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Advances of the small molecule drugs regulating fibroblast-like synovial proliferation for rheumatoid arthritis

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Rheumatoid arthritis (RA) is a type of chronic autoimmune and inflammatory disease. In the pathological process of RA, the alteration of fibroblast-like synoviocyte (FLS) and its related factors is the main influence in the clinic and fundamental research. In RA, FLS exhibits a uniquely aggressive phenotype, leading to synovial hyperplasia, destruction of the cartilage and bone, and a pro-inflammatory environment in the synovial tissue for perpetuation and progression. Evidently, it is a highly promising way to target the pathological function of FLS for new anti-RA drugs. Based on this, we summed up the pathological mechanism of RA-FLS and reviewed the recent progress of small molecule drugs, including the synthetic small molecule compounds and natural products targeting RA-FLS. In the end, there were some views for further action. Compared with MAPK and NF-KB signaling pathways, the JAK/STAT signaling pathway has great potential for research as targets. A small number of synthetic small molecule compounds have entered the clinic to treat RA and are often used in combination with other drugs. Meanwhile, most natural products are currently in the experimental stage, not the clinical trial stage, such as triptolide. There is an urgent need to unremittingly develop new agents for RA.

KEYWORDS

rheumatoid arthritis, fibroblast-like synoviocytes, signaling pathways, small molecule drugs, natural products

1 Introduction

Rheumatoid arthritis (RA) is a type of autoimmune joint disease. It often occurs in women and the elderly. RA might affect 0.5%–1% of the global population (Zhang et al., 2022). Among the multiple factors, genetic and autoimmune along with environmental factors might be the primary causes. It shows the clinical presentation of joint pain, thickening of the synovial membrane, pannus formation, and infiltration of various inflammatory cells in the joint space, leading to the damage of the cartilage as well as bone tissue, even remarkably joint deformity and dysfunction (Smolen et al., 2018). A lot of attention is paid to the treatment of RA because it has high morbidity, might lead to disability, and has poor prognosis (Davis et al., 2012; Almutairi et al., 2021). Currently, non-steroidal anti-inflammatory drugs (NSAIDs), disease-modifying anti-rheumatic drugs

(DMARDs) (synthetic or biologic agents), and glucocorticoids (Lampropoulos et al., 2015; Zhang et al., 2022) are popular in the treatment of RA. With the use of NSAIDs, the risk of cardiovascular disease might occur as well as gastrointestinal side effects, so a comprehensive evaluation is needed (O'Shea et al., 2013). DMARDs such as methotrexate (MTX), while suppressing inflammation and joint destruction, might cause nausea, anorexia, stomatitis, alopecia, myelosuppression, and even liver and pulmonary toxicity in severe cases, which requires careful monitoring. In addition, there are also problems of high expense and gastrointestinal adverse effects for DMARDs (Zhang et al., 2019). Biologic disease-modifying antirheumatic drugs (bDMARDs) show therapeutic effects for RA, but there are some individual differences because of different genetic backgrounds and environmental stimuli (Lampropoulos et al., 2015), and they do not cure the disease (Yamada, 2023). There is an urgent need to continuously develop new anti-RA drugs.

The synovium is considered to be a structure of connective softtissue membrane located in the joint cavity and the fibrocartilage, around arthrosis to provide nutrition and lubrication (Jay et al., 2000). The fibroblast-like synoviocytes (FLSs) are highly specialized mesenchymal cells found in the synovial membrane. In normal physiological regulation, FLS produces joint lubricants, for example, hyaluronic acid which nourishes the cartilage surface and shapes the synovial extracellular matrix (ECM). However, in RA, FLS exhibits a distinctive aggressive phenotype, with this aggressive behavior toward the ECM further exacerbating joint damage (Nygaard and Firestein, 2020). For this reason, one potential strategy for treating RA is the creation of medicines that target FLS (Bartok and Firestein, 2010). It is important to note that several of their monomers appear to have a positive impact on preventing arthritic synovial hyperplasia. They are mainly related to the induction of apoptosis and the inhibition of FLS proliferation. In this review, taking the state of FLS as a starting point, we summarize and discuss the literature on the small molecule drugs of FLS from PubMed, Embase, and other databases in the recent 3 years until 28 February 2023. Specific keywords used are "RA," "FLS," "MAPK," "NF-κB," "JAK/STAT," "Wnt," and "signaling pathways." The small molecule drugs contain organic compounds with low molecular weights, typically ≤1000 Da. Also, these include both synthetic compounds and natural products derived mainly from plants and animals. Publications with incomplete data or conclusions and those not directly related to RA and small molecule compounds are excluded. Here, first, there is an introduction of the pathological mechanisms of RA-FLS. Second, according to the signaling pathways controlling the abnormal behavior of FLS, small molecule drugs of related pathways, especially drugs with high anti-RA-FLS potential, are analyzed in depth. Finally, we list our comments, which we hope will provide directions to developing targeted anti-rheumatic drugs for clinics.

2 FLS involved in the pathogenesis of RA

In RA, FLS proliferation releases several anti-inflammatory cytokines and growth factors, among which are tumor necrosis factor (TNF), interleukin (IL) (such as IL-6, IL-1 β , and IL-17), chemokines, and inflammatory enzymes [such as nitric oxide

synthase (NOS) and cyclooxygenase-2 (COX-2)]. Meanwhile, it provides the inflammatory microenvironment and potentially contributes to the initiation of chronic inflammation in the preliminary stage of RA. In addition, FLS produces large amounts of receptor activator of NF-KB ligand (RANKL), vascular endothelial growth factor (VEGF), matrix metalloproteinases (MMPs), and so on, which causes synovial hyperplasia and arthritic joint destruction (Wang et al., 2012). Worse still, the activated FLS migrates to the cartilage and bone. This migration occurs not only at local sites but also through the bloodstream into distant areas and joints, destroying the cartilage, activating osteoclasts, and enhancing joint destruction in RA (Neumann et al., 2010; Hu et al., 2019). Here, we review the pathological mechanisms of RA from the three perspectives shown in Figure 1: synovial hyperplasia, joint damage, and immune inflammation.

2.1 Synovial hyperplasia

The synovium of RA exhibits endothelial hyperplasia and transformation into pannus tissue that destroys the articular cartilage and bone, with occasional lymphatic-like aggregates. A large number of inflammatory cytokines (IL-1β, TNF-a, etc.) stimulate FLS to proliferate abnormally and exhibit antiapoptosis. The imbalance between FLS anti-apoptotic and proapoptotic factors increases the number of FLS considerably, which directly leads to synovial hyperplasia. The FLS in the synovial lining layer is increased from the normal 1-3 to 10-15 cell layers (Neumann et al., 2010). The proliferated FLS develops into lymphoid-like structures, interacting with immune cells to form lymphoid organs and releasing pro-inflammatory factors and inflammatory mediators. Growth factors, such as platelet-derived growth factor (PDGF), transforming growth factor- β (TGF- β), and stimulatory cytokines in the synovial tissue, induce FLS proliferation through the activation of the signaling pathway. Along with the in situ proliferative capacity of FLS, the expression of anti-apoptotic molecules is also increased. The anti-apoptotic molecule FLICE inhibitory protein (FLIP) apoptosis-triggering suppresses intracellular cystatase-8, decreasing apoptosis and causing synovial proliferation (Bartok and Firestein, 2010).

2.2 Joint damage

Cartilage and bone destruction are hallmarks of RA. MMPs expressed by FLS degrade the chondral matrix, leading to impaired nutrient supply to the articular cartilage and tissue joint destruction.

2.2.1 Chondral matrix degradation

FLS mediates the overproduction of MMPs that interrupts the joint tissue, which contains a structure abundant in collagen and facilitates FLS infestation into the cartilage surface. Mediated by proinflammatory cytokines and toll-like receptors (TLRs), FLS upregulates the expression of MMPs, which activate osteoclasts and directly erode the bone, causing cartilage and bone destruction. Activated osteoclasts can reduce bone mass in the



Pathological mechanisms of RA with FLS (In RA, the proliferation of FLS resulting from synovial hyperplasia releases various anti-inflammatory cytokines and growth factors. Meanwhile, the interaction between FLS and immune cells causes a transformation of regular FLS into an aggressive phenotype, resulting in an abnormal situation of T-cell and B-cell functions related to immune inflammation. Furthermore, FLS secretes pro-inflammatory cytokines into the joint space and invades the adjacent bone tissue through migration, inducing bone erosion and joint destruction. Macrophages also differentiate directly into mature osteoclasts).

periarticular bone early in the lesion, leading to osteoporosis. In addition, the extra expression of MMPs upregulates the levels of inflammatory factors and soluble mediators in the synovial tissue. Also, the factors are bound to receptors of MAPK, JAK/STAT, etc., signaling pathways, promoting and maintaining joint inflammation (Firestein, 2003).

2.2.2 Bone destruction

The migration of FLS is also the process of bone destruction. Due to the cytokines, FLS can migrate into the cartilage and bone, thus exacerbating cartilage destruction (Zeng et al., 2017). FLS produces RANKL in the cartilage or bone. Then, RANKL binds to the receptor activator of NF- κ B (RANK) on osteoclast precursors, inducing osteoclast differentiation, activation, and production. A large number of osteoclasts erode the surface of the adjacent articular cartilage membrane and induce bone destruction. Not only that, RA-FLS hinders the recovery process of bone erosion by hindering osteoblast activation through the secretion of dickkopf-1 (DKK-1). DKK-1 is a crucial regulatory molecule within the Wnt pathway, acting as an inhibitor of osteoclast function (Miao et al., 2013). Under specific microenvironmental conditions, macrophages can also differentiate directly into mature osteoclasts. In addition, inflammatory macrophages are a consistent source of matrix

metalloproteinases, such as MMP-1, MMP-3, MMP-7, MMP-10, MMP-12, MMP-14, and MMP-25, which participate in connective tissue transformation and joint surface erosion observed in RA.

2.3 Immune inflammation

FLS are known to contribute significantly to RA by secreting inflammatory chemokines that interact with synovial infiltrating cells. The chemokines secreted by FLS, including, CXC motif chemokine 8 (CXCL-8), CXCL-10, and CC motif chemokine ligand 2 (CCL2), can recruit a range of immune cells into the synovial tissue. Then, the inflammatory mediators, for example, IL, TNF- α , and TGF- β 1, from these immune cells in turn stimulate FLS activation, resulting in a vicious circle. Macrophages are constantly affected by inflammatory stimuli and participate in the development of chronic synovitis, bone erosion, and cartilage erosion. Macrophages express a lot of molecules on their surface, such as Fc-gamma receptors (FcyRs), TLR, and the major histocompatibility complex class II (MHCII), which in turn, regulate their own activities, activate other cells in the local microenvironment, or attract immune cells outside the joint. TNF-a, IL-6, IL-1β, IL-23, and a wide range of CXCL and CCL chemokines promote and maintain

inflammation by recruiting and activating polymorphonuclear leukocytes, T cells, B cells, or monocytes.

2.3.1 FLS and B cells

There is a bidirectional signaling between FLS and B cells. On one hand, FLS affects the maturation and growth of B cells by secreting cytokines. The etiology of autoimmune disorders involves both humoral immunity and B lymphocytes as significant contributors. The preservation of the B-cell pool and humoral immunity depend on the B-cell-activating factor of the TNF family (BAFF, also known as BLYS) and a proliferation-inducing ligand (APRIL). Taking TLR-3 as an example, TLR-3 triggers not only B-cell-activating BAFF but also APRIL. Both of them participate in the stimulation of B cells, thus prolonging B-cell survival (Bombardieri et al., 2011; Leah, 2011). On the other hand, B cells in turn induce the FLS inflammatory phenotype. In the FLS co-culture experiments with age-associated B cells (ABCs), ABCs induce FLS phenotype excitation through TNF-a inducing the activation of ERK1/2 and JAK-STAT1 signaling pathways, consequently promoting the persistence of RA (Qin et al., 2022).

2.3.2 FLS and T cells

T-cell infiltration and excessive proliferation of FLS are significantly upregulated in RA patients. Both interact during RA inflammation to perpetuate inflammation. RA-FLS can present peptides of inflammatory antigens to antigen-specific T cells, contributing to the auto-reactive immune response in RA (Tran et al., 2007). Then, FLS expresses adhesion molecules, transmitting signals to CD4 T cells, such as vascular cell adhesion molecule-1 (VCAM-1) and intercellular cell adhesion molecule-1 (ICAM-1). Finally, these adhesive molecules interact with integrins, for instance, lymphocyte function-associated antigen 1 (LFA-1), resulting in CD4 T-cell proliferation and IL-17 secretion and exacerbation of the inflammatory response (Mori et al., 2017). At the same time, macrophages express MHCII as antigen-presenting cells, thereby participating in the activation and recruitment of pathogenic T cells. So, there is also an interaction between T cells and FLS (Tran et al., 2008; Tu et al., 2022).

To sum up, FLS can secrete pro-inflammatory cytokines such as TNF- α , IL-1 β , IL-6, and MMP, in the joint space of RA patients and invade the adjacent bone tissue through migration, inducing bone erosion and joint destruction. The interaction between FLS and immune cells causes a transformation of regular FLS into an aggressive phenotype, resulting in abnormal T- and B-cell functions. Also, our body gradually loses its normal immune regulatory and protective ability (Ding et al., 2023). It is evident that FLS is the central effector cell in the pathogenesis. Given that there is no effective treatment targeted at FLS, the inhibition FLS-mediated pro-inflammatory response and subsequent tissue destruction seems to be a feasible strategy for RA (Nygaard and Firestein, 2020). In the next part, we summarize the results in the recent 3 years of small molecule drugs targeted at FLS.

3 Small molecule drugs regulating FLS

In the previous sections, we have clarified that RA-FLS are activated by multiple cytokines involved in the activation of FLS. Targeted pathways of FLS might simultaneously block multiple signaling of cytokine receptors, inhibiting the activation, proliferation, and invasion of FLS and, thus, significantly controlling RA synovial inflammation and joint damage (Mavers et al., 2009; Wendling et al., 2010; Pan et al., 2016). Despite significant breakthroughs in RA therapy, many people with RA have persistent disease. The current RA therapy plans emphasize reducing T-cell and B-cell activity as well as cytokine signaling (Mahmoud et al., 2022). In RA, targeting signal transduction pathways is an emerging treatment option. According to the signaling pathway interacted with FLS, there are mainly MAPK, NF-ĸB, JAK/STAT, PI3K/Akt, and Wnt signaling pathways in Figure 2. So, we present the drugs' research progress which regulates FLS function on the signaling pathways, including the small molecule compounds and natural products. It is aimed to explore promising novel drug development directions and broaden the path of novel targeted FLS.

3.1 Small molecule drugs targeting MAPK regulating FLS

The MAPK signaling pathway is associated with various kinases, such as P38, c-Jun N-terminal kinase (JNK), and extracellular regulated protein kinases (ERKs), which are involved in the proliferation, apoptosis, and migration of FLS, with the addition of cytokine secretion (Harigai et al., 2004; Tang et al., 2019). ERK is involved in the secretion of certain cytokines and cell proliferation and differentiation through the regulation of B-cell lymphoma 2 (Bcl-2). JNK decreases proteoglycan synthesis and enhances MMP-13 synthesis, which are necessary for bone deterioration and joint inflammation. p38 is associated with the cytokine secretion of MMP. Through inhibiting p38, MMP reduces cartilage degradation and inhibits osteoclast formation. Additionally, the MAPK pathway contributes to the FLS's increase in TNF-a expression, amplifying inflammatory signals, inducing FLS proliferation, aggravating inflammation, and damaging joints (Zuo et al., 2015; Kadkhoda et al., 2016). An increasing number of studies have shown that the MAPK pathway is activated in immune and autoimmune response conditions, regulating the cell responses of division, differentiation, apoptosis, inflammation, and stress and also participating in the activation of FLS (Müller-Ladner et al., 2007; Bustamante et al., 2017). In addition, MAPK activates downstream transcription factors that promote synovial cell proliferation and chondrocyte apoptosis. It also leads to high expression of multiple MMPs in synovial cells and chondrocytes and overhydrolysis of the extracellular matrix, resulting in joint damage. Therefore, MAPK is one of the most studied targets to inhibit RA-FLS (Wang et al., 2010).

