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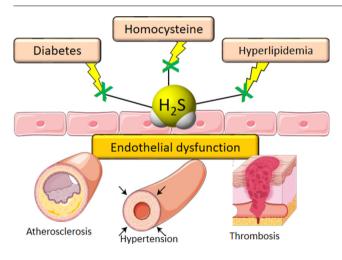
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Role of hydrogen sulfide in endothelial dysfunction: Pathophysiology and therapeutic approaches

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ABSTRACT

The vascular endothelium represents a fundamental mechanical and biological barrier for the maintenance of vascular homeostasis along the entire vascular tree. Changes in its integrity are associated to several cardiovascular diseases, including hypertension, atherosclerosis, hyperhomocysteinemia, diabetes, all linked to the peculiar condition named endothelial dysfunction, which is referred to the loss of endothelial physiological functions, comprehending the regulation of vascular relaxation and/or cell redox balance, the inhibition of leukocyte infiltration and the production of NO. Among the endothelium-released vasoactive factors, in the last years hydrogen sulfide has been viewed as one of the main characters involved in the regulation of endothelium functionality, and many studies demonstrated that H₂S behaves as a vasoprotective gasotransmitter in those cardiovascular diseases where endothelial dysfunction seems to be the central issue.

The role of hydrogen sulfide in endothelial dysfunction-related cardiovascular diseases is discussed in this review, focusing the attention on the possible therapeutic approaches using molecules able to release H₂S.

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Introduction

The vascular endothelium is a monolayer of dynamic cells which cover the inner surface of the entire vascular tree and it

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is known to be an important regulator of the vascular wall homeostasis. In particular, endothelial cells (ECs) represent both mechanical and biological barrier between the vessels and the underlying tissues through which continuous exchange of molecules between the interstitial tissue and blood occurs in the capillaries [1]. The endothelium is continuously exposed to shear stress due to the blood flow and such a mechanical stimulus triggers intracellular and biochemical signals which may alter the basal permeability [2]. Changes in the integrity of the endothelial barrier are related to different pathophysiological processes, including tissue remodeling, repair and inflammation [3]. In addition to its role as a barrier, healthy endothelium is involved in the maintenance of vascular tone, since it mediates smooth muscle relaxation through endothelium-derived hyperpolarizing factors, promotes antioxidant and anti-inflammatory effects through the inhibition of both leukocyte adhesion and migration and prevents smooth muscle cell proliferation and migration. The release and control of several endothelial bioactive molecules have been shown to counteract thrombosis and atherosclerosis [4].

The endothelial dysfunction (ED) refers to the loss of these physiological functions caused by different risk factors, including hyperglycemia, hypercholesterolemia and hyperhomocysteinemia (HHcy) [5]. ECs respond to such pathological conditions by increasing the levels of reactive oxygen species (ROS), and consequently enhancing the oxidative stress along the vascular tree. These conditions trigger several damaging events resulting in the inability of the endothelium to exert its fundamental activities, i.e the regulation of vascular relaxation and/or cell redox balance, leading to inflammatory response that usually promotes endothelial activation (inflammatory response), reduces nitric oxide (NO) production correlated with the impairment of vascular smooth muscle dilation [6].

Many evidence reported the vasoprotective effect of hydrogen sulfide (H₂S) that is an endogenous gaseous molecule, synthesized in mammalian tissues from L-cysteine by two cytosolic pyridoxal-5'-phosphate-dependent enzymes: cystathionine β -synthase (CBS) and cystathionine γ -lyase (CSE) [7]. In addition, 3-mercapto sulfotransferase (3-MST) and cysteine aminotransferase (CAT) contribute to the biosynthesis of H₂S [8]. Even though it has been long considered a toxic gas, recent studies described the fundamental role of H₂S in the regulation of cardiovascular homeostasis; indeed, its deficiency is etiologically associated with several cardiovascular diseases [9].

In addition, H₂S is also emerging as an essential molecule in controlling the homeostasis of endothelial function and an impairment of its endogenous production is related to the pathogenesis of ED [10]. Many studies demonstrated that H₂S behaves as a vasculoprotective gasotransmitter by modulating different cellular pathways and interfering with a variety of vascular diseases. Indeed, H₂S inhibits atherogenic modification of low-density lipoproteins (LDL) [11], prevents monocytes adhesion due to ECs activation [12], clearly promotes vasorelaxing responses [13], decreases intimal hyperplasia by inhibiting vascular smooth muscle cells (VSMC) migration and proliferation [14], limits vascular calcification [15], thrombogenesis and platelet aggregation [16], inhibits macrophage foam cell formation and degranulation [17], limits inflammatory responses and reduces plasma homocysteine (Hcy) levels in experimental animals [18].

Therefore, ED is widely associated with several pathological conditions including diabetes, hypertension, atherosclerosis and HHcy. The role of hydrogen sulfide in such ED-related diseases, is discussed, focusing the attention on the possible therapeutic approaches using molecules able to release H₂S.

Hydrogen sulfide and hypertension-related endothelial dysfunction

The correlation between ED and hypertension is mainly referred to as an alteration of the metabolism of nitric oxide (NO), due to a severe imbalance between the biosynthesis of vasodilator agents and the production of vasoconstrictor substances, which guarantee the control of vascular tone [19].

Indeed, among numerous vasoactive molecules, NO plays a crucial role in mediating vasodilation responses. Many studies demonstrated that the reduction of NO levels in vessels is associated with hypertension. The NO-deficiency impairs vascular homeostasis, alters the endothelium permeability and consequently leads to inflammatory and fibrotic processes, which are responsible for structural alterations, including vascular hypertrophy and increased vascular resistances. Consistently, chronic administration of NOS inhibitors causes sustained hypertension in several pre-clinical models [20,21].

Besides NO, also the endogenous gasotransmitter H_2S is emerging as one of the main characters in modulating the endothelium integrity and functionality. Numerous *in vitro*, *ex vivo* and *in vivo* studies demonstrated the vasoactive role of H_2S , confirming the hypothesis that H_2S importantly contributes to the regulation of vascular tone [22,23]. In fact, like NO, H_2S assures the maintenance of vascular homeostasis by the regulation of the complex balance of several components, including vasodilation responses as widely demonstrated for molecules able to release H_2S [24–36].

The pathogenesis of ED-related hypertension seems to be also related to the reduction of H_2S biosynthesis. Indeed, the downregulation of CSE, the enzyme which is responsible for H_2S production in the vasculature, leads to the increase of blood pressure associated with the reduction of H_2S bioavailability [37–40]; accordingly, CSE-knock-out mice showed an abolished endothelium-dependent relaxation in resistance mesenteric arteries [23].

Furthermore, a human cohort study reported that hypertensive patients showed reduced H₂S plasma levels [41], suggesting that H₂S can be also viewed as a predictive biomarker for the development of high blood pressure [42]. The beneficial effects of H₂S exogenous supply were highlighted by Xiao and colleagues, who reported that 20-week administration of NaHS – which is a molecule able to release H₂S – lowered the arterial pressure and increased the production of NO, enhancing eNOS phosphorylation through the activation of peroxisome proliferator-activated receptor δ / protein kinase B/ AMP-activated protein kinase (PPAR- δ /Akt/AMPK) signaling pathway [43].

 H_2S and NO share several beneficial effects in the cardiovascular system and many studies reported the mutual interaction between H_2S and NO. Indeed, NO increases the uptake of L-cysteine, which is the main substrate of the H_2S generating enzymes CSE and CBS, consequently leading to the increase of H_2S production [44]; moreover, exogenous NO-donors trigger CSE activation in VSMC, increasing the biosynthesis of H_2S [45].

Accordingly, the inhibition of eNOS attenuated the H_2S -induced vasorelaxation in rat aorta [46] and inhibited the expression /activity of CSE in the cardiovascular system [47].

Coletta and colleagues observed that NaHS-mediated vasodilation was attenuated in eNOS-KO mice vessels [27] and, in accordance, NaHS increased the NO production through eNOS sulphydration in rat corpus cavernosum [48,49].

The inhibition of NO synthase, using N(ω)-nitro-L-arginine methyl ester (L-NAME), induces ED-related hypertension, and is a widely used model for studying the effect of several vasoactive molecules, including H₂S.

Indeed, in L-NAME-induced hypertensive rats, the lack of NO biosynthesis was associated with dysfunction of the cysteine/ CSE/H₂S pathway and exogenous administration of H₂S prevented the development of hypertension [50].

Furthermore, H₂S potentiates the NO-mediated vasorelaxing effect through the inhibition of phosphodiesterase-5 (PDE-5), prolonging the half-life of cGMP which is the key messenger in NO vasoactive signals [26].

However, another interaction has been hypothesized: NO and H₂S can chemically interact in the tissues to form nitrosothiols which are endowed with their own signaling pathway [51].

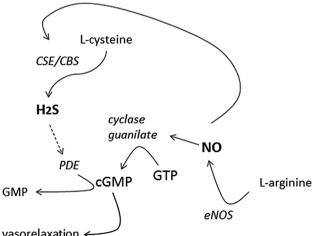
Anyway, such a mutual relationship seems to be missing in the ED condition. In fact, the mutual interplay between NO and H₂S in rat coronary arteries is attenuated in hypertensive conditions. Interestingly, the ED-related hypertensive status highlights reciprocally independent mechanisms of action of NO and H₂S, suggesting that cGMP plays a secondary role in meditating H₂S and NOrelated vasoactive responses. Consequently, in this pathological condition, H₂S plays a compensatory role for the lack of NOendothelium production, assuring the control of vascular tone [52]. Similarly, in ED-related erectile dysfunction, characterized by a decrease in NO bioavailability, H₂S compensates the lack of NO [53] (Fig. 1).

On the other hand, other authors reported that NaHS decreases NO formation, reduces eNOS activity and inhibits the uptake of L-arginine in isolated rat aortae and in ECs [54,55].

Unlike NO, H₂S modulates the vascular tone not only through a cGMP-dependent manner, but also through cGMP-independent mechanisms. Firstly, it has been demonstrated that H₂S directly opens ATP-sensitive potassium (K_{ATP}) channels in VSMC and causes hyperpolarization which is an event responsible for vasorelaxation [24]. In addition, the involvement of voltage-operated potassium (Kv7) channels has been established in endothelium-denuded rat aortic rings. Similarly to retigabine, which is a Kv7.4 opener, also NaHS activates Kv7.4 currents in Chinese hamster ovary cells (CHO) and produces a significant hyperpolarization [28]. Furthermore, other mechanisms have been proposed, including the inhibition of L-type voltage-operated calcium channels [56].

In several studies, the administration of H₂S and H₂S-donors promoted the decrease of blood pressure in hypertensive animals, reversed hypertension-related vascular remodeling and improved the endothelial function in hypertensive humans and rats [13,24].

Zhao et al., in 2001 observed that intravenous administration of NaHS reduced blood pressure, reversed ED, restored NO bioavailability, limited endothelial ROS production in a rat model of Ang-



vasorelaxation

Fig. 1. H₂S increases the cGMP-NO dependent effect by inhibiting the phosphodiesterase-5.

II-induced hypertension [24,57]. The reduction of H_2S plasma levels was also reported in a glucocorticoid-induced hypertension model, typically characterized by ED [38].

The administration of NaHS improves the endothelial function, thanks to the inhibition of oxidative stress and inflammation in spontaneously hypertensive rats. Interestingly, the knocking down of nuclear factor erythroid-2-related factor 2 (Nrf2) abolished such a protective effect [58].

Besides NaHS, polysulfides are considered as natural molecules able to release H₂S in biological environment. Interestingly, diallyl trisulfide (DATS), diallyl disulfide (DADS) and diallyl sulfide (DAS), mainly occurring in plants of the Alliaceae family (for instance garlic), have been demonstrated to be vasorelaxing agents [25] due to their H₂S-generating property. Indeed, the release of H₂S is the key event through which polysulfides exert anti-hypertensive effects in L-NAME-treated Wistar rats [59].

In 2008, GYY4137, a widely studied H₂S slow-releasing compound, has been proven to induce vasodilatation in aortic, renal and cardiac arteries in in vivo L-NAME-induced hypertension model. In particular, unlike NaHS that causes immediate, transient falls in blood pressure, GYY4137 (acutely and chronically administered), promoted a slow-developing fall in blood pressure with minimal effects on the heart rate, in rats [60].

More recently, iminothioether chemical group has been recognized as potential new slow H₂S-releasing moiety endowed with vasorelaxing effects, on aortic and coronary arteries; furthermore, the blood pressure was reduced in L-NAME-treated rats [31].

Interestingly, arylthioamides and, more recently, the natural isothiocyanates occurring in the Brassicaceae family, have been recognized as sulfur compounds endowed with H₂S-releasing properties and with vasoactive effects [29,30].

Martelli and colleagues firstly demonstrated that erucin, the isothiocyanate deriving from arugula (Eruca sativa Mill. belonging to Brassicaceae family) is a smart H₂S-donor, which releases the gasotransmitter only in presence of thiol-groups, or in biological environment. Moreover, erucin showed hyperpolarizing activity on human aortic smooth muscle cells and vasoactive effects in ex vivo models: furthermore, in spontaneously hypertensive rats. erucin promoted anti-hypertensive effects [33]. Furthermore, previous studies reported that 4-month oral treatment with sulforaphane (SFN), which is another natural isothiocyanate, reduced blood pressure values in spontaneously hypertensive rats and increased glutathione levels in aortic smooth muscle cells [61.62].

