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Mini Review Cell death pathology: Perspective for human diseases

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ABSTRACT

Apoptosis, a genetically regulated form of cell death with distinct biochemical and morphological features, plays a relevant physiological and pathological role in the organism, being pivotal in the maintenance of tissue development and homeostasis in the adult as well as in the regulation of immune responses. Deregulation of this process causes several human disorders including cancer, autoimmune and neurodegenerative diseases. Thus, modulation of the apoptotic process and of cell death in general, is a potential therapeutic approach for the treatment of several human pathologies.

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1. Introduction

Since 1965, when the first description of programmed cell death was independently made by Richard Lockshin and John Kerr, searching for the term apoptosis on PubMed yields 206,244 papers. Indeed, during the last year, nearly 20,000 papers have been published using the same search term, indicating that apoptosis remains a major interest in science, accounting for over 3% of all scientific publications. This is what John Kerr [1,2] originally wrote, suggesting the involvement of apoptosis at both physiological and pathological levels:

"Apoptosis seems to be involved in cell turnover in many healthy adult tissues and is responsible for focal elimination of cells during normal embryonic development. It occurs spontaneously in untreated malignant neoplasms, and participates in at least some types of therapeutically induced tumour regression. It is implicated in both physiological involution and atrophy of various tissues and organs. It can also be triggered by noxious agents, both in the embryo and adult animal."

At the present time, a long list of pathologies is associated with deregulation of apoptosis, including cancer (carcinoma, sarcoma, leukemia, lymphoma, and myeloma), autoimmune diseases (systemic lupus erythematosus, immune-mediated thrombocytopenia, autoimmune neutropenia, glomerulonephritis, rheumatoid arthritis, Hashimoto's thyroiditis, insulin dependent diabetes mellitus, and multiple sclerosis) and viral infections (Poxvirus, Adenovirus, and Herpesvirus). In contrast, excessive apoptosis has been associated, for example, with neurodegenerative disorders (Parkinson's, Alzheimer's, Huntington's [3], amyotrophic lateral sclerosis, cerebellar degeneration, status epilepticus [4], prion disease [5], glaucoma [6,7], diabetic retinophaty [8], retinitis pigmentosa [9], and spinal muscular atrophy), AIDS, liver disease, haematological diseases and ischemic injury (hypoxia-ischemia [10], stroke [11], and myocardial infarction). In 45 years of research on cell death, we have also learned that modulation of apoptosis could be useful in the treatment of these different pathologies. Indeed, control of cancer through the induction of apoptosis is the main therapeutic approach in the clinic [12–15]. Correspondingly, the prevention or delay of apoptosis would seem appropriate in diseases where excessive cell death is present, such as neurodegenerative disorders [16,17].

2. Is cell death finished?

For the first time in the last 25 years, the number of publications in cell death has not increased. Does this decline mean the beginning of the end for the cell death field? In fact, the number of papers on, *sensu strictu*, the basic molecular mechanisms of cell death is decreasing. The real reason for this apparent decline is the

Abbreviations: AIDS, acquired immune deficiency syndrome; TIAF1, TGF β induced anti-apoptotic factor 1; TGF β , transforming growth factor beta; AD, Alzheimer disease; APP, amyloid precursor protein; PD, Parkinson's disease; NSC, neural stem cells; AICD, activation induced cell death; HIV, human immunodeficiency virus.

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Table 1

Apoptosis-based clinical trials.

