



## Update of the statements on biology and clinical impact of occult hepatitis B virus infection

Giovanni Raimondo<sup>1,2,\*</sup>, Stephen Locarnini<sup>3</sup>, Teresa Pollicino<sup>1,4</sup>, Massimo Levrero<sup>5</sup>, Fabien Zoulim<sup>5</sup>, Anna S. Lok<sup>6</sup>, and the Taormina Workshop on Occult HBV Infection Faculty Members<sup>#</sup>

### Summary

In October 2018 a large number of international experts with complementary expertise came together in Taormina to participate in a workshop on occult hepatitis B virus infection (OBI). The objectives of the workshop were to review the existing knowledge on OBI, to identify issues that require further investigation, to highlight both existing controversies and newly emerging perspectives, and ultimately to update the statements previously agreed in 2008. This paper represents the output from the workshop. © 2019 European Association for the Study of the Liver. Published by Elsevier B.V. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

### Introduction

The first international workshop on occult hepatitis B virus (HBV) infection (OBI) was held in Taormina (Italy) in 2008 to review the biology and clinical implications of OBI. A panel of international experts produced a document, the Taormina statements and recommendations on OBI that was published in the *Journal of Hepatology* in 2008.<sup>1</sup>

Subsequently, many studies on OBI have been conducted but only a few of the uncertain issues have been resolved. In fact, many aspects of OBI are still controversial, including prevalence, pathobiology and clinical implications. In addition, new challenges have emerged, such as methods and sensitivities of assays for detection and risks of transmission. Thus, it was considered timely to re-visit and discuss the current understanding of OBI, and so 10 years after the first meeting, a new workshop dedicated to OBI was again held in Taormina, on October 1–2, 2018. This workshop included 5 sessions (Virology and Immunology, Diagnosis, Epidemiology, Transmission, and Liver diseases/Therapeutic implications) with presentations by invited experts followed by panel discussions. A sixth session engaged all the participants, with the goal of reaching a consensus and producing an update to the 2008 statements, culminating in this report.

### Definition

- **Occult HBV infection (OBI)** is defined as the presence of replication-competent HBV DNA (i.e. episomal HBV covalently closed circular DNA [cccDNA]) in the liver and/or HBV DNA in the blood of people who test negative for hepatitis B surface antigen (HBsAg) by currently available assays.
- Based on the HBV-specific antibody profiles, OBI may be categorised as (Fig. 1):  
**Seropositive OBI** – hepatitis B core antibody (anti-HBc) and/or hepatitis B surface antibody (anti-HBs) positive.

**Seronegative OBI** – anti-HBc and anti-HBs negative.

Among individuals with OBI, the prevalence of detectable HBV DNA in serum/plasma varies depending on the population studied, the sensitivity of the assay used, and whether blood samples at 1 or more time-points are tested. Many studies have found that HBV DNA is only intermittently detected in serum/plasma,<sup>2–6</sup> and when detectable, the concentration is low, usually less than 200 IU/ml (about 1,000 copies/ml).<sup>6–12</sup>

In people with seropositive-OBI, HBsAg may have become negative either following the resolution of acute hepatitis B (thus, after a few months of HBsAg carriage) or after decades of HBsAg positive (namely, “overt”) chronic HBV infection with or without disease. It is unknown whether patients with chronic HBV infection/disease who become HBsAg negative following antiviral therapy are comparable to patients who spontaneously clear HBsAg from the virological and immunological points of view, e.g. duration of exposure to high viral load and restoration of immune response to HBV. The possible clinical implications of this distinction are also unknown.

People with seronegative OBI (estimated to comprise between 1% and 20% of all individuals with OBI)<sup>7,13–15</sup> might have either progressively lost the hepatitis B antibodies (anti-HBc and anti-HBs) or have been hepatitis B antibody negative since the beginning of the infection. The latter condition has been described in the woodchuck model of occult woodchuck hepatitis virus (WHV) infection.<sup>16</sup>

A subset of people with OBI are infected with HBV S variants carrying mutations in the S gene (‘S-escape’ mutations), resulting in the production of modified HBsAg that is not recognised by some commercially available HBsAg assays. Circulating HBV DNA levels in these people may be compara-

Keywords: Occult HBV infection; OBI; HBV cccDNA; HBV S variants; HBV transmission; HBV reactivation; Hepatocellular carcinoma.

Received 21 January 2019; received in revised form 20 March 2019; accepted 28 March 2019

<sup>1</sup>Division of Clinical and Molecular Hepatology, University of Messina, Messina, Italy;

<sup>2</sup>Department of Clinical and Experimental Medicine, University of Messina, Messina, Italy;

<sup>3</sup>Victorian Infectious Diseases Reference Laboratory at the Doherty Institute, Melbourne, Victoria, Australia;

<sup>4</sup>Department of Human Pathology, University of Messina, Messina, Italy;

<sup>5</sup>Cancer Research Center of Lyon, INSERM U1052, Hospices Civils de Lyon, Lyon University, Lyon, France;

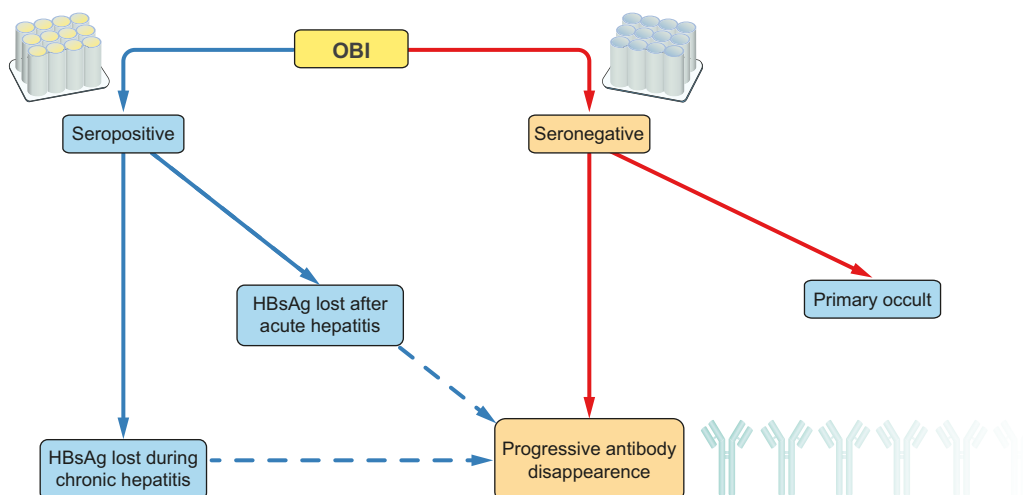
<sup>6</sup>Division of Gastroenterology and Hepatology, University of Michigan, Ann Arbor, MI, USA

<sup>#</sup> See the section on The Taormina Workshop on Occult HBV Infection Faculty Members, near the end of the paper.

\* Corresponding author. Address: Department of Clinical and Experimental Medicine, Division of Clinical and Molecular Hepatology, University Hospital of Messina, Via Conso-lare Valeria 1, 98124 Messina, Italy. Tel.: +39 90 2212392. E-mail address: [raimondo@unime.it](mailto:raimondo@unime.it) (G. Raimondo).

### Key point

In the vast majority of cases, OBI is characterized by the long-lasting persistence of low levels of HBV cccDNA chromatinised episomes in hepatocytes, with a strong suppression of overall replication activity and viral protein expression exerted by the host's defence mechanisms.



**Fig. 1. Schematic representation of HBV serum marker profile in naturally occurring OBI.** AH, acute hepatitis; CH, chronic hepatitis; HBV, hepatitis B virus; OBI, occult HBV infection.

ble to those detected in HBsAg positive individuals. Additional HBV variants with mutations in the S-gene promoter and splice variants have also been reported to affect HBsAg production/secretion and to be responsible for some cases of OBI<sup>17–27</sup> (Fig. 2).

### Virology and immunology

- The molecular basis of OBI is related to the stability and long-term persistence of cccDNA in the nucleus of infected hepatocytes (Fig. 3).

The episomal HBV cccDNA exists as a chromatinised viral minichromosome, which is very stable and long-lasting. Together with the long half-life of hepatocytes, this implies that HBV infection, once initiated, may continue for life even if efficient immune control is achieved.<sup>15,28–36</sup>

- The vast majority of OBI cases have low levels of HBV cccDNA in the liver, and suppression of overall replication activity and viral protein expression is exerted by the host's immunologic and epigenetic mechanisms.

The low level of transcriptionally active cccDNA in OBI cases results in low or undetectable HBV RNA transcription and subsequent protein translation and expression.<sup>15,37</sup> However, cccDNA in OBI cases is fully replication competent.<sup>38</sup> HBV DNA may be integrated into the host's genome and remain in the hepatocytes of HBV-infected individuals after spontaneous or treatment-induced HBsAg clearance. However, integrated HBV DNA is not replication competent and its detection is not required to make a diagnosis of OBI, since OBI is defined as the persistence of replication-competent HBV DNA.

- Immune response to HBV in OBI has not been sufficiently investigated.
- OBI is associated with antiviral immune responses that are believed to be important in maintaining HBV control.