Noteworthy zofenopril, which is a sulfur-containing ACE inhibitor, showed additional beneficial mechanism unrelated to ACE inhibition, but rather to H₂S release. Indeed, zofenopril promoted a more marked anti-hypertensive effect than enalapril when administered in vivo, and it rescued endothelium-dependent vasorelaxation in ex vivo models [63].

Accordingly with the role of H₂S in the regulation of haemodynamic parameters, the novel triphenyl phosphonium derivatized dithiolethione AP39, a slow H₂S-donor, significantly lowered systolic blood pressure, decreased heart rate and arterial stiffness in L-NAME treated rats [64].

Taken together these results reveal the crucial role of H₂S in regulating endothelial function, suggesting that this gasotransmitter, similarly to NO, might be a new tool for treating ED-related hypertension.

Hydrogen sulfide and atherosclerosis-related endothelial dysfunction

Atherosclerosis is a well-known disease triggered and sustained by vascular inflammation and characterized by intimal deposition

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of lipoprotein with consequent accumulation of lipids in arterial wall [65]. The rupture of atherosclerotic plaques or formation of thrombi on the atherosclerosis-induced alteration of vascular wall, are the most recognized causes of stroke or myocardial infarction which, in turn, represents the major cause of death among cardiovascular diseases. The onset of atherosclerosis begins as an immune response associated with endothelial dysfunction/disruption due to mechanical lesions, persistent chronic inflammatory stimuli or oxidative damages [66]. ED leads to the deficiency of those mechanisms deputed to maintain vascular homeostasis and prevents ECs inactivation against the pro-atherogenic stimuli, such as the generation of NO, the most known gasotransmitter. Besides the fundamental role of NO at cardiovascular level, in the last decades other two gasotransmitters have been investigated: carbon monoxide (CO) and H₂S and in particular, the last one was found to derive also from endothelium and to be fundamental in regulating vascular homeostasis [9,23,67]. Furthermore, H₂S preserves the vascular wall integrity, thus preventing the leaking of vascular tree induced by the vascular inflammation. Moreover, it limits the alteration of vascular tissues organization due to the atherosclerosis process [68]. One of the first insights about the role of endogenous H₂S in atherosclerosis derives from the work of authors who focused the attention on the ability of statins, the main class of drugs used to reduce lipid levels in the blood, to promote the generation of endogenous H₂S. In particular, these studies reported that the lipophilic atorvastatin and fluvastatin, but not the hydrophilic pravastatin, induced an enhanced H₂S generation from rats perivascular adipose tissue (by the inhibition of mitochondrial oxidation) and in mice lipopolysaccharide (LPS)-stimulated macrophages (reducing the levels of pro-inflammatory factors such as IL-1 β and monocyte chemoattractant protein-1 (MCP-1)) [69,70]. Besides these observations on the enhanced endogenous H₂S generation, several studies were carried out to investigate the role of H₂S in all the main aspects of atherosclerosis such as oxidation, adhesion, proliferation, HHcy and calcification (Fig. 2).

Using pharmacological/experimental tools like the salt NaHS as H_2S -donor agent, some authors demonstrated that low concentrations of H_2S were able to directly counteract ROS in vascular smooth muscle and, at the same time, to increase the activity of

antioxidant factors and pathways. A very important action of H₂S is the ability to inhibit the hypochlorite-induced modification of LDL into oxidized LDL (ox-LDL). Ox-LDL represents one of the most potent factors for the onset of atherogenic modifications, because they stimulate the expression of adhesion molecules on ECs triggering the inflammatory process and the atherosclerotic progression [11]. But the role of H_2S against the adhesion process is promoted not only through the indirect action on ox-LDL: Wang and colleagues demonstrated that H₂S inhibited the expression of intracellular adhesion molecule-1 (ICAM-1) in TNF-α-stimulated human umbilical vein ECs (HUVEC) and in apolipoprotein-E knockout $(apoE^{-}/^{-})$ mice through the inhibition of IkB degradation and NF-kB nuclear translocation. Targeting adhesion molecules is a fundamental strategy to inhibit the link between endothelium and immune cells like monocytes and T cells, since the inhibition of this first step precludes cell migration and the consequent inflammation which sustains atherosclerotic process [71]. However, H₂S limits the atherosclerotic process also by inhibiting the proliferation of intima and smooth muscle cells. This effect has been demonstrated in rat balloon injured arteries in which NaHS reduced neointimal lesions hyperplasia [72]. Moreover, NaHS also reduced the vascular smooth muscle proliferation, which represents a very harmful phenomenon concurring to the atherosclerotic plaque formation, through the migration of smooth muscle cells from the medial to the sub-endothelial layer where they proliferate, incorporate ox-LDL and thus contribute to the plaque development [73]. Finally, H₂S seems to also have a role in advanced phases of atherosclerosis, including angiosteosis and calcified vessels which show a down-regulation of CSE expression/activity resulting in a reduced production of H₂S. Wu and colleagues administered NaHS in a rat vascular calcification model obtained by the administration of vitamin D3 plus nicotine, and observed reduction of calcium content and accumulation in vessels, lower activation of alkaline phosphatase (a calcification inducer) and down-regulation of the expression of osteopontin gene (osteopontin is a glycoprotein involved in bone biomineralization and highly expressed in calcified plaques) [74]. On these bases, the use of H₂Sdonors seems to be a promising strategy to counteract the different phases of atherosclerotic process and several studies were carried

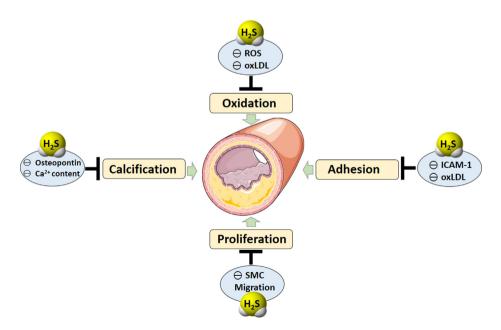


Fig. 2. Hydrogen sulfide counteracts the main aspects of atherosclerosis such as oxidation, adhesion, proliferation and calcification by inhibiting: the increase of ROS and ox-LDL, the expression of ICAM-1, the migration of smooth muscle cells (SMC) from the medial to the sub-endothelial layer where they proliferate, incorporate ox-LDL and so contribute to the plaque development; and finally by inhibiting the expression of osteopontin gene and the increase of vessels calcium content and accumulation.

out using the salt NaHS as H₂S-generating molecule. Indeed, the administration of NaHS prevents the development of atherosclerosis in $apoE^{-}/^{-}$ high fat-fed mice by downregulating the expression of chemokines receptors CX3CR1 and CX3CL1 both in ex vivo explanted plaques and in in vitro treated macrophages [71,75]. Furthermore, Xiong and co-workers in a recent work reported that NaHS enhanced atherosclerotic plaque stability, by increasing cap thickness through the inhibition of VSMCs apoptosis and decrease of the expression of metallopeptidase-9 (MMP-9), an enzyme deputed to collagen degradation, in $apoE^{-}/^{-}$ high fat-fed mice [76]. However, on the basis of current knowledge, the H₂S-donor salts like NaHS, are not the ideal H₂S-donors because of their property to release H₂S in a too much rapid way. Thus, the research of novel slow H₂S-donors and their evaluation on the atherosclerotic process represent a more interesting challenge. Presently, the most investigated slow H₂S-donor is GYY4137. Its anti-atherosclerotic properties has been investigated both in in vitro experiments on murine and human macrophages and in *in vivo* apoE⁻/⁻ high fatfed mice. In macrophages, GYY4137 inhibited foam cell generation induced by the administration of ox-LDL and down-regulated the expression and the activity of pro-inflammatory/atherogenic factors like NF-kB, ICAM-1 and chemokines. In vivo, GYY4137 counteracted the atherosclerotic plaque formation and restored the endothelium-dependent vasorelaxation, also leading to lower ICAM-1, IL-6 and TNF- α expression [77]. A similar investigation on *in vitro* models of macrophages and *in vivo* fat-fed apoE⁻/⁻ mice was carried out also with a particular class of H₂S-donor, i.e. the H₂S-donors hybrids [78–81]. These compounds were synthesized on the basis of previous experience on the NO-donor hybrids, in order to confer the positive effects of the gasotransmitter to selected native molecules. The typical native molecules used in this strategy were well-known drugs, commonly used in clinic, with the aims to improve their pharmacodynamics profile or to correct potential adverse effects of the native compound. This strategy was first applied to non-steroidal anti-inflammatory drugs (NSAIDs) and then applied on many other classes of drugs [82-85]. As concerns the hybrid H₂S-donors, the molecule named ACS14 is an early example. It is an H₂S-releasing aspirin, in which the H₂Sreleasing portion is represented by a dithiolthione moiety. ACS14 was tested both on macrophages stimulated with interferon- γ or LPS and compared with the native molecule aspirin. The results showed that only ACS14, but not aspirin, downregulated CX3CR1 via PPAR-y-dependent mechanism. The same downregulation of CX3CR1 was also found in brachiocephalic artery of $apoE^{-}/^{-}$ high fat-fed mice receiving ACS14 for 12 weeks but not in mice receiving aspirin. This CX3CR1 down-regulation correlated with prevention of atherogenesis and development of atherogenic plaques [86]. Among all the available H_2S -donors, the natural ones, i.e. H₂S-donors deriving from plants, represent a very interesting source of H₂S because of their nutraceutical added value. Garlic polysulfides, such as DADS and DATS, were the first described as natural H₂S-donors [25]. They have been demonstrated to inhibit the ox-LDL-induced expression of adhesion molecule like vascular adhesion molecule-1 (VCAM-1) and E-selectin. Such a property prevents monocyte adhesion in HUVEC cells and protects eNOS against the damage induced by ox-LDL [87,88]. In the last years, another family of plants emerged as a source of H₂S-donor molecules, i.e. the Brassicaceae family (Crucifers) in which the secondary metabolites isothiocyanates were well characterized as H₂Sdonors [33,89-91]. To date, SFN is the most investigated isothiocyanate and some recent articles described its inhibitory action on vascular inflammation and adhesion molecule expression in advanced glycation end products (AGEs)-treated HUVEC-cells and in AGEs-perfused rat aortas [92]. Moreover, in a more specific article, SFN was administered to high fat-fed rabbits and the results showed that SFN reduced the intima/media ratio, the levels of total

cholesterol, LDL, C-reactive protein, lactate dehydrogenase and the NF-kB expression. At the same time, SFN increased high-density lipoproteins (HDL), reduced glutathione levels and improved the aortic endothelium-dependent vasorelaxing effect after administration of acetylcholine, demonstrating an impressive impact on several atherogenic factors [93]. Finally, a very recent and interesting work by Bibli and colleagues investigated the link between H₂S, ED and atherosclerosis, purposing an original mechanism of action. In particular, they observed that both in cultured ECs and in mice, endogenous CSE-derived H₂S induced sulfhydration and dimerization of the RNA-binding protein human antigen R (HuR). Conversely, such a HuR sulfhydration was inhibited in atherosclerosis, leading to increased expression of CD62 antigen-like family member E (CD62E) and cathepsin S mRNAs, which are related to endothelium activation and atherogenic stimuli. As a further confirmation, the administration of the polysulfide SG1002, a slow polysulfide donor, in ECs isolated from CSE knockout mice, restored HuR sulfhydration with consequent decrease in levels of CD62E protein and in monocyte adherence. Moreover, SG1002, administered to apoE⁻/⁻ CSE knockout mice submitted to partial carotid ligation, limited the plaque formation and the HuR link to cathepsin S mRNA, demonstrating an H₂S-induced antiatherogenic effect [94].

Hydrogen sulfide and diabetes-related endothelial dysfunction

It was well established that ED could lead to a series of pathologies including cardiovascular diseases, atherosclerosis, and hypertension and is closely related to diabetes. Indeed, the alteration in endothelial functionality is extremely evident in patients affected by diabetes that is a pathological condition characterized by a high risk of cardiovascular disease, ascribed to the severe adverse effects of oxidative stress and hyperglycemia. Also, prediabetic patients, characterized by impaired glucose tolerance and impaired fasting glucose, develop with more frequency important cardiovascular disease [95]. These findings largely suggest that anomalies in carbohydrate metabolism are responsible for a progressive degeneration of healthy cardiovascular functions, where ED represents the first step of the undesirable events leading to a variety of pathological conditions [96]. ED in diabetes is linked to reduced bioavailability of NO, slightly compensated by endothelium-dependent hyperpolarization and/or production of prostacyclin, and increased production or action of endothelium-derived vasoconstrictors. This condition also leads to microvascular complications, including nephropathy, neuropathy and retinopathy, primarily characterized by abnormal angiogenesis, enhanced oxidative stress, increased production of inflammatory factors, decreased release of NO and impaired endothelial repair.

Additionally, the generation of AGEs is particularly intensified due to presence of elevated concentration of glucose, contributing to vascular complications.

In diabetic patients, ED seems to be a consistent hallmark which leads to an alteration of gasotransmitters i.e. NO, CO and H_2S production [10,97]. In fact, H_2S is mainly produced in ECs and is involved in the fine regulation of endothelial integrity and functions. For this reason, altered H_2S bioavailability has been proposed as a novel marker of ED advancement and prognosis [10]. According to the mentioned observations, an initial dysfunction of ECs triggers metabolic and vascular modifications associated with type 2 diabetes (Fig. 3).