Target	Reagent	Principle	Company	Effects	Trial status
TRAIL	HGS-ETR1	Agonist TRAIL-R1 mAb	HGS	Apoptosis induction	Phase II Completed Carcinoma Non-Small-Cell Lung
	HGS-ETR2	Agonist TRAIL-R2 mAb	HGS	Apoptosis induction	Lymphoma Phase I Kidney Cancer Lymphoma Neuroblastoma Phase I Solid Tumor Non-Hodgkin Lymphoma
	PRO1762	Soluble human Apo2L/TRAIL	Amgen	Apoptosis induction	
	TRA-8	Agonist TRAIL-R2 mAb		Apoptosis induction	Phase II Ovarian Cancer
CD95	CD95-Fc	Humanized CD95 Fc-decoy construct	ApoGenix	Inhibition of CD95 signaling	Phase II Glioblastoma Phase I GvDH
SMAC	AT-406	XIAP-Smac mimetic	Ascenta Therapeutics	Tumor suppression	Phase I Solid Tumor
	AEG35156	XIAP antisense oligonucleotide	Ascenta Therapeutics	Antitumor activity	Lymphoma Phase I/II Terminated Human mammary Carcinoma Phase I Terminated Advanced cancer
	Low MW SMAC mimetc (LBW247)	Inhibitors of XIAP, cIAP-1 and 2			Phase Ib Clinical trials
SURVIVIN	LY2181308	Survivin antisense	Eli Lilly	Antitumor activity	Phase II Completed, In combination therapy in AML Phase I/II withdrawn Hepatocellular Carcinoma
MPT	Lonidamine	Dichlorinated indazole-3-carboxil acid derivate	Threshold Pharmaceuticals	Permeabilize mitochondria	Phase II/III Terminated Beningn
	Cladribine (Litak)	2-chloro-2' dexyadenosine		Apoptosis induction	Approved for Hairy-cell leukemia Phase II/III Photodynamic therapy, Advanced Pancreatic Cancer Phase I/II Dysplastic Nevus Syndrome
	Verterporfin	Porphyrin photosensitizer		Triggering of cytochrome c release	
	Betulinic acid	Pentacyclic triterpenoid		Inducer of apoptosis	
ANTI-APOPTOTIC Bcl2 MEMBERS	Genasense	Bcl-2 antisense oligonucleotides	Genta		Phase III Advanced Melanoma CLL Phase II Non-Hodgkin Lymphoma
p53	INGN201	p53-expressing adenovirus	Introgen Therapeutics	Apoptosis Induction	Phase III Completed In combination therapy in Breast
	SCH58500	p53-expressing adenovirus	Schering-Plough	Apoptosis Induction	Phase I Completed Ovarian Cancer Phase III completed, In combination therapy in Ovarian Cancer
	ONYX-015 Amifostine	p53 delivery adenovirus Restoration of p53	Onyx AstraZeneca	Antitumor activity Restores function of mutant p53	Withdrawn Phase II/IV Completed, Colorectal Head and Neck, Lung Cancer
Proteasome inhibitors	Epoxomicin	Streptomyces epoxyketone	Onyx	Apoptotic effect	Carfilzomib analog of epoxomocin Phase I/II Multiple Myeloma

Example of clinical trials on cell death targets, approved by US NIH (http://clinicaltrials.gov/).

reduced number of basic articles on caspases, on the molecular mechanisms of the Bcl2 family, on IAPs and so on. Conversely, publication on the translational aspects of cell death, such as cardiovascular diseases, neurodegenerative pathology and applied oncology, is in fact increasing. Therefore there is a shift from the basic mechanisms towards its translational aspects. Consequently a large number of publications are no longer classified in the apoptosis category, but fall under categories such as cardiology, immunology, oncology and neurodegeneration. At the same time there is a significant effort to develop pharmacological regulators of cell death in these different pathologies, based on particular mechanisms such as inhibitors of Bcl2, IAPs, caspases, p53 and mitochondrial permeabilization [18–20]. Table 1 shows a simplified highlight of some of the clinical trials using regulators of the cell death pathways.

However, the main mechanisms of cell death have not been fully clarified at the molecular level. For example, the mechanism of action of Bcl2 is far from clear; so is the function of Bax and Bak. Consequently, many questions remain open not only on the Bcl2 family but also on death receptor signaling, IAPs and several caspases. The impression that we know all about cell death should be compared to the late XX century, early XIX century physics, before Albert Einstein and Richard Feynman. So, several crucial mechanistic points await clarification and possibly pharmacological exploitation.

Last but not least, an incremental understanding has occurred in alternative regulatory mechanisms. Autophagy has exploded with a revival of flourishing molecular details [21–28]. And a similar expansion is occurring in less known death pathways such as for example pyroptosis, necroptosis or the Wallerian degeneration.

The consequence of this clinical trend of the cell death field is that it becomes more diffuse and less focused, resulting in less "cell death" meetings and more sessions in all specialist medical meetings. It is however pivotal to maintain a central focus for discussions of distinct pathways, pharmaceutical exploitation and pathological applications.

