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Forcina A. Pre-transplant multidrug-resistant Gram-negative colonization in HSCT

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| 2 3 | Clinical impact of pre-transplant multidrug-resistant Gram-negative colonization in autologous and allogeneic hematopoietic stem cell transplantation. |
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- 1 **Conflict of interest:** The authors declare no competing financial interests.
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3 Highlights

- Mortality due to multidrug-resistant Gram strains (MDR-GNB) is high
 - Pre-transplant MDR-GNB carriers of are often excluded from HSCT
- A pre-transplant MDR-GNB colonization did not impact on clinical outcomes
- Early targeted therapy in carriers may improve OS and reduce mortality
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9 Abstract

Multidrug-resistant Gram-negative bacteria (MDR-GNB) are an emerging cause of 10 morbidity and mortality after hematopoietic stem cell transplantation (HSCT). Three-11 hundred-forty-eight consecutive patients transplanted at our hospital from July 2012 to 12 January 2016 were screened for a pre-transplant MDR-GNB colonization and evaluated 13 for clinical outcomes. A pre-transplant MDR-GNB colonization was found in 16.9% of allo-14 HSCT and in 9.6% of auto-HSCT recipients. Both in auto- and in allo-HSCT, carriers of a 15 MDR-GNB showed no significant differences in overall survival (OS), transplant-related 16 mortality (TRM) nor infection-related mortality (IRM) compared to non-carriers. OS at two 17 years for carriers compared to non-carriers was 85% vs 81% (P=0.262) in auto-HSCT and 18 50% vs 43% (P=0.091) in allo-HSCT.TRM at two years was 14% vs 5%; (P=0.405) in 19 auto-HSCT and 31% vs 25% (P=0.301) in allo-HSCT. IRM at two years was 14% vs 2% 20 (P=0.142) in auto-HSCT and 23% vs 14% (P=0.304) in allo-HSCT. In multivariate analysis, 21 only grade III-IV acute graft-versus-host disease (GVHD) was an independent factor for 22 reduced OS (P<0.001) and increased TRM (P<0.001) and IRM (P<0.001). During the first 23 year after transplant, we collected 73 GNB bloodstream infectious episodes (BSI) in 54 24 patients, 42.4% of which sustained by a MDR-GNB. Rectal swabs positivity associated 25 with the pathogen causing subsequent MDR-GNB BSI in 13/31 (41.9%). Overall, OS at 26 four months from MDR-GNB BSI onset was of 67.9%, with a 14-day attributed mortality of 27

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1 12.9%, not being significantly different between carriers and non-carriers (P=0.207). We 2 conclude that in this extended single-center experience, a pre-transplant MDR-GNB 3 colonization did not significantly influence OS, TRM and IRM both in auto- and allo-HSCT 4 settings and that MDR-GNB attributed mortality can be controlled in carriers when an early 5 pre-emptive antimicrobial therapy is started in case of neutropenic fever.

6

7 Introduction

Infections caused by multidrug-resistant Gram-negative bacteria (MDR-GNB) are a 8 public health challenge worldwide¹⁻⁴. An increasing trend in MDR-GNB infections in 9 hematological malignancies and in hematopoietic stem cell transplantation (HSCT) has 10 been recently reported ^{2,5-7}. The emergence of MDR-GNB is a potential limiting factor in 11 HSCT, facing important clinical issues such as the eligibility to transplant for pre-transplant 12 carriers of a MDR-GNB, the risk of local infection outbreaks and the unacceptably high 13 related mortality. Moreover, the susceptibility pattern to antibiotics and global bacterial 14 epidemiology are changing, as well as the transplanted population and the availability of 15 new drugs^{1, 5, 8-11}. 16

In a recent large prospective Italian multicenter study performed on behalf of Gruppo Italiano Trapianto Midollo Osseo (GITMO) in the HSCT population, the overall survival (OS) at 4 months in patients developing pre-engraftment *Pseudomonas aeruginosa* and carbapenem-resistant (CR) *Klebsiella pneumoniae* (CR-Kp) bloodstream infectious episodes (BSI) was 25% and 40%, respectively, with pre-transplant carriers having a probability of approximately 30% of development a subsequent colonizingpathogen related infection¹². According to some recent studies, patients developing an

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1 MDR-GNB infection after allo-HSCT have a poor outcome, so that their eligibility to 2 allogeneic procedure is generally carefully evaluated for the risk-to-benefit ratio¹³.

The aim of this study was to analyze the impact of a pre-transplant MDR-GNB colonization on overall survival (OS), transplant-related mortality (TRM) and infectionrelated mortality (IRM) in both autologous and allogeneic HSCT population and to analyze the epidemiology of Gram-negative BSI in MDR-GNB carriers and non-carrier patients.

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8 Patients and methods

9 Study population

We analyzed data collected from 348 consecutive adult patients undergoing autologous (94/348, 27%) and allogeneic transplant (254/348, 72.9%), hospitalized in the Hematology and Bone Marrow Transplant Unit at IRCCS San Raffaele Scientific Institute Milan, from July 2012 to January 2016 (43 months). Patients undergoing a second or subsequent transplant during the study period were excluded. The median follow-up for survivors was of 602 days (range 4-1375). Gram-negative BSI episodes occurring during the first year after transplant were also recorded.

