



Immunoactive preparations and regulatory responses in the respiratory tract: potential for clinical application in chronic inflammatory airway diseases

Wojciech Feleszko, Giovanni A Rossi, Rafal Krenke, G Walter Canonica, Laura van Gerven & Oleg Kalyuzhin

To cite this article: Wojciech Feleszko, Giovanni A Rossi, Rafal Krenke, G Walter Canonica, Laura van Gerven & Oleg Kalyuzhin (2020): Immunoactive preparations and regulatory responses in the respiratory tract: potential for clinical application in chronic inflammatory airway diseases, Expert Review of Respiratory Medicine, DOI: [10.1080/17476348.2020.1744436](https://doi.org/10.1080/17476348.2020.1744436)

To link to this article: <https://doi.org/10.1080/17476348.2020.1744436>



© 2020 The Author(s). Published by Informa UK Limited, trading as Taylor & Francis Group.



Accepted author version posted online: 06 Apr 2020.



[Submit your article to this journal](#)



Article views: 78



[View related articles](#)



[View Crossmark data](#)

Publisher: Taylor & Francis & Informa UK Limited, trading as Taylor & Francis Group

Journal: *Expert Review of Respiratory Medicine*

DOI: 10.1080/17476348.2020.1744436

Article type: Review

Immunoactive preparations and regulatory responses in the respiratory tract: potential for clinical application in chronic inflammatory airway diseases

Wojciech Feleszko*¹, Giovanni A Rossi², Rafal Krenke³, G Walter Canonica⁴, Laura van Gerven⁵, Oleg Kalyuzhin^{7,6}

¹Associate Professor of Pediatrics, Department of Pediatric Respiratory Diseases and Allergy, The Medical University of Warsaw, Warsaw, Poland

²Chief Emeritus, Pediatric Pulmonology and Allergy Units, Cystic Fibrosis Regional Centre, IRCCS G. Gaslini, Genoa, Italy

³Full Professor of Respiratory Medicine, Department of Internal Medicine, Pulmonary Diseases & Allergy, Medical University of Warsaw, Warsaw, Poland

⁴Full professor, Personalized Medicine Asthma & Allergy, Clinic-Humanitas University & Research Hospital, Milan, Italy

⁵Associate Professor, Department of Otorhinolaryngology, Head and Neck Surgery, University Hospitals Leuven, Leuven, Belgium

⁶Professor of Department of Clinical Immunology and Allergy, I.M. Sechenov First Moscow State Medical University of the Ministry of Health of the Russian Federation (Sechenov University), Moscow, Russia

*Corresponding author:

Wojciech Feleszko,

Department of Pediatric Respiratory Diseases and Allergy,

The Medical University of Warsaw,

ul. Zwirki i Wigury 63A, PL-02-091 Warsaw, Poland.

Mobile: ++48-693-468-074;

E-mail: wfeleszko@klinikzny.pl or wojciech.feleszko@wum.edu.pl

ACCEPTED MANUSCRIPT

Abstract:

Introduction: The prevalence of chronic inflammatory airway diseases is rising. Their treatment with corticosteroids increases infection risk, while overuse of antimicrobial agents may increase morbidity and antimicrobial resistance. Non-specific immunomodulatory compounds alter immune responses to both infectious and atopic challenges. These compounds may offer an alternative approach for symptom reduction and prophylaxis against both infections and exacerbations in chronic inflammatory airway disease.

Areas covered: We assessed the available data on the efficacy of non-specific immunomodulators including bacterial lysates, synthetic compounds, and vaccines in chronic rhinosinusitis (CRS); allergic and non-allergic rhinitis; chronic obstructive pulmonary disease (COPD), and asthma. A search of PubMed was carried out using the 'Clinical Trials' filter for each condition and immunomodulatory product detailed below, where available, data from meta-analyses were reported.

Expert opinion: Pre-clinical data has revealed a coherent mechanistic path of action for oral immunomodulators on the respiratory immune system, principally via the gut-lung immune axis. In patients with asthma, allergic rhinitis, CRS, and COPD immunomodulatory therapy reduces symptoms, exacerbations, hospitalizations, and drug consumption. However, data are heterogeneous, and study quality remains limited. A lack of high-quality recent trials remains the major unmet research need in the field.

Keywords: immunomodulation; immunostimulation; asthma; chronic obstructive pulmonary disease; chronic rhinosinusitis; allergic rhinitis; bacterial lysates; probiotics

ACCEPTED MANUSCRIPT

Article highlights

- Pre-clinical data have created a detailed picture of the action of immunomodulators on the respiratory immune system principally via the gut-lung immune axis.
- In patients with asthma, allergic rhinitis, chronic rhinosinusitis, and chronic obstructive lung disease immunomodulatory therapy can reduce symptoms, exacerbations, hospitalizations, and drug consumption.
- The results of some available meta-analyses on the clinical effects of immunomodulators have not been addressed in the current guidelines.
- A lack of high-quality trials remains the major unmet research need in the field.

ACCEPTED MANUSCRIPT

1.0 Introduction

1.1 Unmet needs in chronic inflammatory airway diseases

Chronic inflammatory airway diseases can be anatomically grouped into those of the upper (Chronic Rhinosinusitis [CRS], Persistent Allergic Rhinitis [AR] and Persistent Non-Allergic Rhinitis (NAR)) and lower (Chronic Obstructive Pulmonary Disease [COPD] and asthma) airways. Although in the majority of patients with chronic airway diseases symptoms can be effectively controlled by standard therapies, a significant minority remain symptomatic despite recommended pharmacological treatment [1]. Moreover, impaired quality of life, social functioning, sleep, and school/work performance are commonly reported, even by patients with relatively good symptom control [1].

Prevalence of AR and asthma are on the increase, contributing to the allergy pandemic which is, at least partially, an unwanted consequence of increased urbanization worldwide [2]. In a 2014 global survey, approximately 70% of Ear, Nose and Throat specialists stated that the prevalence of AR was 'surely increasing' in their country [3]. Global Burden of Disease Study data indicate that the prevalence of asthma increased by approximately 13% between 1990 and 2015, while the same data source showed a 44% increase in COPD prevalence over the same period [4]. In addition, an allergic phenotype of COPD characterized by increased respiratory symptoms and a higher risk of exacerbations was described during the past decade [5, 6]. COPD is currently the 3rd leading cause of death globally and, like asthma, its prevalence is rising [7]. Importantly, patients with allergic and non-allergic asthma, as well as patients with COPD, show signs of nasal inflammation and report higher levels of nasal symptoms [8]. This reflects a phenomenon known as 'united airway disease' which is indicative of the integrated nature of the upper and lower airways. This heterogeneous condition encompasses multiple endotypes which may be both atopic and non-atopic in nature [8, 9].

Current treatment guidelines for chronic inflammatory airway diseases all rely to a greater or lesser extent on anti-inflammatory therapeutics, particularly corticosteroids, as well as infection control when necessary [10, 11, 12]. Next-generation monoclonal therapies have shown promise in COPD, asthma, CRS, and AR [13, 14, 15, 16]. However, these therapies remain expensive and beyond the reach of developing countries where the majority of the world's population reside and where the prevalence of allergic respiratory disease is likely to rise alongside increasing urbanization [2, 17].

Reliance on corticosteroid therapy, which is common in moderate-to-severe chronic inflammatory airway disease, and during exacerbations, increases the risk of upper and lower respiratory-tract infections, particularly in patients with asthma and COPD [18, 19, 20]. As respiratory infections are a well-known risk factor for exacerbations and worsening of inflammatory airway diseases, a vicious circle may emerge with increased risk of infections due to corticosteroid therapy and further worsening of the disease triggered by infections [21, 22]. An overuse of antimicrobial agents in patients with chronic inflammatory airway diseases is also common and may result in unnecessary or prolonged hospital stays, higher costs, and increased risk of treatment failure [23]. Furthermore, data from animal models suggest that antimicrobial-induced reductions in microbiome diversity are related

to impaired immune function and higher mortality following viral infection [24, 25**]. Emerging evidence from human population studies suggests that community structures of commensal bacterial also modulate susceptibility to infection [26].

1.2 The gut-lung immune axis: infection and inflammation

The “hygiene hypothesis” and other related concepts, including the ‘old friends’ and ‘biodiversity’ hypotheses, have become cornerstones of modern immunology. These theories are supported by two widely accepted concepts that can be briefly outlined as follows: contact with the external environment is essential for the biodiversity of the internal environment; and the biodiversity of the internal environment is essential for proper immune function [27, 28, 29].

Mucosal membranes, in particular those of the gut and respiratory system, represent the major points of contact between the immune system and the environment. A wealth of evidence, extensively reviewed elsewhere, has demonstrated that the mucosal immune system functions as an integrated unit, with substantial crosstalk allowing an integrated immune response against pathogens detected in either mucosa [2, 30, 31, 32]. This so called gut-lung axis was proposed as early as the year 2000, however, the concept that changes in the gut microbiome may impact lung immunity has gained particular traction within the scientific community during the last 5 years [1]. Reduced microbial diversity, so call dysbiosis, in the gut has been linked to lung disease in humans, and increasing evidence suggests that both microbial components (antigens) and metabolites such as short-chain fatty acids have a role in maintaining immune homeostasis [32].

Dendritic cells (DCs) function as immune sentinels within the mucosal membranes, where the detection of live bacteria, viruses, or their associated antigens by pattern recognition receptors (PRR) triggers an immune cascade. The resulting migration of innate and adaptive immune cells via the lymphatic system confers protection against infections throughout the body’s mucosae. In addition, increasing evidence suggests that immune activity at one mucosal site can result in migration of regulatory cells which modulate activity at distant mucosal membranes reducing inflammation and atopic reactions [32, 33, 34, 35] (**Figure 1**).

Non-specific immunomodulatory compounds interact with the innate and acquired immune system, modulating the immune response to various challenges including infectious, atopic, or neoplastic insults. Non-specific immunomodulators include bacterial lysates, synthetic modulators of PRR-mediated signaling, and vaccine adjuvants. Immunoactive preparations have demonstrated the ability to activate DCs in the digestive-tract mucosa resulting in a protective immune response in the lungs [33, 36]. Much of this anti-infective response depends on the creation of an immunostimulatory, proinflammatory milieu [33]. Yet, a growing body of evidence indicates that such preparations can be also effective in the chronic inflammatory airway diseases mentioned above [33]. The explanation behind this apparent mechanistic dichotomy relies on the ability of immunomodulatory compounds to

mimic the activity of naturally occurring microbiota. During infancy, and in response to allergens, atopy-related type 2 T-helper cells (Th2) responses predominate [37, 38]. Exposure to microbiota or immunomodulatory compounds can shift the balance from an atopic Th2 response to a non-atopic type 1 (Th1) response via the activation of DCs and subsequent regulatory T-cell activity [25^{••}, 33^{••}, 35, 39, 40, 41, 42, 43].

In this paper we will summarize the mechanistic and clinical evidence base for the activity of various immunomodulatory compounds in chronic airway diseases. Searches for the treatment and condition were carried out in PubMed using the 'Clinical Trials' filter e.g. (OM-85 [title/abstract] OR bronchovaxom [title/abstract]) AND asthma (OM-85 [title/abstract] OR bronchovaxom [title/abstract]) AND wheeze), supplemented with previous literature reviews and author expertise. Condition-specific mechanistic data will be summarized briefly and, whenever possible, high-quality data from meta-analyses or randomized trials will be reported.

2.0 Probiotics

2.1 Asthma

The mechanistic underpinning of probiotics as an anti-asthma intervention is derived from hypotheses which link a lack of exposure to diverse microbial agents with the risk of developing atopic disease (i.e. the hygiene hypothesis). Numerous bacterial species have been investigated as probiotics, with *Bifidobacterium* and *Lactobacillus* being the major probiotic genera [44, 45, 46]. Several studies in animal models, suggest a beneficial role of probiotics in asthma prevention [47[•], 48].