The high prevalence of OBI worldwide suggests that the immune system is effective in controlling HBV (even if it is not definitively cleared) in the majority of people with OBI.<sup>39</sup> Antiviral immune responses in OBI are continuously stimulated by persistent/intermittent low concentrations of HBV antigens.<sup>40,41</sup> Studies performed after spontaneous resolution of acute or chronic HBV infections indicate that different profiles of antiviral T cell response are associated with the acquisition of HBV control. This difference in efficiency of antiviral protective mechanisms is probably related to the short or long-term duration of exposure of the immune system to high antigen and viral loads.<sup>42</sup> In these studies, however, the presence of OBI was only inferred from anti-HBV antibody positivity but not directly diagnosed by the detection of HBV DNA. In individuals with OBI, the immune response was confirmed by detection of HBV DNA in a single study which showed different profiles of T cell immune response against HBV epitopes in OBI-seropositive and OBI-seronegative individuals. While *ex vivo* responses were similarly weak, *in vitro* T cell expansion following specific peptide stimulation was generally more efficient among individuals with seropositive OBI than seronegative OBI.<sup>43</sup> Not only T cells, but also humoral antibody responses appear to be important in host control of OBI,<sup>44</sup> as indicated by the frequent HBV reactivations observed in patients treated with B-cell selective monoclonal based therapies (anti-CD19/20; e.g. rituximab and ofatumumab). Finally, innate immunity is also likely to contribute to HBV control in occult infection, but no data are available on this issue at present.

## Diagnosis

- Diagnosis of OBI is based on the detection of HBV DNA in the blood or the liver of HBsAg-negative individuals.
  - Detection of HBV DNA in the liver is the gold standard.
  - Detection of HBV DNA in the blood is commonly used.
  - Detection of anti-HBc in the blood is often used as a surrogate.

The diagnosis of OBI is based on the sensitivity of assays used in the detection of HBV DNA and HBsAg. HBsAg assays with inadequate sensitivity or inability to detect HBV S variants may lead to a false negative HBsAg result and misdiagnosis of OBI in people with overt HBV infection. On the other hand, HBV DNA assays with inadequate sensitivity can result in false negative HBV DNA results and may lead to a missed diagnosis of OBI.

The lower limit of detection of most currently available commercial HBsAg assays is 0.05 IU/ml. Recent studies found that between 1% and 48% of samples that tested negative using these assays test positive using more sensitive HBsAg assays with a lower limit of detection of 0.005 IU/ml.<sup>45–47</sup> Besides sensitivity, commercial HBsAg assays differ in their ability to detect S-escape variants.<sup>45,48–50</sup> The use of anti-HBs probes targeting multiple epitopes of HBsAg should be mandatory for all HBsAg assays to ensure the detection of HBV S variants.

Moreover, all serological assays detect excess HBsAg in the presence of immune complexes. Co-occurrence of anti-HBs in excess in those with HBV infection may thus interfere with the detection of HBsAg. In that situation, some people with OBI may become HBsAg positive if tested using assays that can dissociate HBsAg from immune complexes that bind HBsAg to anti-HBs.<sup>51</sup>

The lower limit of detection of most currently available commercial HBV DNA assays is 10–20 IU/ml. It is important that HBV DNA assays have similar performance across HBV genotypes and subtypes. Because HBV DNA is usually present in low concentrations and may only be intermittently detected in people with OBI, testing blood samples collected at more than one time-point, as well as testing DNA extracts from no less than 1 ml of serum or plasma is recommended for the diagnosis of OBI.

In the setting of blood transfusion, assays used for nucleic acid testing (NAT) of blood products have high specificity (99.9%) and a limit of detection of 2–4 IU/ml HBV DNA when applied to individual units. When NAT screening is conducted in minipools of multiple donations (typically, 6–20 donations per pool), the sensitivity decreases according to the dilution factor introduced by the pooling process.<sup>52</sup> One study using a highly sensitive HBV DNA assay with a limit of detection of 3.4 IU/ml identified 3 blood donors

who previously tested negative for HBsAg and HBV DNA and who were shown to have transmitted HBV.<sup>6</sup> As much as 2–24 ml of donor plasma was used for testing and not all archived samples from these donors tested positive for HBV DNA.

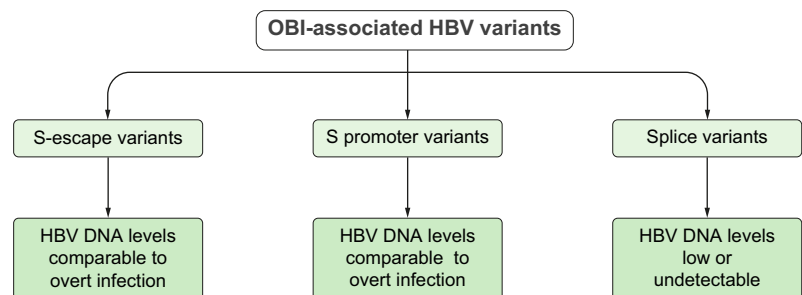
The ideal method of diagnosis of OBI is the detection of replication-competent HBV DNA in the liver. However, standardised and valid assays for HBV DNA detection in the liver are not yet available. Studies using in-house assays have variable sensitivities and specificities. The recommended methods include nested-PCR techniques to amplify at least 3 different viral genomic regions, real-time PCR assays, or droplet digital PCR assays.<sup>1,7,15,53,54</sup> In each case the assay must include primer sets that enable detection of replication-competent HBV DNA. It is important that the liver samples are properly processed to avoid cross-contamination and that appropriate negative controls are included to confirm the specificity of the assays. It is also important that a panel of HBV standards is included to validate the sensitivity of the assay. Given that HBV DNA is present in low concentrations in people with OBI, adequately sized samples, and fresh frozen – but not formalin fixed – liver tissue should be used.

Detection of anti-HBc in the blood may be used as a surrogate marker to identify OBI in blood/organ donors, in people who are about to receive immunosuppressive therapies, and for epidemiological studies. In these settings, liver tissue is often not available, access to tests for HBV DNA in the blood may be limited or delayed, and undetectable HBV DNA in blood tested at one time-point does not rule out OBI. Indeed, HBV reactivation has been reported in HBsAg-negative, anti-HBc-positive individuals who have undetectable HBV DNA in the blood.<sup>55</sup> Similarly, anti-HBc testing may identify some blood donors with OBI who have undetectable HBV DNA in mini-pool NAT.<sup>11</sup> Although earlier anti-HBc assays had high rates of false positive results, the specificity of most currently available commercial assays is very high (≥99%).<sup>56</sup>

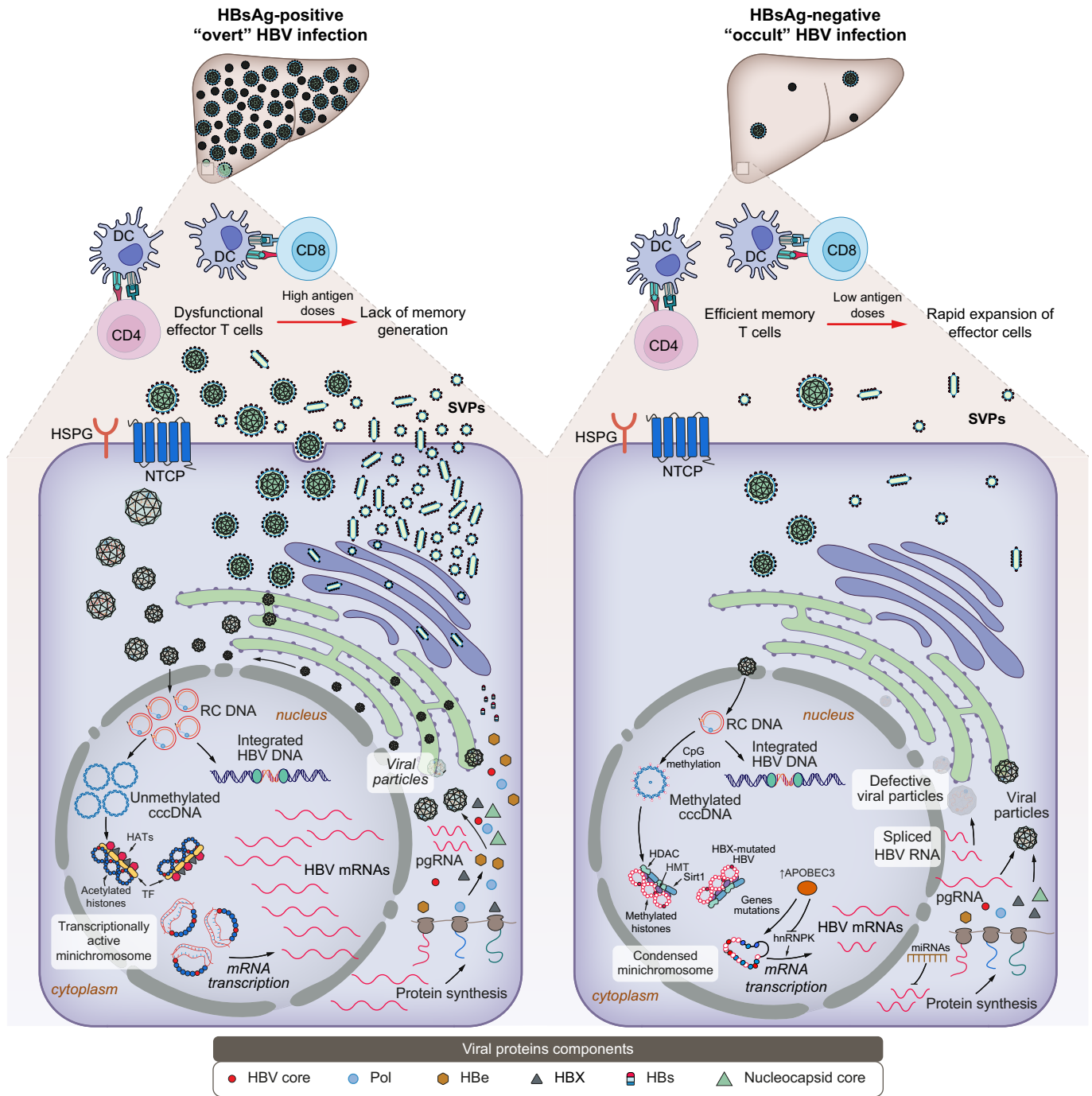
It should be noted that the absence of anti-HBc does not rule out OBI, although the prevalence and

### Key point

The lack of sensitive, standardised, and validated assays for the diagnosis of OBI is a major limitation, and available data across studies cannot be properly compared and combined.