Here, we review the synthetic small molecule compounds and natural products in the recent 3 years targeted to MAPK for FLS in Table 1, and the natural products regulating MAPK are shown in Figure 3. It is important to note that the majority of drugs affected numerous signaling pathways and multiple targets. As an MAPK downstream effector, p38 is considered



a possible target for RA, but only few p38 inhibitors have been tested in humans. Tacrolimus as a macrolide calcineurin inhibitor immunosuppressant drug decreased the production of angiopoietin-1 (Ang1), tyrosine-protein kinase receptor (Tie-2), and VEGF in human FLS by preventing the activation of the IL-1β-mediated JNK and p38 MAPK pathways. Sugiura et al.'s (2020) study was very interesting. They found that glycogen synthase kinase 3 (GSK-3) inhibitors significantly reduced synovial fibroblast migration after 72 h and decreased Akt phosphorylation [Ser (473)] after 48 h in vitro, which might have therapeutic efficacy targeting the invasion and migration of synovial fibroblasts. Also, 3'3-diindolylmethane exhibited the possibility of anti-RA-FLS activitiy in vivo and in vitro (Du et al., 2019). The small molecule compounds reported in recent years that could alter FLS in vivo and in vitro were elutriated extirpate, dasatinib, 4-phenylbutyric acid, and 3-(4-hydroxy-3-methoxyphenyl)-1-3-[1]-phenyl-propenone. Unfortunately, these medications are still in the laboratory stage. Because of their poor performance, p38 inhibitors have limited efficacy in RA treatment. Also, blocking p38's downstream had a compensatory effect on other kinases, so alternative options for p38 have been progressively explored (Guma et al., 2012). Regulation of MAPK kinases upstream of p38, the human mitogen-activated protein kinase kinase (MKK), such as MKK6 and MKK1, could selectively block the production of MMPs and proinflammatory cytokines in FLS (Hammaker et al., 2012). In addition, ubiquitin D might be considered a possible therapeutic target for RA-FLS (Chen et al., 2023).

In natural products in Table 1 and Figure 3, alkaloids and flavonoids were more frequently reported and studied for their effects on the MAPK signaling pathway of FLS. Other categories, such as iridoids and saponins, were also found to have an impact. It is well known that flavonoids possess anti-oxidant and antiinflammatory properties. Flavonoids can inhibit the inflammatory response and reduce the symptoms of inflammation while scavenging free radicals, reducing oxidative stress, and protecting cells from oxidative damage. Flavonoids usually inhibit FLS proliferation, migration, and invasion by inhibiting p38 and JNK. To our surprise, alkaloids also showed up significantly in the treatment of FLS. Preparations of berberine and paclitaxel were available for clinical use, but they have no indication for the treatment of RA.

Triptolide and tetrandrine from *Tripterygium wilfordii* Hook F. and *Stephania tetrandra* root, respectively, have antirheumatic effects in the classic sense. Tripterygium glycoside preparations have been clinically used for the treatment of RA. As the representative, we concentrate on triptolide, which has been studied more and has been proven to have multiple signaling pathways. The treatment with triptolide decreased the expression of phosphorylated JNK that TNF- α -produced, but it had no effect on the expression of phosphorylated p38 or

Name	Source	Targets/ signaling pathways	Estimate	References		
The synthetic small molecule	e compounds					
GSK-3 inhibitors (6-	Serine/threonine protein kinase	JNK, p38, NF-κB	Experimental: NF-κB ↓	Kwon et al. (2014); Sugiura et al.		
thiadiazolidinone-8)			The phosphorylated JNK, c-Jun, ATF-2, p38 ↓	(2020)		
			IL-6 ↓			
			IL-10 ↑			
Tacrolimus	Macrolide antibiotics from <i>Streptomyces</i>	JNK, p38	Clinical: showed higher retention rates combined with bDMARDs	Choe et al. (2012); Kaneko et al. (2021); Terabe et al. (2023)		
			Adverse events stable in long-term observation			
			Effective with acceptable safety			
			Experimental: the expressions of Ang-1, Tie-2, VEGF \downarrow			
3'3-Diindolylmethane	The main product of indole-3-carbinol oligomerization catalyzed by acid	p38, JNK, Akt, mTOR	Experimental: proliferation, migration, and invasion of RA-FLS <i>in vitro</i> ↓	Du et al. (2019)		
			MMP-2, MMP-3, MMP-8, and MMP-9 \downarrow p-p38, JNK \downarrow			
			Akt, mTOR \downarrow			
			Pro-inflammatory cytokines and arthritis severity in mice \downarrow			
Telotristat etiprate	A tryptophan hydroxylase inhibitor	МАРК	Experimental: migration and invasion of RA-FLS <i>in vitro</i> \downarrow	Zhang et al. (2023)		
			Targeting LGALS3			
Dasatinib	A Src kinase inhibitor	MAPK, STATs	Experimental: Src, Fyn, MAPK, STATs ↓	Yalcin Kehribar et al. (2021); Min et al. (2023)		
			MMP-1, MMP-3, MMP-13 in FLS \downarrow			
4-Phenylbutyric acid	An HDAC inhibitor	MAPK, NF-κB	Experimental: p-MAPK, p-NF-κB↓	Choi et al. (2021)		
			MMP-1, MMP-3, COX-2↓			
			Endoplasmic reticulum stress ↓			
3-(4-Hydroxy-3-methoxy-	A benzylideneacetophenone derivative	МАРК	Experimental: IL-8, IL-6, PGE (2) ↓	Sur et al. (2020)		
propenone			Reducing the inflammation in the knee joints in C/K-arthritic rats			
The natural products						
Fangchinoline	A bisbenzylisoquinoline alkaloid from Stephania tetrandra	MAPK, NF-κB	Experimental: inflammatory cytokine secretion and ROS in human FLS ↓	Villa et al. (2020)		
			Phosphorylation of the MAPK and NF- κ B pathway in human FLS \downarrow			
Berberine	An alkaloid from Coptis chinensis	PI3K/Akt, Wnt, RAS/MAPK/FOXO/	Clinical: no indication for treatment of RA	Wang et al. (2019); Shen et al. (2020); Sujitha et al. (2020); Li		
		пlf-1	Experimental: LRP5 protein ↓	et al. (2023); Li et al. (2023)		
			β -Catenin transcription \downarrow p38/ ERK \downarrow			
			Proliferation and adhesion of FLS ↓			
			MMP-1, MMP-3, RANKL, TNF-α↓			

(Continued on following page)

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Name	Source	Targets/ Estimate signaling pathways		References
Paclitaxel	An alkaloid from <i>Taxus chinensis</i>	MAPK, Akt/mTOR	Clinical: no indication for treatment of RA	Chen et al. (2021)
			Experimental: FLS migration dose dependently ↓	
			IL-6, IL-8, RANKL ↓	-
			MMP-8, MMP-9 gene transcription ↓ p-ERK1/2 ↓	_
			p-JNK↓	-
			Akt, p70S6K, 4EBP1, HIF-1α↓	_
Peimine	A steroidal alkaloid from Fritillaria	ERK, JNK, p38	Experimental: TNF-α induced destructive behaviors in MAPK for FLS↓	Zhou et al. (2022)
			RANKL-induced osteoclast formation ↓	-
			Bone-resorption function ↓	
Tetrandrine	An alkaloid from Stephania tetrandra	NF- κ B, Ca ₂ (+),	Experimental: Rac1, Cdc42, RhoA↓	Lv et al. (2015); Zhong et al.
	root	PI3K/Akt, MAPK	MMP-2/9, F-actin, FAK↓	(2019)
			RANKL-induced osteoclastogenesis ↓	
Dehydroevodiamine	A quinazoline alkaloid from Evodiae Fructus	МАРК	Experimental: pro-inflammatory factors in AIA rats ↓	Dai et al. (2022)
			MMP-1, MMP-3 \downarrow p-p38, p-JNK, and p-ERK \downarrow	
Tomatidine	A steroidal alkaloid from the Solanaceae family	МАРК, NF-кВ	Experimental: proliferation and migration of FLS \downarrow	Yu et al. (2021)
			Synovial inflammation and joint destruction in CIA rats \uparrow	
			IL-1β, IL-6, TNF-α ↓	-
			MMP-9, RANKL↓	-
Benzoylaconitine	An alkaloid from Aconitum	MAPK, Akt, NF-κB	Experimental: IL-6, IL-8 ↓	Yu et al. (2020)
			MAPK, p-Akt↓	_
			Degradation of IKB $\alpha \downarrow p$ -p65 and nuclear transposition \downarrow	
Kaempferol	A flavonoid from <i>Kaempferol</i> galanga L.	ERK-1/2, p38, JNK, NF-кВ	Experimental: MAPK activation \downarrow , instead of altering TNF- α receptor activation	Yoon et al. (2013); Pan et al. (2018)
			Phosphorylation of ERK-1/2, p38, JNK ↓	_
			NF-κB ↓	
Orientin	A flavonoid from <i>P. orientale</i>	p38, ERK	Experimental: viability, migration as well as invasion of FLS \downarrow	Ji and Xu (2022)
			TNFα-induced inflammatory makers ↓	

Name	Source	Targets/ Estimate signaling pathways		References
Apigenin-4'-O-alpha-L-	A flavonoid from apigenin derivative	МАРК	Experimental: migration of FLS \downarrow	Cao et al. (2022)
rhamnoside			MMP-1, MMP3, RANKL, TNF-α↓	
			MAPK1, HRAS, ATF-2, p38, JNK↓	-
Naringin	A flavonoid from citrus fruits	PI3K/Akt, ERK	Experimental: inflammation, MMPs ↓	Aihaiti et al. (2021)
			Apoptosis of FLS \uparrow the activation of caspase-3 \uparrow	-
			Bax/Bcl-2 ↑ p- Akt, p-ERK ↓	-
Liquiritin	A flavonoid from the roots of	JNK, P38	Experimental: FLS proliferation ↓	Zhai et al. (2019)
	Glycyrrhiza uralensis		DNA fragmentation in the nucleus \uparrow	-
			Altering the potential of the mitochondrial membrane	
			Bcl-2/Bax ratio ↓	-
			VEGF↓ p-JNK, p-p38↓	-
Neohesperidin	A flavanone glycoside from citrus fruits	МАРК	Experimental: IL-1β, IL-6, IL-8, TNF-α, MMP-3, MMP-9 and MMP-13 in FLSs ↓	Wang et al. (2021)
			МАРК ↓	
			ROS induced by TNF- $\alpha\downarrow$	
Ononin	An isoflavone glycoside from the fruit of <i>Cnidium monnieri</i> (L.) cusson	NF-кВ, МАРК	Experimental: TNF-α mediated cells viability of FLS and MH7A ↓	Meng et al. (2021)
			Cell apoptosis↑	
			IL-1β, IL-6 ↓	
Cyanidin	An anthocyanidin from grapes, bilberry, blackberry, etc.	p38, STAT-3	Experimental: IL-17A induced the migration of monocytes from AA rats ↓	Samarpita and Rasool (2021); Samarpita et al. (2020)
			HSP27, CCR7, CXCR4 ↓	
			RANKL ↓	
			OPG ↑ p38 MAPK ↓	
Cyanidin-3-glucoside	An anthocyanin from berries	p38, ERK and JNK, NF-кB	Experimental: TNF- α , IL-1 β , IL-6 \downarrow p65 \downarrow	Sun and Li (2018)
			Phosphorylation of IκBα, p38, ERK, JNK↓	-
Paris saponin VII Chonglou	A steroidal saponin from <i>Trillium tschonoskii</i> Maxim.	JNK, p38	Experimental: FLS invasion via managing the mitochondrial apoptosis, MAPK pathway	Meng et al. (2021)
			Improving histopathological changes	
			TNF-α, IL-1β, IL-6 ↓	
			Modulating the expressions of apoptosis proteins in AIA rats	

Name	Source	Targets/ signaling pathways	Estimate	References
Gintonin	A ginseng-derived exogenous ligand of lysophosphatidic acid	МАРК, NF-кВ	Experimental: iNOS, IL-6, TNF-α, COX-2↓	Kim et al. (2021); Kim et al. (2021)
			NF- κ B/p65 into the nucleus \downarrow	
Triptolide	An epoxide diterpene lactone from	JNK, MAPK8,	Experimental: p-JNK ↓	Yang et al. (2016); Xie et al.
	Tripterygium wijorati Hook F.	PI3K/AKt	The polymerization of F-actin \downarrow	(2019); Song et al. (2020)
			The activation of MMP-9 ↓	-
			Activating autophagy	-
Geniposide	An iridoid glycoside from Gardenia	JNK, ERK1/2 and	Experimental: proliferation of FLS \downarrow	Li et al. (2018); Bu et al. (2022)
	<i>jasminoides</i> Ellis fruit	p38; P13K; Akt	IFN- γ , IL-17 \downarrow	
			IL-4, TGF β 1↑ p-JNK, p-ERK1/2, p-p38 ↓	-
			p-PI3K, p-Akt ↑	
Gentiopicroside	A secoiridoid glycoside from <i>Gentiana</i>	CD147, p38, NF-кВ	Experimental: proliferation of FLS \downarrow	Jia et al. (2022)
	<i>macropnyua</i> Pail.		MMP secretion↓	
			Regulating the CD147/p38/NF- κ B pathway, p38, Ik κ B α , and p65 \downarrow	
18β-Glycyrrhetinic acid	A triterpene glycoside from <i>Glycyrrhiza</i>	MAPK, NF-κB	Experimental: IL-1 β , IL-6, COX-2 in MH7A \downarrow	Feng et al. (2021)
			Cell viability	-
			Cell apoptosis and G1 phase cell cycle arrest <i>in vitro</i> \uparrow	
			FOXO3 ↑	-
			Liver damage caused by collagen or MTX <i>in vivo</i> ↓	
			Inflammation and proliferation in FLS \downarrow	
Pristimerin	A triterpenoid from Celastraceae and Hippocrateaceae families	MAPK/Erk1/2, PI3K/Akt	Experimental: viability and migration of FLS \downarrow	Lv et al. (2022)
			TNF-a, NO, p-Akt, p-ERK ↓	
Echinocystic acid	A pentacyclic triterpene from <i>Gleditsia</i> sinensis	MAPK, NF-κB	Experimental: arthritis symptoms in SKG mice \downarrow	Cheng et al. (2022)
			TNF-a, IL -6, IL-1 $\beta \downarrow$	-
			P-STAT3↓	-
			MAPK, NF-κB	-
Osthole	A coumarin from <i>Cnidium monnieri</i>	NF-кВ, МАРК	Experimental: IL-1 β , TNF- α , IL-6 \downarrow	Xu et al. (2018); Lin et al. (2023)
	and Angelica pubescens		Proliferation and migration \downarrow	
			TGM2/Myc/WTAP-positive feedback circuit ↓	
Imperatorin	A coumarin from Umbelliferae	p38, ERK NF-кВ	Experimental: proliferation and migration of FLS \downarrow	Lin et al. (2022)
			TNF- α , IL-6, and IL-8 \downarrow p38, ERK \downarrow	
			р-ІкВа ↓	

Name	Source	Targets/ signaling pathways	Estimate	References
Tanshinone IIA	A diterpene quinone from Salvia miltiorrhiza Bunge	MAPK, Akt/mTOR, HIF-1, and NF-кВ	Experimental: FLS proliferation, migration, infiltration time, and dose dependently \downarrow	Du et al. (2020)
			MMPs, pro-inflammatory factors ↓	
Piceatannol	A derivative of resveratrol	MAPK, NF-ĸB	Experimental: Bax, cleaved caspase- 3 ↑	Gao et al. (2022)
			PGE2, IL-6, IL-1β↓	
			COX-2↓	
			MMP-3, MMP-13 ↓	
			MAPK, NF-κB↓	

 \downarrow : suppress, downregulate, inhibit, block, prevent, reduce, decrease; \uparrow : promote, upregulate, active, increase. mTOR, mammalian target of rapamycin; NFATc1, c-Fos and nuclear factor of activated T cells c1; ATF2, activating transcription factor-2; PGE2, prostaglandin E2; ROS, reactive oxygen species; HIF1, hypoxia-inducible factor 1; CIA, collagen-induced arthritis; IkB, inhibitor of κ B; Bcl-2, B-cell lymphoma 2; Bax, Bcl-2-associated X; AA, adjuvant-induced arthritic; OPG, osteoprotegerin; MEKK, mitogen-activated protein kinase kinase; IKK, IkB kinase; TGM2, transglutaminase 2.