A recent meta-analysis that included pooled data from 910 children with mild to moderate asthma demonstrated a higher proportion of children with fewer asthma episodes following treatment with probiotics as compared to controls (risk ratio [RR]: 1.3; 95% confidence interval [CI]: 1.06, 1.59). In addition, the authors reported a probiotic-induced increase in IFN- γ and a decrease in IL-4 levels. However, the above effects were not associated with significant changes in disease severity as measured by childhood asthma control test, asthmatic symptoms during the day or night, number of symptom-free days, forced expiratory volume (expressed as the percentage of predicted), or peak expiratory flow (**Table 1**) [49]. In a large meta-analysis comprising data from 5,157 infants < 1 year old exhibiting asthma or wheezing, probiotic therapy was not found to reduce the overall risk of asthma (RR: 0.94; 95% CI: 0.82, 1.09) or wheeze (RR: 0.97; 95% CI: 0.88, 1.06) (**Table 1**) [50]. However, probiotics administration was associated with a reduction in the incidence of wheeze among infants with atopic disease (n=225; RR: 0.61; 95% CI: 0.42, 0.90) [50]. Finally, in a meta-analysis comprising 4,755 children at high risk of atopy there was no difference in the incidence of asthma and wheezing between infants who received probiotics (gestational or at < 3 months old) compared with those who did not (RR: 0.99; 95% CI: 0.77, 1.27; and RR: 1.02; 95% CI: 0.89, 1.17, respectively) (**Table 1**) [51].

Overall, these results suggest that probiotics lack consistent efficacy in asthma prevention, however, their use may be associated with a reduction in the incidence of wheeze in atopic infants, although larger trials are necessary.

2.1.1 Asthma guidelines

The sole mention of non-specific immunoactive preparations in the current Global Initiative for Asthma (GINA) report refers to the negative results of a meta-analysis on probiotics and asthma development [11]. Favorable statements in the recently published review by the European Academy of Allergy and Clinical Immunology Task Force on Anti-infectives in Asthma may represent the beginning of a shift in perception within the research community. However, the need for higher quality studies is emphasized [55].

2.2 COPD/chronic bronchitis

Few mechanistic studies on the effects of probiotics in COPD have been carried out. However, the NK-1 suppressing effect of cigarette smoke was attenuated in smokers given fermented milk containing *L. casei* [56]. Also, an *in vitro* study showed that phagocytosis of *L. rhamnosus* or *B. breve* by human macrophages was associated with the inhibition of inflammatory response to cigarette smoke [57]. The above-discussed inflammatory cross talk between gut and airways is probably one of the major mechanisms responsible for the ability of probiotics to suppress cigarette-smoke induced airway inflammation [44, 56].

We were unable to find any clinical data on the effect of probiotics on the severity of airway inflammation in patients with COPD/chronic bronchitis.

2.3 Chronic rhinosinusitis

CRS is a chronic inflammatory condition of the nose and paranasal sinuses with a multifactorial etiology and pathophysiology and two major phenotypes; CRS with and without nasal polyps (CRSwNP and CRSsNP, respectively) [58]. Genetic predisposition and environmental factors (exposure to cigarette smoke) and microbial factors (changes in microbiome diversity, biofilm formation) contribute to the onset and course of the disease, influencing epithelial barrier integrity and the adaptive immune response [59, 60, 61, 62, 63].

The conceptual basis for probiotic protection against CRS relies principally on the effects of commensal organisms directly on the mucosa of the upper airway, rather than indirectly via the shared mucosal immune axis [64]. Probiotic species may act as direct competitors to pathogens, both for nutrients and access to cell surface receptors which aid adherence [65]. In addition, production of antibacterial peptides and metabolites may create a more hostile environment for pathogens, for example lowering of pH by lactobacilli has been proposed as an inhibitor of *Pseudomonas aeruginosa*, a common pathogen in CRS [65]. Topical probiotics also have the potential to modulate the immune

system. Intranasal delivery of *Bacillus subtilis* in piglets resulted in increased expression of PRRs, increased numbers of DCs, B cells and T cells, as well as higher levels of IgA [66]. In a randomized trial of a nasal spray containing various lactobacilli and bifidobacteria there was no effect on symptom scores, microbiological flora, or inflammatory mediators. However, the study showed that the topical application was feasible and safe (**Table 2**) [67^{*}]. Oral probiotics have also been investigated in CRS. In a randomized trial, oral *L. rhamnosus* had no effect on sinonasal quality-of-life scores (**Table 2**) [68].

2.3.1 Chronic rhinosinusitis guidelines

Probiotics are not recommended by the current European guidelines for CRS and these agents have not been mentioned in the Canadian or US guidelines [58, 72, 73]. Also, despite acknowledgement and discussion of the importance of the microbiome, a recent international consensus paper on rhinosinusitis does not refer to probiotics [74].

2.4 Allergic rhinitis

Persistent AR occurs for periods longer than four days per week during more than four consecutive weeks. Both symptoms and their impact on quality of life range from mild to severe.

Down regulation of type 2 immune response following intranasal delivery of probiotics has been demonstrated in a mouse model of AR [75, 76, 77]. Additionally, oral intake of probiotic products has been suggested to improve health status in children with AR [76, 78]. Meta-analysis data have demonstrated an effect of probiotics on the type 1:type 2 immune ratio, but no apparent effect on IgE levels [79, 80^{*}, 81^{*}].

In a meta-analysis comprising 2,242 children and adults with AR, probiotic therapy resulted in a significant reduction in total nasal (standardized mean [SMD] difference: -1.23 ; 95% CI: -1.84 , -0.62) and total ocular symptoms scores (SMD: -1.84 ; 95% CI: -2.83 , -0.84). QoL was significantly better in a sub-analysis of the Lp-33 lactobacillus strain. However, total nasal and ocular quality-of-life scores were similar to placebo (**Table 4**) [79]. Conversely, in a meta-analysis of 1,919 patients the positive effect of probiotics on Rhinitis Quality of Life (RQLQ) scores, was significant (SMD: -2.23 ; 95% CI: -4.07 , -0.40) though there was no effect on Rhinitis Total Symptom Scores (RTSS) (SMD -0.36 ; (95% CI: -0.83 , 0.10)) (**Table 4**) [80^{*}]. In their 2015 systematic review (N=1,527), Peng et al. reported a significant combined improvement of the quality-of-life scores and nasal symptom scores of patients with AR (mean difference: -2.97 [95% CI, -4.77 , -1.16]) (**Table 4**) [81^{*}]. There was a high risk of bias in the majority of the studies, highlighting the need for better quality trials [79, 80^{*}].

2.4.1 Allergic rhinitis guidelines

Currently there is no mention of probiotics in the Allergic Rhinitis and its Impact on Asthma (ARIA) guidelines, despite available data from meta-analyses suggesting an impact on symptom control [86].

2.5 Non-allergic rhinitis

NAR is a heterogeneous group of conditions including rhinitis of the elderly, hormonal-induced rhinitis, gustatory rhinitis, occupational rhinitis and idiopathic rhinitis. In the small randomized study, patients received standard of care with or without an oral probiotic supplement containing *L. acidophilus* NCFM, *B. lactis*, and fructo-oligosaccharides. The probiotic group had a lower prevalence of symptoms and endoscopic signs, and a reduction in inflammatory cell infiltrate (**Table 4**) [82].

2.5.1 Allergic rhinitis guidelines

In line with the level of evidence, there is currently no mention of probiotics in the US or European AR guidelines [87, 88].

3.0 OM-85

3.1 Asthma

OM-85 is produced by the alkaline lysis of 21 strains of eight species of common bacterial respiratory-tract pathogens (see footnote **Table 1**). The results of studies on experimental models suggest that OM-85 remains immunologically active following ingestion [89]. The proposed mechanism of OM-85's immunomodulatory activity was recently reviewed [33^{**}]. Briefly, a two-pronged anti-infective and anti-inflammatory mechanistic path has been proposed for OM-85 in wheezing and asthma. This includes reductions in viral infections, and associated exacerbations, likely via release of antimicrobial peptides, activation of macrophages, release of antiviral cytokines, and rapid neutrophil recruitment (**Figure 1**) [33^{**}, 90, 91, 92, 93, 94^{*}]. In addition, OM-85 induces a shift in balance of type 1 and type 2 immune responses reducing the likelihood of allergen-induced airway hyper-responsiveness during exacerbations [33^{**}, 35, 91, 95]. In mouse models of asthma, OM-85-induced: down regulation of type 2 markers on gut DCs; damping of DC-responses to aeroallergens; trafficking of Treg cells to the airway; a shift towards a type 1 cytokine balance; reduced inflammatory cell infiltration; and reductions in serum IgG1 (IgG4 equivalent) and IgE (**Figure 1**). These effects were associated with suppression of mucous metaplasia and mucus hypersecretion, reduced airway hyper-responsiveness, and remodeling [33^{**}, 35, 89, 90, 91, 92, 93, 96^{*}, 97, 98, 99]. Shifts in the cytokine balance in favor of a type 1 rather than type 2 immune response were consistently demonstrated in children with asthma and related conditions [33^{**}, 53^{**}, 99, 100, 101].

A recently published meta-analysis (N=4,851) of the efficacy and safety of OM-85 in pediatric recurrent respiratory-tract infections included 8 studies (702 children) which assessed wheezing episodes. OM-85 therapy resulted in a substantial reduction in the duration of wheezing (mean difference -3.37; 95% CI: -4.25, -2.22) (**Table 1**) [52]. Another recently submitted meta-analysis

investigated the efficacy of bacterial lysates in children with preschool wheezing and asthma. Three trials assessing OM-85 were eligible for data extraction (N=88). The frequency of wheezing episodes and asthma exacerbations was reduced in patients who received OM-85 compared with placebo (**Table 1**). The quality of the trials of bacterial lysates overall was assessed as intermediate [53**].

In the 33 studies which reported adverse events (AEs) in the Yin meta-analysis, OM-85 was well tolerated with mild gastrointestinal symptoms and rash being the most common AEs. Although AEs were more common in patients treated with OM-85 they did not interfere with treatment (RR = 1.39; 95% CI: 1.02, 1.88, P = 0.04) [52]. AEs were not reported separately for OM-85 in the recently submitted meta-analysis, however, AEs for bacterial lysates overall had a similar incidence as those occurring in placebo/standard of care groups [53**].

The above analyzes acknowledged the fact that higher quality trials are required before firm conclusions could be drawn regarding the prophylactic efficacy of OM-85 in asthma/wheezing. The large scale ORal Bacterial EXtracts (ORBEX, NCT02148796; N=986) for the prevention of wheezing lower respiratory-tract illness trial, should contribute towards addressing this need [102]. The trial will address infants and young children at risk of developing chronic airway disease due to familial atopy, some of whom have experienced 1–2 wheezing episodes, and will investigate whether OM-85 prevents or delays wheezing. Results are expected in 2024.

3.1.1 Asthma guidelines

OM-85 is not currently mentioned in the GINA report [11]. As with probiotics, the European Academy of Allergy and Clinical Immunology Task Force on Anti-infectives in Asthma mentions the potential of bacterial lysates alongside the need for higher quality studies before inclusion in guidelines can be recommended [55].

3.2 COPD/chronic bronchitis

A number of studies have evaluated the potential mechanisms of OM-85 prophylaxis against COPD exacerbations. OM-85 therapy increased the CD4+:CD8+ lymphocyte ratio and IFN- γ levels, and exerted a regulatory effect on alveolar macrophage activity and IgA levels in the bronchoalveolar lavage fluid of patients with non-obstructive chronic bronchitis [103]. OM-85 was shown to play a protective role against Rhinovirus-induced cell death in isolated bronchial epithelial cells from COPD patients. Improved antiviral activity due to increased bronchial epithelial cell expression of immune proteins C1q-R and β -defensin as well as activation of Erk1/2 MAPK, an upstream regulator of macrophage activity, likely contribute to OM-85's protective effect [94*]. IL-1 β is a prominent mediator of COPD which drives neutrophilic inflammation and has been implicated in the initiation, maintenance and exacerbation of airway inflammation [104]. Data indicating OM-85 may interfere with inflammasome activation and IL-1 β release induced by alum suggests a potential route for the inhibition of airway inflammation in COPD, particularly during exacerbations. A similar mechanism is likely also at play during asthma exacerbations [90]. Although specific data are sparse, *in vitro* data,

including from peripheral blood mononuclear cells from patients with COPD, suggests that a dual anti-infection/anti-inflammation mechanism similar to that proposed in asthma may play a role in the reduction of exacerbation rates in COPD patients treated with OM-85 (see data below) [90, 94*].

A meta-analysis on the effects of OM-85 therapy in COPD published in 2015 included five randomized trials and 1,190 patients. The exacerbation rate was reduced by 20% (RR: 0.80; 95% CI: 0.65, 0.97) and the incidence of antibiotic use was reduced by 39% (RR: 0.61; 95% CI: 0.48, 0.77) compared with placebo. The authors did not find any change in duration of hospitalization (weighted mean difference: -7.33 days; 95% CI: -18.34, 3.67; P = 0.19), severity of acute exacerbations (weighted mean difference: 0.72 score; 95% CI: -1.55 to 0.11 score; P = 0.09), or total adverse events (RR, 0.94; 95% CI: 0.84 – 1.05; P = 0.24) (**Table 3**). The need for better quality trials was emphasized in the conclusions of this meta-analysis [105**]. In a second more recent meta-analysis (N=1,008) presented at European Academy of Allergy and Clinical Immunology 2019, a 23% reduction in COPD exacerbation rate was reported (RR: 0.77; 95% CI: 0.68, 0.88; P = 0.0002) (**Table 3**) [106].

