ELSEVIER

Contents lists available at ScienceDirect

Brain Stimulation

journal homepage: www.brainstimjrnl.com

Consensus Paper: Probing Homeostatic Plasticity of Human Cortex With Non-invasive Transcranial Brain Stimulation



BRAIN

Anke Karabanov^{a,*}, Ulf Ziemann^b, Masashi Hamada^c, Mark S. George^{d,e}, Angelo Quartarone^f, Joseph Classen^g, Marcello Massimini^h, John Rothwellⁱ, Hartwig Roman Siebner^{a,j}

^a Danish Research Center for Magnetic Resonance, Copenhagen University Hospital Hvidovre, Hvidovre, Denmark

^b Department of Neurology and Stroke, Hertie-Institute for Clinical Brain Research, University of Tübingen, Germany

^c National Hospital for Neurology & Neurosurgery, Queen Square, London, UK

^d Brain Stimulation Laboratory, Medical University of South Carolina, Charleston, SC, USA

^e Ralph H. Johnson VA Medical Center, Charleston, SC, USA

^fDepartment of Neuroscience, University of Messina, Italy

^g Department of Neurology, University Hospital Leipzig, Germany

^h Department of Clinical Sciences, University of Milan, Italy

Sobell Department of Motor Neuroscience and Movement Disorders, Institute of Neurology, University College of London, London, UK

^j Department of Neurology, Copenhagen University Hospital Bispebjerg, Copenhagen, Denmark

ABSTRACT

Homeostatic plasticity is thought to stabilize neural activity around a set point within a physiologically reasonable dynamic range. Over the last ten years, a wide range of non-invasive transcranial brain stimulation (NTBS) techniques have been used to probe homeostatic control of cortical plasticity in the intact human brain. Here, we review different NTBS approaches to study homeostatic plasticity on a systems level and relate the findings to both, physiological evidence from *in vitro* studies and to a theoretical framework of homeostatic function. We highlight differences between homeostatic and other non-homeostatic forms of plasticity and we examine the contribution of sleep in restoring synaptic homeostasis. Finally, we discuss the growing number of studies showing that abnormal homeostatic plasticity may be associated to a range of neuropsychiatric diseases.

© 2015 The Authors. Published by Elsevier Inc. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.0/).

Throughout life the brain flexibly and quickly adapts to environmental changes while at the same time maintaining a relatively stable equilibrium of neural activity over time. At the neural level, synapses can dynamically express lasting changes in synaptic efficacy, long-term potentiation (LTP) or long-term depression (LTD), in response to a change in presynaptic activity [1]. The threshold for induction of LTP and LTD is flexibly adjusted to the level of postsynaptic activity by homeostatic mechanisms [2]. These adjustments of plasticity prevent excessive expression of LTP or LTD and keep neural activity within a useful dynamic range [3,4].

E-mail address: ankenk@drcmr.dk (A. Karabanov).

In humans, a range of non-invasive transcranial brain stimulation (NTBS) techniques has been successfully used to induce cortical plasticity [5–8]. Research has mainly focused on the motor hand area (M1-Hand) and its fast-conducting descending projections to the contralateral hand because M1-Hand can be easily targeted with NTBS due to its relatively superficial position close to the surface of the convexity of the cerebral hemisphere. Moreover, NTBS-induced corticomotor plasticity can be readily probed by measuring the amplitude of motor evoked potentials (MEP) in contralateral hand muscles, although the mechanism of activating corticospinal neurons is complex and not yet fully understood [9]. Several NTBS protocols have been shown to be capable of inducing shifts in corticomotor excitability as indexed by changes in mean MEP amplitude. These changes can outlast the stimulation period for minutes to hours [10], yet both, the magnitude and direction of these excitability changes, display substantial inter-individual variability [11–17]. Depending on the direction of the amplitude changes, these lasting excitability changes have been labeled as

http://dx.doi.org/10.1016/j.brs.2015.06.017

1935-861X/© 2015 The Authors. Published by Elsevier Inc. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.0/).

DOI of original article: 10.1016/j.brs.2015.01.404, 10.1016/j.brs.2015.06.016. AK was supported by the Swedish Research Council (2011-38769-82837-71).

^{*} Corresponding author. Kettegaard Alle 30, 2650 Hvidovre, Denmark. Tel.: +45 27120129.

Definition Box

Metaplasticity: 'plasticity of synaptic plasticity'

Metaplasticity is a higher-order form of synaptic plasticity. The term was originally introduced by W.C. Abraham and M.F. Bear [27]. It refers to synaptic or cellular activity that primes the ability to induce subsequent synaptic plasticity, such as long-term potentiation (LTP) or depression (LTD). The priming event does not necessarily cause a change in the efficacy of normal synaptic transmission. Metaplasticity can be homeostatic or non-homeostatic.

Homeostatic plasticity: 'plasticity stabilizing synaptic plasticity'

The term homeostatic plasticity refers to a range of plasticity mechanisms that stabilize neuronal activity [24]. Homeostatic plasticity counteracts the destabilizing influence of synaptic plasticity and thus, stabilizes neural activity within a physiologically meaningful range. Homeostatic mechanisms can be metaplastic or non-metaplastic.

LTP-like or LTD-like effects [8]. In analogy to homeostatic metaplasticity at the neuronal level, it has been shown that the LTP- and LTD-like changes are subject to homeostatic control. Here, we review the use of NTBS to non-invasively investigate the homeostatic regulation of regional cortical excitability and relate this line of research to homeostatic plasticity described at the neuronal level in invasive non-human animal studies.

Basic principles of synaptic plasticity

The mammalian cortex expresses a wealth of functional and structural mechanisms to change its function in response to experience and use [18]. Functional mechanisms often involve the modification of existing synapses and multiple forms of synaptic plasticity have been demonstrated in vitro and in vivo in excitatory and inhibitory cortical synapses [19–21]. Synapses can strengthen (LTP) or weaken (LTD) their efficacy (i.e., synaptic strength) in response to increases or decreases in their activity and in accordance with Hebb's famous principle of cell assembly [22-25]. Synaptic plasticity is complemented by other forms of plasticity, including plasticity of intrinsic cellular excitability [2,26,27]. These functional mechanisms go hand in hand with structural plasticity, including the formation, removal, and remodeling of synapses and dendritic spines [28]. The abundance of plasticity mechanisms in the mammalian neocortex highlights the changeability of cortical neurons. A critical question is how these multiple processes are integrated at the level of a synapse, a single neuron, intracortical microcircuits, and interacting brain systems. The complexity of mechanisms causing synaptic and cellular plasticity renders it difficult to link plasticity-induced change at the regional or system level to specific synaptic or cellular mechanisms. Yet it is likely that plastic processes at the regional or systems level nevertheless follow the same general principles.

Synaptic plasticity provides a mechanism for learning and enables neurons to dynamically modulate their synaptic strength by relating it to other inputs the cell receives at the same time [29]. Synaptic plasticity provides an efficient positive feedback mechanism, which enforces (LTP) or weakens (LTD) synaptic transmission [30]. At many glutamatergic synapses, the magnitude and temporal dynamics of activity-induced Ca^{2+} influx in the post-synaptic neuron determines whether a given level of presynaptic activity induces LTP or LTD. A fast and large increase in Ca^{2+} triggers LTP, whereas a moderate but more sustained Ca²⁺ influx gives rise to LTD [1,31–34]. The existence of distinct thresholds for LTP and LTD induction that are determined by the dynamics of Ca²⁺ concentrations in the post-synaptic neuron has been nicely illustrated by experiments in rat visual cortex: Artola and coworkers pharmacologically manipulated the level of post-synaptic depolarization by local application of the gamma-aminobutyric acid A (GABAa) receptor antagonist bicuculline. The pharmacological manipulation revealed that the same tetanic stimulation protocol induced either LTP or LTD depending on the level of post-synaptic depolarization: LTD was induced when depolarization exceeded a critical level, but still stayed below the threshold for LTP induction [35]. This study showed that the induction and direction of synaptic plasticity depends on the excitability of the post-synaptic neuron at the time of stimulation.

Homeostatic plasticity

The positive feedback nature of synaptic plasticity that allows the 'rich to get continuously richer' in the case of LTP and 'the poor to get poorer' in the case of LTD [30] challenges the stability of neural networks [1–3,24]: "Unsupervised" synaptic plasticity has the inherent risk to induce extreme neural states, causing excessive firing (in the case of uncontrolled LTP) or complete silencing of neural activity (in the case of uncontrolled LTD). An extensive body of research has demonstrated that a multitude of regulatory cellular mechanisms counteracts the 'runaway' effect of synaptic plasticity. Like LTP and LTD induction many of these mechanisms are triggered by an activity dependent change in intra-cellular Ca²⁺ levels [2,3,24,30,36]. This form of plasticity, commonly referred to as homeostatic plasticity, complements synaptic plasticity and plays a role in stabilizing mean neural activity around a set point within a physiologically reasonable dynamic range.

Net neuronal excitability depends on the interaction between intrinsic firing properties of the neuron and synaptic inputs. Therefore, homeostatic plasticity can be achieved by two fundamentally different mechanisms: synaptic homeostasis regulates excitability by up- or down-regulating synaptic strength, whereas intrinsic homeostasis shifts the relationship between synaptic input and firing by controlling intrinsic excitability [30] (Fig. 1). Even though there is ample evidence that both mechanisms coexist, it is not completely clear to what extent they serve different functions in stabilizing neural circuits and how particular firing patterns or activity levels call the appropriate homeostatic mechanism into action [37–39].

A theoretical model for homeostatic plasticity

Over 30 years ago Bienenstock, Cooper and Munro proposed a theory of how Hebbian plasticity is homeostatically regulated depending on experience-dependent modifications in postsynaptic neuronal activity. The Bienenstock–Cooper–Munro (BCM) theory postulates a "sliding threshold" for bidirectional synaptic plasticity [40,41], predicting that the thresholds for induction of LTP and LTD are dynamically adjusted to the integrated level of previous post-synaptic activity. According to the BCM theory, a history of low post-synaptic activity will lower the synaptic modification threshold for future LTP induction and increase the threshold for LTD. Conversely, a history of high synaptic activity will shift the modification threshold favoring the induction of LTD and increase the threshold for LTP (Fig. 2). The BCM theory has become the most influential model of heterosynaptic homeostatic plasticity and has guided experimental work throughout the last three decades. Even though the BCM theory was first introduced to account for experimental observations in the visual cortex, evidence for a 'sliding threshold' regulating the range of synaptic modification has A. Karabanov et al. / Brain Stimulation 8 (2015) 993–1006



Figure 1. Shows two essentially different mechanisms for the homeostatic regulation. (A) Neuronal activity is governed by both the balance of voltage-gated sodium (Na+) and potassium (K+) channels regulating intrinsic excitability and the weight of excitatory and inhibitory synapses. Neurons react to prolonged sensory deprivation either by increasing the weight of excitatory inputs synaptic homeostasis) (B) or by increasing the amount of inward voltage-dependent currents (intrinsic homeostasis) (C) whereas they react to prolonged sensory activity by increasing the weight of inhibitory inputs (synaptic homeostasis) (D) or by increasing the amount of outward voltage-dependent currents (intrinsic homeostasis) (E).

been obtained in numerous animal and human experiments [42–45] and the rule of a 'sliding threshold' has been established as a key feature of homeostatic plasticity in many brain regions [40].

The threshold for LTP and LTD induction is also modulated under physiological conditions [46,47]. A seminal study by Rioult-Pedotti et al. showed that motor skill learning shares common mechanisms with LTP in the primary motor cortex (M1): when rats had been trained for 5 days on a skilled reaching task, the trained M1 expressed less LTP and more LTD as opposed to the untrained M1 of control rats [48]. This finding shows that the ability to induce LTP and LTD is adjusted by previous learning experience, rendering the induction of LTP more difficult after intensive training.

Approaches to study plasticity in the intact human cortex

The basic mechanisms of plasticity have been primarily investigated *in vitro*. In slice preparations, LTD or LTP are commonly induced by repeated tetanic stimulation of the presynaptic neuron: at many sites, low-frequency stimulation (1-3 Hz) leads to LTD [49] whereas trains of high-frequency stimulation elicits LTP ($\geq 20 \text{ Hz}$) [50]. However, these *in vitro* studies need to be complemented by in vivo studies in animals and humans to probe the functional relevance of synaptic and homeostatic plasticity. This motivates the use of non-invasive transcranial brain stimulation to study plasticity in the intact human cortex.

A range of NTBS protocols have been established over the years to study cortical plasticity [7]. Using stimulation parameters similar to those found effective in slice preparations, both effects reminiscent of early stage LTP and LTD can be observed in the intact human brain [6,51]. Induced plasticity is commonly tested in the fast-conducting corticospinal projections by applying to the M1-Hand. The plasticity is usually probed by measuring the mean amplitude of the motor evoked potential (MEP) with single-pulse transcranial magnetic stimulation (TMS) at constant stimulus intensity before and several times after application of the plasticityinducing NTBS protocol. Serial measurements of mean MEP amplitude offer a feasible and quantitative way to test changes in excitability levels of the corticomotor output pathway. However, it should be noted that the MEP represents a complex composite measure and its amplitude is influenced by multiple physiological factors including the excitability of neural circuits at both the cortical and spinal level [9]. Finally, MEP measurements before and after a plasticity-inducing NTBS protocol restrict the investigation of cortical plasticity to the M1 and any extrapolation of the observed plasticity patterns to other cortical areas need to be made with great caution.

When applying regular trains of repetitive TMS (rTMS), high-frequency rTMS using frequencies of 5 Hz or higher [52] increase excitability in the stimulated M1 [7,53,54], while low-frequency rTMS at a frequency of around 1 Hz [55] decrease corticomotor



Figure 2. Shows the basic concept of metaplasticity following the BCM theory. The modification threshold (θ_{M}), the crossover point from LTD to LTP, is not fixed but varies as a function of post-synaptic activity. Using an LTP-like prime will shift the modification threshold ($\theta_{M'}$) to the right along the *x*-axis, while using an LTD-like prime will shift the modification threshold ($\theta_{M'}$) to the left on the *x*-axis. On the color bar, red codes an LTD response while blue codes an LTP response. (For interpretation of the references to color in this figure legend, the reader is referred to the web version of this article.)

excitability. Patterned rTMS protocols consist of short highfrequency bursts separated by longer inter-burst intervals. They are inspired by patterned burst stimulation protocols applied in cortical slices to induce LTP or LTD [12,46]. Several patterned rTMS protocols have been established, such as patterned paired-pulse protocols [56,57], theta-burst stimulation (TBS) [12,15,58] and quadripulse stimulation [12,15]. The most commonly used TBS protocol applies 50 Hz bursts consisting of three TMS pulses at a burst repetition rate of 5 Hz. TBS of M1 induces generally a lasting increase in MEP amplitude when given intermittently (referred to as intermittent TBS or iTBS), while continuous theta-burst stimulation (cTBS) induced a lasting reduction in MEP amplitude. Quadripulse stimulation (QPS) applies four-pulse bursts at a lower repetition rate than TBS, namely at 0.2 Hz. QPS of M1 at very short inter-stimulus intervals (1.5-10 ms, QPS_{short}) has been shown to increase mean MEP amplitude while QPS of M1 at inter-stimulus intervals of \geq 30 ms (QPS_{long}) decreases MEP amplitude [59].

Other repetitive TMS protocols employ associative stimulation of two neural substrates in a temporally coordinated manner. These paired association stimulation (PAS) protocols use a temporal learning rule in analogy to spike-timing dependent plasticity (STDP). For STDP, the direction of plasticity (LTP or LTD induction) depends on the precise timing of pre- and post-synaptic stimulation. The classic PAS protocol pairs peripheral electrical stimulation with single-pulse TMS of contralateral M1 and repeats these stimulus pairs at a low frequency of 0.1 Hz [14,60–62]. More recent cortico-cortical PAS protocols use dual-site TMS targeting two cortical areas [61–64]. Corticomotor excitability increases after classical PAS, if the afferent stimulus reaches M1 before or at the same time as TMS-induced M1 stimulation. Conversely, corticomotor excitability is reduced, if the afferent stimulation reaches M1 after excitation by TMS.

Also, transcranial direct current stimulation (TDCS) can be used to induce lasting bidirectional excitability changes in the human cortex. By applying a constant low current via small electrodes TDCS can either de- or hyperpolarize a neuron's resting membrane potential: anodal TDCS (aTDCS) is thought to depolarize neurons and thereby increases corticomotor excitability, whereas cathodal TDCS (cTDCS) hyperpolarizes the resting membrane, causing a decreased corticomotor excitability [16].