ERK (Yang et al., 2016) and reduced FLS migration and invasion by targeting the JNK/MAPK signaling pathway (Tang et al., 2020). Triptolide dramatically increased the p-Akt/Akt ratio, and inhibiting the PI3K/Akt signaling pathway in MH7A cells caused autophagy to be triggered, indicating that triptolide repressed autophagy via activating p-Akt/Akt (Xie et al., 2019). Other natural products, such as Paris saponin VII/ Chonglou, geniposide, and gentiopicroside, shown in Table 1, also have the potential to regulate FLS against RA. However, it is currently in the experimental stage.

3.2 Small molecule drugs targeting NF- κB regulating FLS

As a major signaling transcription factor, NF- κ B contributes to synovial inflammation, proliferation, and decay in bones in RA and regulates inflammatory gene expression and cell proliferation. Both innate and adaptive immune cells include NF-B, which is a key mediator of the stimulation of pro-inflammatory genes (Liu et al., 2017). In a normal situation, NF- κ B is bound to its repressor protein I κ B and not activated. The nuclear-localization sequence (NLS) that

TABLE 2 Small molecule drugs targeting NF- $\!\kappa B$ regulating FLS.

Name	Source	Targets/signaling pathways	Estimate	References
The synthetic small mole	cule compounds			
TAK-242	A TLR 4 antagonist	TLR4, TLR3; NF-κB	Experimental: TLR4, TLR3 ↓	Samarpita et al. (2020)
			The migration of NF- κ B to the nucleus	
			IL-8, IL-1, MMP-7 ↓	
CKD-506	A HDAC inhibitor	NF-ĸB	Experimental: MMP-1, MMP-3, IL-6, IL-8 ↓	Park et al. (2020)
			The proliferation of Teff \downarrow	
			Exerting a synergistic effect with MTX	
Oxymatrine hydrazone	Synthesized from oxidized bitter	MEK/1/2, NF-κB	Experimental: IL-1β, IL-6, IL-8 ↓	Zhang et al. (2021)
	ginseng		MMP-1, MMP-13 ↓	
			MEK/1/2 and p65 phosphorylation \downarrow	
Paeoniflorin-6'-O-benzene sulfonate (CP-25)	A paeoniflorin derivative	NF-ĸB, PI3K, GRK2	Experimental: the protein membrane expression and combination↓	Wang et al. (2020); Wang et al. (2023)
Edaravone	Synthetic: 3-methyl-1-phenyl-2- pyrazolin-5-one	NF-кВ, МАРК	Clinical: no indication for the treatment of RA	Zhang et al. (2020); Liu et al. (2023)
			Experimental: altering the antioxidant factors, inflammatory mediators, and pro-inflammatory cytokines [NF-κB, COX-2, and PGE (2)]	
			The level of cytokines and OPN, RANKL, and macrophage M-CSF \downarrow	
Roflumilast	An inhibitor of phosphodiesterase-4	NF-ĸB	Clinical: no indication for the treatment of RA	Zhong et al. (2021)
			Experimental: ROS and MDA in MH7A cells \downarrow	
			IL-6, IL-8, TNF-α↓	
			CCL5, CXCL9, CXCL10 \downarrow	
			MMP-1, MMP-13 ↓	
Sorafenib	A kinase inhibitor	NF-κB, c-Jun	Clinical: no indication for treatment of RA	Wang et al. (2020)
			Experimental: apoptosis in AA FLSs \downarrow	
			Fas, caspase-3, Mcl-1 ↑	
			NF-κB, C-Jun ↓	
Dexmedetomidine	A specific and selective alpha-2 adrenoceptor agonist	NF-ĸB	Clinical: no indication for treatment of RA	Ji et al. (2020)
			Experimental: IL-1β, IL-6, IL-17A, TNF-α, and P-P65↓	
			NLRC5 ↓	
Alogliptin	An important selective dipeptidyl peptidase-4 inhibitor	NF-ĸB	Clinical: no indication for the treatment of RA	Guo et al. (2020)
			Experimental: MMP-3, MMP-13, IL-6, IL-8, and TNF- α p- Jun, p-I κ B β , nuclear translocation of NF- κ B p65 \downarrow	

TABLE 2 (Continued) Small molecule drugs targeting NF- κB regulating FLS.

Name	Source	Targets/signaling Estimate pathways		References	
The natural products					
Diosmetin	A flavonoid from Rutaceae	NF-ĸB	Experimental: proliferation of MH7A cells ↓	Chen et al. (2020)	
			IL-1β, IL-6, IL-8, MMP-1 ↓		
			and NF- κB pathways activation \downarrow		
Mangiferin	A flavonoid of the bisphenirone family from mango leaves	ERK2, p38, NF-кВ	Experimental: MAPKs (ERK2 and p38), NF-кB ↓	Luczkiewicz et al. (2014); Wang et al. (2021)	
Icariin	A flavonoid glycoside from Epimedii Herba	NF-ĸB	Experimental: TRIB1 ↑ by promoting Nrf2 expression regulating the TRIB1/ TLR2/NF-кВ pathway	Wu et al. (2022)	
Isoginkgetin	A biflavonoid from the leaves of the	ІкВβ, р65	Experimental: IL-1β, IL-6, IL-8 ↓	Shao et al. (2022)	
	Ginkgo buoba tree		Migration and invasion of FLS↓ p-IκBα, p-p65, MMP9↓		
Tectoridin	An isoflavone from dry rhizome of	TLR4/NLRP3/NF-κB	Experimental: proliferation of FLS \downarrow	Huang et al. (2022); Niu et al.	
	1115	MAFK	Cleaved caspase-3, Bax ↑	(2022)	
			Bcl-2↓	-	
			Pro-inflammatory cytokines ↓		
			TLR4/NLRP3/NF-κB↓		
			ERK, JNK, p38 \downarrow		
Celastrol	A quinone-methylated triterpenoid from <i>Tripterygium wilfordii</i>	NF-κB, Notch1, ERK,	Experimental: NF-кВ pathway ↓	Gan et al. (2015); Yu et al. (2015); Doss et al. (2016); Fang et al. (2017); An et al. (2020); Yang et al. (2022)	
			NLRP3 inflammasome activation↓		
			ROS ↓	et al. (2022)	
			Changing some chemokine genes expression (CCL2, CXCL10, CXCL12, CCR2 and CXCR4)		
			SYK-MEK-ERK-NF-кB signaling cascade↓		
			Autophagy ↑		
			PI3K/Akt/mTOR↓		
Aucubin	A monoterpenoid from asterids	NF-κB	Experimental: inflammatory factors ↓	Zhang et al. (2022)	
			Bone metabolism factors \downarrow p-Ikk $\alpha/\beta,$ p-IkBa, p-p65 \downarrow		
Heilaohuacid G	A triterpenoid from <i>Kadsura</i> <i>coccinea</i> /heilaohu	NF-κB	Experimental: apoptosis and inflammatory reactions of FLS↓	Yang et al. (2021); Yang et al. (2022)	
Sinomenine	An alkaloid from <i>Sinomenium</i>	NF-κB	Experimental: adenosine receptor \uparrow	Zhou et al. (2017); Yi et al. (2021); Chen et al. (2011): Li et al. (2012):	
			NF-κB activation via α7nAChR↓	Chen et al. (2011); Li et al. (2013); Zhou et al. (2015); Yao et al. (2017)	
			Selective mPGES-1 expression ↓		
			TLR4/MyD88/NF-κB signaling cascade↓		

TABLE 2 (Continued) Small molecule drugs targeting NF- κB regulating FLS.

Name	Source	Targets/signaling pathways	Estimate	References
Magnoflorine	An alkaloid from <i>Clematis</i> manshurica Rupr.	РІЗК/Akt/NF-кВ, Keap1-Nrf2/HO-1	Experimental: proliferation, migration, invasion, and reactive oxygen species levels of MH7A cells ↓	Shen et al. (2022)
			Bax ↑	
			Bcl-2↓ iNOS, COX-2, IL-6, IL-8, MMPs ↓	
			PI3K/Akt/NF-κB↓	
			Keap1-Nrf2/HO-1 ↑	
Curcumin	A polyphenol from turmeric, curcuma longa	NF-κB, AP-1, and p38	Experimental: function of pro- inflammatory mediators↓	Buhrmann et al. (2010); Shang et al. (2016); Mohammadian
			Osteoclastogenic potential	et al. (2022)
			ERK1/2, p38, JNK ↓	-
			RANK, c-Fos, NFATc1 levels↓	
Punicalagin	A polyphenol from pomegranate juice	NF-κB	Experimental: IL-1 beta, IL-6, IL-8 and IL-17A \downarrow	Huang et al. (2021)
			MMP-1 and MMP-13 \downarrow	
			Proliferation and migration of RA FLSs \downarrow phosphorylation of IKK and IkBa \downarrow	
Corilagin	A tannic acid from <i>Geranium</i> wilfordii Maxim.	NF-κΒ p65, ERK, p38, JNK, ΙκΒα	Experimental: Bcl-2, IL-6, IL-8, MMP- 1, MMP-2, MMP-3, MMP-9, COX-2, iNOS ↓	Shen et al. (2022)
			Bax ↑	
			Р-р65/р65, Р-ІкВа/ІкВа, Р-ERK/ERK, Р-JNK/JNK, and Р-р38/р38↓	
			NF- κ B p65 nuclear translocation \downarrow	
			Proliferation, migration, and invasion of FLS \downarrow	
Eugenol	A phenylpropanoid from a variety of aromatic herbal plants such as clove and tulsi	NF-ĸB	Experimental: proliferation, migration, invasion, angiogenesis, and inflammatory response of FLS ↓	Wang et al. (2022)
			NF-κB, COX-2 ↓	
Resveratrol	A phenol from grape	SIRT1, NF-κB	Experimental: SIRT1 and downstream paths \uparrow	Wang et al. (2020); Sheng et al. (2022)
			The striking interplay between the SIRT1 and NF-κB	
Plumbagin	A naphthoquinone from <i>Plumbago</i>	p65	Experimental: viability of human FLS	Shu et al. (2022)
	zeylanica L.		Inflammatory cytokines, MMPs ↓	-
			IкВ, NF-кВ, p65 into the nucleus↓	
Emodin	An anthraquinone from rhubarb, buckthorn, etc.	МАРК, NF-кВ	Experimental: proliferation of the MH7A cell \downarrow	Cao et al. (2022)
			MAPK, PTGS2 ↓	
			CASP3↑	
Aucubin	An iridoid glycoside from <i>Eucommia ulmoides</i> Oliv.	NF-κB	Experimental: migration and invasion of human FLS \downarrow	Zhang et al. (2022)
			NF-кВ -p65 activity of MC3T3-E1 cells $\downarrow p$ -Ікка β , p-Ік β , and p-p65 proteins \downarrow	

pathways	
CantleyosideAn iridoid glycoside from Pterocephalus hookeri (C. B. Clarke) HoeckAMPK/Sirt 1/NF- κ BExperimental: proliferation of human FLS \downarrow Bai et al. (2022)NO, TNF- α , IL-1 β /6, MCP-1 and MMP-1/3/9 \downarrow OCR, ECAR and real-time ATP generation rate p-NF- κ B and true learting and the second sec	

L: suppress, downregulate, inhibit, block, prevent, reduce, decrease; ↑: promote, upregulate, active, increase. HDAC, histone deacetylase; PGE (2), prostaglandin E (2); GRK2, G protein-coupled receptor kinase 2; M-CSF, macrophage colony stimulating factor; MDA, malondialdehyde; TRIB1, Tribbles pseudokinase 1; NFATc1, nuclear factor of activated T cells; NLRP3, NOD-like receptor protein 3; HO-1, heme oxygenase; SIRT1, silent information regulator 1; MCP-1, monocyte chemotactic protein-1; OPN, osteopontin; ATP, adenosine triphosphate; α7nAChR, α7 nicotinic acetylcholine receptor; mPGES-1, microsomal prostaglandin E synthase 1; AP-1, activated protein-1.



belongs to NF-KB is covered by the IKB unable to undergo nuclear translocation. However, in RA due to the activators (TNF-a, IL-17, etc.), IkB is phosphorylated, ubiquitinated by IkB kinase, and eventually degraded by the enzyme, releasing NF-KB. Following that, NF-KB p65 enters the nucleus and combines with target genes (Aupperle et al., 1999). The production of inflammatory mediators such as TNF-a, COX-2, and IL-1ß increases as a result of this nuclear translocation in the synovium. Those activated sustaining states lead to massive abnormal activation of FLS (Saravanan et al., 2014). NF-KB p65 regulates apoptosis and inhibits protein expression, which has an antagonistic effect on apoptosis in FLS (Kadkhoda et al., 2016), leading to synovial hyperplasia and aggravating joint destruction (Yin et al., 2015). In addition, p38 mediates IkB phosphorylation, which is involved in regulating NF-kB activation (Carter et al., 1999; Kaminska, 2005).

The small molecule drugs and natural products targeted at NF- κ B in recent 3 years are summarized in Table 2, and the classification of the natural products is in Figure 3. There have been many studies on small molecule compounds that modulate FLS in the NF- κ B

signaling pathway, such as TAK-242 (Samarpita et al., 2020), CKD-506 (Park et al., 2020), and synthetic derivatives from natural products that also showed the activity of inhibiting proliferation. For example, oxymatrine hydrazone synthesized from oxidized bitter ginseng induced apoptosis and prevented TNF-a-mediated enhanced viability of RA-FLS (Zhang et al., 2021). Paeoniflorin-6'-O-benzene sulfonate (CP-25), a paeoniflorin derivative, had the ability to decrease membrane expression and the combination of these proteins (Wang et al., 2020; Wang et al., 2023). Edaravone, roflumilast, sorafenib, dexmedetomidine, and alogliptin have been used clinically, without the indication for the treatment of RA. The existing experiments showed that they have the anti-proliferation ability of FLS and were worthy of inclusion in the secondary development of drugs. In the natural products in Figure 4, flavonoids still predominated, such as diosmetin, icariin, isoginkgetin, and tectoridin. In a similar situation with the MAPK inhibitions for RA-FLS, these natural products were in the experimental stage. In addition, some inhibitors modulated both NF-KB and MAPK pathways to regulate FLS activity, such as tectoridin and corilagin.

TABLE 3 Small molecule drugs targeting JAK/STAT regulating FLS.

Name	Source	Targets/signaling pathways	Estimate	References
The synthetic s	mall molecule compounds			
Peficitinib	A JAK inhibitor	JAK1, JAK2, JAK3, and Tyk2; STAT3	Clinical: phase II and III clinical trials and extension studies completed	Emori et al. (2020); Gutierrez-Urena et al. (2020); Kitanaga et al. (2020)
			Showed efficacy, safety, and tolerability in monotherapy or csDMARDs	
			Experimental: STAT3 phosphorylation by diversified cytokine concentration- dependently ↓	
			Growth factor-A, MMPs, IL-6, TNFSF11 ↓	
Filgotinib	A selective JAK1 inhibitor	JAK1	Clinical: under clinical trial pending approval for use in RA	Shimizu et al. (2023); Westhovens (2023)
			Dose-related effect was not observed for safety excepting for herpes zoster and the increases of lipids and creatine phosphokinase	
Takinib	A selective TAK1 inhibitor	TAK1, TAK3, JNK, NF-κB	Clinical: JAK-STAT pathways in RA patients ↓	Palmroth et al. (2021); Panipinto et al. (2021); Mardani et al. (2023)
			One case of liver failure	
			Experimental: p-TAK1, no effect for the TAK1 downstream factors ↑	
Baricitinib	A JAK 1 and 2 inhibitor	STAT1, JAK	Clinical: monocyte frequency and p-STAT1 in circulating monocytes served as potential early response markers to baricitinib treatment	Tucci et al. (2022); Weston et al. (2022); Taylor et al. (2023)
			Low-risk-related AESI	
			Low incidence with the dermatologic indications	
			Experimental: OSM-induced JAK signaling ↓	
			IL-6, MCP-1, IP-10 expression in the following stages \downarrow	
Upadacitinib	A selective JAK 1 inhibitor	JAK 1	Clinical: combination with MTX	Panchal et al. (2023); Taldaev et al.
			Maximum adverse events were reported at 12 mg twice daily	(2021)
Tofacitinib	A JAK/STAT inhibitor	STAT6/miR-425-5p/IGF1	Clinical: treatment of RA	Di Benedetto et al. (2021); Palmroth
			Beneficial for RA patients who don't respond to TNF-inhibitors or methotrexate	Liu et al. (2022); Vomero et al. (2022); Ruscitti et al. (2022)
			Modulate autophagy of FLS	
			Experimental: pro-inflammatory cytokines \downarrow collagen I and $\alpha\text{-SMA}$ of RA-FLS \downarrow	
Momelotinib	A competitive JAK1/JAK2 inhibitor	IL-6/JAK1/STAT3	Clinical: no indication for treatment of RA.	Srivastava et al. (2022)
			Experimental: proliferative, migratory of FLS↓	
			PRMT, survivin, HIF-1a \downarrow	
			JAK1 and STAT3 by IL-6/sIL-6R activation↓	
			SOCS3 ↑	

TABLE 3	(Continued)	Small	molecule	drugs	targeting	JAK/STAT	regulating	FLS.