Three cost-effectiveness trials of OM-85 in COPD/chronic bronchitis have been carried out. In an Italian study conducted in the 1990s, the effective cost savings per patient per 6 months were 51,095 Lira, a 60% cost saving, and these savings were found to be associated solely with OM-85's ability to prevent exacerbations [113]. A Canadian study published in 2001 analyzed data collected in 1995. The cost of preventing one day of hospitalization due to a respiratory cause was CND\$45 (95% CI: 18, 210), with an average cost for one day of respiratory hospitalization at \$382 leading to an 88% saving per hospitalization prevented [114]. The authors of a more recent Argentinian study reported a 21% reduction in total costs related to reinfections in a group of COPD patients who received OM-85 prophylaxis (1,356.5 vs 1,708.2 ARS). Similarly, the total cost associated with disease exacerbations was reduced by 24% (1,217.5 vs 1,599.6 ARS) [115].

3.2.1 COPD guidelines

Current GOLD Science Committee Guidelines mention two RCTs of OM-85, both of which were conducted before 2005, noting the reductions in disease severity and exacerbations achieved. The Committee recommends the need for further studies into the long term effects of 'immunoregulators' alongside current maintenance therapies [10].

3.3 Chronic rhinosinusitis

A study examining potential mechanisms for OM-85 prophylaxis in CRS has been carried out using a mouse model [116]. Administration of OM-85 alongside amoxicillin vs amoxicillin alone significantly reduced levels of suppressor of cytokine signaling 1 (SOCS1), SOCS3, TNF- α , and IFN- γ . Both SOCS1 and SOCS3 have been found to be upregulated in CRS, and both TNF- α , and IFN- γ have been shown to be drivers of SOCS1 and SOCS3 expression. The authors suggest that reduced expression of SOCS1 and SOCS3 may ameliorate CRS-related inflammation [116]. A second study showed direct application of OM-85 to the sinus increased ciliary beat frequency via a NO dependent mechanism,

which the authors contend may have protective effects in rhinosinusitis [117]. In an older trial, efficacy of OM-85 in children with CRS correlated with increased IgA levels, presenting a potential anti-infective mechanism [71].

A randomized trial was carried out in children during the acute remission phase of CRS, in which OM-85 significantly improved the visual analogue score for rhinosinusitis, nasal discharge and obstruction scores, number of days with rhinitis attacks per month, and number of days with antibiotic use per month over the following year (**Table 2**) [69]. In a randomized trial in adults with chronic purulent sinusitis, OM-85 improved the rate of infections, symptom scores, and x-ray sinus opacity (**Table 2**). Side-effects were minimal with the same number of patients reporting AEs in the placebo and OM-85 groups (n = 2) [70]. In a 6-month study conducted in 1988, incidence of cough, nasal discharge scores (up to month 5), nasal congestion, episodes of infection, and the duration of antibiotic therapy were reduced in children treated with OM-85 following an acute episode of rhinosinusitis (**Table 2**). No side effects were observed during therapy in either treatment group [71]. Finally, in a Chinese cost-effectiveness analysis of OM-85 therapy, the cost to prevent a full episode of rhinosinusitis exacerbation was RMB 1182.84, while each acute exacerbation of rhinosinusitis cost RMB 1807.21, representing a 35% cost saving [118].

3.3.1 Chronic rhinosinusitis guidelines

Bacterial lysates in general, and OM-85 in particular, are recommended for the treatment of CRS in adults by the European guideline/Consensus Paper with a very high – A grading [58]. A similar recommendation is also given in a Pan-American guideline paper [119].

3.4 Allergic rhinitis

Inflammatory cytokines, infiltration of proinflammatory cells, and increased serum IgG1 and IgE, indicative of a type 2 immune response, are inhibited by OM-85 in a mouse model of AR [33^{**}, 120].

In a study of adult patients with AR, infections were reduced by 33%, although there was no change in the number of exacerbations (**Table 4**) [83]. In a small (N=60) randomized trial recently carried out in China, OM-85 therapy reduced medication use, nasal symptoms, and improved type 1:type 2 cytokine balance [84]. A cost-effectiveness analysis carried out in Argentina showed that total cost per patient per month caused by reinfections and exacerbations were 448.9 ARS and 269.9 ARS, respectively in patients treated with OM-85, compared with 660.40 ARS and 574.40 ARS in the year previous to treatment [115].

3.4.1 Allergic rhinitis guidelines

Currently there is no mention of OM-85 in the Allergic Rhinitis and its Impact on Asthma (ARIA) guidelines or the most recent US allergic rhinitis guidelines [86, 88].

4.0 Pidotimod

4.1 Asthma

Pidotimod is a synthetic thymic dipeptide which appears to share a number of mechanistic similarities with bacterial immunomodulators [33**, 121]. Pidotimod is thought to stimulate two PRRs, toll-like receptor 2 (TLR2) and TLR4, which are expressed on DCs (**Figure 1**) [33**, 122, 123, 124, 125]. Single nucleotide polymorphisms of PRRs have a role in a number of chronic lung diseases, and reduced expression of these receptors has been found to correlate with the severity of atopic disease [126, 127]. In an *ex vivo* study examining the effects of pidotimod on peripheral blood mononuclear cells from children with atopic asthma and healthy controls, the type 2-related cell marker CD30 was down regulated in response to pidotimod in both groups [128]. CD30 has been implicated in the development of atopic asthma, and its expression appears to play a role in type 2 cytokine production by CD4+ and CD8+ human T-cell clones [128, 129, 130, 131]. However, mechanistic data are mixed. In a mouse model of asthma, proinflammatory effects of pidotimod on IgE and eosinophils were demonstrated [132].

To date, there have been no high-quality meta-analyses on the efficacy of pidotimod in asthma. Also, there are no studies assessing the impact of pidotimod on the rate of asthma exacerbations. However, in a prospective multicenter trial, pidotimod reduced the number of respiratory infections during a three-month observation period in a mixed group of children over half of whom had atopic conditions including asthma (**Table 1**). But, there was no longer a significant difference at the 6-month follow-up [54].

4.1.1 Asthma guidelines

There is no mention of pidotimod in current GINA guidelines, however, TLR agonists are mentioned alongside other non-specific immunomodulators by the European Academy of Allergy and Clinical Immunology Task Force on Anti-infectives in Asthma, with the above-mentioned caveats [11, 55].

4.2 COPD/Chronic bronchitis

The potential efficacy of pidotimod in COPD was reviewed in 2015 [125]. Since then, we have not found any new data on the use of this molecule in COPD patients. In an early mechanistic study, pidotimod was found to increase T-cell blastogenesis in patients with COPD, highlighting a potential route for potentiation of immune response in COPD [133].

Three randomized trials of pidotimod have been conducted in Italy in patients with chronic bronchitis. Pidotimod reduced the rate of infectious relapses and improved expectoration characteristics (serous, mucous, mucopurulent, or purulent) (**Table 3**). Side effects were transient and mild in nature and occurred to a similar rate in placebo and pidotimod groups [107, 125]. Similarly, pidotimod added to amoxicillin resulted in faster symptom resolution (dyspnea, cough, sputum, and hyperpyrexia) in patients with chronic bronchitis suffering from bacterial exacerbations (**Table 3**). Six patients had

adverse reactions (nausea; rash; vomiting) at the beginning of treatment resulting in discontinuation of pidotimod, in the placebo group 3 patients withdrew treatment due to AEs [108]. In clinically stable patients with severe COPD (GOLD stage 3), those receiving pidotimod exhibited fewer infectious exacerbations, a longer time to first exacerbation, and a shorter duration of acute infectious episodes. These differences were associated with lower use of antibiotics and less interference with daily activities and work in the pidotimod group (**Table 3**). Therapy was well tolerated with AEs occurring in approximately 6% of both the pidotimod and placebo group [109, 125]. Given the lack of recent clinical trials on pidotimod in COPD, further studies would be helpful in strengthening conclusions regarding efficacy in this group.

4.2.1 COPD guidelines

There is no mention of pidotimod in the current COPD guidelines [10].

5.0 Polyvalent mechanical bacterial lysate (PMBL)

5.1 Asthma

PMBL is a sublingually delivered bacterial lysate produced using a mechanical process that preserves the structure of bacterial antigens [33^{••}, 134]. PMBL increases the number of CD8⁺ T cells, Treg cells and NK cells in children with uncontrolled or partially controlled asthma (**Figure 1**) [33^{••}, 135].

Other, potentially relevant immunological effects on asthma prevention include a pro-type 1 shift in cytokine balance, as well as infection prophylaxis which may protect against exacerbations.

Two trials assessing the efficacy of PMBL in preschool wheezing and asthma (N=136) were included in the previously mentioned meta-analysis on the effect of bacterial lysates on preschool wheezing. The frequency of wheezing episodes and asthma exacerbations was reduced, although the reduction did not reach significance compared with placebo (**Table 1**). AEs were not reported separately for PMBL, however, AEs for bacterial lysates overall had a similar incidence as those occurring in placebo/standard of care groups. The overall quality of the trials of bacterial lysates was assessed as intermediate [53^{••}].

5.1.1 Asthma guidelines

There is no mention of PMBL in current GINA guidelines, however, as mentioned above the European Academy of Allergy and Clinical Immunology Task Force on Anti-infectives in Asthma mentions the potential of bacterial lysates and the need for further research [11, 55].

5.2 COPD/Chronic bronchitis

In elderly patients with COPD receiving PMBL, peripheral blood samples were assessed to establish the phenotypes of circulating peripheral blood mononuclear cells. Changes were evident in B cell

subsets which may be indicative of PMBL-induced maturation, potentially driven by increased numbers of Th (CD3+/CD4+) cells. CD8+ and CD25+ T-cell subsets were unaffected, while the number of Treg cells was reduced and the number of circulating NK cells increased significantly. The authors suggested that these results are indicative of a broad immune response to PMBL involving polyclonal activity [136].

In a pilot study comparing patients treated with an inhaled corticosteroid (ICS) and long acting beta-agonist (LABA) combination (salmeterol/fluticasone) vs patients treated with ICS/LABA plus PMBL, the latter group were characterized by a reduced exacerbation rate, fewer hospitalizations, and a reduced need for both oral corticosteroids and antibiotics (**Table 3**) [110]. In a multicenter phase IV trial of PMBL in patients with moderate to very severe COPD, the primary outcome of a 25% reduction in exacerbation rate was not achieved. However, the number of days with fever and days in poor health were significantly fewer in patients treated with PMBL. Also, the interval between the first and second exacerbations was significantly longer (**Table 3**). Potentially treatment-related AEs were uncommon at 3% with placebo and 0% with PMBL [111*]. Finally, in a randomized trial in elderly patients with COPD, PMBL resulted in increased levels of circulating antibodies and reduced infectious episodes (**Table 3**) [112].

PMBL appears to be effective in the reducing exacerbation rates in COPD, presumably due to a reduction in infectious episodes. We note that the studies on PMBL were better designed than many others in the field (power calculations, CONSORT reporting etc.). We strongly encourage the conduct of similar high-quality studies on immunoactive preparations in order to challenge skepticism within the medical community and produce definitive results.

5.2.1 COPD guidelines

There is no mention of PMBL in the current COPD guidelines [10].

5.3 Allergic rhinitis

In a small randomized trial, 61.5% of patients with AR treated with PMBL showed a significant and clinically relevant improvement (graded by symptom scores) while the rest (38.4%) showed a stationary clinical response (**Table 4**). No cases of negative side effects were reported. The authors also evaluated immunological response, although there were no changes in IgE and IFN- γ concentrations, some patients showed a decrease in the type 2 cytokine IL-4 [85]. A recent open-label study in 38 children suffering from Seasonal Allergic Rhinitis (SAR) caused by grass pollen treated with PMBL and standard treatment during pollen season demonstrated substantial reduction of the symptoms of SAR as compared to the control group [137].

5.3.1 Allergic rhinitis guidelines

Currently there is no mention of PMBL in the Allergic Rhinitis and its Impact on Asthma (ARIA) guidelines or the most recent US allergic rhinitis guidelines [86, 88].