For some but not all of these protocols, it has been shown that changes in MEP amplitude after NTBS of M1 display some features that are reminiscent of LTP or LTD at the synaptic level. The modulation of excitability outlasts stimulation time by at least 30 min, depends on NMDA receptor activity, and originates not from subcortical or spinal excitability changes but from a cortical level [7,53,54]. Therefore the lasting increases or decreases in corticomotor excitability are often called 'LTP-like' or 'LTD-like' plasticity. It is important to note though that despite the resemblance between NTBS-induced 'LTP-like' or 'LTD-like' effects and synaptic LTP or LTD, there are apparent differences: TMS activates a substantial number of axons and leads to a massive stimulation of both inhibitory and excitatory cells, whereas synaptic activity is limited to a very small number of connections in classical in vitro studies of LTP and LTD [65,66]. Therefore, NTBS-induced plasticity is likely a mixture of plasticity induction in a number of different sets of excitatory and inhibitory synapses. Indeed, a simple equalization of synaptic effects and the NTBS induced after effects is certainly an oversimplification [67]. This is why, in the following text, the terms "inhibitory" (LTD-like) or "facilitatory" (LTP-like) are only describing the final outcome of a protocol on cortical excitability. In fact, a "facilitatory" protocol could be caused by a decrease in inhibition instead of up-regulated excitation. Another important point to note is that the knowledge about LTP- and LTD-like effects is nearly exclusively based on NTBS studies targeting M1 and these effects can not be easily extrapolated to other cortical areas.

Testing homeostatic plasticity with NTBS targeting human M1

The BCM theory predicts that high levels of prior activity favor the induction of LTD, while low levels of prior activity favor LTP [68]. In the human M1, homeostatic patterns have been tested using a priming test design, which consists of a "priming" NTBS protocol that triggers a homeostatic response and a "test" NTBS protocol that captures the homeostatic response (for recent review, [69]). The first study that showed bidirectional homeostatic-like plasticity in M1 combined a TDCS protocol to prime the subsequent response of M1 to a 1 Hz rTMS test protocol: In separate sessions, facilitatory aTDCS, inhibitory cTDCS, or sham stimulation were applied prior to a 15 min treatment session of low-intensity 1 Hz TMS. After a facilitatory aTDCS priming session, the subsequent 1 Hz rTMS test session had a marked LTD-like effect, causing a reduction in corticomotor excitability. Conversely, inhibitory priming with cTDCS flipped the effect of the very same 1 Hz rTMS test session, which now produced an increase in corticomotor excitability. When preconditioned by sham TDCS, the 1 Hz protocol did not have an effect on corticomotor excitability [70]. This bidirectional modulation of the subsequent 1 Hz rTMS session by the polarity of TDCS strongly suggests that TDCS triggered a homeostatic mechanism in the primed M1 according to the BCM theory. The observation that in the same individual the same NTBS protocol caused either LTP- or LTD-like effects depending on the history of neural activity (manipulated by TDCS priming) questions the validity of a rigid distinction in "facilitatory" or "inhibitory" NTBS protocols, as if these attributes were stable for a given NTBS protocol and robust against the physiological context.

Many other studies have reported similar homeostatic 'priming' effects on the plasticity-inducing properties of various NTBS protocols [71–76]. The homeostatic pattern that emerged in these studies showed that the priming NTBS would boost the effect of subsequent test NTBS protocol only if the priming NTBS induced the opposite effect on excitability as the test NTBS. Conversely, the priming NTBS would weaken or reverse the effect of subsequent test NTBS, if it had the same effect on excitability as the test NTBS (Fig. 2). A homeostatic reversal of the excitability effect has also been observed when the same NTBS protocol was applied consecutively [73], when two NTBS protocols were applied simultaneously [74], when doubeling the duration of stimulation [77,78] or when omitting breaks in the stimulation [79].

These experiments point to the importance of the interval between priming and test NTBS. Within the framework of the BCM theory, this implies that the temporal dynamics of the primed change in post-synaptic neural activity is critical to shift the sliding threshold in a homeostatic fashion. Yet only one study has tried to systematically investigate the time dependency of homeostatic plasticity by systematically varying the interval between priming and test NTBS and assess the impact of this manipulation on the induction of a homeostatic response [80]. Fricke and coworkers paired two identical 5 min sessions of TDCS. Priming and test TDCS sessions were separated by 0, 3 or 30 min. When priming and test TDCS were given without a break, the TDCS effect was simply prolonged. If the two TDCS sessions were separated by 30 min, there was no priming effect on the plasticity-inducing effect of the test TDCS. Only when the test TDCS started 3 min after the end of priming TDCS, did the two TDCS protocols interact in a homeostatic fashion [80]. This study stresses that there might be a critical time window during which a homeostatic response pattern emerges after priming NTBS. The importance of the interval between NIBS protocols has also been highlighted by several studies showing a (non-homeostatic) prolongation of the inhibitory effects of cTBS when the cTBS protocol is repeated after a 10–15 min break [58,81]. The studies cited above show that the interval between repeated NIBS protocols could have versatile effects on NIBS-induced plasticity. It is, however, important to keep in mind that critical time windows are likely to differ among different priming NIBS protocols [58,81]. A closely related factor that has never been systematically investigated is the "integration time" for the record of prior activity. This has important implications for both the duration of the prime and the interval between prime and test protocol: Am infinite integration of prior activity would prohibit effects caused by short-term priming protocols, whereas a very short integration time would allow extremely short priming interventions to be effective. A better understanding of the homeostatic integration time might be relevant to understand why some priming-test



Figure 3. Shows the bidirectional shift of the LTP-LTD induction curve predicted by the BCM theory (A) and induced by a priming QPS session (B). (A) The LTD–LTP crossover point (θ_M) slides to the right on the *x*-axis if the preceding neuronal activity is high ($\theta_{M'}$), and to the left if preceding activity is low ($\theta_{M''}$). (B) QPS with priming over *M*1. The normalized amplitudes of MEP at 30 min post conditioning as a function of the reciprocal of ISI of QPS (in Hertz) with and without priming over *M*1. QPS-5 ms priming over *M*1 resulted in a rightward shift, whereas QPS-50 ms priming produced a leftward shift of the "LTP-LTD induction curve". The *x*-axis is logarithmically scaled. (Reprinted from Hamada, M. and Ugawa, Y., *Restor. Neurol. Neurosci.*, 28, 419, 2010. With permission from IOS Press and the original authors.)

protocols do not cause homeostatic effects (in these cases the integration time might have been too long).

A relatively new TMS protocol that has proven to be especially helpful for investigating homeostatic effects in M1 is quadruplepulse stimulation (QPS). QPS induces changes in corticomotor excitability by applying trains of four-pulse bursts with an interburst interval of 5 s [59]. Depending on the ISI that separates the four pulses, QPS induces either an LTP-like increase in corticomotor excitability or an LTD-like decrease in corticomotor excitability. An "LTP-LTD induction curve" can be derived by plotting the LTP- or LTD-like effects of the QPS (x-axis) against the frequency of the fourpulse burst [15]. Hamada et al. (2008) showed that this LTP-LTD induction curve can be bi-directionally shifted by a priming QPS protocol (Fig. 3B): A priming QPS with an LTP-inducing high-frequency burst (i.e., QPS with a short ISI of 5 ms) switches the "normal" LTPlike effect of most QPS protocols with short ISIs into an LTD-like effect. An LTP-like effect only persisted for the test QPS protocols with the shortest ISIs. In other words, the priming QPS caused a homeostatic rightward shift of the LTD/LTP induction curve. The opposite effect was produced when an LTD-inducing QPS prime with a lowfrequency burst (i.e., QPS with a long ISI of 50 ms) was used. In this case, priming QPS switched the "normal" LTD-like effect of most QPS protocols with long ISIs into an LTP-like effect, causing a homeostatic Summarizes the results of different studies of homeostatic and non-homeostatic plasticity.