Name	Source	Targets/signaling pathways	Estimate	References			
The natural pro	The natural products						
Matrine	Matrine An alkaloid from genus Sophora JAK/STAT; PI3	JAK/STAT; PI3K/Akt/mTOR;	Experimental: Bcl-2 ↓	Yang et al. (2017); Ao et al. (2022);			
		IGF-p/Smad; Wnt	Bax, caspase-3↑	Lin et al. (2022)			
			JAK2, STAT1, STAT3 phosphorylation \downarrow				
Vitexin	An apigenin flavone glycoside from passion flower, bamboo leaves, and pearl millet	JAK/STAT	Experimental: inflammatory enzyme markers ↓ iNOS ↓	Zhang et al. (2022)			
			JAK/STAT expressions ↓				
			SOCS↑				
Isobavachalcone	A chalcone from <i>Psoralea corylifolia</i> Linn.	PI3K/Akt, JAK/STAT	Experimental: proliferation, migration, and invasion and promoted apoptosis of MH7A cells ↓ p-PI3K, p-STAT3, p-JAK1 SOCS3, p- Akt ↓	Wang et al. (2022)			

1: suppress, downregulate, inhibit, block, prevent, reduce, decrease; ↑: promote, upregulate, active, increase. csDMARDs, conventional synthetic disease-modifying anti-rheumatic drugs; TNFSF11, TNF Superfamily Member 11; AESI, adverse events of special interest; OSM, oncostatin M; α-SMA, smooth muscle alpha-actin; SOCS, suppressor of cytokine signaling; TAK, TGF β-activated kinase.

3.3 Small molecule drugs targeting JAK/ STAT regulating FLS

JAK/STAT signaling has been instrumental in regulating immune and inflammatory responses. The JAK/STAT pathway can be segmented into three components: receptor-associated tyrosine kinase, JAK tyrosine kinase, and STAT transcription factor. The JAK kinase activates JAK upon receptor binding, leading to JAKmediated phosphorylation of STAT. Among the STAT family, STAT1 and STAT3 serve as the primary activators (Kim et al., 2011). The expression and activity of STAT1 are elevated in the initial synovial tissue of RA, while STAT3 facilitates the survival of synovial fibroblasts. Elevated STAT3 expression contributes to the inhibition of programmed cell death-induced anti-apoptotic molecule expression, blocks apoptosis in RA-FLS, and promotes RA synovial thickening (Yang et al., 2017). The JAK/STAT pathway is also involved in regulating the response of RA-FLS to proinflammatory cytokines and plays an essential role in the proinflammatory response and invasive behavior of FLS (Diller et al., 2019).

Inhibitors of JAKs could block the activation of STATs in RA-LS in the synthesis of various drugs and in the study of natural products. We included the synthetic small molecule compounds and natural products in the last 3 years in Table 3. Tofacitinib is a Food and Drug Administration (FDA)- and European Medicines Agency (EMA)-approved JAK inhibitor that effectively treats RA (Vomero et al., 2022). The synthetic small molecule compounds of peficitinib, fingolitinib, takinib, tolvamycin, baricitinib, and abatinib all demonstrated monotherapy effectiveness in clinical trials in RA. The synthetic JAK inhibitors appeared to be an important treatment choice for difficult-to-treat RA patients and researchers (Kubo et al., 2023). Momelotinib had no indication for the treatment of RA in the clinic, but could inhibit the proliferation and migration of FLS (Srivastava et al., 2022). On the contrary, there are few research reports on the natural products in the JAK/STAT signal pathway.

3.4 Small molecule drugs targeting PI3k/Akt regulating FLS

The PI3K/Akt signaling pathway is involved in regulating cell growth, proliferation, differentiation, and survival and is associated with the production of pro-inflammatory cytokines, degrading enzymes of the extracellular matrix, and other factors in FLS. The activation of PI3K induces the phosphorylation of Akt and p-Akt. As a downstream effector, it can be involved in FLS invasion by regulating the transcriptional levels of MMPs. The Akt phosphorylation also activates downstream mTOR complex 1 (mTORC1). mTORC1 translates mRNA into proteins to regulate the cell activities of metabolism, growth, and differentiation and is involved in RA-FLS proliferation and survival (Wendel et al., 2004; Malemud, 2013).

Table 4 is a summary of the synthetic small molecules and natural drugs that have been developed recently that target PI3k/Akt. Metformin, a drug used to treat type 2 diabetes, has been shown to have a protective effect against the development of RA (Liang et al., 2023), and RA-FLS proliferation is inhibited by metformin in a doseand time-dependent manner (Chen et al., 2019). The natural products targeted at PI3k/Akt regulating FLS came from a variety of sources. Against the development of inflammatory arthritis, ginger is a preventive substance. There was evidence that ginger helped reduce RA-related joint pain (Al-Nahain et al., 2014). The active ingredients of ginger, 6-shogaol, and 8-shogaol reduced the production of TNF- α , IL-1 β , IL-6, etc., prevented migration, invasion, and population growth, and ameliorated joint destruction in mice (N. Li et al., 2023; Jo et al., 2022).

3.5 Wnt signaling pathway and relevant drugs regulating FLS

The Wnt signaling cascade participates in regulating the growth, differentiation, production, and apoptosis of osteoblasts. The conventional Wnt/ β -catenin cascade, Wnt/Ca₂⁺ signaling cascade, and Wnt/JNK signaling cascade coordinate with each other to

TABLE 4 Small molecule drugs targeting PI3k/Akt regulating FLS.

Name	Source	Targets/signaling pathways	Estimate	References	
The synthetic small molecule compounds					
Metformin	The biguanide hypoglycemic agents	IGF-IR/PI3K/Akt/m-TOR	Clinical: preventing RA	Liang et al. (2023); Chen et al. (2019);	
			Inflammation, disease severity, and quality of life with high safety \uparrow	Gharib et al. (2021)	
			Experimental: G2/M cell cycle phase arrest ↓		
			mTOR phosphorylation \downarrow		
			Adjusting the p70s6k and 4EBP1 phosphorylation		
The natural products					
Baicalein	A flavone from Scutellaria baicalensis	PI3K/Akt/mTOR	Experimental: apoptotic proteins ↑	Zhang et al. (2022)	
			EMT-related proteins ↓		
			Cell apoptosis ↑		
			Cell migration phosphorylation \downarrow		
			The phosphorylation of PI3K, Akt, and mTOR dose dependently \downarrow		
Nobiletin	A polymethoxylated flavonoid from citrus peels	PI3K/Akt/HIF-1a	Experimental: enhanced the performance in synovial tissue combined with MTX	Liu et al. (2022)	
			P-gp expression \downarrow		
			Contribute to MTX resistance		
Artemisitene	A derivatives of artemisinin from	METTL3/ICAM2/PI3K/	Experimental: progression of FLS \downarrow	Chen et al. (2022)	
	Antemisia annua L	AK/ 0500	N6-methyladenosine modification of ICAM2 mRNA \downarrow		
Shikonin	A naphthoquinone pigment from the root of <i>Lithospermum erythrorhizon</i>	PI3K- Akt -mTOR, MAPK	Experimental: migration, adhesion, and invasion of MH7A cells \downarrow	Lian-Hua et al. (2020); Li et al. (2021)	
			The phosphorylation levels of Akt, JNK, p38, ERK \downarrow		
Cinnamaldehyde	An aldehyde from the bark of <i>Cinnamomum cassia</i>	PI3K/Akt	Experimental: proliferation and metastasis ↓	Li and Wang (2020)	
Daphnetin	A coumarin derivative from <i>Daphne</i>	PI3K/Akt/mTOR	Experimental: inflammatory response \downarrow	Deng et al. (2020)	
	04074		Cytokine expression ↓		
			IL-10 ↑		
6-Shogaol	An alkylphenol from ginger	PI3K/AKT/NF-κB	Experimental: proliferation, migration, and invasion of FLS and MH7A cells \downarrow	Li et al. (2023)	
			IL-1 β , IL-6, IL-8 \downarrow		
			MMP-2, MMP-9 ↓		
			PPAR-γ ↑		
8-Shogaol		TAK1, Akt, MAPK	Experimental: TAK1 activity selectively ↓	Jo et al. (2022)	
			IKK, Akt, MAPK \downarrow		
			Reversing pathologies of joint structure		

↓: suppress, downregulate, inhibit, block, prevent, reduce, decrease; ↑: promote, upregulate, active, increase. METTL3, methyltransferase-like 3.

regulate the dynamic balance between osteoclasts and osteoblasts. Once the balance is disturbed, it might lead to bone erosion and bone destruction (Walsh et al., 2009; De, 2011; Deal, 2012). Studies

had shown that the growth Wnt3a/5a proteins could activate the Wnt signaling cascade as well as downstream genes, thus increasing fibronectin expression and promoting FLS function. The

aforementioned processes also promoted the proliferation of RA synovial tissue without pro-inflammatory factors (Kim et al., 2010; Rabelo Fde et al., 2010; Maeda et al., 2013). Researchers (Cici et al., 2019) suggested that the inflammatory activation of the Wnt pathway might inhibit T-cell function and exacerbate the immune response [181]. In the recent 3 years, we inquired natural products, including paeoniflorin (Yang et al., 2022), 7-hydroxycoumarin (Umbelliferone) (Cai et al., 2022; Cai et al., 2022), and penta-acetyl geniposide (Cai et al., 2021).

4 Conclusion

In this review, we summarized as much as possible the involvement of FLS, covering the RA-FLS pathogenesis, synthetic small molecular compounds, and natural products targeting primary signaling pathways in the last 3 years. Natural products comprise a range of substances derived from diverse natural sources, such as plants, animals, and microorganism. These sources provided valuable resources for the design and development of drugs. From the results, the content of this paper could be continuously extended in the following aspects. 1) For the synthetic small molecule compounds, the popular targeting signaling pathways are still MAPK and NF-KB in the current research stage. We cannot ignore that JAK/STAT has great potential for research studies, due to the fact that several drugs have appeared in the clinic. Moreover, modulation of Wnt signaling might not only repair articular bone damage but also inhibit the production of proinflammatory cytokines, showing a new strategy for RA treatment (Miao et al., 2013; Liu et al., 2019). Typically, these signaling pathways interacted with each other. A small molecule could act through multiple pathways. 2) For the natural products, there was great potential. Researchers have tried to explore drugs targeted to activate FLS to treat RA using traditional human experience and herbs. For example, triptolide has been a hot area of research for several years. Most of the results are currently in the experimental stage, not the clinical trial stage. Fortunately, the source plants of these natural products have been used for RA in clinical studies. 3) The natural products derived from herbal medicine that can regulate RA-FLS abnormalities are mainly

References

Aihaiti, Y., Song Cai, Y., Tuerhong, X., Ni Yang, Y., Ma, Y., Shi Zheng, H., et al. (2021). Therapeutic effects of naringin in rheumatoid arthritis: Network pharmacology and experimental validation. *Front. Pharmacol.* 12, 672054. doi:10.3389/fphar.2021.672054

Al-Nahain, A., Jahan, R., and Rahmatullah, M. (2014). Zingiber officinale: A potential plant against rheumatoid arthritis. *Arthritis* 2014, 159089–159098. doi:10.1155/2014/159089

Almutairi, K., Nossent, J., Preen, D., Keen, H., and Inderjeeth, C. (2021). The global prevalence of rheumatoid arthritis: A meta-analysis based on a systematic review. *Rheumatol. Int.* 41 (5), 863–877. doi:10.1007/s00296-020-04731-0

An, L., Li, Z., Shi, L., Wang, L., Wang, Y., Jin, L., et al. (2020). Inflammation-targeted celastrol nanodrug attenuates collagen-induced arthritis through NF- κ B and Notch1 pathways. *Nano Lett.* 20 (10), 7728–7736. doi:10.1021/acs.nanolett.0c03279

Ao, L., Gao, H., Jia, L., Liu, S., Guo, J., Liu, B., et al. (2022). Matrine inhibits synovial angiogenesis in collagen-induced arthritis rats by regulating HIF-VEGF-Ang and inhibiting the PI3K/Akt signaling pathway. *Mol. Immunol.* 141, 13–20. doi:10.1016/j.molimm.2021.11.002

alkaloids, flavonoids, saponins, phenols, and quinones (Smolen et al., 2018). 4) In addition, we have found many reports on the mechanisms of herbal extract, Chinese herbal compound prescription, and traditional Chinese patent medicines in RA that were worthy of further research.

Author contributions

YT was responsible for writing and drawing by Figdraw. XL drafted the original framework and figures. QD collected and sorted materials. JS and YF provided guidance. LB reviewed writing and drawing. All authors contributed to the article and approved the submitted version.

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Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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Aupperle, K. R., Bennett, B. L., Boyle, D. L., Tak, P. P., Manning, A. M., and Firestein, G. S. (1999). NF-kappa B regulation by I kappa B kinase in primary fibroblast-like synoviocytes. J. Immunol. 163 (1), 427–433. doi:10.4049/jimmunol.163.1.427

Bai, J., Xie, N., Hou, Y., Chen, X., Hu, Y., Zhang, Y., et al. (2022). The enhanced mitochondrial dysfunction by cantleyoside confines inflammatory response and promotes apoptosis of human HFLS-RA cell line via AMPK/Sirt 1/NF-κB pathway activation. *Biomed. Pharmacother.* 149, 112847. doi:10.1016/j.biopha.2022.112847

Bartok, B., and Firestein, G. S. (2010). Fibroblast-like synoviocytes: Key effector cells in rheumatoid arthritis. *Immunol. Rev.* 233 (1), 233–255. doi:10.1111/j.0105-2896.2009.00859.x

Bombardieri, M., Kam, N. W., Brentano, F., Choi, K., Filer, A., Kyburz, D., et al. (2011). A BAFF/APRIL-dependent TLR3-stimulated pathway enhances the capacity of rheumatoid synovial fibroblasts to induce AID expression and Ig class-switching in B cells. *Ann. Rheum. Dis.* 70 (10), 1857–1865. doi:10.1136/ard.2011.150219

Bu, Y., Wu, H., Deng, R., and Wang, Y. (2022). The anti-angiogenesis mechanism of Geniposide on rheumatoid arthritis is related to the regulation of PTEN. *Inflammopharmacology* 30 (3), 1047–1062. doi:10.1007/s10787-022-00975-3

Buhrmann, C., Mobasheri, A., Matis, U., and Shakibaei, M. (2010). Curcumin mediated suppression of nuclear factor- κ B promotes chondrogenic differentiation of mesenchymal stem cells in a high-density co-culture microenvironment. *Arthritis Res. Ther.* 12 (4), R127. doi:10.1186/ar3065

Bustamante, M. F., Garcia-Carbonell, R., Whisenant, K. D., and Guma, M. (2017). Fibroblast-like synoviocyte metabolism in the pathogenesis of rheumatoid arthritis. *Arthritis Res. Ther.* 19 (1), 110. doi:10.1186/s13075-017-1303-3

Cai, L., Mu, Y. R., Liu, M. M., Zhou, M. Y., Meng, B., Liu, F. Y., et al. (2021). Pentaacetyl Geniposide suppresses migration, invasion, and inflammation of TNF- α stimulated rheumatoid arthritis fibroblast-like synoviocytes involving wnt/ β -catenin signaling pathway. *Inflammation* 44 (6), 2232–2245. doi:10.1007/s10753-021-01495-y