6. Conclusion

Although research has been gathering pace in recent years, the data regarding the impact of non-specific immunomodulators on chronic respiratory diseases remain heterogeneous. Initial excitement around probiotics has been tempered by discouraging data in terms of asthma prevention although some efficacy in symptom control is apparent. Although bacterially derived immunomodulators have been used clinically for many years, the past decade has seen a substantially increased understanding of the mucosal immune-derived mechanisms underlying their clinical effects. Positive trial results must be viewed through a prism of skepticism due to the low quality of many studies and there is a tremendous need for methodologically robust, placebo-controlled studies designed according to the established principles for conducting clinical trials (CONSORT criteria). However, in cases where the data are sufficiently robust, expertly conducted meta-analyses have shown consistent efficacy for some of the immunomodulators discussed above.

Despite this, current guideline recommendations appear to be ignorant or dismissive of the recent robust meta-analysis-derived data. Though reductions in symptoms and exacerbations may be moderate in some cases, given the good safety profile of these immunomodulatory preparations, consideration of their utility as potential complimentary therapies which may reduce reliance on corticosteroids must be given more serious consideration.

7. Expert opinion

Following an initial phase of distrust, the growing body of evidence for the efficacy of immunoactive preparations in chronic inflammatory airway diseases has attracted the attention of researchers and physicians. Data supports a cogent mechanistic pathway for oral immunomodulatory products through the activity of the gut-lung immune axis. Efficacy against both infectious and inflammatory insults via dual regulatory and non-specific immune responses also has a clear mechanistic backbone. However, further studies are required to fill in gaps in this picture and identify potential biomarkers of efficacy (see below). It is also necessary to investigate whether the persistent clinical and immunomodulatory effects of the above drugs are associated with epigenetic reprogramming of innate immune memory, so called trained immunity [155].

Despite the substantial strides forward in understanding the mechanism of action of bacterially derived immunoactive preparations, there is little understanding of how the specific composition of these preparations relates to efficacy. Comparative studies on the clinical efficacy and mechanisms of action of immunoactive drugs with different compositions and routes of administration are required (bacterial lysates vs synthetic modulators of PRR-mediated signaling, sublingual bacterial lysates vs oral bacterial lysates, bacterial lysates vs probiotics, PMBLs vs chemical bacterial lysates etc.). The logical aim of these studies is to identify the most biologically active synthetic or purified natural components of bacterial cells mimicking the main immunomodulatory effects of multicomponent

bacterial preparations; while recognizing that this multicomponent nature may, in fact, be intrinsic to their efficacy.

As inflammatory and immune reactions are deeply involved in pathogenesis of chronic obstructive airway diseases, immunoactive preparations which affect different pathways of immune and inflammatory response might favorably change the clinical course of these diseases. The data presented in this paper shows that the use of non-specific immunoactive preparations is associated with some beneficial effects in patients with asthma and COPD. Besides assessing the changes in frequency and severity of exacerbations, a potentially very important and under-researched aspect of the effectiveness of immunoactive preparations are the putative effects on the immune system during remission of chronic disease.

From a clinical point of view, the typical moderate and individually variable effect size of immunomodulator therapies indicates that subgroups of responders and non-responders may exist. Confirmation of the presence of these subgroups followed by investigations into the presence of biomarkers would be worthwhile. We need to be open to investigating immune biomarkers, infectious agents as triggers, and the role of the microbiome in order to identify patient populations who may respond best to therapy.

Another rational direction of further research in this area is comparison of the efficiency of non-specific immunoactive drugs in patients with different phenotypes and endotypes of chronic respiratory disease (allergic or non-allergic, eosinophilic or non-eosinophilic, type 2-dependent or type 2-independent, with polyps or without polyps). Randomized placebo-controlled trials stratified by endotypes/phenotypes of disease, should be a considered part of research strategies.

Conventional approaches to the treatment of asthma and COPD are increasingly being brought into question in favor of the 'treatable traits' concept. According to this approach, identification of specific disease features amenable to targeted therapy can be used to personalize and optimize management strategies. Investigating which treatable traits are most amenable to immunomodulatory therapy could be another fertile avenue of research.

There are still unmet needs in the management of chronic lung disease such as asthma and COPD, including an increasing prevalence of disease and unsatisfactory disease control in patients treated with standard drug regimens. The latter might be at least partially explained by relatively low patient compliance associated with the inhalation therapy which is a cornerstone of both asthma and COPD treatment. Existing therapeutic methods in other chronic respiratory airway diseases are outdated (steroids) or cause unacceptable collateral damage (antibiotics). Thus, other alternative and/or complimentary forms of therapy which are well tolerated and easily accepted by patients are welcome.

Therapies targeting the immune system are increasing important throughout modern medicine, and monoclonal antibody therapies targeting type 2 inflammation already have proven efficacy in both

atopic asthma and atopic dermatitis. However, the expense and burden of adverse events currently restrict these therapies to moderate-to-severe cases of disease. Cost effective therapies without an appreciable side-effect burden, such as immunoactive therapies, are likely to solidify a place in this burgeoning therapeutic landscape presuming the high-quality data necessary for wide acceptance are generated.

We hope that the current strong evidence for specific preparations in particular disease areas, alongside ongoing work, will soon be more widely reflected in recommendations issued by large international scientific societies. Continuing research will help to refine recommendations for the clinical use of non-specific immunoactive preparations in chronic airway diseases and focus the attention of practitioners on the most effective drugs with known cellular and molecular mechanisms of action.

Funding

Editorial and writing support was funded by OM Pharma, part of the Vifor Pharma Group; Authors received no financial support for the writing of this manuscript.

Declaration of interest

G Rossi received lecture and consultation fees from: Menarini Group, Lusofarmaco, Abbott, AbbVie, AstraZeneca, Chiesi Farmaceutici, MSD, Polichem S.A., Pierre Fabre, Vifor Pharma Group. R Krenke received support and/or personal fees from Boehringer Ingelheim, Chiesi, AstraZeneca, Polpharma, outside the submitted work. GW Canonica received research grants, lecturer and/or advisory board fees: A. Menarini Diagnostics, ALK, Allergy Therapeutics PLC, AstraZeneca, Boehringer Ingelheim, Chiesi Farmaceutici, Genentech, Guidotti-Malesci, GSK, Hal Allergy Group, Mylan, Merck, MSD, Mundipharma, Novartis, Regeneron Pharmaceuticals, Roche, Sanofi, Sanofi-Genzyme, Stallergenes-Greer, UCB, Uriach, Valeas, Vifor Pharma Group. L van Gerven received a lecture fee from Menarini. O Kalyuzhin received lecture and consultation fees from Abbott, Sandoz, STADA Arzneimittel AG. The authors have no other relevant affiliations or financial involvement with any organization or entity with a financial interest in or financial conflict with the subject matter or materials discussed in the manuscript apart from those disclosed.

Reviewer disclosures

Peer reviewers on this manuscript have no relevant financial or other relationships to disclose.

References

Papers of special note have been highlighted as either of interest (*) or of considerable interest (**) to readers.

1. Bousquet J, Bachert C, Canonica GW, et al. Unmet needs in severe chronic upper airway disease (SCUAD). *Journal of Allergy and Clinical Immunology*. 2009;124(3):428-433. en.
2. Achakulwisut P, Brauer M, Hystad P, et al. Global, national, and urban burdens of paediatric asthma incidence attributable to ambient NO₂ pollution: estimates from global datasets. *The Lancet Planetary Health*. 2019;3(4):e166-e178. doi: 10.1016/S2542-5196(19)30046-4.
3. Passali D, Cingi C, Staffa P, et al. The International Study of the Allergic Rhinitis Survey: outcomes from 4 geographical regions. *Asia Pac Allergy*. 2018;8(1):e7-e7. doi: 10.5415/apallergy.2018.8.e7. PubMed PMID: 29423374; eng.
4. Collaborators GCRD. Global, regional, and national deaths, prevalence, disability-adjusted life years, and years lived with disability for chronic obstructive pulmonary disease and asthma, 1990-2015: a systematic analysis for the Global Burden of Disease Study 2015. *The Lancet Respiratory medicine*. 2017 Sep;5(9):691-706. doi: 10.1016/s2213-2600(17)30293-x. PubMed PMID: 28822787; PubMed Central PMCID: PMC5573769. eng.
5. Jin J-M, Sun Y-C. Allergy and Chronic Obstructive Pulmonary Disease. *Chinese Medical Journal*. 2017;130(17):2017-2020. en.
6. *Jamieson DB, Matsui EC, Belli A, et al. Effects of Allergic Phenotype on Respiratory Symptoms and Exacerbations in Patients with Chronic Obstructive Pulmonary Disease. *American Journal of Respiratory and Critical Care Medicine*. 2013;188(2):187-192. en.
An analysis of two separate cohorts of COPD patients showing that sensitized COPD patients (allergic phenotype) are more likely to wheeze, to have chronic cough/phlegm, more frequent exacerbations, and require more acute doctor visits. An increased number of sensitizations is associated with a higher risk for adverse health outcomes
7. Global Health Estimates 2016: Deaths by Cause, Age, Sex, by Country and by Region, 2000-2016. Geneva: World Health Organization; 2018.
8. Hens G, Vanaudenaerde BM, Bullens DMA, et al. Original article: Sinonasal pathology in nonallergic asthma and COPD: 'united airway disease' beyond the scope of allergy: Sinonasal pathology in nonallergic asthma and COPD. *Allergy*. 2007;63(3):261-267. en.
9. Yii ACA, Tay TR, Choo XN, et al. Precision medicine in united airways disease: A "treatable traits" approach. *Allergy*. 2018;73(10):1964-1978. en.
10. Singh D, Agusti A, Anzueto A, et al. Global Strategy for the Diagnosis, Management, and Prevention of Chronic Obstructive Lung Disease: the GOLD science committee report 2019. LID - 1900164 [pii] LID - 10.1183/13993003.00164-2019 [doi]. (1399-3003 (Electronic)). eng.
11. Global Initiative for Asthma. Global Strategy for Asthma Management and Prevention, 2018. [June 2019]. Available from: www.ginasthma.org
12. Dykewicz MS, Wallace DV, Baroody F, et al. Treatment of seasonal allergic rhinitis. *Annals of Allergy, Asthma & Immunology*. 2017;119(6):489-511. en.
13. Sridhar S, Liu H, Pham T-H, et al. Modulation of blood inflammatory markers by benralizumab in patients with eosinophilic airway diseases. *Respiratory Research*. 2019;20(1):14. en.
14. Rabe KF, Nair P, Brusselle G, et al. Efficacy and Safety of Dupilumab in Glucocorticoid-Dependent Severe Asthma. *New England Journal of Medicine*. 2018;378(26):2475-2485. en.
15. Bachert C, Hellings PW, Mullol J, et al. Dupilumab improves patient-reported outcomes in patients with chronic rhinosinusitis with nasal polyps and comorbid asthma. *The Journal of Allergy and Clinical Immunology: In Practice*. 2019;7(7):2447-2449. en.
16. Masieri S, Cavaliere C Fau - Begvarfaj E, Begvarfaj E Fau - Rosati D, et al. Effects of omalizumab therapy on allergic rhinitis: a pilot study. (2284-0729 (Electronic)). eng.
17. European Academy of Allergy and Clinical Immunology, Global Atlas of Allergy. June 2019.
18. Fardet L, Petersen I, Nazareth I. Common Infections in Patients Prescribed Systemic Glucocorticoids in Primary Care: A Population-Based Cohort Study. *PLOS Medicine*. 2016;13(5):e1002024. en.
19. **Yang M, Zhang Y, Chen H, et al. Inhaled corticosteroids and risk of upper respiratory tract infection in patients with asthma: a meta-analysis. *Infection*. 2019;47(3):377-385. en.
A systematic review showing that the use of inhaled corticosteroids in asthma patients is associated with significantly increased risk of upper respiratory tract infection
20. Di Pasquale MF, Sotgiu G, Gramegna A, et al. Prevalence and Etiology of Community-acquired Pneumonia in Immunocompromised Patients. *Clinical Infectious Diseases*. 2019;68(9):1482-1493. en.
21. Seemungal TAR, Harper-Owen R, Bhowmik A, et al. Detection of rhinovirus in induced sputum at exacerbation of chronic obstructive pulmonary disease. *European Respiratory Journal*. 2000;16(4):677.