866

Homeostatic plasticity Primary motor cortex Sieber et al. (2004) [7] aTDSC/1 Hz rTMS Shows a full homeostatic interaction between priming and an inhibitory test protocol. Ling et al. (2007) [73] Muller et al. (2007) [73] GHTMS(1 Hz rTMS) The facilitatory priming increases the LTD-like effect of the 12 Hz star priorocol. Muller et al. (2007) [73] Muller et al. (2007) [74] The Sitter et al. (2007) [74] A Personal function of the test PAS: prime Nisseh et al. (2007) [74] Todd et al. (2009) [75] The facilitatory priming increases the LTD-like effect of the test PAS: prime A Personal function of the test PAS: prime Nisseh et al. (2004) [72] Todd et al. (2009) [73] The facilitatory priming increases the LTD-like effect of the test PAS: prime A Personal function of the test PAS: prime Nisteh et al. (2004) [75] Todd et al. (2008) [75] The facilitatory prime in a prime/set priority PAS: prime Gentro et al. (2008) [76] CHTSG (MS: prime) The facilitatory prime in a prime/set priority PAS: prime interaction the test protocol in tracortical and train defect of the test PAS: prime induced and train the protocol is prioroged, no activity prime is needed to induce an ITD. Bit effect CHTSG (MS: prime) CHTSG (MS: prime) The interaction of the test PAS: prime induced and train the protocol is prioroged, no activity prime is needed to induce an ITD.			Study	Priming/Test	Main findings
CIUSAN IPP INDS protocol. yer et al. (2003) [71] File TMS1 IPP INDS protocol. Muller et al. (2007) [73] PASUP-PASUP APPASUP Muller et al. (2007) [73] PASUP-PASUP APPASUP Nitsch et al. (2007) [74] PASUP-PASUP APPASUP Nitsch et al. (2007) [74] PASUP-PASUP APPASUP Nitsch et al. (2007) [74] PASUP-PASUP APPASUP Todd et al. (2007) [74] PASUP-PASUP Approxemation (File Concurrently when given as a prime/ter prococol bith TCS Todd et al. (2009) [75] 2 Hz or 6 Hz rTMS/GIBS The rTMS priming did not effect her TBS effect, but the TBS prime did increase the ingibinory effect of (TBSUN) profiles (File SUP) (FILE	Homeostatic plasticity	Primary motor cortex	Siebner et al. (2004) [70]	aTDSC/1 Hz rTMS	Shows a full homeostatic interaction between priming and an inhibitory test
her et al. (2004) [17] hang et al. (2007) [73] Muller et al. (2007) [73] Nitsche et al. (2007) [74] Todd et al. (2009) [75] Centurer et al. (2009) [75] Centurer et al. (2009) [75] Centurer et al. (2009) [75] Centurer et al. (2009) [76] Centurer et al. (2009) [77] Centurer et al. (2009) [77] Centurer et al. (2009) [78] Centurer et al. (2008) [77] Nuscle et al. (2001) [78] Centurer et al. (2008) [77] Nuscle et al. (2001) [78] Centurer et al. (2008) [78] Centurer et al. (2009) [78] Centurer et al. (2009) [78] Centurer et al. (2008) [78] Centure				cides/i Hz rims	protocol.
Lang et al. (2004) [72] Lang et al. (2004) [72] DLS/SP 127 IMS One of the first studies to show a Juli noneestatic interaction between priming Muller et al. (2007) [73] PMS_mp PMS_mp A PMS_mp prime increases the IDP sile effect of the test PMS_m, an PMS_m prime Muller et al. (2007) [74] PMS_mp PMS_mp A PMS_mp prime increases the facilitation PMS effect of CMS. PMS_mp PMS_mp CDS(JPMS_mp A PMS_mp prime increases the facilitation PMS effect. Todd et al. (2008) [75] Todd et al. (2008) [77] THE (TMS/CTBS THESS) The rFMS priming did not effect the cTBS effect, but the rTBS prime did increases the facilitation PMS effect. Status et al. (2001) [78] CTBS(short)[PMS_mp The rFMS priming did not effect the cTBS effect, but the rTBS prime did increases Genttor et al. (2008) [77] CTBS(short)[PMS_mp The rFMS prime indices and the PMS_mp inhibition without change to infracortical circuits. Gamboa et al. (2010) [78] CTBS (double duration) TTBS (double duration) TTBS (double duration) Rothkegel et al. (2011) [78] A PMS_mp protocol is pro			lyer et al. (2003) [71]	6 HZ TIMS/I HZ TIMS	The facilitatory priming increases the LTD-like effect of the THz test protocol.
click_Sp Hz rMS and an iduitatory rest protocol. APS_mp Prime inclustory rest protocol. Muller et al. (2007) [74] PMS_mp *PKS_mp APS_mp prime inclustory rest protocol. Nitche et al. (2007) [74] PMS_mp *PKS_mp decreases the 1TF-like effect of the test PMS_m, an PAS_mp prime decreases the 1TF-like effect of the test PMS_mp. Nitche et al. (2009) [75] 2.1k or 6 Hz rMS(TS monocurrently when given a 3 prime/fect of cross file cr. but the TBS prime did increase the 1TF-like effect of the rest PMS_mp. monocurrently when given a 3 prime/fect of cross file cr. but the TBS prime did increase the 1TF-like effect of the rest PMS_mp. Todd et al. (2009) [75] 2.1k or 6 Hz rMS(TBS mBS(TF)PMS_mp. The rTMS priming did not effect the CTBS effect, but the TBS prime did increase the 1TF-like effect of the rest PMS_mp. Gentor et al. (2008) [77] CTBS (dott)[PAS_mp. The rTMS prime did increase the 1TF-like effect of the rest PMS_mp. Gambaa et al. (2010) [78] CTBS (dott)[PAS_mp. The did bit protocol is proinged, no activity prime is enced to induce a trivity. Gambaa et al. (2010) [78] PTMS protocol when prime did increase the 1TF-like effect of the rest PMS muscle activity. The rTMS protocol when prime did increase the 1TF-like effect of the rest PMS muscle activity. Muscle activity/FMS protocol when prime did increase the 1TF-like effect of the rest PMS muscle activity. The rTMS protocol when prime did increase the 1TF-like effect of the rest PMS muscle activity. The rTMS protocol when prime did increase the 1TF-like effect of the rest PMS muscle activity.			Lang et al. (2004) [72]	aTDCS/5 Hz rTMS	One of the first studies to show a full homeostatic interaction between priming
Muller et al. (2007) [74] PAstrp -PAstrp PAstrp -PAstrp PAstrp -PAstrp PAstrp -PAstrp PAstrp -PAstrp PAstrp -PAstrp PAstrp Pastrp Prime increases the LIP-side effect of the test PAstrn, an PAstrp prime ecceases the LIP-side effect of the test PAstrn, and PAstrp prime ecceases the LIP-side effect of the test PAstrn, and PAstrp prime ecceases the LIP-side effect of the test PAstrn, and PAstrp Prime ecceases the LIP-side effect of the test PAstrn, and PAstrp Prime ecceases the LIP-side effect of the test PAstrn, and PAstrp Prime ecceases the LIP-side effect of the test PAstrn, and PAstrp Prime ecceases the LIP-side effect of the test PAstrn, and PAstrp Prime ecceases the LIP-side effect of the test PAstrn, and PAstrp Prime ecceases the LIP-side effect of the test PAstrn, and LIP-side effect of the test PAstrn, and LIP-side effect of the past PAstrn				cIDCS/5 Hz rIMS	and an facilitatory test protocol.
PAstront PrAstront			Muller et al. (2007) [73]	PAS _{LTP} -PAS _{LTP}	A PAS _{LTD} prime increases the LTP-like effect of the test PAS _{LTP} , an PAS _{LTP} prime
Nitsche et al. (2007) [74] AlbCs/PStTP A nonstatut effect was only observed where given concurrently when given as prime/test protocol bith TDCS prime induction concurrently and bits protocol bith TDCS effect. Todd et al. (2009) [75] 21k or 61k2 (TMS) (TDS prime) The rTMS prime induced bith TDCS effect. Ni et al. (2014) [76] CTBS (Short) [PAS;TP The rTMS prime induced bith TDCS effect. Genime et al. (2008) [77] CTBS (Short) [PAS;TP The rTMS prime induced bith TDCS in outcome the PAS;TP facilitation and led to reduced SICL and LICL and L				PAS _{LTD} -PAS _{LTP}	decreases the LIP-like effect of the test PAS _{LTP} .
cibc.s/rAs_rm cibc.s/rAs_rm concurrently when given as a prime/rest protocol bith IDCS rest of a random set of iter TMS/rBS protocols and increase the ingibitory effect. protocols and increase the ingibitory effect. rest of iter TMS/rBS Ni et al. (2014) [76] 21k or 6 hir TMS/rBS The rTMS priming did not effect the cTBS effect. Ni et al. (2014) [76] crts of hir TMS/rBS The rTMS priming did not effect the cTBS effect. The rTMS priming did not effect the cTBS effect. Gentner et al. (2008) [77] Muscle activity/rBS (20 s) Short rTBS did only induces an ITD-like effect when given is a standart 5 Hz protocol to the activity of the standart Gamboa et al. (2010) [78] effect (2010) [78] Both TTBS effect. Gamboa et al. (2010) [78] fifter intervals at itervals at itervals at itervals at a standart 5 Hz protocol the facilitation effect is turned to an inhibition. Both TTBS exerces their effect when given for double the standart diducation. Rothkegel et al. (2011) [80] at CTBS creates at a standart 5 Hz protocol the facilitation effect is turned to an inhibition. When the protocols are given without a break (doubling their length) a protocol the facilitation effect is turned to an inhibition. Intracortical networks Deelgen et al. (2011) [86] TTBS/cTBS No effect of priming on SICI and SICF. Fricke et al. (2011) [86] rTBS/cTBS No effect of priming on SICI and SICF. CrtDCS/sTDCS Intercergional cortical networks and outside MT			Nitsche et al. (2007) [74]	aIDCS/PAS _{LTP}	A homestatic effect was only observed when the protocols where given
einfer & prime(%) or einfer & prime(%) or rodo et al. (2009) [75] 2 thz or 6 itz crite(%) or rTSS(CTBS Ni et al. (2014) [76] CTBS(short)[PAS _{LTP} CTBS(short)[PAS _{LTP} CTBS(short)[PAS _{LTP} CTBS(short)[PAS _{LTP} CTBS(short)[PAS _{LTP} CTBS(short)[PAS _{LTP} CTBS(short)[PAS _{LTP} CTBS(short)[PAS _{LTP} CTBS(short)[PAS _{LTP} CTBS(short)[PAS _{LTP} CTBS(double duration) Rothkegel et al. (2010) [78] Fricke et al. (2010) [78] CTBS(double duration) TTBS(double duration) TTBS(TTBS Sober et al. (2011) [80] TTBS(TTBS Sober et al. (2011) [80] TTBS(TTBS				CIDCS/PAS _{LTP}	concurrently when given as a prime/test prococol bith TDCS
Todd et al. (2009) [75]2 Hz or 6 Hz rMS/CTBSThe rTMS priming did not effect the cTBS effect, but the iTBS prime did increase the ingibiary effect of cTBS.Ni et al. (2014) [76]CTBS/Short)/RAStrp crBS/Short//RAStrpThe cTBS prime enhanced the PAStrp ficilitation and led to reduced SCI and UCI crBS/Short//RAStrp crBS (40 s)Gentner et al. (2008) [77]Muscle activity/CTBS (20 s)Short CTBS did only induces an LTD-like effect when prime dby muscle activity, when the protocol is prolonged, no activity prime is needed to induce an LTD- like effect.Gamboa et al. (2010) [78]CTBS (double duration) TTBS (double duration)Both TTBS and cTBS reverse their effect when given for double the standart duration.Rothkegel et al. (2010) [78]TTBS (double duration) TTBS (double duration)Both TTBS and cTBS exerse their effect is sen, when the break is 20 min the protocols at different intervalsHamada et al. (2008) [59]QPS/QPSHigh-freq. QPS priming induces the opposite effect to morecolar activity in diverse is a homeostatic inghtward shift of the LTD/LTP induction curve. Low homeostatic inghtward shift of the LTD/LTP induction curve.Intracortical networksDoelrgen et al. (2001) [86]TTBS/CTBS TTBS/CTBSNo effect of priming on SICI and SICF. CTDS/CTDCS at different intervalsInteracortical networks and outside MIPoter-Nerger et al. (2009) [89]1Hz rTMS/PAStrp TTBS/CTBSNo effect of priming on SICI and SICF. SICI is only altered when prime and test protocol over M1 increases M1 excitability.Intercortical networks and outside MIQPS/QPS1Hz rTMS/PAStrp TTBS/CTBSNo effect of priming on SICI and SICF. SICI is only a				concurrently	protocols did increase the facilitatory PAS effect.
InteractionTBS/CFBSthe inglitory effect of CFBS.Ni et al. (2014) [76]CFBS (born)[PASTITGentner et al. (2008) [77]Muscle activity/CFBS (20 s)Gamboa et al. (2010) [78]CFBS (do s)Gamboa et al. (2010) [78]CFBS (do s)Rothkegel et al. (2010) [79]CFBS (do s)Britz CS/STOCSCFBS (do s)Britz CS/STOCSCFBS (do s)Britz CS/STOCSBroin TBS and CFBS reverse their effect when given for double the standart durationBritz CS/STOCSTDS/STOCSBritz CS/STOCSCFDS (CFCS do s)Intracortical networksDoeltgen et al. (2011) [80]Intracortical networksDoeltgen et al. (2011) [80]Interregional cortical networks and outside M1Poter-Nerger et al. (2011) [80]Interregional cortical networks and outside M1Poter-Nerger et al. (2009) [89]Interregional cortical networks and outside M1Poter-Nerger et al. (2009) [80]Interregional cortical networks a			Todd et al. (2009) [75]	2 Hz or 6 Hz rTMS/cTBS	The rTMS priming did not effect the cTBS effect, but the iTBS prime did increase
Ni et al. (2014) [76]CTBS (short) [PASTIP CTBS (short) [PASTIP Muscle activity) CTBS (40 s)The CTBS prime enhanced the PASTIP facilitation and led to reduced SICI and LICI ad abalished the PASTIP facilitation and led to reduced SICI and LICI adaption without change to intracortical incuits.Genther et al. (2008) [77]CTBS (60 s)Short CTBS (did only induces an LTD-like effect when prime by muscle activity, when the protocol is prolonged, no activity prime is needed to induce an LTD like effect.Gamboa et al. (2010) [78]CTBS (double duration) TTBS (double duration) aTDCS (ATDCS at DCS (ATDCS at different intervalsBoth RSB and CTBS reverse their effect when given for double the standart or an inhibition.Fricke et al. (2011) [80]TDCS (ATDCS at different intervalsWhen omitting breaks in a standart 5 Hz protocol the facilitation effect is turned uration.Intracortical networksDoelgen et al. (2008) [59]QPS (QPSWhen the protocols are given without a break (doubling their length) a prolongation of the test" effect is seen, when the break is 20 min the protocols do no nitheract but when given with a 3 min break between test and prime the protocol are given with a 3 min break between test and prime there is a homeostatic inflward shift of the LTP-LTD induction curve.Intracortical networksDoelgen et al. (2011) [86] Fricke et al. (2011) [86]TTBS (CTBS TTDS (CTCS TTDS (CTCS TTDS (CTCS TTDS (CTCS TTDS (CTCS)No effect of priming on SICI and SICF.Intracortical networksDoelgen et al. (2009) [80]TTBS (CTBS (TTDS (CTCS) TTDS (CTCS)No effect of priming on SICI and SICF.Interregional cortical networks and outside M1Potter-Nerger et al. (200				iTBS/cTBS	the ingibitory effect of cTBS.
CHES(AND)[PPS_mo and abolished the PAS_mo inhibition without change to intracortical circuits. Muscle activity/CHES (20 s) CHES (40 s) bort CHES (40 or) CHES (40 s) and abolished the PAS_mo inhibition without change to intracortical circuits. Bort CHES (40 s) CHES (40 s) bort CHES (40 or) induces an LTD-like effect when prime db ymscle activity, when the protocol is prolonged, no activity prime is needed to induce an LTD. Interegional cortical Bort CHES (40 s) bort CHES (40 or) induces an LTD-like effect when prime db ymscle activity, when the protocol is prolonged, no activity prime is needed to induce an LTD. Interegional cortical Interregional cortical networks and outside Mi Genther et al. (2010) [79] 5 Hz TMS protocol with or without breaks Scheepee effect when given for double the standart dufferent intervals Interregional cortical networks and outside Mi Polegen et al. (2011) [80] THES/CHES cortEX When the protocol sare given without a break between test and prime three is a homeostatic intervals Interregional cortical networks and outside Mi Doelgen et al. (2011) [80] THES/CHES cortEX No effect of priming conses a homeostatic rightward shift of the LTD/LTP induction curve. Low-freq. QPS priming induces the opposite effect (CHES/CHES cortEX) No effect of priming on SICI and SICF. Interregional cortical networks and outside Mi Potter-Nerger et al. (2009) [89] 1 Hz rTMS/ 1 Hz rTMS/ HAmada et al. (2009) [41] QPS/QPS Hightspresting crtES/CHES cortEX SICI			Ni et al. (2014) [76]	cTBS(short)/PAS _{LTP}	The cTBS prime enhanced the PAS _{LTP} facilitation and led to reduced SICI and LICI
Gentner et al. (2008) [77] KuSS (20 s) CTBS (40 s) Short CTBS did only induces an LTD-like effect when prime by muscle activity. when the protocol is prolonged, no activity prime is needed to induce an LTD. like effect. Gamboa et al. (2010) [79] 5 Hz TMS protocol with a trianon rTBS (double duration) rites (double duration) Both TBS and CTBS reverse their effect when given for double the standart duration. Rothkegel et al. (2010) [79] 5 Hz TMS protocol with out without breaks When omitting breaks in a standart 5 Hz protocol the facilitation effect is turned to an inhibition. Pricke et al. (2011) [80] aTDCS/aTDCS at different intervals When the protocols are given without a break (doubling their length) a prolongation of the 'tst' effect is see, monestatic interaction. Hamada et al. (2008) [59] QPS/QPS High-freq. QPS priming causes a homeostatic rightward shift of the LTD/LTP induction curve. Low -freq. QPS priming induces the opposite effect (homeostatic leftward shift of the LTP-LTD induction curve). Intracortical networks Doeltgen et al. (2001) [80] TTBS/CTBS ritek et al. (2011) [80] TTBS/CTBS ritek et al. (2001) [70] No effect of priming on SICI and SICF. (TDCS/TDCS rCTDCS/TDCS rCTDS/TBS No effect of priming on SICI and SICF. Interregional cortical networks and outside M1 Potter-Nerger et al. (2009) [89] TTBS/MPS/Stm 5 Hz rTMS/PAStm 7TBS/TTBS No effect of priming on SICI and SICF. SICI is only altered when prime and test protocol over M1 increases M1 excitability. SICI and SICF. SICI is only altere				cTBS(short)/PAS _{LTD}	and abolished the PAS _{LTD} inhibition without change to intracortical circuits.
CTBS (40 s) when the protocol is prolonged, no activity prime is needed to induce an LTD. like effect. Gamboa et al. (2010) [78] CTBS (40 s) when the protocol is prolonged, no activity prime is needed to induce an LTD. like effect. Rothkegel et al. (2010) [79] STH2: (MS) protocol with or without breaks in a standart 5 H2 protocol the facilitation effect is turned to an inhibition. Both TTBS and CTBS reverse their effect when given for double the standart duration. Pricke et al. (2011) [80] STH2: (MS) protocol with or without breaks in a standart 5 H2 protocol the facilitation effect is turned to an inhibition. When mitting breaks in a standart 5 H2 protocol the facilitation effect is turned to an inhibition. When the protocols are given without a break (doubling their length) a TDCS/aTDCS at different intervals When the protocol are given with a 3 min break between test and prime three is a homeostatic inghtward shift of the LTD/LTP induction curve. Low-freq. QPS priming induces the opposite effect (homeostatic leftward shift of the LTD-LTD induction curve.). Intracortical networks Doeltgen et al. (2011) [86] TTBS/CTBS No effect of priming on SICI and SICF. Stehner et al. (2004) [70] ATDCS/ATDCS No effect of priming on SICI and SICF. SICE is only altered when prime and test protocol are identical. Interregional cortical networks and outside M1 Potter-Nerger et al. (2009) [89] H2 rTMS/PAStrap SICE is only altered when prime and test protocol over M1 increases M1 excitability. SICE is			Gentner et al. (2008) [77]	Muscle activity/cTBS (20 s)	Short cTBS did only induces an LTD-like effect when primed by muscle activity,
Gamboa et al. (2010) [78] CBS (double duration) TBS (double duration) or without breaks without breaks Both TBS and CTBS reverse their effect when given for double the standart duration. Rothkegel et al. (2010) [79] 5 Hz rTMS protocol with or without breaks When omitting breaks in a standart 5 Hz protocol the facilitation effect is turned to an inhibition. Fricke et al. (2011) [80] ATDCS/ATDCS at different intervals When the protocols are given without a break (doubling their length) a prolongation of the 'test' effect is seen, when the break is 20 min the protocols at a different intervals Hamada et al. (2008) [59] OPS/OPS High-freq, QPS priming induces the opposite effect (homeostatic interaction. Intracortical networks Doeltgen et al. (2011) [86] iTBS/CTBS No effect of priming on SICI and SICF. Siehner et al. (2004) [70] atDCS/ATDCS at different intervals No effect of priming on SICI and SICF. Murakami et al. (2009) [89] Hz rTMS/PAS.rm CTBS/(TBS TBS/CTBS SICI is only altered when prime and test protocol are identical. No effect of priming on SICI and SICF. TTMS/(TBS) Hz rTMS to the dPMC prior to a PAS.rm protocol over M1 increases M1 excitability. Interregional cortical networks and outside M1 Hamada et al. (2009) [80] Hz rTMS/TBS 1 Hz rTMS to the dPMC prior to a PAS.rm protocol over M1 suppressed M1 excitability. Hamada et al. (2009) [90] Hz rTMS// Hz rTMS/				cTBS (40 s)	when the protocol is prolonged, no activity prime is needed to induce an LTD. like effect.
Rothkegel et al. (2010) [79] FitZ (double duration) bit BC (double duration) without breaks duration. bit BC (double duration) bit bit MS protocol with or without breaks When omitting breaks in a standart 5 Hz protocol the facilitation effect is turned to an inhibition. Fricke et al. (2011) [80] ATDCS/ATDCS at different intervals When the protocols are given without a break (doubling their length) a cTDCS/CTDCS at different intervals When the protocols are given without a break (doubling their length) a cTDCS/CTDCS Intracortical networks Hamada et al. (2008) [59] QPS/QPS High-freq. QPS priming causes a homeostatic rightward shift of the LTD/LTP induction curve. Low-freq. QPS priming induces the opposite effect (homeostatic letward shift of the LTP-LTD induction curve). Intracortical networks Doeltgen et al. (2011) [86] ITBS/CTBS No effect of priming on SICI and SICF. Siebner et al. (2004) [70] ATDCS/ATDCS at different intervals siebner et al. (2004) [70] No CTDCS/Hz TMS (TBS/TBS (TBS/TBS (TBS/TBS (TBS/TBS) No effect of priming on SICI and SICF. Interregional cortical networks and outside M1 Potter-Nerger et al. (2009) [89] 1 Hz rTMS/PAStrp 1 Hz rTMS/PAStrp 1 Hz rTMS/PAStrp 5 Hz rTMS/PAStrp 1 Hz rTMS to the dPMC prior to a PAStrp protocol over M1 increases M1 excitability. Hamada et al. (2009) [90] Hz rTMS/TBS Homeostatic modulation of M1 excitability when a priming qPS prime is given to the constater and M1. Hamada et al. (2009) [90] Hz r			Gamboa et al. (2010) [78]	cTBS (double duration)	Both iTBS and cTBS reverse their effect when given for double the standart
Rothkegel et al. (2010) [79]5 Hz rTMS protocol with or withou breaksWhen omitting breaks in a standart 5 Hz protocol the facilitation effect is turned to an inhibition.Fricke et al. (2011) [80]aTDCS/aTDCS at different intervals at different intervalsWhen the protocols are given without a break (doubling their length) a prolongation of the "test" effect is seen, when the break is 20 min the protocols do not interzation.Intracortical networksDoeltgen et al. (2008) [59]QPS/QPSHigh-freq. QPS priming causes a homeostatic rightward shift of the LTD/LTP induction curve. Low-freq. QPS priming induces the opposite effect (homeostatic letward shift of the LTD-LTD induction curve). No effect of priming on SICI and SICF. TCCS/cTDCS at different intervalsIntracortical networksDoeltgen et al. (2011) [80]iTBS/cTBSNo effect of priming on SICI and SICF. CTDCS/cTDCS at different intervalsSiebner et al. (2004) [70]aTDCS or cTDCS/1 Hz rTMSNo effect of priming on SICI and SICF. SICI is only altered when prime and test protocol are identical. ITBS/TBS TBS/TBS cTBS/TBS 				iTBS (double duration)	duration.
MICS/aTDCS aTDCS/aTDCSUban minitum.Fricke et al. (2011) [80]aTDCS/aTDCS at different intervals at different intervalsWhen the protocols are given without a break (doubling their length) a prolongation of the 'test' effect is seen, when the break is 20 min the protocols do not interact but when given with a 3 min break between test and prime there is a homeostatic interaction.Intracortical networksDoeltgen et al. (2001) [86]TITS/CTBS Fricke et al. (2011) [80]No effect of priming on SICI and SICF. (homeostatic lefter of priming on SICI and SICF. cTDCS/aTDCS rDCS/aTDCSNo effect of priming on SICI and SICF. (bornestatic lefter of priming on SICI and SICF. (bornestatic lefter of priming on SICI and SICF. cTDCS/aTDCSInterregional cortical networks and outside M1Potter-Nerger et al. (2009) [89]1 Hz rTMS/PASLTP TMS/PASLTPNo effect of priming on SICI and SICF. still remain therewalsInterregional cortical networks and outside M1Potter-Nerger et al. (2009) [89]1 Hz rTMS/PASLTP Hamada et al. (2009) [89]1 Hz rTMS/PASLTP trMS/PASLTP1 Hz rTMS to the dPMC prior to a PASLTP protocol over M1 increases M1 excitability.Hamada et al. (2009) [40]QPS/QPS1 Hz rTMS/PASLTP trMS/PASLTP5 Hz rTMS to the dPMC prior to a PASLTP protocol over M1 suppressed M1 excitability.Hamada et al. (2009) [41]QPS/QPS1 Hz rTMS/ Homeostatic modulation of M1 excitability when a priming QPS prime is given to the CMAHamada et al. (2009) [90]1 Hz rTMS/ Hz rTMS/ Homeostatic modulation of M1 excitability when a priming rMS prime is given to the SMA			Rothkegel et al. (2010) [79]	5 Hz rTMS protocol with or	When omitting breaks in a standart 5 Hz protocol the facilitation effect is turned
Picke et al. (2011) [80]allCS/aTDCS classeswhile the protocols are given Windout a break (doubing the length 4) prolocols are given Windout a Deck (doubing the length 4) prolocols are given Windout a Deck (doubing the length 4) prolocols are given Windout a Deck (doubing the length 4) prolocols are given Windout a Deck (doubing the length 4) prolocols are given Windout a Deck (doubing the length 4) prolocols are given Windout a Deck (doubing the length 4) prolocols are given Windout a Deck (doubing the length 4) prolocols are given Windout a Deck (doubing the length 4) prolocols are given Windout a Deck (doubing the length 4) prolocols are given Windout a Deck (doubing the length 4) prolocols are given Windout a Deck (doubing the length 4) prolocols are given Windout a Deck (doubing the length 4) prolocols are given Windout a Deck (doubing the length 4) prolocols are given Windout a Deck (doubing the length 4) prolocols are given Windout a Deck (doubing the length 4) prolocols are given Windout a Deck (doubing the length 4) prolocols are given Windout a Deck (doubing the length 4) prolocols are given Windout a Deck (doubing the length 4) prolocol are length			Frields at al. (2011) [00]	without Dreaks	to dil illilidition.
CHCS.(CHCS)phologization of the test effect is seen, when the photocolsdifferent intervalsdifferent intervalsdifferent intervalsdifferent intervalsHamada et al. (2008) [59]QPS/QPSHigh-freq. QPS priming causes a homeostatic rightward shift of the LTD/LTPinduction curve. Low-freq. QPS priming causes a homeostatic rightward shift of the LTD/LTPinduction curve. Low-freq. QPS priming causes a homeostatic rightward shift of the LTD/LTPinduction curve. Low-freq. QPS priming causes a homeostatic rightward shift of the LTD/LTPinduction curve. Low-freq. QPS priming causes a homeostatic rightward shift of the LTD/LTPinduction curve. Low-freq. QPS priming causes a homeostatic rightward shift of the LTD/LTPinduction curve. Low-freq. QPS priming causes a homeostatic rightward shift of the LTD/LTPinduction curve. Low-freq. QPS priming causes a homeostatic rightward shift of the LTD/LTPinduction curve. Low-freq. QPS priming causes a homeostatic rightward shift of the LTD/LTPinduction curve. Low-freq. QPS priming causes a homeostatic rightward shift of the LTD/LTPinduction curve. Low-freq. QPS priming causes a homeostatic rightward shift of the LTD/LTPinduction curve. Low-freq. QPS priming causes and back and the defect of priming on SICI and SICF.Siebner et al. (2004) [70]aTDCS/GTDSrights/rightsrights/rightsrights/rightsrights/rightsrights/rightsrights/rightsrights/rightsrights/rightsrights/rightsrights/rightsrights/rights/rightsrights/rightsrigh			FIICKE ET al. (2011) [80]	aTDCS/aTDCS	prolongation of the 'tast' offact is seen, when the break is 20 min the protocols
Hamada et al. (2008) [59]QPS/QPSHigh-freq. QPS priming causes a homeostatic rightward shift of the LTD/LTP induction curve. Low-freq. QPS priming induces the opposite effect (homeostatic leftward shift of the LTP-LTD induction curve).Intracortical networksDoeltgen et al. (2011) [86]iTBS/CTBSNo effect of priming on SICI and SICF. CTDCS/CTDCSFricke et al. (2011) [80]aTDCS/aTDCS TCDS/CTDCSNo effect of priming on SICI and SICF. (TDCS/CTDCS)Siebner et al. (2004) [70]aTDCS or cTDCS/1 Hz rTMS (TBS/CTBS)No effect of priming on SICI and SICF. SICI is only altered when prime and test protocol are identical. iTBS/TTBS CTBS/TTBS TBS/CTBSInterregional cortical networks and outside M1Potter-Nerger et al. (2009) [89]1 Hz rTMS/PAS _{LTP} 1 Hz rTMS to the dPMC prior to a PAS _{LTP} protocol over M1 increases M1 excitability.Hamada et al. (2009) [44]QPS/QPS 1 Hz rTMS/ITBS (TMS/ CTMS/ITBS1 Hz rTMS to the dPMC prior to a PAS _{LTP} protocol over M1 suppressed M1 excitability.Hamada et al. (2009) [90]1 Hz rTMS/ITBS 1 Hz rTMS/ITBS5 Hz rTMS how conduction of M1 excitability when a priming QPS prime is given to the contralateral M1.				at different intervals	do not interact but when given with a 3 min break between test and prime there is a homosectitic interaction
IntractionDescriptionConstraintIntracortical networksDeeltgen et al. (2011) [86]iTBS/CTBSNo effect of priming on SICI and SICF.Fricke et al. (2011) [80]aTDCS/aTDCSNo effect of priming on SICI and SICF.CTDSC/cTDCSat different intervalsSiebner et al. (2004) [70]aTDCS or CTDCS/1 Hz rTMSNo effect of priming on SICI and SICF.Murakami et al. (2012) [88]CTBS/CTBSSICI is only altered when prime and test protocol are identical.Interregional cortical networks and outside M1Potter-Nerger et al. (2009) [89]1 Hz rTMS/PASLTD1 Hz rTMS to the dPMC prior to a PASLTP protocol over M1 increases M1 excitability.Hamada et al. (2009) [44]QPS/QPSHomeostatic modulation of M1 excitability when a priming QPS prime is given to the SMAHomeostatic modulation of M1 excitability when a priming rTMS prime is given to the contralateral M1.			Hamada et al. (2008) [59]	QPS/QPS	High-freq. OPS priming causes a homeostatic rightward shift of the LTD/LTP
Intracortical networks Doeltgen et al. (2011) [86] iTBS/CTBS No effect of priming on SICI and SICF. Fricke et al. (2011) [80] aTDCS/aTDCS No effect of priming on SICI and SICF. CTDCS/CTDCS at different intervals No effect of priming on SICI and SICF. Siebner et al. (2004) [70] aTDCS or cTDCS/1 Hz rTMS No effect of priming on SICI and SICF. Murakami et al. (2012) [88] CTBS/CTBS SICI is only altered when prime and test protocol are identical. iTBS/TTBS cTBS/TTBS SICI is only altered when prime and test protocol over M1 increases M1 networks and outside M1 Potter-Nerger et al. (2009) [89] 1 Hz rTMS/PASLTD 1 Hz rTMS to the dPMC prior to a PASLTD protocol over M1 increases M1 excitability. 5 Hz rTMS/PASLTD 5 Hz rTMS (PASLTD) 5 Hz rTMS to the dPMC prior to a PASLTD protocol over M1 suppressed M1 excitability. 5 Hz rTMS/PASLTD 1 Hz rTMS to the dPMC prior to a PASLTD protocol over M1 suppressed M1 excitability. 5 Hz rTMS/ Hamada et al. (2009) [44] QPS/QPS 1 Hz rTMS/ Ragert et al. (2009) [90] 1 Hz rTMS/iTBS Homeostatic modulation of M1 excitability when a priming rTMS prime is given to the contralateral M1.					induction curve. Low-freq. QPS priming induces the opposite effect
Intracortical networks Doeltgen et al. (2011) [86] iTBS/CTBS No effect of priming on SICI and SICF. Fricke et al. (2011) [80] aTDCS/aTDCS rDCS/cTDCS at different intervals Siebner et al. (2004) [70] aTDCS or cTDCS/1 Hz rTMS No effect of priming on SICI and SICF. Murakami et al. (2012) [88] CTBS/CTBS rTBS/CTBS rTBS/TTBS rTBS/TBS rTBS/TBS rTBS/TBS rTBS/TBS rTBS/TBS rTBS/TBS rTBS/TBS rTBS/CTBS rTBS/TBS rTMS/PASLTP Hamada et al. (2009) [44] QPS/QPS Hamada et al. (2009) [44] QPS/QPS Hamada et al. (2009) [44] PS/QPS Hamada et al. (2009) [40] 1 Hz rTMS/ Ragert et al. (2009) [90] 1 Hz rTMS/TBS Hz rTMS/TBS Hz rTMS/A Ragert et al. (2009) [90] 1 Hz rTMS/TBS Hz rTMS/TBS Hz rTMS/A Ragert et al. (2009) [90] 1 Hz rTMS/FTBS Hz rTMS/FTBS Hz rTMS/FTBS Hz rTMS/FTBS Homeostatic modulation of M1 excitability when a priming rTMS prime is given to the Contralateral M1.					(homeostatic leftward shift of the LTP-LTD induction curve).
Fricke et al. (2011) [80] aTDCS/aTDCS cTDCS/CTDCS at different intervals No effect of priming on SICI and SICF. Siebner et al. (2004) [70] aTDCS or cTDCS/1 Hz rTMS Murakami et al. (2012) [88] No effect of priming on SICI and SICF. Interregional cortical networks and outside M1 Potter-Nerger et al. (2009) [89] 1 Hz rTMS/PASLTD 5 Hz rTMS/PASLTD No effect of priming on SICI and SICF. Interregional cortical networks and outside M1 Potter-Nerger et al. (2009) [89] 1 Hz rTMS/PASLTD 5 Hz rTMS/PASLTD 1 Hz rTMS to the dPMC prior to a PASLTD protocol over M1 increases M1 excitability. Hamada et al. (2009) [44] QPS/QPS 1 Hz rTMS/ Homeostatic modulation of M1 excitability when a priming QPS prime is given to the SMA Ragert et al. (2009) [90] 1 Hz rTMS/iTBS Homeostatic modulation of M1 excitability when a priming rTMS prime is given to the contralateral M1.		Intracortical networks	Doeltgen et al. (2011) [86]	iTBS/cTBS	No effect of priming on SICI and SICF.
CTDCS/CTDCS at different intervals Siebner et al. (2004) [70] Murakami et al. (2012) [88] Interregional cortical networks and outside M1 Potter-Nerger et al. (2009) [89] Networks and outside M1 Potter-Nerger et al. (2009) [89] THZ rTMS/PASLTP Namada et al. (2009) [44] Ragert et al. (2009) [90] Hamada et al. (2009) [90] Hamada et al. (2009) [90] Harrow Cortical Name and et al. (2009) [90] Hamada et al. (2009) [90] Harrow Cortical Name and et al. (2009) [90] Hamada et al. (2009) [90] Harrow Cortical Name and et al. (2009) [90] Harrow Cortica			Fricke et al. (2011) [80]	aTDCS/aTDCS	No effect of priming on SICI and SICF.
at different intervals Siebner et al. (2004) [70] aTDCS or cTDCS/1 Hz rTMS Murakami et al. (2012) [88] CTBS/CTBS CTBS/CTBS TTMS/PAS _{LTP} to the dPMC prior to a PAS _{LTP} protocol over M1 increases M1 excitability. 5 Hz rTMS to the dPMC prior to a PAS _{LTP} protocol over M1 suppressed M1 excitability. 5 Hz rTMS to the dPMC prior to a PAS _{LTP} protocol over M1 suppressed M1 excitability. 5 Hz rTMS to the dPMC prior to a PAS _{LTP} protocol over M1 suppressed M1 excitability. 5 Hz rTMS/ TMS/				cIDCS/cIDCS	
Siebner et al. (2004) [70] AIDCS or CIDCS/1 Hz rTMS Murakami et al. (2012) [88] CTBS TBS/CTBS/CTBS TBS/CTBS/CTBS TBS/CTBS/CTBS/CTBS TBS/CTBS/CTBS/CTBS/CTBS/CTBS/CTBS/CTBS/C				at different intervals	
Murakami et al. (2012) [88] iTBS/iTBS/iTBS iTBS/iTBS/iTBS iTBS/iTBS/iTBS iTBS/iTBS/iTBS iTBS/iTBS/iTBS iTBS/iTBS/iTBS/iTBS iTBS/iTBS/iTBS/iTBS/iTBS iTBS/iTBS/iTBS/iTBS/iTBS/iTBS/iTBS/iTBS/			Siebner et al. (2004) [70]	alDCS or clDCS/1 Hz rIMS	No effect of priming on SICI and SICF.
Interregional cortical networks and outside M1 Potter-Nerger et al. (2009) [89] 1 Hz rTMS/PASLTD 1 Hz rTMS to the dPMC prior to a PASLTD protocol over M1 increases M1 excitability. Interregional cortical networks and outside M1 Potter-Nerger et al. (2009) [41] QPS/QPS 1 Hz rTMS to the dPMC prior to a PASLTD protocol over M1 increases M1 excitability. Hamada et al. (2009) [44] QPS/QPS Homeostatic modulation of M1 excitability when a priming QPS prime is given to the SMA Ragert et al. (2009) [90] 1 Hz rTMS/iTBS Homeostatic modulation of M1 excitability when a priming rTMS prime is given to the contralateral M1.			Murakami et al. (2012) [88]	clBS/clBS	SICI is only altered when prime and test protocol are identical.
Interregional cortical networks and outside M1 Potter-Nerger et al. (2009) [89] 1 Hz rTMS/PASLTD 1 Hz rTMS to the dPMC prior to a PASLTD protocol over M1 increases M1 Networks and outside M1 5 Hz rTMS/PASLTP 1 Hz rTMS to the dPMC prior to a PASLTP protocol over M1 increases M1 Networks and outside M1 6 Hz rTMS/PASLTP 5 Hz rTMS/PASLTP Hamada et al. (2009) [44] QPS/QPS Homeostatic modulation of M1 excitability when a priming QPS prime is given to the SMA Ragert et al. (2009) [90] 1 Hz rTMS/iTBS Homeostatic modulation of M1 excitability when a priming rTMS prime is given to the contralateral M1.				11BS/11BS	
Interregional cortical networks and outside M1 Potter-Nerger et al. (2009) [89] 1 Hz rTMS/PAS _{LTP} 1 Hz rTMS to the dPMC prior to a PAS _{LTD} protocol over M1 increases M1 excitability. 5 Hz rTMS to the dPMC prior to a PAS _{LTP} protocol over M1 suppressed M1 excitability. Hamada et al. (2009) [44] QPS/QPS 1 Hz rTMS/ Ragert et al. (2009) [90] 1 Hz rTMS/iTBS 1 Hz rTMS/iTBS 1 Hz rTMS/iTBS 1 Hz rTMS/iTBS				c1BS/11BS	
Interregional cortical networks and outside M1 Potter-Nerger et al. (2009) [89] 1 Hz TIMS/PASLTD 1 Hz TIMS to the dPMC prior to a PASLTD protocol over M1 increases M1 excitability. networks and outside M1 5 Hz rTMS/PASLTP excitability. 6 Hz TIMS to the dPMC prior to a PASLTD protocol over M1 increases M1 excitability. 5 Hz rTMS/PASLTP 1 Hz rTMS/ Hamada et al. (2009) [44] QPS/QPS 1 Hz rTMS/iTBS 1 Hz rTMS/ to the SMA Ragert et al. (2009) [90] 1 Hz rTMS/iTBS Homeostatic modulation of M1 excitability when a priming rTMS prime is given to the contralateral M1.			Detter News et al. (2000) [00]	11BS/CIBS	1 II. TMC to the JDMC minute a DAC
Hamada et al. (2009) [44] QPS/QPS Homeostatic modulation of M1 excitability when a priming QPS prime is given to the SMA Ragert et al. (2009) [90] 1 Hz rTMS/ Homeostatic modulation of M1 excitability when a priming rTMS prime is given to the contralateral M1.		Interregional cortical networks and outside M1	Potter-Nerger et al. (2009) [89]	1 Hz rTMS/PAS _{LTD} 5 Hz rTMS/PAS _{LTP}	I HZ FIMS to the dPMC prior to a PAS _{LTD} protocol over MT increases MT excitability.
excitability. Hamada et al. (2009) [44] 1 Hz rTMS/ Ragert et al. (2009) [90] 1 Hz rTMS/iTBS Prime is given to the SMA Homeostatic modulation of M1 excitability when a priming rTMS prime is given to the contralateral M1.				,	5 Hz rTMS to the dPMC prior to a PAS _{LTP} protocol over M1 suppressed M1
Hamada et al. (2009) [44]QPS/QPSHomeostatic modulation of M1 excitability when a priming QPS prime is given to the SMARagert et al. (2009) [90]1 Hz rTMS/iTBSHomeostatic modulation of M1 excitability when a priming rTMS prime is given to the contralateral M1.					excitability.
Ragert et al. (2009) [90] 1 Hz rTMS/iTBS Homeostatic modulation of M1 excitability when a priming rTMS prime is given to the contralateral M1.			Hamada et al. (2009) [44]	QPS/QPS 1 Hz rTMS/	Homeostatic modulation of M1 excitability when a priming QPS prime is given to the SMA
to the contralateral M1.			Ragert et al. (2009) [90]	1 Hz rTMS/iTBS	Homeostatic modulation of M1 excitability when a priming rTMS prime is given
					to the contralateral M1.
Bliem et al. (2008) [91] PAS/20 Hz HFS Homeostatic plasticity in primary sensorimotor cortex.			Bliem et al. (2008) [91]	PAS/20 Hz HFS	Homeostatic plasticity in primary sensorimotor cortex.
Gartica Jossi et al. (2014) [92] 5 Hz r1MS/20 Hz HFS Homeostatic plasticity in primary sensorimotor cortex.			Gartica Tossi et al. (2014) [92]	5 Hz rTMS/20 Hz HFS	Homeostatic plasticity in primary sensorimotor cortex.
BOCCI et al. (2014) [93] IDCS/rTMS Homeostatic plasticity in primary visual cortex.			BOCCI et al. (2014) [93]	IDCS/FIMS	Homeostatic plasticity in primary visual cortex.