Cai, L., Zhou, M. Y., Hu, S., Liu, F. Y., Wang, M. Q., Wang, X. H., et al. (2022). Umbelliferone inhibits migration, invasion and inflammation of rheumatoid arthritis fibroblast-like synoviocytes and relieves adjuvant-induced arthritis in rats by blockade of wnt/ β -catenin signaling pathway. *Am. J. Chin. Med.* 50 (7), 1945–1962. doi:10.1142/ S0192415X22500835

Cai, L., Zong, P., Zhou, M. Y., Liu, F. Y., Meng, B., Liu, M. M., et al. (2022). 7-Hydroxycoumarin mitigates the severity of collagen-induced arthritis in rats by inhibiting proliferation and inducing apoptosis of fibroblast-like synoviocytes via suppression of Wnt/ β -catenin signaling pathway. *Phytomedicine* 94, 153841. doi:10. 1016/j.phymed.2021.153841

Cao, C., Zeng, L., and Rong, X. (2022). Therapeutic mechanism of emodin for treatment of rheumatoid arthritis: A network pharmacology-based analysis. *Nan Fang. Yi Ke Da Xue Xue Bao* 42 (6), 913–921. doi:10.12122/j.issn.1673-4254.2022.06.16

Cao, D., Fan, Q., Li, Z., Chen, M., Jiang, Y., Lin, R., et al. (2022). Transcriptomic profiling revealed the role of apigenin-4'-O- α -L-rhamnoside in inhibiting the activation of rheumatoid arthritis fibroblast-like synoviocytes via MAPK signaling pathway. *Phytomedicine* 102, 154201. doi:10.1016/j.phymed.2022.154201

Carter, A. B., Knudtson, K. L., Monick, M. M., and Hunninghake, G. W. (1999). The p38 mitogen-activated protein kinase is required for NF-kappaB-dependent gene expression. The role of TATA-binding protein (TBP). *J. Biol. Chem.* 274 (43), 30858–30863. doi:10.1074/jbc.274.43.30858

Chen, D. P., Wong, C. K., Leung, P. C., Fung, K. P., Lau, C. B., Lau, C. P., et al. (2011). Anti-inflammatory activities of Chinese herbal medicine sinomenine and Liang Miao San on tumor necrosis factor- α -activated human fibroblast-like synoviocytes in rheumatoid arthritis. *J. Ethnopharmacol.* 137 (1), 457–468. doi:10.1016/j.jep.2011. 05.048

Chen, H., Tao, L., Liang, J., Pan, C., and Wei, H. (2023). Ubiquitin D promotes the progression of rheumatoid arthritis via activation of the p38 MAPK pathway. *Mol. Med. Rep.* 27 (2), 53. doi:10.3892/mmr.2023.12940

Chen, J., Lin, X., He, J., Liu, D., He, L., Zhang, M., et al. (2022). Artemisitene suppresses rheumatoid arthritis progression via modulating METTL3-mediated N6-methyladenosine modification of ICAM2 mRNA in fibroblast-like synoviocytes. *Clin. Transl. Med.* 12 (12), e1148. doi:10.1002/ctm2.1148

Chen, K., Lin, Z. W., He, S. M., Wang, C. Q., Yang, J. C., Lu, Y., et al. (2019). Metformin inhibits the proliferation of rheumatoid arthritis fibroblast-like synoviocytes through IGF-IR/PI3K/AKT/m-TOR pathway. *Biomed. Pharmacother.* 115, 108875. doi:10.1016/j.biopha.2019.108875

Chen, X., Lin, H., Chen, J., Wu, L., Zhu, J., Ye, Y., et al. (2021). Paclitaxel inhibits synoviocyte migration and inflammatory mediator production in rheumatoid arthritis. *Front. Pharmacol.* 12, 714566. doi:10.3389/fphar.2021.714566

Chen, Y., Wang, Y., Liu, M., Zhou, B., and Yang, G. (2020). Diosmetin exhibits antiproliferative and anti-inflammatory effects on TNF- α -stimulated human rheumatoid arthritis fibroblast-like synoviocytes through regulating the Akt and NF- κ B signaling pathways. *Phytother. Res.* 34 (6), 1310–1319. doi:10.1002/ptr.6596

Cheng, Y. C., Zhang, X., Lin, S. C., Li, S., Chang, Y. K., Chen, H. H., et al. (2022). Echinocystic acid ameliorates arthritis in SKG mice by suppressing Th17 cell differentiation and human rheumatoid arthritis fibroblast-like synoviocytes inflammation. J. Agric. Food Chem. 70 (51), 16176–16187. doi:10.1021/acs.jafc.2c05802

Choe, J. Y., Lee, S. J., Park, S. H., and Kim, S. K. (2012). Tacrolimus (FK506) inhibits interleukin-1 β -induced angiopoietin-1, Tie-2 receptor, and vascular endothelial growth factor through down-regulation of JNK and p38 pathway in human rheumatoid fibroblast-like synoviocytes. *Jt. Bone Spine* 79 (2), 137–143. doi:10.1016/j.jbspin. 2011.03.018

Choi, Y., Lee, E. G., Jeong, J. H., and Yoo, W. H. (2021). 4-Phenylbutyric acid, a potent endoplasmic reticulum stress inhibitor, attenuates the severity of collagen-induced arthritis in mice via inhibition of proliferation and inflammatory responses of synovial fibroblasts. *Kaohsiung J. Med. Sci.* 37 (7), 604–615. doi:10.1002/kjm2.12376

Cici, D., Corrado, A., Rotondo, C., and Cantatore, F. P. (2019). Wnt signaling and biological therapy in rheumatoid arthritis and spondyloarthritis. *Int. J. Mol. Sci.* 20 (22), 5552. doi:10.3390/ijms20225552

Dai, Y., Sheng, J., He, S., Wu, Q., Wang, Y., and Su, L. (2022). Dehydroevodiamine suppresses inflammatory responses in adjuvant-induced arthritis rats and human fibroblast-like synoviocytes. *Bioengineered* 13 (1), 268–279. doi:10.1080/21655979.2021.1999554

Davis, J. M., 3rd, and Matteson, E. L.American College of Rheumatology; European League Against Rheumatism (2012). My treatment approach to rheumatoid arthritis. *Mayo Clin. Proc.* 87 (7), 659–673. doi:10.1016/j.mayocp.2012.03.011 De, A. (2011). Wnt/Ca2+ signaling pathway: A brief overview. Acta Biochim. Biophys. Sin. (Shanghai) 43 (10), 745–756. doi:10.1093/abbs/gmr079

Deal, C. (2012). Bone loss in rheumatoid arthritis: Systemic, periarticular, and focal. Curr. Rheumatol. Rep. 14 (3), 231–237. doi:10.1007/s11926-012-0253-7

Deng, H., Zheng, M., Hu, Z., Zeng, X., Kuang, N., and Fu, Y. (2020). Effects of daphnetin on the autophagy signaling pathway of fibroblast-like synoviocytes in rats with collagen-induced arthritis (CIA) induced by TNF-a. *Cytokine* 127, 154952. doi:10. 1016/j.cyto.2019.154952

Di Benedetto, P., Ruscitti, P., Berardicurti, O., Panzera, N., Grazia, N., Di Vito Nolfi, M., et al. (2021). Blocking Jak/STAT signalling using tofacitinib inhibits angiogenesis in experimental arthritis. *Arthritis Res. Ther.* 23 (1), 213. doi:10. 1186/s13075-021-02587-8

Diller, M., Hasseli, R., Hülser, M. L., Aykara, I., Frommer, K., Rehart, S., et al. (2019). Targeting activated synovial fibroblasts in rheumatoid arthritis by peficitinib. *Front. Immunol.* 10, 541. doi:10.3389/fimmu.2019.00541

Ding, Q., Hu, W., Wang, R., Yang, Q., Zhu, M., Li, M., et al. (2023). Signaling pathways in rheumatoid arthritis: Implications for targeted therapy. *Signal Transduct. Target Ther.* 8 (1), 68. doi:10.1038/s41392-023-01331-9

Doss, H. M., Ganesan, R., and Rasool, M. (2016). Trikatu, an herbal compound ameliorates rheumatoid arthritis by the suppression of inflammatory immune responses in rats with adjuvant-induced arthritis and on cultured fibroblast like synoviocytes via the inhibition of the NFkB signaling pathway. *Chem. Biol. Interact.* 258, 175–186. doi:10.1016/j.cbi.2016.09.003

Du, H., Wang, Y., Zeng, Y., Huang, X., Liu, D., Ye, L., et al. (2020). Tanshinone IIA suppresses proliferation and inflammatory cytokine production of synovial fibroblasts from rheumatoid arthritis patients induced by TNF- α and attenuates the inflammatory response in AIA mice. *Front. Pharmacol.* 11, 568. doi:10.3389/fphar.2020.00568

Du, H., Zhang, X., Zeng, Y., Huang, X., Chen, H., Wang, S., et al. (2019). A novel phytochemical, DIM, inhibits proliferation, migration, invasion and TNF- α induced inflammatory cytokine production of synovial fibroblasts from rheumatoid arthritis patients by targeting MAPK and AKT/mTOR signal pathway. *Front. Immunol.* 10, 1620. doi:10.3389/fimmu.2019.01620

Emori, T., Kasahara, M., Sugahara, S., Hashimoto, M., Ito, H., Narumiya, S., et al. (2020). Role of JAK-STAT signaling in the pathogenic behavior of fibroblast-like synoviocytes in rheumatoid arthritis: Effect of the novel JAK inhibitor peficitinib. *Eur. J. Pharmacol.* 882, 173238. doi:10.1016/j.ejphar.2020.173238

Fang, Z., He, D., Yu, B., Liu, F., Zuo, J., Li, Y., et al. (2017). High-throughput study of the effects of celastrol on activated fibroblast-like synoviocytes from patients with rheumatoid arthritis. *Genes (Basel)* 8 (9), 221. doi:10.3390/genes8090221

Feng, Y., Mei, L., Wang, M., Huang, Q., and Huang, R. (2021). Anti-inflammatory and pro-apoptotic effects of 18beta-glycyrrhetinic acid *in vitro* and *in vivo* models of rheumatoid arthritis. *Front. Pharmacol.* 12, 681525. doi:10.3389/fphar.2021.681525

Firestein, G. S. (2003). Evolving concepts of rheumatoid arthritis. *Nature* 423 (6937), 356-361. doi:10.1038/nature01661

Gan, K., Xu, L., Feng, X., Zhang, Q., Wang, F., Zhang, M., et al. (2015). Celastrol attenuates bone erosion in collagen-Induced arthritis mice and inhibits osteoclast differentiation and function in RANKL-induced RAW264.7. *Int. Immunopharmacol.* 24 (2), 239–246. doi:10.1016/j.intimp.2014.12.012

Gao, X., Kang, X., Lu, H., Xue, E., Chen, R., Pan, J., et al. (2022). Piceatannol suppresses inflammation and promotes apoptosis in rheumatoid arthritis-fibroblast-like synoviocytes by inhibiting the NF- κ B and MAPK signaling pathways. *Mol. Med. Rep.* 25 (5), 180. doi:10.3892/mmr.2022.12696

Gharib, M., Elbaz, W., Darweesh, E., Sabri, N. A., and Shawki, M. A. (2021). Efficacy and safety of Metformin use in rheumatoid arthritis: A randomized controlled study. *Front. Pharmacol.* 12, 726490. doi:10.3389/fphar.2021.726490

Guma, M., Hammaker, D., Topolewski, K., Corr, M., Boyle, D. L., Karin, M., et al. (2012). Antiinflammatory functions of p38 in mouse models of rheumatoid arthritis: Advantages of targeting upstream kinases MKK-3 or MKK-6. *Arthritis Rheum.* 64 (9), 2887–2895. doi:10.1002/art.34489

Guo, Q., Zhang, S., Huang, J., and Liu, K. (2020). Alogliptin inhibits IL-1β-induced inflammatory response in fibroblast-like synoviocytes. *Int. Immunopharmacol.* 83, 106372. doi:10.1016/j.intimp.2020.106372

Gutierrez-Urena, S. R., Amaya-Cabrera, E. L., Uribe-Martinez, J. F., Ventura-Valenzuela, M. E., Rosal-Arteaga, C., Martinez-Bonilla, G. E., et al. (2020). Peficitinib hydrobromide to treat rheumatoid arthritis. *Drugs Today (Barc)* 56 (8), 505–514. doi:10.1358/dot.2020.56.8.3123469

Hammaker, D., Topolewski, K., Edgar, M., Yoshizawa, T., Fukushima, A., Boyle, D. L., et al. (2012). Decreased collagen-induced arthritis severity and adaptive immunity in MKK-6-deficient mice. *Arthritis Rheum.* 64 (3), 678–687. doi:10.1002/art.33359

Harigai, M., Hara, M., Kawamoto, M., Kawaguchi, Y., Sugiura, T., Tanaka, M., et al. (2004). Amplification of the synovial inflammatory response through activation of mitogen-activated protein kinases and nuclear factor kappaB using ligation of CD40 on CD14+ synovial cells from patients with rheumatoid arthritis. *Arthritis Rheum.* 50 (7), 2167–2177. doi:10.1002/art.20340

Hu, X., Tang, J., Zeng, G., Hu, X., Bao, P., Wu, J., et al. (2019). RGS1 silencing inhibits the inflammatory response and angiogenesis in rheumatoid arthritis rats through the

inactivation of Toll-like receptor signaling pathway. J. Cell Physiol. 234 (11), 20432-20442. doi:10.1002/jcp.28645

Huang, M., Wu, K., Zeng, S., Liu, W., Cui, T., Chen, Z., et al. (2021). Punicalagin inhibited inflammation and migration of fibroblast-like synoviocytes through NF- κ B pathway in the experimental study of rheumatoid arthritis. *J. Inflamm. Res.* 14, 1901–1913. doi:10.2147/JIR.S302929

Huang, Q., Xiao, X., Yu, J., Yang, Y., Yu, J., Liu, Y., et al. (2022). Tectoridin exhibits anti-rheumatoid arthritis activity through the inhibition of the inflammatory response and the MAPK pathway *in vivo* and *in vitro*. *Arch. Biochem. Biophys.* 727, 109328. doi:10.1016/j.abb.2022.109328

Jay, G. D., Britt, D. E., and Cha, C. J. (2000). Lubricin is a product of megakaryocyte stimulating factor gene expression by human synovial fibroblasts. *J. Rheumatol.* 27 (3), 594–600.

Ji, W., and Xu, W. (2022). Orientin inhibits the progression of fibroblast-like synovial cells in rheumatoid arthritis by regulating MAPK-signaling pathway. *Allergol. Immunopathol. Madr.* 50 (6), 154–162. doi:10.15586/aei.v50i6.742

Ji, Y. R., Chen, Y., Chen, Y. N., Qiu, G. L., Wen, J. G., Zheng, Y., et al. (2020). Dexmedetomidine inhibits the invasion, migration, and inflammation of rheumatoid arthritis fibroblast-like synoviocytes by reducing the expression of NLRC5. *Int. Immunopharmacol.* 82, 106374. doi:10.1016/j.intimp.2020.106374

Jia, N., Ma, H., Zhang, T., Wang, L., Cui, J., Zha, Y., et al. (2022). Gentiopicroside attenuates collagen-induced arthritis in mice via modulating the CD147/p38/NF-κB pathway. *Int. Immunopharmacol.* 108, 108854. doi:10.1016/j.intimp.2022.108854

Jo, S., Samarpita, S., Lee, J. S., Lee, Y. J., Son, J. E., Jeong, M., et al. (2022). 8-Shogaol inhibits rheumatoid arthritis through targeting TAK1. *Pharmacol. Res.* 178, 106176. doi:10.1016/j.phrs.2022.106176

Kadkhoda, Z., Amirzargar, A., Esmaili, Z., Vojdanian, M., and Akbari, S. (2016). Effect of TNF- α blockade in gingival crevicular fluid on periodontal condition of patients with rheumatoid arthritis. *Iran. J. Immunol.* 13 (3), 197–203.