22. Bjerregaard A, Laing IA, Poulsen N, et al. Characteristics associated with clinical severity and inflammatory phenotype of naturally occurring virus-induced exacerbations of asthma in adults. *Respiratory Medicine*. 2017;123:34-41. en.
23. Stefan MS, Shieh M-S, Spitzer KA, et al. Association of Antibiotic Treatment With Outcomes in Patients Hospitalized for an Asthma Exacerbation Treated With Systemic Corticosteroids. *JAMA Internal Medicine*. 2019;179(3):333-339. en.
24. Abt Michael C, Osborne Lisa C, Monticelli Laurel A, et al. Commensal Bacteria Calibrate the Activation Threshold of Innate Antiviral Immunity. *Immunity*. 2012;37(1):158-170. en.
25. *Li N, Ma W-T, Pang M, et al. The Commensal Microbiota and Viral Infection: A Comprehensive Review. *Frontiers in Immunology*. 2019;10:1551.
A comprehensive recent review of the relationship between microbiota and infection which covers aspects of immunity which will aid understanding of immunoactive preparations
26. Lee KH, Gordon A, Shedden K, et al. The respiratory microbiome and susceptibility to influenza virus infection. *PLOS ONE*. 2019;14(1):e0207898. en.
27. Strachan DP. Hay fever, hygiene, and household size. *BMJ*. 1989;299(6710):1259-1260. en.
28. **Haahtela T. A biodiversity hypothesis. *Allergy*. 2019:1-12. en.
An important review stressing the key role of biodiversity in natural environments as a fundamental factor leading to the loss of immunoprotective factors. Advice for city planning, food and energy production, and nature conservation based on the biodiversity hypothesis are postulated
29. Rook GAW. Hygiene Hypothesis and Autoimmune Diseases. *Clinical Reviews in Allergy & Immunology*. 2012;42(1):5-15. en.
30. Marsland BJ, Trompette A, Gollwitzer ES. The Gut-Lung Axis in Respiratory Disease. (2325-6621 (Electronic)). eng.
31. Anand S, Mande SS. Diet, Microbiota and Gut-Lung Connection. *Frontiers in Microbiology*. 2018;9:2147.
32. Dang AT, Marsland BJ. Microbes, metabolites, and the gut-lung axis. *Mucosal Immunology*. 2019 2019/07/01;12(4):843-850. doi: 10.1038/s41385-019-0160-6.
33. Esposito S, Soto-Martinez ME, Feleszko W, et al. Nonspecific immunomodulators for recurrent respiratory tract infections, wheezing and asthma in children: a systematic review of mechanistic and clinical evidence. *Current Opinion in Allergy and Clinical Immunology*. 2018;18(3):198-209. en.
A comprehensive systematic clinical and mechanistic review of non-specific immunoactive preparations in pediatric recurrent respiratory disease and wheezing/asthma
34. Keely S, Talley NJ, Hansbro PM. Pulmonary-intestinal cross-talk in mucosal inflammatory disease. *Mucosal Immunology*. 2012 2012/01/01;5(1):7-18. doi: 10.1038/mi.2011.55.
35. Navarro S, Cossalter G, Chiavaroli C, et al. The oral administration of bacterial extracts prevents asthma via the recruitment of regulatory T cells to the airways. *Mucosal Immunology*. 2011;4(1):53-65. en.
36. Feleszko W, Jaworska J, Hamelmann E. Toll-like receptors—novel targets in allergic airway disease (probiotics, friends and relatives). *European Journal of Pharmacology*. 2006;533(1-3):308-318. en.
37. Debock I, Flamand V. Unbalanced Neonatal CD4(+) T-Cell Immunity. *Frontiers in immunology*. 2014;5:393-393. doi: 10.3389/fimmu.2014.00393. PubMed PMID: 25221551; eng.
38. Pulendran B, Artis D. New paradigms in type 2 immunity. *Science*. 2012;337(6093):431-435. doi: 10.1126/science.1221064. PubMed PMID: 22837519; eng.
39. Spisek R, Brazova J, Rozkova D, et al. Maturation of dendritic cells by bacterial immunomodulators. *Vaccine*. 2004 Jul 29;22(21-22):2761-2768. doi: 10.1016/j.vaccine.2004.01.006. PubMed PMID: WOS:000223001900015.
40. Hu X, Zheng W, Wang L, et al. The detailed analysis of the changes of murine dendritic cells (DCs) induced by thymic peptide pidotimod(PTD). *Human Vaccines & Immunotherapeutics*. 2012 Sep;8(9):1250-1258. doi: 10.4161/hv.20579. PubMed PMID: WOS:000310580000021.
41. Bystron J, Hermanova Z, Szotkovska J, et al. Comparison of the effect of ribosomal immunotherapy on plasma levels of total IgE and cytokines IL-4, IL-5, IL-12 and IFN gamma in adult atopic and non-atopic patients during the pollen season. *Clinical Drug Investigation*. 2004 2004;24(12):755-760. doi: 10.2165/00044011-200424120-00007. PubMed PMID: WOS:000225958000006.
42. Gaboriau-Routhiau V, Rakotobe S, Lécuyer E, et al. The Key Role of Segmented Filamentous Bacteria in the Coordinated Maturation of Gut Helper T Cell Responses. *Immunity*. 2009;31(4):677-689. en.
43. Kearney SC, Dziekiewicz M, Feleszko W. Immunoregulatory and immunostimulatory responses of bacterial lysates in respiratory infections and asthma. *Annals of Allergy Asthma & Immunology*. 2015 May;114(5):364-369. doi: 10.1016/j.anai.2015.02.008. PubMed PMID: WOS:000353878500003.
44. Mortaz E, Adcock IM, Folkerts G, et al. Probiotics in the Management of Lung Diseases. *Mediators of Inflammation*. 2013;2013:1-10. en.

45. Papizadeh M, Rohani M, Nahrevanian H, et al. Probiotic characters of Bifidobacterium and Lactobacillus are a result of the ongoing gene acquisition and genome minimization evolutionary trends. *Microbial Pathogenesis*. 2017;111:118-131. en.
46. Obieglo K, van Wijck Y, de Kleijn S, et al. Microorganism-induced suppression of allergic airway disease: novel therapies on the horizon? *Expert Review of Respiratory Medicine*. 2014;8(6):717-730. en.
47. *Feleszko W, Jaworska J, Rha RD, et al. Probiotic-induced suppression of allergic sensitization and airway inflammation is associated with an increase of T regulatory-dependent mechanisms in a murine model of asthma. *Clinical & Experimental Allergy*. 2007;37(4):498-505. en.
A pioneering study demonstrating immunomodulatory effects of orally-applied bacterial immunomodulators (probiotic bacteria) on the development of Treg cells in an animal model of asthma
48. Anatriello E, Cunha M, Nogueira J, et al. Oral feeding of Lactobacillus bulgaricus N45.10 inhibits the lung inflammation and airway remodeling in murine allergic asthma: Relevance to the Th1/Th2 cytokines and STAT6/T-bet. *Cellular Immunology*. 2019;341:103928. en.
49. Lin J, Zhang Y, He C, et al. Probiotics supplementation in children with asthma: A systematic review and meta-analysis: Probiotics in children with asthma. *Journal of Paediatrics and Child Health*. 2018;54(9):953-961. en.
50. Wei X, Jiang P, Liu J, et al. Association between probiotic supplementation and asthma incidence in infants: a meta-analysis of randomized controlled trials. (1532-4303 (Electronic)). eng.
51. Zuccotti G, Meneghin F, Aceti A, et al. Probiotics for prevention of atopic diseases in infants: systematic review and meta-analysis. *Allergy*. 2015;70(11):1356-1371. en.
52. Yin J, Xu B, Zeng X, et al. Broncho-Vaxom in pediatric recurrent respiratory tract infections: A systematic review and meta-analysis. *International Immunopharmacology*. 2018;54:198-209. en.
53. **Boer Gd, J Ž, K S, et al. Bacterial lysate add-on therapy for the prevention of wheezing episodes and asthma exacerbations: a systematic review and meta-analysis [Submitted for publication].
An up-to-date meta-analysis of randomized-controlled trials with the use of bacterial-derived immunostimulants in children and adolescents with wheeze and asthma, showing a decrease in the number of wheezing episodes and asthma exacerbations
54. Namazova-Baranova LS, Alekseeva AA, Kharit SM, et al. Efficacy and safety of pidotimod in the prevention of recurrent respiratory infections in children: a multicentre study. *International Journal of Immunopathology and Pharmacology*. 2014 Jul-Sep;27(3):413-419. PubMed PMID: WOS:000342397100011.
55. Edwards MR, Walton RP, Jackson DJ, et al. The potential of anti-infectives and immunomodulators as therapies for asthma and asthma exacerbations. *Allergy*. 2018;73(1):50-63. en.
56. Morimoto K, Takeshita T, Nanno M, et al. Modulation of natural killer cell activity by supplementation of fermented milk containing Lactobacillus casei in habitual smokers. *Preventive Medicine*. 2005;40(5):589-594. en.
57. Mortaz E, Adcock IM, Ricciardolo FLM, et al. Anti-Inflammatory Effects of Lactobacillus Rahmnosus and Bifidobacterium Breve on Cigarette Smoke Activated Human Macrophages. *PLOS ONE*. 2015;10(8):e0136455. en.
58. Fokkens WJ, Lund VJ, Mullol J, et al. EPOS 2012: European position paper on rhinosinusitis and nasal polyps 2012. A summary for otorhinolaryngologists. *Rhinology journal*. 2012;50(1):1-12.
59. Reh DD, Higgins TS, Smith TL. Impact of tobacco smoke on chronic rhinosinusitis: a review of the literature. *International Forum of Allergy & Rhinology*. 2012;2(5):362-369. en.
60. Krysko O, Holtappels G, Zhang N, et al. Alternatively activated macrophages and impaired phagocytosis of S. aureus in chronic rhinosinusitis: Phagocytosis and macrophage phenotype in CRS. *Allergy*. 2011;66(3):396-403. en.
61. Foreman A, Holtappels G, Psaltis AJ, et al. Adaptive immune responses in Staphylococcus aureus biofilm-associated chronic rhinosinusitis: Immune responses in S. aureus biofilm-associated CRS. *Allergy*. 2011;66(11):1449-1456. en.
62. Soyka MB, Wawrzyniak P, Eiwegger T, et al. Defective epithelial barrier in chronic rhinosinusitis: The regulation of tight junctions by IFN-γ and IL-4. *Journal of Allergy and Clinical Immunology*. 2012;130(5):1087-1096. en.
63. Van Zele T, Gevaert P, Holtappels G, et al. Local immunoglobulin production in nasal polyposis is modulated by superantigens. *Clinical & Experimental Allergy*. 2007;37(12):1840-1847. en.
64. Cervin AU. The Potential for Topical Probiotic Treatment of Chronic Rhinosinusitis, a Personal Perspective. *Frontiers in Cellular and Infection Microbiology*. 2018;7:530.
65. Stubbendieck RM, Straight PD. Multifaceted Interfaces of Bacterial Competition. *Journal of Bacteriology*. 2016;198(16):2145-2155. en.
66. Yang Y, Jing Y, Yang J, et al. Effects of intranasal administration with *Bacillus subtilis* on immune cells in the nasal mucosa and tonsils of piglets. *Experimental and Therapeutic Medicine*. 2018;15:5189-5198.