Interaction of motor learning and		Ziehmann et al. (2004) [8]	Thumb abduction/PAS _{LTP} Thumb abduction/PAS _{LTD}	Motor learning can act as a priming intervention for subsequent NIBS and induce homeostatic effects.
homeostatic plasticity		Lepage et al. (2012) [95]	Motor observation/PAS _{LTP}	Observation of a motor training task is sufficient to prevent subsequent induction of LTP-like PAS effects.
		Rosenkranz et al. (2007) [85]	Novel vs. well-practiced thumb abduction/PAS	The effect of motor learning as a 'primer' depends on the learning phase: homeostatic effects only observed when 'priming' involved a novel motor task.
		Elahi et al. (2014) [98]	PAS/thumb abduction	NIBS can act as a primer on motor learning.
		Jung et al. (2009) [99]	PAS _{LTD} /thumb abduction task PAS _{LTP} /thumb abduction task	PAS given 90 min before the learning task shows a "classic" homeostatic interaction, when given directly before the task both PAS _{LTP} and PAS _{LTD} facilitate learning
		Teo et al. (2011) [100]	iBTS/thumb abduction	Priming with iTBS boosts performance in a subsequent ballistic motor learning task. The effect of priming iBTS can be blocked by nicotine administration.
		Kuo et al. (2008) [101]	TDCS/serial reaction time task	No homeostatic effect between TDCS and motor learning found.
		Rosenkranz et al. (2014) [111]	Hand immobilization/PAS	Eight hours of hand immobilization significantly reduce the inhibitory effects of PAS-10 ms while enhancing the facilitatory effects of PAS-25ms.
Non-homeostatic plasticity		Nitsche et al. (2003) [102] Anatal et al. (2004) [103] Reis and Fritsch (2011) [105] Stagg et al. (2011) [107]	Concurrent motor learning and TDCS	'Gating': studies have reported reinforcing effects between voluntary motor activity and TDCS when applied <i>concurrently</i> .
		Devendahl et al. (2010) [114]	0.1 Hz rTMS/PAS	'Anti-gating': a very low-frequency prime abolished the ability to induce LTP- and LTD-like with subsequent PAS
		Huang et al. (2010) [119]	iTBS/cTBS cTBS/iTBS	The LTP-like effect induced by iTBS is abolished (de-potentiated), when a short train of cTBS followed the protocol.
				The LTD-like effect induced by cTBS is abolished (de-depressed), if followed by a short train of iTBS.
		Ni et al. (2014) [76]	PAS _{LTP} /cTBS (short) PAS _{LTD} /cTBS (short)	De-potentiating effect of a short inhibitory follow-up.
		Goldsworthy et al. (2014) [120]	cTBS/voluntary contraction	De-depressing effect on a short facilitatory follow up on an inhibitory protocol.
		Cantarero et al. (2013) [96]	Motor learning task/cTBS	Occlusion of LTP-like effect and motor skill retention after short inhibitory
		Cantarero et al. (2013) [97]		protocol.
		Lepage et al. (2012) [95]	Motor observation/PAS _{LTP}	Observation of a motor training task is sufficient to prevent subsequent induction of LTP-like PAS effects.
Homeostatic Plasticity in pathological states	Focal hand dystonia	Quartarone et al. (2005) [138]	TDCS/1 Hz rTMS	The 'homeostatic' response pattern of healthy controls is absent in the affected hand of writer's cramp patients.
		Kang et al. (2011) [139]	PAS _{LTP} —thumb abduction PAS _{LTD} —thumb abduction	In contrast to healthy controls the writer's cramp patients do not show any modulation of learning-dependent plasticity.
	Parkinson's disease	Huang et al. (2011) [148]	TBS	Patients with levodopa-induced dyskinesia showed normal potentiation but were unresponsive to the de-potentiation protocol.

