

# CO Disrupts the Reduced H-Cluster of FeFe Hydrogenase. A Combined DFT and Protein Film Voltammetry Study

Carole Baffert,<sup>†</sup> Luca Bertini,<sup>‡</sup> Thomas Lautier,<sup>§</sup> Claudio Greco,<sup>‡</sup> Kateryna Sybirna,<sup>⊥</sup> Pierre Ezanno,<sup>†</sup> Emilien Etienne,<sup>†</sup> Philippe Soucaille,<sup>§</sup> Patrick Bertrand,<sup>†</sup> Hervé Bottin,<sup>⊥</sup> Isabelle Meynial-Salles,<sup>§</sup> Luca De Gioia,<sup>\*,‡</sup> and Christophe Léger<sup>\*,†</sup>

<sup>†</sup>Bioenergetics and Engineering of Proteins, CNRS, UPR 9036, 31 chemin Joseph Aiguier, 13402 Marseille Cedex 20, France

<sup>‡</sup>Department of Biotechnology and Biosciences, Università degli Studi di Milano-Bicocca, Milan 20126, Italy

<sup>§</sup>Université de Toulouse, INSA, UPS, INP, LISBP, INRA:UMR792,135 CNRS:UMR 5504, Avenue de Rangueil, 31077 Toulouse, France

<sup>⊥</sup>iBiTec-S, SB2SM, LPB (URA CNRS 2096), DSV, CEA, 91191 Gif-sur-Yvette, France

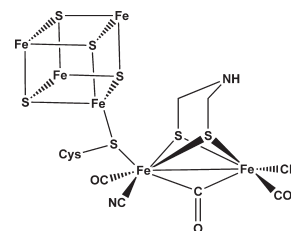
**S** Supporting Information

**ABSTRACT:** Carbon monoxide is often described as a competitive inhibitor of FeFe hydrogenases, and it is used for probing H<sub>2</sub> binding to synthetic or *in silico* models of the active site H-cluster. Yet it does not always behave as a simple inhibitor. Using an original approach which combines accurate electrochemical measurements and theoretical calculations, we elucidate the mechanism by which, under certain conditions, CO binding can cause permanent damage to the H-cluster. Like in the case of oxygen inhibition, the reaction with CO engages the entire H-cluster, rather than only the Fe<sub>2</sub> subsite.

Hydrogenases (H<sub>2</sub>ases), the enzymes that catalyze the biological production and oxidation of H<sub>2</sub>, are classified according to the metal content of their active site. In FeFe-H<sub>2</sub>ases, H<sub>2</sub> activation occurs at the H-cluster, which consists of a [Fe<sub>2</sub>(CO)<sub>3</sub>(CN)<sub>2</sub>(μ-dtma)] subsite (dtma = dithiomethylamine) covalently bound to a [4Fe4S] cluster (Scheme 1). During catalysis, the 2Fe subsite exists in the Fe<sup>II</sup>Fe<sup>I</sup> and Fe<sup>I</sup>Fe<sup>I</sup> states (Hox and Hred, respectively).<sup>1</sup> The cubane is reduced in the so-called “super-red” state (Hsred).<sup>2</sup> In theoretical and experimental studies of the H-cluster, CO is often considered as an innocent ligand which competes for hydrogen, although early experiments with the H<sub>2</sub>ase from *Clostridium pasteurianum* (Cp) showed that CO may also alter the enzyme in an irreversible manner.<sup>3</sup> Recent experiments clarified this by demonstrating that CO inhibition is fully reversible when the enzyme is oxidizing H<sub>2</sub>, not when it is producing H<sub>2</sub>.<sup>4</sup> Here we elucidate the nature of the irreversible damage induced by exogenous CO.