**Fig. 2. HBV genetic variants leading to the synthesis of HBsAg unrecognised by available assays or affecting its production/secretion.** HBV, hepatitis B virus; HBsAg, HBV surface antigen.



**Fig. 3. Schematic comparison of overt and occult HBV infection.** Differences in amounts of total viral DNA, transcripts, proteins, formation and secretion of virions are exemplified. Diversity in packaging and transcriptional activities of HBV minichromosomes is also represented. Modified from Pollicino T. and Raimondo G., *Journal of Hepatology* 2015.<sup>131</sup> cccDNA, covalently closed circular DNA; DC, dendritic cell; HATs, histone acetyltransferases; HDAC, histone deacetylase; HMT, histone methyltransferase; HBV, hepatitis B virus; HBsAg, HBV surface antigen; HSPG, heparan sulfate proteoglycan; NTCP, Na<sup>+</sup>-taurocholate cotransporting polypeptide; pgRNA, pregenomic RNA; rcDNA, relaxed circular DNA; SVPs, sub-viral particles; TF, transcription factor.

clinical significance of seronegative OBI in humans are unknown.

**Epidemiology**

- Defining the epidemiology of OBI can be challenging as it relies on the performance and sensitivity of HBsAg and HBV DNA detection

assays; it also varies with the presence of risk factors for HBV exposure, the presence and severity of liver disease, the prevalence of HBV in the general population of a given country, and the definition used for OBI.

- The majority of prevalence studies have been conducted on blood donors and patients with liver disease. The OBI prevalence in these



groups is related to the prevalence of overt HBV infection in that geographical area and the population studied.

- Due to methodological limitations, OBI prevalence in the general population is still largely undefined.
- The prevalence of OBI is higher in patients with chronic liver disease and may be as high as 40% to 75% in those with HBsAg-negative hepatocellular carcinoma (HCC).
- OBI is rarely detected amongst blood donors, with HBV DNA detection rates in HBsAg-negative samples typically being less than 0.5%.

The prevalence of OBI varies greatly across the world and across patient populations, with higher rates reported in Asia.<sup>13</sup> Yet, despite high endemicity, a low prevalence of OBI has been found by various groups in Asia and in Africa.<sup>57–60</sup> Prevalence rates have varied from as low as <1% to as high as 87% but these results need to be interpreted with caution<sup>2,59</sup> because a number of factors can influence rates of OBI including the particular risk group studied, sampling issues, assay sensitivity, and the prevalence of HBsAg in the geographical region in which the study was conducted. Higher rates have also been found in individuals with risk factors for HBV infection, e.g. those coinfecting with hepatitis C virus (HCV) (15%–33%)<sup>7,61</sup> or human immunodeficiency virus (HIV) (10%–45%),<sup>62–65</sup> people who inject drugs (45%),<sup>66</sup> and people on dialysis (27%).<sup>67</sup> Prevalence rates are also higher in patients with HCC (62%),<sup>68</sup> cryptogenic cirrhosis (32%),<sup>69</sup> or those who have undergone liver transplantation (64%).<sup>70</sup> In carefully conducted studies of blood donors,<sup>71</sup> HBV DNA was detected in 0% to 4.6% of those who were HBsAg-negative and anti-HBc positive, with or without anti-HBs, with a median prevalence of 1%.<sup>11,22,55,56,67,68,72,73</sup> There is a single study that tested HBV DNA in the liver to determine the prevalence of OBI in patients with no liver disease. In this study, HBV DNA was detected in 16% of Italians with normal liver histology who underwent abdominal surgery from 2002 to 2006.<sup>74</sup>

As discussed earlier, the performance and sensitivity of the HBsAg and HBV DNA assays, the characteristics of the study population, the prevalence of HBsAg positive infection in the general population, and the criteria used to define OBI can all influence OBI prevalence rates. Thus, it is difficult to compare data between studies or to perform meta-analyses across studies.

## Transmission

### Blood transfusion

- HBsAg-negative, HBV DNA positive blood components have to be considered infectious.
- HBV transmission from OBI blood donors is still a major health issue in low- and middle-income countries, where anti-HBc and/or NAT are not implemented.

- A residual risk of transfusion-transmitted OBI exists even in developed countries, because the minimal HBV DNA infectious dose is below the limit of detection of the current NAT assays.
- The incidence of transfusion-related transmission of HBV from OBI donors might be underestimated.

Several and often concomitant reasons may be underestimated due to: a) undetectable or intermittently detectable HBV DNA in donors; b) difficulty and reluctance to trace recipients; c) lack or limited volume of donor archive samples; d) HBV infection in recipients without clinically evident acute hepatitis, which generally goes unnoticed, and may represent the majority of cases of transfusion-related transmission from OBI donors.<sup>75</sup>

The presence of anti-HBs in recipients prior to transfusion significantly reduces the risk of infection.<sup>76</sup> A recent study from Candotti and colleagues<sup>6</sup> investigated 3 repeat HBsAg-negative donors from Slovenia who had undetectable HBV DNA by highly sensitive NAT and who transmitted HBV to 9 recipients following transfusion of blood components. This study has enabled a revised estimation of the minimal HBV infectious dose from the previous 20 IU/ml to approximately 3.0 IU/ml of HBV DNA. The NAT sensitivity required to prevent HBV transmission by transfusion would need to be lowered from the current 3.4 IU/ml to a new lower limit of detection of 0.15 IU/ml.

Transfusion transmission of HBV could be reduced by implementing anti-HBc screening (if donor loss rate is not too high) or HBV NAT with a lower limit of detection of 0.15 IU/ml (technologically demanding), NAT screening of individual donation (rather than mini-pool) with larger volumes, or pathogen reduction strategies.

### Liver transplantation

HBV transmission can occur from an OBI-seropositive liver donor to a recipient who is HBV susceptible. These recipients should receive lifelong prophylaxis with nucleos(t)ide analogues (NUCs) to prevent hepatitis B.

HBV persists in the liver of people who had been infected even after HBsAg clearance. Thus, an OBI liver donor can transmit HBV infection to an HBsAg-negative, anti-HBc negative and anti-HBs negative recipient with possible development of hepatitis B.<sup>77,78</sup> Long-term prophylactic antiviral therapy with a NUC, such as entecavir or tenofovir, is recommended. However, while HBsAg positive infection is prevented, antiviral prophylaxis may not prevent the development of an OBI in the recipient.<sup>79,80</sup> HBV DNA can be detected in the liver of patients who received liver transplantation for hepatitis B and who received anti-HBV prophylaxis. Despite the absence of detectable

### Key point

The prevalence of OBI varies greatly across the world, with a higher prevalence in areas of elevated HBV endemicity and among those subjects at higher risk of parenteral transmission of the viruses.

HBsAg and HBV DNA in serum, OBI of the liver grafts is frequent. Thus, lifelong prophylaxis with NUCs is recommended for all patients who had liver transplantation for hepatitis B.<sup>80–82</sup>

#### Mother-to-child transmission

- OBI may occur in newborns from HBsAg positive mothers despite proper active/passive immunoprophylaxis at birth.<sup>83–91</sup>

HBV vaccination is one of the most important and most successful accomplishments in medical science. The World Health Organization has proposed goals to eliminate HBV by 2030.<sup>92</sup> Elimination of mother-to-child transmission of HBV is one of the most important tactics to achieve this goal. OBI in newborns, detection of anti-HBc but not HBsAg after the age of one when passive transfer of maternal antibodies should have disappeared, occurs when immunoprophylaxis fails to completely prevent HBV infection but succeeds in modulating the infection to prevent progression to chronic HBV infection.

#### Clinical implications

- In the vast majority of cases, OBI does not appear to lead to any clinical sequelae. However, OBI may result in transmission of HBV infection to blood or organ transplant recipients, and reactivation of HBV replication in patients receiving cancer chemotherapy or other immunosuppressive therapies.