 H_2S exhibits positive effects on vascular endothelium, emerging as a central regulator of vascular disease in diabetes, due to its crucial role in maintaining healthy functions in ECs [98,99]. In addition, H_2S contributes to wound healing and VSMC relaxation, thus decreasing blood pressure and platelet aggregation

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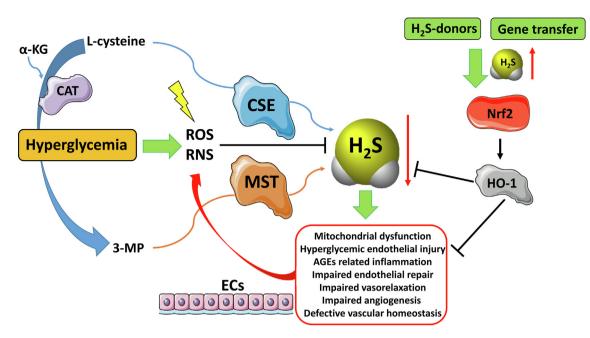


Fig. 3. Schematic representation of the effects of hyperglycemia on ECs. This event caused ED due to the formation ROS/RNS, leading to a decrease of gasotransmitter H_2S . H_2S deficiency is implied in a series of alteration of cellular processes highlighted in a red line box. The exogenous H_2S supplementation counteracts these dysfunctions stimulating the expression of HO-1 in an Nrf2-dependent manner. The picture was prepared by using Smart SERVIER MEDICAL ART provided under Creative Commons Attribution 3.0 Unported License. ROS/RNS (Reactive Oxygen Species/Reactive Nitrogen Species); α -KT (α -ketoglutarate); CAT (cysteine aminotransferase); CSE (cystathionine γ -lyase); MST (3-mercapto sulfortansferase); 3-MP (3-mercaptopyruvate); ECs (endothelial cells); Nrf2 (nuclear factor erythroid 2-related factor 2); HO-1 (heme oxygenase 1); AGEs (advanced glycation end products). (For interpretation of the references to color in this figure legend, the reader is referred to the web version of this article.)

[100,101]. It also exerts potent anti-apoptotic, antioxidant, angiogenic responses and anti-inflammatory. H₂S exerts several of its pharmacological effects by targeting proteins through Ssulfhydration. In this reaction, an atom of sulfur is added to the thiol groups of reactive cysteine residues resulting in the formation of hydropersulfides [102]. More recently, H₂S has been demonstrated to limit ED, nephropathy, cardiomyopathy, retinopathy in experimental animal models of diabetes [103–106], underlining the protective nature of this molecule [107]. Reliably, a shortfall in H₂S homeostasis is involved in the pathogenesis of hyperglycemic endothelial injury. Consequently, the use of H₂Sreleasing compounds or gene therapy to increase the level of endogenous H₂S could help in restoring endothelial function and antagonizing the progression of related diseases. For these reasons, we discuss about the potential role of H₂S-releasing agents as a novel therapeutic option for the treatment of diabetes-related ED.

In a ground-breaking study, Xie and colleagues [108] highlighted the significant role of H₂S in diabetes. They demonstrated, that this gasotransmitter counteracts diabetes-accelerated atherosclerosis, in a model of streptozotocin-induced diabetic high fat-fed mice. In the mentioned study they demonstrated that chronic administration of GYY4137, a slow-releasing H₂S-donor, reduced atherosclerotic lesion size in atheroprone animals, independently by any change in circulating blood cholesterol and glucose. Xie and colleagues also observed that the anti-atherosclerotic effect of GYY4137 is linked to a reduction in the amount of macrophages within the plaque, in the expression of the adhesion receptors and the production of superoxide. Similar effects also emerged when peritoneal macrophages or ECs are treated with ox-LDL and high concentrations of glucose, which is a condition that mimics the in vivo environment encountered in diabetes. In addition, the researchers observed that GYY4137 is able to activate the transcription of Nrf2 through the specific S-sulfhydration of amino acidic residue cysteine 151 (Cys151) in Kelch-like ECH-associated protein 1 (Keap1). Keap1 decrease the activity of Nrf2 by promoting its degradation through the ubiquitin-proteasome pathway. Fascinatingly, in diabetic mice the antiatherogenic action of GYY4137 is completely abolished when Nrf2 is deleted or silenced, highlighting that the action is mediated by Nrf2. Moreover, the compound increases the trascription of HO-1 in an Nrf2dependent manner, and inhibition or depletion of HO-1 completely reduces the beneficial actions of GYY4137, indicating that HO-1 is involved in the antiatherogenic effects of H₂S. Notably, Xie and coworkers detected significantly lower levels of plasma H₂S in diabetic mice, and demonstrated that the administration of GYY4137 restored physiological levels of H₂S. In patients with diabetes and other diabetic animal models a reduction in circulating H₂S has also been reported [109,110], confirming an H₂S deficiency in diabetic pathology. The cause for this decline is still unclear but may be associated to alterations in the global activity of H₂S-generating enzymes, the microbial reduction of sulfate in the intestine, the generation of H₂S from other sources and/or the metabolism of H₂S in diabetes. Clearly, more detailed studies are needed to solve this issue and to discover optimal circulating concentrations of H₂S needed to maintain vascular homeostasis in diabetes.

Gero and coworkers [111] studied the therapeutic potential of two novel mitochondria-targeted hydroxythiobenzamide and anethole dithiolethione H₂S-donors (AP123 and AP39 respectively) in diabetic cardiovascular complications, which are, at least in part, promoted by mitochondrial ROS production in ECs. High concentration of glucose in the bloodstream is responsible for superoxide production in the mitochondria and triggers modifications in the mitochondrial membrane potential, leading to mitochondrial dysfunction. Since H₂S supplementation reduces the mitochondrial oxidant production and shows efficacy against diabetic vascular damage in vivo, the role of AP39 and AP123 has been evaluated in preventing hyperglycemia-induced oxidative stress and metabolic alterations in microvascular ECs. AP39 and AP123 reduced hyperglycemia-induced mitochondrial membrane hyperpolarization and repressed the mitochondrial ROS production. Both H₂Sreleasing compounds improved the cellular metabolism and enlarged the electron transport at respiratory complex III. The high

potency and long-lasting effect provoked by these H₂S-donors suggest that these classes of compounds could be useful for the treatment of diabetic vascular complications.

Another work by Ng and colleagues [112] highlighted how NaHS chronic treatment is able to decrease the oxidative stress and exerts vasoprotective effects, improving endothelial function in streptozotocin-induced diabetes C57BL6/J mice. In particular, NaHS treatment restored endothelial function reverting diabetesinduced vascular dysfunction by improving NO efficacy and reducing superoxide production in the mouse aorta.

In another study, Ma and colleagues [113] demonstrated that the exogenous supplementation of H_2S ameliorated diabetesassociated cognitive decline (DACD). This disorder is related to ED and represents one of the complications of diabetes often leading to cognitive impairment and an augmented risk of dementia. The researchers investigated the anti-inflammatory and antiapoptotic effects of H_2S on DACD and demonstrated that H_2S improved the spatial learning and memory abilities of the diabetic mice by modulating the IL-23/IL-17 axis and the mitochondrial apoptotic pathway, which were discovered to be associated with DACD. Accordingly, H_2S treatment may help preventing the progression of apoptotic hippocampal neurons and EC in diabetic mice and could represent an innovative therapeutic strategy for the treatment of DACD.

Recently, H₂S-donors derived from natural source are attracting the attention of researchers for the discovery of novel therapeutics strategies to treat different diseases including diabetes and diabetes-related ED. In particular, organosulfur compounds (OSCs) occurring in plant extracts can behave as cardio protective agents in type 2 diabetes, due to their immunomodulatory, antioxidant, anti-inflammatory and hypoglycemic effects. OSCs contained in garlic (Allium sp.), due to their properties, seem to be a valuable dietary support in type 2 diabetes, as reported in several studies. Garlic OSCs are able to produce H₂S, and several their beneficial effects are probably due to this property. In fact, recently, many studies reported the relevant effects of exogenous and endogenous H₂S in diabetes, including in vivo and in vitro experiments and clinical trials as nicely reviewed by Melino and colleagues [114]. Briefly, garlic has been demonstrated to exert beneficial effects in diabetes, including hypocholesterolemia, hyperinsulinemia, antilipid peroxidation, hypoglycemia, hypotriglyceridemia and antiglycation [114]. As reported by Padiya and colleagues [115], garlic extract improved insulin sensitivity in a model of insulinresistance, associated with metabolic syndromes in fructose-fed diabetic rats treated with garlic homogenate. In particular, they showed a significant decrease in serum glucose, triglycerides and uric acid levels. Garlic also normalized the serum levels of endogenous NO and H₂S after fructose feeding, suggesting also an improvement in the endothelium functionality and an attenuated metabolic syndrome.

More recently, Thomson and coworkers showed the antidiabetic and antioxidant potential of aged garlic extract in streptozotocin-induced diabetic rats. The diabetic rats showed an elevated blood glucose, serum cholesterol and triglycerides, erythrocyte glycated hemoglobin and kidney and liver lipid peroxidation. Interestingly, the treatment with aged garlic extract positively reverted the diabetic alterations to significantly lower levels than those measured in diabetic rats.

Baluchnejadmojarad and colleagues [116] investigated the antidiabetic, antioxidant, anti-inflammatory and normotropic property of S-allyl cysteine (SAC), the most abundant organosulfur bioactive molecule in aged garlic extract. The mentioned work was designed to assess the therapeutic potential of SAC in limiting memory and learning deficits in streptozotocin-induced diabetic rats and to evaluate the involvement of toll-like receptor 4 (TLR4), Nrf2, nuclear factor-kappa B (NF- κ B), and HO-1 signaling

cascades. SAC treated rats showed a decrease of serum glucose, and an improvement in spatial recognition memory in several *in vivo* tests. Moreover, SAC reduced lipid peroxidation marker malondialdehyde (MDA), acetylcholinesterase activity, and augmented antioxidant defensive system including catalase, superoxide dismutase (SOD), and reduced glutathione in hippocampal lysate. In addition, SAC lowered hippocampal TLR4, NF-kB and TNF- α and prevented reduction of HO-1 and Nrf2 in diabetic rats. Taken together, chronic SAC treatment ameliorated cognitive deficits in diabetic rats. Unfortunately, these encouraging results were not found during a recent clinical trial.

Also, sulfur-containing compounds deriving from the Brassicaceae family could represent valuable starting point for drug development against diabetes and its related diseases. In fact, SFN, an isothiocvanate found in Broccoli, has been proposed as potential antidiabetic agents. In a clinical study SFN was administered (112 or 225 umol/day for 4 weeks) to diabetic patients. The results showed reduction in fasting glucose, inflammatory markers and serum insulin. Moreover, in the same randomized doubleblind placebo-controlled clinical trial, a decrease in serum triglycerides was observed. Furthermore, they reported a lowered density of lipoprotein ox-LDL/LDL ratio, reduced atherogenic index of the plasma and an increase in HDL concentration in patients receiving the highest dose of SFN [117,118]. Axelsson and colleagues observed that SFN reduced hepatic glucose production and improved glucose control in patients with type 2 diabetes. Patients were treated with broccoli sprout extract containing SFN which interestingly, reduced glycated hemoglobin (HbA1c) and fasting blood glucose in obese patients with dysregulated type 2 diabetes [119]. Moreover, SFN was shown to arrest the progression of type 1 diabetes in streptozotocin treated mice by inhibiting oxidative stress-induced beta cell damage and protecting ECs against diabetes-induced complications [120]. SFN is also known to prevent aortic damage in streptozotocin-induced diabetes in type 1 diabetes mice [121].

Additionally, many authors demonstrated that inhibition of AGEs formation may be a valid therapeutic strategy to improve vascular complications in diabetes. AGEs directly blocks NO activity and produces ROS in vascular endothelium. The diabetic condition is characterized by low levels of NO at the vascular level due to oxidative stress and increased AGEs levels which contribute to the worsening of the endothelium functionality. Diabetic rats treated with SFN showed a significant decreased in vascular oxidative stress, and concomitantly increased NO bioavailability, partially explaining the beneficial effects on vascular function [122].

Summarizing, in diabetes, H₂S-donors showed a wide spectrum of beneficial effects counteracting diabetic complications. The evidence on the beneficial effects of H₂S-donors included the restoration of H₂S level, since in diabetes there is a decrease of H₂S bioavailability [123]. In addition, in overweight and obese patients, the H₂S levels in plasma are reduced, and this event represents a feature of metabolic syndrome and it is commonly observed in type 2 diabetes [40]. H₂S or its donors have an antiatherogenic property and act by inhibiting the oxidation of LDL as a result of scavenging the free radicals (notably hypochlorous acid and hydrogen peroxide), inhibition of the myeloperoxidase enzyme, and inhibition of the foam cell formation by several mechanisms [124]. Based on this evidence, the use of H₂S-releasing compounds is a highly feasible approach that can be used to augment H₂S levels in diabetic patients. Along with inorganic salts and natural H₂S-donors, many synthetic compounds have been developed, possessing better H₂S release kinetics and pharmacokinetic profiles [91]. Alternatively, endogenous circulating levels of H₂S may be increased by gene delivery of H₂S-generating enzymes or by the supplementation of dietary sulfur that is readily converted to H₂S by the gut microbiome. Strategies employing these approaches in

clinical study, could provide in the near future innovative drug candidates for the prevention and treatment of diabetes and its related diseases.

Hydrogen sulfide and hyperhomocysteinemia-related endothelial dysfunction

Hcy is a thiol-containing amino acid deriving from essential amino acid methionine through remethylation reaction, mainly occurring in the liver. Hcy metabolism, described more in detail in Fig. 4, involves three different enzymes, such as betaine-Hcy methyltransferase (BHMT), CBS and CSE [125].

