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Genetic testing for Ebstein anomaly

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Abstract

Ebstein anomaly (EA) is a rare congenital tricuspid valve malformation, characterized by downward displacement of the septal leaflet and an atrialized right ventricle. About 80% of cases of EA are non-syndromic; in the other 20%, the anomaly is associated with a chromosomal or Mendelian syndrome. The prevalence of EA is estimated at about 1 per 20,000 live births, and accounts for less than 1% of all congenital heart defects. EA has autosomal dominant inheritance. Likely causative genes are: NKX2-5, MYH7 and TPM1. This Utility Gene Test was developed on the basis of an analysis of the literature and existing diagnostic protocols. It is useful for confirming diagnosis, as well as for differential diagnosis, potential risk assessment and access to clinical trials.

Keywords: Ebstein anomaly, EBTNA UTILITY GENE TEST

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General information about the disease

Ebstein anomaly (EA) is a rare congenital tricuspid valve malformation, characterized by downward displacement of the septal leaflet and an atrialized right ventricle with great variability in clinical, anatomical, echocardiographic and electrocardiographic features, all of which influence prognosis. It may manifest with severe symptoms at birth or may remain asymptomatic until adulthood. It is sometimes associated with septal defects, transposition of large vessels, preexcitation syndromes, or left ventricular non-compaction (1, 2). Isolated forms of EA are non-syndromic, but 20% also have extracardiac defects associated with chromosomal and Mendelian genetic disorders, such as Apert, Noonan, CHARGE, Holt-Oram, Cornelia de Lange, and Kabuki syndromes and VACTERL association (3).

Morphological characteristics of EA include: adherence of the septal and posterior leaflets to the underlying myocardium; downward (apical) displacement of the functional annulus; dilation of the "atrialized" portion of the right ventricle, with various degrees of hypertrophy and thinning of the wall; redundancy, fenestrations and tethering of the anterior leaflet; dilation of the right atrioventricular junction (true tricuspid annulus) (4,5).

Patients with EA have conduction system abnormalities, which are at least partly due to compression of the atrioventricular node by the septal malformation, accessory pathways, and abnormalities of the right bundle branch. Right bundle branch block is frequent in humans with EA and this block has also been noted in animals with EA (6-8).

The prevalence of EA is estimated at about 1 in 20,000 live births, with equal occurrence in males and females. It accounts for less than 1% of all congenital heart defects (9,10). Advances in diagnostics now make it possible to detect EA in the fetal or neonatal period. Prognosis is poor: mortality is 29% at 1 year, 34% at 10 years, and 56% at 30 years (11, 12).

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The diagnostic work-up includes clinical assessment to identify symptoms, echocardiogram, electrocardiogram, chest radiogram, integration of septal and color Doppler with two-dimensional echocardiography, spin-echo MR imaging, CT, diagnostic catheterization and genetic testing.

Differential diagnosis should consider EA caused by Down syndrome and other chromosomal disorders.

Non-syndromic EA has autosomal dominant inheritance with variable expressivity and incomplete penetrance.

Candidate genes for EA (OMIM disease 224700)

- NKX2-5 (OMIM gene 600584) (10)
- MYH7 (OMIM gene 160760) (10)
- TPM1 (OMIM gene 191010) (13)

In humans, mutations in the *NKX2-5* gene provided the first evidence that genetic factors are etiologically crucial in non-syndromic congenital heart diseases (14). The most common phenotypes are secundum atrial septal defect and atrioventricular conduction disturbance, but cases of EA are also reported (15, 16).

MYH7 mutations are predominantly found in EA associated with left ventricular non-compaction cardiomyopathy (LVNC) (17); a convincing hypothesis regarding this association is that LVNC is the result of altered cell regulation during wall formation (18) and EA may be caused by an arrest in directional growth during this process (19, 20).

TPM1 encodes α -tropomyosin, which is involved in myocardial contraction, as well as in stabilization of nonmuscle cytoskeletal actin filaments. Nijak et al. reported an LVNC-EA family with a TPM1 mutation, and established an association between EA and TPM1-related LVNC (21).

Pathogenic variants may include missense and nonsense mutations and small deletions.

Aims of the test

- To determine the gene defect responsible for the disease;
- To confirm clinical diagnosis;
- To assess the recurrence risk and perform genetic counselling for at-risk/affected individuals.

Test characteristics

Specialist centers/ Published Guidelines

The test is listed in the Orphanet database and is offered by 2 accredited medical genetic laboratories in the EU, and in the GTR database, offered by 11 accredited medical genetic laboratories in the US.

Guidelines for clinical use of the test are described in Genetics Home Reference (ghr.nlm.nih.gov).