One thing is definitely clear. The fashion of cell death has ended, and, as expected, the cover page has been temporarily been taken by stem cells and a molecular revival of cancer metabolism. But is not this positive?

Hence, the question "is cell death finished?" is erroneously proposed. The more appropriate formulation should be "how is cell death evolving?", or "has cell death become translational?"

3. Apoptosis in cancer

If we ask a student an example of pathology that is associated with defective cell death, the most frequent answer would be cancer. Indeed, evasion of apoptosis is one of the basic features of cancer [29]. A large number of papers have described the underlying molecular mechanisms [30-32]. According to the histological National Cancer Institute classification, cancer can be classified in five major categories: carcinoma, sarcoma, myeloma, leukemia and lymphoma. Focusing only on 2010, apoptosis is present in all cancer types, being predominant in carcinoma [33-37] and leukemia [38-40] (Fig. 1A). Thus, despite the fact that leukemia accounts for just 3% of all cancer cases [41], it is the second most investigated cancer from the apoptotic point of view. Looking at carcinomas, that accounts for about 80% of all cancer cases; the number of publications in 2010 in each cancer hallmark (evading apoptosis, tissue invasion and metastasis, angiogenesis and increased proliferation) is shown in Fig. 1B. We would like to think that this picture could help us understand the state of the art in this field and its future direction. A primary role, in our experience, is within the p53 family [42-47].

Table 1 shows some example of ongoing clinical trials based on regulators of cell death. Killing malignant cells is the main target of cancer therapy, but this could be done not just by inducing apoptosis, but also through the modulation of other cancer hallmarks.

4. Apotosis in neurodegenerative disorders

Neurodegeneration includes pathology that results in loss of neurons and is manifest as a broad group of neurological disorders. Genetic [48,49] and environmental factors [50–53] are the main causes of neurodegenerative diseases. In general, we can consider cell death (neuronal death) as the mechanism underlying these conditions and understanding the pathways that regulate cell death could help to find the road leading to the development of therapy [54–56].

In Alzheimer disease (AD), the extracellular accumulation of fibrillar amyloid β and the intracellular deposition of neurofibrillary tangles are the main features [57,58]. Recently, alteration of TGF- β signaling has been proposed as a further mechanism that could regulate amyloid- β aggregation and plaque formation and Lee et al. have found aggregates of TGF β -induced anti-apoptotic factor 1 (TIAF1 in the hippocampus of AD patients) [59]. Activation



Fig. 1. Current trend of cancer related papers on cell death. (A) Numbers of publications in 2010 on apoptosis in different type of cancers. The search was performed as following: apoptosis or "cell death" with the specific histological type, according to the NCI classification (http://training.seer.cancer.gov/disease/categories/classification.html). (B) Numbers of carcinoma-related scientific papers published in 2010 focused on the selective mechanisms.

of the caspase machinery precedes tangle formation [60] and capsase-6 as well as caspase-3 [61] have been implicated in AD pathology. Indeed, the neuritic beading induced by amyloid precursor protein (APP) is dependent on caspase-6, since it is inhibited by z-VEID-fmk (a specific caspase-6 inhibitor) or by overexpressing a caspase-6 dominant negative [62]. Plaque formation is also secondary to the accumulation and activation of microglia in the CA1 area of hippocampus in a mouse model of AD [63].

Parkinson's disease (PD) is the second most common pathology affecting the central nervous system. A pivotal role in the pathogenesis of PD is played by the α -synuclein protein, and the gene encoding α -synuclein has been found mutated [64,65] or amplified (triplication) [66] in at least some cases of PD. Moreover, ectopic expression of α -synuclein leads to cell death through a mechanism that remains poorly understood [67]. Experimental evidence suggests that the 14-3-3 proteins, negative regulators of cell death, are deregulated and co-aggregated with α -synuclein Lewy bodies in PD [68]. In a recent paper, Yacoubian et al. using two different models of PD, confirm a strong neuroprotective effect of 14-3-3 proteins, in particular of 14-3-3 θ , $-\epsilon$ and $-\gamma$ isoforms [69]. Because PPAR- γ ligands can induce 14-3-3 expression [70], they could therefore be used as neuroprotective agents in PD therapy.