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18 Microbiological definitions

MDR-GNB colonization was defined as the isolation of the microorganism from rectal swabs, a non-sterile body site, in absence of clinical signs or symptoms of active infection. A GNB BSI was defined as the isolation of the microorganism from one or more blood cultures; a recurrent GBN BSI was defined as the isolation of the microorganism in a patient who previously experienced a GNB BSI and cleared it with documented negative

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blood cultures after the first episode. Multi-drug resistance was defined as acquired non susceptibility to at least 1 agent in 3 or more antimicrobial categories¹⁴.

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4 Microbiological surveillance and transplant procedures

5 All candidate patients to a first HSCT were screened with a rectal swab before 6 transplant to detect ESBL-producers and carbapenem-resistant Enterobacteriaceae and 7 non-fermenting GNB. Cultures were performed with MacConkey agar plates. Selective 8 chromogenic media plates were not routinely used, so our screening policy did not cover 9 vancomycin-resistant *Enterococcus faecium* (VRE) colonization.

Rectal swabs were then regularly repeated with weekly frequency until discharge. 10 Patients were isolated in single rooms with high-efficiency particulate air filtration from the 11 beginning of conditioning. When MDR-GNB colonization was detected, infection-control 12 strategies were adopted as previously described¹⁵. Transplants were conducted according 13 to local procedures and treatment protocols. All patients received primary antibacterial 14 prophylaxis (usually with levofloxacin) until engraftment. In case of fever, standard blood 15 cultures were performed with at least two sets of aerobic and anaerobic bottles collected 16 from central lines and peripheral blood during the acute febrile episode. 17

Empirical antibiotic therapy was usually administered intravenously following our institutional antimicrobial guidelines. At the early onset of neutropenic fever in MDR-GNB carriers, the de-escalation approach was preferred, starting promptly a combination therapy based on the results of the antimicrobial susceptibility test of the rectal swab isolate. In all cases of GNB positivity on blood cultures patients were treated accordingly as most appropriate.

24

25 Statistical methods

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Continuous variables were summarized according to median and range. Categorical 1 2 variables were summarized according to their frequency distributions. Comparison of medians of continuous variables between groups was made according to Mann-Whitney U 3 test. Comparison of proportions of categorical variables between groups was made 4 according to Fischer exact test. The probability of OS was estimated using the Kaplan-5 Meyer estimator. Cumulative incidences (CI) were estimated for graft-versus-host disease 6 7 (GVHD), TRM and IRM to accommodate competing risks. Relapse or progression of the original disease were competing risks for TRM, death without relapse was a competing 8 risk for relapse, death from any cause was a competing risk for engraftment. 9

10

Recurrence of original disease, GVHD and death from any other cause were 11 competing risks for IRM. Both relapse or progression and death from any causes were 12 competing risks for GVHD. Univariate comparisons of survival curves were made using the 13 log-rank test, while the Gray's test was used for univariate comparisons of CI functions. 14 The association between time to death and colonization status and other relevant 15 variables was evaluated in a multivariate analysis using Cox's proportional hazards 16 regression analysis. Potential covariates included in the model were: pre-transplant gut 17 18 colonization, patient age (continuous variable), type of diagnosis (myeloid vs lymphoid origin), refined disease risk index (DRI) at transplantation, donor type, Sorror Comorbidity 19 index score, GVHD prophylaxis backbone, development of grade III-IV acute GVHD (time 20 dependent variable). Affirmation of the proportional hazard assumption was met for all 21 variables. All tests were two-sided. The type I error rate was fixed at 0.05 for determination 22 of factors associated with time to event. Statistical analyses were performed with the 23 SPSS (SPSS Inc./IBM, Armonk, NY, USA) and R (R Development Core Team, Vienna) 24 software packages. 25

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1 Results

Patients' characteristics are summarized in Table 1. We found no significant
differences between carriers and non-carriers regarding age, sex, Sorror HCT Comorbidity
Index, CMV host/donor serostatus matching distribution, disease type, disease status at
transplant or donor type as reported in Table 2.

6

7 OS, TRM and IRM in auto-HSCT population

Only nine out of 94 auto-HSCT recipients (9.6%) were found to be colonized by a 8 MDR-GNB before transplant. Distribution of MDR-GNB genus and species and resistance 9 patterns in pre-transplant carriers of auto-HSCT and allo-HSCT recipients are summarized 10 in **Table 3**. The 2-yrs OS in auto-HSCT recipients was 82%, overall. We did not find any 11 significant differences in 2-yrs OS (85% vs 81%; P=0.262), 2-yrs TRM (14% vs 5%; 12 P=0.405) and 2-yrs IRM (14% vs 2%; P=0.142) comparing MDR-GNB carriers to non-13 carriers (Figure 1). Multivariate analysis was not performed in this setting due to the low 14 numbers of patients. 15

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17 OS, TRM, IRM and GVHD in allo-HSCT population

Forty-three out of 254 (16.9%) patients were found to be colonized by a MDR-GNB 18 before allo-HSCT. The 2-yrs OS after allo-HSCT was 49%. We did not find any significant 19 difference in 2-yrs OS when pre-transplant MDR-GNB carriers to non-carriers were 20 compared (50% vs 43%; P=0.091). Interestingly, in multivariate analysis, only the time-21 dependent development of acute GVHD (aGVHD) of grade \geq III (P<0.0001) and a refined 22 disease-risk index (DRI) high or very high (P<0.0001) were significantly associated with a 23 reduced OS. In particular, a pre-transplant MDR-GNB colonization was not associated with 24 a reduced OS (HR=1.417; P=0.129). Transplant-related mortality at two years was 25% 25 26 overall, and comparable between pre-transplant MDR-GNB carriers and non-carriers (31%)