A still widely debated topic is whether OBI may accelerate the progression toward cirrhosis and the development of HCC in patients with chronic liver disease caused by other factors (e.g. HCV, alcohol, non-alcoholic steatohepatitis).<sup>13,93,94</sup> While many studies, including some studies from Europe and the United States where the prevalence of HBV infection in the general population is low, have shown a significant association between OBI (as determined by detection of HBV DNA in the liver) and HCV-related cirrhosis as well as HCV-related HCC,<sup>95–97</sup> other studies in these locations found no association.<sup>98</sup> Most studies have found a high prevalence of anti-HBc among patients with HCV-related HCC, but not all studies have shown a significantly higher prevalence compared to patients with chronic HCV and no HCC.<sup>97,99–102</sup> Similarly, among the few studies where detection of HBV DNA in liver was performed, some but not all studies showed a difference in detection rates between patients with HCV, with and without HCC.<sup>15,96,98</sup>

In the woodchuck model, the clearance of the WHV surface antigen is invariably associated with persistent detection of WHV DNA in the liver as well as mild persistent necroinflammation and a high rate of HCC development.<sup>103–105</sup> Both virus- and host-related differences may explain the higher association rates compared to HBV infection in humans.

OBI retains several of the oncogenic mechanisms of overt HBV, including production of pro-oncogenic proteins and the propensity of the viral DNA to integrate into the host's genome.<sup>15,106,107</sup>

Further studies on molecular epidemiology and onco-pathogenesis are required to confirm the role of OBI in HCC development, and to determine the mechanisms by which it might exert a pro-oncogenic activity.

#### HBV reactivation

The definition of HBV reactivation in patients with OBI generally includes: i) HBsAg seroreversion and/or an increase of serum HBV DNA by at least 1 log above the lower limit of detection of the assay in a person who had previously undetectable HBsAg and HBV DNA in serum, and ii) a more than 1 log increase in serum HBV DNA in people who had detectable HBV DNA at baseline.

People with OBI can experience reactivation of HBV replication when they receive cancer chemotherapy or other immunosuppressive therapies. Although the incidence is lower than in those with chronic HBV infection, HBV reactivation can occur in up to 40% of people with OBI when potent immunosuppressive therapies are used. The risk is high (>10%) in patients receiving anti-CD20 containing regimens and myeloablative regimens for hematopoietic stem cell transplantation.<sup>108–111</sup> The risk is low (<1%) to moderate (1–10%) in people who receive other cancer chemotherapies, high dose corticosteroids, or anti-rejection therapies for solid organ transplantation.<sup>112</sup> Earlier studies suggested that the risk is modest in individuals with OBI receiving tumor necrosis factor inhibitors, but recent studies found that the risk of HBV reactivation is very low in patients receiving these therapies.<sup>103</sup> Similarly, the risk of HBV reactivation is very low in individuals with OBI receiving direct-acting antiviral therapy for hepatitis C.<sup>113–115</sup>

Given the shared transmission routes for HIV and HBV, and the immune impairment produced by HIV, OBI and HBV reactivations were more frequently reported in patients with acquired immunodeficiency syndrome. Following the widespread use of potent antiretroviral therapies, including antiretroviral agents with anti-HBV activity, reactivation of OBI has become negligible in the HIV population receiving appropriate therapy. However, HBV reactivation can occur in patients coinfecting with HIV when antiretroviral regimens are modified and drugs active against HBV are withdrawn.

Most studies on HBV reactivation in people with OBI relied on detection of anti-HBc. In studies where HBV DNA in the blood is tested, the risk of HBV reactivation is higher in those with detectable HBV DNA but the risk is also present in those with undetectable HBV DNA in serum.<sup>55</sup> Anti-HBs antibody – when present – may progressively decrease during immunosuppressive therapy,

#### Key point

Although OBI potentially maintains the pro-oncogenic properties usually attributed to the HBsAg-positive HBV infection, its role as a risk factor for hepatocellular carcinoma development still needs to be confirmed.

and HBV reactivation can also occur in people who are anti-HBs and anti-HBc positive.<sup>55,116,117</sup> Prophylactic antiviral therapy with NUCs with a high barrier to resistance, *i.e.*, entecavir or tenofovir, should be used in all patients with OBI at high risk of HBV reactivation. Those at moderate risk may receive prophylactic antiviral therapy and if not, they should be closely monitored and antiviral therapy initiated at the earliest sign of HBV reactivation. Those at low risk do not require prophylactic antiviral therapy but they need to be monitored. Risk stratification, indications for prophylactic antiviral therapy and frequency of monitoring are described in professional society guidelines.<sup>118–120</sup>

### Antiviral therapy

- Currently, antiviral therapy is not recommended for individuals with OBI.

The proposed definition of HBV functional cure, clearance of HBsAg, may suggest a conversion from overt HBV infection to OBI, but a key differentiation is that the definition of HBV functional cure requires that HBV DNA is not detected in blood. While low amounts of HBV DNA can persist in the liver, replication is suppressed – possibly by the host immune response. The risk of HCC and liver mortality is lower in patients with chronic HBV infection who have cleared HBsAg.<sup>121</sup>

The current drug discovery efforts and clinical trials aim at developing novel antiviral strategies to achieve a cure for HBV. Sterilising cure with eradication of cccDNA and integrated HBV DNA is likely not possible. Thus, current efforts focus on achieving functional cure defined as HBsAg clearance in a high proportion of patients after a finite course of therapy.<sup>122</sup> Patients with overt HBV infection who achieve functional cure would be HBsAg negative, anti-HBc positive with or without anti-HBs. While HBV DNA is not detected in the blood, the majority if not all of these patients would still have detectable HBV DNA in the liver. Multiple studies have shown that spontaneous or treatment-induced HBsAg clearance in patients with chronic HBV infection result in decreased necroinflammation of the liver and, in turn, in a reduced risk of cirrhosis, HCC, and HBV-related mortality.<sup>118,119</sup>

The development of quantitative and more sensitive assays based on digital droplet PCR for cccDNA detection has shown the possibility of detecting as little as 1 cccDNA copy/10<sup>5</sup> liver cells<sup>54,123</sup> in patients with chronic hepatitis B and viral suppression during NUC therapy, as well as in liver donors with OBI. If these results are confirmed, these assays will help to evaluate antivirals aimed at HBV cure, while highlighting the very high bar required to demonstrate eradication of HBV.

To eliminate HBV from people with OBI, HBV-infected hepatocytes would either need to be

eliminated or cured. Several paths could theoretically be investigated:

- Elimination of cccDNA within infected hepatocytes (*i.e.* curing infected cells) through cccDNA targeting strategies such as the gene editing approaches including the CRISPR/Cas9 technologies,<sup>124,125</sup> or cytokine-mediated degradation of cccDNA;<sup>126–128</sup>
- Specific killing of infected hepatocytes using strategies aimed at restoring HBV-specific T cell responses (check point inhibitors, restoration of HBV-specific T cell metabolism), therapeutic vaccination strategies, engineered T cell therapies such as chimeric antigen receptor (CAR-T) cells technologies<sup>129</sup> or HBV-T cell receptor (TCR) engineered T cells<sup>130</sup> to kill the residual infected cells in the liver.

This would require not only the adaptation of these cutting-edge technologies to this clinical application, but also more understanding of the biology of cccDNA and immune control, as well as of the number of infected hepatocytes in the setting of OBI.

More research will be necessary to achieve these goals, but such research is clearly warranted in light of the expected clinical benefit in terms of prevention of viral reactivation, transmission, and complications of the underlying liver disease.

### Further research studies

Several clinical and biological aspects of OBI need to be further explored. In this context, a fundamental objective is to definitively clarify whether, how, and in which circumstances OBI might be involved in liver injury and/or hepatocarcinogenesis. Studies aimed at understanding the immunological mechanisms that drive the development of OBI would be of seminal importance, as they might help in understanding the deficiencies in host immune control which lead to the development of productive HBV infection, and provide insights into new directions to cure HBV infection.

The 2018 OBI workshop speakers identified the following questions that should be addressed to resolve the controversies and uncertainties surrounding prevalence, clinical implications, and virologic/immunologic mechanisms of OBI.

### Epidemiology and clinical research:

- Adopt standardised methods of reporting for studies on OBI such that results across studies can be compared and combined.
- Development of more sensitive, standardised, and validated assays for detection of HBV DNA in the blood and liver.
- Development of more sensitive, standardised, and validated assays for detection of HBsAg in the blood, including detection of HBV S variants, and HBsAg present in immune complexes with anti-HBs. Determine the clinical

### Key point

Clinical implications of OBI concern: (i) HBV transmission, inducing a typical hepatitis B in the recipients, by transfusion of blood products or by liver transplantation; (ii) HBV reactivation with consequent development of hepatitis B, as in cases undergoing potent immune-suppressive therapies.

cal implications of low concentrations of circulating HBsAg, in particular the risk of transmission, liver disease progression, and HCC.

- Determine the prevalence of OBI in different parts of the world, specifically among blood donors – using standardised definitions and appropriate assays.
- Define the risk of transmission of HBV from OBI blood, organ, tissue, and cell donors, and determine the best strategies to prevent such transmissions, tailoring them to local prevalence and available resources.
- Define the clinical significance of OBI, in particular its role in HCC and in accelerating (or not) progression to cirrhosis in patients with other identifiable causes of liver disease as well as those with no identifiable cause (cryptogenic liver disease).
- Harmonise international guidelines on recommendations for preventing HBV reactivation in people with OBI.