Several observations indicate that adult neurogenesis is impaired in neurodegenerative disorders, suggesting that the postmitotic neuron is not the only target affected by these disorders [71]. Indeed, the proliferation and differentiation of neural stem cells (NSC) into mature neurons is reduced by amyloid- β in human AD. The p73 gene [72,73], a member of p53 family, is implicated in the pathogenesis of AD [74–77]. In 2010, several reports identified p73 as a positive regulator of self-renewal with essential roles both in the maintenance of embryonic and adult neurogenesis, and by inhibiting premature senescence of NSC [78–81]. Moreover, this positive regulation of self-renewal of NCS by p73 is independent of p53.

5. Apoptosis in immune diseases

Physiologically, the immune system is one of the largest users of apoptosis, both in terms of the generation of mature immunologically active cells and in limiting the extent of an immune response [82]. Thus, apoptosis plays a role in both positive and negative selection of immune cells in the thymus, for example by eliminating autoreactive cells, thereby establishing tolerance to self-tissues. Indeed, it has been estimated that 90% of immature thymocytes are eliminated by apoptosis during thymic education. Moreover, the clonal expansion of cells during an immune response is curtailed by apoptosis (Activation Induced Cell Death; AICD) once the response has eliminated its stimulus. In addition, cytotoxic T and NK cells kill virus-infected or transformed target cells by inducing apoptotic cell death.

Defective cell death in the immune system can result in autoimmune disorders. For example, the CD95/CD95L system is a potent inducer of apoptosis on activated T lymphocytes and it has been demonstrated that alteration in Fas-mediated apoptosis is the basic pathology underlying the autoimmune disease systemic lupus erythematosus [83]. In apoptosis mediated by CD95, both extrinsic and intrinsic pathways are activated, depending on the cell context [84–86]. A layer of complexity in this system has been added by the observations that, in some cellular contexts, caspase cleavage of Golgi proteins is required for CD95/CD95L mediated apoptosis [87,88].

Acquired immunodeficiency syndrome (AIDS) as a consequence of human immunodeficiency virus (HIV) infection is the most dramatic example of immune system pathology linked to excessive cell death [89]. An update on the current understanding of the role and mechanisms of accelerated apoptosis of T cells in the immunopathogenesis of HIV infection has now been provided by Cummins et al. and may be a useful stimulus to revisit our concepts of treatment for this devastating disease [90].