Using protein film voltammetry (PFV)<sup>5</sup> and methods described before,<sup>6,7</sup> we have quantitatively examined the kinetics of CO inhibition of the FeFe H<sub>2</sub>ases from *Clostridium acetobutylicum* (Ca) and *Chlamydomonas reinhardtii* (Cr). We found that inhibition proceeds by reversible formation of an inactive enzyme–CO complex, which irreversibly inactivates in a first-order irreversible process in the range of potential that favors the formation of Hred. This behavior is explained by DFT calculations:

**Scheme 1.** Active Site (H-Cluster) of FeFe Hydrogenase



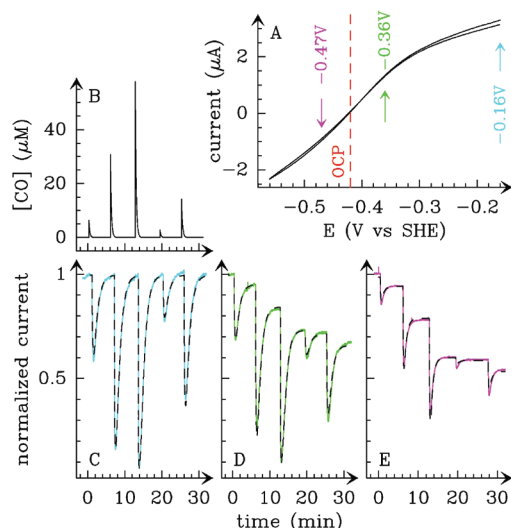
CO binding to Hred leads to cleavage of the bond between the Fe and the cysteine S that bridges the Fe<sub>2</sub> and [4Fe4S] subsites (Scheme 1). The CO-bound, disrupted 2Fe cluster is so stable that this process is irreversible. However, CO dissociation can occur if the Hred-CO state is taken to very low potential *before* the H-cluster has fallen apart, because the Hsred-CO species does not rupture and easily releases CO.

The procedures we used to purify the FeFe hydrogenases from Ca (hydA, Strep-tagged, and homologously expressed) and Cr (hydA1, Strep-tagged, and heterologously expressed in *Shewanella oneidensis*) have been described in refs 7b (and references therein) and 7c, respectively.

Panel A in Figure 1 shows a steady-state voltammogram recorded with Ca FeFe H<sub>2</sub>ase adsorbed at a rotating disk graphite electrode (see methods in Supporting Information (SI)). The open circuit potential (OCP) equates the reduction potential of the H<sup>+</sup>/H<sub>2</sub> couple and marks the limit between oxidative and reductive catalysis (H<sub>2</sub> oxidation versus production). The vertical arrows indicate the potentials that were used in the chronoamperometric experiments shown in panels C–E. In each case, the enzyme activity (the current) was monitored as a function of time, in the dark, while aliquots of a solution saturated with CO were repeatedly injected in the electrochemical cell. The change in CO concentration against time is shown in panel B. After each injection, CO is flushed away by the stream of H<sub>2</sub>, and its concentration decays quickly (typical time constant τ = 15 s),

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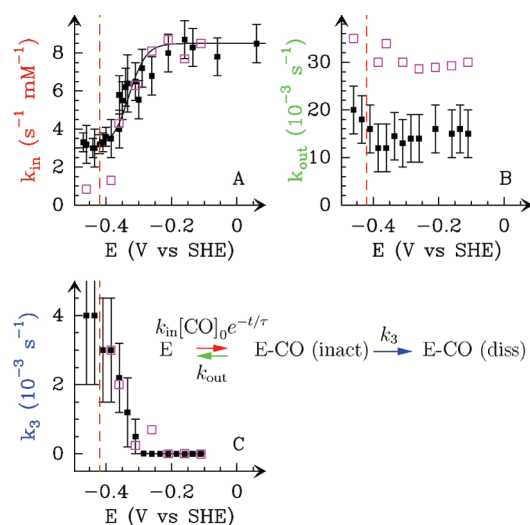