### Basic/translational research:

- Virology – Identify the molecular mechanisms implicated in the occurrence of OBI, and define how frequently OBI is due to infection with HBV variants.
- Immunology – Identify what is different between immune control in OBI versus overt HBV infection, and determine how to harness the immune control mechanisms in OBI to achieve functional HBV cure. Establish what disturbs immune control in OBI, leading to HBV reactivation.

### Conclusions

The primary objectives of the 2018 Taormina workshop were to review the existing knowledge about OBI and – through vigorous discussion among a large number of international experts with complementary expertise – to both update the previous statements and to highlight emerging perspectives that need to be more extensively investigated.

OBI is a frequent condition that deserves attention from the scientific, medical, and public health communities. We hope these communities will collaborate to address the key questions we identified and that the answers to many of these questions will be satisfactorily addressed at the next workshop on Occult HBV Infection a decade from now.

### Financial support

The authors received no financial support to produce this manuscript.

### Conflict of interest

The authors declare no conflicts of interest that pertain to this work.

Please refer to the accompanying [ICMJE disclosure](#) forms for further details.

### The Taormina Workshop on Occult HBV Infection Faculty Members

**Jean-Pierre Allain** (Department of Haematology, University of Cambridge, United Kingdom); **Thomas Berg** (Department of Hepatology, University Hospital, Leipzig, Germany); **Antonio Bertolotti** (Singapore Institutes for Clinical Sciences, Singapore); **Maurizia Rossana Brunetto** (Internal Medicine, Department of Clinical and Experimental Medicine, Pisa University and Liver Unit, University Hospital of Pisa, Italy); **Raffaele Bruno** (Division of Infectious Diseases, University of Pavia, Italy); **Ding-Shinn Chen** (Department of Internal Medicine, National Taiwan University College of Medicine, Taiwan); **Nicola Coppola** (Infectious Disease Unit, University of Campania, Italy); **Markus Cornberg** (Department of Gastroenterology, Hepatology and Endocrinology, Hannover Medical School, Hannover; Germany); **Antonio Craxi** (Division of Gastroenterology and Hepatology, University Hospital of Palermo, Italy); **Maura Dandri** (Department of Internal Medicine, University Medical Center Hamburg-Eppendorf, Hamburg, Germany); **Vito Di Marco** (University Hospital of Palermo, Italy); **Carlo Ferrari** (Department of Medicine and Surgery, University of Parma, Italy); **Giovanni Battista Gaeta** (Division of Infectious Diseases, University of Campania, Naples, Italy); **Dieter Glebe** (Justus-Liebig University of Giessen, Institute of Medical Virology, National Reference Centre for Hepatitis B and D Viruses, Biomedical Research Centre Seltersberg, Giessen, Germany); **Luca G. Guidotti** (IRCCS San Raffaele Scientific Institute, Milan, Italy Milan, Italy); **Anna Kramvis** (Hepatitis Virus Diversity Research Unit, Department of Internal Medicine, University of the Witwatersrand, Johannesburg, South Africa; ORCID ID <https://orcid.org/0000-0001-6006-376>); **Pietro Lampertico** (Division of Gastroenterology and Hepatology, Fondazione IRCCS Ca' Granda Ospedale Maggiore Policlinico Milan, Italy); **Chengyao Li** (Department of Transfusion Medicine, School of Laboratory Medicine and Biotechnology, Southern Medical University, Guangzhou, China); **Jake Liang** (NIDDK-NIH, USA); **Alfredo Marzano** (Division of Gastroenterology, University Hospital of Torino, Italy); **Thomas I. Michalak** (Faculty of Medicine, Memorial University, St. John's, NL, Canada); **Jean-Michel Pawlotsky** (Department of Virology & INSERM U955, Henri Mondor Hospital, University of Paris-Est, Créteil, France); **Daniele Prati** (Department of Transfusion Medicine and Hematology, Fondazione IRCCS Ca' Granda Ospedale Maggiore Policlinico, Milan, Italy); **Massimo Puoti** (Division of Infectious Diseases, Niguarda Hospital, Milan, Italy); **Didier Samuel** (Centre Hépatobiliaire, University Hôpital Paul Brousse, France); **Vincent Soriano** (UNIR Health Science School & Infectious Diseases & Tropical Medicine, La Paz University Madrid, Spain); **Giovanni Squadrito** (Division of Clinical and Molecular Hepatology, Department of Clinical and Experimental Medicine, University of Messina, Italy); **Camille Sureau** (Transfusion Institut National de la Transfusion Sanguine INSERM U134, Paris, France); **Christian Trepo** (Internal Medicine Department, Hospices Civils



de Lyon, Lyon, France); **Man-Fung Yuen** (Department of Medicine, The University of Hong Kong, Queen Mary Hospital, Hong Kong, China).

**Supplementary data**

Supplementary data to this article can be found online at <https://doi.org/10.1016/j.jhep.2019.03.034>.