Test strategy

A multi-gene next generation sequencing panel is used for the detection of nucleotide variations in coding exons and flanking introns of the above genes.

Potentially causative variants and regions with low cover-

age are Sanger-sequenced. Sanger sequencing is also used for family segregation studies.

To perform molecular diagnosis, a single sample of biological material is normally sufficient. This may be 1 ml peripheral blood in a sterile tube with 0.5 ml $\rm K_3EDTA$ or 1 ml saliva in a sterile tube with 0.5 ml ethanol 95%. Sampling rarely has to be repeated.

Gene-disease associations and the interpretation of genetic variants are rapidly developing fields. It is therefore possible that the genes mentioned in this note may change as new scientific data is acquired. It is also possible that genetic variants today defined as of "unknown or uncertain significance" may acquire clinical importance.

Genetic test results

Positive

Identification of pathogenic variants in the above genes confirms the clinical diagnosis and is an indication for family studies.

A pathogenic variant is known to be causative for a given genetic disorder based on previous reports, or predicted to be causative based on loss of protein function or expected significant damage to proteins or protein/protein interactions. In this way it is possible to obtain a molecular diagnosis in new/other subjects, establish the risk of recurrence in family members and plan preventive and/or therapeutic measures.

Inconclusive

Detection of a variant of unknown or uncertain significance (*VUS*): a new variation without any evident pathogenic significance or a known variation with insufficient evidence (or with conflicting evidence) to indicate it is *likely benign* or *likely pathogenic* for a given genetic disorder. In these cases, it is advisable to extend testing to the patient's relatives to assess variant segregation and clarify its contribution. In some cases, it could be necessary to perform further examinations/tests or to do a clinical reassessment of pathological signs.

Negative

The absence of variations in the genomic regions investigated does not exclude a clinical diagnosis but suggests the possibility of:

- alterations that cannot be identified by sequencing, such as large rearrangements that cause loss (deletion) or gain (duplication) of extended gene fragments;
- sequence variations in gene regions not investigated by this test, such as regulatory regions (5' and 3' UTR) and deep intronic regions;
- variations in other genes not investigated by the present test.

Unexpected

Unexpected results may emerge from the test, for example information regarding consanguinity, absence of family correlation or other genetically-based diseases.

Risk for progeny

In autosomal dominant transmission, the probability that an affected carrier transmit the variant to his/her children is 50% in any pregnancy, irrespective of the sex of the child conceived.

Limits of the test

The test is limited by current scientific knowledge regarding the gene and disease.

Analytical sensitivity (proportion of positive tests when the genotype is truly present) and specificity (proportion of negative tests when the genotype is not present)

NGS: Analytical sensitivity >99.99%, with a minimum coverage of 10X; Analytical specificity 99.99%.

SANGER: Analytical sensitivity >99.99%; Analytical specificity 99.99%.

Clinical sensitivity (proportion of positive tests if the disease is present) and clinical specificity (proportion of negative tests if the disease is not present)

Clinical sensitivity: variations in the aforementioned genes are linked to EA, but may be individual variations (identified in one or a few families) and total epidemiological data is therefore not available.

Clinical sensitivity will be estimated based on internal cases. (22)

Clinical specificity: data not available.

Prescription appropriateness

The genetic test is appropriate when:

- a) the patient meets the diagnostic criteria for EA;
- b) the sensitivity of the test is greater than or equal to that of tests described in the literature.

Clinical utility

Clinical management	Utility
Confirmation of clinical diagnosis	Yes
Differential diagnosis	Yes
Couple risk assessment	Yes

Availability of clinical trials can be checked on-line at https://clinicaltrials.gov/

References

- Nihoyannopoulos P, McKenna WJ, Smith G, Foale R. Echocardiographic assessment of the right ventricle in Ebstein's anomaly: relation to clinical outcome. J Am Coll Cardiol 1986; 8(3): 627-35.
- Dima-Cozma C, Cojocaru DC, Chiriac S, Negru R, Mitu F. Ebstein's anomaly. Rev Med Chir Soc Med Nat lasi 2013; 117(3): 674-79.
- Digilio MC, Silvestri LM, Dallapiccola B, Marino B. Genetics of Ebstein Anomaly. In: Giamberti A, Chessa M. (eds) The Tricuspid Valve in Congenital Heart Disease. Springer, Milano 2014.