67. *Mårtensson A, Abolhalaj M, Lindstedt M, et al. Clinical efficacy of a topical lactic acid bacterial microbiome in chronic rhinosinusitis: A randomized controlled trial: Nasal effects of a LAB microbiome in CRSsNP. *Laryngoscope Investigative Otolaryngology*. 2017;2(6):410-416. en.
Although efficacy results were disappointing this paper demonstrates the feasibility of the topical application of probiotics for upper respiratory-tract disease
68. Mukerji SS, Pynnonen MA, Kim HM, et al. Probiotics as adjunctive treatment for chronic rhinosinusitis: A randomized controlled trial. *Otolaryngology–Head and Neck Surgery*. 2009;140(2):202-208. en.
69. Chen J, Zhou Y, Nie J, et al. Bacterial lysate for the prevention of chronic rhinosinusitis recurrence in children. *The Journal of Laryngology & Otology*. 2017;131(6):523-528. en.
70. Heintz B, Schlenter W, Kirsten R, et al. Clinical efficacy of Broncho-Vaxom in adult patients with chronic purulent sinusitis--a multi-centric, placebo-controlled, double-blind study. 1989:530-534. eng.
71. Zagar S, Löfler-Badzek D. Broncho-Vaxom® in Children with Rhinosinusitis: A Double-Blind Clinical Trial. *ORL*. 1988;50(6):397-404. en.
72. Kaplan A. Canadian guidelines for chronic rhinosinusitis: Clinical summary. (1715-5258 (Electronic)). eng.
73. Rosenfeld RM, Piccirillo JF, Chandrasekhar SS, et al. Clinical practice guideline (update): adult sinusitis. (1097-6817 (Electronic)). eng.
74. Orlandi RR, Kingdom TT, Hwang PH. International Consensus Statement on Allergy and Rhinology: Rhinosinusitis Executive Summary: ICAR Executive Summary. *International Forum of Allergy & Rhinology*. 2016;6(S1):S3-S21. en.
75. Felice GD, Barletta B, Butteroni C, et al. Use of Probiotic Bacteria for Prevention and Therapy of Allergic Diseases: Studies in Mouse Model of Allergic Sensitization. *Journal of Clinical Gastroenterology*. 2008;42:S130-S132. en.
76. Özdemir Ö. Various effects of different probiotic strains in allergic disorders: an update from laboratory and clinical data: Various effects of probiotics in allergy. *Clinical & Experimental Immunology*. 2010;160(3):295-304. en.
77. Ren J, Zhao Y, Huang S, et al. Immunomodulatory effect of *Bifidobacterium breve* on experimental allergic rhinitis in BALB/c mice *Experimental and Therapeutic Medicine*. 2018;16:3996-4004.
78. Giovannini M, Agostoni C, Riva E, et al. A Randomized Prospective Double Blind Controlled Trial on Effects of Long-Term Consumption of Fermented Milk Containing *Lactobacillus casei* in Pre-School Children With Allergic Asthma and/or Rhinitis. *Pediatric Research*. 2007;62(2):215-220.
79. Güvenç IA, Muluk NB, Mutlu FŞ, et al. Do Probiotics have a role in the Treatment of Allergic Rhinitis? A Comprehensive Systematic Review and Metaanalysis. *American Journal of Rhinology & Allergy*. 2016;30(5):e157-e175. en.
80. *Zajac AE, Adams AS, Turner JH. A systematic review and meta-analysis of probiotics for the treatment of allergic rhinitis: Probiotics for the treatment of AR. *International Forum of Allergy & Rhinology*. 2015;5(6):524-532. en.
Study showing potential for quality-of-life improvements with probiotics in patients with allergic rhinitis
81. *Peng Y, Li A, Yu L, et al. The Role of Probiotics in Prevention and Treatment for Patients with Allergic Rhinitis: A Systematic Review. *American Journal of Rhinology & Allergy*. 2015;29(4):292-298. en.
Study showing potential for quality-of-life improvements with probiotics in allergic rhinitis
82. Gelardi M, De Luca C, Taliente S, et al. Adjuvant treatment with a symbiotic in patients with inflammatory non-allergic rhinitis. *J Biol Regul Homeost Agents*. 2017;31:201-206.
83. Koatz AM, Coe NA, Cicerán A, et al. Clinical and Immunological Benefits of OM-85 Bacterial Lysate in Patients with Allergic Rhinitis, Asthma, and COPD and Recurrent Respiratory Infections. *Lung*. 2016;194(4):687-697. en.
84. Meng Q, Li P, Li Y, et al. Broncho-vaxom alleviates persistent allergic rhinitis in patients by improving Th1/Th2 cytokine balance of nasal mucosa. *Rhinology journal*. 2019;0(0):0-0.
85. Banche G, Allizond V, Mandras N, et al. Improvement of clinical response in allergic rhinitis patients treated with an oral immunostimulating bacterial lysate: *in vivo* immunological effects. *International Journal of Immunopathology and Pharmacology*. 2007;20(1):129-138. en.
86. Brożek JL, Bousquet J, Agache I, et al. Allergic Rhinitis and its Impact on Asthma (ARIA) guidelines—2016 revision. *Journal of Allergy and Clinical Immunology*. 2017;140(4):950-958. en.
87. Hellings PW, Klimek L, Cingi C, et al. Non-allergic rhinitis: Position paper of the European Academy of Allergy and Clinical Immunology. *Allergy*. 2017;72(11):1657-1665. en.
88. Seidman MD, Gurgel RK, Lin SY, et al. Clinical practice guideline: Allergic rhinitis. (1097-6817 (Electronic)). eng.
89. Manolova V, Flace A, Jeandet P, et al. Biomarkers Induced by the Immunomodulatory Bacterial Extract OM-85: Unique Roles for Peyer's Patches and Intestinal Epithelial Cells. *Journal of Clinical & Cellular Immunology*. 2017;08:494.

90. Dang AT, Pasquali C, Ludigs K, et al. OM-85 is an immunomodulator of interferon-beta production and inflammasome activity. *Scientific Reports*. 2017 Mar 6;7. doi: 10.1038/srep43844. PubMed PMID: WOS:000395621500001.
91. Huber M, Mossmann H, Bessler WG. Th1-orientated immunological properties of the bacterial extract OM-85-BV. *European Journal of Medical Research*. 2005 May 20;10(5):209-217. PubMed PMID: WOS:000229542300007.
92. Luan H, Zhang Q, Wang L, et al. OM85-BV Induced the Productions of IL-1 beta, IL-6, and TNF-alpha via TLR4-and TLR2-Mediated ERK1/2/NF-kappa B Pathway in RAW264.7 Cells. *Journal of Interferon and Cytokine Research*. 2014 Jul;34(7):526-536. doi: 10.1089/jir.2013.0077. PubMed PMID: WOS:000339152400006.
93. Liao J-Y, Zhang T. Influence of OM-85 BV on hBD-1 and immunoglobulin in children with asthma and recurrent respiratory tract infection. *Zhongguo dang dai er ke za zhi = Chinese journal of contemporary pediatrics*. 2014 2014-May;16(5):508-12. PubMed PMID: MEDLINE:24857002.
94. *Roth M, Pasquali C, Stolz D, et al. Broncho Vaxom (OM-85) modulates rhinovirus docking proteins on human airway epithelial cells via Erk1/2 mitogen activated protein kinase and cAMP. *PLOS ONE*. 2017;12(11):e0188010. en.
An ex vivo study on bronchial epithelial cells (BEC), showing that OM-85 significantly reduces rhinovirus-induced BEC death and virus replication via increase of C1q-R and β -defensin as well as reduced expression of ICAM1
95. Strickland DH, Judd S, Thomas JA, et al. Boosting airway T-regulatory cells by gastrointestinal stimulation as a strategy for asthma control. *Mucosal Immunology*. 2011;4(1):43-52. en.
96. *Fu R, Li J, Zhong H, et al. Broncho-Vaxom Attenuates Allergic Airway Inflammation by Restoring GSK3 beta-Related T Regulatory Cell Insufficiency. *Plos One*. 2014 Mar 25;9(3). doi: 10.1371/journal.pone.0092912. PubMed PMID: WOS:000333675600103.
A study showing that oral administration of a bacterial lysate preparation is associated with expansion of Treg cells capable of attenuating airway inflammation in asthmatic mice models
97. Rodrigues A, Gualdi LP, De Souza RG, et al. Bacterial extract (OM-85) with human-equivalent doses does not inhibit the development of asthma in a murine model. *Allergologia Et Immunopathologia*. 2016 Nov-Dec;44(6):504-511. doi: 10.1016/j.aller.2016.04.010. PubMed PMID: WOS:000388157400004.
98. Hua Zhong JW, Yin Yao, Ran Fu, Hang Li, Qingling Fu, Weiping Wen. A bacterial extract of OM-85 Broncho-Vaxom suppresses ovalbumin-induced airway inflammation and remodeling in a mouse chronic allergic asthma model. *Int J Clin Exp Pathol*. 2017;10:8.
99. Lu Y, Li Y, Xu L, et al. Bacterial Lysate Increases the Percentage of Natural Killer T Cells in Peripheral Blood and Alleviates Asthma in Children. *Pharmacology*. 2015 2015;95(3-4):139-144. doi: 10.1159/000377683. PubMed PMID: WOS:000354663600005.
100. Chen ZGJ, J.Z.; Li M; Chen, W.; Chen, Y.F; Chen, F.H.; Chen, H. Effect and analysis of clinical efficacy of immunomodulator on serum levels of IL-4 and IFN-gamma in asthmatic children. *J Sun Yat-sen Univ Med Sci* 2009;30:3. Chinese.
101. Han RF, Li HY, Wang JW, et al. Study on clinical effect and immunologic mechanism of infants capillary bronchitis secondary bronchial asthma treated with bacterial lysates Broncho-Vaxom. *European Review for Medical and Pharmacological Sciences*. 2016 May;20(10):2151-2155. PubMed PMID: WOS:000382459500032.
102. Clinicaltrials.gov: NCT02148796. [cited 2019 November]. Available from: <https://clinicaltrials.gov/ct2/show/NCT02148796>
103. Emmerich B, Pachmann K, Milatovic D, et al. Influence of OM-85 BV on Different Humoral and Cellular Immune Defense Mechanisms of the Respiratory Tract. *Respiration*. 1992;59(3):19-23. en.
104. Zou Y, Chen X, Liu J, et al. Serum IL-1 β and IL-17 levels in patients with COPD: associations with clinical parameters. *International Journal of Chronic Obstructive Pulmonary Disease*. 2017:1247-1254. en.
105. **Pan L, Jiang X-G, Guo J, et al. Effects of OM-85 BV in patients with chronic obstructive pulmonary disease: A systematic review and meta-analysis. *Journal of Clinical Pharmacology*. 2015 Oct;55(10):1086-1092. doi: 10.1002/jcph.518. PubMed PMID: WOS:000360831700003.
Most recent, fully published meta-analysis of trials demonstrating efficacy of OM-85 for exacerbation reduction in COPD
106. Boer G, Braunstahl G. Bacterial lyses as add-on therapy in obstructive lung diseases: A systematic review and a meta-analysis. Abstract TP0797. *European Academy of Allergy and Clinical Immunology (EAACI)*; 01-05 June; Lisbon, Portugal2019.
107. Bisetti A, Ciappi G Fau - Bariffi F, Bariffi F Fau - Catena E, et al. Evaluation of the efficacy of pidotimod in the exacerbations in patients affected with chronic bronchitis. (0004-4172 (Print)). eng.
108. Pozzi E, Dolcetti A, Orlandi O, et al. Pidotimod in the treatment of patients affected by bacterial exacerbations of chronic bronchitis. 1994:1495-1498. eng.
109. Ciaccia A. Pidotimod activity against chronic bronchitis exacerbations. (0004-4172 (Print)). eng.