leftward shift of the LTP-LTD induction curve. The bidirectional shifts in the LTP-LTD induction curve nicely demonstrated the existence of a "sliding modification threshold" as predicted by the BCM theory [45]. Table 1 summarizes the results of different studies of homeostatic plasticity and other forms of metaplasticity.

Homeostatic plasticity in cortical networks

Intra-cortical homeostatic plasticity in the motor cortex

The MEP is a complex measure of corticospinal excitability and is influenced by spinal excitability as well as by various intracortical circuits projecting onto the corticospinal motor neurons [9,81,82]. This means that homeostatic plasticity might not only affect corticospinal neurons directly but might also act on intracortical circuits within M1.

Intracortical excitability can be measured by using paired-pulse TMS paradigms, which apply a conditioning (CS) and test stimulus (TS) through the same coil [83]. While several studies have shown motor-training induced plasticity of these intracortical inhibitory circuits [84,85], very few studies have investigated homeostatic effects in intracortical circuits. The results of these studies are not yet fully conclusive: Several studies using facilitatory and inhibitory TDCS primed 1-Hz rTMS [70], facilitatory-facilitatory TDCS and inhibitory-inhibitory TDCS [80] and iTBS primed cTBS [86] found no consistent homeostatic changes in intracortical inhibitory GABAAergic circuits in M1 underlying short interval intracortical inhibition (SICI) [87]. A more systematic investigation of homeostatic effects in intracortical inhibitory circuits demonstrated homeostatic plasticity-like effects on SICI: Murakami and colleagues [88] applied 'facilitatory' intermittent theta-burst stimulation (iTBS) or 'inhibitory' continuous theta-burst stimulation (cTBS) to induce a homeostatic response in intracortical inhibitory circuits. They found that a priming TBS protocol altered the responsiveness of the inhibitory SICI circuits to a test TBS only when the second TBS protocol was identical to the priming protocol (iTBS \rightarrow iTBS or cTBS \rightarrow cTBS). The normal direction of TBS-induced SICI aftereffects was reversed by priming with identical TBS, suggesting homeostatic regulation of excitability in inhibitory circuits. However, even in that study homeostatic metaplasticity was less consistently expressed in the intracortical inhibitory circuits than in the excitatory corticospinal pathway. In contrast to homeostasis in the corticospinal pathway alternating TBS protocols (the iTBS \rightarrow cTBS or cTBS \rightarrow iTBS) failed to trigger a homeostatic response in inhibitory circuits.

Facilitatory circuits within M1 have been even more sparsely studied than intracortical inhibition and no consistent homeostatic effects have been demonstrated so far on intracortical facilitation [80,86]. The few data presently available suggest that homeostatic plasticity is less consistently expressed. Alternatively, homeostatic plasticity in intracortical circuits upstream to the corticospinal motor neuron may simply be more difficult to capture with MEP measurements. Subtle homeostatic changes may have an effect size that remains within the noise level of normal fluctuations in MEP amplitude. More robust homeostatic effects in intracortical circuits are likely to be paralleled by concurrent homeostatic changes in the corticospinal neurons. In that case, the presence of homeostatic changes in MEP amplitude evoked by single-pulse TMS may mask homeostatic effects in upstream intracortical circuits as probed with double-pulse TMS.

Inter-cortical homeostatic plasticity

Homeostatic interactions can also occur in interregional networks. Several studies have shown that a homeostatic response can be elicited in M1 when the priming protocol is given over a secondary motor area to activate cortico-cortical projections to M1. Potter-Nerger and coworkers demonstrated homeostatic priming on PAS to left M1 after rTMS priming was applied to ipsilateral dorsal premotor cortex (dPMC). Thus, inhibitory 1 Hz rTMS of dPMC prior to an inhibitory PAS protocol over M1 increased M1 excitability, whereas facilitatory 5 Hz rTMS of dPMC prior to a facilitatory PAS protocol over M1 suppressed M1 excitability [89]. Homeostatic modulation of M1 excitability was also demonstrated when a priming QPS session was given to the supplementary motor area [44] or when a priming 1 Hz rTMS was given to the contralateral M1 [90]. Taken together, these findings indicate that homeostatic interactions can be elicited through different input channels in the human M1.

Studies using other measures of cortical excitability, such as somatosensory evoked potentials (SSEP) or visual evoked potentials (VEP), have shown that homeostatic metaplasticity can also be expressed in other cortical areas. SSEP recordings provided evidence for homeostatic plasticity in primary somatosensory cortex [91,92]. Both SSEP applied NIBS before high-frequency (20-Hz) tactile electrical stimulation of the contralateral median nerve in order to demonstrate a homeostatic response in the somatosensory cortex. In primary visual cortex, the VEP revealed a homeostatic reaction to a combined TDCS-rTMS protocol [93]. Identifying additional neurophysiological markers of brain plasticity such as recordings of TMS-evoked cortical potentials with combined TMS-EEG [94] might facilitate investigations into homeostatic effects expressed in other cortical areas.

Homeostatic plasticity and motor learning

Motor learning can induce plasticity under physiological conditions and many studies have shown that brain stimulation and motor learning can interact homeostatically. Early studies showed that a simple motor learning task could act as a 'primer' for subsequent PAS protocols. Ziemann and coworkers [8] showed that motor learning prevented the induction of subsequent LTP-like PAS effects while enhancing subsequent LTD-like effects. More recent work suggests that observation of a motor training task is sufficient to prevent subsequent induction of LTP-like PAS effects [95] and that the temporary occlusion of LTP-like plasticity after motor learning is likely to be a mechanism necessary for successful skill retention. Retention for a simple motor task after learning was proportional to the magnitude of LTP occlusion during a subsequent NTBS protocol and that the amount of occlusion was predictive of resilience against interference of subsequent learning [96,97]. Interestingly, the effect of motor learning as a 'primer' depends on the learning phase: the observed homeostatic effects on subsequent PAS protocols were only observed when 'priming' involved training a novel motor task, while 'priming' with a well-practiced task did not significantly modulate subsequent PAS [85]. Homeostatic interactions between a facilitatory PAS response and a motor learning task can also be seen if the learning task follows the PAS intervention [98].

While these studies clearly demonstrate that learning may have a homeostatic impact on plasticity induced by NTBS, the evidence for a reverse interaction, a homeostatic effect of NTBS on plasticity induced by subsequent motor learning is less consistent. According to the BCM theory, one might expect an inhibitory NTBS protocol to facilitate a subsequent motor leaning task. Jung and Ziemann [99] studied motor learning of rapid thumb abduction movements. The training session was primed with a PAS protocol which ended 0 min or 90 min before training began. When PAS was given directly before training, both the inhibitory and excitatory PAS protocol enhanced motor learning, indicating a non-homeostatic interaction. However, the same PAS protocols given 90 min before learning gave rise to a "classic" homeostatic interaction. In that condition, excitability-decreasing PAS still had a beneficial effect on motor learning, but excitability-increasing PAS impaired motor learning. These results once again stress the importance of timing between priming and test protocols and suggest that nonhomeostatic mechanisms may play a role, especially when the interval between priming stimulation and motor training is short.

Studying homeostatic plasticity in the context of motor learning is difficult, since synaptic strengthening is likely not the only factor influencing the learning rate. A more recent study found that priming with iTBS boosted performance in a subsequent ballistic motor learning task [100]. In that study, the beneficial effect of priming iTBS was blocked by the administration of nicotine. Behavioral analysis and modeling suggested that the iTBS prime facilitated performance by increasing motor output variability. The hypothesis was that the motor system could then explore the task workspace more quickly to find the optimal way to perform the task. The authors hypothesized that nicotine blocked this effect, presumably by reducing the signal-to-noise ratio in cerebral cortex [100]. This and other mechanisms may explain why other studies, which assessed the priming effects of brain stimulation on motor learning, failed to reveal homeostatic effects [101].

Many studies consistently show that NTBS protocols that are sub-threshold for inducing action potentials in the cortex, in particular TDCS, can enhance motor learning when the NTBS protocol is given concurrently with the learning task [102–107]. Although most NTBS protocols that were applied during motor training enhanced motor learning in a non-homeostatic fashion, homeostatic interaction might well occur. However, this should not be called "metaplasticity," because priming and test intervention are not separated in time [3]. An optimal exploitation of homeostatic mechanisms to boost motor learning will require a better understanding of the mechanisms by which the various NTBS protocols modulate motor learning.

It is worth mentioning, that homeostatic interactions between voluntary movement and NTBS are not restricted to motor learning. Several studies have demonstrated plasticity interactions (homeostatic and non-homeostatic) when simple voluntary muscle contractions where performed prior, during or after an NTBS protocol [77,108–110]. Also restricting movement can have a homeostatic influence on NTBS-induced plasticity: 8-h of hand immobilization did significantly reduce the inhibitory effects of PAS-10 while enhancing the facilitatory effects of PAS-25 [111].

Gating vs. homeostatic plasticity

The interactions between motor training and concurrent NTBS often follow non-homeostatic rules (i.e., the priming intervention does not have a homeostatic effect on the test procedure). A complementary mechanism by which NTBS might increase the beneficial effects of motor learning is 'gating'. Many studies have reported gating interactions between voluntary motor activity and NTBS when NTBS was applied *concurrently* with a motor task [102,103,105,107].

Gating mechanisms may also increase the efficacy of NTBS of the M1 to produce LTP-like or LTD-like effects. Gating may be provoked by several mechanisms such as increasing net calcium influx into the targeted cortical neurons, shifting intrinsic excitability of the targeted neurons (e.g. sub-threshold depolarization during anodal TDCS), or transiently suppressing the efficacy of intracortical inhibitory circuits. It has been shown that NTBS can induce acute disinhibition, thereby potentially gate the plasticityinducing effects of NTBS. For instance, a short period of sub-threshold 5 Hz rTMS can cause a transient suppression of short-latency intracortical inhibition in the stimulated M1-HAND along with a increase in regional cerebral blood flow [52]. Further, a temporary ischemic nerve block of the distal upper limb caused acute disinhibition in the contralateral sensorimotor cortex and boosted training-induced learning of ballistic elbow flexion movements [112]. Together, these studies suggest that "gating" might play a role in determining the efficacy of a given NTBS protocol, but studies are lacking which systematically study the relationship between acute NTBS-induced disinhibition and the efficacy to induce LTP- or LTD-like effects. It is important to point out that gating is a non-homeostatic mechanism, because it does not alter the threshold for expressing LTP or LTD [7]. Yet gating may promote the induction of LTP-like effects in neural circuits targeted by NTBS or learning and indirectly facilitate a homeostatic response.