About the 5–10% of the total daily cellular production of Hcy is not metabolized within the cells but is released into the bloodstream compartment. In healthy subjects the concentration of free Hcy is maintained below 15 μ mol/L thanks to a constant renal clearance [126]. If the regulation of intracellular Hcy levels is disrupted, Hcy extrusion to the plasma compartment increases and consequently high levels of Hcy are accumulated in the bloodstream [127]. The free Hcy excess into the blood prevents cell toxicity, but on the other hand, exposes vascular tissue to a possible deleterious effect [128].

High tissue and plasma Hcy levels (i.e. HHcy) is an independent risk factor for cerebrovascular, cardiovascular and peripheral artery disease [129]. The pathogenesis of HHcy is mainly due to impaired renal clearance, malnutrition that may cause vitamin B12 and folate deficiency, methionine-rich meat diet or polymorphism in CBS gene [125]. HHcy-related vascular diseases are strictly linked to ED since elevated levels of Hcy have been reported to be directly harmful for the ECs both in *in vitro* and *in vivo* models. Consequently, Hcy exerts its adverse effects by disturbing endothelial function [130]. Accordingly, many studies demonstrate a clear association between HHcy and coronary artery disease, atherosclerosis, hypertension and neurodegenerative diseases [131]. Indeed, HHcy can be considered as a predictive biomarker of cardiovascular mortality in patients affected by coronary artery disease, and plasma levels of Hcy correlate with the severity of atherosclerosis [132].

The HHcy-related ED is a complex mechanism that triggers secondary cardiovascular diseases including atherosclerosis, hypertension and neurodegeneration by inducing oxidative stress and inflammatory status. HHcy increases ROS production through different mechanism, as Hcy autoxidation. Indeed, Hcy can bind plasma proteins or a second Hcy molecule through a disulfide bridge. In these cases, the free thiol group of Hcy undergoes to autoxidation, generating H₂O₂, ROS, hydroxyl radical and superoxide anion which can in turn react with NO to produce peroxynitrite, leading to a decrease in the bioavailability of NO [133]. Secondary, HHcy creates a disequilibrium between antioxidant and oxidant enzymes, i.e. inhibition of SOD or activation of NADPH oxidases [134]. Furthermore, both in vitro and in vivo experiments have reported that HHcv can block the glutathione peroxidase activity and HO-1, contributing to ROS accumulation and exacerbating the ECs damage [135,136].

Beside the dramatic increase of the oxidative stress, HHcy also promotes a proinflammatory effect, through the activation of those genes that regulate the expression of chemokines, cytokines and adhesion molecules, such as NF-kB: Hcy and ROS promote NF-kB translocation into the nucleus where activates specific target genes. In particular, HHcy-related NF-kB activation has been demonstrated to promote the expression of ICAM-1, MCP-1, VCAM-1 and E-selectin [137]. HHcy induces vascular inflammation also involving migration of leukocytes from the vessels to the underlying tissues by increasing the expression of CD11B/CD18 proteins which are responsible to promote the interaction between the endothelium and the inflammatory cells [138]. Moreover, HHcy causes vascular fibrosis due to activation of matrix metalloproteinases (MMPs), leading to the alteration of extracellular matrix (ECM) metabolism and promotion of collagen deposition [139] (see Fig. 5).

The functional importance of the endothelium also lies in the maintenance of antithrombotic surface for blood flow and hemostasis by controlling the equilibrium between the production

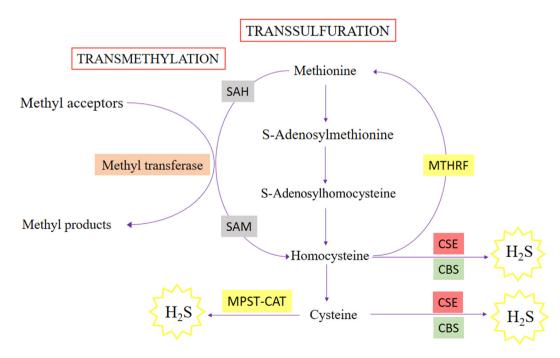


Fig. 4. Homocysteine (Hcy) metabolism in physiological conditions. Hcy derives from the hydrolysis of S-Adenosyl-L-homocysteine (SAH) and is metabolized through *trans*-sulfuration pathway and remethylated to methionine via the remethylation pathway. When methionine is in excess, Hcy goes to *trans*-sulfuration pathway which is catalyzed by cystathionine β-synthase (CBS) and cystathionine γ-lyase (CSE).

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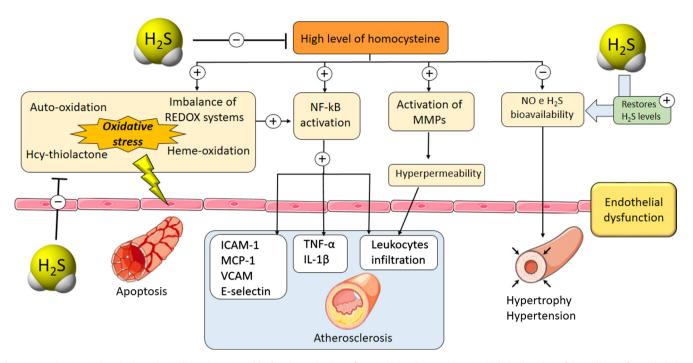


Fig. 5. Hyperhomocysteinemia (HHcy) condition is responsible for the activation of several signaling pathways which lead to harmful conditions for endothelium functionality. Firstly, HHcy creates a severe oxidative stress due to the formation of reactive intermediates (Hcy-thiolactone, Hcy persulfide etc.) which contribute to the imbalance of crucial REDOX enzyme systems (inhibition of SOD or activation of NADPH oxidases). Furthermore, HHcy and ROS activate NF-kB signaling pathway resulting in a marked inflammatory response through the production of cytokines (TNF- α , Il-1 β); simultaneously pro-atherosclerotic mediators are overexpressed (ICAM-1, MCP-1, VCAM, E-selectin) contributing to the formation of atherosclerotic lesion. HHcy also activates MMPs increasing endothelium permeability which facilitates leukocyte infiltration in the underlying tissues. Furthermore, HHcy is responsible for a decrease in the bioavailability of fundamental vasodilator molecules, including NO and H₂S promoting vascular hypertrophy and hypertension. In this context, H₂S is able to beneficially act at different levels: H₂S is a well-known ROS scavenger, thus exogenous administration of H₂S-donor significantly reduces the oxidative stress, limiting the vascular inflammation. Furthermore, H₂S directly reduces the Hcy plasma levels, thus preventing the activation of the downstream pathways. Finally, H₂S limits the HHcy-induced hypertension and vascular hypertrophy.

of pro- and anti-coagulant molecules. HHcy promotes the activation of coagulation process by inhibiting thrombomodulindependent protein C activation, impairing production of von Willebrand factor (VWF) and activating factor V [140]. Consistently, high levels of Hcy have been reported in patients affected by acute coronary artery disease and venous thrombosis [141].

HHcy is also involved in atherosclerosis disease. Indeed, the apoptotic process occurring in ECs is an important hallmark of atherosclerosis and leads to the generation and the disruption of atherosclerotic plaque. In this process, HHcy mediates apoptotic cell death of smooth muscle and ECs [142]. Other studies demonstrated the contribution of HHcy-induced inflammation in the atherosclerotic plaques using hyperhomocysteinemic apoE⁻/⁻-deficient mice, revealing the activation of NF- κ B and downstream proinflammatory mediators in atherosclerotic lesions [143]. Moreover, Hcy high plasma levels induced ECs apoptosis due to endoplasmic reticulum stress, enhancement in ROS production and Hcy-thiolactone generation [144].

The clear association between cardiovascular disease and HHcy contributed to highlighting the great importance of ECs and demonstrated the interference of Hcy in maintaining the endothelium functionality. In particular, HHcy reduces NO bioavailability [145], interferes with the H₂S signaling pathway [146], enhances oxidative stress [147], perturbs lipoprotein metabolism and protein N-homocysteinylation [148].

Among the vasodilators endothelial factor, H_2S emerged as an important gasotransmitter regulating endothelium functionality and cardiovascular homeostasis and many evidences demonstrate that HHcy leads to severe deficiency in H_2S bioavailability. Accordingly, exogenous supplement of H_2S is likely to be considered a pharmacological tool for limiting the HHcy-related ED.

In particular, impaired H₂S bioavailability has been related to prognosis and progression of HHcv-related ED, since high levels of Hcv has been demonstrated to suppress expression/activity of the H₂S-generating enzymes CBS and CSE, reducing the endogenous biosynthesis of H₂S in cultured ECs [149]. The impairment of H₂S production linked to high level of Hcy has been demonstrated by Man-Hong and colleagues who showed that intracerebroventricular injection of Hcy caused ER stress in the cerebral ECs, worsened the learning and memory, and downregulated CBS expression resulting in a limited generation of H₂S. Since H₂S plays an important role in regulating ER stress, this study suggested that Hcy-induced loss of learning and memory is related to the lowered bioavailability of endogenous H₂S and the increase in ER stress [150]. Indeed, the restoration of H₂S levels with NaHS treatment significantly limited cerebrovascular neurodegeneration and dysfunction. Such a protection is associated with the reduction of oxidative stress, associated with decreased of malondialdehvde and superoxide, and with the simultaneous increased of glutathione, SOD, catalase [151,152]. The above-mentioned studies demonstrated that H₂S supplement with the administration of NaHS increases the levels of antioxidant molecules. This is probably due to the H₂S ability in enhancing the activity of γ -glutamylcysteine synthetase and upregulating transport of cysteine, strictly related to the synthesis of glutathione [153]. Furthermore, H₂S may also behave like a scavenger towards the HHcy-induced hemoglobin oxidation species, limiting the lipid peroxidation in vessel endothelium [154].

Different *in vitro* cell culture experiments showed that the treatment with NaHS significantly reduced RNS/ROS levels and normalized redox enzyme levels in VSMC and bEnd3 ECs (brain ECs) pretreated with methionine, which is the precursor of Hcy

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[155,156]. Further investigations highlighted the crucial role of mitochondria in Hcy-induced RNS/ROS production leading to mitochondria toxicity and cell death through the activation of N-methyl-D-aspartate-R1 (NMDA-R1) in ECs. Moreover, Hcy treatment lead to an impairment in ATP mitochondrial production. The treatment with NaHS restored the ATP levels and prevented the NMDA-R1 activation, thus protecting ECs from Hcy damage [157]. The authors also demonstrated that the epigenetical modulation of CSE expression enhanced the beneficial role of endogenous H₂S in inhibiting HHcy-mitochondrial toxicity and reducing mitochondrial superoxide formation [157]. In addition, Hcy enhanced ROS production in isolated mitochondria from mouse aortic ECs, triggering the mitophagy process. Interestingly, epigenetical modulation of H₂S generating enzymes (i.e. CSE or CBS), limited mitophagy by inhibiting ROS production [156].

Another study reported that dietary supplementation with methionine in drinking water promoted the increase of Hcy and inflammatory cytokines (TNF- α , IL-1 β) plasma levels, associated with impairment of endogenous H₂S production in C57BL/6 mice. Moreover, HHcy significantly downregulated CSE expression in the intestinal macrophages extracted from the HHcy-mice compared to control mice. To clarify whether H₂S impairment was correlated with the increase of pro-inflammatory cytokines, methionine-fed mice were treated with GYY4137, an H₂S-releasing molecule, and DL-propargylglycine or D-penicillamine, which are two CSE inhibitors: GYY4137 markedly decreased TNF- α and IL-1 β amount in the plasma of methionine-treated mice; on the contrary, DL-propargylglycine and D-penicillamine exacerbated the increase of these cytokines level [158,159].

In addition, Hcy sustains and promotes ED and causes vascular hyperpermeability by increasing the activity of MMPs which are clearly involved in the atherosclerotic plaque formation, since they can facilitate vascular remodeling by digesting the matrix detaching the intima from the media tunica and allow smooth-muscle cell migration from the media to the intima. H₂S is able to inhibit Hcy-induced MMPs activity leading to a decrease of smooth muscle hyperproliferation and suppression of vascular remodeling and inflammation [160,161].

Beside the downregulation of the H_2S generating enzymes, Hcy can be also considered as a scavenger of hydrosulfide anion to form Hcy persulfide. In HHcy conditions, the accumulation of Hcy persulfide weakened the cardioprotective effect of NaHS in HHcy animals subjected to ischemia–reperfusion injury [162].

HHcy has been demonstrated to reduce NO biosynthesis by inhibiting eNOS and activating arginase enzyme. *In vitro* and *in vivo* studies reported that H₂S is able to counteract such a NO decrease by mainly increasing eNOS mRNA synthesis, maintaining soluble guanylate cyclase and inhibiting PDE-5 activity [26,52].

Although the promising results about the and effectiveness of NaHS in reducing the HHcy-related ED, this H₂S-donor is characterized by a fast H₂S donation which makes it unusable in the clinical practice due to its toxicity and short half-life. Consequently, the research in this field focused on developing safe H₂S-based drugs, with reduced toxicity and enhanced efficacy, in particular the H₂S prodrug SG1002 is likely to be a very promising pharmacological therapy for heart failure. SG1002 is an inorganic mixture (sodium polysulthionate) containing S₈, Na₂SO₄, Na₂S₂O₃, Na₂S₃O₆, Na₂S₄O₆, and Na₂S₅O₆ and promoted cardiac remodeling and afterload in the CBS+/- hyperhomocysteinemic mouse model. CBS+/mice developed several symptoms of early and late cardiac modification, including hypertrophy and fibrosis that lead in enhanced afterload on the heart. These tissue modifications were limited by dietary supply of the SG1002 which normalized the histological and molecular measurements of hypertrophy and fibrosis, since SG1002 significantly decreased collagen accumulation in the left ventricle of hyperhomocysteinemic mice, inactivated MMP and limited cardiomyocyte size [163].