**Figure 1.** (A) Steady-state voltammogram for *Ca* FeFe H<sub>2</sub>ase, 1 atm of H<sub>2</sub>, pH 7, 30 °C, 20 mV/s, 3000 rpm. The open circuit potential (OCP) is indicated by a dashed red line. (B) CO concentration against time. (C–E) Normalized current traces showing the activity changes that result from the sequence of injections shown in panel B, recorded at  $E = -0.16$  V (C),  $-0.36$  V (D), and  $-0.47$  V (E), in the dark, 1 atm of H<sub>2</sub>, pH 7, 30 °C. See raw data in SI Figure S1. The dashed lines are the best fit to the model shown in Figure 2.<sup>6,7</sup> In these experiments, the concentration of inhibitor varies, and the activity is a complex function of time (even when  $k_3 = 0$ , see eq 2 in ref 9); the amplitude of the activity change increases with (but is not merely proportional to) the amount of inhibitor injected.

whereas recovery of activity is limited by CO release from the enzyme (typical time constant  $\sim 1$  min).<sup>7a,7b</sup> Correction for the effect of film desorption is described in SI section III.<sup>8</sup> Inhibition by CO is fully reversible at  $-0.16$  V (Figure 1C) and only partly reversible at  $-0.36$  and  $-0.47$  V (Figure 1D,E). Experiments with the enzyme from *Cr* gave similar results (SI, Figure S2).

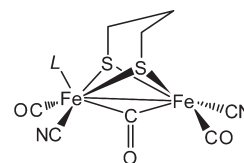
We fitted the chronoamperometric data (dashed lines in Figure 1) using the kinetic model we introduced before for interpreting oxygen inhibition.<sup>6,7a,7b</sup> As depicted in Figure 2, this model assumes that the formation of an inactive CO adduct (second-order rate constant  $k_{in}$ ) is followed by either CO release (rate  $k_{out}$ ) or irreversible inactivation (rate  $k_3$ ). This is supported by the finding from independent crystallographic and spectroscopic investigations that exogenous CO binds at a single position on the 2Fe subsite.<sup>10</sup> Figure 2 shows the dependence on electrode potential of the rate constants determined by fitting data such as those in Figure 1C–E. The values of the “in” and “out” rates are consistent with those determined in early experiments carried out with the enzyme from *Cp* without precise potential control.<sup>11</sup>  $k_{out}$  is constant over a large range of electrode potential, whereas  $k_{in}$  decreases and  $k_3$  increases in the potential range where we expect Hred to be the dominant species in the steady state. Indeed, the change in  $k_{in}$  is well described by an  $n = 1$  sigmoid centered on  $E = -0.33$  V (black line in Figure 2A), close to the equilibrium potential of the Hox/Hred couple ( $-0.4$  V at pH 8 in *Cr*<sup>2</sup>). This suggests that Hred is the species that reacts irreversibly with CO.

The loss of activity after prolonged exposure to CO at  $-0.47$  V is irreversible: it cannot be recovered by removing CO from the solution and poisoning the electrode at either very low or high potential, e.g., at  $-0.76$  V for 60 s (SI Figure S3, panels E, F) or at  $-0.76$  or  $-0.16$  V for 10 min (not shown). However, Figure S3



**Figure 2.** CO inhibition kinetic parameters obtained by fitting data obtained with *Ca* (black) and *Cr* (purple) H<sub>2</sub>ases (1 atm of H<sub>2</sub>, pH 7, 30 °C). The values of  $k_{in}$  obtained with *Cr* have been divided by 10. The vertical dashed line marks the OCP.

### Scheme 2. General Structure of Models 1–6 Used To Study Reaction with CO<sup>a</sup>



<sup>a</sup> The models formally correspond to the Fe<sup>II</sup>Fe<sup>I</sup> (1–3) or Fe<sup>I</sup>Fe<sup>I</sup> (4–6) redox states of the dinuclear cluster. See text for the nature of L. The formal valences of the cubane irons are 2Fe<sup>II</sup>2Fe<sup>III</sup>. Some properties of models 1, 3, 4, and 6 have been discussed in ref 12.

also demonstrates that the H-cluster can be rescued *in extremis* if the electrode potential is taken to a *very low* value (below  $-0.6$  V, where Hsred should be the dominant species) *before* the irreversible reaction has time to proceed (Figure S3, panels C, D); in this nonphysiological range of potential, the data could not be accurately analyzed (SI Figure S5), possibly because of an additional inactivation process.