**References**

*Author names in bold designate shared co-first authorship*

- [1] Raimondo G, Allain JP, Brunetto MR, Buendia MA, Chen DS, Colombo M, et al. Statements from the Taormina expert meeting on occult hepatitis B virus infection. *J Hepatol* 2008;49:652–657.
- [2] Kazemi-Shirazi L, Petermann D, Muller C. Hepatitis B virus DNA in sera and liver tissue of HBsAg negative patients with chronic hepatitis C. *J Hepatol* 2000;33:785–790.
- [3] Kannangai R, Vivekanandan P, Netski D, Mehta S, Kirk GD, Thomas DL, et al. Liver enzyme flares and occult hepatitis B in persons with chronic hepatitis C infection. *J Clin Virol* 2007;39:101–105.
- [4] Chemin I, Guillaud O, Queyron PC, Trepo C. Close monitoring of serum HBV DNA levels and liver enzymes levels is most useful in the management of patients with occult HBV infection. *J Hepatol* 2009;51:824–825.
- [5] Saitta C, Musolino C, Marabello G, Martino D, Leonardi MS, Pollicino T, et al. Risk of occult hepatitis B virus infection reactivation in patients with solid tumours undergoing chemotherapy. *Dig Liver Dis* 2013;45:683–686.
- [6] Candotti D, Assennato SM, Laperche S, Allain JP, Levicnik-Stezinar S. Multiple HBV transfusion transmissions from undetected occult infections: revisiting the minimal infectious dose. *Gut* 2018.
- [7] Cacciola I, Pollicino T, Squadrito G, Cerenzia G, Orlando ME, Raimondo G. Occult hepatitis B virus infection in patients with chronic hepatitis C liver disease. *N Engl J Med* 1999;341:22–26.
- [8] Brechot C, Thiers V, Kremersdorf D, Nalpas B, Pol S, Paterlini-Brechot P. Persistent hepatitis B virus infection in subjects without hepatitis B surface antigen: clinically significant or purely “occult”? *Hepatology* 2001;34:194–203.
- [9] Hollinger FB, Sood G. Occult hepatitis B virus infection: a covert operation. *J Viral Hepat* 2010;17:1–15.
- [10] Xiao X, Zhai J, Zeng J, Tian C, Wu H, Yu Y. Comparative evaluation of a triplex nucleic acid test for detection of HBV DNA, HCV RNA, and HIV-1 RNA, with the Procleix Tigris System. *J Virol Methods* 2013;187:357–361.
- [11] Spreafico M, Berzuini A, Foglieni B, Candotti D, Raffaele L, Guarnori I, et al. Poor efficacy of nucleic acid testing in identifying occult HBV infection and consequences for safety of blood supply in Italy. *J Hepatol* 2015;63:1068–1076.
- [12] Mortensen E, Kamali A, Schirmer PL, Lucero-Obusan C, Winston CA, Oda G, et al. Are current screening protocols for chronic hepatitis B virus infection adequate? *Diagn Microbiol Infect Dis* 2016;85:159–167.
- [13] Torbenson M, Thomas DL. Occult hepatitis B. *Lancet Infect Dis* 2002;2:479–486.
- [14] Hsu HY, Chang MH, Ni YH, Chiang CL, Wu JF, Chen HL, et al. Chronologic changes in serum hepatitis B virus DNA, genotypes, surface antigen mutants and reverse transcriptase mutants during 25-year nationwide immunization in Taiwan. *J Viral Hepat* 2017;24:645–653.
- [15] Pollicino T, Squadrito G, Cerenzia G, Cacciola I, Raffa G, Craxi A, et al. Hepatitis B virus maintains its pro-oncogenic properties in the case of occult HBV infection. *Gastroenterology* 2004;126:102–110.
- [16] Michalak TI, Mulrooney PM, Coffin CS. Low doses of hepadnavirus induce infection of the lymphatic system that does not engage the liver. *J Virol* 2004;78:1730–1738.
- [17] Yamamoto K, Horikita M, Tsuda F, Itoh K, Akahane Y, Yotsumoto S, et al. Naturally occurring escape mutants of hepatitis B virus with various mutations in the S gene in carriers seropositive for antibody to hepatitis B surface antigen. *J Virol* 1994;68:2671–2676.
- [18] Hou J, Karayiannis P, Waters J, Luo K, Liang C, Thomas HC. A unique insertion in the S gene of surface antigen-negative hepatitis B virus Chinese carriers. *Hepatology* 1995;21:273–278.
- [19] Carman WF, Van Deursen FJ, Mimms LT, Hardie D, Coppola R, Decker R, et al. The prevalence of surface antigen variants of hepatitis B virus in Papua New Guinea, South Africa, and Sardinia. *Hepatology* 1997;26:1658–1666.
- [20] Chaudhuri V, Tayal R, Nayak B, Acharya SK, Panda SK. Occult hepatitis B virus infection in chronic liver disease: full-length genome and analysis of mutant surface promoter. *Gastroenterology* 2004;127:1356–1371.
- [21] Hass M, Hannoun C, Kalinina T, Sommer G, Manegold C, Gunther S. Functional analysis of hepatitis B virus reactivating in hepatitis B surface antigen-negative individuals. *Hepatology* 2005;42:93–103.
- [22] Candotti D, Lin CK, Belkhir D, Sakuldamrongpanich T, Biswas S, Lin S, et al. Occult hepatitis B infection in blood donors from South East Asia: molecular characterisation and potential mechanisms of occurrence. *Gut* 2012;61:1744–1753.
- [23] **Huang CH, Yuan Q**, Chen PJ, Zhang YL, Chen CR, Zheng QB, et al. Influence of mutations in hepatitis B virus surface protein on viral antigenicity and phenotype in occult HBV strains from blood donors. *J Hepatol* 2012;57:720–729.
- [24] Biswas S, Candotti D, Allain JP. Specific amino acid substitutions in the S protein prevent its excretion in vitro and may contribute to occult hepatitis B virus infection. *J Virol* 2013;87:7882–7892.
- [25] Huang FY, Wong DK, Seto WK, Zhang AY, Lee CK, Lin CK, et al. Sequence variations of full-length hepatitis B virus genomes in Chinese patients with HBsAg-negative hepatitis B infection. *PLoS ONE* 2014;9:e99028.
- [26] Ponde RA. Molecular mechanisms underlying HBsAg negativity in occult HBV infection. *Eur J Clin Microbiol Infect Dis* 2015;34:1709–1731.
- [27] El Chaar M, Candotti D, Crowther RA, Allain JP. Impact of hepatitis B virus surface protein mutations on the diagnosis of occult hepatitis B virus infection. *Hepatology* 2010;52:1600–1610.
- [28] Mason AL, Xu L, Guo L, Kuhns M, Perrillo RP. Molecular basis for persistent hepatitis B virus infection in the liver after clearance of serum hepatitis B surface antigen. *Hepatology* 1998;27:1736–1742.
- [29] Bock CT, Schwinn S, Locarnini S, Fyfe J, Manns MP, Trautwein C, et al. Structural organization of the hepatitis B virus minichromosome. *J Mol Biol* 2001;307:183–196.
- [30] Zoulim F. New insight on hepatitis B virus persistence from the study of intrahepatic viral cccDNA. *J Hepatol* 2005;42:302–308.
- [31] Pollicino T, Belloni L, Raffa G, Pediconi N, Squadrito G, Raimondo G, et al. Hepatitis B virus replication is regulated by the acetylation status of hepatitis B virus cccDNA-bound H3 and H4 histones. *Gastroenterology* 2006;130:823–837.
- [32] Werle-Lapostolle B, Bowden S, Locarnini S, Wursthorn K, Petersen J, Lau G, et al. Persistence of cccDNA during the natural history of chronic hepatitis B and decline during adefovir dipivoxil therapy. *Gastroenterology* 2004;126:1750–1758.
- [33] Levrero M, Pollicino T, Petersen J, Belloni L, Raimondo G, Dandri M. Control of cccDNA function in hepatitis B virus infection. *J Hepatol* 2009;51:581–592.
- [34] Locarnini S, Zoulim F. Molecular genetics of HBV infection. *Antivir Ther* 2010;15(Suppl 3):3–14.
- [35] Glebe D, Bremer CM. The molecular virology of hepatitis B virus. *Semin Liver Dis* 2013;33:103–112.
- [36] Dandri M, Locarnini S. New insight in the pathobiology of hepatitis B virus infection. *Gut* 2012;61(Suppl 1):i6–i17.
- [37] Wong DK, Huang FY, Lai CL, Poon RT, Seto WK, Fung J, et al. Occult hepatitis B infection and HBV replicative activity in patients with cryptogenic cause of hepatocellular carcinoma. *Hepatology* 2011;54:829–836.
- [38] Pollicino T, Raffa G, Costantino L, Lisa A, Campello C, Squadrito G, et al. Molecular and functional analysis of occult hepatitis B virus isolates from patients with hepatocellular carcinoma. *Hepatology* 2007;45:277–285.
- [39] Bes M, Vargas V, Piron M, Casamitjana N, Esteban JI, Vilanova N, et al. T cell responses and viral variability in blood donation candidates with occult hepatitis B infection. *J Hepatol* 2012;56:765–774.
- [40] Penna A, Artini M, Cavalli A, Levrero M, Bertoletti A, Pilli M, et al. Long-lasting memory T cell responses following self-limited acute hepatitis B. *J Clin Invest* 1996;98:1185–1194.
- [41] Reherrmann B, Ferrari C, Pasquinelli C, Chisari FV. The hepatitis B virus persists for decades after patients’ recovery from acute viral hepatitis