Also naturally occurring isothiocyanates may be exploited for the treatment of HHcy-related ED, since many studies demonstrated their ability to donate H_2S [89]. Indeed, preliminary data reported that SFN, which is a natural isothiocyanate able widely present in the *Brassicaceae* family, exerts protective effects on Hcy-induced oxidative stress *in vitro* by reducing ER stress and behaving as ROS scavenger [164].

Conclusions

The ED refers to as the loss of ECs physiological functions caused by different risk factors, including hyperglycemia, hypercholesterolemia and HHcy. ECs respond to such pathological conditions by enhancing the oxidative stress along the vascular tree, leading to several damaging events resulting in the inability of the endothelium to exert its fundamental activities, i.e. the regulation of vascular relaxation and/or cell redox balance, NO production and vascular smooth muscle dilation. ED comprehends the alteration of fundamental aspects in cell and tissue metabolism, in regulating their homeostasis and development, and in strengthening their ability to react to stress situations. The complexity of this phenomenon involves a large number of actors and effectors and among these, H₂S emerged as one of the main characters in the regulation of endothelial functionality. Indeed, H₂S controls the homeostasis of endothelial function and an impairment of its endogenous production is related to the pathogenesis of ED. For this reason, strategies employing compounds able to donate H₂S could provide in the near future innovative drug candidates for the prevention and treatment of ED in different pathological conditions, such as hypertension, diabetes and atherosclerosis.

Thanks to the intense efforts that have recently been implemented by biomedical research in this field, we presently know that many compounds are actually druggable H₂S-donors and the searching for molecules that may exhibit a satisfactory and physiological-like H₂S generation represents a challenging and timely issue. It is noteworthy that several drugs are currently under evaluation in promising clinical studies. Among the several molecules, a very significant part is represented by sulfur compounds of natural origin which could represent a useful pharmacological tool to be used in therapy but could also provide a precious template for the design of new H₂S-donor molecules with improved pharmacodynamic and/or pharmacokinetic characteristics.

Although the data reported in the literature demonstrate that molecules able to donate H_2S may represent a useful tool in the protection of ECs in different pathological conditions, further clinical studies are needed. Indeed, there are no specific clinical trials involving receiving H_2S -donors patients affected by those pathologies that leads to ED. These studies would be very useful, to investigate the reduction, or the slowing down of ED related to specific pathological conditions (diabetes, HHcy, atherosclerosis or hypertension).

Compliance with Ethics Requirements

This article does not contain any studies with human or animal subjects.

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Declaration of Competing Interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

References

- Kruger-Genge A, Blocki A, Franke RP, Jung F. Vascular endothelial cell biology: an update. Int J Mol Sci 2019;20. doi: <u>https://doi.org/10.3390/ijms20184411</u>.
- [2] Charbonier FW, Zamani M, Huang NF. Endothelial cell mechanotransduction in the dynamic vascular environment. Adv Biosyst 2019;3. doi: <u>https://doi.org/10.1002/adbi.201800252</u>.
- [3] Deanfield JE, Halcox JP, Rabelink TJ. Endothelial function and dysfunction: testing and clinical relevance. Circulation 2007;115:1285–95. doi: <u>https://doi.org/10.1161/CIRCULATIONAHA.106.652859</u>.
- [4] Favero G, Paganelli C, Buffoli B, Rodella LF, Rezzani R. Endothelium and its alterations in cardiovascular diseases: life style intervention. Biomed Res Int 2014;2014:. doi: <u>https://doi.org/10.1155/2014/801896</u>801896.
- [5] Cheng HM, Koutsidis G, Lodge JK, Ashor A, Siervo M, Lara J. Tomato and lycopene supplementation and cardiovascular risk factors: a systematic review and meta-analysis. Atherosclerosis 2017;257:100–8. doi: <u>https://doi.org/10.1016/i.atherosclerosis.2017.01.009</u>.
- [6] Kvietys PR, Granger DN. Role of reactive oxygen and nitrogen species in the vascular responses to inflammation. Free Radic Biol Med 2012;52:556–92. doi: <u>https://doi.org/10.1016/j.freeradbiomed.2011.11.002</u>.
- [7] Meier M, Janosik M, Kery V, Kraus JP, Burkhard P. Structure of human cystathionine beta-synthase: a unique pyridoxal 5'-phosphate-dependent heme protein. EMBO J 2001;20:3910–6. doi: <u>https://doi.org/10.1093/emboi/ 20.15.3910</u>.
- [8] Stipanuk MH. Sulfur amino acid metabolism: pathways for production and removal of homocysteine and cysteine. Annu Rev Nutr 2004;24:539–77. doi: <u>https://doi.org/10.1146/annurev.nutr.24.012003.132418</u>.
- [9] Martelli A, Testai L, Marino A, Breschi MC, Da Settimo F, Calderone V. Hydrogen sulphide: biopharmacological roles in the cardiovascular system and pharmaceutical perspectives. Curr Med Chem 2012;19:3325–36. doi: https://doi.org/10.2174/092986712801215928.
- [10] Wang R, Szabo C, Ichinose F, Ahmed A, Whiteman M, Papapetropoulos A. The role of H2S bioavailability in endothelial dysfunction. Trends Pharmacol Sci 2015;36:568–78. doi: <u>https://doi.org/10.1016/j.tips.2015.05.007</u>.
- [11] Laggner H, Muellner MK, Schreier S, Sturm B, Hermann M, Exner M, et al. Hydrogen sulphide: a novel physiological inhibitor of LDL atherogenic modification by HOCI. Free Radic Res 2007;41:741–7. doi: <u>https://doi.org/ 10.1080/10715760701263265</u>.
- [12] Perna AF, Sepe I, Lanza D, Capasso R, Zappavigna S, Capasso G, et al. Hydrogen sulfide reduces cell adhesion and relevant inflammatory triggering by preventing ADAM17-dependent TNF-alpha activation. J Cell Biochem 2013;114:1536–48. doi: <u>https://doi.org/10.1002/jcb.24495</u>.
- [13] Kanagy NL, Szabo C, Papapetropoulos A. Vascular biology of hydrogen sulfide. Am J Physiol Cell Physiol 2017;312:C537–49. doi: <u>https://doi.org/10.1152/aipcell.00329.2016</u>.
- [14] Wang Y, Wang X, Liang X, Wu J, Dong S, Li H, et al. Inhibition of hydrogen sulfide on the proliferation of vascular smooth muscle cells involved in the modulation of calcium sensing receptor in high homocysteine. Exp Cell Res 2016;347:184–91. doi: <u>https://doi.org/10.1016/i.vexcr.2016.08.004</u>.
- [15] Zavaczki E, Jeney V, Agarwal A, Zarjou A, Oros M, Katko M, et al. Hydrogen sulfide inhibits the calcification and osteoblastic differentiation of vascular smooth muscle cells. Kidney Int 2011;80:731–9. doi: <u>https://doi.org/ 10.1038/ki.2011.212</u>.
- [16] Emerson M. Hydrogen sulfide and platelets: a possible role in thrombosis. Handb Exp Pharmacol 2015;230:153–62. doi: <u>https://doi.org/10.1007/978-3-319-18144-8_7</u>.
- [17] Marino A, Martelli A, Citi V, Fu M, Wang R, Calderone V, et al. The novel H2 S donor 4-carboxy-phenyl isothiocyanate inhibits mast cell degranulation and renin release by decreasing intracellular calcium. Br J Pharmacol 2016;173:3222–34. doi: <u>https://doi.org/10.1111/bph.13583</u>.
- [18] Weber GJ, Pushpakumar S, Tyagi SC, Sen U. Homocysteine and hydrogen sulfide in epigenetic, metabolic and microbiota related renovascular hypertension. Pharmacol Res 2016;113:300–12. doi: <u>https://doi.org/ 10.1016/j.phrs.2016.09.002</u>.
- [19] Vanhoutte PM, Shimokawa H, Tang EH, Feletou M. Endothelial dysfunction and vascular disease. Acta Physiol (Oxf) 2009;196:193–222. doi: <u>https://doi.org/10.1111/i.1748-1716.2009.01964.x</u>.
- [20] Majzunova M, Kvandova M, Berenyiova A, Balis P, Dovinova I, Cacanyiova S. Chronic NOS inhibition affects oxidative state and antioxidant response differently in the kidneys of young normotensive and hypertensive rats. Oxid Med Cell Longev 2019;2019:5349398. doi: <u>https://doi.org/10.1155/2019/</u> 5349398.
- [21] Berenyiova A, Dovinova I, Kvandova M, Kristek F, Jansen E, Majzunova M, et al. The effect of chronic NO synthase inhibition on the vasoactive and

structural properties of thoracic aorta, NO synthase activity, and oxidative stress biomarkers in young SHR. Oxid Med Cell Longev 2018;2018:2502843. doi: https://doi.org/10.1155/2018/2502843.

- [22] Wang R. Hydrogen sulfide: a new EDRF. Kidney Int 2009;76:700-4. doi: https://doi.org/10.1038/ki.2009.221.
- [23] Yang G, Wu L, Jiang B, Yang W, Qi J, Cao K, et al. H2S as a physiologic vasorelaxant: hypertension in mice with deletion of cystathionine gammalyase. Science 2008;322:587–90. doi: <u>https://doi.org/</u> 10.1126/science.1162667.
- [24] Zhao W, Zhang J, Lu Y, Wang R. The vasorelaxant effect of H(2)S as a novel endogenous gaseous K(ATP) channel opener. EMBO J 2001;20:6008–16. doi: <u>https://doi.org/10.1093/emboj/20.21.6008</u>.
- [25] Benavides GA, Squadrito GL, Mills RW, Patel HD, Isbell TS, Patel RP, et al. Hydrogen sulfide mediates the vasoactivity of garlic. Proc Natl Acad Sci U S A 2007;104:17977–82. doi: <u>https://doi.org/10.1073/pnas.0705710104</u>.
- [26] Bucci M, Papapetropoulos A, Vellecco V, Zhou Z, Pyriochou A, Roussos C, et al. Hydrogen sulfide is an endogenous inhibitor of phosphodiesterase activity. Arterioscler Thromb Vasc Biol 2010;30:1998–2004. doi: <u>https://doi.org/10.1161/ATVBAHA.110.209783</u>.
- [27] Coletta C, Papapetropoulos A, Erdelyi K, Olah G, Modis K, Panopoulos P, et al. Hydrogen sulfide and nitric oxide are mutually dependent in the regulation of angiogenesis and endothelium-dependent vasorelaxation. Proc Natl Acad Sci U S A 2012;109:9161–6. doi: <u>https://doi.org/10.1073/pnas.1202916109</u>.
- [28] Martelli A, Testai L, Breschi MC, Lawson K, McKay NG, Miceli F, et al. Vasorelaxation by hydrogen sulphide involves activation of Kv7 potassium channels. Pharmacol Res 2013;70:27–34. doi: <u>https://doi.org/10.1016/j.phrs.2012.12.005</u>.
- [29] Martelli A, Testai L, Citi V, Marino A, Pugliesi I, Barresi E, et al. Arylthioamides as H2S donors: I-cysteine-activated releasing properties and vascular effects in vitro and in vivo. ACS Med Chem Lett 2013;4:904–8. doi: <u>https://doi.org/ 10.1021/ml400239a</u>.
- [30] Martelli A, Testai L, Citi V, Marino A, Bellagambi FG, Ghimenti S, et al. Pharmacological characterization of the vascular effects of aryl isothiocyanates: is hydrogen sulfide the real player?. Vascul Pharmacol 2014;60:32–41. doi: <u>https://doi.org/10.1016/j.vph.2013.11.003</u>.
- [31] Barresi E, Nesi G, Citi V, Piragine E, Piano I, Taliani S, et al. Iminothioethers as hydrogen sulfide donors: from the gasotransmitter release to the vascular effects. J Med Chem 2017;60:7512–23. doi: <u>https://doi.org/10.1021/acs.jmedchem.7b00888</u>.
- [32] Martelli A, Citi V, Calderone V. Vascular effects of H2S-donors: fluorimetric detection of H2S generation and ion channel activation in human aortic smooth muscle cells. Methods Mol Biol 2007;2019:79–87. doi: <u>https://doi.org/10.1007/978-1-4939-9528-8 6</u>.
- [33] Martelli A, Piragine E, Citi V, Testai L, Pagnotta E, Ugolini L, et al. Erucin exhibits vasorelaxing effects and antihypertensive activity by H2 S-releasing properties. Br J Pharmacol 2019. doi: <u>https://doi.org/10.1111/bph.14645</u>.
- [34] Testai L, Marino A, Piano I, Brancaleone V, Tomita K, Di Cesare Mannelli L, et al. The novel H2S-donor 4-carboxyphenyl isothiocyanate promotes cardioprotective effects against ischemia/reperfusion injury through activation of mitoKATP channels and reduction of oxidative stress. Pharmacol Res 2016;113:290–9. doi: <u>https://doi.org/10.1016/j. phrs.2016.09.006.</u>
- [35] Severino B, Corvino A, Fiorino F, Luciano P, Frecentese F, Magli E, et al. 1,2,4-Thiadiazolidin-3,5-diones as novel hydrogen sulfide donors. Eur J Med Chem 2018;143:1677-86. doi: <u>https://doi.org/10.1016/i.eimech.2017.10.068</u>.
- [36] Citi V, Corvino A, Fiorino F, Frecentese F, Magli E, Perissutti E, et al. Structureactivity relationships study of isothiocyanates for H2S releasing properties: 3-Pyridyl-isothiocyanate as a new promising cardioprotective agent. J of Adv Res 2020. doi: <u>https://doi.org/10.1016/i.jare.2020.02.017</u>.
- [37] Huang P, Chen S, Wang Y, Liu J, Yao Q, Huang Y, et al. Down-regulated CBS/ H2S pathway is involved in high-salt-induced hypertension in Dahl rats. Nitric Oxide 2015;46:192–203. doi: <u>https://doi.org/10.1016/j.niox.2015.01.004</u>.
- [38] Emmanuele di Villa Bianca R, Mitidieri E, Donnarumma E, Tramontano T, Brancaleone V, Cirino G, et al. Hydrogen sulfide is involved in dexamethasone-induced hypertension in rat. Nitric Oxide. 46 2015:80–6. doi: <u>https://doi.org/10.1016/j.niox.2014.11.013</u>.
- [39] Sun NL, Xi Y, Yang SN, Ma Z, Tang CS. Plasma hydrogen sulfide and homocysteine levels in hypertensive patients with different blood pressure levels and complications. Zhonghua Xin Xue Guan Bing Za Zhi 2007;35:1145–8.
- [40] Whiteman M, Gooding KM, Whatmore JL, Ball CI, Mawson D, Skinner K, et al. Adiposity is a major determinant of plasma levels of the novel vasodilator hydrogen sulphide. Diabetologia 2010;53:1722–6. doi: <u>https://doi.org/ 10.1007/s00125-010-1761-5</u>.
- [41] Kutz JL, Greaney JL, Santhanam L, Alexander LM. Evidence for a functional vasodilatatory role for hydrogen sulphide in the human cutaneous microvasculature. J Physiol 2015;593:2121–9. doi: <u>https://doi.org/10.1113/ IP270054</u>.
- [42] Ahmad A, Sattar MA, Rathore HA, Khan SA, Lazhari MI, Afzal S, et al. A critical review of pharmacological significance of Hydrogen Sulfide in hypertension. Indian J Pharmacol 2015;47:243–7. doi: <u>https://doi.org/10.4103/0253-7613.157106</u>.
- [43] Xiao L, Dong JH, Teng X, Jin S, Xue HM, Liu SY, et al. Hydrogen sulfide improves endothelial dysfunction in hypertension by activating peroxisome proliferator-activated receptor delta/endothelial nitric oxide synthase