The molecular origin of the observed behaviors is revealed by the results of DFT calculations carried out on models of the Hox, Hred, and Hsred forms of the active site (see SI for details on models and calculations). We first examine CO binding to a series of Fe<sup>II</sup>Fe<sup>I</sup> and Fe<sup>I</sup>Fe<sup>I</sup> models of the H-cluster (Scheme 2 and SI Table S1) characterized by the general structure  $[(\mu\text{-pdt})\text{Fe}_2(\text{CO})_3(\text{CN})_2(\text{L})]$ , where pdt = propanedithiolate and L = CH<sub>3</sub>S<sup>−</sup> (models 1 and 4), CH<sub>3</sub>SH (2 and 5), or  $[\text{Fe}_4\text{S}_4(\text{CH}_3\text{S})_4]^{2-}$  (3 and 6).

The most relevant DFT results are as follows: (i)  $\Delta G^0$  values for CO binding to Fe<sup>II</sup>Fe<sup>I</sup> models (from  $-6.4$  to  $-10.8$  kcal/mol) match remarkably well the experimental value for CO binding to Hox:  $-8$  kcal/mol for *Ca* FeFe hydrogenase, deduced from the ratio  $k_{out}/k_{in} = 1.8 \mu\text{M}$  at high potential in Figure 2A,B;  $-9$  kcal/mol for the enzyme from *Cr*, deduced from  $k_{out}/k_{in} = 0.4 \mu\text{M}$ . (ii) CO binds more strongly to the Fe<sup>II</sup>Fe<sup>I</sup> model 1 than to the corresponding Fe<sup>I</sup>Fe<sup>I</sup> model 4, but (iii) the reaction of CO with Hred models 5 and 6 leads to *irreversible* cleavage (see



(+0.3 Å), is not cleaved upon CO binding, indicating that in the H<sub>red</sub> state the structural integrity of the H-cluster is maintained.

Scheme 4 summarizes our findings. Most importantly, we conclude that CO binding to H<sub>red</sub> irreversibly inactivates the enzyme by breaking down the H-cluster, giving the inactive, inert compound  $[(\mu\text{-dtma})\text{Fe}_2(\text{CO})_4(\text{CN})_2]^{2-}$ . Although we do not have specific information about the fate of the stable, broken parts of the H cluster, we consider unlikely that they dissociate from the enzyme, because this would require a major conformation change to open a pathway through the protein matrix (as occurs for H-cluster maturation).<sup>16</sup> It will be interesting to perform further experiments to trap and spectroscopically characterize the inactivated active site. EPR could be used with the enzyme from Cr that has no electron-transferring FeS clusters, but the H<sub>red</sub> state will have to be characterized first,<sup>2</sup> and we have no indication that the dissociated cubane will be paramagnetic. However, it is likely that the CO-induced disruption of the H-cluster should have consequences on the FTIR signature of either enzyme.

It is remarkable that the reaction of CO binding engages the entire H-cluster, rather than only the Fe<sub>2</sub> subsite. This is reminiscent of the recent finding that O<sub>2</sub> inhibition involves initial attack of the distal Fe ion, followed by damage to the cubane.<sup>7a,17</sup>

## ■ ASSOCIATED CONTENT

**S Supporting Information.** Complete ref 6, experimental and numerical methods, raw data corresponding to Figure 1, electrochemical signals obtained with Cr FeFe hydrogenase, results of electrochemical experiments at very low potential, Table S1, description of the possible pathways for the reaction of Fe<sup>I</sup>Fe<sup>I</sup> models with CO, other numerical details, including total energy and optimized atomic coordinates of all models discussed in the paper. This material is available free of charge via the Internet at <http://pubs.acs.org>.

## ■ AUTHOR INFORMATION

### Corresponding Author

luca.degioia@unimib.it; christophe.leger@ifr88.cnrs-mrs.fr

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