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vs 25%; P=0.301). In multivariate analysis, only the development of aGVHD of grade ≥III 1 2 (HR=7.933; P<0.001) and age > 53 years (HR=1.029; P=0.005) were independent risk factors for increased TRM. As for OS, also for TRM a pre-transplant colonization status 3 was not significantly associated with an increased mortality (HR=1.428; P=0.268). 4 Infection-related mortality at two years was 16% in the allo-HSCT population. A pre-5 transplant MDR-GNB colonization was not associated with a significant increase of IRM. 6 being of 23% in carriers and 14% in non-carriers (P=0.304) (Figure 2). Multivariate 7 analysis for IRM confirmed again that only the development of aGVHD of grade ≥III 8 (HR=18.690; P<0.0001) and the presence of a myeloid-lineage disease (HR=1.450; 9 P=0.034), were the independent risk factors for an increased IRM. A pre-transplant MDR-10 GNB colonization status was not significantly associated with an increased IRM 11 (HR=1.690; P=0.215). Incidence of aGVHD of grade \geq III by day 100 was 13% in the entire 12 allo-HSCT population, with no significant differences according to the pre-transplant 13 colonization status, being 21% in carriers and 12% in non-carriers, respectively (P=0.152). 14 Moreover, multivariate analysis showed that a pre-transplant MDR-GNB colonization did 15 not influence the likelihood of developing severe aGVHD (HR=1.948; P=0.069). Results of 16 multivariate analysis are summarized in Table 4. 17

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19 Multidrug-resistant Gram-negative bloodstream infections in HSCT

Patients were observed for one year after transplant and all the BSI caused by Gram-negative bacteria were recorded during this time. Overall, we registered 73 GNB-BSI in 54 patients. Fifteen patients experienced recurrent GNB-BSI either from the same or from a different GNB. Median time from transplant to GNB-BSI onset was 16 days (range 0-308). Among non-colonized patients (n=296), 41/296 developed a GNB-BSI: of these, 30/296 (10%) patients developed a non-MDR GNB-BSI (in details: 16 BSI were sustained by multi-sensible E. Coli, 8 by Pseudomonas spp, 2 by Klebsiella spp, 1 by

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Acinetobacter baumanii, 3 by other GNB), while 11/296 (4%) developed a MDR-GNB BSI
(in details: 6 BSI were sustained by ESBL-producing E. Coli, 3 by ESBL-producing Kp, 1
by CR-Enterobacter cloacae, 1 by CR-Kp).

Among pre-transplant colonized patients (n=52), 13/52 developed a GNB-BSI: of 4 these, 4/52 (8%) patients developed a non-MDR GNB-BSI (in details: 3 BSI were 5 sustained by multi-sensible E. Coli and 1 by multi-sensible Pseudomonas aeruginosa), 6 while 9/52 (17%) patients had a MDR GNB-BSI (in details: BSI were sustained by CR-Kp, 7 2 by Stenotrophomonas maltophilia, 1 by ESBL-producing E. Coli, 1 by CR- Pseudomonas 8 aeruginosa). According to these distribution, pre-transplant carriers had a significant 9 increased number of MDR-GNB BSI compared to non-carriers (9/52 colonized vs 11/296 10 non-colonized, P=0.0009). 11

Twenty-seven of 31 (87%) MDR-GNB BSI occurred in allo-HSCT, while only 4/31 (12.9%) 12 in auto-HSCT. In the entire population, MDR GNB BSI were sustained by carbapenem-13 resistant *Pseudomonas aeruginosa* (5/15) or CR-Kp (10/15), with an attributable mortality 14 of 20% (only three patients died at day six, nine and ten, respectively from BSI onset). 15 Details of microbial isolates and distribution of MDR-GNB BSI according to transplant type 16 are summarized in Figure 3. Thirteen out of 31 (41.9%) of MDR-GNB BSI episodes 17 occurred in acute leukemia patients and 70% had active diseases at the onset of sepsis. 18 Median neutrophil count at MDR-GNB BSI onset was 0x10⁹/L (range 0-6.2). 19

Rectal swabs positivity associated with the pathogen causing subsequent MDR-GNB BSI in 13/31 (41.9%). In particular, 9/13 (69.2%) MDR-GNB BSI were sustained by CR-Kp. Overall, OS at four months from MDR-GNB BSI onset was of 67.9%, with a 14-day crude mortality from BSI onset of 12.9% (4/31). For CR-Kp BSI, OS at four months was of 63%, with a 14-day attributed mortality of 20% (2/10). Interestingly, 14-day attributed mortality after MDR-GNB BSI onset was not significantly different between carriers (n=13) and non-carriers (n=18) (P=0.207), despite in non-carriers, BSI were sustained by ESBL⁺

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E. Coli or ESBL⁺ Kp (9/18) in 50% of episodes, while in carriers up to 76.9% (10/13) MDRGNB BSI were caused by CR-Kp (9/10) or CR-*Pseudomonas aeruginosa* (1/10). Details of
first and second-line antimicrobial treatments in carriers and non-carriers are detailed in **Table 5**.