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signaling. J Hypertens 2018;36:651–65. doi: <u>https://doi.org/10.1097/</u> <u>HIH.000000000001605</u>.

- [44] Wang R. Two's company, three's a crowd: can H2S be the third endogenous gaseous transmitter?. FASEB J 2002;16:1792–8. doi: <u>https://doi.org/10.1096/ fi.02-0211hyp</u>.
- [45] Zhao W, Ndisang JF, Wang R. Modulation of endogenous production of H2S in rat tissues. Can J Physiol Pharmacol 2003;81:848–53. doi: <u>https://doi.org/ 10.1139/y03-077</u>.
- [46] Zhao W, Wang R. H(2)S-induced vasorelaxation and underlying cellular and molecular mechanisms. Am J Physiol Heart Circ Physiol 2002;283:H474–480. doi: <u>https://doi.org/10.1152/ajpheart.00013.2002</u>.
- [47] Hosoki R, Matsuki N, Kimura H. The possible role of hydrogen sulfide as an endogenous smooth muscle relaxant in synergy with nitric oxide. Biochem Biophys Res Commun 1997;237:527–31. doi: <u>https://doi.org/10.1006/ bbrc.1997.6878</u>.
- [48] Altaany Z, Ju Y, Yang G, Wang R. The coordination of S-sulfhydration, Snitrosylation, and phosphorylation of endothelial nitric oxide synthase by hydrogen sulfide. Sci Signal 7 2014:ra87. doi: <u>https://doi.org/10.1126/ scisignal.2005478</u>.
- [49] Meng J, Ganesan Adaikan P, Srilatha B. Hydrogen sulfide promotes nitric oxide production in corpus cavernosum by enhancing expression of endothelial nitric oxide synthase. Int J Impot Res 25 2013:86–90. doi: <u>https://doi.org/10.1038/ijir.2012.39</u>.
- [50] Zhong G, Chen F, Cheng Y, Tang C, Du J. The role of hydrogen sulfide generation in the pathogenesis of hypertension in rats induced by inhibition of nitric oxide synthase. J Hypertens 2003;21:1879–85. doi: <u>https://doi.org/ 10.1097/00004872-200310000-00015</u>.
- [51] Cacanyiova S, Berenyiova A, Kristek F. The role of hydrogen sulphide in blood pressure regulation. Physiol Res 2016;65:S273–89.
- [52] Testai L, D'Antongiovanni V, Piano I, Martelli A, Citi V, Duranti E, et al. Different patterns of H2S/NO activity and cross-talk in the control of the coronary vascular bed under normotensive or hypertensive conditions. Nitric Oxide 2015;47:25–33. doi: <u>https://doi.org/10.1016/j.niox.2015.03.003</u>.
- [53] Yetik-Anacak G, Dikmen A, Coletta C, Mitidieri E, Dereli M, Donnarumma E, et al. Hydrogen sulfide compensates nitric oxide deficiency in murine corpus cavernosum. Pharmacol Res 2016;113:38–43. doi: <u>https://doi.org/10.1016/j. phrs.2016.08.015</u>.
- [54] Geng B, Cui Y, Zhao J, Yu F, Zhu Y, Xu G, et al. Hydrogen sulfide downregulates the aortic L-arginine/nitric oxide pathway in rats. Am J Physiol Regul Integr Comp Physiol 2007;293:R1608–1618. doi: <u>https://doi.org/10.1152/ aipregu.00207.2006</u>.
- [55] Chai Q, Lu T, Wang XL, Lee HC. Hydrogen sulfide impairs shear stress-induced vasodilation in mouse coronary arteries. Pflugers Arch 2015;467:329–40. doi: <u>https://doi.org/10.1007/s00424-014-1526-y</u>.
- [56] Tian XY, Wong WT, Sayed N, Luo J, Tsang SY, Bian ZX, et al. NaHS relaxes rat cerebral artery in vitro via inhibition of l-type voltage-sensitive Ca2+ channel. Pharmacol Res 2012;65:239–46. doi: <u>https://doi.org/10.1016/j.phrs.2011.11.006</u>.
- [57] Al-Magableh MR, Kemp-Harper BK, Hart JL. Hydrogen sulfide treatment reduces blood pressure and oxidative stress in angiotensin II-induced hypertensive mice. Hypertens Res 2015;38:13–20. doi: <u>https://doi.org/ 10.1038/hr.2014.125</u>.
- [58] Li J, Teng X, Jin S, Dong J, Guo Q, Tian D, et al. Hydrogen sulfide improves endothelial dysfunction by inhibiting the vicious cycle of NLRP3 inflammasome and oxidative stress in spontaneously hypertensive rats. J Hypertens 2019;37:1633–43. doi: <u>https://doi.org/10.1097/</u> HIH.00000000002101.
- [59] Sharma DK, Manral A, Saini V, Singh A, Srinivasan BP, Tiwari M. Novel diallyldisulfide analogs ameliorate cardiovascular remodeling in rats with L-NAME-induced hypertension. Eur J Pharmacol 2012;691:198–208. doi: https://doi.org/10.1016/j.ejphar.2012.07.022.
- [60] Li L, Whiteman M, Guan YY, Neo KL, Cheng Y, Lee SW, et al. Characterization of a novel, water-soluble hydrogen sulfide-releasing molecule (GYY4137): new insights into the biology of hydrogen sulfide. Circulation 2008;117:2351–60. doi: <u>https://doi.org/10.1161/</u> <u>CIRCULATIONAHA.107.753467.</u>
- [61] Wu L, Juurlink BH. The impaired glutathione system and its up-regulation by sulforaphane in vascular smooth muscle cells from spontaneously hypertensive rats. J Hypertens 2001;19:1819–25. doi: <u>https://doi.org/ 10.1097/00004872-200110000-00016</u>.
- [62] Senanayake GV, Banigesh A, Wu L, Lee P, Juurlink BH. The dietary phase 2 protein inducer sulforaphane can normalize the kidney epigenome and improve blood pressure in hypertensive rats. Am J Hypertens 2012;25:229–35. doi: <u>https://doi.org/10.1038/ajh.2011.200</u>.
- [63] Bucci M, Vellecco V, Cantalupo A, Brancaleone V, Zhou Z, Evangelista S, et al. Hydrogen sulfide accounts for the peripheral vascular effects of zofenopril independently of ACE inhibition. Cardiovasc Res 2014;102:138–47. doi: https://doi.org/10.1093/cvr/cvu026.
- [64] Tomasova L, Pavlovicova M, Malekova L, Misak A, Kristek F, Grman M, et al. Effects of AP39, a novel triphenylphosphonium derivatised anethole dithiolethione hydrogen sulfide donor, on rat haemodynamic parameters and chloride and calcium Cav3 and RyR2 channels. Nitric Oxide 2015;46:131–44. doi: https://doi.org/10.1016/j.niox.2014.12.012.
- [65] Libby P, Bornfeldt KE, Tall AR. Atherosclerosis: successes, surprises, and future challenges. Circ Res 2016;118:531–4. doi: <u>https://doi.org/10.1161/</u> <u>CIRCRESAHA.116.308334</u>.

- [66] Libby P. Inflammation in atherosclerosis. Nature 2002;420:868–74. doi: https://doi.org/10.1038/nature01323.
- [67] Martelli A, Testai L, Breschi MC, Blandizzi C, Virdis A, Taddei S, et al. Hydrogen sulphide: novel opportunity for drug discovery. Med Res Rev 2012;32:1093–130. doi: <u>https://doi.org/10.1002/med.20234</u>.
- [68] Monti M, Terzuoli E, Ziche M, Morbidelli L. H2S dependent and independent anti-inflammatory activity of zofenoprilat in cells of the vascular wall. Pharmacol Res 2016;113:426–37. doi: <u>https://doi.org/10.1016/j.phrs.2016.09.017</u>.
- [69] Wojcicka G, Jamroz-Wisniewska A, Atanasova P, Chaldakov GN, Chylinska-Kula B, Beltowski J. Differential effects of statins on endogenous H2S formation in perivascular adipose tissue. Pharmacol Res 2011;63:68–76. doi: <u>https://doi.org/10.1016/j.phrs.2010.10.011</u>.
- [70] Xu Y, Du HP, Li J, Xu R, Wang YL, You SJ, et al. Statins upregulate cystathionine gamma-lyase transcription and H2S generation via activating Akt signaling in macrophage. Pharmacol Res 2014;87:18–25. doi: <u>https://doi.org/10.1016/j. phrs.2014.06.006</u>.
- [71] Wang Y, Zhao X, Jin H, Wei H, Li W, Bu D, et al. Role of hydrogen sulfide in the development of atherosclerotic lesions in apolipoprotein E knockout mice. Arterioscler Thromb Vasc Biol 2009;29:173–9. doi: <u>https://doi.org/10.1161/ ATVBAHA.108.179333</u>.
- [72] Ling K, Xu A, Chen Y, Chen X, Li Y, Wang W. Protective effect of a hydrogen sulfide donor on balloon injury-induced restenosis via the Nrf2/HIF-1alpha signaling pathway. Int J Mol Med 2019;43:1299–310. doi: <u>https://doi.org/ 10.3892/ijmm.2019.4076</u>.
- [73] Qiao W, Chaoshu T, Hongfang J, Junbao D. Endogenous hydrogen sulfide is involved in the pathogenesis of atherosclerosis. Biochem Biophys Res Commun 2010;396:182–6. doi: <u>https://doi.org/10.1016/j.bbrc.2010.04.061</u>.
- [74] Wu SY, Pan CS, Geng B, Zhao J, Yu F, Pang YZ, et al. Hydrogen sulfide ameliorates vascular calcification induced by vitamin D3 plus nicotine in rats. Acta Pharmacol Sin 2006;27:299–306. doi: <u>https://doi.org/10.1111/j.1745-7254.2006.00283.x</u>.
- [75] Zhang H, Guo C, Wu D, Zhang A, Gu T, Wang L, et al. Hydrogen sulfide inhibits the development of atherosclerosis with suppressing CX3CR1 and CX3CL1 expression. PLoS ONE 2012;7:. doi: <u>https://doi.org/10.1371/journal.pone.0041147</u>e41147.
- [76] Xiong Q, Wang Z, Yu Y, Wen Y, Suguro R, Mao Y, et al. Hydrogen sulfide stabilizes atherosclerotic plaques in apolipoprotein E knockout mice. Pharmacol Res 2019;144:90–8. doi: <u>https://doi.org/10.1016/j. phrs.2019.04.006.</u>
- [77] Liu Z, Han Y, Li L, Lu H, Meng G, Li X, et al. The hydrogen sulfide donor, GYY4137, exhibits anti-atherosclerotic activity in high fat fed apolipoprotein E(-/-) mice. Br J Pharmacol 2013;169:1795–809. doi: <u>https://doi.org/10.1111/ bph.12246</u>.
- [78] Sestito S, Pruccoli L, Runfola M, Citi V, Martelli A, Saccomanni G, et al. Design and synthesis of H2S-donor hybrids: a new treatment for Alzheimer's disease?. Eur J Med Chem 2019;184:. doi: <u>https://doi.org/10.1016/j. eimech.2019.111745</u>111745.
- [79] Ercolano G, De Cicco P, Frecentese F, Saccone I, Corvino A, Giordano F, et al. Anti-metastatic properties of naproxen-HBTA in a murine model of cutaneous melanoma. Front Pharmacol 2019;10:66. doi: <u>https://doi.org/ 10.3389/fphar.2019.00066</u>.
- [80] Sestito S, Daniele S, Pietrobono D, Citi V, Bellusci L, Chiellini G, et al. Memantine prodrug as a new agent for Alzheimer's Disease. Sci Rep 2019;9:4612. doi: <u>https://doi.org/10.1038/s41598-019-40925-8</u>.
- [81] Rapposelli S, Gambari L, Digiacomo M, Citi V, Lisignoli G, Manferdini C, et al. A Novel H2S-releasing Amino-Bisphosphonate which combines bone anticatabolic and anabolic functions. Sci Rep 2017;7:11940. doi: <u>https://doi.org/ 10.1038/s41598-017-11608-z</u>.
- [82] Martelli A, Rapposelli S, Calderone V. NO-releasing hybrids of cardiovascular drugs. Curr Med Chem 2006;13:609–25. doi: <u>https://doi.org/10.2174/</u>092986706776055634.
- [83] Calderone V, Martelli A, Testai L, Citi V, Breschi MC. Using hydrogen sulfide to design and develop drugs. Expert Opin Drug Discov 2016;11:163–75. doi: <u>https://doi.org/10.1517/17460441.2016.1122590</u>.
- [84] Martelli A, Testai L, Anzini M, Cappelli A, Di Capua A, Biava M, et al. The novel anti-inflammatory agent VA694, endowed with both NO-releasing and COX2selective inhibiting properties, exhibits NO-mediated positive effects on blood pressure, coronary flow and endothelium in an experimental model of hypertension and endothelial dysfunction. Pharmacol Res 2013;78:1–9. doi: https://doi.org/10.1016/j.phrs.2013.09.008.
- [85] Anzini M, Di Capua A, Valenti S, Brogi S, Rovini M, Giuliani G, et al. Novel analgesic/anti-inflammatory agents: 1,5-diarylpyrrole nitrooxyalkyl ethers and related compounds as cyclooxygenase-2 inhibiting nitric oxide donors. J Med Chem 2013;56:3191–206. doi: <u>https://doi.org/10.1021/jm301370e</u>.
- [86] Zhang H, Guo C, Zhang A, Fan Y, Gu T, Wu D, et al. Effect of S-aspirin, a novel hydrogen-sulfide-releasing aspirin (ACS14), on atherosclerosis in apoEdeficient mice. Eur J Pharmacol 2012;697:106–16. doi: <u>https://doi.org/ 10.1016/j.eiphar.2012.10.005</u>.
- [87] Lei YP, Chen HW, Sheen LY, Lii CK. Diallyl disulfide and diallyl trisulfide suppress oxidized LDL-induced vascular cell adhesion molecule and Eselectin expression through protein kinase A- and B-dependent signaling pathways. J Nutr 2008;138:996–1003. doi: <u>https://doi.org/10.1093/jn/ 138.6.996</u>.
- [88] Lei YP, Liu CT, Sheen LY, Chen HW, Lii CK. Diallyl disulfide and diallyl trisulfide protect endothelial nitric oxide synthase against damage by