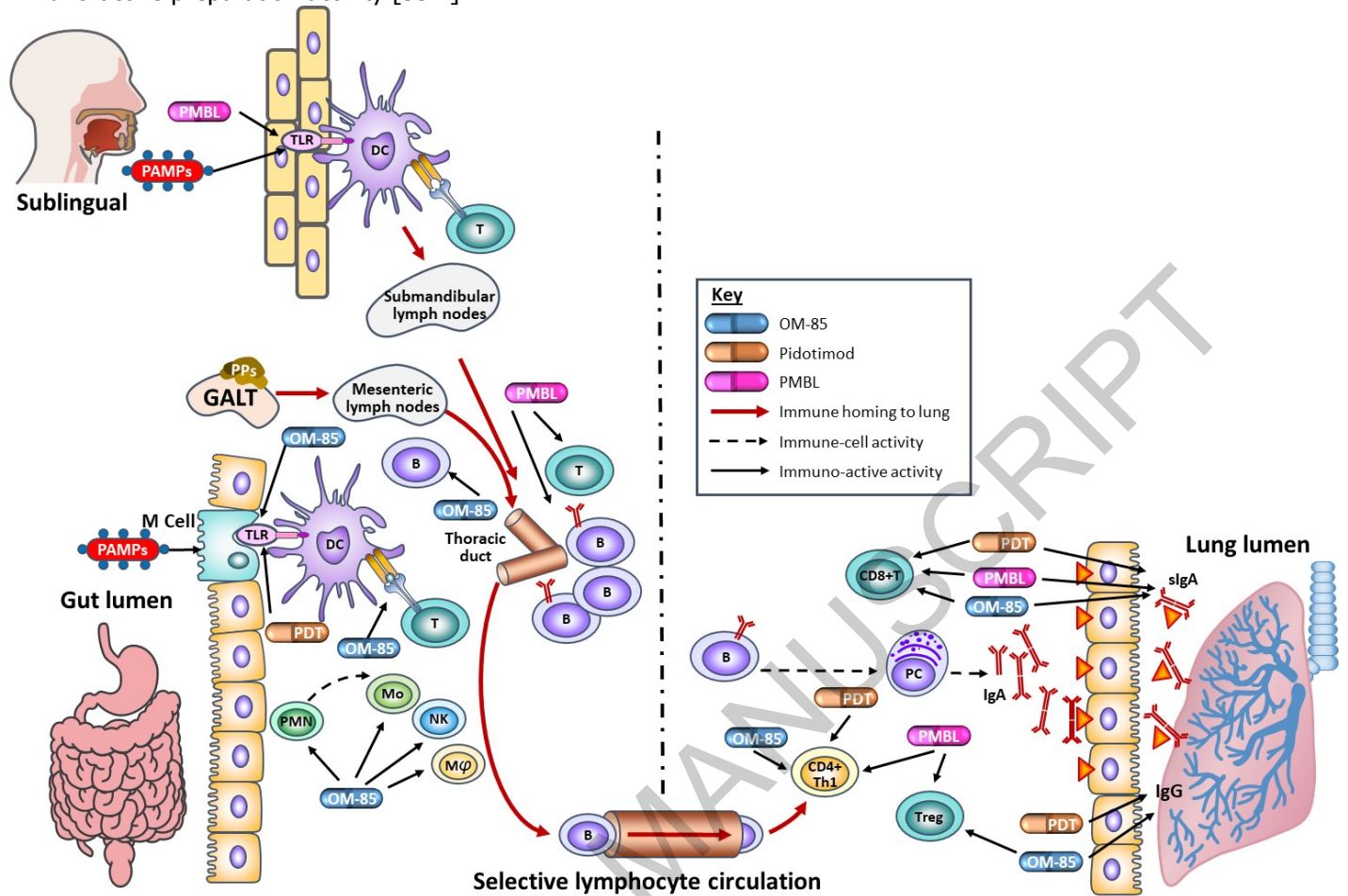
110. Cazzola M, Noschese P, Di Perna F. Value of adding a polyvalent mechanical bacterial lysate to therapy of COPD patients under regular treatment with salmeterol/fluticasone. *Therapeutic advances in respiratory disease*. 2009 Apr;3(2):59-63. doi: 10.1177/1753465809104677. PubMed PMID: MEDLINE:19443519.
111. *Braido F, Melioli G, Cazzola M, et al. Sub-lingual administration of a polyvalent mechanical bacterial lysate (PMBL) in patients with moderate, severe, or very severe chronic obstructive pulmonary disease (COPD) according to the GOLD spirometric classification: A multicentre, double-blind, randomised, controlled, phase IV study (AIACE study: Advanced Immunological Approach in COPD Exacerbation). *Pulmonary Pharmacology & Therapeutics*. 2015;33:75-80. en.
Despite the primary endpoint of this trial not being met, the study design shows that high-quality randomized trials are achievable within the field of immunoactive-preparation research
112. Ricci R, Palmero C, Bazurro G, et al. The administration of a polyvalent mechanical bacterial lysate in elderly patients with COPD results in serological signs of an efficient immune response associated with a reduced number of acute episodes. *Pulmonary Pharmacology & Therapeutics*. 2014 Feb;27(1):109-113. doi: 10.1016/j.pupt.2013.05.006. PubMed PMID: WOS:000331007200016.
113. Bergemann R, Brandt A, Zoellner U, et al. Preventive treatment of chronic bronchitis: a meta-analysis of clinical trials with a bacterial extract (OM-85 BV) and a cost-effectiveness analysis. *Monaldi archives for chest disease = Archivio Monaldi per le malattie del torace*. 1994 Sep;49(4):302-7. PubMed PMID: MEDLINE:8000415.
114. Collet JP, Ducruet T, Haider S, et al. Economic impact of using an immunostimulating agent to prevent severe acute exacerbations in patients with chronic obstructive pulmonary disease. *Canadian respiratory journal*. 2001 2001;8(1):27-33. PubMed PMID: MEDLINE:11223497.
115. Koatz A, Zakin L, Ciceran A. Cost consequence of preventive treatment with OM-85 bacterial lysate compared to the same patients without OM-85 the previous year in allergic rhinitis, asthma and COPD in Argentina. *Value in Health*. 2015;18(7):A498. en.
116. Tao Y, Yuan T, Li X, et al. Bacterial extract OM-85 BV protects mice against experimental chronic rhinosinusitis. *International Journal of Clinical and Experimental Pathology*. 2015 2015;8(6):6800-6806. PubMed PMID: WOS:000359277700088.
117. Triantafillou V, Workman AD, Patel NN, et al. Broncho-Vaxom® (OM-85 BV) soluble components stimulate sinonasal innate immunity. *International Forum of Allergy & Rhinology*. 2019;9(4):370-377. en.
118. Xuan J, Wang L, Yin H, et al. The cost-effectiveness of OM-85 in managing respiratory tract infections in China. *Journal of Medical Economics*. 2015 Mar;18(3):167-172. doi: 10.3111/13696998.2014.971159. PubMed PMID: WOS:000350545300001.
119. Dibildox-Martinez J, Mayorga Butron JL, Macías Fernández LA, et al. Pan-American Clinical Guideline on Rhinosinusitis. *Otolaryngology–Head and Neck Surgery*. 2012;147(2_suppl):P253-P253 en.
120. Han L, Zheng C-P, Sun Y-Q, et al. A bacterial extract of OM-85 Broncho-Vaxom prevents allergic rhinitis in mice. *American Journal of Rhinology & Allergy*. 2014 Mar-Apr;28(2):110-116. doi: 10.2500/ajra.2013.27.4021. PubMed PMID: WOS:000334052100011.
121. Cazzola M, Capuano A, Rogliani P, et al. Bacterial lysates as a potentially effective approach in preventing acute exacerbation of COPD. *Current Opinion in Pharmacology*. 2012 Jun;12(3):300-308. doi: 10.1016/j.coph.2012.01.019. PubMed PMID: WOS:000306160600011.
122. Esposito S, Garziano M, Rainone V, et al. Immunomodulatory activity of pidotimod administered with standard antibiotic therapy in children hospitalized for community-acquired pneumonia. *Journal of Translational Medicine*. 2015 Sep 3;13. doi: 10.1186/s12967-015-0649-z. PubMed PMID: WOS:000360523300002.
123. Carta S, Silvestri M, Rossi GA. Modulation of airway epithelial cell functions by Pidotimod: NF-kappa B cytoplasmic expression and its nuclear translocation are associated with an increased TLR-2 expression. *Italian Journal of Pediatrics*. 2013 May 10;39. doi: 10.1186/1824-7288-39-29. PubMed PMID: WOS:000322573300001.
124. Kim C-H, Kim D-J, Lee S-J, et al. Toll-like receptor 2 promotes bacterial clearance during the initial stage of pulmonary infection with *Acinetobacter baumannii*. *Molecular Medicine Reports*. 2014;9(4):1410-1414. en.
125. Ferrario BE, Garuti S, Braido F, et al. Pidotimod: the state of art. *Clinical and molecular allergy : CMA*. 2015 2015;13(1):8-8. doi: 10.1186/s12948-015-0012-1. PubMed PMID: MEDLINE:25999796.
126. Singh DP, Bagam P, Sahoo MK, et al. Immune-related gene polymorphisms in pulmonary diseases. *Toxicology*. 2017;383:24-39. en.
127. Gaballah HH, Gaber RA, Sharshar RS, et al. NOD2 expression, DNA damage and oxido-inflammatory status in atopic bronchial asthma: Exploring their nexus to disease severity. *Gene*. 2018;660:128-135. en.
128. Gourgiotis D, Papadopoulos NG, Bossios A, et al. Immune modulator pidotimod decreases the in vitro expression of CD30 in peripheral blood mononuclear cells of atopic asthmatic and normal children. *Journal of Asthma*. 2004 2004;41(3):285-287. doi: 10.1081/jas-120026085. PubMed PMID: WOS:000222409900005.

129. Polte T, Behrendt A, Hansen G. Direct evidence for a critical role of CD30 in the development of allergic asthma. *Journal of Allergy and Clinical Immunology*. 2006;118(4):942-948. en.
130. Del Prete G, De Carli M, Almerigogna F, et al. Preferential expression of CD30 by human CD4+ T cells producing Th2-type cytokines. *The FASEB Journal*. 1995;9(1):81-86. en.
131. Manetti R, Annunziato F, Biagiotti R, et al. CD30 expression by CD8+ T cells producing type 2 helper cytokines. Evidence for large numbers of CD8+CD30+ T cell clones in human immunodeficiency virus infection. *The Journal of Experimental Medicine*. 1994;180(6):2407-2411. en.
132. Fu L-Q, Li Y-L, Fu A-K, et al. Pidotimod exacerbates allergic pulmonary infection in an OVA mouse model of asthma. *Molecular Medicine Reports*. 2017;16(4):4151-4158. en.
133. Benetti G, Illeni M, Passera A, et al. Ex vivo evaluation of pidotimod activity in patients with chronic obstructive pulmonary disease. (0004-4172 (Print)). eng.
134. Cazzola M, Anapurapu S, Page CP. Polyvalent mechanical bacterial lysate for the prevention of recurrent respiratory infections: A meta-analysis. *Pulmonary Pharmacology & Therapeutics*. 2012;25(1):62-68. en.
135. Bartkowiak-Emeryk M. The influence of polyvalent mechanical bacterial lysate on immunological parameters in asthmatic children. . Abstract 0078. *European Academy of Allergy and Clinical Immunology Congress*; 17-21 June 2017; Helsinki2017.
136. Lanzilli G, Traggiai E, Braido F, et al. Administration of a polyvalent mechanical bacterial lysate to elderly patients with COPD: Effects on circulating T, B and NK cells. *Immunology Letters*. 2013 Jan;149(1-2):62-67. doi: 10.1016/j.imlet.2012.11.009. PubMed PMID: WOS:000315704000009.
137. Janeczek K, Emeryk A, Rapiejko P. Effect of polyvalent bacterial lysate on the clinical course of pollen allergic rhinitis in children. *Advances in Dermatology and Allergology*. 2019;36(4):504-505.
138. Luca S, Mihaescu T. History of BCG Vaccine. (1841-9038 (Print)). eng.
139. Marchant A, Goetghebuer T, Ota MO, et al. Newborns develop a Th1-type immune response to *Mycobacterium bovis* bacillus Calmette-Guérin vaccination. 1999;163:2249-2255. eng.
140. Shen H, Huang H, Wang J, et al. Neonatal vaccination with *Bacillus Calmette-Guérin* elicits long-term protection in mouse-allergic responses. *Allergy*. 2008;63(5):555-563. en.
141. El-Zein M, Conus F, Benedetti A, et al. Association Between *Bacillus Calmette-Guérin* Vaccination and Childhood Asthma in the Quebec Birth Cohort on Immunity and Health. *American Journal of Epidemiology*. 2017;186(3):344-355 %U <https://academic.oup.com/aje/article/186/3/344/3797130>. en.
142. Holt PG, Jones CA. The development of the immune system during pregnancy and early life. *Allergy*. 2000;55(8):688-697. en.
143. El-Zein M, Parent ME, Benedetti A, et al. Does BCG vaccination protect against the development of childhood asthma? A systematic review and meta-analysis of epidemiological studies. *International Journal of Epidemiology*. 2010;39(2):469-486. en.
144. Linehan MF, Frank TL, Hazell ML, et al. Is the prevalence of wheeze in children altered by neonatal BCG vaccination? *Journal of Allergy and Clinical Immunology*. 2007;119(5):1079-1085 %U <https://linkinghub.elsevier.com/retrieve/pii/S0091674907003612>. en.
145. Linehan MF, Nurmatov U, Frank TL, et al. Does BCG vaccination protect against childhood asthma? Final results from the Manchester Community Asthma Study retrospective cohort study and updated systematic review and meta-analysis. *Journal of Allergy and Clinical Immunology*. 2014;133(3):688-695.e14 %U <https://linkinghub.elsevier.com/retrieve/pii/S0091674913012657>. en.
146. Kim SW, Yeo SW. The Effect of *Bacillus Calmette-Guérin* in a Mouse Model of Allergic Rhinitis. *Otolaryngology-Head and Neck Surgery*. 2007;136(5):720-725 %U <http://journals.sagepub.com/doi/10.1016/j.otohns.2006.10.016>. en.
147. Soysal A, Bahçeciler N, Barlan I, et al. Lack of an inverse association between tuberculosis infection and atopy: By T-cell-based immune assay (RD1-ELISpot). *Pediatric Allergy and Immunology*. 2008;19(8):709-715 %U <http://doi.wiley.com/10.1111/j.1399-3038.2007.00708.x>. en.
148. Cavallo GP, Elia M, Giordano D, et al. Decrease of Specific and Total IgE Levels in Allergic Patients After BCG Vaccination: Preliminary Report. *Archives of Otolaryngology-Head & Neck Surgery*. 2002;128(9):1058 %U <http://archotol.jamanetwork.com/article.aspx?doi=10.1001/archotol.128.9.1058>. en.
149. Ahmadiafshar A, MR P, Moosavinasab N, et al. A study of relation between BCG scar and atopy in schoolchildren of Zanjan City. 2005;4:185-188. eng.
150. Li J, Zhou Z, An J, et al. Absence of Relationships Between Tuberculin Responses and Development of Adult Asthma With Rhinitis and Atopy. *Chest*. 2008;133(1):100-106 %U <https://linkinghub.elsevier.com/retrieve/pii/S0012369215489640>. en.
151. Eifan AO, Akkoc T, Ozdemir C, et al. No association between tuberculin skin test and atopy in a bacillus Calmette-Guérin vaccinated birth cohort. *Pediatric Allergy and Immunology*. 2009;20(6):545-550 %U <http://doi.wiley.com/10.1111/j.1399-3038.2008.00846.x>. en.
152. Li J, Df L, Li S-Y, et al. Efficacy of intramuscular BCG polysaccharide nucleotide on mild to moderate bronchial asthma accompanied with allergic rhinitis: a randomized, double blind, placebo-controlled study. 2005;118:1595-1603. eng.