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References

- J.F. Kerr, A.H. Wyllie, A.R. Currie, Apoptosis: a basic biological phenomenon with wide-ranging implications in tissue kinetics, Br. J. Cancer 26 (1972) 239– 257.
- [2] G. Kroemer, L. Galluzzi, P. Vandenabeele, et al., Classification of cell death: recommendations of the Nomenclature Committee on Cell Death 2009, Cell Death Differ. 16 (2009) 3–11.
- [3] J. Sassone, C. Colciago, P. Marchi, et al., Mutant Huntingtin induces activation of the Bcl-2/adenovirus E1B 19-kDa interacting protein (BNip3), Cell Death Dis. 1 (2010) e7.
- [4] T. Engel, K. Tanaka, E.M. Jimenez-Mateos, et al., Loss of p53 results in protracted electrographic seizures and development of an aggravated epileptic phenotype following *status epilepticus*, Cell Death Dis. 1 (2010) e79.
- [5] G. Di Guardo, G. Marruchella, Prions and neuronal death, Cell Death Dis 1 (2010) e6.
- [6] N. Calandrella, C. De Seta, G. Scarsella, G. Risuleo, Carnitine reduces the lipoperoxidative damage of the membrane and apoptosis after induction of cell stress in experimental glaucoma, Cell Death Dis. 1 (2010) e62.
- [7] M. Almasieh, Y. Zhou, M.E. Kelly, C. Casanova, A. Di Polo, Structural and functional neuroprotection in glaucoma: role of galantamine-mediated activation of muscarinic acetylcholine receptors, Cell Death Dis. 1 (2010) e27.
- [8] L. Perrone, T.S. Devi, K.I. Hosoya, T. Terasaki, L.P. Singh, Inhibition of TXNIP expression *in vivo* blocks early pathologies of diabetic retinopathy, Cell Death Dis. 1 (2010) e65.
- [9] J. Sancho-Pelluz, M.V. Alavi, A. Sahaboglu, et al., Excessive HDAC activation is critical for neurodegeneration in the rd1 mouse, Cell Death Dis. 1 (2010) e24.
- [10] S. Chi, W. Cai, et al., *Baifuzi* reduces transient ischemic brain damage through an interaction with the STREX domain of BK_{Ca} channels, Cell Death Dis. 1 (2010) e13.
- [11] K.R. Francis, L. Wei, Human embryonic stem cell neural differentiation and enhanced cell survival promoted by hypoxic preconditioning, Cell Death Dis. 1 (2010) e22.
- [12] M. Degli Esposti, Bcl-2 antagonists and cancer: from the clinic back to the bench, Cell Death Dis. 1 (2010) e37.
- [13] C.R. Reis, A.M. van der Sloot, A. Natoni, et al., Rapid and efficient cancer cell killing mediated by high-affinity death receptor homotrimerizing TRAIL variants, Cell Death Dis. 1 (2010) e83.
- [14] S. Schneider-Jakob, N. Corazza, A. Badmann, et al., Synergistic induction of cell death in liver tumor cells by TRAIL and chemotherapeutic drugs via the BH3only proteins Bim and Bid, Cell Death Dis. 1 (2010) e86.
- [15] O. Rigaud, N.O. Fortunel, P. Vaigot, et al., Exploring ultrashort high-energy electron-induced damage in human carcinoma cells, Cell Death Dis. 1 (2010) e73.
- [16] M.B. Sättler, M. Bähr, Future neuroprotective strategies, Exp. Neurol. 225 (2010) 40-47.
- [17] Q. Li, H. Li, K. Roughtonet, et al., Lithium reduces apoptosis and autophagy after neonatal hypoxia–ischemia, Cell Death Dis. 1 (2010) e56.
- [18] U. Fischer, K. Schulze-Osthoff, Apoptosis-based therapies and drug targets, Cell Death and Differentiation 12 (2005) 942–961.
- [19] J.C. Reed Drug, Insight: cancer therapy strategies based on restoration of endogenous cell death mechanisms, Nat. Clin. Pract. Oncol. 3 (2006) 388–398.
- [20] J.C. Reed, Apoptosis-based therapies, Nat. Rev. Drug Discov. 1 (2002) 111-121.
- [21] C. Liang, Negative regulation of autophagy, Cell Death Diff. 17 (2010) 1807– 1815.
- [22] M.A. Delgado, V. Deretic, Toll-like receptors in control of immunological autophagy, Cell Death Diff. 16 (2009) 976–983.
- [23] T. Noda, N. Fujita, T. Yoshimori, The late stages of autophagy: how does the end begin? Cell Death Diff 16 (2009) 984–990.
- [24] A. Longatti, S.A. Tooze, Vesicular trafficking and autophagosome formation, Cell Death Diff. 16 (2009) 956–965.
- [25] A. Eisenberg-Lerner, S. Bialik, H.U. Simon, A. Kimchi, Life and death partners: apoptosis, autophagy and the cross-talk between them, Cell Death Diff. 16 (2009) 966–975.
- [26] T. Vellai, Autophagy genes and ageing, Cell Death Diff. 16 (2009) 94-102.
- [27] N. Kourtis, N. Tavernarakis, Autophagy and cell death in model organisms, Cell
- Death Diff. 16 (2009) 21–30.
 [28] M.C. Maiuri, E. Tasdemir, A. Criollo, et al., Control of autophagy by oncogenes and tumor suppressor genes, Cell Death Diff. 16 (2009) 87–93.
- [29] D. Hanahan, R.A. Weinberg, The hallmarks of cancer, Cell 100 (2000) 57–70.
- [30] F.H. Igney, P.H. Krammer, Death and anti-death: tumour resistance to apoptosis, Nat. Rev. Cancer 2 (2002) 277–288.

