**, *4*, 891–910. [[CrossRef](#)]
70. Jung, S.-J.; Kim, D.-S.; Park, W.-J.; Lee, H.; Choi, I.-J.; Park, J.-Y.; Lee, J.-H. Mutation of the TERT promoter leads to poor prognosis of patients with non-small cell lung cancer. *Oncol. Lett.* **2017**, *14*, 1609–1614. [[CrossRef](#)]
71. Vinagre, J.; Almeida, A.; Pópulo, H.; Batista, R.; Lyra, J.; Pinto, V.; Coelho, R.; Celestino, R.; Prazeres, H.; Lima, L.; et al. Frequency of TERT promoter mutations in human cancers. *Nat. Commun.* **2013**, *4*, 2185. [[CrossRef](#)]
72. Thomas, D.; Sagar, S.; Liu, X.; Lee, H.-R.; Grunkemeyer, J.A.; Grandgenett, P.M.; Caffrey, T.; O'Connell, K.A.; Swanson, B.; Marcos-Silva, L.; et al. Isoforms of MUC16 activate oncogenic signaling through EGF receptors to enhance the progression of pancreatic cancer. *Mol. Ther.* **2021**, *29*, 1557–1571. [[CrossRef](#)] [[PubMed](#)]
73. Lu, M.; Zhao, B.; Liu, M.; Wu, L.; Li, Y.; Zhai, Y.; Shen, X. Pan-cancer analysis of SETD2 mutation and its association with the efficacy of immunotherapy. *npj Precis. Oncol.* **2021**, *5*, 51. [[CrossRef](#)] [[PubMed](#)]
74. Oh, J.-H.; Jang, S.J.; Kim, J.; Sohn, I.; Lee, J.-Y.; Cho, E.J.; Chun, S.-M.; Sung, C.O. Spontaneous mutations in the single TTN gene represent high tumor mutation burden. *npj Genomic Med.* **2020**, *5*, 33. [[CrossRef](#)]
75. Lu, N.; Liu, J.; Xu, M.; Liang, J.; Wang, Y.; Wu, Z.; Xing, Y.; Diao, F. CSMD3 is Associated with Tumor Mutation Burden and Immune Infiltration in Ovarian Cancer Patients. *Int. J. Gen. Med.* **2021**, *Volume 14*, 7647–7657. [[CrossRef](#)]
76. Sun, Y.; Li, L.; Yao, W.; Liu, X.; Yang, Y.; Ma, B.; Xue, D. USH2A Mutation is Associated with Tumor Mutation Burden and Antitumor Immunity in Patients with Colon Adenocarcinoma. *Front. Genet.* **2021**, *12*, 762160. [[CrossRef](#)]
77. Liu, Z.; Liu, L.; Jiao, D.; Guo, C.; Wang, L.; Li, Z.; Sun, Z.; Zhao, Y.; Han, X. Association of RYR2 Mutation with Tumor Mutation Burden, Prognosis, and Antitumor Immunity in Patients with Esophageal Adenocarcinoma. *Front. Genet.* **2021**, *12*, 669694. [[CrossRef](#)] [[PubMed](#)]
78. Lamberti, G.; Andriani, E.; Sisi, M.; Federico, A.D.; Ricciuti, B. Targeting DNA damage response and repair genes to enhance anticancer immunotherapy: Rationale and clinical implication. *Futur. Oncol.* **2020**, *16*, 1751–1766. [[CrossRef](#)] [[PubMed](#)]