5

6 **Discussion**

In this single-center study we analyzed the clinical outcome of 348 patients
 undergoing autologous or allogeneic HSCT from July 2012 to January 2016 according to
 their pre-transplant multidrug-resistant Gram-negative bacteria (MDR-GNB) colonization
 status, and collected all Gram-negative bloodstream infectious episodes (GNB-BSI) during
 the first year after transplant.

Although recent data suggest an important role also of Gram-positive bacteria such as VRE or methicillin-resistant *Staphylococcus aureus* in allo-HSCT¹⁶, their incidence in our cohort was very low (<1%), making the epidemiological scenario largely dominated by Gram-negative rods, particularly MDR-GNB, the latter becoming the main focus of our study.

In patients receiving auto-HSCT, a rapid immune recovery and lack of 17 pharmacological immunosuppression make TRM and IRM generally low¹⁷. In the auto-18 HSCT cohort, transplant outcomes did not significantly differ according to the pre-19 transplant MDR-GNB colonization, at variance to what has been recently reported by 20 Scheich et al¹⁸ in the same setting. Noticeably, MDR-GNB pre-transplant carriers were 21 only 9.6% of the transplanted population, a proportion significantly lower than that of allo-22 transplanted patients (16.9%). The reason for this may be due to the different 23 characteristics of the two populations. Autologous HSCT is usually reserved to patients 24 with MM and NHL, who receive the majority of chemotherapy cycles in an outpatient 25

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regimen and are less exposed to prolonged hospitalization periods and broad-spectrum
 antibiotics.

During the past decades, allo-HSCT transplant activity has increased exponentially¹⁹ 3 compared to auto-HSCT, and is now a potentially curative option also for older patients 4 without expecting a substantial increase in TRM even when HLA-haploidentical donors are 5 used²⁰. However, in allo-HSCT, IRM is a cause of major concerns, especially in the recent 6 era of emerging MDR bacteria²¹. In our study, despite some high-risk factors both for TRM 7 and for disease relapse (e.g. prevalence of AML patients and of patients with advanced 8 diseases, use of HLA-haploidentical donors), the 2-yrs TRM in allo-HSCT was 25%, 9 comparable to that reported by other Centers using standard allo-HSCT procedures^{22,23} 10 and 2-yrs IRM was 16%, without a substantial increase in MDR-GNB pre-transplant 11 carriers compared to non-carriers and again not significant different from those reported in 12 literature²⁴. Nonetheless, the incidence of post-transplant MDR-GNB BSI in carriers was 13 significantly higher than in carriers (17% vs 4%, P=0.0009), suggesting that the acquisition 14 of a MDR GNB colonization may be a surrogate marker more advanced diseases. 15

In our experience, 42.4% of GNB-BSI were due to a MDR strain, with carbapenem-16 resistant Klebsiella pneumoniae (CR-Kp) and ESBL⁺ Escherichia coli being the most 17 frequent pathogens. The majority of patients with a MDR-GNB BSI had acute leukemia 18 (50%) and grade III-IV neutropenia (74%). Recently, it has been reported that MDR 19 bacterial colonization, evaluated through rectal swabs, decreases OS by increasing TRM 20 and the probability of infectious complications after allo-HSCT, despite the use of 21 antibiotics targeting the gut-colonizing pathogens²⁵. On the other hand, Heidenreich et al. 22 in a retrospective single-center study demonstrated that MDR-GNB carriers other than 23 from *Pseudomonas aeruginosa* (evaluated also in urinary samples or other body regions) 24 and treated with imipenem/cilastatin as first line therapy, did not a negative impact on 25 TRM and OS²⁶. 26

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For these reasons, several transplant centers still consider MDR-GNB colonization as 1 a relative contraindication to transplant, especially in allo-HSCT, due to the high-mortality 2 rate in case of neutropenic MDR-GNB BSI and the likelihood of pre-engraftment BSI 3 development in carriers. At our Institution, when combinations of pre-emptive measures 4 were applied, MDR GNB BS could be successfully managed and TRM and OS in carriers 5 were not significantly increased compared to non-carriers. However, it should be noticed 6 that MDR-GNB BSI often arose in patients showing concomitant dismal features (i.e. 7 active disease (88%), severe aGVHD (60%), no neutrophil engraftment (20%). In this 8 study, we showed that, both in auto-HSCT and allo-HSCT, a pre-transplant colonization is 9 not per se significantly associated with long-term reduced OS, TRM or IRM. Moreover, the 10 14-day attributed mortality after MDR-GNB BSI onset was not significantly different 11 between carriers and non-carriers (P=0.207), despite in non-carriers MDR-GNB BSI were 12 mostly sustained by ESBL⁺ E. Coli or ESBL⁺ Kp, while in carriers up to 76.9% MDR-GNB 13 BSI were caused by CR-Kp or CR-Pseudomonas aeruginosa, which are known from 14 previous studies to be associated with higher mortality (ranging from 50-70%)¹² compared 15 to ESBL⁺ producing pathogens. This lack of a significant mortality difference may be 16 related to an intensive Institutional policy of early de-escalation approach of antimicrobials 17 in carriers, with the initiation of a targeted first-line therapy at the onset of neutropenic 18 fever, whenever possible, in order control the infectious mortality due to MDR pathogens. 19

Therefore, we think that in this cohort, the adoption of pre-transplant active surveillance and the early initiation of targeted combined antimicrobial therapy in MDR-GNB carriers have successfully impacted on clinical outcomes, thus making OS, TRM and IRM not significantly different between carriers and non-carriers, while the onset of severe GVHD remains the major risk factor for a detrimental outcome.