- [31] S.W. Tait, D.R. Green, Caspase-independent cell death: leaving the set without the final cut, Oncogene 27 (2008) 6452–6461.
- [32] M. Fricker, J. O'Prey, A.M. Tolkovsky, K.M. Ryan, Phosphorylation of Puma modulates its apoptotic function by regulating protein stability, Cell Death Dis. 1 (2010) e59.
- [33] M.S. Goldberg, D. Xing, Y. Ren, et al., Nanoparticle-mediated delivery of siRNA targeting Parp1 extends survival of mice bearing tumors derived from Brca1deficient ovarian cancer cells, Proc. Natl. Acad. Sci. USA 108 (2011) 745–750.
- [34] M. Flourakis, V. Lehen'kyi, B. Beck, et al., Orai1 contributes to the establishment of an apoptosis-resistant phenotype in prostate cancer cells, Cell Death Dis. 1 (2010) e75.
- [35] N. Bhatnagar, X. Li, S.K.R. Padi, Q. Zhang, M.-S. Tang, B. Guo, Downregulation of miR-205 and miR-31 confers resistance to chemotherapy-induced apoptosis in prostate cancer cells, Cell Death Dis. 1 (2010) e105.
- [36] Y. Wang, P. Nangia-Makker, V. Balan, V. Hogan, A. Raz, Calpain activation through galectin-3 inhibition sensitizes prostate cancer cells to cisplatin treatment, Cell Death Dis. 1 (2010) e101.
- [37] N. Fenouille, A. Puissant, M. Dufies, et al., Persistent activation of the Fyn/ERK kinase signaling axis mediates imatinib resistance in chronic myelogenous leukemia cells through upregulation of intracellular SPARC, Cancer Res. 70 (2010) 9659–9670.
- [38] B. Accordi, V. Espina, V.M. Giordan, et al., Functional protein network activation mapping reveals new potential molecular drug targets for poor prognosis pediatric BCP-ALL, PLoS One 5 (2010) e13552.
- [39] N. Heidari, M.A. Hicks, H. Harada, GX15-070 (obatoclax) overcomes glucocorticoid resistance in acute lymphoblastic leukemia through induction of apoptosis and autophagy, Cell Death Dis. 1 (2010) e76.
- [40] M. Wemeau, O. Kepp, A. Tesnière, et al., Calreticulin exposure on malignant blasts predicts a cellular anticancer immune response in patients with acute myeloid leukemia, Cell Death Dis. 1 (2010) e104.
- [41] A. Jemal, R. Siegel, E. Ward, et al., Cancer statistics, 2008, CA Cancer J. Clin. 58 (2008) 71–96.
- [42] S. Ramadan, A. Terrinoni, M.V. Catani, et al., p73 induces apoptosis by different mechanisms, Biochem. Biophys. Res. Commun. 331 (2005) 713–717.
- [43] T. Schilling, A. Kairat, G. Melino, et al., Interference with the p53 family network contributes to the gain of oncogenic function of mutant p53 in hepatocellular carcinoma, Biochem. Biophys. Res. Commun. 394 (2010) 817– 823.
- [44] T. Schilling, E.S. Schleithoff, A. Kairat, et al., Active transcription of the human FAS/CD95/TNFRSF6 gene involves the p53 family, Biochem. Biophys. Res. Commun. 387 (2009) 399–404.