Kaminska, B. (2005). MAPK signalling pathways as molecular targets for antiinflammatory therapy--from molecular mechanisms to therapeutic benefits. *Biochim. Biophys. Acta* 1754 (1-2), 253–262. doi:10.1016/j.bbapap.2005.08.017

Kaneko, Y., Kawahito, Y., Kojima, M., Nakayama, T., Hirata, S., Kishimoto, M., et al. (2021). Efficacy and safety of tacrolimus in patients with rheumatoid arthritis - a systematic review and meta-analysis. *Mod. Rheumatol.* 31 (1), 61–69. doi:10.1080/14397595.2020.1719607

Kim, J., Kim, J., Kim, D. W., Ha, Y., Ihm, M. H., Kim, H., et al. (2010). Wnt5a induces endothelial inflammation via beta-catenin-independent signaling. *J. Immunol.* 185 (2), 1274–1282. doi:10.4049/jimmunol.1000181

Kim, M., Sur, B., Villa, T., Nah, S. Y., and Oh, S. (2021). Inhibitory activity of gintonin on inflammation in human IL-1β-stimulated fibroblast-like synoviocytes and collageninduced arthritis in mice. J. Ginseng Res. 45 (4), 510–518. doi:10.1016/j.jgr.2020.12.001

Kim, M., Sur, B., Villa, T., Yun, J., Nah, S. Y., and Oh, S. (2021). Gintonin regulates inflammation in human IL-1β-stimulated fibroblast-like synoviocytes and carrageenan/ kaolin-induced arthritis in rats through LPAR2. *J. Ginseng Res.* 45 (5), 575–582. doi:10.1016/j.jgr.2021.02.001

Kim, S. K., Park, K. Y., Yoon, W. C., Park, S. H., Park, K. K., Yoo, D. H., et al. (2011). Melittin enhances apoptosis through suppression of IL-6/sIL-6R complex-induced NF- κ B and STAT3 activation and Bcl-2 expression for human fibroblast-like synovicytes in rheumatoid arthritis. *Jt. Bone Spine* 78 (5), 471-477. doi:10.1016/j.jbspin.2011.01.004

Kitanaga, Y., Imamura, E., Nakahara, Y., Fukahori, H., Fujii, Y., Kubo, S., et al. (2020). *In vitro* pharmacological effects of peficitinib on lymphocyte activation: A potential treatment for systemic sclerosis with JAK inhibitors. *Rheumatol. Oxf.* 59 (8), 1957–1968. doi:10.1093/rheumatology/kez526

Kubo, S., Nakayamada, S., and Tanaka, Y. (2023). JAK inhibitors for rheumatoid arthritis. *Expert Opin. Investig. Drugs* 32 (4), 333-344. doi:10.1080/13543784.2023. 2199919

Kwon, Y. J., Yoon, C. H., Lee, S. W., Park, Y. B., Lee, S. K., and Park, M. C. (2014). Inhibition of glycogen synthase kinase- 3β suppresses inflammatory responses in rheumatoid arthritis fibroblast-like synoviocytes and collagen-induced arthritis. *Jt. Bone Spine* 81 (3), 240–246. doi:10.1016/j.jbspin.2013.09.006

Lampropoulos, C. E., Orfanos, P., Bournia, V. K., Karatsourakis, T., Mavragani, C., Pikazis, D., et al. (2015). Adverse events and infections in patients with rheumatoid arthritis treated with conventional drugs or biologic agents: A real world study. *Clin. Exp. Rheumatol.* 33 (2), 216–224.

Leah, E. (2011). Crosstalk in RA synovia-TLR3-BAFF axis sustains B-cell activation. *Nat. Rev. Rheumatol.* 7 (10), 559. doi:10.1038/nrrheum.2011.122

Li, F., Dai, M., Wu, H., Deng, R., Fu, J., Zhang, Z., et al. (2018). Immunosuppressive effect of Geniposide on mitogen-activated protein kinase signalling pathway and their cross-talk in fibroblast-like synoviocytes of adjuvant arthritis rats. *Molecules* 23 (1), 91. doi:10.3390/molecules23010091

Li, J., Pang, J., Liu, Z., Ge, X., Zhen, Y., Jiang, C. C., et al. (2021). Shikonin induces programmed death of fibroblast synovial cells in rheumatoid arthritis by inhibiting energy pathways. *Sci. Rep.* 11 (1), 18263. doi:10.1038/s41598-021-97713-6

Li, N., Li, X., Deng, L., Yang, H., Gong, Z., Wang, Q., et al. (2023). 6-Shogaol inhibits the proliferation, apoptosis, and migration of rheumatoid arthritis fibroblast-like synoviocytes via the PI3K/AKT/NF- κ B pathway. *Phytomedicine* 109, 154562. doi:10. 1016/j.phymed.2022.154562

Li, X., He, L., Hu, Y., Duan, H., Li, X., Tan, S., et al. (2013). Sinomenine suppresses osteoclast formation and *Mycobacterium tuberculosis* H37Ra-induced bone loss by modulating RANKL signaling pathways. *PLoS One* 8 (9), e74274. doi:10.1371/journal. pone.0074274

Li, X., and Wang, Y. (2020). Cinnamaldehyde attenuates the progression of rheumatoid arthritis through down-regulation of PI3K/AKT signaling pathway. *Inflammation* 43 (5), 1729–1741. doi:10.1007/s10753-020-01246-5

Li, Z., Chen, M., Wang, Z., Fan, Q., Lin, Z., Tao, X., et al. (2023). Berberine inhibits RA-FLS cell proliferation and adhesion by regulating RAS/MAPK/FOXO/HIF-1 signal pathway in the treatment of rheumatoid arthritis. *Bone Jt. Res.* 12 (2), 91–102. doi:10. 1302/2046-3758.122.BJR-2022-0269.R1

Lian-Hua, H. E., Qian-Qian, W., Cong-Cong, S., Na, L., and Chun-Fang, L. (2020). Effect of shikonin on function of rheumatoid arthritis fibroblast like synoviocytes. *Zhongguo Zhong Yao Za Zhi* 45 (19), 4712–4718. doi:10.19540/j.cnki.cjcmm. 20200506.401

Liang, J., Cai, Y., Zhang, J., Jing, Z., Lv, L., Zhang, G., et al. (2023). Metformin treatment reduces the incidence of rheumatoid arthritis: A two-sample mendelian randomized study. *J. Clin. Med.* 12 (7), 2461. doi:10.3390/jcm12072461

Lin, W., Chen, G., Mao, Y., Ma, X., Zhou, J., Yu, X., et al. (2022). Imperatorin inhibits proliferation, migration, and inflammation *via* blocking the NF- κ B and MAPK pathways in rheumatoid fibroblast-like synoviocytes. *ACS Omega* 7 (34), 29868–29876. doi:10.1021/acsomega.2c02766

Lin, X., Chen, J., Tao, C., Luo, L., He, J., and Wang, Q. (2023). Osthole regulates N6methyladenosine-modified TGM2 to inhibit the progression of rheumatoid arthritis and associated interstitial lung disease. *MedComm* 4(2), e219. doi:10.1002/mco2.219

Lin, Y., He, F., Wu, L., Xu, Y., and Du, Q. (2022). Matrine exerts pharmacological effects through multiple signaling pathways: A comprehensive review. *Drug Des. Devel Ther.* 16, 533–569. doi:10.2147/DDDT.S349678

Liu, J., Zhao, N., Su, S. H., Gao, Y., and Qi, B. (2023). Anti-arthritic effect of edaravone against complete freund adjuvant induced arthritis via osteoclast differentiation and HIF-1α-VEGF-ANG-1 Axis. *Drug Des. Devel Ther.* 17, 519–534. doi:10.2147/DDDT. S391606

Liu, R., Song, Y., Li, C., Zhang, Z., Xue, Z., Huang, Q., et al. (2022). The naturally occurring flavonoid nobiletin reverses methotrexate resistance via inhibition of P-glycoprotein synthesis. J. Biol. Chem. 298 (4), 101756. doi:10.1016/j.jbc.2022.101756

Liu, T., Zhang, L., Joo, D., and Sun, S. C. (2017). NF-κB signaling in inflammation. Signal Transduct. Target Ther. 2, 17023. doi:10.1038/sigtrans.2017.23

Liu, X. G., Zhang, Y., Ju, W. F., Li, C. Y., and Mu, Y. C. (2019). MiR-21 relieves rheumatoid arthritis in rats via targeting Wnt signaling pathway. *Eur. Rev. Med. Pharmacol. Sci.* 23 (3), 96–103. doi:10.26355/eurrev_201908_18635

Liu, Y., Peng, J., Xiong, X., Cheng, L., and Cheng, X. (2022). Tofacitinib enhances IGF1 via inhibiting STAT6 transcriptionally activated-miR-425-5p to ameliorate inflammation in RA-FLS. *Mol. Cell Biochem.* 477 (10), 2335–2344. doi:10.1007/s11010-022-04444-x

Luczkiewicz, P., Kokotkiewicz, A., Dampc, A., and Luczkiewicz, M. (2014). Mangiferin: A promising therapeutic agent for rheumatoid arthritis treatment. *Med. Hypotheses* 83 (5), 570-574. doi:10.1016/j.mehy.2014.08.021

Lv, M., Liang, Q., Luo, Z., Han, B., Ni, T., Wang, Y., et al. (2022). UPLC-LTQ-Orbitrap-Based cell metabolomics and network pharmacology analysis to reveal the potential antiarthritic effects of pristimerin: *In vitro*, in silico and *in vivo* study. *Metabolites* 12 (9), 839. doi:10.3390/metabo12090839

Lv, Q., Zhu, X. Y., Xia, Y. F., Dai, Y., and Wei, Z. F. (2015). Tetrandrine inhibits migration and invasion of rheumatoid arthritis fibroblast-like synoviocytes through down-regulating the expressions of Rac1, Cdc42, and RhoA GTPases and activation of the PI3K/Akt and JNK signaling pathways. *Chin. J. Nat. Med.* 13 (11), 831–841. doi:10.1016/S1875-5364(15)30087-X

Maeda, K., Takahashi, N., and Kobayashi, Y. (2013). Roles of Wnt signals in bone resorption during physiological and pathological states. *J. Mol. Med. Berl.* 91 (1), 15–23. doi:10.1007/s00109-012-0974-0

Mahmoud, D. E., Kaabachi, W., Sassi, N., Tarhouni, L., Rekik, S., Jemmali, S., et al. (2022). The synovial fluid fibroblast-like synoviocyte: A long-neglected piece in the puzzle of rheumatoid arthritis pathogenesis. *Front. Immunol.* 13, 942417. doi:10.3389/ fimmu.2022.942417

Malemud, C. J. (2013). Intracellular signaling pathways in rheumatoid arthritis. J. Clin. Cell Immunol. 4, 160. doi:10.4172/2155-9899.1000160

Mardani, M., Mohammadshahi, J., Abolghasemi, S., and Teimourpour, R. (2023). Drug-induced liver injury due to tofacitinib: A case report. *J. Med. Case Rep.* 17 (1), 97. doi:10.1186/s13256-023-03821-4

Mavers, M., Ruderman, E. M., and Perlman, H. (2009). Intracellular signal pathways: Potential for therapies. *Curr. Rheumatol. Rep.* 11 (5), 378–385. doi:10.1007/s11926-009-0054-9 Meng, M., Yue, Z., Chang, L., Liu, Y., Hu, J., Song, Z., et al. (2021). Anti-rheumatoid arthritic effects of Paris saponin VII in human rheumatoid arthritis fibroblast-like synoviocytes and adjuvant-induced arthritis in rats. *Front. Pharmacol.* 12, 683698. doi:10.3389/fphar.2021.683698

Meng, Y., Ji, J., Xiao, X., Li, M., Niu, S., He, Y., et al. (2021). Ononin induces cell apoptosis and reduces inflammation in rheumatoid arthritis fibroblast-like synoviocytes by alleviating MAPK and NF-κB signaling pathways. *Acta Biochim. Pol.* 68 (2), 239–245. doi:10.18388/abp.2020_5528

Miao, C. G., Yang, Y. Y., He, X., Li, X. F., Huang, C., Huang, Y., et al. (2013). Wnt signaling pathway in rheumatoid arthritis, with special emphasis on the different roles in synovial inflammation and bone remodeling. *Cell Signal* 25 (10), 2069–2078. doi:10.1016/j.cellsig.2013.04.002

Min, H. K., Kim, S. H., Won, J. Y., Kim, K. W., Lee, J. Y., Lee, S. H., et al. (2023). Dasatinib, a selective tyrosine kinase inhibitor, prevents joint destruction in rheumatoid arthritis animal model. *Int. J. Rheum. Dis.* 26 (4), 718–726. doi:10.1111/1756-185X. 14627

Mohammadian Haftcheshmeh, S., Khosrojerdi, A., Aliabadi, A., Lotfi, S., Mohammadi, A., and Momtazi-Borojeni, A. A. (2021). Immunomodulatory effects of curcumin in rheumatoid arthritis: Evidence from molecular mechanisms to clinical outcomes. *Rev. Physiol. Biochem. Pharmacol.* 179, 1–29. doi:10.1007/112_2020_54

Mori, M., Hashimoto, M., Matsuo, T., Fujii, T., Furu, M., Ito, H., et al. (2017). Cellcontact-dependent activation of CD4(+) T cells by adhesion molecules on synovial fibroblasts. *Mod. Rheumatol.* 27 (3), 448–456. doi:10.1080/14397595.2016.1220353

Müller-Ladner, U., Ospelt, C., Gay, S., Distler, O., and Pap, T. (2007). Cells of the synovium in rheumatoid arthritis. Synovial fibroblasts. *Arthritis Res. Ther.* 9 (6), 223. doi:10.1186/ar2337

Neumann, E., Lefèvre, S., Zimmermann, B., Gay, S., and Müller-Ladner, U. (2010). Rheumatoid arthritis progression mediated by activated synovial fibroblasts. *Trends Mol. Med.* 16 (10), 458–468. doi:10.1016/j.molmed.2010.07.004

Niu, X., Song, H., Xiao, X., Yang, Y., Huang, Q., Yu, J., et al. (2022). Tectoridin ameliorates proliferation and inflammation in TNF- α -induced HFLS-RA cells via suppressing the TLR4/NLRP3/NF- κ B signaling pathway. *Tissue Cell* 77, 101826. doi:10.1016/j.tice.2022.101826

Nygaard, G., and Firestein, G. S. (2020). Restoring synovial homeostasis in rheumatoid arthritis by targeting fibroblast-like synoviocytes. *Nat. Rev. Rheumatol.* 16 (6), 316–333. doi:10.1038/s41584-020-0413-5

O'Shea, J. J., Laurence, A., and McInnes, I. B. (2013). Back to the future: Oral targeted therapy for RA and other autoimmune diseases. *Nat. Rev. Rheumatol.* 9 (3), 173–182. doi:10.1038/nrrheum.2013.7

Palmroth, M., Kuuliala, K., Peltomaa, R., Virtanen, A., Kuuliala, A., Kurttila, A., et al. (2021). Tofacitinib suppresses several JAK-STAT pathways in rheumatoid arthritis *in vivo* and baseline signaling profile associates with treatment response. *Front. Immunol.* 12, 738481. doi:10.3389/fimmu.2021.738481

Pan, D., Li, N., Liu, Y., Xu, Q., Liu, Q., You, Y., et al. (2018). Kaempferol inhibits the migration and invasion of rheumatoid arthritis fibroblast-like synoviocytes by blocking activation of the MAPK pathway. *Int. Immunopharmacol.* 55, 174–182. doi:10.1016/j. intimp.2017.12.011

Pan, F., Zhu, L., Lv, H., and Pei, C. (2016). Quercetin promotes the apoptosis of fibroblast-like synoviocytes in rheumatoid arthritis by upregulating lncRNA MALAT1. *Int. J. Mol. Med.* 38 (5), 1507–1514. doi:10.3892/ijmm.2016.2755

Panchal, V., Vyas, B. H., Sivasubramanian, B. P., Panchal, K., and Patel, H. (2023). A meta-analysis evaluating the effectiveness and safety of upadacitinib in treating rheumatoid arthritis in patients with inadequate response to disease-modifying anti-rheumatic drugs. *Cureus* 15 (1), e34384. doi:10.7759/cureus.34384