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26 Conflict of interests

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- 1 The authors declare no competing financial interests.
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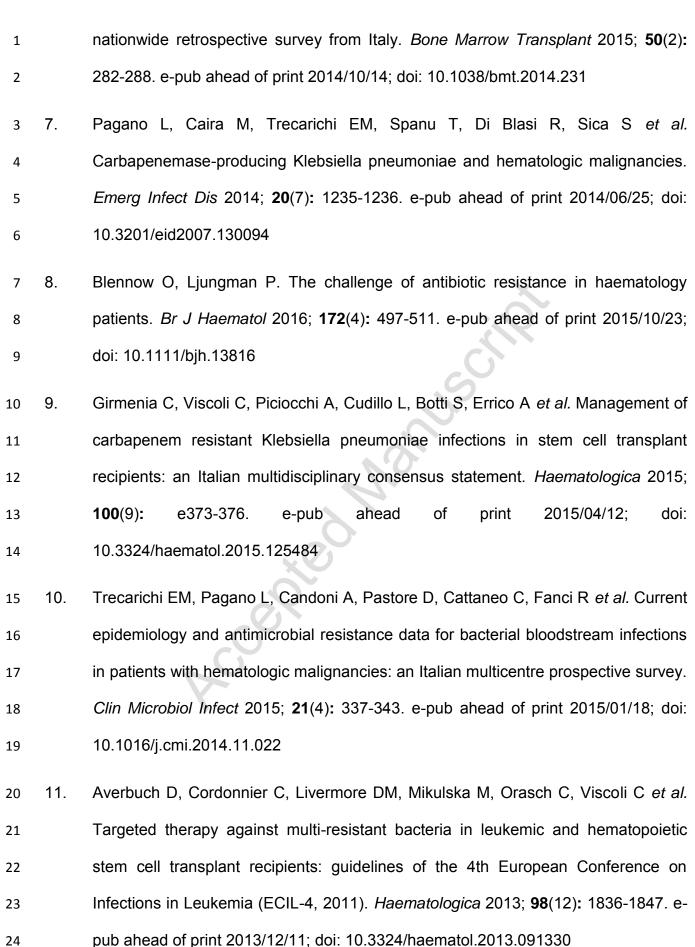
9 Author contribution

- 10 Conception and design: Alessandra Forcina, Francesca Lorentino, Vincenzo Marasco
- 11 and Fabio Ciceri
- 12 **Provision of study material or patients:** All authors
- 13 Collection and assembly of data: Alessandra Forcina, Vincenzo Marasco, Francesca
- 14 Lorentino, Magda Marcatti
- 15 Data analysis and interpretation: All authors
- 16 Manuscript writing: All authors
- 17 Final approval of manuscript: All authors

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Forcina A. Pre-transplant multidrug-resistant Gram-negative colonization in HSCT

Figure 1. Autologous HSCT. (A) Kaplan-Meier curves of the estimated 2-yrs OS for pretransplant MDR-GNB carriers (dotted line) compared non-carriers (black line); (B) Kaplan-Meier curves of the estimated 2-yrs TRM for pre-transplant MDR-GNB carriers (dotted line) compared non-carriers (black line); (C) Kaplan-Meier curves of the estimated 2-yrs IRM for pre-transplant MDR-GNB carriers (dotted line) compared non-carriers (black line).

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Figure 2. Allogeneic HSCT. (A) Kaplan-Meier curves of the estimated 2-yrs OS for pretransplant MDR-GNB carriers (dotted line) compared non-carriers (black line); (B) KaplanMeier curves of the estimated 2-yrs TRM for pre-transplant MDR-GNB carriers (dotted
line) compared non-carriers (black line); (C) Kaplan-Meier curves of the estimated 2-yrs
IRM for pre-transplant MDR-GNB carriers (dotted line) compared non-carriers (black line).
(D) Cumulative incidence of 100-day acute GVHD for pre-transplant MDR-GNB carriers
(dotted line) compared non-carriers (black line).

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Figure 3. Distribution of MDR-GNB BSI in auto-HSCT (n=4) and in allo-HSCT (n=27) patients according to microbiological etiology. ESBL⁺ EC = extended-spectrum beta lactamase producing *E. Coli*; CR-Kp = Carbapenem-resistant *Klebsiella pneumoniae;* CR-PA = carbapenem-resistant *Pseudomonas aeruginosa*; STN = *Stenotrophomonas maltophilia*. Other MDR-GNB included: 1 CR-*Enterobacter cloacae*; 1 *Ochrobactrum antrhopi*; 1 *Sphingomonas paucimobilis*.