- Edwards WD. Embryology and pathologic features of Ebstein's anomaly. Prog Pediatr Cardiol 1993; 2:5–15.
- 5. Attenhofer Jost CH, Connolly HM, Dearani JA, Edwards WD, Danielson GK. Ebstein's anomaly. Circulation 2007; 115(2): 277-85.
- Takemura N, Machida N, Nakagawa K, Amasaki H, Washizu M, Hirose H. Ebstein's anomaly in a beagle dog. J Vet Med Sci 2003; 65(4): 531-33.
- 7. Choi R, Lee SK, Moon HS, Park IC, Hyun C. Ebstein's anomaly with an atrial septal defect in a jindo dog. The Canadian Veterinary Journal 2009; 50(4): 405-10.
- 8. Yuan SM. Ebstein's Anomaly: Genetics, Clinical Manifestations, and Management. Pediatr Neonatol 2017; 58(3): 211-215.
- Correa-Villaseñor A, Ferencz C, Neill CA, Wilson PD, Boughman JA. Ebstein's malformation of the tricuspid valve: genetic and environmental factors. The Baltimore-Washington Infant Study Group. Teratology 1994; 50(2): 137-47.
- Andelfinger GU. Human Genetics of Ebstein Anomaly. In: Rickert-Sperling S., Kelly R., Driscoll D. (eds) Congenital Heart Diseases: The Broken Heart. Springer, Vienna 2016.
- 11. Celermajer DS, Bull C, Till JA, Cullen S, Vassillikos VP, Sullivan ID, Allan L, Nihoyannopoulos P, Somerville J, Deanfield JE. Ebstein's anomaly: Presentation and outcome from fetus to adult. J Am Coll Cardiol 1994; 23(1): 170–76.
- Flores Arizmendi A, Fernandez Pineda L, Quero Jimenez C, Maitre Azcarate MJ, Herráiz Sarachaga I, Urroz E, Pérez de León J, Luis Moya J, Quero Jiménez M. The clinical profile of Ebstein's malformation as seen from the fetus to the adult in 52 patients. Cardiol Young 2004; 14(1): 55–63.
- Kelle AM, Bentley SJ, Rohena LO, Cabalka AK, Olson TM. Ebstein anomaly, left ventricular non-compaction, and early onset heart failure associated with a de novo α-tropomyosin gene mutation. Am J Med Genet A 2016; 170(8): 2186-90.
- 14. Schott JJ, Benson DW, Basson CT, Pease W, Silberbach GM, Moak JP, Maron BJ, Seidman CE, Seidman JG. Congenital heart disease caused by mutations in the transcription factor NKX2-5. Science 1998; 281(5373): 108 11.
- Elliott DA, Kirk EP, Yeoh T, Chandar S, McKenzie F, Taylor P, Grossfeld P, Fatkin D, Jones O, Hayes P, Feneley M, Harvey RP. Cardiac homeobox gene NKX2-5 mutations and congenital heart disease: associations with atrial septal defect and hypoplastic left heart syndrome. J Am Coll Cardiol 2003; 41(11): 2072 76.
- McElhinney DB, Geiger E, Blinder J, Woodrow Benson D, & Goldmuntz E. NKX2.5 mutations in patients with congenital heart disease. J Am Coll Cardiol 2003; 42(9): 1650 – 55
- Postma AV, van Engelen K, van de Meerakker J, Rahman T, Probst S, Baars MJ, Bauer U, Pickardt T, Sperling SR, Berger F, Moorman AF, Mulder BJ, Thierfelder L, Keavney B, Goodship J, Klaassen S. Mutations in the sarcomere gene MYH7 in Ebstein anomaly. Circ Cardiovasc Genet 2011; 4(1):43-50.
- Chen H, Zhang W, Li D, Cordes TM, Mark Payne R, Shou W. Analysis of ventricular hypertrabeculation and noncompaction using genetically engineered mouse models. Pediatr Cardiol 2009; 30(5):626–34.
- de Lange FJ, Moorman AFM, Anderson RH, Männer J, Soufan AT, de Gier-de Vries C, Schneider MD, Webb S, van den Hoff MJ, Christoffels Vet al. Lineage and morphogenetic analysis of the cardiac valves. Circ Res 2004; 95(6):645–54.
- Meilhac SM, Esner M, Kerszberg M, Moss JE, Buckingham ME. Oriented clonal cell growth in the developing mouse myocardium underlies cardiac morphogenesis. J Cell Biol 2004; 164(1):97–09.
- Nijak A, Alaerts M, Kuiperi C, Corveleyn A, Suys B, Paelinck B, Saenen J, Van Craenenbroeck E, Van Laer L, Loeys B, Verstraeten A. Left ventricular non-compaction with Ebstein anomaly attributed to a TPM1 mutation. Eur J Med Genet 2018; 61(1):8-10.
- Chen B, Gagnon M, Shahangian S, Anderson NL, Howerton DA, Boone JD; Centers for Disease Control and Prevention (CDC).
 Good Laboratory Practices for Molecular Genetic Testing for Heritable Diseases and Conditions. MMWR Recomm Rep 2009; 58 (RR-6):1-37.

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