Panipinto, P. M., Singh, A. K., Shaikh, F. S., Siegel, R. J., Chourasia, M., and Ahmed, S. (2021). Takinib inhibits inflammation in human rheumatoid arthritis synovial fibroblasts by targeting the janus kinase-signal transducer and activator of transcription 3 (JAK/STAT3) pathway. *Int. J. Mol. Sci.* 22 (22), 12580. doi:10.3390/ ijms222212580

Park, J. K., Jang, Y. J., Oh, B. R., Shin, J., Bae, D., Ha, N., et al. (2020). Therapeutic potential of CKD-506, a novel selective histone deacetylase 6 inhibitor, in a murine model of rheumatoid arthritis. *Arthritis Res. Ther.* 22 (1), 176. doi:10.1186/s13075-020-02258-0

Qin, Y., Cai, M. L., Jin, H. Z., Huang, W., Zhu, C., Bozec, A., et al. (2022). Ageassociated B cells contribute to the pathogenesis of rheumatoid arthritis by inducing activation of fibroblast-like synoviocytes via TNF- α -mediated ERK1/2 and JAK-STAT1 pathways. *Ann. Rheum. Dis.* 81 (11), 1504–1514. doi:10.1136/ard-2022-222605

Rabelo Fde, S., da Mota, L. M., Lima, R. A., Lima, F. A., Barra, G. B., de Carvalho, J. F., et al. (2010). The Wnt signaling pathway and rheumatoid arthritis. *Autoimmun. Rev.* 9 (4), 207–210. doi:10.1016/j.autrev.2009.08.003

Ruscitti, P., Liakouli, V., Panzera, N., Angelucci, A., Berardicurti, O., Di Nino, E., et al. (2022). Tofacitinib may inhibit myofibroblast differentiation from rheumatoid-fibroblast-like synoviocytes induced by TGF-beta and IL-6. *Pharm. (Basel)* 15 (5), 622. doi:10.3390/ph15050622

Samarpita, S., Ganesan, R., and Rasool, M. (2020). Cyanidin prevents the hyperproliferative potential of fibroblast-like synoviocytes and disease progression

via targeting IL-17A cytokine signalling in rheumatoid arthritis. Toxicol. Appl. Pharmacol. 391, 114917. doi:10.1016/j.taap.2020.114917

Samarpita, S., Kim, J. Y., Rasool, M. K., and Kim, K. S. (2020). Investigation of toll-like receptor (TLR) 4 inhibitor TAK-242 as a new potential anti-rheumatoid arthritis drug. *Arthritis Res. Ther.* 22 (1), 16. doi:10.1186/s13075-020-2097-2

Samarpita, S., and Rasool, M. (2021). Cyanidin attenuates IL-17A cytokine signaling mediated monocyte migration and differentiation into mature osteoclasts in rheumatoid arthritis. *Cytokine* 142, 155502. doi:10.1016/j.cyto.2021.155502

Saravanan, S., Islam, V. I., Babu, N. P., Pandikumar, P., Thirugnanasambantham, K., Chellappandian, M., et al. (2014). Swertiamarin attenuates inflammation mediators via modulating NF- κ B/I κ B and JAK2/STAT3 transcription factors in adjuvant induced arthritis. *Eur. J. Pharm. Sci.* 56, 70–86. doi:10.1016/j.ejps.2014.02.005

Shang, W., Zhao, L. J., Dong, X. L., Zhao, Z. M., Li, J., Zhang, B. B., et al. (2016). Curcumin inhibits osteoclastogenic potential in PBMCs from rheumatoid arthritis patients via the suppression of MAPK/RANK/c-Fos/NFATc1 signaling pathways. *Mol. Med. Rep.* 14 (4), 3620–3626. doi:10.3892/mmr.2016.5674

Shao, N., Feng, Z., and Li, N. (2022). Isoginkgetin inhibits inflammatory response in the fibroblast-like synoviocytes of rheumatoid arthritis by suppressing matrix metallopeptidase 9 expression. *Chem. Biol. Drug Des.* 99 (6), 923–929. doi:10.1111/ cbdd.14049

Shen, P., Jiao, Y., Miao, L., Chen, J. H., and Momtazi-Borojeni, A. A. (2020). Immunomodulatory effects of berberine on the inflamed joint reveal new therapeutic targets for rheumatoid arthritis management. J. Cell Mol. Med. 24 (21), 12234–12245. doi:10.1111/jcmm.15803

Shen, Y., Fan, X., Qu, Y., Tang, M., Huang, Y., Peng, Y., et al. (2022). Magnoflorine attenuates inflammatory responses in RA by regulating the PI3K/Akt/NF- κ B and Keap1-Nrf2/HO-1 signalling pathways *in vivo* and *in vitro*. *Phytomedicine* 104, 154339. doi:10.1016/j.phymed.2022.154339

Shen, Y., Teng, L., Qu, Y., Liu, J., Zhu, X., Chen, S., et al. (2022). Anti-proliferation and anti-inflammation effects of corilagin in rheumatoid arthritis by downregulating NF- κ B and MAPK signaling pathways. *J. Ethnopharmacol.* 284, 114791. doi:10.1016/j.jep.2021. 114791

Sheng, S., Wang, X., Liu, X., Hu, X., Shao, Y., Wang, G., et al. (2022). The role of resveratrol on rheumatoid arthritis: From bench to bedside. *Front. Pharmacol.* 13, 829677. doi:10.3389/fphar.2022.829677

Shimizu, T., Kawashiri, S. Y., Morimoto, S., Kawazoe, Y., Kuroda, S., Kawasaki, R., et al. (2023). Efficacy and safety of selective JAK 1 inhibitor filgotinib in active rheumatoid arthritis patients with inadequate response to methotrexate: Comparative study with filgotinib and tocilizumab examined by clinical index as well as musculoskeletal ultrasound assessment (TRANSFORM study): Study protocol for a randomized, open-label, parallel-group, multicenter, and non-inferiority clinical trial. *Trials* 24 (1), 161. doi:10.1186/s13063-023-07176-5

Shu, C., Chen, J., Lv, M., Xi, Y., Zheng, J., and Xu, X. (2022). Plumbagin relieves rheumatoid arthritis through nuclear factor kappa-B (NF- κ B) pathway. *Bioengineered* 13 (5), 13632–13642. doi:10.1080/21655979.2022.2081756

Smolen, J. S., Aletaha, D., Barton, A., Burmester, G. R., Emery, P., Firestein, G. S., et al. (2018). Rheumatoid arthritis. *Nat. Rev. Dis. Prim.* 4, 18001. doi:10.1038/nrdp.2018.1

Song, X., Zhang, Y., Dai, E., Wang, L., and Du, H. (2020). Prediction of triptolide targets in rheumatoid arthritis using network pharmacology and molecular docking. *Int. Immunopharmacol.* 80, 106179. doi:10.1016/j.intimp.2019.106179

Srivastava, S., Samarpita, S., Ganesan, R., and Rasool, M. (2022). CYT387 inhibits the hyperproliferative potential of fibroblast-like synoviocytes via modulation of IL-6/ JAK1/STAT3 signaling in rheumatoid arthritis. *Immunol. Invest.* 51 (6), 1582–1597. doi:10.1080/08820139.2021.1994589

Sugiura, T., Kamino, H., Nariai, Y., Murakawa, Y., Kondo, M., Kawakami, M., et al. (2020). Screening of a panel of low molecular weight compounds that inhibit synovial fibroblast invasion in rheumatoid arthritis. *J. Immunol.* 205 (12), 3277–3290. doi:10. 4049/jimmunol.1901429

Sujitha, S., Dinesh, P., and Rasool, M. (2020). Berberine encapsulated PEG-coated liposomes attenuate Wnt1/ β -catenin signaling in rheumatoid arthritis via miR-23a activation. *Eur. J. Pharm. Biopharm.* 149, 170–191. doi:10.1016/j.ejpb.2020.02.007

Sun, Y., and Li, L. (2018). Cyanidin-3-glucoside inhibits inflammatory activities in human fibroblast-like synoviocytes and in mice with collagen-induced arthritis. *Clin. Exp. Pharmacol. Physiol.* 45 (10), 1038–1045. doi:10.1111/1440-1681.12970

Sur, B., Kim, M., Villa, T., and Oh, S. (2020). Benzylideneacetophenone derivative alleviates arthritic symptoms via modulation of the MAPK signaling pathway. *Molecules* 25 (15), 3319. doi:10.3390/molecules25153319

Taldaev, A., Rudnev, V. R., Nikolsky, K. S., Kulikova, L. I., and Kaysheva, A. L. (2021). Molecular modeling insights into upadacitinib selectivity upon binding to JAK protein family. *Pharm. (Basel)* 15 (1), 30. doi:10.3390/ph15010030

Tang, M., Zhu, W. J., Yang, Z. C., and He, C. S. (2019). Brucine inhibits TNF-ainduced HFLS-RA cell proliferation by activating the JNK signaling pathway. *Exp. Ther. Med.* 18 (1), 735–740. doi:10.3892/etm.2019.7582

Tang, Y., Liu, Q., Feng, Y., Zhang, Y., Xu, Z., Wen, C., et al. (2020). Tripterygium ingredients for pathogenicity cells in rheumatoid arthritis. *Front. Pharmacol.* 11, 583171. doi:10.3389/fphar.2020.583171

Taylor, P. C., Bieber, T., Alten, R., Witte, T., Galloway, J., Deberdt, W., et al. (2023). Baricitinib safety for events of special interest in populations at risk: Analysis from randomised trial data across rheumatologic and dermatologic indications. *Adv. Ther.* 40 (4), 1867–1883. doi:10.1007/s12325-023-02445-w

Terabe, K., Takahashi, N., Asai, S., Hirano, Y., Kanayama, Y., Yabe, Y., et al. (2023). Effectiveness of tacrolimus concomitant with biological disease-modifying antirheumatic drugs in patients with rheumatoid arthritis. *Mod. Rheumatol.* 33 (2), 292–301. doi:10.1093/mr/roac025

Tran, C. N., Davis, M. J., Tesmer, L. A., Endres, J. L., Motyl, C. D., Smuda, C., et al. (2007). Presentation of arthritogenic peptide to antigen-specific T cells by fibroblast-like synoviocytes. *Arthritis Rheum.* 56 (5), 1497–1506. doi:10.1002/art.22573

Tran, C. N., Thacker, S. G., Louie, D. M., Oliver, J., White, P. T., Endres, J. L., et al. (2008). Interactions of T cells with fibroblast-like synoviocytes: Role of the B7 family costimulatory ligand B7-H3. *J. Immunol.* 180 (5), 2989–2998. doi:10.4049/jimmunol. 180.5.2989

Tu, J., Huang, W., Zhang, W., Mei, J., and Zhu, C. (2022). Two main cellular components in rheumatoid arthritis: Communication between T cells and fibroblast-like synoviocytes in the joint synovium. *Front. Immunol.* 13, 922111. doi:10.3389/fimmu.2022.922111

Tucci, G., Garufi, C., Pacella, I., Zagaglioni, M., Pinzon Grimaldos, A., Ceccarelli, F., et al. (2022). Baricitinib therapy response in rheumatoid arthritis patients associates to STAT1 phosphorylation in monocytes. *Front. Immunol.* 13, 932240. doi:10.3389/fimmu.2022.932240

Villa, T., Kim, M., and Oh, S. (2020). Fangchinoline has an anti-arthritic effect in two animal models and in IL-1 β -stimulated human FLS cells. *Biomol. Ther. Seoul.* 28 (5), 414–422. doi:10.4062/biomolther.2020.113

Vomero, M., Caliste, M., Barbati, C., Speziali, M., Celia, A. I., Ucci, F., et al. (2022). Tofacitinib decreases autophagy of fibroblast-like synoviocytes from rheumatoid arthritis patients. *Front. Pharmacol.* 13, 852802. doi:10.3389/fphar.2022.852802

Walsh, N. C., Reinwald, S., Manning, C. A., Condon, K. W., Iwata, K., Burr, D. B., et al. (2009). Osteoblast function is compromised at sites of focal bone erosion in inflammatory arthritis. *J. Bone Min. Res.* 24 (9), 1572–1585. doi:10.1359/jbmr.090320

Wang, C. H., Yao, H., Chen, L. N., Jia, J. F., Wang, L., Dai, J. Y., et al. (2012). CD147 induces angiogenesis through a vascular endothelial growth factor and hypoxiainducible transcription factor 1α-mediated pathway in rheumatoid arthritis. *Arthritis Rheum.* 64 (6), 1818–1827. doi:10.1002/art.34341

Wang, D. D., Jiang, M. Y., Wang, W., Zhou, W. J., Zhang, Y. W., Yang, M., et al. (2020). Paeoniflorin-6'-O-benzene sulfonate down-regulates CXCR4-G $\beta\gamma$ -PI3K/AKT mediated migration in fibroblast-like synoviocytes of rheumatoid arthritis by inhibiting GRK2 translocation. *Biochem. Biophys. Res. Commun.* 526 (3), 805–812. doi:10.1016/j. bbrc.2020.03.164

Wang, G., Xie, X., Yuan, L., Qiu, J., Duan, W., Xu, B., et al. (2020). Resveratrol ameliorates rheumatoid arthritis via activation of SIRT1-Nrf2 signaling pathway. *Biofactors* 46 (3), 441-453. doi:10.1002/biof.1599

Wang, H., Mei, D., Liang, F. Q., Xue, Z. Y., Wang, P., Liu, R. J., et al. (2023). BAFF promotes FLS activation through BAFFR-mediated non-canonical NF-kB pathway and the effects of CP-25. *Inflammation* 46, 861–875. doi:10.1007/s10753-022-01774-2

Wang, H., Tu, S., Yang, S., Shen, P., Huang, Y., Ba, X., et al. (2019). Berberine modulates LPA function to inhibit the proliferation and inflammation of FLS-RA via p38/ERK MAPK pathway mediated by LPA(1). *Evid. Based Complement. Altern. Med.* 2019, 2580207. doi:10.1155/2019/2580207

Wang, J., Lian, J., Kong, X., and Lin, N. (2010). Effects of triptolide on cell proliferation and regulation of Ras-MAPKs pathway in synoviocytes induced by tumor necrosis factor. *Zhongguo Zhong Yao Za Zhi* 35 (7), 888–891. doi:10.4268/ cicmm20100719

Wang, M., Dai, T., Li, S., and Wang, W. (2022). Eugenol suppresses the proliferation and invasion of TNF-a-induced fibroblast-like synoviocytes via regulating NF-kB and COX-2. *Biochem. Biophys. Res. Commun.* 612, 63–69. doi:10.1016/j.bbrc.2022.04.074

Wang, R., Liu, J., Wang, Z., Wu, X., Guo, H., Jiao, X., et al. (2021). Mangiferin exert protective effects on joints of adjuvant-induced arthritis rats by regulating the MAPKs/NF- κ B pathway of fibroblast-like synoviocytes. *Int. Immunopharmacol.* 101, 108352. doi:10.1016/j.intimp.2021.108352

Wang, S., Du, Q., Sun, J., Geng, S., and Zhang, Y. (2022). Investigation of the mechanism of Isobavachalcone in treating rheumatoid arthritis through a combination strategy of network pharmacology and experimental verification. *J. Ethnopharmacol.* 294, 115342. doi:10.1016/j.jep.2022.115342

Wang, X. H., Dai, C., Wang, J., Liu, R., Li, L., and Yin, Z. S. (2021). Therapeutic effect of neohesperidin on TNF-α-stimulated human rheumatoid arthritis fibroblast-like synoviocytes. *Chin. J. Nat. Med.* 19 (10), 741–749. doi:10.1016/S1875-5364(21) 60107-3

Wang, Z. Z., Huang, T. Y., Gong, Y. F., Zhang, X. M., Feng, W., and Huang, X. Y. (2020). Effects of sorafenib on fibroblast-like synoviocyte apoptosis in rats with adjuvant arthritis. *Int. Immunopharmacol.* 83, 106418. doi:10.1016/j.intimp.2020.106418

Wendel, H. G., De Stanchina, E., Fridman, J. S., Malina, A., Ray, S., Kogan, S., et al. (2004). Survival signalling by Akt and eIF4E in oncogenesis and cancer therapy. *Nature* 428 (6980), 332–337. doi:10.1038/nature02369