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Forcina A. Pre-transplant multidrug-resistant Gram-negative colonization in HSCT

1 Table 1. Patients characteristics (n=348)

| Characteristic | Allo-HSCT (n=254) | Auto-HSCT (n=94) |
|---|-------------------------|---|
| Age, median years (range) | 53 (16-77) | 54 (19-78) |
| Male, sex n (%) | 154 (60.6) | 58 (61.7) |
| Underlying disease: n. (%) | | |
| Acute myeloid leukemia | 138 (54.3) | 6 (6.3) |
| Acute lymphoid leukemia | 13 (5.1) | 0 (0) |
| Myelodisplastic syndromes | 19 (7.4) | 1 (1) |
| Chronic myeloproliferative | 9 (3.5) | 1 (1) |
| Non Hodgkin's lymphoma | 27 (10.6) | 35 (37.2) |
| Hodgkin's lymphoma | 16 (6.2) | 7 (7.4) |
| Multiple myeloma | 25 (9.8) | 35 (37.2) |
| Other diseases | 7 (2.7) | 9 (9.5) |
| Sorror HCT-CI, median (range) | 2 (0-9) | , , , , |
| Donor type: n. (%) | | · |
| HLA-Identical | 43 (16.9) | / |
| Matched Unrelated | 86 (33.8) | , , |
| HLA-Haploidentical | 120 (47.2) | , |
| Cord blood | 5 (1.9) | , , |
| CMV host/donor serostatus | | , |
| Negative/Negative | 26 (10.2) | / |
| Positive/Positive | 149 (58.6) | / |
| Positive/Negative | 12 (4.7) | / |
| Negative/Positive | 66 (25.9) | / |
| Disease Risk Index | | , |
| Very High | 18 (7) | / |
| High | 85 (33.4) | / |
| Low-intermediate | 144 (56.6) | / |
| Not applicable | 7 (2.7) | / |
| Pretransplantation conditioning intensity: n, (%) | , (2.7) | 7 |
| Myeloablative | 165 (64.9) | 68 (72.3) |
| Reduced intensity | 89 (35) | 26 (27.6) |
| Phase of the underlying disease at transplant: n. | 00 (00) | 20 (27.0) |
| (%) | 110 (43.3) | 42 (44.7) |
| Malignancies in complete remission | 113 (44.4) | 46 (48.9) |
| Malignancies not in complete remission/active | 31 (12.2) | 6 (6.4) |
| Non-malignant stable/ chronic diseases | 51 (12.2) | 0 (0.4) |
| T-cell depletion | | |
| No | 23 (9) | 1 |
| ATG | 96 (37.8) | / |
| PT-Cy | 90 (37.8) 121 (47.6) | / |
| • | • • | / |
| ATG + PT-Cy Pro_transplant MDP_GNP_carriers: n_(%) | 14 (5.5) 42 (16 9) | |
| Pre-transplant MDR-GNB carriers: n, (%) | 43 (16.9) | 9 (9.6) |

2 Abbreviations: ATG = antilymphocyte globulin; PTCy= post-transplant Cyclophosphamide; MDR-GNB= multi-drug

3 resistant Gram-negative bacteria.

Forcina A. Pre-transplant multidrug-resistant Gram-negative colonization in HSCT

Table 2. Patients characteristics according to the colonization status in allo-HSCT and in auto-HSCT

| | Allo-HSCT Auto-HSCT | | | | | |
|---------------------------------|---------------------|-----------------------------|---------|-------------------|------------------------|---------|
| Characteristic | Carriers (n=43) | Non- carriers (n=211) | P value | Carriers (n=9) | Non-carriers (n=85) | P value |
| Age, median | 47 | 53 | 0.315 | 61 | 53 | 0.148 |
| (range) | (21-77) | (16-77) | | (31-71) | (18-77) | |
| Sorror HCT-CI, median | 2 (0-9) | 2 (0-8) | 0.166 | - | - | - |
| (range) | | | | | | |
| Sex, Male | 26 | 128 | 1 | 6 | 52 | 1 |
| Underlying disease | | | 0.058 | | | 0.45 |
| Acute lymphoid leukemia | 1 | 12 | | - | - | |
| Acute myeloid leukemia | 31 | 107 | | 1 | 5 | |
| Myelodisplastic | 0 | 19 | | - | 1 | |
| syndromes | 1 | 8 | | - | 1 | |
| Chronic | 1 | 24 | | 2 | 33 | |
| myeloproliferative | 3 | 13 | | <u>.</u> O` | 7 | |
| Multiple myeloma | 6 | 21 | | 6 | 29 | |
| Hodgkin's lymphoma | - | - | | - | 9 | |
| Non Hodgkin's lymphoma other | | | \sim | | | |
| Donor type | | | 0.725 | - | _ | - |
| HLA-Identical | 9 | 34 | 0,720 | | | |
| Matched Unrelated | 13 | 73 | | | | |
| HLA-Haploidentical | 21 | 99 | | | | |
| Cord blood | 0 | 5 | | | | |
| Disease Risk Index | | | 0.749 | - | _ | - |
| Low-intermediate | 24 | 120 | | | | |
| High | 15 | 70 | | | | |
| Very high | 4 | 14 | | | | |
| Disease status at | 6 | | 0.078 | | | 1 |
| transplant | 1 | 30 | | 0 | 6 | |
| Upfront | 21 | 89 | | 4 | 38 | |
| Malignancies in CR | 21 | 98 | | 5 | 41 | |
| Malignancies not CR | | | | | | |
| CMV host/donor | | | 0.586 | - | - | - |
| serostatus | 3 | 23 | | | | |
| Negative/negative | 40 | 188 | | | | |
| Others | | | | | | |

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Forcina A. Pre-transplant multidrug-resistant Gram-negative colonization in HSCT