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oxidized low-density lipoprotein. Mol Nutr Food Res 2010;54(Suppl 1): S42–52. doi: https://doi.org/10.1002/mnfr.200900278.

- [89] Citi V, Martelli A, Testai L, Marino A, Breschi MC, Calderone V. Hydrogen sulfide releasing capacity of natural isothiocyanates: is it a reliable explanation for the multiple biological effects of Brassicaceae?. Planta Med 2014;80:610-3. doi: <u>https://doi.org/10.1055/s-0034-1368591</u>.
- [90] Citi V, Piragine E, Pagnotta E, Ugolini L, Di Cesare Mannelli L, Testai L, et al. Anticancer properties of erucin, an H2 S-releasing isothiocyanate, on human pancreatic adenocarcinoma cells (AsPC-1). Phytother Res 2019;33:845–55. doi: <u>https://doi.org/10.1002/ptr.6278</u>.
- [91] Martelli A, Citi V, Testai L, Brogi S, Calderone V. Organic isothiocyanates as hydrogen sulfide donors. Antioxid Redox Signal 2020;32:110–44. doi: <u>https:// doi.org/10.1089/ars.2019.7888</u>.
- [92] Matsui T, Nakamura N, Ojima A, Nishino Y, Yamagishi SI. Sulforaphane reduces advanced glycation end products (AGEs)-induced inflammation in endothelial cells and rat aorta. Nutr Metab Cardiovasc Dis 2016;26:797–807. doi: <u>https://doi.org/10.1016/j.numecd.2016.04.008</u>.
- [93] Shehatou GS, Suddek GM. Sulforaphane attenuates the development of atherosclerosis and improves endothelial dysfunction in hypercholesterolemic rabbits. Exp Biol Med (Maywood) 2016;241:426–36. doi: <u>https://doi.org/10.1177/1535370215609695</u>.
- [94] Bibli SI, Hu J, Sigala F, Wittig I, Heidler J, Zukunft S, et al. Cystathionine gamma lyase sulfhydrates the RNA binding protein human antigen R to preserve endothelial cell function and delay atherogenesis. Circulation 2019;139:101-14. doi: <u>https://doi.org/10.1161/</u> <u>CIRCULATIONAHA.118.034757.</u>
- [95] Avogaro A, Albiero M, Menegazzo L, de Kreutzenberg S, Fadini GP. Endothelial dysfunction in diabetes: the role of reparatory mechanisms. Diabetes Care 2011;34(Suppl 2):S285–290. doi: <u>https://doi.org/10.2337/dc11-s239</u>.
- [96] Avogaro A, Fadini GP, Gallo A, Pagnin E, de Kreutzenberg S. Endothelial dysfunction in type 2 diabetes mellitus. Nutr Metab Cardiovasc Dis 2006;16 (Suppl 1):S39-45. doi: <u>https://doi.org/10.1016/j.numecd.2005.10.015</u>.
- [97] Fadini GP, Sartore S, Agostini C, Avogaro A. Significance of endothelial progenitor cells in subjects with diabetes. Diabetes Care 2007;30:1305–13. doi: <u>https://doi.org/10.2337/dc06-2305</u>.
- [98] van den Born Joost C, Hammes Hans-Peter, Greffrath Wolfgang, van Goor Harry, Hillebrands Jan-Luuk. Gasotransmitters in vascular complications of diabetes. Diabetes 2016;65(2):331–45. doi: <u>https://doi.org/10.2337/db15-1003</u>.
- [99] Durante W. Hydrogen sulfide therapy in diabetes-accelerated atherosclerosis: a whiff of success. Diabetes 2016;65:2832–4. doi: <u>https:// doi.org/10.2337/dbi16-0042</u>.
- [100] Kolluru GK, Shen X, Bir SC, Kevil CG. Hydrogen sulfide chemical biology: pathophysiological roles and detection. Nitric Oxide 2013;35:5–20. doi: https://doi.org/10.1016/j.niox.2013.07.002.
- [101] Polhemus DJ, Lefer DJ. Emergence of hydrogen sulfide as an endogenous gaseous signaling molecule in cardiovascular disease. Circ Res 2014;114:730-7. doi: <u>https://doi.org/10.1161/CIRCRESAHA.114.300505</u>.
- [102] Mustafa AK, Gadalla MM, Sen N, Kim S, Mu W, Gazi SK, et al. H2S signals through protein S-sulfhydration. Sci Signal 2009;2:ra72. doi: <u>https://doi.org/ 10.1126/scisignal.2000464</u>.
- [103] Suzuki K, Olah G, Modis K, Coletta C, Kulp G, Gero D, et al. Hydrogen sulfide replacement therapy protects the vascular endothelium in hyperglycemia by preserving mitochondrial function. Proc Natl Acad Sci U S A 2011;108:13829–34. doi: <u>https://doi.org/10.1073/pnas.1105121108</u>.
- [104] Zhou X, Feng Y, Zhan Z, Chen J. Hydrogen sulfide alleviates diabetic nephropathy in a streptozotocin-induced diabetic rat model. J Biol Chem 2014;289:28827–34. doi: <u>https://doi.org/10.1074/jbc.M114.596593</u>.
- [105] Si YF, Wang J, Guan J, Zhou L, Sheng Y, Zhao J. Treatment with hydrogen sulfide alleviates streptozotocin-induced diabetic retinopathy in rats. Br J Pharmacol 2013;169:619–31. doi: <u>https://doi.org/10.1111/bph.12163</u>.
- [106] Zhou X, An G, Lu X. Hydrogen sulfide attenuates the development of diabetic cardiomyopathy. Clin Sci (Lond) 2015;128:325–35. doi: <u>https://doi.org/ 10.1042/CS20140460</u>.
- [107] Xiao Q, Ying J, Xiang L, Zhang C. The biologic effect of hydrogen sulfide and its function in various diseases. Medicine (Baltimore) 2018;97:. doi: <u>https://doi.org/10.1097/MD.00000000013065</u>e13065.
- [108] Xie L, Gu Y, Wen M, Zhao S, Wang W, Ma Y, et al. Hydrogen sulfide induces keap1 S-sulfhydration and suppresses diabetes-accelerated atherosclerosis via Nrf2 activation. Diabetes 2016;65:3171–84. doi: <u>https://doi.org/10.2337/ db16-0020</u>.
- [109] Szabo C. Roles of hydrogen sulfide in the pathogenesis of diabetes mellitus and its complications. Antioxid Redox Signal 2012;17:68–80. doi: <u>https://doi.org/10.1089/ars.2011.4451</u>.
- [110] Jain SK, Bull R, Rains JL, Bass PF, Levine SN, Reddy S, et al. Low levels of hydrogen sulfide in the blood of diabetes patients and streptozotocin-treated rats causes vascular inflammation?. Antioxid Redox Signal 2010;12:1333–7. doi: <u>https://doi.org/10.1089/ars.2009.2956</u>.
- [111] Gero D, Torregrossa R, Perry A, Waters A, Le-Trionnaire S, Whatmore JL, et al. The novel mitochondria-targeted hydrogen sulfide (H2S) donors AP123 and AP39 protect against hyperglycemic injury in microvascular endothelial cells in vitro. Pharmacol Res 2016;113:186–98. doi: <u>https://doi.org/10.1016/j. phrs.2016.08.019</u>.
- [112] Ng HH, Yildiz GS, Ku JM, Miller AA, Woodman OL, Hart JL. Chronic NaHS treatment decreases oxidative stress and improves endothelial function in

diabetic mice. Diab Vasc Dis Res 2017;14:246–53. doi: <u>https://doi.org/</u> 10.1177/1479164117692766.