Wendling, D., Prati, C., Toussirot, E., and Herbein, G. (2010). Targeting intracellular signaling pathways to treat rheumatoid arthritis: Pandora's box? *Jt. Bone Spine* 77 (2), 96–98. doi:10.1016/j.jbspin.2010.01.004

Westhovens, R. (2023). Filgotinib in rheumatoid arthritis. *Expert Rev. Clin. Immunol.* 19 (2), 135–144. doi:10.1080/1744666X.2023.2149495

Weston, S., Macdonald, J. L., Williams, L. M., Roussou, E., Kang, N. V., Kiriakidis, S., et al. (2022). The JAK inhibitor baricitinib inhibits oncostatin M induction of proinflammatory mediators in *ex-vivo* synovial derived cells. *Clin. Exp. Rheumatol.* 40 (9), 1620–1628. doi:10.55563/clinexprheumatol/cfsajk

Wu, Z. M., Xiang, Y. R., Zhu, X. B., Shi, X. D., Chen, S., Wan, X., et al. (2022). Icariin represses the inflammatory responses and survival of rheumatoid arthritis fibroblast-like synoviocytes by regulating the TRIB1/TLR2/NF-kB pathway. *Int. Immunopharmacol.* 110, 108991. doi:10.1016/j.intimp.2022.108991

Xie, C., Jiang, J., Liu, J., Yuan, G., and Zhao, Z. (2019). Triptolide suppresses human synoviocyte MH7A cells mobility and maintains redox balance by inhibiting autophagy. *Biomed. Pharmacother.* 115, 108911. doi:10.1016/j.biopha.2019.108911

Xu, R., Liu, Z., Hou, J., Huang, T., and Yang, M. (2018). Osthole improves collageninduced arthritis in a rat model through inhibiting inflammation and cellular stress. *Cell Mol. Biol. Lett.* 23, 19. doi:10.1186/s11658-018-0086-0

Xu, Z., Shang, W., Zhao, Z., Zhang, B., Liu, C., and Cai, H. (2022). Curcumin alleviates rheumatoid arthritis progression through the phosphatidylinositol 3-kinase/protein kinase B pathway: An *in vitro* and *in vivo* study. *Bioengineered* 13 (5), 12899–12911. doi:10.1080/21655979.2022.2078942

Yalcin Kehribar, D., Ozgen, M., Yolbas, S., Yildirim, A., Onalan Etem, E., Ciftci, O., et al. (2021). The inhibition of Src kinase suppresses the production of matrix metalloproteinases in from synovial fibroblasts and inhibits MAPK and STATs pathways. *Turk J. Med. Sci.* 51 (4), 2142–2149. doi:10.3906/sag-2008-274

Yamada, H. (2023). The search for the pathogenic T cells in the joint of rheumatoid arthritis: Which T-cell subset drives autoimmune inflammation? *Int. J. Mol. Sci.* 24 (8), 6930. doi:10.3390/ijms24086930

Yang, F., Shen, J., and Cai, H. (2022). Paeoniflorin inhibits Wnt1/beta-catenin pathway and promotes apoptosis of fibroblast-like synoviocytes in patients with rheumatoid arthritis by upregulating lncRNA MALAT1. *Xi Bao Yu Fen Zi Mian Yi Xue Za Zhi* 38 (8), 692–698.

Yang, J., Liu, J., Li, J., Jing, M., Zhang, L., Sun, M., et al. (2022). Celastrol inhibits rheumatoid arthritis by inducing autophagy via inhibition of the PI3K/AKT/mTOR signaling pathway. *Int. Immunopharmacol.* 112, 109241. doi:10.1016/j.intimp.2022. 109241

Yang, Y., Dong, Q., and Li, R. (2017). Matrine induces the apoptosis of fibroblast-like synoviocytes derived from rats with collagen-induced arthritis by suppressing the activation of the JAK/STAT signaling pathway. *Int. J. Mol. Med.* 39 (2), 307–316. doi:10.3892/ijmm.2016.2843

Yang, Y. P., Jian, Y. Q., Liu, Y. B., Ismail, M., Xie, Q. L., Yu, H. H., et al. (2021). Triterpenoids from kadsura coccinea with their anti-inflammatory and inhibited proliferation of rheumatoid arthritis-fibroblastoid synovial cells activities. *Front. Chem.* 9, 808870. doi:10.3389/fchem.2021.808870

Yang, Y. P., Jian, Y. Q., Liu, Y. B., Xie, Q. L., Yu, H. H., Wang, B., et al. (2022). Heilaohuacid G, a new triterpenoid from Kadsura coccinea inhibits proliferation, induces apoptosis, and ameliorates inflammation in RA-FLS and RAW 264.7 cells via suppressing NF-?B pathway. *Phytother. Res.* 36 (10), 3900–3910. doi:10.1002/ptr.7527

Yang, Y., Ye, Y., Qiu, Q., Xiao, Y., Huang, M., Shi, M., et al. (2016). Triptolide inhibits the migration and invasion of rheumatoid fibroblast-like synoviocytes by blocking the activation of the JNK MAPK pathway. *Int. Immunopharmacol.* 41, 8–16. doi:10.1016/j. intimp.2016.10.005

Yao, R. B., Zhao, Z. M., Zhao, L. J., and Cai, H. (2017). Sinomenine inhibits the inflammatory responses of human fibroblast-like synoviocytes *via* the TLR4/MyD88/ NF-κB signaling pathway in rheumatoid arthritis. *Pharmazie* 72 (6), 355–360. doi:10. 1691/ph.2017.6946

Yi, L., Ke, J., Liu, J., Lai, H., Lv, Y., Peng, C., et al. (2021). Sinomenine increases a denosine A_{2A} receptor and inhibits NF-KB to inhibit arthritis in adjuvant-inducedarthritis rats and fibroblast-like synoviocytes through a7nAChR. *J. Leukoc. Biol.* 110 (6), 1113–1120. doi:10.1002/JLB.3MA0121-024RRRR

Yin, G., Wang, Y., Cen, X. M., Yang, M., Liang, Y., and Xie, Q. B. (2015). Lipid peroxidation-mediated inflammation promotes cell apoptosis through activation of NF- κ B pathway in rheumatoid arthritis synovial cells. *Mediat. Inflamm.* 2015, 460310. doi:10.1155/2015/460310

Yoon, H. Y., Lee, E. G., Lee, H., Cho, I. J., Choi, Y. J., Sung, M. S., et al. (2013). Kaempferol inhibits IL-1 β -induced proliferation of rheumatoid arthritis synovial fibroblasts and the production of COX-2, PGE2 and MMPs. *Int. J. Mol. Med.* 32 (4), 971–977. doi:10.3892/ijmm.2013.1468

Yu, H. H., Li, M., Li, Y. B., Lei, B. B., Yuan, X., Xing, X. K., et al. (2020). Benzoylaconitine inhibits production of IL-6 and IL-8 via MAPK, Akt, NF-xB signaling in IL-1β-induced human synovial cells. *Biol. Pharm. Bull.* 43 (2), 334–339. doi:10.1248/bpb.b19-00719

Yu, X., Zhou, J., Zhao, F., Liu, X., Mao, Y., Diao, L., et al. (2021). Tomatidine suppresses the destructive behaviors of fibroblast-like synoviocytes and ameliorates type

II collagen-induced arthritis in rats. Front. Pharmacol. 12, 670707. doi:10.3389/fphar. 2021.670707

Yu, Y., Koehn, C. D., Yue, Y., Li, S., Thiele, G. M., Hearth-Holmes, M. P., et al. (2015). Celastrol inhibits inflammatory stimuli-induced neutrophil extracellular trap formation. *Curr. Mol. Med.* 15 (4), 401–410. doi:10.2174/1566524015666150505160743

Zeng, S., Wang, K., Huang, M., Qiu, Q., Xiao, Y., Shi, M., et al. (2017). Halofuginone inhibits TNF- α -induced the migration and proliferation of fibroblast-like synoviocytes from rheumatoid arthritis patients. *Int. Immunopharmacol.* 43, 187–194. doi:10.1016/j. intimp.2016.12.016

Zhai, K. F., Duan, H., Cui, C. Y., Cao, Y. Y., Si, J. L., Yang, H. J., et al. (2019). Liquiritin from Glycyrrhiza uralensis attenuating rheumatoid arthritis via reducing inflammation, suppressing angiogenesis, and inhibiting MAPK signaling pathway. *J. Agric. Food Chem.* 67 (10), 2856–2864. doi:10.1021/acs.jafc.9b00185

Zhang, D., Ning, T., and Wang, H. (2022). Vitexin alleviates inflammation and enhances apoptosis through the regulation of the JAK/STAT/SOCS signaling pathway in the arthritis rat model. *J. Biochem. Mol. Toxicol.* 36 (12), e23201. doi:10.1002/jbt. 23201

Zhang, G., Liu, B., Zeng, Z., Chen, Q., Feng, Y., and Ning, X. (2021). Oxymatrine hydrazone (OMTH) synthesis and its protective effect for rheumatoid arthritis through downregulation of MEK/NF-κB pathway. *Environ. Toxicol.* 36 (12), 2448–2453. doi:10.1002/tox.23357

Zhang, L., Lin, Y., Xu, X., Liu, H., Wang, X., and Pan, J. (2023). Telotristat Etiprate alleviates rheumatoid arthritis by targeting LGALS3 and affecting MAPK signaling. *Intractable Rare Dis. Res.* 12 (1), 45–57. doi:10.5582/irdr.2022.01121

Zhang, Q., Liu, J., Zhang, M., Wei, S., Li, R., Gao, Y., et al. (2019). Apoptosis induction of fibroblast-like synoviocytes is an important molecular-mechanism for herbal medicine along with its active components in treating rheumatoid arthritis. *Biomolecules* 9 (12), 795. doi:10.3390/biom9120795

Zhang, X., Guan, X., Piao, Y., Che, X., Si, M., and Jin, J. (2022). Baicalein induces apoptosis of rheumatoid arthritis synovial fibroblasts through inactivation of the PI3K/

Akt/mTOR pathway. Evid. Based Complement. Altern. Med. 2022, 3643265. doi:10. 1155/2022/3643265

Zhang, X., Ye, G., Wu, Z., Zou, K., He, X., Xu, X., et al. (2020). The therapeutic effects of edaravone on collagen-induced arthritis in rats. *J. Cell Biochem.* 121 (2), 1463–1474. doi:10.1002/jcb.29382

Zhang, Y., Tang, L. D., Wang, J. Y., Wang, H., Chen, X. Y., Zhang, L., et al. (2022). Anti-inflammatory effects of aucubin in cellular and animal models of rheumatoid arthritis. *Chin. J. Nat. Med.* 20 (6), 458–472. doi:10.1016/S1875-5364(22)60182-1

Zhong, B., Guo, S., Yang, Z., Han, L., Du, J., Chen, J., et al. (2021). Roflumilast reduced the IL-18-induced inflammatory response in fibroblast-like synoviocytes (FLS). ACS Omega 6 (3), 2149–2155. doi:10.1021/acsomega.0c05281

Zhong, Z., Qian, Z., Zhang, X., Chen, F., Ni, S., Kang, Z., et al. (2019). Tetrandrine prevents bone loss in ovariectomized mice by inhibiting RANKL-induced osteoclastogenesis. *Front. Pharmacol.* 10, 1530. doi:10.3389/fphar.2019.01530

Zhou, H., Liu, J. X., Luo, J. F., Cheng, C. S., Leung, E. L., Li, Y., et al. (2017). Suppressing mPGES-1 expression by sinomenine ameliorates inflammation and arthritis. *Biochem. Pharmacol.* 142, 133–144. doi:10.1016/j.bcp.2017.07.010

Zhou, J., Mao, Y., Shi, X., Zhang, Y., Yu, X., Liu, X., et al. (2022). Peimine suppresses collagen-induced arthritis, activated fibroblast-like synoviocytes and TNF α -induced MAPK pathways. *Int. Immunopharmacol.* 111, 109181. doi:10.1016/j.intimp.2022. 109181

Zhou, Y. R., Zhao, Y., Bao, B. H., and Li, J. X. (2015). SND-117, a sinomenine bivalent alleviates type II collagen-induced arthritis in mice. *Int. Immunopharmacol.* 26 (2), 423–431. doi:10.1016/j.intimp.2015.04.006

Zuo, J., Xia, Y., Li, X., Ou-Yang, Z., and Chen, J. W. (2015). Selective modulation of MAPKs contribute to the anti-proliferative and anti-inflammatory activities of 1,7dihydroxy-3,4-dimethoxyxanthone in rheumatoid arthritis-derived fibroblast-like synoviocyte MH7A cells. *J. Ethnopharmacol.* 168, 248–254. doi:10.1016/j.jep.2015. 03.069

Glossary

Glossaly		TGF-β	Transforming growth factor-β
D A	Dhaumataid arthritis	NFATc1	Nuclear factor of activated T cells
KA NEATal	c Fee and pucker factor of activated T calls cl	FLIP	Anti-apoptotic molecule FLICE inhibitory protein
FIG	Eibenhant like amoring to	NLRP3	NOD-like receptor protein 3
ATE2	A stirsting transmitting faster 2	TLRs	Toll-like receptors
AIF2	Activating transcription factor-2	HO-1	Heme oxygenase
NSAIDS	Non-steroidal anti-inflammatory drugs	DKK-1	Dickkopf-1
PGE (2)	Prostagiandin E (2)	SIRT1	Silent information regulator 1
DMARDs	Disease-modifying anti-rheumatic drugs	CXCL-8	CXC motif chemokine 8
ROS	Reactive oxygen species	MCP-1	Monocyte chemotactic protein-1
MTX	Methotrexate	CCL2	CC motif chemokine ligand 2
HIF1	Hypoxia-inducible factor 1	OPN	Osteopontin
bDMARDs	Biologic disease-modifying anti-rheumatic drugs	MHCII	Major histocompatibility complex class II
CIA	Collagen-induced arthritis	ATP	Adenosine triphosphate
ECM	Extracellular matrix	FcyRs	Fc-gamma receptors
IκB	Inhibitor of KB	a7nAChR	α7-nicotinic acetylcholine receptor
MAPK	Mitogen-activated protein kinase	APRIL	A proliferation-inducing ligand
Bcl-2	B-cell lymphoma-2	mPGES-1	Microsomal prostaglandin E synthase 1
NF-ĸB	Nuclear factor kappa-B	ABCs	Age-associated B cells
Bax	Bcl-2-associated X	AP-1	Activated protein-1
JAK	Janus kinase	VCAM-1	Vascular cell adhesion molecule-1
AA	Adjuvant-induced arthritic	FDA	Food and Drug Administration
STAT	Signal transducers and activators of transcription	ICAM-1	Intercellular cell adhesion molecule-1
OPG	Osteoprotegerin	EMA	European Medicines Agency
TNF	Tumor necrosis factor	LFA-1	Lymphocyte function-associated antigen
MEKK	Mitogen-activated protein kinase kinase	csDMARDs	Conventional synthetic disease-modifying anti-rheumatic drugs
IL	Interleukin	JNK	c-Jun N-terminal kinase
IKK	IKB kinase	TNFSF11	TNF Superfamily Member 11
NOS	Nitric oxide synthase	ERK	Extracellular regulated protein kinase
TGM2	Transglutaminase 2	AESI	Adverse events of special interest
COX-2	Cyclooxygenase-2	Bcl-2	B-cell lymphoma 2
NLS	Nuclear-localization sequence	OSM	Oncostatin M
RANK	Receptor activator of NF-ĸB	Ang1	Angiopoietin-1
HDAC	Histone deacetylases	α-SMA	Smooth muscle alpha-actin
RANKL	Receptor activator of NF-KB ligand	Tie-2	Tyrosine-protein kinase receptor
GRK2	G protein-coupled receptor kinase 2	SOCS	Suppressor of cytokine signaling
VEGF	Vascular endothelial growth factor	GSK-3	Glycogen synthase kinase 3
M-CSF	Macrophage colony stimulating factor	ТАК	TGF β-activated kinase
MMPs	Matrix metalloproteinases	мкк	Mitogen-activated protein kinase kinase
MDA	Malondialdehyde	mTORC1	mTOR complex 1
PDGF	Platelet-derived growth factor	mTOR	Mammalian target of rapamycin
TRIB1	Tribbles pseudokinase 1	mion	manimuman target of rapaniyent