Table 3. Distribution of MDR-GNB genus and species and resistance patterns in pre-transplant 1

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| 2 | carriers in auto-HSCT and allo-HSCT | | | | | | |
|---|--|------------------|-----------------|--|--|--|--|
| | Pathogen and resistance pattern | Allo-HSCT (n=43) | Auto-HSCT (n=9) | | | | |
| | ESBL-producing Escherichia coli, total n (%) | 10 (23.2) | / | | | | |
| | CR Klebsiella pneumoniae, total n (%) | 16 (37.2) | 4 (44.4) | | | | |
| | Other CR Enterobacteriacee, total n (%) | 2 (4.6) | 1 (11.1) | | | | |
| | MDR Pseudomonas aeruginosa, total n (%) | 7 (16.2) | 1 (11.1) | | | | |
| | Stenotrophomonas maltophilia, total n (%) | 4 (9.3) | 1 (11.1) | | | | |
| | Other MDR-GNB, total n (%) | 4 (9.3) | 2 (22.2) | | | | |

3 Abbreviations: ESBL = extended-spectrum beta lactamase producing; MDR = multi-drug resistant; MDRO-GN= multi-

4 drug resistant Gram-negative organisms; CR: resistant to carbapenems, defined as a MIC for imipenem and/or 5 meropenem and/or ertapenem > 1 mg/L;

- 1 **Table 4.** Multivariate analysis for 2-yrs OS, 2-yrs TRM, 2-yrs IRM and 100-days severe acute GVHD in the
- 2 allo-HSCT population (n=254)

| OS | HR | (95 % CI) | P-value |
|-------------------------------|--------|----------------|----------|
| aGVHD ≥ grade III | 3.177 | (2.060-4.901) | < 0.0001 |
| DRI | | | |
| High vs low-intermediate | 2.661 | (1.781-3.977) | < 0.0001 |
| Very high vs low-intermediate | 5.361 | (2.928-9.816) | < 0.0001 |
| MDR-GNB carrier | 1.417 | (0.904-2.221) | 0.129 |
| TRM | HR | (95 % CI) | P-value |
| aGVHD ≥ grade III | 7.933 | (4.666-13.487) | < 0.0001 |
| Disease type | | | 0.055 |
| Myeloid vs lymphoid | 0.580 | (0.333-1.011) | |
| MDR-GNB carrier | 1.428 | (0.760-2.682) | 0.268 |
| Age ≥ 53yrs | 1.029 | (1.009-1.050) | 0.005 |
| IRM | HR | (95 % CI) | P-value |
| MDR-GNB carrier | 1.690 | (0.737-0.737) | 0.215 |
| Age > 53 yrs | 1.022 | (0.998-1.046) | 0.074 |
| aGVHD ≥ grade III | 18.690 | (7.867-44.401) | < 0.0001 |
| Disease type | | | |
| Myeloid vs Lymphoid | 1.450 | (1.029-2.044) | 0.034 |
| aGVHD ≥ grade III | HR | (95 % CI) | P-value |
| MDR-GNB carrier | 1.864 | (0.914-3.800) | 0.087 |
| Donor type | | (0.086-1.574) | 0.178 |
| HLA-Haploidentical vs CBU | 0.368 | | |
| MUD vs CBU | 0.275 | (0.062-1.219) | 0.089 |
| HLA-identical vs CBU | 0.034 | (0.003-0.382) | 0.006 |

Forcina A. Pre-transplant multidrug-resistant Gram-negative colonization in HSCT

- 1 Abbreviations: OS= overall survival; TRM= transplant-related mortality; IRM= infection-related mortality; MDR-GNB=
- 2 multi-drug resistant Gram-negative bacteria; aGVHD = acute graft versus host disease; DRI = disease risk index; MUD=
- 3 matched unrelated donor; CBU=cord blood unit
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- 1 **Table 5**. Details of microbiological isolates on blood cultures and antimicrobial therapy in pre-transplant
- 2 carriers and non-carriers

| Pt n° | Type of HSCT | Rectal swab isolate | BC isolate | I line therapy | II line therapy | Outcome | Cause of death |
|----------|--------------------|---------------------------|----------------------|----------------------------------|------------------------------|---------|-------------------|
| 1 | Allo | No | ESBL ⁺ EC | PIP/TAZ | AMIK | alive | |
| 2 | Allo | CR-KP | CR-KP | MER, TIGE, COL, CLINDA | | dead | PD |
| 3 | Allo | No | CR-KP | MER, GENTA, TIGE | COL | alive | |
| 4 | Allo | STN | STN | MER, VANCO | AMIK | dead | GVHD |
| 5 | Allo | No | ESBL ⁺ EC | PIP/TAZ | MER, TIGE, AMIK | alive | |
| 6 | Allo | No | ESBL ⁺ EC | PIP/TAZ | ВВК8 | alive | |
| 7 | Allo | No | ENTB | CIPRO | > | dead | PD |
| 8* | Allo | CR-KP | CR-KP | MER, TIGE, COL | ERT, GENTA | alive | |
| 8 | Allo | CR-KP | CR-KP | MER, TIGE, COL, ERT, GENTA | | dead | sepsis |
| 9 | Allo | ESBL ⁺ EC | ESBL ⁺ EC | MER, AMIK | | dead | sepsis |
| 10 | Allo | No | ESBL ⁺ EC | PIP/TAZ, METRO | MER | dead | PD |
| 11 | Allo | STN | STN | MER, LIN, AMIK | TIGE, TMP/SMZ, PIP/TAZ | dead | PD |
| 12* | Allo | CR-KP | CR-KP | AMIK, TIGE, GENTA, MER | ERT, COL | alive | |
| 12 | | | ESBL ⁺ KP | CEFT, MER | | alive | |
| 13* | Allo | No | MDR-PA | MER, AMIK, COL | | alive | |
| 13 | | | MDR-PA | CEF, CIPRO, AMIK | | dead | sepsis in PD |
| 14 | Allo | No | ESBL ⁺ EC | MER | | dead | GVHD |
| 15 | Allo | No | MDR-PA | LEVO | AMIK, IMIP | dead | GVHD |
| 16 | Allo | No | Sphingomonas | LEVO | CEFT, VANCO, | dead | sepsis |