- [113] Ma S, Zhong D, Ma P, Li G, Hua W, Sun Y, et al. Exogenous hydrogen sulfide ameliorates diabetes-associated cognitive decline by regulating the mitochondria-mediated apoptotic pathway and IL-23/IL-17 expression in db/db mice. Cell Physiol Biochem 2017;41:1838–50. doi: <u>https://doi.org/ 10.1159/000471932</u>.
- [114] Melino S, Leo S, Toska-Papajani V. Natural hydrogen sulfide donors from Allium sp. as a nutraceutical approach in type 2 diabetes prevention and therapy. Nutrients 2019;11. doi: <u>https://doi.org/10.3390/nu11071581</u>.
- [115] Padiya R, Khatua TN, Bagul PK, Kuncha M, Banerjee SK. Garlic improves insulin sensitivity and associated metabolic syndromes in fructose fed rats. Nutr Metab (Lond) 2011;8:53. doi: <u>https://doi.org/10.1186/1743-7075-8-53</u>.
- [116] Baluchnejadmojarad T, Kiasalari Z, Afshin-Majd S, Ghasemi Z, Roghani M. Sallyl cysteine ameliorates cognitive deficits in streptozotocin-diabetic rats via suppression of oxidative stress, inflammation, and acetylcholinesterase. Eur J Pharmacol 2017;794:69–76. doi: <u>https://doi.org/10.1016/j. eiphar.2016.11.033</u>.
- [117] Bahadoran Z, Mirmiran P, Hosseinpanah F, Rajab A, Asghari G, Azizi F. Broccoli sprouts powder could improve serum triglyceride and oxidized LDL/ LDL-cholesterol ratio in type 2 diabetic patients: a randomized double-blind placebo-controlled clinical trial. Diabetes Res Clin Pract 2012;96:348–54. doi: https://doi.org/10.1016/j.diabres.2012.01.009.
- [118] Bahadoran Z, Tohidi M, Nazeri P, Mehran M, Azizi F, Mirmiran P. Effect of broccoli sprouts on insulin resistance in type 2 diabetic patients: a randomized double-blind clinical trial. Int J Food Sci Nutr 2012;63:767–71. doi: <u>https://doi.org/10.3109/09637486.2012.665043</u>.
- [119] Axelsson AS, Tubbs E, Mecham B, Chacko S, Nenonen HA, Tang Y, et al. Sulforaphane reduces hepatic glucose production and improves glucose control in patients with type 2 diabetes. Sci Transl Med 2017;9. doi: <u>https:// doi.org/10.1126/scitranslmed.aah4477</u>.
- [120] Bai Y, Wang X, Zhao S, Ma C, Cui J, Zheng Y. Sulforaphane protects against cardiovascular disease via Nrf2 activation. Oxid Med Cell Longev 2015;2015:. doi: <u>https://doi.org/10.1155/2015/407580</u>407580.
- [121] Leoncini E, Malaguti M, Angeloni C, Motori E, Fabbri D, Hrelia S. Cruciferous vegetable phytochemical sulforaphane affects phase II enzyme expression and activity in rat cardiomyocytes through modulation of Akt signaling pathway. J Food Sci 2011;76:H175–181. doi: <u>https://doi.org/10.1111/j.1750-3841.2011.02311.x</u>.
- [122] Pereira A, Fernandes R, Crisostomo J, Seica RM, Sena CM. The Sulforaphane and pyridoxamine supplementation normalize endothelial dysfunction associated with type 2 diabetes. Sci Rep 2017;7:14357. doi: <u>https://doi.org/ 10.1038/s41598-017-14733-x</u>.
- [123] Brancaleone V, Roviezzo F, Vellecco V, De Gruttola L, Bucci M, Cirino G. Biosynthesis of H2S is impaired in non-obese diabetic (NOD) mice. Br J Pharmacol 2008;155:673–80. doi: <u>https://doi.org/10.1038/bip.2008.296</u>.
- [124] Du J, Huang Y, Yan H, Zhang Q, Zhao M, Zhu M, et al. Hydrogen sulfide suppresses oxidized low-density lipoprotein (ox-LDL)-stimulated monocyte chemoattractant protein 1 generation from macrophages via the nuclear factor kappaB (NF-kappaB) pathway. J Biol Chem 2014;289:9741–53. doi: https://doi.org/10.1074/jbc.M113.517995.
- [125] Kumar A, Palfrey HA, Pathak R, Kadowitz PJ, Gettys TW, Murthy SN. The metabolism and significance of homocysteine in nutrition and health. Nutr Metab (Lond) 2017;14:78. doi: <u>https://doi.org/10.1186/s12986-017-0233-z</u>.
- [126] Long Y, Nie J. Homocysteine in renal injury. Kidney Dis (Basel) 2016;2:80–7. doi: <u>https://doi.org/10.1159/000444900</u>.
- [127] Kerkeni M, Tnani M, Chuniaud L, Miled A, Maaroufi K, Trivin F. Comparative study on in vitro effects of homocysteine thiolactone and homocysteine on HUVEC cells: evidence for a stronger proapoptotic and proinflammative homocysteine thiolactone. Mol Cell Biochem 2006;291:119–26. doi: <u>https:// doi.org/10.1007/s11010-006-9204-9</u>.
- [128] Ciaccio M, Bivona G, Bellia C. Therapeutical approach to plasma homocysteine and cardiovascular risk reduction. Ther Clin Risk Manag 2008;4:219–24. doi: <u>https://doi.org/10.2147/tcrm.s1807</u>.
 [129] Ganguly P, Alam SF. Role of homocysteine in the development of
- [129] Ganguly P, Alam SF. Role of homocysteine in the development of cardiovascular disease. Nutr J 2015;14:6. doi: <u>https://doi.org/10.1186/1475-2891-14-6</u>.
- [130] Zhang K, Kaufman RJ. From endoplasmic-reticulum stress to the inflammatory response. Nature 2008;454:455–62. doi: <u>https://doi.org/</u> <u>10.1038/nature07203</u>.
- [131] Chrysant SG, Chrysant GS. The current status of homocysteine as a risk factor for cardiovascular disease: a mini review. Expert Rev Cardiovasc Ther 2018;16:559–65. doi: <u>https://doi.org/10.1080/14779072.2018.1497974</u>.
- [132] Girelli D, Martinelli N, Olivieri O, Pizzolo F, Friso S, Faccini G, et al. Hyperhomocysteinemia and mortality after coronary artery bypass grafting. PLoS ONE 2006;1:. doi: <u>https://doi.org/10.1371/journal.pone.0000083</u>e83.
- [133] Weiss N. Mechanisms of increased vascular oxidant stress in hyperhomocysteinemia and its impact on endothelial function. Curr Drug Metab 2005;6:27–36. doi: <u>https://doi.org/10.2174/1389200052997357</u>.
- [134] Liu HH, Shih TS, Huang HR, Huang SC, Lee LH, Huang YC. Plasma homocysteine is associated with increased oxidative stress and antioxidant enzyme activity in welders. Sci World J 2013;2013:. doi: <u>https://doi.org/ 10.1155/2013/370487</u>370487.
- [135] Mani M, Golmohammadi T, Khaghani S, Zamani Z, Azadmanesh K, Meshkani R, et al. Homocysteine induces heme oxygenase-1 expression via

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transcription factor Nrf2 activation in HepG2 cell. Iran Biomed J 2013;17:93–100. doi: https://doi.org/10.6091/ibj.1158.2013.

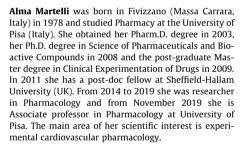
- [136] Handy DE, Zhang Y, Loscalzo J. Homocysteine down-regulates cellular glutathione peroxidase (GPx1) by decreasing translation. J Biol Chem 2005;280:15518–25. doi: <u>https://doi.org/10.1074/jbc.M501452200</u>.
- [137] Wang G, Siow YL, O K. Homocysteine stimulates nuclear factor kappaB activity and monocyte chemoattractant protein-1 expression in vascular smooth-muscle cells: a possible role for protein kinase C. Biochem J 2000;352 (Pt 3):817–26.
- [138] Guo X, Dudman NP. Homocysteine induces expressions of adhesive molecules on leukocytes in whole blood. Chin Med J (Engl) 2001;114:1235–9.
- [139] Bonnans C, Chou J, Werb Z. Remodelling the extracellular matrix in development and disease. Nat Rev Mol Cell Biol 2014;15:786–801. doi: https://doi.org/10.1038/nrm3904.
- [140] Gerdes VE, Hovinga HA, ten Cate H, Macgillavry MR, Leijte A, Reitsma PH, et al. Homocysteine and markers of coagulation and endothelial cell activation. J Thromb Haemost 2004;2:445–51. doi: <u>https://doi.org/10.1111/ i.1538-7836.2004.00674.x</u>.
- [141] Sule AA, Chin TJ, Khien LH. Recurrent unprovoked venous thromboembolism in a young female patient with high levels of homocysteine. Int J Angiol 2012;21:95–8. doi: <u>https://doi.org/10.1055/s-0032-1315626</u>.
- [142] Ago T, Kitazono T, Kuroda J, Kumai Y, Kamouchi M, Ooboshi H, et al. NAD(P)H oxidases in rat basilar arterial endothelial cells. Stroke 2005;36:1040–6. doi: https://doi.org/10.1161/01.STR.0000163111.05825.0b.
- [143] Hofmann MA, Lalla E, Lu Y, Gleason MR, Wolf BM, Tanji N, et al. Hyperhomocysteinemia enhances vascular inflammation and accelerates atherosclerosis in a murine model. J Clin Invest 2001;107:675–83. doi: https://doi.org/10.1172/JCI10588.
- [144] Olas B, Kontek B. Hydrogen sulfide decreases the plasma lipid peroxidation induced by homocysteine and its thiolactone. Mol Cell Biochem 2015;404:39–43. doi: <u>https://doi.org/10.1007/s11010-015-2364-8</u>.
- [145] Stanger O, Weger M. Interactions of homocysteine, nitric oxide, folate and radicals in the progressively damaged endothelium. Clin Chem Lab Med 2003;41:1444–54. doi: <u>https://doi.org/10.1515/CCLM.2003.222</u>.
- [146] Pushpakumar S, Kundu S, Sen U. Endothelial dysfunction: the link between homocysteine and hydrogen sulfide. Curr Med Chem 2014;21:3662–72. doi: https://doi.org/10.2174/0929867321666140706142335.
- [147] Tyagi N, Sedoris KC, Steed M, Ovechkin AV, Moshal KS, Tyagi SC. Mechanisms of homocysteine-induced oxidative stress. Am J Physiol Heart Circ Physiol 2005;289:H2649-2656. doi: <u>https://doi.org/10.1152/ajpheart.00548.2005</u>.
- [148] Zang T, Pottenplackel LP, Handy DE, Loscalzo J, Dai S, Deth RC, et al. Comparison of protein N-homocysteinylation in rat plasma under elevated homocysteine using a specific chemical labeling method. Molecules 2016;21. doi: https://doi.org/10.3390/molecules21091195.
- [149] Cheng X. Updating the relationship between hyperhomocysteinemia lowering therapy and cardiovascular events. Cardiovasc Ther 2013;31: e19–26. doi: <u>https://doi.org/10.1111/1755-5922.12014</u>.
- [150] Li MH, Tang JP, Zhang P, Li X, Wang CY, Wei HJ, et al. Disturbance of endogenous hydrogen sulfide generation and endoplasmic reticulum stress in hippocampus are involved in homocysteine-induced defect in learning and memory of rats. Behav Brain Res 2014;262:35–41. doi: <u>https://doi.org/ 10.1016/j.bbr.2014.01.001</u>.
- [151] Kamat PK, Kalani A, Givvimani S, Sathnur PB, Tyagi SC, Tyagi N. Hydrogen sulfide attenuates neurodegeneration and neurovascular dysfunction induced by intracerebral-administered homocysteine in mice. Neuroscience 2013;252:302–19. doi: <u>https://doi.org/10.1016/j.neuroscience.2013.07.051</u>.
- [152] Nath N, Prasad HK, Kumar M. Cerebroprotective effects of hydrogen sulfide in homocysteine-induced neurovascular permeability: involvement of oxidative stress, arginase, and matrix metalloproteinase-9. J Cell Physiol 2019;234:3007–19. doi: <u>https://doi.org/10.1002/jcp.27120</u>.
- [153] Kimura Y, Kimura H. Hydrogen sulfide protects neurons from oxidative stress. FASEB J 2004;18:1165–7. doi: <u>https://doi.org/10.1096/fi.04-1815fie</u>.
 [154] Potor L, Nagy P, Mehes G, Hendrik Z, Jeney V, Petho D, et al. Hydrogen sulfide
- [154] Potor L, Nagy P, Mehes G, Hendrik Z, Jeney V, Petho D, et al. Hydrogen sulfide abrogates hemoglobin-lipid interaction in atherosclerotic lesion. Oxid Med Cell Longev 2018;2018:3812568. doi: <u>https://doi.org/10.1155/2018/ 3812568</u>.
- [155] Yan SK, Chang T, Wang H, Wu L, Wang R, Meng QH. Effects of hydrogen sulfide on homocysteine-induced oxidative stress in vascular smooth muscle cells. Biochem Biophys Res Commun 2006;351:485–91. doi: <u>https://doi.org/ 10.1016/j.bbrc.2006.10.058</u>.
- [156] Tyagi N, Moshal KS, Sen U, Vacek TP, Kumar M, Hughes Jr WM, et al. H2S protects against methionine-induced oxidative stress in brain endothelial cells. Antioxid Redox Signal 2009;11:25–33. doi: <u>https://doi.org/10.1089/ ars.2008.2073</u>.
- [157] Kamat PK, Kalani A, Tyagi SC, Tyagi N. Hydrogen sulfide epigenetically attenuates homocysteine-induced mitochondrial toxicity mediated through NMDA receptor in mouse brain endothelial (bEnd3) cells. J Cell Physiol 2015;230:378–94. doi: <u>https://doi.org/10.1002/jcp.24722</u>.

- [158] Li JJ, Li Q, Du HP, Wang YL, You SJ, Wang F, et al. Homocysteine triggers inflammatory responses in macrophages through inhibiting CSE-H2S signaling via DNA hypermethylation of CSE promoter. Int J Mol Sci 2015;16:12560–77. doi: <u>https://doi.org/10.3390/ijms160612560</u>.
- [159] Brancaleone V, Esposito I, Gargiulo A, Vellecco V, Asimakopoulou A, Citi V, et al. D-Penicillamine modulates hydrogen sulfide (H2S) pathway through selective inhibition of cystathionine-gamma-lyase. Br J Pharmacol 2016;173:1556–65. doi: <u>https://doi.org/10.1111/bph.13459</u>.
- [160] Moshal KS, Sen U, Tyagi N, Henderson B, Steed M, Ovechkin AV, et al. Regulation of homocysteine-induced MMP-9 by ERK1/2 pathway. Am J Physiol Cell Physiol 2006;290:C883–891. doi: <u>https://doi.org/10.1152/ aipcell.00359.2005</u>.
- [161] Bescond A, Augier T, Chareyre C, Garcon D, Hornebeck W, Charpiot P. Influence of homocysteine on matrix metalloproteinase-2: activation and activity. Biochem Biophys Res Commun 1999;263:498–503. doi: <u>https://doi.org/10.1006/bbrc.1999.1391</u>.
- [162] Nakano S, Ishii I, Shinmura K, Tamaki K, Hishiki T, Akahoshi N, et al. Hyperhomocysteinemia abrogates fasting-induced cardioprotection against ischemia/reperfusion by limiting bioavailability of hydrogen sulfide anions. J Mol Med (Berl) 2015;93:879–89. doi: <u>https://doi.org/10.1007/s00109-015-1271-5</u>.
- [163] Kar S, Shahshahan HR, Kambis TN, Yadav SK, Li Z, Lefer DJ, et al. Hydrogen Sulfide Ameliorates Homocysteine-Induced Cardiac Remodeling and Dysfunction. Front Physiol 2019;10:598. doi: <u>https://doi.org/10.3389/ fphys.2019.00598</u>.
- [164] Lucarini E, Micheli L, Trallori E, Citi V, Martelli A, Testai L, et al. Effect of glucoraphanin and sulforaphane against chemotherapy-induced neuropathic pain: Kv7 potassium channels modulation by H2 S release in vivo. Phytother Res 2018;32:2226–34. doi: <u>https://doi.org/10.1002/ptr.6159</u>.



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