- despite active maintenance of a cytotoxic T-lymphocyte response. *Nat Med* 1996;2:1104–1108.
- [42] Boni C, Laccabue D, Lampertico P, Giuberti T, Vigano M, Schivazappa S, et al. Restored function of HBV-specific T cells after long-term effective therapy with nucleos(t)ide analogues. *Gastroenterology* 2012;143, 963–73 e9.
- [43] Zerbini A, Pilli M, Boni C, Fiscaro P, Penna A, Di Vincenzo P, et al. The characteristics of the cell-mediated immune response identify different profiles of occult hepatitis B virus infection. *Gastroenterology* 2008;134:1470–1481.
- [44] Loomba R, Liang TJ. Hepatitis B reactivation associated with immune suppressive and biological modifier therapies: current concepts, management strategies, and future directions. *Gastroenterology* 2017;152:1297–1309.
- [45] Yang R, Song G, Guan W, Wang Q, Liu Y, Wei L. The Lumipulse G HBsAg-Quant assay for screening and quantification of the hepatitis B surface antigen. *J Virol Methods* 2016;228:39–47.
- [46] Seto WK, Tanaka Y, Wong DK, Lai CL, Shinkai N, Yuen JC, et al. Evidence of serologic activity in chronic hepatitis B after surface antigen (HBsAg) seroclearance documented by conventional HBsAg assay. *Hepatology* 2012;7:98–105.
- [47] Ozeki I, Nakajima T, Suii H, Tatsumi R, Yamaguchi M, Kimura M, et al. Analysis of hepatitis B surface antigen (HBsAg) using high-sensitivity HBsAg assays in hepatitis B virus carriers in whom HBsAg seroclearance was confirmed by conventional assays. *Hepatology* 2018;48: E263–E274.
- [48] Shinkai N, Matsuura K, Sugauchi F, Watanabe T, Murakami S, Iio E, et al. Application of a newly developed high-sensitivity HBsAg chemiluminescent enzyme immunoassay for hepatitis B patients with HBsAg seroclearance. *J Clin Microbiol* 2013;51:3484–3491.
- [49] Zhang K, Liu Y, Chen R, Li Q, Xu Z, Si L, et al. Antigenicity reduction contributes mostly to poor detectability of HBsAg by hepatitis B virus (HBV) S-gene mutants isolated from individuals with occult HBV infection. *J Med Virol* 2018;90:263–270.
- [50] Deguchi M, Kagita M, Yoshioka N, Tsukamoto H, Takao M, Tahara K, et al. Evaluation of the highly sensitive chemiluminescent enzyme immunoassay “Lumipulse HBsAg-HQ” for hepatitis B virus screening. *J Clin Lab Anal* 2018;32 e22334.
- [51] Matsumoto A, Imaizumi M, Tanaka Y, Nishiguchi S, Yatsuhashi H, Ishida T, et al. Novel and highly sensitive immunoassay for total hepatitis B surface antigen, including that complexed with hepatitis B surface antibody. *J Gastroenterol* 2017;52:376–384.
- [52] Candotti D, Laperche S. Hepatitis B virus blood screening: need for reappraisal of blood safety measures? *Front Med (Lausanne)* 2018;5:29.
- [53] Akram A, Islam SMR, Munshi SU, Tabassum S. Detection of hepatitis B virus DNA among chronic and potential occult HBV patients in resource-limited settings by loop-mediated isothermal amplification assay. *J Viral Hepat* 2018;25:1306–1311.
- [54] Caviglia GP, Abate ML, Tandoi F, Ciancio A, Amoroso A, Salizzoni M, et al. Quantitation of HBV cccDNA in anti-HBc-positive liver donors by droplet digital PCR: a new tool to detect occult infection. *J Hepatology* 2018;69:301–7.
- [55] Cholongitas E, Haidich AB, Apostolidou-Kiouti F, Chalevas P, Papatheodoridis GV. Hepatitis B virus reactivation in HBsAg-negative, anti-HBc-positive patients receiving immunosuppressive therapy: a systematic review. *Ann Gastroenterol* 2018;31:480–490.
- [56] White R, Delieu E, Perry KR, Parry JV. Four anti-HBc assays: medicines and Healthcare products Regulatory Agency; 2003.
- [57] Behzad-Behbahani A, Mafi-Nejad A, Tabei SZ, Lankarani KB, Torab A, Moaddeb A. Anti-HBc & HBV-DNA detection in blood donors negative for hepatitis B virus surface antigen in reducing risk of transfusion associated HBV infection. *Indian J Med Res* 2006;123:37–42.
- [58] El-Zayadi AR, Ibrahim EH, Badran HM, Saeid A, Moneib NA, Shemis MA, et al. Anti-HBc screening in Egyptian blood donors reduces the risk of hepatitis B virus transmission. *Transfus Med* 2008;18:55–61.
- [59] Yuen MF, Lee CK, Wong DK, Fung J, Hung I, Hsu A, et al. Prevalence of occult hepatitis B infection in a highly endemic area for chronic hepatitis B: a study of a large blood donor population. *Gut* 2010;59:1389–1393.
- [60] Zheng X, Ye X, Zhang L, Wang W, Shuai L, Wang A, et al. Characterization of occult hepatitis B virus infection from blood donors in China. *J Clin Microbiol* 2011;49:1730–1737.
- [61] Kao JH, Chen PJ, Lai MY, Chen DS. Occult hepatitis B virus infection and clinical outcomes of patients with chronic hepatitis C. *J Clin Microbiol* 2002;40:4068–4071.
- [62] Gupta S, Singh S. Occult hepatitis B virus infection in ART-naive HIV-infected patients seen at a tertiary care centre in north India. *BMC Infect Dis* 2010;10:53.
- [63] Lo Re 3rd V, Frank I, Gross R, Dockter J, Linnen JM, Giachetti C, et al. Prevalence, risk factors, and outcomes for occult hepatitis B virus infection among HIV-infected patients. *J Acquir Immune Defic Syndr* 2007;44:315–320.
- [64] Bell TG, Makondo E, Martinson NA, Kramvis A. Hepatitis B virus infection in human immunodeficiency virus infected southern African adults: occult or overt—that is the question. *PLoS ONE* 2012;7 e45750.
- [65] Mudawi H, Hussein W, Mukhtar M, Yousif M, Nemer O, Glebe D, et al. Overt and occult hepatitis B virus infection in adult Sudanese HIV patients. *Int J Infect Dis* 2014;29:65–70.
- [66] Torbenson M, Kannangai R, Astemborski J, Strathdee SA, Vlahov D, Thomas DL. High prevalence of occult hepatitis B in Baltimore injection drug users. *Hepatology* 2004;39:51–57.
- [67] Di Stefano M, Volpe A, Stallone G, Tartaglia L, Prato R, Martinelli D, et al. Occult HBV infection in hemodialysis setting is marked by presence of isolated antibodies to HBcAg and HCV. *J Nephrol* 2009;22:381–386.
- [68] Hassan ZK, Hafez MM, Mansor TM, Zekri AR. Occult HBV infection among Egyptian hepatocellular carcinoma patients. *Virol J* 2011;8:90.
- [69] Chan HL, Tsang SW, Leung NW, Tse CH, Hui Y, Tam JS, et al. Occult HBV infection in cryptogenic liver cirrhosis in an area with high prevalence of HBV infection. *Am J Gastroenterol* 2002;97:1211–1215.
- [70] Ghisetti V, Marzano A, Zamboni F, Barbui A, Franchello A, Gaia S, et al. Occult hepatitis B virus infection in HBsAg negative patients undergoing liver transplantation: clinical significance. *Liver Transpl* 2004;10:356–362.
- [71] Hollinger FB. Hepatitis B virus infection and transfusion medicine: science and the occult. *Transfusion* 2008;48:1001–1026.
- [72] Liu CJ, Chen DS, Chen PJ. Epidemiology of HBV infection in Asian blood donors: emphasis on occult HBV infection and the role of NAT. *J Clin Virol* 2006;36(Suppl 1):S33–S44.
- [73] Candotti D, Grabarczyk P, Ghiazza P, Roig R, Casamitjana N, Iudicone P, et al. Characterization of occult hepatitis B virus from blood donors carrying genotype A2 or genotype D strains. *J Hepatology* 2008;49:537–547.
- [74] Raimondo G, Navarra G, Mondello S, Costantino L, Colloredo G, Cucinotta E, et al. Occult hepatitis B virus in liver tissue of individuals without hepatic disease. *J Hepatology* 2008;48:743–746.
- [75] Locarnini S, Raimondo G. How infectious is the hepatitis B virus? Readings from the occult. *Gut* 2018.
- [76] Allain JP, Mihaljevic I, Gonzalez-Fraile MI, Gubbe K, Holm-Harritshoj L, Garcia JM, et al. Infectivity of blood products from donors with occult hepatitis B virus infection. *Transfusion* 2013;53:1405–1415.
- [77] Chazouilleres O, Mamish D, Kim M, Carey K, Ferrell L, Roberts JP, et al. “Occult” hepatitis B virus as source of infection in liver transplant recipients. *Lancet* 1994;343:142–146.
- [78] Cholongitas E, Papatheodoridis GV, Burroughs AK. Liver grafts from anti-hepatitis B core positive donors: a systematic review. *J Hepatology* 2010;52:272–279.
- [79] Cheung CK, Lo CM, Man K, Lau GK. Occult hepatitis B virus infection of donor and recipient origin after liver transplantation despite nucleoside analogue prophylaxis. *Liver Transpl* 2010;16:1314–1323.
- [80] Coffin CS, Mulrooney-Cousins PM, van Marle G, Roberts JP, Michalak TI, Terrault NA. Hepatitis B virus quasispecies in hepatic and extrahepatic viral reservoirs in liver transplant recipients on prophylactic therapy. *Liver Transpl* 2011;17:955–962.
- [81] Roche B, Feray C, Gigou M, Roque-Afonso AM, Arulnaden JL, Delvart V, et al. HBV DNA persistence 10 years after liver transplantation despite successful anti-HBs passive immunoprophylaxis. *Hepatology* 2003;38:86–95.
- [82] Hussain M, Soldevila-Pico C, Emre S, Luketic V, Lok ASGroup NH-OS. Presence of intrahepatic (total and ccc) HBV DNA is not predictive of HBV recurrence after liver transplantation. *Liver Transpl* 2007;13:1137–1144.
- [83] Shahmoradi S, Yahyapour Y, Mahmoodi M, Alavian SM, Fazeli Z, Jazayeri SM. High prevalence of occult hepatitis B virus infection in children born to HBsAg-positive mothers despite prophylaxis with hepatitis B vaccination and HBIG. *J Hepatology* 2012;57:515–521.
- [84] Hsu HY, Chang MH, Ni YH, Chiang CL, Wu JF, Chen HL. Universal infant immunization and occult hepatitis B virus infection in children and adolescents: a population-based study. *Hepatology* 2015;61:1183–1191.