It is also important to note that not all interactions between consecutively paired protocols depend on homeostatic effects and that several forms of non-homeostatic metaplasticity have been observed using brain stimulation: A very low frequency (0.1 Hz) rTMS prime given to M1 abolished the ability to induce LTP- and LTD-like effects in the primed M1 with subsequent PAS [113,114]. The prime alone did not alter corticospinal excitability as measured by MEP amplitude, but increased short-interval and long-interval intracortical inhibition in the stimulated M1. Increased excitability of intracortical inhibitory circuits caused by the priming protocol might have prevented the PAS protocol from inducing LTP- or LTDlike changes by reducing the ability of afferent volleys, evoked by the peripheral stimulus, to interact with the TMS pulse given over M1-HAND in a Hebbian fashion. If the afferent volley has less "access" to the corticospinal excitability, potentially by reducing the output neurons due to excessive intracortical inhibition, the calcium influx in the corticospinal neurons during PAS [113,114]may drop below the threshold for inducing LTP or LTD-like plasticity. A reduction of the calcium influx caused by increased activity of intracortical inhibitory circuits does not invoke homeostatic regulation because the threshold for LTP and LTD induction is not principally shifted. Such a mechanism rather represents an 'antigating' effect that reduces the efficacy of NTBS without shifting the threshold for expressing LTP and LTD [113,114]. However, the notion of 'gating' and 'anti-gating' remains to be thoroughly tested in future studies.

Another non-homeostatic form of metaplasticity is depotentiation (or de-depression). De-potentiation erases previously induced LTP (or LTD) and may be the key mechanism for retrograde inference with learning. There is ample evidence for depotentiation and de-depression in the animal literature, which implicates this form of metaplasticity as a factor in learning reversal and forgetting [115–117]. Metaplasticity patterns resembling depotentiation and de-depression were observed in an experiment that combined iTBS and cTBS [118]: The normal LTP-like effect induced by facilitatory iTBS was abolished (de-potentiated), when a short train of inhibitory cTBS followed the iTBS protocol. Vice versa, the LTD-like effect normally induced by a cTBS protocol was abolished (de-depressed), if followed by a short train of facilitatory iTBS. When given alone, the short TBS trains did not change corticomotor excitability. This shows that the de-potentiating (or de-depressing) protocol itself does not need to have any discernable effect when applied alone. Only when given within a certain time window after an LTP- or LTD-inducing protocol are these effects visible. The early phases of LTP and LTD induction are more vulnerable to the effect of interfering stimuli than later phases, when synaptic changes in synaptic efficacy have been stabilized [118]. Other recent studies have confirmed the de-potentiating effect of an inhibitory followup on facilitatory NTBS protocols [76] and the de-depressing effect on a facilitatory follow up on an inhibitory protocol [119].

This study also suggests that repeated cTBS application also seems to protect against de-depression: When pairing two cTBS protocols, separated by a 10 min-break, the induced LTD-like effect was resistant against de-depression.

These examples show that there are many non-homeostatic forms of cortical plasticity and metaplasticity that might shape the efficacy of NTBS to induce LTP- or LTD-like effects. Hence, researchers investigating metaplasticity need to be careful when labeling a modulation of NTBS-induced plasticity as "homeostatic." An effect is only likely to be homeostatic, if the priming intervention alters the LTP-LTD induction curve in a way that the changes in LTD-LTP induction threshold favor the induction of plasticity opposite to the priming protocol (Figs. 2 and 3). As mentioned earlier, the temporal relationship between the priming and test protocols is crucial for the induction of both homeostatic and nonhomeostatic metaplasticity. Future studies need to explore the interplay between these non-homeostatic and homeostatic forms of cortical plasticity. Currently, there is a growing interest in therapeutic multi-session NTBS applications aiming to extend the duration of excitability changes. When designing such therapeutic protocols it is especially important to better understand possible homeostatic interactions in order to avoid creating an excitability effect in opposition to the therapeutic goal.

Mechanisms regulating metaplasticity

One of the key predictions of the original BCM theory is that the activity dependent threshold is calculated from a running timeaverage of post-synaptic action potential activity. More recent BCM models have, however, started to question the role of postsynaptic action potentials and focused on the time-averaged free calcium concentration as the biological signal controlling homeostatic metaplasticity [29,120]. Recent *in vitro* experiments confirmed that homeostatic plasticity in the hippocampus did not depend on somatic action potentials, but was determined by calcium release from intra-cellular stores, triggered by muscarinic acetylcholine receptors [121]. In addition to intra-cellular Ca²⁺ stores, Ca²⁺ can also enter the cell via NMDA receptors or via L-type voltage-gated Ca²⁺ channels. Homeostatic modulation of highfrequency tetanic stimulation was also observed when pharmacologically reducing Ca²⁺ via those routes [122–124].

A study combining an acute pharmacological intervention with cTBS showed that the magnitude of Ca²⁺ signaling is also highly relevant for the induction of LTP- and LTD-like phenomena in humans [125]. When the duration of cTBS was shortened from 40 s to 20 s, cTBS was shown to induce a facilitatory effect on corticomotor excitability. These LTP-like effects of short cTBS on corticomotor excitability were reversed when healthy volunteers were treated with nimodipine, an L-type voltage-gated \mbox{Ca}^{2+} channel antagonist. Pharmacological blockade of the NMDA receptor by dextromethorphan did not cause a homeostatic effect, but dextromethorphan abolished both the LTD-like effect of cTBS produced by nimodipine and the normal LTP-like effect of cTBS alone in M1. This study also suggested that the homeostatic effects induced by voluntary activity might be mediated by L-type voltage-gated calcium channels. It is likely that the effects of other interventional NTBS protocols are also strongly influenced by Ca²⁺ dynamics, but might be sensitive to manipulation of Ca^{2+} influx via different routes. This remains a relevant topic for future research.

At the cellular level, a complex machinery of transcriptional as well as pre- and post-synaptic molecular signaling mechanisms can induce and shape homeostatic mechanisms. These mechanisms include secreted molecules such as the brain-derived neurotrophic factor (BDNF) or the tumor necrosis factor (TNF), cell adhesion molecules (e.g. integrins, ephrins, cadherins), different kinases (CaMKs, CaMKII) and transcription factors such as Arg3.1 (for a detailed review on the molecular mechanisms of homeostatic plasticity the reader is referred to [126]).

Synaptic homeostasis and sleep

While we focused on the ability of NTBS to probe and shape homeostatic plasticity in previous sections, this section summarizes the contribution of sleep to homeostatic control and how this can be studied with NTBS. Neurons can undergo specific plastic changes during learning and behavior, they also have many ways to keep overall synaptic weights and post-synaptic activity levels under control. It has been proposed that irrespective of the specific mechanism involved, achieving this control may require the alternation between wakefulness and sleep [127]. Specifically, according to the "synaptic homeostasis hypothesis," the fundamental function of sleep is the restoration of synaptic homeostasis, which is challenged by synaptic strengthening triggered by learning during wakefulness [127]. In this framework, sleep is the price we pay for having a plastic brain that is able to learn and adapt to the everchanging demands of the environment. Since neurons signal suspicious coincidences and salient events by increasing their firing, learning should happen primarily through synaptic potentiation. Moreover, synaptic potentiation should occur mainly during wakefulness in order to be adaptive, when the brain interacts with the external environment, not during sleep when it is disconnected. Hence, wakefulness is associated with synaptic potentiation and net synaptic weight increases over the wakening hours. Increased synaptic strength during waking has obvious benefits but also various costs at the cellular and systems level; for example, it implies higher energy consumption and demand for the synthesis and delivery of synaptic supplies; in addition, it reduces the selectivity of neuronal responses and saturates the ability to learn. For this reason, neurons must eventually re-normalize total synaptic strength in order to restore cellular functions as well as selectivity. Indeed, the other main tenet of the synaptic homeostasis hypothesis is that re-normalization of synaptic strength occurs primarily during sleep, when the brain is spontaneously active offline, not in wake when a neuron's inputs are biased by a particular situation.

It is important to note that homeostatic plasticity, as described in previous sections, and synaptic sleep homeostasis are related but separate phenomena. Whereas the primary variable regulated by homeostatic plasticity is the level of neural activity [38], sleep homeostasis primarily acts on global synaptic strength. An intriguing hypothesis is that synaptic re-normalization during sleep may be brought about by slow waves and by the underlying alternation between burst firing and neuronal silence. While the relevance and the details of this mechanism remain unknown, experimental studies in animal models show that overall synaptic weights increase during wakefulness but decrease during sleep. For example, structural evidence demonstrates that the strength, the size and number of synapses in the brain of Drosophila flies increase after a period of wakefulness and decrease only when animals are allowed to sleep [128] From a molecular point of view, the levels of GluA1containing AMPA receptors (a molecular marker of synaptic potentiation) were found to be 30-40% higher after wakefulness than after sleep in rats [129]. Electrophysiologically, the slope of the early (monosynaptic) response evoked by electrical stimulation delivered in the rat cerebral cortex, a classic marker of synaptic strength in vivo, increases with time spent awake and decreases with time spent asleep [130].

In humans, a similar shift of the excitation/inhibition balance toward excitation was documented by two TMS-MEPs studies [131,132] that detected a significant decrease of short-term intracortical inhibition occurring, at the group level, after 24 h of sleep deprivation. This shift in excitatory/inhibitory balance does also affect efficiency of NTBS-induced plasticity: A systematic comparison of the effect of diurnal rhythm on facilitatory PAS showed that the facilitatory effects of PAS were greater in the evening. This study also confirmed that intracortical inhibition was reduced in the evening [133]. While providing some information on the nature of cortical plastic changes, these NTBS studies confirm the idea that in humans, sleep may contribute to keep the overall weight of cortical synapses under control.

An important practical implication is that synaptic sleep homeostasis needs to be taken into account whenever interventional NTBS protocols are given over consecutive days or weeks. In these studies, the sleep quality might have substantial impact on the emergence of cumulative NTBS effects.

Homeostatic plasticity in pathological states

Synaptic homeostasis has been demonstrated to be a fundamental mechanism within brain circuits, operating in different species including humans [2,38,127,134–136], but much less is known about the significance of dysfunctional homeostatic plasticity for the pathogenesis and pathophysiology of brain diseases. In this review, we focus on a series of experiments, which have used NTBS to probe homeostatic plasticity in focal dystonia and discuss the future potential of NTBS to study homeostatic plasticity in neuropsychiatric disorders.

Focal dystonia

Using TDCS as conditioning protocol and low-frequency (1 Hz) rTMS as test protocol, Quartarone et al. found that the 'homeostatic' response pattern of healthy controls was absent in the affected hand of writer's cramp patients [137,138]. In dystonic patients, aTDCS to M1 increased MEP amplitude as in normal controls, but the subsequent 1 Hz rTMS did not produce an LTD-like effect. Thus despite producing an LTP-like effect, aTDCS failed to trigger a homeostatic response that sensitized M1 to the LTD-inducing effect of 1 Hz rTMS.

A subsequent study addressed the question whether patients with focal hand dystonia would show an enhancement of motor learning induced plasticity after priming with an excitabilityreducing NTBS protocol as previously shown in healthy individuals [99]. While the healthy control group showed a homeostatic enhancement of learning-dependent plasticity following an excitability-reducing prime and a homeostatic suppression of learning-dependent plasticity following an excitability-increasing prime, the writer's cramp patients did not show any modulation of learning-dependent plasticity and the lack of homeostatic modulation was correlated with the clinical severity of the dystonia [139]. These results suggest that focal hand dystonia is associated with a dysfunctional homeostatic regulation of plasticity, which might set the frame for aberrant sensorimotor plasticity. Several NTBS studies have shown that patients with focal hand dystonia show excessive sensorimotor plasticity with lack of somatotopic specificity [140,141]. However, due to the large variability of PAS responses, both in patients and healthy subjects, there is considerable overlap between patient and healthy data. The questions the validity of excessive, nonfocal PAS effects as a general 'dystonic fingerprint' [142]. Large multicenter studies and a stronger focus on individual plasticity profiles will help to clarify the role of dysfunctional homeostatic plasticity in dystonia.

Additionally, it should be noted that focal dystonia is also characterized by deficient inhibition within intracortical circuits [143]. This might explain that also the response to NTBS protocols

such as iTBS, which are not involving a sensorimotor component, is abnormal in these patients. However, iTBS induced plasticity is absent or reduced in focal dystonia, not excessive and non-focal as for PAS [144]. Deficient intracortical inhibition might also produce an abnormal "gating" of the LTP-inducing effects of NTBS and hereby introduce a bias toward producing LTP-like rather than LTDlike effects in M1.

Parkinson's disease

There is ample evidence for altered LTP- and LTD-like plasticity in Parkinson's disease (PD) [145–147] and recent research suggests that abnormalities in plasticity may depend on disease state and I-DOPA administration [145]. Despite the relatively large number of NTBS studies investigating synaptic plasticity in PD, homeostatic plasticity has not been systematically investigated. Huang et al. studied non-homeostatic metaplasticity in patients with and without levodopa-induced dyskinesia (LID). PD patients without LIDs had normal potentiation and de-potentiation, when they took their full dose of levodopa. Patients with levodopa-induced LIDs were studied while being on half their usual dose of levodopa to prevent emergence of overt dyskinesias during testing. LID patients showed normal potentiation but were unresponsive to the depotentiation protocol [148]. Given this altered non-homeostatic metaplasticity in LID patients, it is possible that homeostatic plasticity might also be affected in PD.

Psychiatric disorders

Several lines of research suggest that both the NMDA- and GABA-ergic transmitter systems that participate in cortical plasticity are also involved in the pathophysiology of various psychiatric disorders such as schizophrenia (SCZ), major depressive disorder (MDD) and bipolar disorder [149–153]. Except for dysfunctional GABA and glutamatergic neurotransmission, key features of these disorders are abnormalities in the expression of several proteins which are important for synaptic plasticity and homeostatic plasticity (e.g. BDNF, dybindin, neurexin) [154–156].

Disrupted plasticity is an established part of the pathophysiology in schizophrenia (SCZ), and several neurophysiological experiments using a range of plasticity-inducing NTBS protocols have shown that LTP- and LTD-like effects are reduced in SCZ [152,157,158]. SCZ patients also demonstrate less use-dependent plasticity. By measuring the spontaneous direction of TMS-induced thumb movements before and after 30-min training in thumb abduction, Daskalakis and coworkers [159] found that M1 excitability was affected less in SCZ patients than healthy controls. Impaired cortical plasticity has also been reported in patients with major depressive disorder (MDD) who have reduced plasticity in response to TMS [160] and visual evoked potentials [161].

As in PD although there is ample evidence for altered LTP- and LTD-like plasticity in SCZ and MDD, direct examples of impaired homeostatic plasticity are rare. On a molecular level, evidence exists linking various psychiatric diseases such as SCZ, MDD and other disorders to dysfunctional homeostatic synaptic plasticity involving a wide array of genes and molecules required for homeostatic synaptic plasticity [155,156]. However, even though these molecular findings have led to a conceptual framework that places homeostatic dysfunction at the heart of a wide array of neurologic and psychiatric diseases there is, to the authors knowledge, no direct investigation of homeostatic regulation in psychiatric patient populations. Considering the links between the pathophysiology of a variety of psychiatric disorders and synaptic processes necessary for homeostatic control, it will be a future challenge to understand how these mechanisms work together in the intact human brain.

A systematic investigation of homeostatic plasticity in various psychiatric disorders will help to start understanding how homeostatic responses orchestrates systemic functions in the brain.

Dysfunctional synaptic plasticity and homeostatic plasticity in various disorders could have an impact on the design of future clinical trials. At the moment, treatment trials for several psychiatric disorders involve the application of plasticity-inducing NTBS protocols to counteract hypo- or hyperactivity of different brain areas. If, indeed, plasticity in these disorders is fundamentally changed, we cannot assume that the plasticity-enhancing effect of brain stimulation techniques, observed in healthy subjects, can be directly translated to patient populations. Indeed Barr et al. showed that one session of 20 Hz rTMS had opposing effects in SCZ patients and healthy volunteers: rTMS inhibited gamma—oscillatory activity in patients, who had a greater activity at baseline, while the same rTMS protocol potentiated gamma—oscillatory activity in healthy controls with relatively lower oscillations at baseline, suggesting a homeostatic interaction [162].