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| | | | paucimobilis | | MER | | |
|-----|------|--------|-----------------------|---------------------------------|----------------|--------|-----------------|
| 17 | Allo | CR-KP | CR-KP | COL, GENTA, MER, LIN | TIGE | dead | PD |
| 18 | Allo | No | Ochrobactrum antrhopi | TIGE, MER | | alive | |
| 18 | Allo | No | STN | AMOX/CLAV | | alive | |
| 19 | Allo | No | MDR-PA | CEF | MER | dead | sepsis in PD |
| 20 | Allo | No | ESBL ⁺ KP | MER | DAPTO, AMIK | alive | |
| 21* | Allo | CR-KP | CR-KP | MER, ERT, GENTA | TIGE, COL, RIF | alive§ | |
| 21 | | | CR-KP | MER, ERT, GENTA, COL, RIF | il? | alive§ | |
| 21 | | | CR-KP | MER, ERT, GENTA, COL, RIF | 5 | dead | sepsis |
| 22 | Auto | MDR-PA | MDR-PA | PIP/TAZ, GENTA | | alive | |
| 23 | Auto | No | ESBL ⁺ KP | PIP/TAZ | AMIK, MER | dead | sepsis in PD |
| 24 | Auto | CR-KP | CR-KP | MER, TIG, GENTA, COL | | alive | |
| 25 | Auto | | ESBL ⁺ EC | LEVO, CEFT | MER | alive | |

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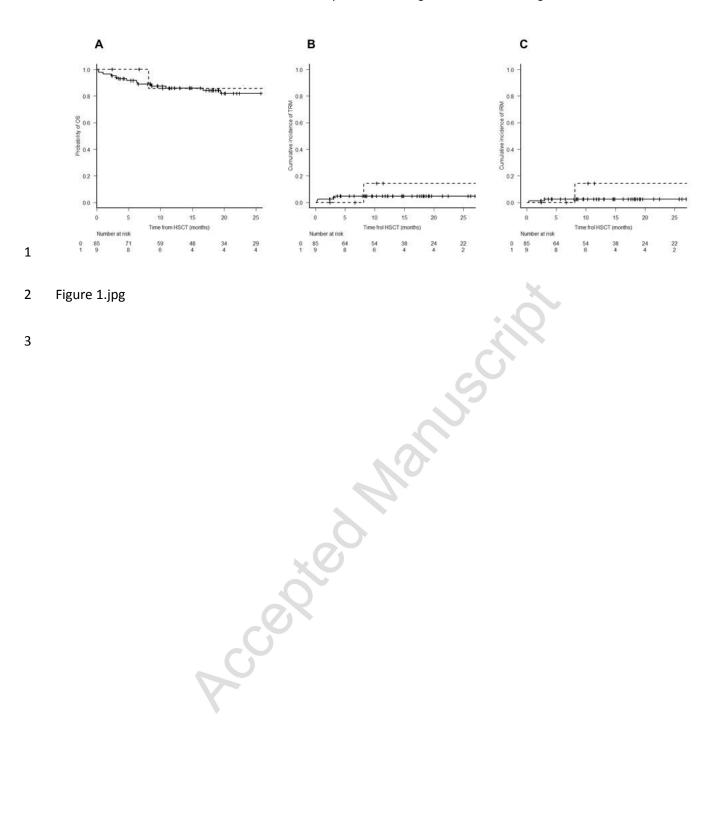
Abbreviations: BC = Blood culture; ESBL⁺ = Extended-spectrum beta-lactamase producing; CR = Carbapenem-resistant; MDR = multidrug-resistant; EC = *Escherichia coli*: KP=*Klebsiella pneumoniae*; STN = *Strenotrophomonas maltophilia*; ENTB =Enterobacteriaceae; PIP/TAZ = piperacillin/tazobactam; AMIK= amikacin; MER = meropenem, CLINDA = clindamycin, TIGE = tigecycline; COL = colistin; GENTA = gentamycin; VANCO = vancomycin; CIPRO= ciprofloxacin; ERT = ertapenem; METRO = metronidazole; LIN = linezolid; TMP/SMZ = trimethoprim/sulfamethoxazole; CEFT = ceftazidim; CEF = cefepim; LEVO = levofloxacin; IMIP = imipenem; AMOX/CLAV = amoxicillin/clavulanate; RIF =

7 rifampicin; PD = progression of disease; GVHD = graft-vs-host-disease.

8 *patients experiencing recurrent BSI.

9 § alive until subsequent BSI.

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