- [45] G. Melino, V. De Laurenzi, K.H. Vousden, p73: friend or foe in tumorigenesis, Nat. Rev. Cancer 2 (2002) 605–615.
- [46] G. Melino, F. Bernassola, M. Ranalli, et al., p73 induces apoptosis via PUMA transactivation and Bax mitochondrial translocation, J. Biol. Chem. 279 (2004) 8076–8083.
- [47] J.G. Gong, A. Costanzo, H.Q. Yang, et al., Regulation of the p53 homolog p73 by c-Abl tyrosine kinase in cell death response to cisplatin, Nature 399 (1999) 806–809.
- [48] C.M. Tanner, R. Ottman, S.M. Goldman, et al., Parkinson disease in twins: an etiologic study, JAMA 281 (1999) 341–346.
- [49] M. Hutton, J. Perez-Tur, J. Hardy, Genetics of Alzheimer's disease, Essays Biochem. 33 (1998) 117-131.
- [50] L.T. Kurtland, Amyotrophic lateral sclerosis and Parkinson's disease complex on Guam linked to an environmental neurotoxin, Trends Neurosci. 11 (1988) 51–54.
- [51] S. Przedborski, M. Vila, MPTP: a review of its mechanisms of neurotoxicity, Clin. Neuro. Res. 1 (2001) 407–418.
- [52] C.M. Tanner, The role of environmental toxins in the etiology of Parkinson's disease, Trends Neurosci. 12 (1989) 49–54.
- [53] C. Corona, F. Masciopinto, E. Silvestri, et al., Dietary zinc supplementation of 3xTg-AD mice increases BDNF levels and prevents cognitive deficits as well as mitochondrial dysfunction, Cell Death Dis. 1 (2010) e91.
- [54] J.B. Martin, Molecular basis of the neurodegenerative disorders, N. Engl. J. Med. 340 (1999) 1970–1980.
- [55] W.H. Dribben, L.N. Eisenman, S. Mennerick, Magnesium induces neuronal apoptosis by suppressing excitability, Cell Death Dis. 1 (2010) e63.
- [56] J. Nicolai, S. Burbassi, J. Rubin, O. Meucci, CXCL12 inhibits expression of the NMDA receptor's NR2B subunit through a histone deacetylase-dependent pathway contributing to neuronal survival, Cell Death Dis. 1 (2010) e33.
- [57] C.L. Masters, G. Simms, N.A. Weinman, et al., Amyloid plaque core protein in Alzheimer disease and Down syndrome, Proc. Natl. Acad. Sci. USA 82 (1985) 4245–4249.
- [58] J. Gotz, F. Chen, J. van Dorpe, R.M. Nitsch, Formation of neurofibrillary tangles in P3011 tau transgenic mice induced by Abeta 42 fibrils, Science 293 (2001) 1491–1495.
- [59] M.-H. Lee, S.-R. Lin, J.-Y. Chang, et al., TGF- β induces TIAF1 self-aggregation via type II receptor-independent signaling that leads to generation of amyloid β plaques in Alzheimer's disease, Cell Death Dis. 1 (2010) e110.
- [60] J. Avila, Alzheimer disease: caspases first, Nat. Rev. Neurol. 6 (2010) 587-588.
- [61] M. D'Amelio, V. Cavallucci, S. Middei, et al., Caspase-3 triggers early synaptic dysfunction in a mouse model of Alzheimer's disease, Nat. Neurosci. 14 (2011) 69–76.