Conclusions and perspectives

Homeostatic metaplasticity plays a critical role in stabilizing neural activity around a set point and is defined by inducing a shift in the stimulus—response curve of the firing neuron and is controlled by the intra-cellular Ca²⁺ levels. The use of NTBS allows homeostatic effects to be investigated on a systems level and in interaction with physiological conditions. Since NTBS activates a massive number of neurons, inducing action potentials in a mixture of inhibitory and excitatory cells, NTBS-induced plasticity cannot be equated with *in vitro* studies on synaptic plasticity. Additionally, the traditional measure of NTBS-induced excitability, the MEP, has confined most investigations of homeostatic effects in the intact human brain to the primary motor cortex.

In the future a combination of NTBS with other brain mapping techniques will allow investigation of homeostatic phenomena to expand to cortical areas outside M1. A careful investigation of the network effects and the combination of NTBS with neuroimaging, pharmacology and animal studies will help to reveal more insights into the neural mechanisms underlying homeostasis at a systems level.

Systematic investigation of individual differences in NTBS response will, in the future, allow researchers to move toward the use of individually adjusted protocols that take relevant neurophysiological state markers into consideration. These custom made protocols may decrease inter-individual variance and make NTBS an even more powerful tool. The study of homeostatic plasticity in patients with neurological and psychiatric diseases is still very limited and future research should tackle this issue since it might give some insight into contribution of dysfunctional regulation of cortical plasticity to these conditions.

Acknowledgments

Thanks to Ms. Friederike Irmen for help with the table.

References

- Abbott LF, Nelson SB. Synaptic plasticity: taming the beast. Nat Neurosci 2000;(3 Suppl):1178–83.
- [2] Turrigiano GG, Nelson SB. Homeostatic plasticity in the developing nervous system. Nat Rev Neurosci 2004;5(2):97–107.
- [3] Abraham WC. Metaplasticity: tuning synapses and networks for plasticity. Nat Rev Neurosci 2008;9(5):387.
- [4] Hulme SR, Jones OD, Abraham WC. Emerging roles of metaplasticity in behaviour and disease. Trends Neurosci 2013;36(6):353–62.
- [5] Karabanov A, Ziemann U, Classen J, Siebner H. Understanding Homeostatic Plasticity. In: Miniussi C, Paulus W, Rossini P, editors. Transcranial Brain Stimulation. Frontiers in Neuroscience: CRC Press; 2012. p. 230–44.

- [6] Classen J. Plasticity. Handb Clin Neurol 2013;116:525-34.
- [7] Ziemann U, Siebner HR. Modifying motor learning through gating and homeostatic metaplasticity. Brain Stimul 2008;1(1):60–6.
- [8] Ziemann U, Ilic TV, Pauli C, Meintzschel F, Ruge D. Learning modifies subsequent induction of long-term potentiation-like and long-term depressionlike plasticity in human motor cortex. J Neurosci 2004;24(7):1666-72.
- [9] Groppa S, Oliviero A, Eisen A, et al. A practical guide to diagnostic transcranial magnetic stimulation: report of an IFCN committee. Clin Neurophysiol 2012;123(5):858–82.
- [10] Siebner HR, Rothwell J. Transcranial magnetic stimulation: new insights into representational cortical plasticity. Exp Brain Res 2003;148(1):1–16.
- [11] Pascual-Leone A, Valls-Sole J, Wassermann EM, Hallett M. Responses to rapid-rate transcranial magnetic stimulation of the human motor cortex. Brain 1994;117(Pt 4):847–58.
- [12] Huang YZ, Edwards MJ, Rounis E, Bhatia KP, Rothwell JC. Theta burst stimulation of the human motor cortex. Neuron 2005;45(2):201–6.
- [13] Thickbroom GW, Byrnes ML, Edwards DJ, Mastaglia FL. Repetitive pairedpulse TMS at I-wave periodicity markedly increases corticospinal excitability: a new technique for modulating synaptic plasticity. Clin Neurophysiol 2006;117(1):61–6.
- [14] Stefan K, Kunesch E, Cohen LG, Benecke R, Classen J. Induction of plasticity in the human motor cortex by paired associative stimulation. Brain 2000;123(Pt 3):572–84.
- [15] Hamada M, Hanajima R, Terao Y, et al. Quadro-pulse stimulation is more effective than paired-pulse stimulation for plasticity induction of the human motor cortex. Clin Neurophysiol 2007;118(12):2672–82.
- [16] Nitsche MA, Cohen LG, Wassermann EM, et al. Transcranial direct current stimulation: State of the art 2008. Brain Stimul 2008;1:206–23.
- [17] Hamada M, Murase N, Hasan A, Balaratnam M, Rothwell JC. The role of interneuron networks in driving human motor cortical plasticity. Cereb Cortex 2013;23(7):1593–605.
- [18] Feldman DE. Synaptic mechanisms for plasticity in neocortex. Annu Rev Neurosci 2009;32:33–55.
- [19] Sjostrom PJ, Rancz EA, Roth A, Hausser M. Dendritic excitability and synaptic plasticity. Physiol Rev 2008;88(2):769–840.
- [20] Foeller E, Feldman DE. Synaptic basis for developmental plasticity in somatosensory cortex. Curr Opin Neurobiol 2004;14(1):89–95.
- [21] Hensch TK. Critical period plasticity in local cortical circuits. Nat Rev Neurosci 2005;6(1):877–88.
- [22] Lisman J, Lichtman JW, Sanes JR. LTP: perils and progress. Nat Rev Neurosci 2003;4(1):926–9.
- [23] Malenka RC, Bear MF. LTP and LTD: an embarrassment of riches. Neuron 2004;44(1):5-21.
- [24] Turrigiano GG, Nelson SB. Hebb and homeostasis in neuronal plasticity. Curr Opin Neurobiol 2000;10(3):358–64.
- [25] Hebb D. The Organization of Behavior. New York: Wiley & Sons; 1949.
- [26] Kim SJ, Linden DJ. Ubiquitous plasticity and memory storage. Neuron 2007;56(4):582–92.
- [27] Abraham WC, Bear MF. Metaplasticity: the plasticity of synaptic plasticity. Trends Neurosci 1996;19(4):126–30.
- [28] Alvarez VA, Sabatini BL. Anatomical and physiological plasticity of dendritic spines. Annu Rev Neurosci 2007;30:79–97.
- [29] Shouval HZ, Bear MF, Cooper LN. A unified model of NMDA receptordependent bidirectional synaptic plasticity. Proc Natl Acad Sci U S A 2002;99(16):10831–6.
- [30] Turrigiano GG. The self-tuning neuron: synaptic scaling of excitatory synapses. Cell 2008;135(3):422–35.
- [31] Tsumoto T. Long-term potentiation and long-term depression in the neocortex. Prog Neurobiol 1992;39(2):209–28.
- [32] Lisman J. A mechanism for the Hebb and the anti-Hebb processes underlying learning and memory. Proc Natl Acad Sci U S A 1989;86(23):9574–8.
- [33] Artola A, Singer W. Long-term depression of excitatory synaptic transmission and its relationship to long-term potentiation. Trends Neurosci 1993;16(11):480–7.
- [34] Yang SN, Tang YG, Zucker RS. Selective induction of LTP and LTD by postsynaptic [Ca²⁺]i elevation. J Neurophysiol 1999;81(2):781–7.
- [35] Artola A, Brocher S, Singer W. Different voltage-dependent thresholds for inducing long-term depression and long-term potentiation in slices of rat visual cortex. Nature 1990;347(6288):69–72.
- [36] Bear MF. Bidirectional synaptic plasticity: from theory to reality. Philos Trans R Soc Lond B Biol Sci 2003;358(1432):649–55.
- [37] Turrigiano G. Too many cooks? Intrinsic and synaptic homeostatic mechanisms in cortical circuit refinement. Annu Rev Neurosci 2011;34:89–103.
- [38] Turrigiano G. Homeostatic synaptic plasticity: local and global mechanisms for stabilizing neuronal function. Cold Spring Harb Perspect Biol 2012;4(1):a005736.
- [39] Nelson SB, Turrigiano GG. Strength through diversity. Neuron 2008;60(3):477–82.
- [40] Cooper LN, Bear MF. The BCM theory of synapse modification at 30: interaction of theory with experiment. Nat Rev Neurosci 2012;13(11):798-810.
- [41] Bienenstock EL, Cooper LN, Munro PW. Theory for the development of neuron selectivity: orientation specificity and binocular interaction in visual cortex. J Neurosci 1982;2(1):32–48.
- [42] Kirkwood A, Rioult MC, Bear MF. Experience-dependent modification of synaptic plasticity in visual cortex. Nature 1996;381(6582):526-8.

- [43] Wang H, Wagner JJ. Priming-induced shift in synaptic plasticity in the rat hippocampus. J Neurophysiol 1999;82(4):2024–8.
- [44] Hamada M, Hanajima R, Terao Y, et al. Primary motor cortical metaplasticity induced by priming over the supplementary motor area. J Physiol 2009;587(Pt 20):4845–62.
- [45] Hamada M, Ugawa Y. Quadripulse stimulation—a new patterned rTMS. Restor Neurol Neurosci 2010;28(4):419—24.
- [46] Hess G, Aizenman CD, Donoghue JP. Conditions for the induction of longterm potentiation in layer II/III horizontal connections of the rat motor cortex. J Neurophysiol 1996;75(5):1765–78.
- [47] Castro-Alamancos MA, Donoghue JP, Connors BW. Different forms of synaptic plasticity in somatosensory and motor areas of the neocortex. J Neurosci 1995;15(7 Pt 2):5324–33.
- [48] Rioult-Pedotti MS, Friedman D, Donoghue JP. Learning-induced LTP in neocortex. Science 2000;290(5491):533–6.
- [49] Massey PV, Bashir ZI. Long-term depression: multiple forms and implications for brain function. Trends Neurosci 2007;30(4):176–84.
- [50] Cooke SF, Bliss TV. Plasticity in the human central nervous system. Brain 2006;129(Pt 7):1659–73.
- [51] Sharma N, Classen J, Cohen LG. Neural plasticity and its contribution to functional recovery. Handb Clin Neurol 2013;110(2):3–12.
- [52] Takano B, Drzezga A, Peller M, et al. Short-term modulation of regional excitability and blood flow in human motor cortex following rapid-rate transcranial magnetic stimulation. Neuroimage 2004;23(3):849–59.
- [53] Quartarone A, Siebner HR, Rothwell JC. Task-specific hand dystonia: can too much plasticity be bad for you? Trends Neurosci 2006;29(4):192–9.
- [54] Hallett M. Transcranial magnetic stimulation: a primer. Neuron 2007;55(2): 187–99.
- [55] Chen R, Classen J, Gerloff C, et al. Depression of motor cortex excitability by low-frequency transcranial magnetic stimulation. Neurology 1997;48(5): 1398–403.
- [56] Sommer M, Tergau F, Wischer S, Paulus W. Paired-pulse repetitive transcranial magnetic stimulation of the human motor cortex. Exp Brain Res 2001;139(4):465–72.
- [57] Cash RF, Murakami T, Chen R, Thickbroom GW, Ziemann U. Augmenting Plasticity Induction in Human Motor Cortex by Disinhibition Stimulation. Cereb Cortex 2014 [Epub ahead of print].
- [58] Nyffeler T, Wurtz P, Luscher HR, et al. Extending lifetime of plastic changes in the human brain. Eur J Neurosci 2006;24(10):2961–6.
- [59] Hamada M, Terao Y, Hanajima R, et al. Bidirectional long-term motor cortical plasticity and metaplasticity induced by quadripulse transcranial magnetic stimulation. J Physiol 2008;586(16):3927–47.
- [60] Classen J, Wolters A, Stefan K, et al. Paired associative stimulation. Suppl Clin Neurophysiol 2004;57:563–9.
- [61] Rizzo V, Siebner HS, Morgante F, Mastroeni C, Girlanda P, Quartarone A. Paired associative stimulation of left and right human motor cortex shapes interhemispheric motor inhibition based on a Hebbian mechanism. Cereb Cortex 2009;19(4):907–15.
- [62] Chao CC, Karabanov AN, Paine R, et al. Induction of Motor Associative Plasticity in the Posterior Parietal Cortex-Primary Motor Network. Cereb Cortex 2015;25:365–73.
- [63] Arai N, Muller-Dahlhaus F, Murakami T, et al. State-dependent and timingdependent bidirectional associative plasticity in the human SMA-M1 network. J Neurosci 2011;31(43):15376–83.
- [64] Buch ER, Johnen VM, Nelissen N, O'Shea J, Rushworth MF. Noninvasive associative plasticity induction in a corticocortical pathway of the human brain. J Neurosci 2011;31(48):17669–79.
- [65] Funke K, Benali A. Modulation of cortical inhibition by rTMS findings obtained from animal models. J Physiol 2011;589(Pt 18):4423–35.
- [66] Pell GS, Roth Y, Zangen A. Modulation of cortical excitability induced by repetitive transcranial magnetic stimulation: influence of timing and geometrical parameters and underlying mechanisms. Prog Neurobio 2011;93(1):59–98.
- [67] Carson RG, Kennedy NC. Modulation of human corticospinal excitability by paired associative stimulation. Front Hum Neurosci 2013;7:823.
- [68] Ziemann U, Paulus W, Nitsche MA, et al. Consensus: Motor cortex plasticity protocols. Brain Stimul 2008;1(3):164–82.
- [69] Muller-Dahlhaus F, Ziemann U. Metaplasticity in Human Cortex. Neuroscientist 2014;21:185–202.
- [70] Siebner HR, Lang N, Rizzo V, et al. Preconditioning of low-frequency repetitive transcranial magnetic stimulation with transcranial direct current stimulation: evidence for homeostatic plasticity in the human motor cortex. J Neurosci 2004;24(13):3379–85.
- [71] Iyer MB, Schleper N, Wassermann EM. Priming stimulation enhances the depressant effect of low-frequency repetitive transcranial magnetic stimulation. J Neurosci 2003;23(34):10867–72.
- [72] Lang N, Siebner HR, Ernst D, et al. Preconditioning with transcranial direct current stimulation sensitizes the motor cortex to rapid-rate transcranial magnetic stimulation and controls the direction of after-effects. Biol Psychiatry 2004;56(9):634–9.
- [73] Muller JF, Orekhov Y, Liu Y, Ziemann U. Homeostatic plasticity in human motor cortex demonstrated by two consecutive sessions of paired associative stimulation. Eur J Neurosci 2007;25(11):3461–8.