153. Obihara CC, Beyers N, Gie RP, et al. Inverse association between Mycobacterium tuberculosis infection and atopic rhinitis in children. *Allergy*. 2005;60(9):1121-1125 %U <http://doi.wiley.com/10.1111/j.1398-9995.2005.00834.x>. en.
154. Singh M, Das RR, Kumar L, et al. Bacille Calmette-Guérin Vaccination is Associated with Lower Prevalence of Allergic Diseases in Indian Children. *American Journal of Rhinology & Allergy*. 2013;27(4):e107-e112 %U <http://journals.sagepub.com/doi/10.2500/ajra.2013.27.3940>. en.
155. Mulder WJM, Ochando J, Joosten LAB, et al. Therapeutic targeting of trained immunity. *Nature Reviews Drug Discovery*. 2019;18(7):553-566. en.

ACCEPTED MANUSCRIPT

Figure 1: Illustration of immune activity in the mucosa-associated lymphoid tissue, showing detection of PAMPs at distant mucosa and homing of immune cells to the lung via the shared mucosal immune axes, as well as sites of immuno-active preparation activity [33**]



B, B cell; GALT, gut-associated immune tissue; Mφ, macrophage; Mo, monocyte; NK, natural killer cell; PAMPs, pathogen-associated molecular patterns; PC plasma cell; PMN, polymorphonuclear neutrophil; PPs, Peyer's patches; T, T-cell; Treg, regulatory T cell.

Table 1: Summary of showing improvement, worsening, or no change in clinical outcomes of immunomodulator therapy in asthma/wheezing

Intervention	Study	Design	Studies included	Species included	Population	Outcome: improvement (↔), worsening (↓), or no change (↔)
Probiotics	Lin et al. 2018 [49]	Meta-analysis	11	Mixed cultures of <i>Lactobacillus sp.</i> , <i>Bifidobacterium sp.</i> , and less commonly, <i>Streptococcus sp.</i> ; Mixed cultures of <i>Bifidobacterium sp.</i> ; Single <i>Lactobacillus sp. cultures</i> ^a	Children; N=910	<ul style="list-style-type: none"> ↑ Children with fewer episodes of asthma ↔ C-ACT ↔ Asthmatic symptoms (day or night) ↔ Symptom-free days ↔ Forced expiratory volume ↔ Peak expiratory flow
Probiotics	Wei et al. 2019 [50]	Meta-analysis	19	Single <i>Lactobacillus sp.</i> cultures; Single <i>Bifidobacterium sp.</i> cultures; Mixed cultures of <i>Lactobacillus sp.</i> ; Mixed cultures of <i>Bifidobacterium sp.</i> ; Mixed cultures of <i>Lactobacillus sp.</i> , <i>Bifidobacterium sp.</i> and <i>Propionibacterium</i> ^a	Children; N=5,157	<ul style="list-style-type: none"> ↔ Asthma risk ↔ Wheeze risk ↑ Wheeze incidence in infants with atopic disease
Probiotics	Zuccotti et al. 2015 [51]	Meta-analysis	17	Mixed cultures of <i>Lactobacillus sp.</i> and <i>Bifidobacterium sp.</i> ; single <i>Lactobacillus sp. cultures</i> ^a	Children; N=4,755	<ul style="list-style-type: none"> ↔ Asthma prevention ↔ Wheezing prevention ↓ Eczema development ↔ Rhinoconjunctivitis
OM-85	Yin et al. 2018 [52]	Meta-analysis	8 Wheezing (53 total)	See footnote ^b	Children; n=702 (N=4,851)	<ul style="list-style-type: none"> ↑ Duration of wheezing
OM-85	De Boer et al. 2019 [53**]	Meta-analysis	3 (5 total)	See footnote ^b	Children; N=88	<ul style="list-style-type: none"> ↑ Wheezing episodes and asthma exacerbations
Pidotimod	Namazova-Baranova et al. 2014 [54]	Prospective multicenter trial	N/A	N/A	'Frail' children including those with atopic diseases (≈55%); N=157	<ul style="list-style-type: none"> ↑ Acute respiratory infections ↑ Antibacterial therapy ↑ Disease severity ↑ Complications
PMBL	De Boer et al. 2019 [53**]	Meta-analysis	2 (5 total)	See footnote ^c	Children; N=137	<ul style="list-style-type: none"> ↔ Wheezing episodes and asthma exacerbations

^aSee original references for full list of specific cultures from studies included in meta-analyses; ^b*Haemophilus influenzae* (1 strain), *Streptococcus pneumoniae* (4 strains), *Klebsiella pneumoniae subsp. pneumoniae* (2 strains), *Klebsiella pneumoniae subsp. ozaenae* (1 strains), *Staphylococcus aureus* (6 strains), *Streptococcus pyogenes* (1 strains), *Streptococcus sanguinis* (3 strains), *Moraxella (Branhamella) catarrhalis* (3 strains) [90]; ^c*Staphylococcus*

aureus, *S. pyogenes*, *Streptococcus viridans*, *K. ozaenae*, *H. influenzae* serotype B, *M. catarrhalis* and *S. pneumoniae* [110]. CT/C-ACT, Asthma Control Test/Childhood Asthma Control Test; SABA, short-acting β agonists.

ACCEPTED MANUSCRIPT

Table 3: Summary of showing improvement, worsening, or no change in clinical outcomes of immunomodulator therapy in COPD

Intervention	Study	Design	Studies included	Population	Outcome: improve change (↔)
OM-85	Pan et al., 2015 [105**]	Meta-analysis	5	Adults; N=1,190	<ul style="list-style-type: none"> ↑ Exacerbation rate ↑ Antibiotic use ↔ Hospitalizations ↔ Severity of acute ↔ Total adverse events
OM-85	De Boer et al. 2019 [106]	Meta-analysis	4	Adults; N=1,008	<ul style="list-style-type: none"> ↑ Exacerbations
Pidotimod	Bisetti et al. 1994 [107]	Randomized trial	N/A	Adults; N=181	<ul style="list-style-type: none"> ↑ Infectious relapse ↑ Expectoration clearance
Pidotimod	Pozzi et al. 1994 [108]	Randomized trial	N/A	Adults > 40 years old; N=137	<ul style="list-style-type: none"> ↑ Patients with de ↑ Sputum volume ↑ Faster decrease ↑ Time to recover ↑ Immunity (skin ↑ Investigator glo
Pidotimod	Ciaccia et al., 1994 [109]	Randomized trial	N/A	Adults > 40 years old; N=580	<ul style="list-style-type: none"> ↑ Exacerbations ↑ Time to first exa ↑ Duration of infe ↑ Number of days ↑ Days of work m ↑ Investigator glo
PMBL	Cazzola et al., 2009 [110]	Randomized trial	N/A	Adults ≥ 50 years old; N=63	<ul style="list-style-type: none"> ↑ Exacerbations ↑ Rate of exacerb ↔ Exacerbation sy ↑ Exacerbations re ↑ Hospitalizations ↑ Duration of anti ↑ Duration of cort
PMBL	Braido et al., 2015 [111*]	Randomized trial	N/A	Adults > 40 years old; N=288	<ul style="list-style-type: none"> ↔ 25% reduction i ↔ Time to first exa ↑ Time between f ↑ Days with fever ↑ Days in poor he ↔ All-cause hospit ↑ Hospitalization f
PMBL	Ricci et al., 2014 [112]	Randomized trial	N/A	Adults ≥ 65 years; N=28	<ul style="list-style-type: none"> ↑ Infectious episo

Table 4: Summary of showing improvement, worsening, or no change in clinical outcomes of immunomodulator therapy in allergic and non-allergic rhinitis

ACCEPTED MANUSCRIPT

Intervention	Condition	Study	Design	Studies included	Species included	Population	Outcome: improvement (), worsening (), or no change ()
Probiotics	Allergic Rhinitis	Güvenç et al., 2016 [79]	Meta-analysis	22	Single <i>Lactobacillus sp. cultures</i> ^a ; Single <i>Tetragenococcus sp. culture</i> ; Single <i>Bifidobacterium sp. culture</i> ; Single <i>E. coli sp. culture</i> . Mixed culture of <i>Lactobacillus sp.</i> and live yoghurt ^a	Children and adults; N=2,242	↑ Nasal symptom score ↑ Ocular symptom score ↑ QoL (LP-33 subgroup)
Probiotics	Allergic Rhinitis	Zajac et al., 2015 [80*]	Meta-analysis	23	Sixteen studies used <i>Lactobacillus sp.</i> , six used <i>Bifidobacterium</i> ; <i>E. coli (Nissle 1917)</i> , <i>Tetragenococcus halophilus (Th221)</i> , and <i>Bacillus clausii</i> were used in single studies	Adult; N=1,919	↑ Rhinitis Quality of Life scores ↔ Rhinitis Total Symptom Scores
Probiotics	Allergic Rhinitis	Peng et al., 2015 [81*]	Meta-analysis	11	Mixed cultures of <i>Lactobacillus sp.</i> <i>Bifidobacterium sp.</i> and, less commonly, <i>Propionibacterium</i> , and <i>Streptococcus</i> ; Single <i>Lactobacillus sp. cultures</i> ; Single <i>Bifidobacterium sp. cultures</i> ^a	Adults and children; N=1,527	↑ QoL and symptoms scores
Probiotics	Non-allergic rhinitis	Gelardi et al. 2017 [82]	Randomized trial	N/A	<i>Lactobacillus acidophilus</i> and <i>Bifidobacterium lactis</i>	Adults; N=93	↑ Patients with nasal obstruction ↑ Patients with rhinorrhea ↑ Patients with endoscopic signs
OM-85	Allergic Rhinitis	Koatz et al., 2016 [83]	Prospective trial	N/A	See footnote ^b	Adult; N=84 (allergic rhinitis n=29)	↔ Number of exacerbations ↑ RTIs
OM-85	Allergic Rhinitis	Meng et al. 2019 [84]	Randomized trial	N/A	See footnote ^b	Adult; N=60 (9 patients with mild asthma)	↑ Medication use ↑ Nasal symptoms scores ↑ Itching

							symptom scores ↑ Type 2/Type 1 cytokine balance Eosinophils
PMBL	Allergic Rhinitis	Banche et al. 2007 [85]	Randomized trial	N/A	See footnote ^c	Adult; N=41	↑ Nasal blockage and rhinorrhea Ocular symptoms ↑ Asthmatic symptoms

^aSee original references for full list of specific cultures from studies included in meta-analyses;

^b*Haemophilus influenzae* (1 strain), *Streptococcus pneumoniae* (4 strains), *Klebsiella pneumoniae subsp. pneumoniae* (2 strains), *Klebsiella pneumoniae subsp. ozaenae* (1 strains), *Staphylococcus aureus* (6 strains), *Streptococcus pyogenes* (1 strains), *Streptococcus sanguinis* (3 strains), *Moraxella (Branhamella) catarrhalis* (3 strains) [90]; ^c*Staphylococcus aureus*, *S. pyogenes*, *Streptococcus viridans*, *K. ozaenae*, *H. influenzae serotype B*, *M. catarrhalis* and *S. pneumoniae* [110]. BCG-PSN, BCG polysaccharide nucleotide

ACCEPTED MANUSCRIPT