- [62] S.N. Sivananthan, A.W. Lee, C.G. Goodyer, A.C. LeBlanc, Familial amyloid precursor protein mutants cause caspase-6-dependent but amyloid β-peptideindependent neuronal degeneration in primary human neuron cultures, Cell Death Dis. 1 (2010) e100.
- [63] J.J. Rodríguez, J. Witton, M. Olabarria, H.N. Noristani, A. Verkhratsky, Increase in the density of resting microglia precedes neuritic plaque formation and microglial activation in a transgenic model of Alzheimer's disease, Cell Death Dis. 1 (2010) e1.
- [64] M.H. Polymeropoulos, C. Lavedan, E. Leroy, et al., Mutation in the alphasynuclein gene identified in families with Parkinson's disease, Science 276 (1997) 2045–2047.
- [65] A. Athanassiadou, G. Voutsinas, L. Psiouri, et al., Genetic analysis of families with Parkinson disease that carry the Ala53Thr mutation in the gene encoding alpha-synuclein, Am. J. Hum. Genet. 65 (1999) 555–558.
- [66] A.B. Singleton, M. Farrer, A. Johnson, et al., Alpha-Synuclein locus triplication causes Parkinson's disease, Science 302 (2003) 841.
- [67] W. Zhou, J. Schaack, W.M. Zawada, C.R. Freed, Overexpression of human alphasynuclein causes dopamine neuron death in primary human mesencephalic culture, Brain Res. 926 (2002) 42–50.
- [68] Y. Kawamoto, I. Akiguchi, S. Nakamura, Y. Honjyo, H. Shibasaki, et al., 14–3-3 proteins in Lewy bodies in Parkinson disease and diffuse Lewy body disease brains, J. Neuropathol. Exp. Neurol. 61 (2002) 245–253.
- [69] T.A. Yacoubian, S.R. Slone, A.J. Harrington, et al., Differential neuroprotective effects of 14-3-3 proteins in models of Parkinson's disease, Cell Death Dis. 1 (2010) e2.
- [70] J.S. Wu, W.M. Cheung, Y.S. Tsai, et al., Ligand-activated peroxisome proliferator-activated receptor-gamma protects against ischemic cerebral infarction and neuronal apoptosis by 14-3-3epsilon upregulation, Circulation 119 (2009) 1124–1134.
- [71] N.J. Haughey, A. Nath, S.L. Chan, A.C. Borchard, M.S. Rao, et al., Disruption of neurogenesis by amyloid beta-peptide, and perturbed neural progenitor cell homeostasis, in models of Alzheimer's disease, J. Neurochem. 83 (2002) 1509– 1524.
- [72] R. Killick, M. Niklison-Chirou, M.R. Tomasini, et al., P73: a multifunctional protein in neurobiology, Mol. Neurobiol. 43 (2011) 139–146.
- [73] A. Rufini, M. Agostini, F. Grespi, et al., p73 in Cancer Genes, Cancer. 2 (2011) 491–502.
- [74] C. Wilson, S. Henry, M.A. Smith, R. Bowser, The p53 homologue p73 accumulates in the nucleus and localizes to neurites and neurofibrillary tangles in Alzheimer disease brain, Neuropathol. Appl. Neurobiol. 30 (2004) 19–29.
- [75] R. Scacchi, G. Gambina, G. Moretto, R.M. Corbo, Association study between P53 and P73 gene polymorphisms and the sporadic late-onset form of Alzheimer's disease, J. Neural. Transm. 116 (2009) 1179–1184.
- [76] C. Hooper, R. Killick, M. Tavassoli, G. Melino, S. Lovestone, TAp73alpha induces tau phosphorylation in HEK293a cells via a transcription-dependent mechanism, Neurosci Lett. 401 (2006) 30–34.
- [77] M.K. Wetzel, S. Naska, C.L. Laliberté, et al., p73 regulates neurodegeneration and phospho-tau accumulation during aging and Alzheimer's disease, Neuron 11 (2008) 708–721.
- [78] M. Agostini, P. Tucci, H. Chen, et al., p73 regulates maintenance of neural stem cell, Biochem. Biophys. Res. Commun. 403 (2010) 13–17.
- [79] F. Talos, A. Abraham, A.V. Vaseva, et al., p73 Is an essential regulator of neural stem cell maintenance in embryonal and adult CNS neurogenesis, Cell Death Differ. 17 (2010) 1816–1829.
- [80] M. Fujitani, G.I. Cancino, C.B. Dugani, et al., TAp73 acts via the bHLH Hey2 to promote long-term maintenance of neural precursors, Curr. Biol. 20 (2010) 2058–2065.
- [81] L. González-Cano, M. Herreros-Villanueva, R. Fernandez-Alonso, et al., p73 deficiency results in impaired self renewal and premature neuronal differentiation of mouse neural progenitors independently of p53. Cell Death Dis. 1 (2010) e109.
- [82] J.J. Cohen, Programmed cell death in the immune system, Adv. Immunol. 50 (1991) 55–85.
- [83] J. Cheng, T. Zhou, C. Liu, et al., Protection from Fas-mediated apoptosis by a soluble form of the Fas molecule, Science 263 (1994) 1759–1762.
- [84] A. Strasser, P.J. Jost, S. Nagata, The many roles of FAS receptor signaling in the immune system, Immunity 30 (2009) 180–192.
- [85] P.H. Krammer, CD95's deadly mission in the immune system, Nature 407 (2000) 789–795.
- [86] D. Tischner, C. Woess, E. Ottina. A. Villunger, Bcl-2-regulated cell death signalling in the prevention of autoimmunity, Cell Death Dis. 1 (2010) e48.
- [87] R.S. Maag, M. Mancini, A. Rosen, C.E. Machamer, Caspase-resistant golgin-160 disrupts apoptosis induced by secretory pathway stress and ligation of death receptors, Mol. Biol. Cell 16 (2005) 3019–3027.
- [88] J.P.X. Cheng, V.M.S. Betin, H. Weir, et al., Caspase cleavage of the Golgi stacking factor GRASP65 is required for Fas/CD95-mediated apoptosis, Cell Death Dis. 1 (2010) e82.
- [89] M. Rothen, S. Gratzl, H.H. Hirsch, C. Moroni, Apoptosis in HIV-infected individuals is an early marker occurring independently of high viremia, AIDS Res. Hum. Retroviruses 13 (1997) 771–779.
- [90] N.W. Cummins, A.D. Badley, Mechanisms of HIV-associated lymphocyte apoptosis, Cell Death Dis. 1 (2010) e99.