- [74] Nitsche MA, Roth A, et al. Timing-dependent modulation of associative plasticity by general network excitability in the human motor cortex. J Neurosci 2007;27(14):3807–12.
- [75] Todd G, Flavel SC, Ridding MC. Priming theta-burst repetitive transcranial magnetic stimulation with low- and high-frequency stimulation. Exp Brain Res 2009;195(2):307–15.
- [76] Ni Z, Gunraj C, Kailey P, Cash RF, Chen R. Heterosynaptic modulation of motor cortical plasticity in human. J Neurosci 2014;34(21):7314–21.
- [77] Gentner R, Wankerl K, Reinsberger C, Zeller D, Classen J. Depression of human corticospinal excitability induced by magnetic theta-burst stimulation: evidence of rapid polarity-reversing metaplasticity. Cereb Cortex 2008;18(9):2046–53.
- [78] Gamboa OL, Antal A, Moliadze V, Paulus W. Simply longer is not better: reversal of theta burst after-effect with prolonged stimulation. Exp Brain Res 2010;204(2):181–7.
- [79] Rothkegel H, Sommer M, Paulus W. Breaks during 5Hz rTMS are essential for facilitatory after effects. Clin Neurophysiol 2010;121(3):426–30.
- [80] Fricke K, Seeber AA, Thirugnanasambandam N, Paulus W, Nitsche MA, Rothwell JC. Time course of the induction of homeostatic plasticity generated by repeated transcranial direct current stimulation of the human motor cortex. J Neurophysiol 2011;105(3):1141–9.
- [81] Karabanov A, Siebner HR. Unravelling homeostatic interactions in inhibitory and excitatory networks in human motor cortex. J Physiol 2012;590(Pt 22):5557–8.
- [82] Di Lazzaro V, Ziemann U, Lemon RN. State of the art: Physiology of transcranial motor cortex stimulation. Brain Stimul 2008;1(4):345–62.
- [83] Reis J, Swayne OB, Vandermeeren Y, et al. Contribution of transcranial magnetic stimulation to the understanding of cortical mechanisms involved in motor control. J Physiol 2008;586(2):325–51.
- [84] Classen J, Liepert J, Wise SP, Hallett M, Cohen LG. Rapid plasticity of human cortical movement representation induced by practice. J Neurophysiol 1998;79(2):1117–23.
- [85] Rosenkranz K, Kacar A, Rothwell JC. Differential modulation of motor cortical plasticity and excitability in early and late phases of human motor learning. J Neurosci 2007;27(22):12058–66.
- [86] Doeltgen SH, Ridding MC. Low-intensity, short-interval theta burst stimulation modulates excitatory but not inhibitory motor networks. Clin Neurophysiol 2011;122(7):1411–6.
- [87] Kujirai T, Caramia MD, Rothwell JC, et al. Corticocortical inhibition in human motor cortex. J Physiol 1993;471:501–19.
- [88] Murakami T, Muller-Dahlhaus F, Lu MK, Ziemann U. Homeostatic metaplasticity of corticospinal excitatory and intracortical inhibitory neural circuits in human motor cortex. J Physiol 2012;590(Pt 22):5765–81.
- [89] Potter-Nerger M, Fischer S, Mastroeni C, et al. Inducing homeostatic-like plasticity in human motor cortex through converging corticocortical inputs. J Neurophysiol 2009;102(6):3180–90.
- [90] Ragert P, Camus M, Vandermeeren Y, Dimyan MA, Cohen LG. Modulation of effects of intermittent theta burst stimulation applied over primary motor cortex (M1) by conditioning stimulation of the opposite M1. J Neurophysiol 2009;102(2):766-73.
- [91] Bliem B, Muller-Dahlhaus JF, Dinse HR, Ziemann U. Homeostatic metaplasticity in the human somatosensory cortex. J Cogn Neurosci 2008;20(8):1517–28.
- [92] Gatica Tossi MA, Stude P, Schwenkreis P, Tegenthoff M, Dinse HR. Behavioural and neurophysiological markers reveal differential sensitivity to homeostatic interactions between centrally and peripherally applied passive stimulation. Eur J Neurosci 2013;38(6):2893–901.
- [93] Bocci T, Caleo M, Tognazzi S, et al. Evidence for metaplasticity in the human visual cortex. J Neural Transm 2014;121(3):221–31.
- [94] Rajji TK, Sun Y, Zomorrodi-Moghaddam R, et al. PAS-induced potentiation of cortical-evoked activity in the dorsolateral prefrontal cortex. Neuropsychopharmacology 2013;38(12):2545–52.
- [95] Lepage JF, Morin-Moncet O, Beaule V, de Beaumont L, Champoux F, Theoret H. Occlusion of LTP-like plasticity in human primary motor cortex by action observation. PLoS One 2012;7(6):e38754.
- [96] Cantarero G, Lloyd A, Celnik P. Reversal of long-term potentiation-like plasticity processes after motor learning disrupts skill retention. J Neurosci 2013;33(31):12862–9.
- [97] Cantarero G, Tang B, O'Malley R, Salas R, Celnik P. Motor learning interference is proportional to occlusion of LTP-like plasticity. J Neurosci 2013;33(11):4634–41.
- [98] Elahi B, Hutchison WD, Daskalakis ZJ, Gunraj C, Chen R. Dose-response curve of associative plasticity in human motor cortex and interactions with motor practice. J Neurophysiol 2014;111(3):594–601.
- [99] Jung P, Ziemann U. Homeostatic and nonhomeostatic modulation of learning in human motor cortex. J Neurosci 2009;29(17):5597–604.
- [100] Teo JT, Swayne OB, Cheeran B, Greenwood RJ, Rothwell JC. Human theta burst stimulation enhances subsequent motor learning and increases performance variability. Cereb Cortex 2011;21(7):1627–38.
- [101] Kuo MF, Unger M, Liebetanz D, et al. Limited impact of homeostatic plasticity on motor learning in humans. Neuropsychologia 2008;46(8):2122–8.
- [102] Nitsche MA, Schauenburg A, Lang N, et al. Facilitation of implicit motor learning by weak transcranial direct current stimulation of the primary motor cortex in the human. J Cogn Neurosci 2003;15(4):619–26.

- [103] Antal A, Nitsche MA, Kincses TZ, Kruse W, Hoffmann KP, et al. Facilitation of visuo-motor learning by transcranial direct current stimulation of the motor and extrastriate visual areas in humans. Eur J Neurosci 2004;19(10):2888–92.
- [104] Reis J, Schambra HM, Cohen LG, et al. Noninvasive cortical stimulation enhances motor skill acquisition over multiple days through an effect on consolidation. Proc Natl Acad Sci U S A 2009;106(5):1590–5.
- [105] Reis J, Fritsch B. Modulation of motor performance and motor learning by transcranial direct current stimulation. Curr Opin Neurol 2011;24(6):590–6.
- [106] Schambra HM, Abe M, Luckenbaugh DA, Reis J, Krakauer JW, et al. Probing for hemispheric specialization for motor skill learning: a transcranial direct current stimulation study. J Neurophysiol 2011;106(2):652–61.
- [107] Stagg CJ, Jayaram G, Pastor D, Kincses ZT, Matthews PM, et al. Polarity and timing-dependent effects of transcranial direct current stimulation in explicit motor learning. Neuropsychologia 2011;49(5):800–4.
- [108] Todd G, Rogasch NC, Flavel SC, Ridding MC. Voluntary movement and repetitive transcranial magnetic stimulation over human motor cortex. J Appl Physiol 2009;106(5):1593–603.
- [109] Huang YZ, Rothwell JC, Edwards MJ, Chen RS. Effect of physiological activity on an NMDA-dependent form of cortical plasticity in human. Cereb Cortex 2008;18(3):563–70.
- [110] Fujiwara T, Rothwell JC. The after effects of motor cortex rTMS depend on the state of contraction when rTMS is applied. Clin Neurophysiol 2004;115(7): 1514–8.
- [111] Rosenkranz K, Seibel J, Kacar A, Rothwell J. Sensorimotor deprivation induces interdependent changes in excitability and plasticity of the human hand motor cortex. J Neurosci 2014;34(21):7375–82.
- [112] Ziemann U, Wittenberg GF, Cohen LG. Stimulation-induced within-representation and across-representation plasticity in human motor cortex. J Neurosci 2002;22(13):5563–71.
- [113] Delvendahl I, Jung NH, Mainberger F, Kuhnke NG, Cronjaeger M, Mall V. Occlusion of bidirectional plasticity by preceding low-frequency stimulation in the human motor cortex. Clin Neurophysiol 2010;121(4):594–602.
- [114] Siebner HR. A primer on priming the human motor cortex. Clin Neurophysiol 2010;121(4):461–3.
- [115] Larson J, Xiao P, Lynch G. Reversal of LTP by theta frequency stimulation. Brain Res 1993;600(1):97–102.
- [116] Kulla A, Manahan-Vaughan D. Depotentiation in the dentate gyrus of freely moving rats is modulated by D1/D5 dopamine receptors. Cereb Cortex 2000;10(6):614–20.
- [117] Huang CC, Liang YC, Hsu KS. Characterization of the mechanism underlying the reversal of long term potentiation by low frequency stimulation at hippocampal CA1 synapses. J Biol Chem 2001;276(51):48108–17.
- [118] Huang YZ, Rothwell JC, Lu CS, Chuang WL, Lin WY, Chen RS. Reversal of plasticitylike effects in the human motor cortex. J Physiol 2010;588(Pt 19):3683–93.
- [119] Goldsworthy MR, Muller-Dahlhaus F, Ridding MC, Ziemann U. Resistant Against De-depression: LTD-Like Plasticity in the Human Motor Cortex Induced by Spaced cTBS. Cereb Cortex 2015;25:1724–34.
- [120] Yeung LC, Shouval HZ, Blais BS, Cooper LN. Synaptic homeostasis and input selectivity follow from a calcium-dependent plasticity model. Proc Natl Acad Sci U S A 2004;101(41):14943–8.
- [121] Hulme SR, Jones OD, Ireland DR, Abraham WC. Calcium-dependent but action potential-independent BCM-like metaplasticity in the hippocampus. J Neurosci 2012;32(20):6785–94.
- [122] Mizuno T, Kanazawa I, Sakurai M. Differential induction of LTP and LTD is not determined solely by instantaneous calcium concentration: an essential involvement of a temporal factor. Eur J Neurosci 2001;14(4):701–8.
- [123] Cummings JA, Mulkey RM, Nicoll RA, Malenka RC. Ca²⁺ signaling requirements for long-term depression in the hippocampus. Neuron 1996;16(4):825–33.
- [124] Hirsch JC, Crepel F. Blockade of NMDA receptors unmasks a long-term depression in synaptic efficacy in rat prefrontal neurons in vitro. Exp Brain Res 1991;85(3):621–4.
- [125] Wankerl K, Weise D, Gentner R, Rumpf JJ, Classen J. L-type voltage-gated Ca²⁺ channels: a single molecular switch for long-term potentiation/longterm depression-like plasticity and activity-dependent metaplasticity in humans. | Neurosci 2010;30(18):6197–204.
- [126] Pozo K, Goda Y. Unraveling mechanisms of homeostatic synaptic plasticity. Neuron 2010;66(3):337–51.
- [127] Tononi G, Cirelli C. Sleep and the price of plasticity: from synaptic and cellular homeostasis to memory consolidation and integration. Neuron 2014;81(1):12–34.
- [128] Gilestro GF, Tononi G, Cirelli C. Widespread changes in synaptic markers as a function of sleep and wakefulness in Drosophila. Science 2009;324(5923): 109–12.
- [129] Bushey D, Tononi G, Cirelli C. Sleep and synaptic homeostasis: structural evidence in Drosophila. Science 2011;332(6037):1576–81.
- [130] Vyazovskiy VV, Cirelli C, Pfister-Genskow M, Faraguna U, Tononi G. Molecular and electrophysiological evidence for net synaptic potentiation in wake and depression in sleep. Nat Neurosci 2008;11(2):200–8.
 [131] Civardi C, Boccagni C, Vicentini R, et al. Cortical excitability and sleep
- [131] Civardi C, Boccagni C, Vicentini R, et al. Cortical excitability and sleep deprivation: a transcranial magnetic stimulation study. J Neurol Neurosurg Psychiatry 2001;71(6):809–12.
- [132] Kreuzer P, Langguth B, Popp R, et al. Reduced intra-cortical inhibition after sleep deprivation: a transcranial magnetic stimulation study. Neurosci Lett 2011;493(3):63–6.

- [133] Sale MV, Ridding MC, Nordstrom MA. Cortisol inhibits neuroplasticity induction in human motor cortex. J Neurosci 2008;28(33):8285–93.
- [134] Grunwald ME, Mellem JE, Strutz N, Maricq AV, Kaplan JM. Clathrin-mediated endocytosis is required for compensatory regulation of GLR-1 glutamate receptors after activity blockade. Proc Natl Acad Sci U S A 2004;101(9):3190–5.
- [135] Marder E, Goaillard JM. Variability, compensation and homeostasis in neuron and network function. Nat Rev Neurosci 2006;7(7):563–74.
- [136] Vitureira N, Letellier M, Goda Y. Homeostatic synaptic plasticity: from single synapses to neural circuits. Curr Opin Neurobiol 2012;22(3):516–21.
- [137] Quartarone A, Pisani A. Abnormal plasticity in dystonia: Disruption of synaptic homeostasis. Neurobiology 2011;42(2):162–70.
- [138] Quartarone A, Rizzo V, Bagnato S, et al. Homeostatic-like plasticity of the primary motor hand area is impaired in focal hand dystonia. Brain 2005;128(Pt 8):1943–50.
- [139] Kang JS, Terranova C, Hilker R, Quartarone A, Ziemann U. Deficient homeostatic regulation of practice-dependent plasticity in writer's cramp. Cereb Cortex 2011;21(5):1203–12.
- [140] Quartarone A, Sant'Angelo A, Battaglia F, et al. Enhanced long-term potentiation-like plasticity of the trigeminal blink reflex circuit in blepharospasm. J Neurosci 2006;26(2):716–21.
- [141] Weise D, Schramm A, Beck M, Reiners K, Classen J. Loss of topographic specificity of LTD-like plasticity is a trait marker in focal dystonia. Neurobiol Dis 2011;42(2):171–6.
- [142] Sadnicka A, Hamada M, Bhatia KP, Rothwell JC, Edwards MJ. A reflection on plasticity research in writing dystonia. Mov Disord 2014;29(8):980–7.
- [143] Siebner HR, Tormos JM, Ceballos-Baumann AO, et al. Low-frequency repetitive transcranial magnetic stimulation of the motor cortex in writer's cramp. Neurology 1999;52(3):529–37.
- [144] Belvisi D, Suppa A, Marsili L, et al. Abnormal experimentally- and behaviorally-induced LTP-like plasticity in focal hand dystonia. Exp Neurol 2013;240:64–74.
- [145] Koch G. Do studies on cortical plasticity provide a rationale for using noninvasive brain stimulation as a treatment for Parkinson's disease patients? Front Neurol 2013;4:180.
- [146] Morgante F, Espay AJ, Gunraj C, Lang AE, Chen R. Motor cortex plasticity in Parkinson's disease and levodopa-induced dyskinesias. Brain 2006;129(Pt 4):1059–69.
- [147] Bagnato S, Agostino R, Modugno N, Quartarone A, Berardelli A. Plasticity of the motor cortex in Parkinson's disease patients on and off therapy. Mov Disord 2006;21(5):639–45.
- [148] Huang YZ, Rothwell JC, Lu CS, Chuang WL, Chen RS. Abnormal bidirectional plasticity-like effects in Parkinson's disease. Brain 2011;134(Pt 8):2312–20.
- [149] Radhu N, Ravindran LN, Levinson AJ, Daskalakis ZJ. Inhibition of the cortex using transcranial magnetic stimulation in psychiatric populations: current and future directions. J Psychiatry Neurosci 2012;37(6):369–78.
- [150] Daskalakis ZJ, Christensen BK, Chen R, Fitzgerald PB, Zipursky RB, et al. Evidence for impaired cortical inhibition in schizophrenia using transcranial magnetic stimulation. Arch Gen Psychiatry 2002;59(4):347–54.
- [151] Benes FM, Berretta S. GABAergic interneurons: implications for understanding schizophrenia and bipolar disorder. Neuropsychopharmacology 2001;25(1):1–27.
- [152] Fitzgerald PB, Brown TL, Marston NA, et al. Reduced plastic brain responses in schizophrenia: a transcranial magnetic stimulation study. Schizophr Res 2004;71(1):17–26.
- [153] Greenberg BD, Ziemann U, Harmon A, Murphy DL, Wassermann EM. Decreased neuronal inhibition in cerebral cortex in obsessive-compulsive disorder on transcranial magnetic stimulation. Lancet 1998;352(9131): 881–2.
- [154] Stefansson H, Sarginson J, Kong A, et al. Association of neuregulin 1 with schizophrenia confirmed in a Scottish population. Am J Hum Genet 2003;72(1):83–7.
- [155] Duman RS, Aghajanian GK. Synaptic dysfunction in depression: potential therapeutic targets. Science 2012;338(6103):68–72.
- [156] Wondolowski J, Dickman D. Emerging links between homeostatic synaptic plasticity and neurological disease. Front Cell Neurosci 2013;7:223.
- [157] Hasan A, Nitsche MA, Herrmann M, et al. Impaired long-term depression in schizophrenia: a cathodal tDCS pilot study. Brain Stimul 2012;5(4): 475–83.
- [158] Hasan A, Nitsche MA, Rein B, et al. Dysfunctional long-term potentiation-like plasticity in schizophrenia revealed by transcranial direct current stimulation. Behav Brain Res 2011;224(1):15–22.
- [159] Daskalakis ZJ, Christensen BK, Fitzgerald PB, Chen R. Dysfunctional neural plasticity in patients with schizophrenia. Arch Gen Psychiatry 2008;65(4):378–85.
- [160] Player MJ, Taylor JL, Weickert CS, et al. Neuroplasticity in depressed individuals compared with healthy controls. Neuropsychopharmacology 2013;38(11):2101–8.
- [161] Normann C, Schmitz D, Furmaier A, Doing C, Bach M. Long-term plasticity of visually evoked potentials in humans is altered in major depression. Biol Psychiatry 2007;62(5):373–80.
- [162] Barr MS, Farzan F, Arenovich T, Chen R, Fitzgerald PB, et al. The effect of repetitive transcranial magnetic stimulation on gamma oscillatory activity in schizophrenia. PLoS One 2011;6:e22627.