- [85] **Chen ZX, Gu GF, Bian ZL**, Cai WH, Shen Y, Hao YL, et al. Clinical course and perinatal transmission of chronic hepatitis B during pregnancy: a real-world prospective cohort study. *J Infect* 2017;75:146–154.
- [86] **Amponsah-Dacosta E, Lebelo RL, Rakgole JN, Selabe SG, Gededzha MP, Mayaphi SH**, et al. Hepatitis B virus infection in post-vaccination South Africa: occult HBV infection and circulating surface gene variants. *J Clin Virol* 2015;63:12–17.
- [87] **Foad H, Maklad S, Mahmoud F, El-Karakasy H**. Occult hepatitis B virus infection in children born to HBsAg-positive mothers after neonatal passive-active immunoprophylaxis. *Infection* 2015;43:307–314.
- [88] **Mu SC, Lin YM, Jow GM, Chen BF**. Occult hepatitis B virus infection in hepatitis B vaccinated children in Taiwan. *J Hepatol* 2009;50:264–272.
- [89] **Sadeghi A, Yahyapour Y, Poortahmasebi V, Shahmoradi S, Roggendorf M, Karimzadeh H**, et al. Clearance of HBV DNA in immunized children born to HBsAg-positive mothers, years after being diagnosed with occult HBV infection. *J Viral Hepat* 2016;23:282–285.
- [90] **Lu Y, Liu YL, Nie JJ, Liang XF, Yan L, Wang FZ**, et al. Occult HBV infection in immunized neonates born to HBsAg-positive mothers: a prospective and follow-up study. *PLoS ONE* 2016;11 e0166317.
- [91] **Zhou S, Li T, Allain JP, Zhou B, Zhang Y, Zhong M**, et al. Low occurrence of HBsAg but high frequency of transient occult HBV infection in vaccinated and HBIG-administered infants born to HBsAg positive mothers. *J Med Virol* 2017;89:2130–2137.
- [92] **Organization WH**. *Global Hepatitis Report 2017*. Licence: CC BY-NC-SA 3.0 IGO. Geneva, 2017.
- [93] **Covolo L, Pollicino T, Raimondo G, Donato F**. Occult hepatitis B virus and the risk for chronic liver disease: a meta-analysis. *Dig Liver Dis* 2013;45:238–244.
- [94] **Raimondo G, Caccamo G, Filomia R, Pollicino T**. Occult HBV infection. *Semin Immunopathol* 2013;35:39–52.
- [95] **Shetty K, Hussain M, Nei L, Reddy KR, Lok AS**. Prevalence and significance of occult hepatitis B in a liver transplant population with chronic hepatitis C. *Liver Transpl* 2008;14:534–540.
- [96] **Squadrito G, Cacciola I, Alibrandi A, Pollicino T, Raimondo G**. Impact of occult hepatitis B virus infection on the outcome of chronic hepatitis C. *J Hepatol* 2013;59:696–700.
- [97] **Wang H, Swann R**, Thomas E, Innes HA, Valerio H, Hayes PC, et al. Impact of previous hepatitis B infection on the clinical outcomes from chronic hepatitis C? A population-level analysis. *J Viral Hepat* 2018;25:930–938.
- [98] **Lok AS, Everhart JE, Di Bisceglie AM, Kim HY, Hussain M, Morgan TR**, et al. Occult and previous hepatitis B virus infection are not associated with hepatocellular carcinoma in United States patients with chronic hepatitis C. *Hepatology* 2011;54:434–442.
- [99] **Ikeda K, Marusawa H, Osaki Y, Nakamura T, Kitajima N, Yamashita Y**, et al. Antibody to hepatitis B core antigen and risk for hepatitis C-related hepatocellular carcinoma: a prospective study. *Ann Intern Med* 2007;146:649–656.
- [100] **Coppola N, Onorato L, Sagnelli C, Sagnelli E, Angelillo IF**. Association between anti-HBc positivity and hepatocellular carcinoma in HBsAg-negative subjects with chronic liver disease: a meta-analysis. *Medicine (Baltimore)* 2016;95 e4311.
- [101] **Stroffolini T, Almasio PL, Persico M, Bollani S, Benvegna L, Di Costanzo G**, et al. Lack of correlation between serum anti-HBc detectability and hepatocellular carcinoma in patients with HCV-related cirrhosis. *Am J Gastroenterol* 2008;103:1966–1972.
- [102] **Kitab B, Ezzikouri S, Alaoui R, Nadir S, Badre W, Trepo C**, et al. Occult HBV infection in Morocco: from chronic hepatitis to hepatocellular carcinoma. *Liver Int* 2014;34:e144–e150.
- [103] **Korba BE, Wells FV, Baldwin B, Cote PJ, Tennant BC, Popper H**, et al. Hepatocellular carcinoma in woodchuck hepatitis virus-infected woodchucks: presence of viral DNA in tumor tissue from chronic carriers and animals serologically recovered from acute infections. *Hepatology* 1989;9:461–470.
- [104] **Michalak TI, Pardoe IU, Coffin CS, Churchill ND, Freake DS, Smith P**, et al. Occult lifelong persistence of infectious hepadnavirus and residual liver inflammation in woodchucks convalescent from acute viral hepatitis. *Hepatology* 1999;29:928–938.
- [105] **Mulrooney-Cousins PM, Michalak TI**. Persistent occult hepatitis B virus infection: experimental findings and clinical implications. *World J Gastroenterol* 2007;13:5682–5686.
- [106] **Saitta C, Tripodi G, Barbera A, Bertuccio A, Smedile A, Ciancio A**, et al. Hepatitis B virus (HBV) DNA integration in patients with occult HBV infection and hepatocellular carcinoma. *Liver Int* 2015;35:2311–2317.
- [107] **Brechot C**. Pathogenesis of hepatitis B virus-related hepatocellular carcinoma: old and new paradigms. *Gastroenterology* 2004;127: S56–S61.
- [108] **Hsu C, Tsou HH, Lin SJ, Wang MC, Yao M, Hwang WL**, et al. Chemotherapy-induced hepatitis B reactivation in lymphoma patients with resolved HBV infection: a prospective study. *Hepatology* 2014;59:2092–2100.
- [109] **Seto WK, Chan TS, Hwang YY, Wong DK, Fung J, Liu KS**, et al. Hepatitis B reactivation in patients with previous hepatitis B virus exposure undergoing rituximab-containing chemotherapy for lymphoma: a prospective study. *J Clin Oncol* 2014;32:3736–3743.
- [110] **Hammond SP, Borchelt AM, Ukomadu C, Ho VT, Baden LR, Marty FM**. Hepatitis B virus reactivation following allogeneic hematopoietic stem cell transplantation. *Biol Blood Marrow Transplant* 2009;15:1049–1059.
- [111] **Seto WK, Chan TS, Hwang YY, Wong DK, Fung J, Liu KS**, et al. Hepatitis B reactivation in occult viral carriers undergoing hematopoietic stem cell transplantation: a prospective study. *Hepatology* 2017;65:1451–1461.
- [112] **Paul S, Saxena A, Terrin N, Viveiros K, Balk EM, Wong JB**. Hepatitis B virus reactivation and prophylaxis during solid tumor chemotherapy: a systematic review and meta-analysis. *Ann Intern Med* 2016;164:30–40.
- [113] **Pauly MP, Tucker LY, Szpakowski JL, Ready JB, Baer D, Hwang J**, et al. Incidence of hepatitis B virus reactivation and hepatotoxicity in patients receiving long-term treatment with tumor necrosis factor antagonists. *Clin Gastroenterol Hepatol* 2018;16: 1964–73 e1.
- [114] **Mucke MM, Backus LI, Mucke VT, Coppola N, Preda CM, Yeh ML**, et al. Hepatitis B virus reactivation during direct-acting antiviral therapy for hepatitis C: a systematic review and meta-analysis. *Lancet Gastroenterol Hepatol* 2018;3:172–180.
- [115] **Tamori A, Abiru S, Enomoto H, Kioka K, Korenaga M, Tani J**, et al. Low incidence of hepatitis B virus reactivation and subsequent hepatitis in patients with chronic hepatitis C receiving direct-acting antiviral therapy. *J Viral Hepat* 2018;25:608–611.
- [116] **Onozawa M, Hashino S, Izumiyama K, Kahata K, Chuma M, Mori A**, et al. Progressive disappearance of anti-hepatitis B surface antigen antibody and reverse seroconversion after allogeneic hematopoietic stem cell transplantation in patients with previous hepatitis B virus infection. *Transplantation* 2005;79:616–619.
- [117] **Vigano M, Vener C, Lampertico P, Annaloro C, Pichoud C, Zoulim F**, et al. Risk of hepatitis B surface antigen seroreversion after allogeneic hematopoietic SCT. *Bone Marrow Transplant* 2011;46:125–131.
- [118] **Terrault NA, Lok ASF, McMahon BJ, Chang KM, Hwang JP, Jonas MM**, et al. Update on prevention, diagnosis, and treatment of chronic hepatitis B: AASLD 2018 hepatitis B guidance. *Hepatology* 2018;67:1560–1599.
- [119] **European Association for the Study of the Liver**. Electronic address eee, **European Association for the Study of the Liver**. *EASL 2017 Clinical Practice Guidelines on the management of hepatitis B virus infection*. *J Hepatol* 2017;2017(67):370–398.
- [120] **Sarin SK, Kumar M, Lau GK, Abbas Z, Chan HL, Chen CJ**, et al. Asian-Pacific clinical practice guidelines on the management of hepatitis B: a 2015 update. *Hepatol Int* 2016;10:1–98.
- [121] **Yip TC, Wong GL, Chan HL, Tse YK, Lam KL, Lui GC**, et al. HBsAg seroclearance further reduces hepatocellular carcinoma risk after complete viral suppression with nucleos(t)ide analogues. *J Hepatol* 2019;70:361–370.
- [122] **Lok AS, Zoulim F, Dusheiko G, Ghany MG**. Hepatitis B cure: from discovery to regulatory approval. *Hepatology* 2017;66:1296–1313.
- [123] **Inchauspé A, Locatelli M, Lebosse F, Diederichs A, Freydisier-Berthet A, Alam A**, et al. Droplet digital PCR quantitation of HBV cccDNA pool and transcriptional activity in long-term nucleos(t)ide analogue treated patients. In: *Liver. JHEAftSot*, editor. ILC2018. Paris, France: Elsevier Inc; 2018. p. S481.
- [124] **Seeger C, Sohn JA**. Targeting hepatitis B virus with CRISPR/Cas9. *Mol Ther Nucleic Acids* 2014;3 e216.
- [125] **Seeger C, Sohn JA**. Complete spectrum of CRISPR/Cas9-induced mutations on HBV cccDNA. *Mol Ther* 2016;24:1258–1266.
- [126] **Lucifora J, Xia Y, Reisinger F, Zhang K, Stadler D, Cheng X**, et al. Specific and nonhepatotoxic degradation of nuclear hepatitis B virus cccDNA. *Science* 2014;343:1221–1228.
- [127] **Xia Y, Stadler D, Lucifora J, Reisinger F, Webb D, Hosel M**, et al. Interferon-gamma and tumor necrosis factor-alpha produced by T cells reduce the HBV persistence form, cccDNA, without cytolysis. *Gastroenterology* 2016;150:194–205.

## Review

- [128] Koh S, Kah J, Tham CYL, Yang N, Ceccarello E, Chia A, et al. Nonlytic lymphocytes engineered to express virus-specific T-cell receptors limit HBV infection by activating APOBEC3. *Gastroenterology* 2018;155(180–93) e6.
- [129] Gehring A, Protzer U. Targeting innate and adaptive immune responses to cure chronic HBV infection. *Gastroenterology* 2018.
- [130] Bertoletti A, Le Bert N. Immunotherapy for chronic hepatitis B virus infection. *Gut Liver*. 2018;12:497–507.
- [131] Pollicino T, Raimondo G. Occult hepatitis B infection. *J Hepatol* 2014;61:688–689.